Oxidant/Antioxidant Imbalance and the Risk of Alzheimer's Disease

Ahmed E. Abdel Moneim^{*}

Department of Zoology & Entomology, Faculty of Science, Helwan University, Cairo, Egypt

Abstract: Alzheimer's disease (AD) is the most common form of dementia characterized by progressive loss of memory and other cognitive functions among older people. Senile plaques and neurofibrillary tangles are the most hallmarks lesions in the brain of AD in addition to neurons loss. Accumulating evidence has shown that oxidative stress-induced damage may play an important role in the initiation and progression of AD pathogenesis. Redox impairment occurs when there is an imbalance between the production and quenching of free radicals from oxygen species. These reactive oxygen species augment the formation and aggregation of amyloid- β and tau protein hyperphosphorylation and vice versa. Currently, there is no available treatments can modify the disease. However, wide varieties



of antioxidants show promise to delay or prevent the symptoms of AD and may help in treating the disease. In this review, the role of oxidative stress in AD pathogenesis and the common used antioxidant therapies for AD will summarize.

Keywords: Alzheimer's disease, antioxidants, oxidative stress.

ALZHEIMER'S DISEASE

Alzheimer's disease (AD) is the most common form of dementia characterized by progressive loss of memory and other cognitive functions among older people, AD is placing a considerable emotional and financial burden on patients, their families, caregivers and society, as more people live long enough to become affected. AD begins slowly. It first affects the brain regions of CA1 region of the hippocampus, prefrontal cortex and pyramidal cells in lamina II of the entorhinal cortex that control memory, thought and language. Over time, symptoms get worse. Patients may not recognize family members or have trouble writing, reading or even speaking. AD typically begins after age 60 with death occurring on average a decade or so passes after diagnosis [1].

Pathology

Histopathology of post-mortem brains obtained from AD clinically characterized patients provided the first clues to the mechanisms of disease and potential interventions. It led to the description of the disease a century ago by Alois Alzheimer [1], and the identification of the AD hallmark lesions. The histopathological changes include extracellular deposits of amyloid- β (A β) forming senile plaques and intracellular neurofibrillary tangles (NFT) formed by accumulation of abnormal hyperphosphorylated filaments of tau in pyramidal neurons. Besides these features, a large body of evidence indicates prominent activation of inflammatory processes and the innate immune response activation. Classic senile plaques are spherical structures consisting of a central core of A β fibrous protein that is surrounded by degenerating or

dystrophic nerve endings. The A β protein contains a 40 or 42 amino acid peptide of A β that is derived from proteolytic processing of a larger amyloid precursor protein (APP) molecule via two pathways: the α pathway and the β pathway. APP is degraded by α -secretase to produce a nonamyloidogentic molecules, whereas the sequential enzymatic actions of beta-site APP-cleaving enzyme 1 (BACE-1), a β secretase, generated small APPB (sAPPB) in the extracullar space. sAPP β is subsequently degraded by γ -secretase, a protein complex contains presenilin 1 at its catalytic core, to release AB and APP intracellular C-terminal domain (AICD) [2]. It is believed that the most toxic A β 40–42 peptides are resulted by the abnormal processing of the APP molecule [3]. However, the produced $A\beta$ is degraded by many enzymes includes, but not limited to, insulin-degrading enzyme (IDE) and neprilysin (NEP). NEP and IDE are reduced in AD [4]. The imbalance between AB production and clearance causes $A\beta$ to accumulate in the extracellular space. AB1-42 readily forms insoluble clumps and initiates a cascade of events leading to apoptosis and neuronal dysfunction or death. This process called the amyloid hypothesis (Fig. 1).

On the other hand, NFT is a phosphorylated tau protein (p Tau) and found in the intracellular space of the neurons and are composed of paired helical filaments and straight filaments of hyperphosphorylated microtubule-associated protein (MAP), tau protein. The intracellular deposition of NFT causes destroying the normal cytoskeletal architecture with subsequent neuronal cell death. This process called the tau hypothesis.

The most consistent neurochemical change associated with AD has been the well-documented decline in cholinergic activity that has inspired many attempts to treat AD with cholinergic drugs based on cholinergic hypothesis. However, additional alerts in Ca²⁺ homeostasis, catecholamine

^{*}Address correspondence to this author at the Biomedical Research Center. Health Sciences Technology Park, University of Granada, Avda. del Conocimiento s/n, 18100 Armilla, Granada, Spain; Tel: (+34) 611302236; E-mail: aest1977@hotmail.com



Fig. (1). Amyloid hypothesis. During AD development, amyloid precursor protein is cleavage to produce β amyloid peptide that aggregates and accumulates to form amyloid- β plaques. This plaques cause neurotoxicity or microglia activation, which in turn microglia release ROS and many pro-inflammatory cytokines such as NO, PGE₂, IL-1, IL-6, and TNF- α that accelerate cholinergic neuron damage. These proinflammatory cytokines subsequently activate astrocytes that also produce more cytokines to amplify the inflammatory signals and result in neuroinflammation and neurodegeneration.

(norepinephrine), serotonin, glutamate and neuropeptides [corticotrophin-releasing factors (CRF) and somatostatin (SRIF)] have also been described [5].

Genetics

Mutations in any one of a number of different singlegene on chromosomes 1, 14, and 21 can cause familial Alzheimer's disease (FAD) of early-onset Alzheimer's. Mutations of APP gene on chromosome 21 cause the formation of abnormal APP. A mutation of presenilin 1 (PS-1) gene on chromosome 14 causes abnormal PS-1 to be made, and a mutation of presenilin 2 (PS-2) gene on chromosome 1 leads to abnormal PS-2. However, the genetic causes of late-onset Alzheimer's, which develops after age 65, are not yet completely understood [6], but they likely include a combination of age, genetic, environmental, and lifestyle factors that influence a person's risk for developing the disease. A gene called Apolipoprotein E (ApoE) found on chromosome 19 appears to be a risk factor for the late-onset form of Alzheimer's. ApoE comes in several different alleles, or forms. The three forms of ApoE are ApoE ε^2 , ApoE ε^3 , and ApoE ε4 [7]. While inheritance of ApoE ε4 increases the risk AD development, ApoE ε 2 substantially protects against it.

Epidemiology

AD is a multifactorial disease and no specific environmental exposure has been found to be consistently associated with AD onset. Furthermore, the strong association between AD and increasing age may partially reflect the cumulative effect of different factors including genetic susceptibility, depression, traumatic head injuries, exposure to toxins and electromagnetic fields and vascular factors including midlife high blood pressure, cerebral and cardiovascular disease, smoking, obesity and diabetes as increasing disease risk factors, while anti-inflammatory medications and low to moderate alcohol consumption seem to reduce the disease risk. In addition, education and occupational history status are highly correlated (e.g., low education versus high education and unskilled versus skilled worker) with either a higher prevalence [8] or incidence of AD [9]. Low cognitive activities, poor social network or social disengagement have been shown to increase the risk of dementia in the elderly [10]. In supporting of these factors, the high level of complex mental activities, more frequently participating in mentally stimulating and physically activities are correlated with a reduced the AD risk [11].

CURRENT ANTI-ALZHEIMER'S TREATMENT

Currently, no drug treatments are available that can slow or stop Alzheimer's disease progression. However, scientists around the world are studying dozens of treatment strategies that may have the potential to delay or prevent the symptoms of AD [12].

There are two main types of medication can help for a time with memory symptoms and other cognitive changes that associated with Alzheimer's disease; cholinesterase inhibitors and *N*-methyl-D-aspartate (NMDA) receptor antagonists. The two types of drugs are worked in different ways. Cholinesterase inhibitors include rivastigmine, donepezil and galantamine to treat mild to moderate Alzheimer's, whereas; the NMDA receptor antagonist is memantine to treat moderate to severe Alzheimer's.

Cholinesterase inhibitors help by preventing acetylcholinesterase from breaking down acetylcholine at the nerve ending. Increased concentration of acetylcholine leads to improve the communication between the nerve cells that use acetylcholine as a neurotransmitter, which may in turn temporarily improve or stabilize the symptoms of Alzheimer's disease [13-14]. This class of medications may be used for three years, possibly longer.

The action of NMDA receptor antagonist is quite different from, and more complex than, that of cholinesterase inhibitors. It is thought to work by blocking glutamate. Glutamate, the major excitatory neurotransmitter in CNS, is released in excessive amounts and this causes the neurons to be damaged further. Memantine can protect neurons by blocking these effects of excess glutamate [15].

OXIDATIVE STRESS AND AD

Oxidative stress (OS) reflects the imbalance between the production and quenching of free radicals from oxygen species in the biological system. The disturbances in the normal redox status of cell can cause an increment in reactive oxygen species (ROS). These ROS play a key role in many chronic diseases including cancer, mitochondrial diseases [16], neurodegenerative diseases [17, 18]. Accumulating evidence has shown that the presence of extensive OS is a characteristic of AD brains in addition to the established pathology of senile plaques and NFT [19]. The interventions between OS and other key events in AD, which amplify the complexity of this issue was summarized in Fig. (2).



Fig. (2). Interventions between oxidative stress and the other key factors in AD.

Nucleic Acids Oxidation

Excess ROS or free radicals can oxidize nucleic acid which 8-hydroxyguanosine and 8-hydroxydeoxyguanosine (8-OHdG) are formed as markers of DNA, RNA and mitochondrial DNA (mtDNA) oxidative damage. Nucleic acids damage is thought to play a key step in neuronal loss associated with aging and many neurodegeneration diseases [20]. mtDNA is highly susceptible to oxidative stress because of its vicinity to ROS generation, the absence of histones and has limited repairing mechanisms. mtDNA damage could potentially cause bioenergetic and nerve dysfunctions. Interestingly, mtDNA damage is observed before A β deposition and neuronal degeneration [21]. Indeed, DNA oxidative damage is a feature of AD and considered as an early event in AD progression [20, 22].

