

Review Article

Complications of Corneal Collagen Cross-Linking

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Received 28 February 2011; Revised 17 October 2011; Accepted 20 October 2011

Academic Editor: Antonio Leccisotti

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Cross-linking of corneal collagen (CXL) is a promising approach for the treatment of keratoconus and secondary ectasia. Several long-term and short-term complications of CXL have been studied and documented. The possibility of a secondary infection after the procedure exists because the patient is subjected to epithelial debridement and the application of a soft contact lens. Formation of temporary corneal haze, permanent scars, endothelial damage, treatment failure, sterile infiltrates, and herpes reactivation are the other reported complications of this procedure. Cross-linking is a low-invasive procedure with low complication and failure rate but it may have direct or primary complications due to incorrect technique application or incorrect patient's inclusion and indirect or secondary complications related to therapeutic soft contact lens, patient's poor hygiene, and undiagnosed concomitant ocular surface diseases.

1. Introduction

Keratoconus is a progressive, bilateral, often asymmetrical, and noninflammatory corneal ectasia. Prevalence of keratoconus is 1 : 2000 [1] and is usually diagnosed during the second and third decade of life. Currently available treatments for keratoconus (rigid contact lens, lamellar Keratoplasty, intacs) largely involve interventions which are done for tectonic, optical, or refractive purpose. Unfortunately, neither of those options treats the underlying cause of ectasia, and therefore cannot stop the progression of keratoconus.

Corneal collagen cross-linking (CXL) based on the combined use of the photosensitizer riboflavin and UVA light of 370 nm was introduced by Wollensak et al. from Germany in 2003. CXL is the only available treatment directed at the underlying pathology in keratoconic cornea, which is stromal biomechanical and structural instability leading to progressive ectasia. CXL induces covalent inter- and intrafibrillar collagen cross-links creating an increase in biomechanical rigidity of human cornea by about 300%. The cross-linking effect is maximal only in the anterior stroma. Corneal collagen cross-linking (CXL) is currently under investigation to determine if it can slow, stabilize, or even possibly reverse the progression of corneal ectasia in patients with

keratoconus [2]. The present paper is a review of literature on CXL complications.

2. Corneal Collagen Cross-Linking with Riboflavin and UVA

The main indication for CXL in ophthalmology has been the management of corneal ectasia, such as halting the progression of keratoconus. In addition, CXL has been proposed as a treatment modality for iatrogenic keratectasia [3], infectious melting keratitis [4], and bullous keratopathy [5]. The latter application utilizes the antioedematous effect of cross-linkage on the stroma. CXL with riboflavin and UVA has been sequentially combined with other modalities, namely, intrastromal ring segments [6] and photorefractive keratectomy (PRK) [7] for the treatment of keratoconus.

UVA irradiation can cause keratocyte and corneal endothelial cell destruction or death, as well as possible lens and retinal damage [8] as it has a toxic effect on cell viability, but there have been no reported complications on the endothelial cell count, lens, or retina due to the limitation of UVA transmission through the cornea [9]. It had also been suggested that CXL treatment be restricted to the anterior 250 μm to 350 μm of the stroma. Thus, CXL is not

recommended for patients whose corneas are thinner than 400 μm [10] because 85% to 90% of the UVA radiation is absorbed in the anterior 400 μm of the cornea; the procedure should not harm the patient's corneal endothelium, lens, and retina [11].

3. Technique

A standard CXL procedure begins with the administration of an anaesthetic, followed by debridement of the central 7 mm to 9 mm of the cornea to allow uniform diffusion of the riboflavin into the stroma [11]. Next, riboflavin 0.1% suspended in a dextran T500 20% solution is applied and allowed to permeate the cornea before UVA irradiation. UVA radiation of 370 nm wavelength and an irradiance of 3 mW/cm² at a distance of 5.4 mm from the cornea is applied for a period of 30 min, delivering a dose of 5.4 J/cm² [12]. Antibiotic eye drops are instilled as prophylaxis and a bandage contact lens is inserted, which is then removed at the followup visit once epithelial healing is complete.

