




Microbial Diversity in Commercial Bee Pollen from Europe, Chile, and Mexico, Based on 16S rRNA Gene Amplicon Metagenome Sequencing

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ABSTRACT Bee pollen is a highly nutritive natural foodstuff. Because of its use as a comestible, the association of bacteria with bee pollen is commercially and biologically important. We report here the bacterial diversity of seven bee pollen samples (five from Europe, one from Chile, and one from Mexico) based on 16S rRNA gene amplicon metagenome sequencing.

Bee pollen has for years been touted as a highly nutritious food (1). It is also used in the rearing of insects of agricultural interest, such as honey bees (*Apis* spp.), bumblebees (*Bombus* spp.), and syrphids (*Sphaerophoria* spp.) (2, 3). Therefore, the commercial demand for bee pollen by beekeepers has increased in recent years. Countries, such as Spain, China, Australia, and Argentina, are the main producers of bee pollen (4). One issue which affects the utility of bee pollen as a human foodstuff and in the commercial context is the association of microbes with the pollen. The structural and nutritional characteristics of bee pollen, as well as the potential for contact with the environment and with other insects during production, make bee pollen a favorable microhabitat for many microorganisms (5). Microbial diversity in pollen samples has been studied in specific areas (6). The objective of the present study was to analyze the bacterial diversity present in commercial bee pollen from Europe, Chile, and Mexico.

Bee pollen samples were obtained from three different suppliers. We worked with seven such samples, five samples from Europe, one sample from Chile, and one sample from Mexico. DNA extraction for each sample was carried out with a ZymoBIOMICS DNA minikit (Zymo Research, Irvine, CA, USA). Amplicons of the region V3-V4 for 16S rRNA genes were amplified with primers 337F/805R (7) using the Illumina MiSeq platform at Macrogen, Inc. (Seoul, Republic of Korea). The total numbers of raw sequences were 179,758 to 214,249 for the 16S rRNA gene libraries. The sequences were subjected to low-quality trimming, merging of paired reads, and elimination of reads shorter than 50 bp. To identify operational taxonomic unit (OTUs), from 122,465 to 151,548 filtered reads from 16S rRNA gene metagenomic libraries were clustered, and the processed sequences were uploaded to the service cloud for the analysis of bacterial diversity with the 16S Biodiversity Tool and classified with RDP Tools version

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2.12 (8, 9), with 97% identity. Bioinformatic analysis was carried out using Geneious software version 9.

Based on the amplicon analysis, the most prevalent phylum in all samples was the *Cyanobacteria*, with a relative abundance of 50% to 76%. The relative abundances of other phyla were 12% to 28% for *Proteobacteria*, 1% to 3% for *Thermotogae*, and 1% to 37% for *Firmicutes*. Bacterial orders with a relative abundance of greater than 1% were as follows: in the sample from Chile, *Rhodobacterales* (10%), *Rhodospirillales* (8%), *Rickettsiales* (1%), *Lactobacillales* (2%), and *Petrotogales* (1%); in the sample from Mexico, *Rhodospirillales* (5%), *Rhodobacterales* (4%), *Rickettsiales* (2%), *Pseudomonadales* (1%), *Lactobacillales* (10%), and *Petrotogales* (2%); and in the samples from Europe, *Lactobacillales* (4% to 37%), *Rhodobacterales* (6% to 18%), *Rhodospirillales* (3% to 8%), *Pseudomonadales* (1% to 3%), *Petrotogales* (1% to 3%), and *Rickettsiales* (1% to 2%).

These results extend our knowledge of the diversity of bacteria associated with bee pollen and may be useful in developing procedures for microbial control during pollen production.

Accession number(s). The sequences obtained in the present study were deposited in the Sequence Read Archive (SRA) via the National Center for Biotechnology Information (NCBI) under the numbers [SRP132301](https://doi.org/10.1016/j.biocontrol.2012.06.001) (Chilean sample), [SRP132302](https://doi.org/10.1016/j.biocontrol.2012.06.001) (Mexican sample), and [SRP132303](https://doi.org/10.1016/j.biocontrol.2012.06.001) (European samples, [SRX3654272](https://doi.org/10.1016/j.biocontrol.2012.06.001), [SRX3655212](https://doi.org/10.1016/j.biocontrol.2012.06.001), [SRX3655293](https://doi.org/10.1016/j.biocontrol.2012.06.001), [SRX3655335](https://doi.org/10.1016/j.biocontrol.2012.06.001), and [SRX3655336](https://doi.org/10.1016/j.biocontrol.2012.06.001)).

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