Proteins Oxidation

It has been demonstrated that the levels of 3-nitrotyrosine and protein carbonyls, which are resulted from protein oxidation are elevated in brains of patients who suffered from AD [23]. Proteins oxidation causes advanced glycation end products (AGEs) that can be a factor in the development or even worsening of many diseases. AGEs chemically are posttranslation modified proteins that are formed when adding nonenzymatically monosaccharides to the amino group of the protein. This protein undergoes further modification via oxidation, condensation, and dehydration to produce the AGEs. This reaction, which is catalyzed by transition metals such as iron to form enediol radical, which produces free radicals by reducing molecular oxygen. AGEs may speed up oxidative damage via either direct radical production by chemical oxidation and degradation of AGEs, indirect oxidative stress via AGE-receptor (RAGE) binding and activation of signaling pathways such as upregulation of the transcription factor nuclear factor- κB (NF- κB), or by interacting with microglia in an acute phase reaction that results in a respiratory burst and potentiate free radical production that leads to deficits in learning and memory [24]. AGEs colocalized with the senile plaques of AD-affected brains [25]. Moreover, the senile plaques contain 3 times more AGEs than that of the age-matched brains control. AGE modified AB to form AB-AGEs and accelerates the aggregation and accumulation of nonfibrillar soluble A β in vitro, which suggests that this process may also occur in vivo and creates vicious cycles or positive feedback loops [26-27]. Interestingly, AB is considered as a ligand of RAGE this in turn mediates Aβ-induced oxidative damage. In addition to the accumulation of free radical damage, alterations in the antioxidant enzymes activity or expression such as superoxide dismutase (SOD) and catalase have been observed in both CNS and peripheral tissues of AD patients [28]. Moreover, the increased oxidative damage to lipids and proteins and the decline of glutathione and radical detoxifying enzymes activity are more localized to the synapses and correlate with the severity of the disease, suggesting that oxidative stress could be involved in AD-related synaptic loss [29].

Lipid Peroxidation

CNS is a major target for lipid peroxidation. In the brain, low concentrations of the endogenous antioxidant component glutathione and the antioxidant enzyme catalase, a high metabolic rate (consumes about 20–30% of inspired oxygen), and a high proportion of polyunsaturated fatty acids (PUFAs) make this organ an ideal target for oxidative damage [30]. As a result of PUFAs attacked by free radical,

malondialdehyde (MDA) and 4-hydroxy-2,3-nonenal (HNE) are formed beside to acrolein as a reactive substance. In AD brains elevated MDA, HNE and acrolein has been identified. Moreover, lipid peroxidation markers noted in patients with mild cognitive impairment, suggesting that lipid peroxidation is an early event in AD progression. Furthermore, MDA is also found in different brain regions and cerebrospinal fluid (CSF) of AD patients [31]. Lipid peroxidation reacts with macromolecules causing impairment of the function of membrane proteins such as the neuronal glucose transporter (GLUT 3), reduction of glucose metabolism by inhibiting enolase, inhibition of glutamate transporters, inhibition of Na^{+}/K^{+} ATPases, inhibition of antioxidant enzymes as SOD 1 and hemeoxygenase 1, activation of kinases, and dysregulation of ionic transfers and calcium homeostasis [32]. Dis-ruption of Ca^{2+} homeostasis, due to increase in intracellular Ca²⁺, could cause a cascade of intracellular events as ROS generation and cellular death by apoptosis, and it also worth noting that AD shows Ca²⁺-dependent cell death [33].

Metals Homeostasis Disturbance

As mention above and recent evidences suggest that disruption of metal homeostasis may also contribute to oxidative damage [34-35]. During aging metals such zinc, iron and copper accumulate in the brain which act as antioxidants. Metal dependent enzymatic processes are important for brain metabolism and metal dyshomehostasis is linked to AD progression. Zinc, iron and copper are able to interact with secretase that promoting APP cleavage, senile plaque formation, facilitating Aß aggregation and hyperphosphorylation of tau protein [35, 34]. Furthermore, copper, zinc and iron bind to A β triggering signaling cascades that amplify oxidative damage [34]. In addition, synaptic zinc has been associated with increasing plaque burden in brain of AD mouse models [36]. There is evidence that disruption of zinc homeostasis may play an important role in microtubule and tau pathology [37]. Regarding this fact, divalent metal ion chelators such as clioquinol and desferrioxamine have had some success in altering the progression of AD [38-39] by facilitating solubilization of AB plaques. However, zinc might at low concentration actually protects the neurons by blocking A β channels or compete with Cu for A β binding [2] and partially prevents the cognition loss.

Mitochondrial Dysfunction

Mitochondrial dysfunction appears to play a prominent role in the early events of AD progression [40]. Regarding this fact, a decreased in oxidative phoshorylation genes expression of mitochondria was noted in the neocortex of AD brain and this decreased was correlated with the severity of dementia [32]. There are evidences that both phosphorylated tau protein and A^β deactivated complexes I and IV, respectively. Indeed, AD markers are alerted mitochondrial oxidative phosphorylation (OXPHOS) system. Furthermore, loss of mitochondrial integrity plays an important role in synaptic dysfunction [41]. Moreover, deposition of AB leads to more mitochondrial damage [42] by interacting with A β -binding alcohol dehydrogenase the mitochondrial protein (ABAD, a neuronal mitochondrial enzyme exacerbates A\beta-mediated mitochondrial and neuronal dysfunction). The formed complex prevents the binding of NAD⁺ to ABAD, thereby changing mitochondrial membrane permeability and reducing the activity of respiratory enzymes causing ROS generation. Also, mitochondrial mobility has also been altered in AD causing a mitochondrial reduction in neurites [43]. Aβ plaques induce a reduction in motile mitochondria [44-45]. In addition, Aβ significantly alerted mitochondrial fission and fusion by changing the expression of almost proteins that regulate this process.

Moreover, in spite of the growing number of data concerning the central role of mitochondria in apoptosis signaling. A reduction in mitochondrial membrane potential induce mitochondrial permeability transition pore (mtPTP) as an early universal event of apoptosis. The liberation of mitochondrial cytochrome c and apoptosis-inducing factor (AIF) from the mitochondrial intermembrane space into the cytoplasm are also the most key events in activating the cascade of reactions that leading to cell death. Such alterations in mitochondrial structure have been noted as a causative mechanism in AD pathogenesis [46].

CAN MICROBIOME BE LINKED TO AD?

Nowadays, accumulating evidences are linked between microbiome and homeostatic status of CNS. For example, gastrointestinal (GI) tract contains gram-positive organisms are capable to metabolize glutamate to GABA crosses blood brain barrier [47], GI microbiome can significantly alert BDNF expression and reduce its level in hippocampus and cortex [48], GI microbiome can generate β -N-methylamino-L-alanine (BMAA), neurotoxin induced NMDA activation, glutathione depletion, oxidative stress and intra-neuronal protein misfolding [49], some bacteria and fungus can produce amyloids against human defenses, those amyloids can aggregate into oligomers promote amyloid fibril formation [50], lastly, some types of bacteria can produce phenol soluble modulins (PSMs; some peptides), those PSMs activate strongly formyl-peptide receptor 2 (FPR2) of neutrophil [51]. Interestingly, FPR2 can bind to a wide variety of amyloid-like ligands [52].

CAN ANTIOXIDANTS BE CONSIDERED AS A THERAPEUTIC TARGET IN AD?

Antioxidants are molecules that inhibit the oxidation of other molecules. Antioxidants are widely used and have been investigated for the prevention of many diseases. In the recent years, many natural compounds with antioxidant properties have been investigated as adjuvant therapies for AD and other neurodegenerative diseases. It has been reported that antioxidants such as vitamin E or α -tocopherol, β -carotene, vitamin C, lipoic acid and *N*-acetylcysteine and others may offer protection against extracellular and intracellular ROS and H₂O₂-cell-damaging compounds that are generated as byproducts of normal cell functioning before these radicals damage cells or activate microglia through their action as intracellular second messengers [53].

As accumulating evidences have concerned oxidative stress in AD initiation and progression, the possibility of using natural antioxidants for prevention and treatment of AD has attracted considerable attention. Furthermore, antioxidant therapy is considered a promising low risk therapeutic strategy for AD. This review will mainly focus on the

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recent development of common used antioxidant therapy for AD and thus will provide indication for the future potential antioxidant therapeutic methods for AD patients. The potential functions of the different antioxidants are summarized in (Table 1).

Table 1.	The potential functions of some antioxidants in Al	Z-
	heimer's disease.	

Antioxidant	Potential functions
Melatonin	Enhance the rest-activity rhythm and improved sleep quality [54]. Prevent Aβ fibrillogenesis and aggregation [55].
Estrogens	Enhance the uptake of aggregated Aβ into microglia [56]. Reduce Aβ production [57].
Selenium	Stimulate mitochondrial biogenesis signaling and enhance mitochondrial functional performance [58]. Reduce tau level [59].
Polyphenols	Ellagic acid and punicalagin: Potent β-secretace inhibitors [60]. Curcumin: Inhibit cytokines that initial amyloid production [60]. Epigallocatechin-3-gallate: Reduce Aβ production and plaque deposition in brains [61].
Vitamins	Vitamin E: Improve cognitive performance [62-64] and suppress tau-induced neurotoxicity [65]. β -Carotene: Increase choline acetyltransferase activ- ity [66]. Vitamin B12: Enhance neurochemistry [67]. 1,25Dihydroxy-vitamin D3: Increase the phagocytic clearance of amyloid plaques [68].
Docosahexaenoic acid	Maintain integrity and neuronal function. Limit the production and accumulation of neurotoxic $A\beta$ from its APP v [69-70]
CoQ ₁₀	Improve brain bioenergetics [71]. Attenuate $A\beta$ overproduction and intracellular $A\beta$ deposition in the cortex [72].

ROLE OF MELATONIN IN AD

Melatonin appears as unique for several reasons. Melatonin is an ubiquitously compound synthesized in the pineal gland and other body organs and tissues [73] suggesting that melatonin involves in a number of not yet defined activities at the cellular level and acts as a hormone. The majority of melatonin directly released from the pineal gland via the pineal recess to the cerebrospinal fluid (CSF) [74], moreover, melatonin production decreases with the aging, a fact which has been suggested to a the major predisposing factor in age-associated degenerative diseases [75, 76].

Melatonin (Fig. 3) and other structurally related indolic compounds proved to be more potent than classical antioxidants [77-78]. Melatonin can directly detoxify both reactive oxygen and nitrogen species or indirectly by regulating the enzymatic activity that promotes the overall antioxidative

defense systems [73]. In addition, metabolites formed from the interaction between melatonin and free radicals, including N1-acetyl-N2-formyl-5-methoxykynuramine (AFMK) and N1-acetyl-5-methoxykynuramine (AMK), are also efficient free radical scavengers. Moreover, melatonin exerts anti-excitatory effects, this effect is supported by observations showing that melatonin keep neurons away from AB toxicity via GABA receptor activation. This sedating effects of melatonin and its related compounds display particular chronobiological activity that make them capable of correcting the circadian rhythm disorders seen in AD patients [75]. In this regard, 10 patients have mild cognitive impairment (MCI) treated with melatonin (6 mg/day for 10 days) exhibited significantly better rest-activity rhythm with improved sleep quality, the patients also have ability to remember previously learned items along with a significant reduction in depressed mood [54].