4. Complications of CXL

Several long-term and short-term complications of CXL have been studied and documented [13, 14] which may be direct or primary due to incorrect technique application or incorrect patient's inclusion or indirect or secondary complications related to therapeutic soft contact lens, patient's poor hygiene, and undiagnosed concomitant ocular surface diseases (dry eye, blepharitis, etc.).

4.1. Postoperative Infection/Ulcer. Debriding the corneal epithelium theoretically exposes the cornea to microbial infection. Bacterial keratitis has been reported 3 days following treatment in which scraping revealed an *E. coli* infection [15]. Acanthamoeba keratitis due to eye washing under tap water as the patient was unaware of a bandage contact lens being inserted has been reported [16]. Poor contact lens hygiene resulting in polymicrobial keratitis caused by *streptococcus salivarius*, *streptococcus oralis*, and coagulase-negative *staphylococcus* sp. has been reported recently [17]. A patient with no history of herpetic keratitis developed herpes simplex keratitis geographical ulcer and iritis five days after treatment [18]. *Staphylococcus epidermidis* keratitis has also been reported 2 days after treatment [19]. Diffuse lamellar keratitis (stage 3) has been reported following treatment in a case of post-LASIK ectasia [20]. Severe keratitis with patient's contact lens and cornea scrapings positive for *pseudomonas aeruginosa* has also been reported recently [21]. Reactivated herpetic keratitis and neurodermatitis have also been reported following CXL [18, 22]. One study reported four cases of severe keratitis in a group of 117 keratoconic eyes treated with standard CXL [23].

Keratitis can occur following CXL because of presence of an epithelial defect, use of soft bandage contact lens, and topical corticosteroids in the immediate postoperative period. In cases of corneal infection after CXL, contact with the infectious agent likely occurred during the early

postoperative period rather than during surgery because CXL not only damages keratocytes, but it also kills bacteria and fungi. This effect is used to advantage when CXL is performed for infectious keratitis.

4.2. Corneal Haze. In a recently published retrospective study of 163 eyes with grade I–III keratoconus, approximately 9% of the 127 patients developed clinically significant haze after 1-year followup. The subset of patients developing steroid resistant haze appeared to have more advanced keratoconus, as reflected in a lower mean corneal thickness and higher keratometry value of the apex compared with the control group [24]. An older age, grade III or IV keratoconus (according to Krumeich's classification), and preoperative reticular pattern of stromal microstriae observed preoperatively by in vivo confocal microscopy [14] are considered risk factors for corneal haze post cross-linking. Advanced keratoconus should be considered at higher risk of haze development after CXL due to low corneal thickness and high corneal curvature [24].

After collagen cross-linking using riboflavin and UV-A, a lacunar honeycomb-like hydration pattern can be found in the anterior stroma with the maximum cross-linking effect, which is because of the prevention of interfibrillar cross-linking bonds in the positions of the apoptotic keratocytes [25]. The polygonal cross-linking network might contribute favorably to the biomechanical elasticity of the cross-linked cornea and to the demarcation of the anterior stroma after CXL on biomicroscopy [25], thus making lacunar edema a positive sign of efficient cross-linking. Another study documented stromal haze in 5 of 44 patients within 6 months of undergoing CXL. There has been a debate as to whether stromal haze is a normal finding after CXL because of its frequency [26].

Koller et al. [27] evaluated anterior stromal haze, which was graded on a scale used in cases after PRK [28]; the mean grade was 0.78, 0.18, and 0.06 at 1 month, 6 months, and 12 months, respectively. Previous confocal microscopy studies [26] report that a dense extracellular matrix compatible with clinical haze forms between 2 months and 3 months postoperatively.

The haze after CXL differs from the haze after PRK in stromal depth. Whereas haze after PRK is strictly subepithelial, haze after CXL extends into the anterior stroma to approximately 60% depth, which is on average equal to an absolute depth of 300 μm . Haze after CXL is different in clinical character from haze after other procedures, such as excimer laser photorefractive keratotomy. The former is a dustlike change in the corneal stroma or a midstromal demarcation line, whereas the latter has a more reticulated subepithelial appearance [29]. The haze may be associated with the depth of CXL into the stroma as well as the amount of keratocyte loss [26, 27].

