

# Multifunctional $\gamma\delta$ T cells and their receptors for targeted anticancer immunotherapy

Wouter Scheper, Cordula Gründer and Jürgen Kuball\*

Department of Hematology and Immunology; University Medical Center; Utrecht, The Netherlands

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**Abbreviations:** allo-SCT, allogeneic stem cell transplantation; CMV, cytomegalovirus; CTE, combinatorial- $\gamma\delta$ TCR-chain exchange; GVHD, graft *versus* host disease; TCR, T-cell receptor

Human  $\gamma\delta$  T cells possess broad antitumor reactivity and are involved in the control of viral infections. We have recently described multifunctional  $\gamma\delta$  T cells induced by cytomegalovirus after allogeneic stem cell transplantation, placing  $\gamma\delta$  T cells and their receptors in the spotlight for the development of novel anticancer immunotherapies.

Over the last few decades, cytomegalovirus (CMV) reactivation was considered a major life-threatening complication of allogeneic stem cell transplantation (allo-SCT). Nowadays, a sensitive monitoring for early CMV reactivation combined with the availability of effective antiviral treatments has rendered the CMV-related death of transplanted patients a rare event. Fortuitously, such an improved control over CMV reactivation has facilitated observational studies in large cohorts of transplanted patients, highlighting a surprising beneficial association between CMV reactivation and a reduced risk of leukemic relapse.<sup>1</sup> So far, however, how viral reactivation would provide a protection from leukemic relapse has remained unclear. Among various possibilities, it has been proposed that natural killer (NK) cells may cross-react with CMV-infected cells and tumor cells by responding to CMV-infected residual AML blasts.<sup>1</sup> In a recent issue of *Leukemia*, we propose an additional and perhaps even more physiologically relevant explanation for this apparent paradox, i.e., that  $\gamma\delta$  T cells play a pivotal role in the CMV-induced clearance of residual tumor cells.<sup>2</sup> We observed that these unconventional T cells not only expand in patients that reactivate CMV upon allo-SCT but also react against

both CMV-infected and leukemic cells. Thus, we propose that multifunctional  $\gamma\delta$  T cells could substantially contribute to the protection from leukemic relapse that is associated with CMV reactivation after allo-SCT.

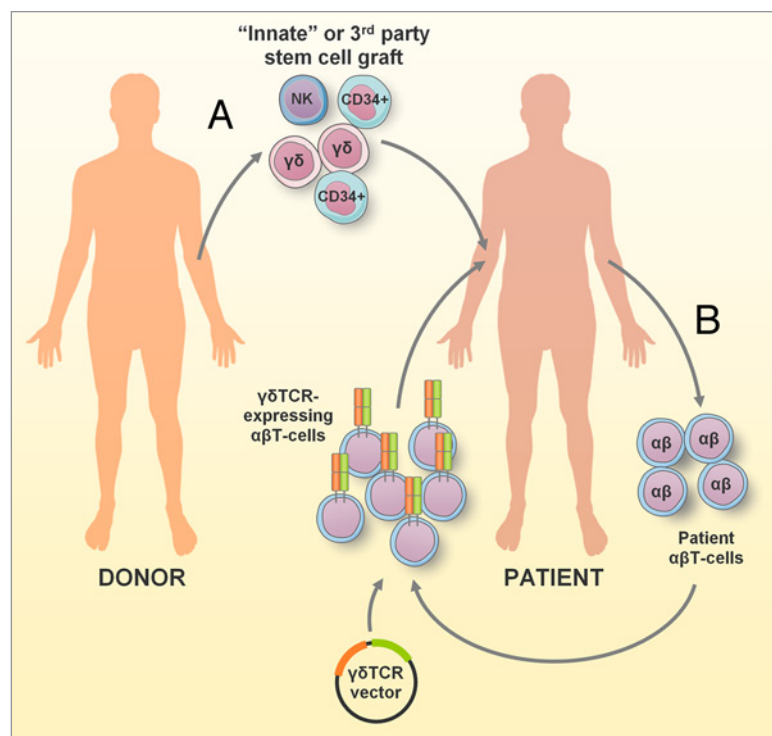
In humans, circulating  $\gamma\delta$  T cells are a minor population that mostly expresses T-cell receptors (TCRs) containing the V $\delta$ 2 and V $\gamma$ 9 gene segments (so-called V $\delta$ 2<sup>pos</sup>  $\gamma\delta$  T cells).<sup>3</sup> In contrast, epithelial  $\gamma\delta$  T cells mainly express TCRs composed of V $\delta$ 1 or V $\delta$ 3 chains (V $\delta$ 2<sup>neg</sup>  $\gamma\delta$  T cells) optionally in combination with CD8 $\alpha\alpha$ . Over the last decade, many studies have implicated V $\delta$ 2<sup>neg</sup>  $\gamma\delta$  T cells in the anti-CMV response as well as in antitumor immunosurveillance, but the first report on the cross-reactivity of these cells against CMV and cancer originated from the isolation of V $\delta$ 2<sup>neg</sup>  $\gamma\delta$  T-cell clones from kidney transplant recipients.<sup>4</sup> At least in some of these clones, the double reactivity was mediated by a  $\gamma\delta$  TCR recognizing a common stress antigen that was upregulated on both CMV-infected and transformed intestinal epithelial cells,<sup>5</sup> explaining why CMV infection alone could induce an immune cell population reacting against both CMV and cancer. Conversely, in our study, gene transfer experiments using  $\gamma\delta$  TCRs isolated from cross-reactive V $\delta$ 2<sup>neg</sup>