The anti-fibrillogenic activity of melatonin and its metabolites were observed not only in vitro but also in vivo [79, 75]. Evidence derived from transgenic mouse model indicates that melatonin administrated at early phase regulates APP and A^β metabolism with little anti-amyloid at the late phase [80]. The possible mechanism beyond this action is the ability of melatonin to inhibit glycogen synthase kinase-3 (GSK-3) and up-regulate the c-Jun N-terminal kinase resulted in matrix metalloproteinases activation that degrades AB. It has been demonstrated that melatonin directly interacts with A β and prevents its aggregation [55]. Melatonin could promote the conversion of β -sheets into random coils and inhibit progressive β-sheet and/or amyloid fibrils by disrupting the imidazole-carboxylate salt bridges. Melatonin may not only reduce $A\beta$ neurotoxicity, but also facilitate the clearance of AB peptide by increasing the proteolytic degradation via increased insulin-degrading enzyme (IDE) activity [80].

Oral administration of melatonin attenuated A β -induced proinflammatory cytokines, by inhibiting NF- κ B binding to DNA and suppressing inducible nitric oxide synthase (iNOS) gene expression in brain of rat [81]. As a consequence, melatonin and its metabolites may improve the clinical course of AD.

Besides melatonin traditional role as an antioxidant and free radical scavenger, melatonin maintains mitochondrial homeostasis and inhibits mitochondrial cell death pathways by lowering electron leakage, inhibiting the opening of mtPTP, thus maintaining the mitochondrial respiratory electron flux. In addition, administration of melatonin inhibited the Aβ-induced mitochondria-related factor up-regulation as Bax and suppressed caspase-3 activity and melatonin may also initiate the survival signal pathways. Taken together, the above mentioned evidences suggest that melatonin serves as a potential antioxidant therapeutic strategy for AD [32].

ROLE OF ESTROGEN IN AD

The antioxidant property of estrogens is attributed to the novel redox cycling of catechol estrogens [82]. Estrogens are one of the best-studied classes of molecules for their potential role in neuroprotection. Furthermore, estrogenic prevention and a disease-modifying therapy against AD are well studied. Estrogens are powerful neuroprotective agents against oxidative stress and excitatory neurotoxicity. These activities are attributed to the capability of estrogens to maintain proper mitochondrial functions and suppress mitochondrial apoptosis-related proteins. The ability of estrogens to protect neurons from a number of toxic insults, including A β peptide have been extensively assessed [83]. Estrogens significantly regulate A β processing and deposition.

Data from ex vivo experiment showed that each of the three major circulating estrogens; E2, estrone and estriol (Fig. 4) can reduce $A\beta$ fibrillation. Also, estrogen enhances the uptake of A β into human cortical microglia [56] and prevents AB aggregation while concomitantly increases soluble APPα [84], accelerates APP trafficking in the trans-Golgi network [57] and reduces the expression of BACE-1 [83], thereby reduces the production of A β . Moreover, 17 β estradiol and the selective estrogen receptor modulators such as raloxifene and tamoxifen in human neuroblast long-term cell cultures showed neuroprotective effects by increasing resistance against AB-induced toxicity [85]. Furthermore, data form in vivo study conducted with transgenic mouse containing the human Swedish APP mutation showed that E2 reduced Aß [86]. Further, in transgenic mouse with double mutations in APP and PS-1 genes, E2 treatment showed a significant reduction in brain AB [87]. However, Green et al. [88] showed that E2 has no effect on A β in PDAPP transgenic mouse. Interestingly, in one clinical trial, estrogen treatment improved memory function in women with AD by its ability to maintain and sustain neuronal viability [89]. However, estrogen is ineffective in reversing AD process.

ROLE OF SELENIUM IN AD

Selenium (Se), a vital trace element that is abundant in the brain, mainly exerts its antioxidative effect through selenoproteins, such as glutathione peroxidase, thioredoxin reductase, selenoprotein P, selenoprotein R, and selenoprotein M [90]. There has been heightened interest in the role of Se in health and neurologic disorders including AD [91-92]. It has been reported that the level of Se declines with age [93] and Se deficiency might increase the risk of AD [94]. AD patients demonstrated significantly lower Se levels in plasma, erythrocytes, and nails when compared with the control group [95]. Se intake may slow down the onset of cognitive decline associated with AD [59].

Connections have been observed between Se and risk factors of AD. Relations have been shown between Se and ApoE and presenilin 2, both genetic risk factors for AD [96]. Several autopsy studies using human postmortem brain tissue samples from AD patients and healthy control patients investigated whether AD is associated with altered levels of Se [92]. Supplementation of sodium selenite significantly stimulated mitochondrial biogenesis signaling and enhanced mitochondrial functional performance in murine hippocampal neuronal cells [58]. Sodium selenite also prevented cognitive deficits and oxidative damage in a rat model of AD [97]. Furthermore, sodium selenate was reported to mitigate tau pathology, neurodegeneration and functional deficits through the activation of protein phosphatase 2A in AD mice, significantly boosting phosphatase activity [98]. Meanwhile, organo-selenium decreased amyloid burden and prevented RNA and DNA oxidative damage in APP/PS1 mice [99]. Although the effect of selenium on AD has been investigated in a number of studies, moreover, selenomethionine (Se-Met) treatment reduced the levels of tau. Se-Met treatment also reduced the phosphorylation of tau at the site Ser404 [59].

ROLE OF METAL ION CHELATION IN AD

Interactions between metal ions and $A\beta$ are one of the currently accepted hypotheses "metal hypothesis of AD". Further, abnormal metal ion homeostasis is connected with the AD neuropathogenesis. Based on this fact, prevention of metal-A β interactions and restoration of metal ion homeostasis in the brain via metal chelation therapy has been proposed in order to reduce metal-A β species neurotoxicity [100].

To date, strategy of metal chelator has been used as agent for metal ion chelation therapy in AD. Desferrioxamine B (Desferal®; a drug used to treat Fe overload) was the first compound used to treat metal overload in the CNS and to dissolve amyloid aggregates. Desferrioxamine B significantly improved the behavioral and cognitive declines of AD patients [101]. Clioquinol [CQ, a classic metal chelator for Cu (II) and Zn(II)] also used to prevent A β plaques forma-



 17β -EstradiolEstroneEstriolFig. (4). Estrogens chemical structures. 17β -Estradiol ($C_{18}H_{24}O_2$), estrone ($C_{18}H_{22}O_2$) and estriol ($C_{18}H_{24}O_3$).

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tion. However, the use of these synthetic metal chelators also led to various disadvantages: (i) its hydrophilic and charged characters disable the blood brain barrier crossing (ii) it is rapidly degraded and caused miss location of metal ions *in vivo*, (iii) it cause significant side effects such as anemia due to its strong affinity for Fe(III), and (iv) some of metal chelators as CQ may cause neurotoxicity and mutagenicity.

Hence, the search for new metal chelators without side effects continues to grow. Natural metal chelators such as *Linum usitatissimum*, *Punica granatum*, and garlic extract are effective in preventing heavy metals-associated toxicities [102-103]. Hence, the use of natural compounds or their active components have proven to be a promising approach for safer metal chelation.

ROLE OF POLYPHENOLS IN AD

Polyphenols are secondary metabolites of plant that constitute one of the most common and widespread groups of substances in plants and apparently act as defense (against herbivores, microbes, viruses or competing plants) and signal compounds, as well as protecting the plant from ultraviolet radiation and oxidants and may contribute to flavor, color and oxidative stability in plants. The term "phenolic" or "polyphenol" can be precisely defined chemically as a substance which possesses an aromatic ring bearing one (monophenol) or more (polyphenol) hydroxyl substituents, including functional derivatives (esters, methyl ethers, glycosides, etc.). The main classes under polyphenols are phenolic acids, flavonoids, stilbenes and lignans. Pomegranates, apples, grapes, green tea and many other plant sources are the subject of increasing scientific interest because of their antioxidant and anti-inflammatory properties with possible health benefits. Many of polyphenols received intensive studies for their potential disease prevention or treatment effects and are worthy of consideration for AD.

Punica granatum (pomegranate) has the potential to suppress the AD pathogenic cascade at multiple sites. Pomegranate juice decreased amyloid load and improved behavior in APPsw (double Swedish APP mutation; Tg2576) transgenic mouse model of AD [61]. Ellagic acid, punicalagin and punicalin (Fig. 5) of pomegranate were found to be powerful β -secretace inhibitors [60]. Also, pomegranate shows a direct radical scavenging activity with lipid peroxidation inhibiting property, particularly metal catalyzed peroxidation [103]. Pomegranate is also known as a good inhibitor of gene expression of inflammatory cytokines such as IL-6, IL-8, vascular endothelial growth factor (VEGF) and prostaglandin E2 (PGE2), COX-2, and iNOS by influence of inhibition of JUN and NF-kB-mediated gene transcription [104-105]. All of these inflammatory cytokines have been implicated in A β toxicity, indicating the multi-target inter-



Punicalin

Epigallocatechin-3-gallate

Fig. (5). Chemical structures of Ellagic acid ($C_{14}H_6O_8$), punicalagin ($C_{48}H_{28}O_{30}$), punicalin ($C_{34}H_{22}O_{22}$) and epigallocatechin-3-gallate ($C_{22}H_{18}O_{11}$).

vention of pomegranate in AD. In addition, pomegranate polyphenols attenuate disruption of mitochondrial membrane [106]. Furthermore, pomegranate has other proven antiamyloid activities. Pomegranate decreased soluble Aß levels and AB deposition by inhibiting BACE1 [60]. Ellagic acid, a phenol found abundantly in pomegranate inhibits Aß plaques formation and Aβ toxicity *in vitro* [107]. Another polyphenol found in pomegranate, epigallocatechin-3-gallate (EGCG; Fig. 5), reduces A β formation and A β plaques deposition in the brain of transgenic mouse containing the human Swedish APP mutation [61]. This anti-amyloid activity of pomegranate remains effective in aged mice, even after amyloid deposition continues over time. Interestingly, pomegranate was shown to decrease A β plaques load and improve memory impairment in behavioral performance testes in AD transgenic mice [108] even in response to acute A^β brain injection [109].

ROLE OF TRADITIONAL HERBS IN AD

Aged garlic extract (AGE) has demonstrated beneficial effects in AD models [110]. AGE and its active ingredients S-Allyl-L-Cysteine (SAC; Fig. 6) treatments not only decreased AB plaques loads in the brains of APP transgenic mice, but also ameliorated tau pathology by inhibiting GSK-3β and increased levels of synaptic protein markers as synaptosomal-associated protein 25 (SNAP-25) [111]. Previous research has demonstrated that AGE protects the cellular structures from Aβ-mediated neurotoxicity [112]. Interestingly, SAC was shown to have Aβ disaggregation property in vitro by activating peroxisome proliferators-activated receptors- α (PPAR- α) in microglia and macrophages that involved in A_β clearance [113]. In APP transgenic mice, four months of AGE and SAC treatments significantly reduced both A β load and A β plaques numbers in the brain versus non-treated controls [114]. In addition, AGE treatment resulted in a significant reduction in the intracellular APP level. Moreover, SAC treatment can prevent A\beta-mediated neurodegeneration in hippocampus by preventing endoplasmic reticulum (ER) stress and improve memory deficits [115]. Mechanistically, SAC prevents Aβ-induced neuroinflammation and toxicity by inhibiting NF-kB activation and also reverses ROS-mediated decline in cholinergic function of the neurons by increasing in levels of neuronal acetylcholine transferase activity [110].