Greenstein et al. [30] studied the natural course after CXL and found a significant postoperative increase in haze measured by both Scheimpflug densitometry and slit lamp assessment. The increase peaked at 1 month and plateaued between 1 month and 3 months. Between 3 months and

6 months, the cornea began to clear and there was a significant decrease in CXL-associated corneal haze which usually does not require treatment except for some low dose steroid medication in some cases. From 6 months to 1 year postoperatively, there continued to be a decrease in haze measurements. Typically late permanent scarring should be differentiated from the early postoperative temporary haze [31] which is often paracentral and compatible with good visual results. It may not be actually related to CXL itself but rather to the ongoing disease process and corneal remodeling.

Haze formation after CXL may be a result of back-scattered and reflected light, which decreases corneal transparency [32]. In vitro and ex vivo studies [33, 34] show that CXL leads to an immediate loss of keratocytes in the corneal stroma. In a confocal microscopy study, Mazzotta et al. [35] found that in eyes with keratoconus, activated keratocytes repopulated the corneal stroma starting at 2 months and that the repopulation was almost complete at 6 months. It is possible that these activated keratocytes contribute to the development of CXL-associated corneal haze. Other factors that may contribute to CXL-associated corneal haze include stromal swelling pressure changes [36], proteoglycan-collagen interactions [37], and glycosaminoglycan hydration [38]. Further study is needed to elucidate the pathophysiology of the development and time course of CXL-associated corneal haze.

4.3. Endothelial Damage. The endothelial damage threshold was shown to be at an irradiance of 0.35 mW/cm^2 , which is approximately twice compared with the 0.18 mW/cm^2 that reaches the corneal endothelium when using the currently recommended protocol [10]. It may be due to a stromal thickness less than $400 \mu\text{m}$ or incorrect focusing. If the procedure is done on a thinner cornea, it may lead to perforation. The recommended safety criteria must be observed because UV irradiation has potential to damage various intraocular structures.

4.4. Peripheral Sterile Infiltrates. Sterile corneal stromal infiltrates occur as a result of enhanced cell-mediated immunity to staphylococcal antigens deposited at high concentrations in areas of static tear pooling [39]. Sterile infiltration after CXL may be related to staphylococcal antigen deposition in areas of static tear pooling beneath the bandage contact lens [39].

4.5. Herpes Reactivation. Reactivation of HSV has been reported after emotional stress, trauma, fever, and laser surgery. These established clinical triggers are thought to be mediated by the adrenergic and sensory nervous systems. Exposure to UV light can also induce oral and genital herpes in humans and ocular herpes in animal models. Development of herpes keratitis and iritis after riboflavin-UVA treatment has been reported [18]. It seems that UVA light could be a potent stimulus to trigger/induce reactivation of latent HSV infections even in patients with no history of clinical herpes virus ocular infections. Significant corneal

epithelial/stromal trauma or actual damage of the corneal nerves could be the mechanism of HSV reactivation. The use of topical corticosteroids and mechanical trauma caused by epithelial debridement may be additional risk factors. Prophylactic systemic antiviral treatment in patients with a history of herpetic disease after cross-linking with UVA might decrease the possibility of recurrence.

4.6. Treatment Failure. CXL failure is largely defined as keratoconic progression following treatment. One study of 117 eyes from 99 patients who underwent CXL documented a failure rate of 7.6% at one-year followup [27]. The results also indicated that 2.9% of eyes lost two or more lines of snellen visual acuity. Age older than 35 years, cornea thickness $<400 \mu\text{m}$, and a preoperative CDVA better than 20/25 were identified as significant risk factors for complication. A high preoperative maximum keratometry reading was a significant risk factor for failure. Sterile infiltrates were seen in 7.6% of eyes and central stromal scars in 2.8%. The researchers concluded that changing the inclusion criteria may significantly reduce the complications and failures of CXL. Risk factors for CXL failure included a preoperative patient age of 35 years or older, spectacle-corrected visual acuity better than 20/25, and a maximum keratometry reading greater than 58.00 D [27].

5. Conclusion

Apart from haze and stromal hyperdensity after CXL with early or late onset as direct complication of the treatment, no other direct or primary complications of the procedure have been reported. Complications described in the literature are in the major part of indirect origin (infections, therapeutic contact lens, previous surgery (lasik), coexisting disorders of ocular surface, incorrect patient inclusion in the treatment, technical problems with UVA solid state emitter, wrong technique application, bad focusing, tilting, defocus, etc.). Therefore, only surgeons with sufficient experience in the management of corneal wound healing should perform this procedure. More studies are necessary to identify rare complications and to establish a list of indications regarding patient age, diagnosis, and the stage of keratectasia. The role of the UV light on the immune mechanisms of the cornea and its effect on corneal wound healing warrant further investigation. Repeat cross-linking treatments may become necessary in the long term. Considering that the turnover rate of stromal collagen fibres is several years, prospective studies with a followup of at least eight to ten years will be necessary.

References

- [1] Y. S. Rabinowitz, "Keratoconus," *Survey of Ophthalmology*, vol. 42, no. 4, pp. 297–319, 1998.
- [2] E. Spörl, M. Huhle, M. Kasper, and T. Seiler, "Artificial stiffening of the cornea by induction of intrastromal cross-links," *Ophthalmology*, vol. 94, no. 12, pp. 902–906, 1997.
- [3] H. P. Iseli, M. A. Thiel, F. Hafezi, J. Kampmeier, and T. Seiler, "Ultraviolet a/riboflavin corneal cross-linking for infectious