Fig. (6). Chemical structure of S-Allyl-L-Cysteine ($C_6H_{11}NO_2S$).

Silymarin is a mixture of four flavonolignane diastereomers; silibinin (or silybin), isosilybin, silydianin and silychristin, the major ingredients of the milk thistle extract (*Silybum marianum*). Silymarin showed anti-amyloid property *in vivo* and significantly reduced the A β plaque burden associated with microglial activation, A β plaques formation and disturbed behavior in APP transgenic mice. However, this anti-amyloid property of Silymarin is not attributed to β secretase inhibition. Silymarin might act also to enhance neuronal cell viability by activating protein kinase B and inhibiting caspase-3 as well as attenuate A β neurotoxicity in AD model mice [116]. In addition, silibinin (Fig. 7) prevents memory impairment and oxidative damage induced by A β in mice [117].



Fig. (7). Chemical structure of silibinin $(C_{25}H_{22}O_{10})$.

ROLE OF VITAMINS IN AD

 α -Tocopherol (vitamin E), L-ascorbic acid (vitamin C), β -carotene (a precursor form of vitamin A) and vitamin D are organic compound with antioxidant properties, which decrease free-radical-mediated damage in neuronal cells and help to inhibit dementia and cognitive impairment [32]. Therefore, it has been postulated that vitamins could be used as important therapeutic strategies.

There are two groups of vitamin E with different ten forms; five as tocotrienols and five as tocopherols and identified by α -, β -, γ -, δ - and ϵ -. The most biologically active form of vitamin E is α -tocopherol (Fig. 8). The work of Sano et al. [118] gave impetus to the idea of vitamin E as the treatment of AD. At the present days, the same group has reported that vitamin E benefits in patients with mild to moderate AD were seen by slowing functional decline [119]. Vitamin E lowered the oxidation of blood glutathione and the peroxidation of plasma lipids that cause an improvement in AD. In vivo studies, vitamin E has been shown to prevent the toxic effects of A β and improve cognitive performance [62-64]. In the Chicago Health and Aging Project, higher intakes of vitamin E from natural sources were associated with decline in Alzheimer's disease incidence [120]. Similarly, in the Rotterdam study, high vitamin E intake was associated with reduced the incidence of dementia [63]. Furthermore, Dias-Santagataet et al. [121] reported that α tocopherol administration significantly prevented tauinduced neurotoxicity in Drosophila, and similar beneficial outcomes were recently reported by other researchers using tau pathology in transgenic mouse model [65], which underscored the therapeutic value of vitamin E. However, vitamin E should come from foods, rather than supplements, where, vitamin E from supplements has not been shown to reduce AD risk [122]. Mechanistically, the potential effect of vitamin E in AD is remain elusive, however, vitamin E may exert this potential by preventing A β -induced ROS, protecting against oxidation-mediated decline in neurotransmissionassociated protein, inhibiting inflammatory cytokines those participated in neuroinflammation and activating PP2A.

Vitamin B12 (Fig. 9) is essential for the health of the brain and nervous system and for blood cell formation [123]. However, vitamin B12 is lowered in elderly adults especially in males. Moore *et al.* [124] showed that low serum levels of vitamin B12 are allied with neurodegenera-

tive disease and cognitive impairment and that vitamin B12 therapy does not improve cognition in patients without preexisting deficiency. Vitamin B12 supplementation increased the activity of choline acetyltransferase in cholinergic neurons in cats [66] and improved cognitive performance in AD patients possibly by its ability to reduce homocysteine levels [125]. Hyperhomocysteinemia was implicated in neurotoxicity by overstimulation of NMDA receptors or by increasing the vulnerability of hippocampal neurons to excitotoxicity and A β toxicity.



Fig. (8). Chemical structure of α -tocopherol form of vitamin E (C₂₉H₅₀O₂).



Fig. (9). Chemical structure of vitamin B12 (C₆₃H₈₈CoN₁₄O₁₄P).

A relationship between dietary carotenoids and agerelated cognitive function has been reported. Perrig *et al.* [126] noted that the higher level of β -carotene in plasma was associated with better memory performances (priming, working-memory, free recall, recognition and vocabulary test) in old and very old subjects. β -Carotene (a major precursor to vitamin A; Fig. **10**) might have beneficial effects via its antioxidant or A β anti-oligomerisation effects [127-128]. Certain carotenoids also may modulate the functional properties of synaptic membranes [128], enhance gap junctional communication [67].

Vitamin D is a steroidal hormone and exerts its effects via vitamin D receptor (VDR) that located in the nucleus. Traditionally, vitamin D regulates the metabolism of bone, however, the recent studies shown that VDR are abundantly present in neurons and glial cells in CNS and participated in some physiological processes. The active form of vitamin D, 1,25 dihydroxy-vitamin D3 (1,25-D3 or cholecalciferol; Fig. 10), upregulates neurotrophin expression, such as nerve growth factor (NFG), neurotrophin 3 (NT3), and glial-derived neurotrophic factor (GDNF) [129] those affects the survival and differentiation of neurons. Consistently, hypovitaminosis D is associated with prevalent cognitive impairment and AD dementia in elder [68]. Mechanistically, vitamin D reduced the risk of AD and other neurodegenerative diseases through several mechanisms including neuro-protection and synaptic plasticityenhancing effects through detoxification pathways by inhibiting iNOS and enhancing antioxidant system, regulates calcium homeostasis and protects neurons from excess calcium, inhibits neuroinflammation by down regulating NFκB activity and modulates angiogenesis by down regulating angiogenin 2 and vascular endothelial factor expression (VEGF) [130-131]. Furthermore, vitamin D activates glyoxalase 1 that catalyze methylglyoxal, the major precursor to AGE formation [132]. In vivo, vitamin D inhibits β secretase, affects the expression and processing or APP and increases the phagocytic clearance of A^β plaques by NGFstimulated astrocytes [133-135]. Moreover, in vitro, vitamin D reduces AB-induced inflammation and apoptosis in primary cortical neurons [136].

ROLE OF OMEGA-3 IN AD

Omega-3 fatty acids are polyunsaturated fatty acids with a double bond after the third carbon atom in its carbon chain from the terminal methyl end and commonly found naturally in marine and plant oils. The backbone of omega-3 is lino-



Fig. (10). Chemical structures of β -carotene (C₄₀H₅₆) and 1,25Dihydroxy-vitamin D3 (C₂₇H₄₀O).

lenic acid and human must be obtained it in diet, human cannot synthesize it. Consuming omega-3 fatty acids versus other fatty acids reduce the risk of cancer, cardiovascular disease, inflammation, and neurological disorders.

Docosahexaenoic acid (DHA; Fig. 11) is the most important components of omega-3 fatty acid with potent anti-inflammatory and antioxidant prosperities. In brain, DHA is modified to neuroprotectin 1 (NPD1) through the action of phospholipase A₂ and lipoxygenase. NPD1 has been shown to have potent anti-inflammatory and neuroprotective effects in neural systems [137] by regulating the redox state of neurons. The action of NPD1 includes up regulation of Bcl-2 family, down regulation of pro-apoptotic proteins and suppressing the production of prostaglandin that participated in neuronal damage. Furthermore, there is a growing body of evidence that DHA modifies the expression of many genes that regulate a variety of biological functions including neurogenesis and neuronal functions and survival that important for cognitive health [138]. Thus, the intake of DHA can reduced the risk of AD [139]. For this, it is not surprising that, supplementation of aged transgenic mouse containing the human Swedish APP mutation with DHA-depleting rich safflower oil diet exhibited oxidative damage and significant (70-95%) loss of postsynaptic proteins that are seriously depleted in AD brains by 70–90% [140].



Fig. (11). Chemical structure of docosahexaenoic acid (DHA; $C_{22}H_{32}O_2$).

DHA exerts protective effects against neurotoxicity induced by A β [141, 142]. DHA limits the production and accumulation of neurotoxic A β from its APP [69, 70] by facilitating the interaction of α -secretase with APP to produce non-amyloidogenic sAPP α . Elevated sAPP α levels were associated with substantial protection against mitochondrial dysfunction and apoptosis [143]. Further, DHA prevents the action of γ -secretase by forming shield over the essential recognition sequence site for γ -secretase, and also inhibits A β formation and fibrillation. Additionally, omega-3 effectively reduces cholesterol and prevents its oxidation. High cholesterol levels in the brains directly stimulate β - and γ secretase activities [144].

Additionally, dietary DHA increased cerebral acetylcholine levels *in vivo* [145, 146], and improved memory through the influence of DHA on the phospholipids of neuronal membranes. Thus, DHA increases the learning ability in rats [147] and prevents the loss of neurons longevity, discrimination-learning ability and memory in aged rats [148].

Finally, DHA has a number of potential mechanisms as an antioxidant by binding to membrane and trapping the generated ROS [149] and enhancing the activity of endogenous antioxidant system [150]. Furthermore, DHA may reduce the generated ROS by increasing nitric oxide (NO) synthesis, which may decrease the cellular oxygen pool and subsequently reduce ROS and lipid peroxidation [151]. Additionally, NO significantly increases blood flow and supply of nourishment and facilitates the removal of toxic metabolites and proteins from the brain.

ROLE OF COQ₁₀ IN AD

In 1955, Festenstein *et al.* [152], the British scientists in Morton's Laboratory in Liverpool isolated a new unsaponifiable lipid from the intestinal mucosa of horses. The substance was identified later as a quinone and was found to be distributed in the most animal tissues, Morton named it ubiquinone "the name derived from ubiquitous quinone that meaning everywhere present quinine". In David Green's Laboratory at the Wisconsin University in USA after two years, Crane *et al.* [153] observed a novel quinone in the inner membrane of mitochondria and named it coenzyme Q because of its important role in the electron transport chain and ATP synthesis.

Coenzyme Q_{10} (Co Q_{10} ; Fig. 12) exerts a potential neuroprotective effect. In brain, Co Q_{10} protects the neuronal cells by increasing the stability of the cell membranes, decreases free radical that may attack DNA, and shows capability to recycle and regenerate the other antioxidant molecules, such as tocopherol and ascorbic acid [154].



Fig. (12). Chemical structure of coenzyme Q_{10} ($C_{59}H_{90}O_4$).