- keratitis associated with corneal melts," *Cornea*, vol. 27, no. 5, pp. 590–594, 2008.
- [4] G. Wollensak, H. Aurich, C. Wirbelauer, and D. T. Pham, "Potential use of riboflavin/UVA cross-linking in bullous keratopathy," *Ophthalmic Research*, vol. 41, no. 2, pp. 114–117, 2009.
 - [5] N. Ehlers and J. Hjortdal, "Riboflavin-ultraviolet light induced cross-linking in endothelial decompensation," *Acta Ophthalmologica*, vol. 86, no. 5, pp. 549–551, 2008.
 - [6] A. J. Kanellopoulos and P. S. Binder, "Collagen cross-linking (CCL) with sequential topography-guided PRK: a temporizing alternative for keratoconus to penetrating keratoplasty," *Cornea*, vol. 26, no. 7, pp. 891–895, 2007.
 - [7] A. Caporossi, S. Baiocchi, C. Mazzotta, C. Traversi, and T. Caporossi, "Parasurgical therapy for keratoconus by riboflavin-ultraviolet type A rays induced cross-linking of corneal collagen. Preliminary refractive results in an Italian study," *Journal of Cataract and Refractive Surgery*, vol. 32, no. 5, pp. 837–845, 2006.
 - [8] E. Spoerl, M. Mrochen, D. Sliney, S. Trokel, and T. Seiler, "Safety of UVA-riboflavin cross-linking of the cornea," *Cornea*, vol. 26, no. 4, pp. 385–389, 2007.
 - [9] P. Vinciguerra, E. Albè, A. M. Mahmoud, S. Trazza, F. Hafezi, and C. J. Roberts, "Intra- and postoperative variation in ocular response analyzer parameters in keratoconic eyes after corneal cross-linking," *Journal of Refractive Surgery*, vol. 26, no. 9, pp. 669–676, 2010.
 - [10] G. Wollensak, E. Spoerl, M. Wilsch, and T. Seiler, "Endothelial cell damage after riboflavin-ultraviolet-A treatment in the rabbit," *Journal of Cataract and Refractive Surgery*, vol. 29, no. 9, pp. 1786–1790, 2003.
 - [11] G. Wollensak, "Crosslinking treatment of progressive keratoconus: new hope," *Current Opinion in Ophthalmology*, vol. 17, no. 4, pp. 356–360, 2006.
 - [12] G. Wollensak, E. Spoerl, and T. Seiler, "Riboflavin/ultraviolet-A-induced collagen crosslinking for the treatment of keratoconus," *American Journal of Ophthalmology*, vol. 135, no. 5, pp. 620–627, 2003.
 - [13] F. Raiskup-Wolf, A. Hoyer, E. Spoerl, and L. E. Pillunat, "Collagen crosslinking with riboflavin and ultraviolet-A light in keratoconus: long-term results," *Journal of Cataract and Refractive Surgery*, vol. 34, no. 5, pp. 796–801, 2008.
 - [14] A. Caporossi, C. Mazzotta, S. Baiocchi, and T. Caporossi, "Long-term results of riboflavin ultraviolet a corneal collagen cross-linking for keratoconus in Italy: the Siena Eye Cross Study," *American Journal of Ophthalmology*, vol. 149, no. 4, pp. 585–593, 2010.
 - [15] M. Pollhammer and C. Cursiefen, "Bacterial keratitis early after corneal crosslinking with riboflavin and ultraviolet-A," *Journal of Cataract and Refractive Surgery*, vol. 35, no. 3, pp. 588–589, 2009.
 - [16] P. Rama, F. Di Matteo, S. Matuska, G. Paganoni, and A. Spinelli, "Acanthamoeba keratitis with perforation after corneal crosslinking and bandage contact lens use," *Journal of Cataract and Refractive Surgery*, vol. 35, no. 4, pp. 788–791, 2009.
 - [17] K. V. Zamora and J. J. Males, "Polymicrobial keratitis after a collagen cross-linking procedure with postoperative use of a contact lens," *Cornea*, vol. 28, no. 4, pp. 474–476, 2009.
 - [18] G. D. Kymionis, D. M. Portaliou, D. I. Bouzoukis et al., "Herpetic keratitis with iritis after corneal crosslinking with riboflavin and ultraviolet A for keratoconus," *Journal of Cataract and Refractive Surgery*, vol. 33, no. 11, pp. 1982–1984, 2007.
 - [19] J. J. Pérez-Santonja, A. Artola, J. Javaloy, J. L. Alió, and J. L. Abad, "Microbial keratitis after corneal collagen crosslinking," *Journal of Cataract and Refractive Surgery*, vol. 35, no. 6, pp. 1138–1140, 2009.
 - [20] G. D. Kymionis, D. I. Bouzoukis, V. F. Diakonis, D. M. Portaliou, A. I. Pallikaris, and S. H. Yoo, "Diffuse lamellar keratitis after corneal crosslinking in a patient with post-laser in situ keratomileusis corneal ectasia," *Journal of Cataract and Refractive Surgery*, vol. 33, no. 12, pp. 2135–2137, 2007.
 - [21] N. Sharma, P. Maharana, G. Singh, and J. S. Titiyal, "Pseudomonas keratitis after collagen crosslinking for keratoconus: case report and review of literature," *Journal of Cataract and Refractive Surgery*, vol. 36, no. 3, pp. 517–520, 2010.
 - [22] E. Coskunseven, M. R. Jankov, and F. Hafezi, "Contralateral eye study of corneal collagen cross-linking with riboflavin and UVA irradiation in patients with keratoconus," *Journal of Refractive Surgery*, vol. 25, no. 4, pp. 371–376, 2009.
 - [23] C. Koppen, J. C. Vryghem, L. Gobin, and M. J. Tassignon, "Keratitis and corneal scarring after UVA/riboflavin cross-linking for keratoconus," *Journal of Refractive Surgery*, vol. 25, no. 9, pp. S819–S823, 2009.
 - [24] F. Raiskup, A. Hoyer, and E. Spoerl, "Permanent corneal haze after riboflavin-UVA-induced cross-linking in keratoconus," *Journal of Refractive Surgery*, vol. 25, no. 9, pp. S824–S828, 2009.
 - [25] G. Wollensak and H. Herbst, "Significance of the lacunar hydration pattern after corneal cross linking," *Cornea*, vol. 29, no. 8, pp. 899–903, 2010.
 - [26] C. Mazzotta, A. Balestrazzi, S. Baiocchi, C. Traversi, and A. Caporossi, "Stromal haze after combined riboflavin-UVA corneal collagen cross-linking in keratoconus: in vivo confocal microscopic evaluation," *Clinical and Experimental Ophthalmology*, vol. 35, no. 6, pp. 580–582, 2007.
 - [27] T. Koller, M. Mrochen, and T. Seiler, "Complication and failure rates after corneal crosslinking," *Journal of Cataract and Refractive Surgery*, vol. 35, no. 8, pp. 1358–1362, 2009.
 - [28] K. D. Hanna, Y. M. Pouliquen, G. O. Waring, M. Savoldelli, F. Fantes, and K. P. Thompson, "Corneal wound healing in monkeys after repeated excimer laser photorefractive keratectomy," *Archives of Ophthalmology*, vol. 110, no. 9, pp. 1286–1291, 1992.
 - [29] J. D. Carr, R. Patel, and P. S. Hersh, "Management of late corneal haze following photorefractive keratectomy," *Journal of Refractive Surgery*, vol. 11, no. 3, pp. S309–S313, 1995.
 - [30] S. A. Greenstein, K. L. Fry, J. Bhatt, and P. S. Hersh, "Natural history of corneal haze after collagen crosslinking for keratoconus and corneal ectasia: scheimpflug and biomicroscopic analysis," *Journal of Cataract and Refractive Surgery*, vol. 36, no. 12, pp. 2105–2114, 2010.
 - [31] L. S. Lim, R. Beuerman, L. Lim, and D. T. H. Tan, "Late-onset deep stromal scarring after riboflavin—UV-A corneal collagen cross-linking for mild keratoconus," *Archives of Ophthalmology*, vol. 129, no. 3, pp. 360–362, 2011.
 - [32] C. J. Connon, J. Marshall, A. L. Patmore, A. Brahma, and K. M. Meek, "Persistent haze and disorganization of anterior stromal collagen appear unrelated following phototherapeutic keratectomy," *Journal of Refractive Surgery*, vol. 19, no. 3, pp. 323–332, 2003.
 - [33] G. Wollensak, E. Spoerl, M. Wilsch, and T. Seiler, "Keratocyte apoptosis after corneal collagen cross-linking using riboflavin/UVA treatment," *Cornea*, vol. 23, no. 1, pp. 43–49, 2004.
 - [34] J. S. Dhaliwal and S. C. Kaufman, "Corneal collagen cross-linking: a confocal, electron, and light microscopy study of eye bank corneas," *Cornea*, vol. 28, no. 1, pp. 62–67, 2009.

- [35] C. Mazzotta, A. Balestrazzi, C. Traversi et al., "Treatment of progressive keratoconus by riboflavin-UVA-induced cross-linking of corneal collagen: ultrastructural analysis by Heidelberg retinal tomograph II in vivo confocal microscopy in humans," *Cornea*, vol. 26, no. 4, pp. 390–397, 2007.
- [36] C. H. Dohlman, B. O. Hedbys, and S. Mishima, "The swelling pressure of the corneal stroma," *Investigative Ophthalmology*, vol. 1, pp. 158–162, 1962.
- [37] Y. M. Michelacci, "Collagens and proteoglycans of the corneal extracellular matrix," *Brazilian Journal of Medical and Biological Research*, vol. 36, no. 8, pp. 1037–1046, 2003.
- [38] G. Wollensak, H. Aurich, D. T. Pham, and C. Wirbelauer, "Hydration behavior of porcine cornea crosslinked with riboflavin and ultraviolet A," *Journal of Cataract and Refractive Surgery*, vol. 33, no. 3, pp. 516–521, 2007.
- [39] R. I. Angunawela, F. Arnalich-Montiel, and B. D. S. Allan, "Peripheral sterile corneal infiltrates and melting after collagen crosslinking for keratoconus," *Journal of Cataract and Refractive Surgery*, vol. 35, no. 3, pp. 606–607, 2009.