CoQ₁₀ is a potent antioxidant in mitochondrial membranes [155]. More specifically, CoQ₁₀ facilitates efficient transport of electrons and defends mitochondria against oxidative injury. However, the levels of CoQ₁₀ have been reported to decline with age, making mitochondria increasingly vulnerable and hazardous [156]; this malfunction may contribute to the development of neurodegenerative diseases. As such, considerable interest has been placed on the therapeutic use of CoQ10 in AD [157]. Nevertheless, the preclinical and clinical evaluation of CoQ10 has been hindered by its hydrophobicity, severely limiting its use in vitro and in vivo [158]. CoQ_{10} was able to modestly restore autophagic activity [159] and exerts anti-inflammatory effects by inhibiting NF-κB [160]. CoQ₁₀ administration to aged APP/SP-1 transgenic mice led to a reduction in cortical levels of $A\beta_{42}$ and reduction of oxidative stress markers [161]. Moreover, CoQ₁₀ supplementation improved brain bioenergetics [71] and partially prevented AB overproduction and deposition in the cortex of transgenic mice brain [72] and suppressed brain AGEs levels.

CLINICAL TRIALS WITH ANTIOXIDANTS THERA-PIES IN AD

Unfortunately, the outcomes of many clinical trials with different antioxidants demonstrated no or minimal effects. For example,

- *Curcuma longa* exerts potent antioxidant and antiinflammatory properties and capable of inhibiting $A\beta$ aggregation *in vitro*, but when it tested in clinical trial showed no significant effect [162]. Low absorption of curcumin through gastrointestinal tract may be a cause for that, however, a carrier mediated transport or nanotechnology based delivery system can potentiate its effect [163].
- Ginkgo biloba has many properties including antioxidant effect and used in Chinese medicines. However, when Ginkgo biloba standardized extract (EGb 761) tested in a large randomized controlled trial did not show any difference in cognitive decline from the control group [164].
- CoQ10 or its analog are potent antioxidants, however, when CoQ10 tested in 563 patients with mild AD for one year did not show any changes in cognitive decline compared to the placebo group [165].

CONCLUSION

According to the predominantly symptoms those caused by acetycholine signaling dysfunction, the cholinergic hypothesis was conceived at the beginning. However, amyloid and tau hypothesizes are used nowadays based on the AD pathogenesis. APP was degraded with β -secretase to sAPP that subsequently degraded by γ -secretase to release A β , amyloid hypothesis. While in tau hypothesis, NFT was formed as a result of tau protein hyperphosphorylation. Even though the toxicity of A β is well established in AD pathogenesis, oxidative stress may play an important role in the initiation and progression of AD. Furthermore, oxidative stress amplifies the key events in AD.

Currently, acetylcholinesterase inhibitors and NMDA receptor antagonists alone or in combination are used to treat mild to moderate AD symptoms. High intakes of antioxidants low the risk for AD, and show beneficial effects in the prevention of the disease in several *in vitro* studies.

Unfortunately, the outcome of clinical trials with antioxidants demonstrated minimal effect. For this reason, when it comes to the use of these studies as examples to deny the oxidative-stress hypothesis of AD, at best they can only be considered inconclusive. Since, AD is a heterogeneous disorder, multimodal strategies using different molecular targets and delivery methods with antioxidants should be examined.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

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REFERENCES

- Citron M. Alzheimer's disease: strategies for disease modification. Nat Rev Drug Discov 9(5): 387-398 (2010).
- [2] Querfurth HW, LaFerla FM. Alzheimer's Disease. New Engl J Med 362(4): 329-344 (2010).
- [3] Cruts M, Van Broeckhoven C. Molecular genetics of Alzheimer's disease. Ann Med 30(6): 560-565 (1998).

- [4] Dong S, Duan Y, Hu Y, Zhao Z. Advances in the pathogenesis of Alzheimer's disease: a re-evaluation of amyloid cascade hypothesis. Transl Neurodegener 1(1): 18 (2012).
- [5] Yaari R, Corey-Bloom J. Alzheimer's disease. Semin Neurol 27(1): 32-41 (2007).
- [6] Rovelet-Lecrux A, Hannequin D, Raux G, Le Meur N, Laquerriere A, Vital A, et al. APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy. Nat Genet 38(1): 24-26 (2006).
- [7] Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, *et al.* Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. Science 261(5123): 921-923 (1993).
- [8] Hurt LS, Ronsmans C, Saha S. Effects of education and other socioeconomic factors on middle age mortality in rural Bangladesh. J Epidemiol Community Health 58(4): 315-320 (2004).
- [9] Roe CM, Xiong C, Miller JP, Morris JC. Education and Alzheimer disease without dementia: support for the cognitive reserve hypothesis. Neurology 68(3): 223-228 (2007).
- [10] Fabrigoule C. Do leisure activities protect against Alzheimer's disease? Lancet Neurol 1(1):11 (2002).
- Tolppanen AM, Taipale H, Koponen M, Lavikainen P, Tanskanen A, Tiihonen J, *et al.* Use of existing data sources in clinical epidemiology: Finnish health care registers in Alzheimer's disease research the Medication use among persons with Alzheimer's disease (MEDALZ-2005) study. Clin Epidemiol 5: 277-285 (2013).
- [12] Alzheimer's-Association 2012 Alzheimer's disease facts and figures. Alzheimers Dement 8(2):131-168 (2012).
- [13] Abdel Moneim AE. Citrus peel extract attenuates acute cyanide poisoning-induced seizures and oxidative stress in rats. CNS Neurol Disord Drug Targets 13(4): 638-646 (2014).
- [14] El-Khadragy MF, Al-Olayan EM, Abdel Moneim AE. Neuroprotective effects of *Citrus reticulata* in scopolamineinduced dementia oxidative stress in rats. CNS Neurol Disord Drug Targets 13(4): 684-690 (2014).
- [15] Rogawski MA, Wenk GL. The Neuropharmacological Basis for the Use of Memantine in the Treatment of Alzheimer's Disease. CNS Drug Reviews 9(3): 275-308 (2003).
- [16] Enns GM, Kinsman SL, Perlman SL, Spicer KM, Abdenur JE, Cohen BH, *et al.* Initial experience in the treatment of inherited mitochondrial disease with EPI-743. Mol Genet Metab 105(1): 91-102 (2012).
- [17] Jomova K, Vondrakova D, Lawson M, Valko M. Metals, oxidative stress and neurodegenerative disorders. Mol Cell Biochem 345(1-2): 91-104 (2010).
- [18] Abdel Moneim AE. The Neuroprotective Effects of Purslane (*Portulaca oleracea*) on Rotenone- Induced Biochemical Changes and Apoptosis in Brain of Rat. CNS Neurol Disord Drug Targets 12(6): 830-841 (2013).
- [19] Zhao Y, Zhao B. Oxidative stress and the pathogenesis of Alzheimer's disease. Oxid Med Cell Longev 2013: 316523 (2013).
- [20] Moreira PI, Nunomura A, Nakamura M, Takeda A, Shenk JC, Aliev G, et al. Nucleic acid oxidation in Alzheimer disease. Free Radic Biol Med 44(8): 1493-1505 (2008).
- [21] Aliyev A, Chen SG, Seyidova D, Smith MA, Perry G, de la Torre J, et al. Mitochondria DNA deletions in atherosclerotic hypoperfused brain microvessels as a primary target for the development of Alzheimer's disease. J Neurol Sci 229-230: 285-292 (2005).
- [22] Bradley-Whitman MA, Timmons MD, Beckett TL, Murphy MP, Lynn BC, Lovell MA. Nucleic acid oxidation: an early feature of Alzheimer's disease. J Neurochem 128(2): 294-304 (2014).
- [23] Beal MF. Oxidatively modified proteins in aging and disease. Free Radic Biol Med 32(9): 797-803 (2002).
- [24] Munch G, Gerlach M, Sian J, Wong A, Riederer P Advanced glycation end products in neurodegeneration: more than early markers of oxidative stress? Ann Neurol 44 (3 Suppl 1): S85-88 (1998).
- [25] Smith MA, Taneda S, Richey PL, Miyata S, Yan SD, Stern D, Sayre LM, Monnier VM, Perry G Advanced Maillard reaction end products are associated with Alzheimer disease pathology. Proc Natl Acad Sci USA 91 (12): 5710-5714 (1994).
- [26] Tuppo EE, Forman LJ Free radical oxidative damage and Alzheimer's disease. J Am Osteopath Assoc 101 (12 Suppl Pt 1): S11-15 (2001).

- [27] Vitek MP, Bhattacharya K, Glendening JM, Stopa E, Vlassara H, Bucala R, Manogue K, Cerami A Advanced glycation end products contribute to amyloidosis in Alzheimer disease. Proc Natl Acad Sci USA 91 (11): 4766-4770 (1994).
- [28] Padurariu M, Ciobica A, Hritcu L, Stoica B, Bild W, Stefanescu C Changes of some oxidative stress markers in the serum of patients with mild cognitive impairment and Alzheimer's disease. Neurosci Lett 469(1): 6-10 (2010).
- [29] Ansari MA, Scheff SW Oxidative stress in the progression of Alzheimer disease in the frontal cortex. J Neuropathol Exp Neurol 69(2): 155-167 (2010).
- [30] Abdel Moneim AE. The neuroprotective effect of berberine in mercury-induced neurotoxicity in rats. Metab Brain Dis (2015).
- [31] Williams TI, Lynn BC, Markesbery WR, Lovell MA. Increased levels of 4-hydroxynonenal and acrolein, neurotoxic markers of lipid peroxidation, in the brain in Mild Cognitive Impairment and early Alzheimer's disease. Neurobiol Aging 27(8): 1094-1099 (2006).
- [32] Feng Y, Wang X. Antioxidant therapies for Alzheimer's disease. Oxid Med Cell Longev 2012: 472932. (2012).
- [33] Wei H, Xie Z. Anesthesia, calcium homeostasis and Alzheimer's disease. Curr Alzheimer Res 6(1): 30-35 (2009).
- [34] Greenough MA, Camakaris J, Bush AI. Metal dyshomeostasis and oxidative stress in Alzheimer's disease. Neurochem Int 62 (5): 540-555 (2013).
- [35] Gonzalez-Dominguez R, Garcia-Barrera T, Gomez-Ariza JL. Homeostasis of metals in the progression of Alzheimer's disease. Biometals 27(3): 539-549 (2014).
- [36] Lee JY, Mook-Jung I, Koh JY. Histochemically reactive zinc in plaques of the Swedish mutant beta-amyloid precursor protein transgenic mice. J Neurosci 19(11): RC10 (1999).
- [37] Craddock TJ, Tuszynski JA, Chopra D, Casey N, Goldstein LE, Hameroff SR, et al. The zinc dyshomeostasis hypothesis of Alzheimer's disease. PLoS One 7(3): e33552 (2012).
- [38] Lannfelt L, Blennow K, Zetterberg H, Batsman S, Ames D, Harrison J, et al. Safety, efficacy, and biomarker findings of PBT2 in targeting Abeta as a modifying therapy for Alzheimer's disease: a phase IIa, double-blind, randomised, placebo-controlled trial. Lancet Neurol 7(9):779-786 (2008).
- [39] Madeo J, Elsayad C. The Role of Oxidative Stress in Alzheimer's Disease. J Alzheimers Dis Parkinsonism 3(116). (2013).
- [40] Silva DF, Selfridge JE, Lu J, EL, Cardoso SM, Swerdlow RH. Mitochondrial abnormalities in Alzheimer's disease: possible targets for therapeutic intervention. Adv Pharmacol 64: 83-126 (2012).
- [41] Calkins MJ, Manczak M, Reddy PH. Mitochondria-Targeted Antioxidant SS31 Prevents Amyloid Beta-Induced Mitochondrial Abnormalities and Synaptic Degeneration in Alzheimer's Disease. Pharmaceuticals (Basel) 5(10): 1103-1119 (2012).
- [42] Hauptmann S, Scherping I, Drose S, Brandt U, Schulz KL, Jendrach M, et al. Mitochondrial dysfunction: an early event in Alzheimer pathology accumulates with age in AD transgenic mice. Neurobiol Aging 30(10): 1574-1586 (2009).
- [43] Zhu X, Perry G, Smith MA, Wang X. Abnormal mitochondrial dynamics in the pathogenesis of Alzheimer's disease. J Alzheimers Dis 33(1): S253-262 (2013).
- [44] Wang X, Perry G, Smith MA, Zhu X. Amyloid-beta-derived diffusible ligands cause impaired axonal transport of mitochondria in neurons. Neurodegener Dis 7(1-3): 56-59 (2010).
- [45] Garcia-Escudero V, Martin-Maestro P, Perry G, Avila J. Deconstructing mitochondrial dysfunction in Alzheimer disease. Oxid Med Cell Longev 2013:162152 (2013).
- [46] Moreira PI, Carvalho C, Zhu X, Smith MA, Perry G. Mitochondrial dysfunction is a trigger of Alzheimer's disease pathophysiology. Biochimica et Biophysica Acta (BBA) - Mol Basis Dis 1802(1): 2-10 (2010).
- [47] Barrett E, Ross RP, O'Toole PW, Fitzgerald GF. Stanton Cgamma-Aminobutyric acid production by culturable bacteria from the human intestine. J Appl Microbiol 113(2): 411-417 (2012).
- [48] Lu B, Nagappan G, Guan X, Nathan PJ, Wren P. BDNF-based synaptic repair as a disease-modifying strategy for neurodegenerative diseases. Nat Rev Neurosci 14(6): 401-416 (2013).
- [49] Brenner SR. Blue-green algae or cyanobacteria in the intestinal micro-flora may produce neurotoxins such as Beta-N-Methylamino-L-Alanine (BMAA) which may be related to

development of amyotrophic lateral sclerosis, Alzheimer's disease and Parkinson-Dementia-Complex in humans and Equine Motor Neuron Disease in horses. Med Hypotheses 80 (1): 103 (2013).

- [50] Schwartz K, Boles BR. Microbial amyloids--functions and interactions within the host. Curr Opin Microbiol 16(1): 93-99 (2013).
- [51] Rautenberg M, Joo HS, Otto M, Peschel A. Neutrophil responses to staphylococcal pathogens and commensals via the formyl peptide receptor 2 relates to phenol-soluble modulin release and virulence. FASEB J 25(4): 1254-1263 (2011).
- [52] Hill JM, Bhattacharjee S, Pogue AI, Lukiw WJ The gastrointestinal tract microbiome and potential link to Alzheimer's disease. Front Neurol 5: 43 (2014).
- [53] Pagani L, Eckert A. Amyloid-Beta interaction with mitochondria. Int J Alzheimers Dis 2011 (2011).
- [54] Jean-Louis G, von Gizycki H, Zizi F. Melatonin effects on sleep, mood, and cognition in elderly with mild cognitive impairment. J Pineal Res 25(3):177-183 (1998).
- [55] Lin L, Huang QX, Yang SS, Chu J, Wang JZ, Tian Q. Melatonin in Alzheimer's disease. Int J Mol Sci 14(7): 14575-14593 (2013).
- [56] Li R, Shen Y, Yang LB, Lue LF, Finch C, Rogers J. Estrogen enhances uptake of amyloid beta-protein by microglia derived from the human cortex. J Neurochem 75(4): 1447-1454 (2000).
- [57] Greenfield JP, Leung LW, Cai D, Kaasik K, Gross RS, Rodriguez-Boulan E, et al. Estrogen lowers Alzheimer beta-amyloid generation by stimulating trans-Golgi network vesicle biogenesis. J Biol Chem 277(14): 12128-12136 (2002).
- [58] Mendelev N, Mehta SL, Idris H, Kumari S, Li PA. Selenite stimulates mitochondrial biogenesis signaling and enhances mitochondrial functional performance in murine hippocampal neuronal cells. PLoS One 7(10): e47910 (2012).
- [59] Song G, Zhang Z, Wen L, Chen C, Shi Q, Zhang Y, et al. Selenomethionine ameliorates cognitive decline, reduces tau hyperphosphorylation, and reverses synaptic deficit in the triple transgenic mouse model of Alzheimer's disease. J Alzheimers Dis 41(1):85-99 (2014).
- [60] Kwak HM, Jeon SY, Sohng BH, Kim JG, Lee JM, Lee KB, et al. beta-Secretase (BACE1) inhibitors from pomegranate (Punica granatum) husk. Arch Pharm Res 28(12):1328-1332 (2005).
- [61] Hartman RE, Shah A, Fagan AM, Schwetye KE, Parsadanian M, Schulman RN, et al. Pomegranate juice decreases amyloid load and improves behavior in a mouse model of Alzheimer's disease. Neurobiol Dis 24(3): 506-515 (2006).
- [62] Montiel T, Quiroz-Baez R, Massieu L, Arias C. Role of oxidative stress on beta-amyloid neurotoxicity elicited during impairment of energy metabolism in the hippocampus: protection by antioxidants. Exp Neurol 200(2): 496-508 (2006).
- [63] Devore EE, Grodstein F, van Rooij FJ, Hofman A, Stampfer MJ, Witteman JC, *et al.* Dietary antioxidants and long-term risk of dementia. Arch Neurol 67(7): 819-825 (2010).
- [64] Giraldo E, Lloret A, Fuchsberger T, Vina J. Aβ and tau toxicities in Alzheimer's are linked via oxidative stress-induced p38 activation: protective role of vitamin E. Redox Biol 2: 873-877 (2014).
- [65] Nakashima H, Ishihara T, Yokota O, Terada S, Trojanowski JQ, Lee VM, et al. Effects of alpha-tocopherol on an animal model of tauopathies. Free Radic Biol Med 37(2): 176-186 (2004).
- [66] Nadeau A, Roberge AG. Effects of vitamin B12 supplementation on choline acetyltransferase activity in cat brain. Int J Vitam Nutr Res 58(4): 402-406 (1988).
- [67] Stahl W, Sies H. Effects of carotenoids and retinoids on gap junctional communication. Biofactors 15(2-4): 95-98 (2001).
- [68] Littlejohns TJ, Henley WE, Lang IA, Annweiler C, Beauchet O, Chaves PH, et al. Vitamin D and the risk of dementia and Alzheimer disease. Neurology 83(10): 920-928 (2014).
- [69] Cole GM, Ma QL, Frautschy SA. Omega-3 fatty acids and dementia. Prostaglandins Leukot Essent Fatty Acids 81(2-3): 213-221 (2009).
- [70] Green KN, Martinez-Coria H, Khashwji H, Hall EB, Yurko-Mauro KA, Ellis L, *et al.* Dietary docosahexaenoic acid and docosapentaenoic acid ameliorate amyloid-beta and tau pathology via a mechanism involving presenilin 1 levels. J Neurosci 27(16):4385-4395 (2007).
- [71] Horecky J, Gvozdjakova A, Kucharska J, Obrenovich ME, Palacios HH, Li Y, *et al.* Effects of coenzyme Q and creatine supplementation on brain energy metabolism in rats exposed to

chronic cerebral hypoperfusion. Curr Alzheimer Res 8(8): 868-875 (2011).

- [72] Yang X, Yang Y, Li G, Wang J, Yang ES. Coenzyme Q10 attenuates beta-amyloid pathology in the aged transgenic mice with Alzheimer presenilin 1 mutation. J Mol Neurosci 34 (2): 165-171 (2008).
- [73] Abdel Moneim AE, Ortiz F, Leonardo-Mendonca RC, Vergano-Villodres R, Guerrero-Martinez JA, Lopez LC, et al. Protective effects of melatonin against oxidative damage induced by Egyptian cobra (*Naja haje*) crude venom in rats. Acta Trop 143: 58-65 (2015).
- [74] Tricoire H, Moller M, Chemineau P, Malpaux B. Origin of cerebrospinal fluid melatonin and possible function in the integration of photoperiod. Reprod Suppl 61: 311-321 (2003).
- [75] Cardinali DP, Furio AM, Brusco LI. Clinical aspects of melatonin intervention in Alzheimer's disease progression. Curr Neuropharmacol 8(3): 218-227 (2010).
- [76] Srinivasan V, Pandi-Perumal SR, Cardinali DP, Poeggeler B, Hardeland R. Melatonin in Alzheimer's disease and other neurodegenerative disorders. Behav Brain Funct 2: 15 (2006).
- [77] Poeggeler B, Miravalle L, Zagorski MG, Wisniewski T, Chyan YJ, Zhang Y, et al. Melatonin reverses the profibrillogenic activity of apolipoprotein E4 on the Alzheimer amyloid Abeta peptide. Biochemistry 40(49): 14995-15001 (2001).
- [78] Al-Olayan EM, El-Khadragy MF, Abdel Moneim AE. The protective properties of melatonin against aluminium-induced neuronal injury. Int J Exp Path (2015).
- [79] Olcese JM, Cao C, Mori T, Mamcarz MB, Maxwell A, Runfeldt MJ, et al. Protection against cognitive deficits and markers of neurodegeneration by long-term oral administration of melatonin in a transgenic model of Alzheimer disease. J Pineal Res 47(1): 82-96 (2009).
- [80] Cardinali D, Vigo D, Olivar N, Vidal M, Brusco L. Melatonin Therapy in Patients with Alzheimer's Disease. Antioxidants 3(2): 245-277 (2014).
- [81] Rosales-Corral S, Tan DX, Reiter RJ, Valdivia-Velazquez M, Martinez-Barboza G, Acosta-Martinez JP, et al. Orally administered melatonin reduces oxidative stress and proinflammatory cytokines induced by amyloid-beta peptide in rat brain: a comparative, *in vivo* study versus vitamin C and E. J Pineal Res 35(2): 80-84 (2003).
- [82] Prokai L, Prokai-Tatrai K, Perjesi P, Zharikova AD, Perez EJ, Liu R, et al. Quinol-based cyclic antioxidant mechanism in estrogen neuroprotection. Proc Natl Acad Sci USA 100(20): 11741-11746 (2003).
- [83] Simpkins JW, Perez E, Wang X, Yang S, Wen Y, Singh M. The potential for estrogens in preventing Alzheimer's disease and vascular dementia. Ther Adv Neurol Disord 2(1): 31-49 (2009).
- [84] Jaffe AB, Toran-Allerand CD, Greengard P, Gandy SE. Estrogen regulates metabolism of Alzheimer amyloid beta precursor protein. J Biol Chem 269(18): 13065-13068 (1994).
- [85] Benvenuti S, Luciani P, Vannelli GB, Gelmini S, Franceschi E, Serio M, et al. Estrogen and selective estrogen receptor modulators exert neuroprotective effects and stimulate the expression of selective Alzheimer's disease indicator-1, a recently discovered antiapoptotic gene, in human neuroblast long-term cell cultures. J Clin Endocrinol Metab 90 (3): 1775-1782 (2005).
- [86] Levin-Allerhand JA, Lominska CE, Wang J, Smith JD. 17Alphaestradiol and 17beta-estradiol treatments are effective in lowering cerebral amyloid-beta levels in AbetaPPSWE transgenic mice. J Alzheimers Dis 4(6): 449-457 (2002).
- [87] Carroll JC, Rosario ER, Chang L, Stanczyk FZ, Oddo S, LaFerla FM, *et al.* Progesterone and estrogen regulate Alzheimer-like neuropathology in female 3xTg-AD mice. J Neurosci 27(48): 13357-13365 (2007).
- [88] Green PS, Bales K, Paul S, Bu G. Estrogen therapy fails to alter amyloid deposition in the PDAPP model of Alzheimer's disease. Endocrinol 146(6): 2774-2781 (2005).
- [89] Asthana S, Craft S, Baker LD, Raskind MA, Birnbaum RS, Lofgreen CP, et al. Cognitive and neuroendocrine response to transdermal estrogen in postmenopausal women with Alzheimer's disease: results of a placebo-controlled, double-blind, pilot study. Psychoneuroendocrinol 24(6): 657-677 (1999).
- [90] Kryukov GV, Castellano S, Novoselov SV, Lobanov AV, Zehtab O, Guigo R, *et al.* Characterization of mammalian selenoproteomes. Science 300(5624): 1439-1443 (2003).

- [91] Pinton S, Bruning CA, Sartori Oliveira CE, Prigol M, Nogueira CW. Therapeutic effect of organoselenium dietary supplementation in a sporadic dementia of Alzheimer's type model in rats. J Nutr Biochem 24(1): 311-317 (2013).
- [92] Loef M, Schrauzer GN, Walach H. Selenium and Alzheimer's disease: a systematic review. J Alzheimers Dis 26(1): 81-104 (2011).
- [93] Savarino L, Granchi D, Ciapetti G, Cenni E, Ravaglia G, Forti P, et al. Serum concentrations of zinc and selenium in elderly people: results in healthy nonagenarians/centenarians. Exp Gerontol 36(2): 327-339 (2001).
- [94] Akbaraly TN, Hininger-Favier I, Carriere I, Arnaud J, Gourlet V, Roussel AM, *et al.* Plasma selenium over time and cognitive decline in the elderly. Epidemiology 18(1): 52-58 (2007).
- [95] Cardoso BR, Ong TP, Jacob-Filho W, Jaluul O, Freitas MI, Cozzolino SM. Nutritional status of selenium in Alzheimer's disease patients. Br J Nutr 103(6): 803-806 (2010).
- [96] Gao S, Jin Y, Hall KS, Liang C, Unverzagt FW, Ma F, et al. Selenium level is associated with apoE epsilon4 in rural elderly Chinese. Public Health Nutr 12(12): 2371-2376 (2009).
- [97] Ishrat T, Parveen K, Khan MM, Khuwaja G, Khan MB, Yousuf S, et al. Selenium prevents cognitive decline and oxidative damage in rat model of streptozotocin-induced experimental dementia of Alzheimer's type. Brain Res 1281: 117-127 (2009).
- [98] van Eersel J, Ke YD, Liu X, Delerue F, Kril JJ, Gotz J, et al. Sodium selenate mitigates tau pathology, neurodegeneration, and functional deficits in Alzheimer's disease models. Proc Natl Acad Sci USA 107(31): 13888-13893 (2010).
- [99] Lovell MA, Xiong S, Lyubartseva G, Markesbery WR. Organoselenium (Sel-Plex diet) decreases amyloid burden and RNA and DNA oxidative damage in APP/PS1 mice. Free Radic Biol Med 46(11): 1527-1533 (2009).
- [100] Budimir A. Metal ions, Alzheimer's disease and chelation therapy. Acta Pharm 61(1): 1-14 (2011).
- [101] Crapper McLachlan DR, Dalton AJ, Kruck TP, Bell MY, Smith WL, Kalow W, *et al.* Intramuscular desferrioxamine in patients with Alzheimer's disease. Lancet 337(8753): 1304-1308 (1991).
- [102] Abdel Moneim AE. Flaxseed oil as a neuroprotective agent on lead acetate-induced monoamineric alterations and neurotoxicity in rats. Biol Trace Elem Res 148(3): 363-370 (2012).
- [103] Abdel Moneim AE. Evaluating the potential role of pomegranate peel in aluminum-induced oxidative stress and histopathological alterations in brain of female rats. Biol Trace Elem Res 150(1-3): 328-336 (2012).
- [104] Kim SJ, Jeong HJ, Lee KM, Myung NY, An NH, Yang WM, et al. Epigallocatechin-3-gallate suppresses NF-kappaB activation and phosphorylation of p38 MAPK and JNK in human astrocytoma U373MG cells. J Nutr Biochem 18 (9): 587-596 (2007).
- [105] Han X, Shen T, Lou H. Dietary Polyphenols and Their Biological Significance. Internat J Mol Sci 8(9): 950-988 (2007).
- [106] Jung JY, Mo HC, Yang KH, Jeong YJ, Yoo HG, Choi NK, et al. Inhibition by epigallocatechin gallate of CoCl2-induced apoptosis in rat PC12 cells. Life Sci 80(15): 1355-1363 (2007).
- [107] Feng Y, Yang SG, Du XT, Zhang X, Sun XX, Zhao M, et al. Ellagic acid promotes Abeta42 fibrillization and inhibits Abeta42induced neurotoxicity. Biochem Biophys Res Commun 390(4): 1250-1254 (2009).
- [108] Rojanathammanee L, Puig KL, Combs CK. Pomegranate polyphenols and extract inhibit nuclear factor of activated T-cell activity and microglial activation *in vitro* and in a transgenic mouse model of Alzheimer disease. J Nutr 143(5): 597-605 (2013).
- [109] Choi SJ, Lee JH, Heo HJ, Cho HY, Kim HK, Kim CJ, et al. Punica granatum protects against oxidative stress in PC12 cells and oxidative stress-induced Alzheimer's symptoms in mice. J Med Food 14(7-8): 695-701 (2011).
- [110] Ray B, Chauhan NB, Lahiri DK. Oxidative insults to neurons and synapse are prevented by aged garlic extract and S-allyl-L-cysteine treatment in the neuronal culture and APP-Tg mouse model. J Neurochem 117(3): 388-402 (2011).
- [111] Ray B, Chauhan NB, Lahiri DK. The "aged garlic extract:" (AGE) and one of its active ingredients S-allyl-L-cysteine (SAC) as potential preventive and therapeutic agents for Alzheimer's disease (AD). Curr Med Chem 18(22): 3306-3313.
- [112] Peng Q, Buz'Zard AR, Lau BH. Neuroprotective effect of garlic compounds in amyloid-beta peptide-induced apoptosis *in vitro*. Med Sci Monit 8(8): BR328-337 (2002).

- [113] Gupta VB, Rao KS. Anti-amyloidogenic activity of S-allyl-Lcysteine and its activity to destabilize Alzheimer's beta-amyloid fibrils *in vitro*. Neurosci Lett 429(2-3): 75-80 (2007).
- [114] Chauhan NB. Effect of aged garlic extract on APP processing and tau phosphorylation in Alzheimer's transgenic model Tg2576. J Ethnopharmacol 108(3): 385-394 (2006).
- [115] Kosuge Y, Koen Y, Ishige K, Minami K, Urasawa H, Saito H, et al. S-allyl-L-cysteine selectively protects cultured rat hippocampal neurons from amyloid beta-protein- and tunicamycin-induced neuronal death. Neuroscience 122(4): 885-895 (2003).
- [116] Murata N, Murakami K, Ozawa Y, Kinoshita N, Irie K, Shirasawa T, et al. Silymarin attenuated the amyloid beta plaque burden and improved behavioral abnormalities in an Alzheimer's disease mouse model. Biosci Biotechnol Biochem 74(11): 2299-2306 (2010).
- [117] Lu P, Mamiya T, Lu LL, Mouri A, Zou L, Nagai T, et al. Silibinin prevents amyloid beta peptide-induced memory impairment and oxidative stress in mice. Br J Pharmacol 157(7): 1270-1277 (2009).
- [118] Sano M, Ernesto C, Thomas RG, Klauber MR, Schafer K, Grundman M, et al. A controlled trial of selegiline, alphatocopherol, or both as treatment for Alzheimer's disease. The Alzheimer's Disease Cooperative Study. N Engl J Med 336(17):1216-1222 (1997).
- [119] Dysken MW, Sano M, Asthana S, Vertrees JE, Pallaki M, Llorente M, et al. Effect of vitamin E and memantine on functional decline in Alzheimer disease: the TEAM-AD VA cooperative randomized trial. JAMA 311(1): 33-44 (2014).
- [120] Morris MC, Evans DA, Tangney CC, Bienias JL, Wilson RS, Aggarwal NT, et al. Relation of the tocopherol forms to incident Alzheimer disease and to cognitive change. Am J Clin Nutr 81(2): 508-514 (2005).
- [121] Dias-Santagata D, Fulga TA, Duttaroy A, Feany MB. Oxidative stress mediates tau-induced neurodegeneration in Drosophila. J Clin Invest 117(1): 236-245 (2007).
- [122] Barnard ND, Bush AI, Ceccarelli A, Cooper J, de Jager CA, Erickson KI, et al. Dietary and lifestyle guidelines for the prevention of Alzheimer's disease. Neurobiol Aging 35(2): S74-78 (2014).
- [123] Chhillar N, Singh NK, Banerjee BD, Bala K, Basu M, Sharma D. Intergenotypic variation of Vitamin B12 and Folate in AD: In north indian population. Ann Indian Acad Neurol 17(3): 308-312 (2014).
- [124] Moore E, Mander A, Ames D, Carne R, Sanders K, Watters D. Cognitive impairment and vitamin B12: a review. Int Psychogeriatr 24(4): 541-556 (2012).
- [125] Morris MC, Schneider JA, Tangney CC. Thoughts on B-vitamins and dementia. J Alzheimers Dis 9(4): 429-433 (2006).
- [126] Perrig WJ, Perrig P, Stahelin HB. The relation between antioxidants and memory performance in the old and very old. J Am Geriatr Soc 45(6): 718-724 (1997).
- [127] Kesse-Guyot E, Andreeva VA, Ducros V, Jeandel C, Julia C, Hercberg S, *et al.* Carotenoid-rich dietary patterns during midlife and subsequent cognitive function. Br J Nutr 111(5): 915-923 (2014).
- [128] Johnson EJ, Vishwanathan R, Johnson MA, Hausman DB, Davey A, Scott TM, *et al.* Relationship between Serum and Brain Carotenoids, alpha-Tocopherol, and Retinol Concentrations and Cognitive Performance in the Oldest Old from the Georgia Centenarian Study. J Aging Res 2013: 951786 (2013).
- [129] Annweiler C, Montero-Odasso M, Hachinski V, Seshadri S, Bartha R, Beauchet O. Vitamin D concentration and lateral cerebral ventricle volume in older adults. Mol Nutr Food Res 57(2): 267-276 (2013).
- [130] Balion C, Griffith LE, Strifler L, Henderson M, Patterson C, Heckman G, et al. Vitamin D, cognition, and dementia: a systematic review and meta-analysis. Neurology 79(13): 1397-1405 (2012).
- [131] Lu'o'ng KV, Nguyen LT. The role of vitamin D in Alzheimer's disease: possible genetic and cell signaling mechanisms. Am J Alzheimers Dis Other Demen 28(2):126-136 (2013).
- [132] Bierhaus A, Humpert PM, Morcos M, Wendt T, Chavakis T, Arnold B, et al. Understanding RAGE, the receptor for advanced glycation end products. J Mol Med (Berl) 83 (11): 876-886 (2005).
- [133] Masoumi A, Goldenson B, Ghirmai S, Avagyan H, Zaghi J, Abel K, et al. 1alpha,25-dihydroxyvitamin D3 interacts with curcuminoids to stimulate amyloid-beta clearance by macrophages

of Alzheimer's disease patients. J Alzheimers Dis 17(3): 703-717 (2009).

- [134] Mizwicki MT, Menegaz D, Zhang J, Barrientos-Duran A, Tse S, Cashman JR, et al. Genomic and nongenomic signaling induced by 1alpha,25(OH)2-vitamin D3 promotes the recovery of amyloidbeta phagocytosis by Alzheimer's disease macrophages. J Alzheimers Dis 29 (1): 51-62 (2012).
- [135] Yu J, Gattoni-Celli M, Zhu H, Bhat NR, Sambamurti K, Gattoni-Celli S, *et al.* Vitamin D3-enriched diet correlates with a decrease of amyloid plaques in the brain of AbetaPP transgenic mice. J Alzheimers Dis 25(2): 295-307 (2011).
- [136] Dursun E, Gezen-Ak D, Yilmazer S. A novel perspective for Alzheimer's disease: vitamin D receptor suppression by amyloidbeta and preventing the amyloid-beta induced alterations by vitamin D in cortical neurons. J Alzheimers Dis 23 (2): 207-219 (2011).
- [137] Mukherjee PK, Marcheselli VL, Serhan CN, Bazan NG. Neuroprotectin D1: a docosahexaenoic acid-derived docosatriene protects human retinal pigment epithelial cells from oxidative stress. Proc Natl Acad Sci USA 101(22): 8491-8496 (2004).
- [138] Jose Gagliardi R. Omega 3 fatty acid for the prevention of cognitive decline and dementia. Sao Paulo Med J 130(6): 419 (2012).
- [139] Cole GM, Lim GP, Yang F, Teter B, Begum A, Ma Q, et al. Prevention of Alzheimer's disease: Omega-3 fatty acid and phenolic anti-oxidant interventions. Neurobiol Aging 26(1): 133-136 (2005).
- [140] Sima AA, Li ZG. Diabetes and Alzheimer's disease is there a connection? Rev Diabet Stud 3(4): 161-168 (2006).
- [141] Grimm MO, Kuchenbecker J, Grosgen S, Burg VK, Hundsdorfer B, Rothhaar TL, *et al.* Docosahexaenoic acid reduces amyloid beta production via multiple pleiotropic mechanisms. J Biol Chem 286(16): 14028-14039 (2011).
- [142] Hashimoto M, Hossain S. Neuroprotective and ameliorative actions of polyunsaturated fatty acids against neuronal diseases: beneficial effect of docosahexaenoic acid on cognitive decline in Alzheimer's disease. J Pharmacol Sci 116 (2): 150-162 (2011).
- [143] Eckert GP, Chang S, Eckmann J, Copanaki E, Hagl S, Hener U, et al. Liposome-incorporated DHA increases neuronal survival by enhancing non-amyloidogenic APP processing. Biochim Biophys Acta 1808(1): 236-243 (2011).
- [144] Stefani M, Liguri G. Cholesterol in Alzheimer's disease: unresolved questions. Curr Alzheimer Res 6(1): 15-29 (2009).
- [145] Lim SY, Suzuki H. Intakes of dietary docosahexaenoic acid ethyl ester and egg phosphatidylcholine improve maze-learning ability in young and old mice. J Nutr 130 (6): 1629-1632 (2000).
- [146] Minami M, Kimura S, Endo T, Hamaue N, Hirafuji M, Togashi H, et al. Dietary docosahexaenoic acid increases cerebral acetylcholine levels and improves passive avoidance performance in stroke-prone spontaneously hypertensive rats. Pharmacol Biochem Behav 58(4): 1123-1129 (1997).
- [147] Yokota A. Relationship between polyunsaturated fatty acid (PUFA) and learning ability in the brain of rat fetus and newborn. Nihon Sanka Fujinka Gakkai Zasshi 45(1): 15-22 (1993).
- [148] Yamamoto N, Okaniwa Y, Mori S, Nomura M, Okuyama H. Effects of a high-linoleate and a high-alpha-linolenate diet on the learning ability of aged rats. Evidence against an autoxidationrelated lipid peroxide theory of aging. J Gerontol 46(1): B17-22 (1991).
- [149] Yavin E, Brand A, Green P. Docosahexaenoic acid abundance in the brain: a biodevice to combat oxidative stress. Nutr Neurosci 5(3): 149-157 (2002).
- [150] Hossain MS, Hashimoto M, Gamoh S, Masumura S. Antioxidative effects of docosahexaenoic acid in the cerebrum versus cerebellum and brainstem of aged hypercholesterolemic rats. J Neurochem 72(3): 1133-1138 (1999).
- [151] Green P, Glozman S, Yavin E. Ethyl docosahexaenoate-associated decrease in fetal brain lipid peroxide production is mediated by activation of prostanoid and nitric oxide pathways. Biochim Biophys Acta 1531 (1-2): 156-164 (2001).
- [152] Festenstein GN, Heaton FW, Lowe JS, Morton RA. A constituent of the unsaponifiable portion of animal tissue lipids (lambda max. 272 m mu). Biochem J 59(4): 558-566 (1955).
- [153] Crane FL, Hatefi Y, Lester RL, Widmer C. Isolation of a quinone from beef heart mitochondria. Biochim Biophys Acta 25(1): 220-221 (1957).

- [154] Crane FL. Biochemical functions of coenzyme Q10. J Am Coll Nutr 20(6): 591-598 (2001).
- [155] Forsmark-Andree P, Lee CP, Dallner G, Ernster L. Lipid peroxidation and changes in the ubiquinone content and the respiratory chain enzymes of submitochondrial particles. Free Radic Biol Med 22(3): 391-400 (1997).
- [156] Ernster L, Dallner G. Biochemical, physiological and medical aspects of ubiquinone function. Biochim Biophys Acta 1271(1): 195-204 (1995).
- [157] Matthews RT, Yang L, Browne S, Baik M, Beal MF. Coenzyme Q10 administration increases brain mitochondrial concentrations and exerts neuroprotective effects. Proc Natl Acad Sci USA 95(15): 8892-8897 (1998).
- [158] Bergamini C, Moruzzi N, Sblendido A, Lenaz G, Fato R. A water soluble CoQ10 formulation improves intracellular distribution and promotes mitochondrial respiration in cultured cells. PLoS One 7(3): e33712 (2012).
- [159] Ma D, Stokes K, Mahngar K, Domazet-Damjanov D, Sikorska M, Pandey S. Inhibition of stress induced premature senescence in presenilin-1 mutated cells with water soluble Coenzyme Q10. Mitochondrion 17: 106-115 (2014).

- [160] Sharma SK, El Refaey H, Ebadi M. Complex-1 activity and 18F-DOPA uptake in genetically engineered mouse model of Parkinson's disease and the neuroprotective role of coenzyme Q10. Brain Res Bull 70(1): 22-32 (2006).
- [161] Dumont M, Kipiani K, Yu F, Wille E, Katz M, Calingasan NY, et al. Coenzyme Q10 decreases amyloid pathology and improves behavior in a transgenic mouse model of Alzheimer's disease. J Alzheimers Dis 27(1): 211-223 (2011).
- [162] Ray B, Bisht S, Maitra A, Lahiri DK. Neuroprotective and neurorescue effects of a novel polymeric nanoparticle formulation of curcumin (NanoCurc) in the neuronal cell culture and animal model: implications for Alzheimer's disease. J Alzheimers Dis 23(1): 61-77 (2011).
- [163] Ray B, Lahiri DK. Neuroinflammation in Alzheimer's disease: different molecular targets and potential therapeutic agents including curcumin. Curr Opin Pharmacol 9(4): 434-444 (2009).
- [164] Snitz BE, O'Meara ES, Carlson MC, Arnold AM, Ives DG, Rapp SR, et al. Ginkgo biloba for preventing cognitive decline in older adults: a randomized trial. JAMA 302(24): 2663-2670 (2009).
- [165] Thal LJ, Grundman M, Berg J, Ernstrom K, Margolin R, Pfeiffer E, et al. Idebenone treatment fails to slow cognitive decline in Alzheimer's disease. Neurology 61 (11):1498-1502 (2003).

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