$\gamma\delta$  T-cell clones demonstrated the crucial involvement of the  $\gamma\delta$  TCR in tumor reactivity but not in the recognition of CMV-infected cells, suggesting that—at least in these V $\delta$ 2<sup>neg</sup>  $\gamma\delta$  T-cell clones—anti-CMV reactivity relied on receptors other than the  $\gamma\delta$  TCR. This brings up a major issue in the  $\gamma\delta$  T-cell research field, that is, the poor understanding of the mechanisms of  $\gamma\delta$  T-cell activation as well of the antigens recognized by  $\gamma\delta$  TCRs. In this respect, one important finding of our study is the identification of CD8 $\alpha\alpha$  as a co-stimulatory molecule for the activation of defined  $\gamma\delta$  TCRs. The expression of CD8 $\alpha\alpha$  on  $\gamma\delta$  T cells has previously been described, yet so far there were no reports on its function. In  $\alpha\beta$  T cells, the CD8 $\alpha\beta$  heterodimer serves as co-receptor for the  $\alpha\beta$  TCR, restricting its interaction to antigens presented on MHC Class I molecules. Conversely,  $\gamma\delta$  TCRs recognize antigens independently of MHC molecules, suggesting that the co-activating function of CD8 $\alpha\alpha$  is likely to rely on alternative mechanisms. The precise mechanisms whereby CD8 $\alpha\alpha$  delivers co-stimulatory signals in this setting remain to be elucidated. Nevertheless, we observed a striking increase of circulating CD8 $\alpha\alpha$ <sup>+</sup>  $\gamma\delta$  T cells in CMV-reactivating individuals in our patient cohort as well as in an additional independent cohort

\*Correspondence to: Jürgen Kuball; Email: j.h.e.kuball@umcutrecht.nl

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**Figure 1.** Antitumor strategies based on  $\gamma\delta$  T cells. (A) The cell preparation for an “innate allogeneic stem cell transplantation” (allo-SCT) from conventional or third party donors may selectively contain or be enriched for  $\gamma\delta$  T cells to provide anti-cytomegalovirus (CMV) and antitumor protection in the absence of graft versus host disease (GVHD). (B) In a complementary “autologous engineered transplantation,” T cells are isolated from cancer patients, expanded and engineered to express  $\gamma\delta$  TCRs (optimized by combinatorial- $\gamma\delta$ TCR-chain exchange) ex vivo. Reprogrammed T cells are subsequently re-infused into the patient, where they specifically recognize and kill tumor cells.

of congenitally CMV-infected neonates, implying that CD8 $\alpha\alpha$  expression by  $\gamma\delta$  T cells represents a pathophysiologically relevant phenomenon in vivo.

The demonstration that a proportion of  $\gamma\delta$  T cells can cross-recognize CMV and a broad panel of hematological cancer cells make them particularly attractive for clinical applications, such as adoptive cell transfer-based immunotherapy. In the context of allo-SCT, a situation favoring the reactivity of re-infused cells against CMV and leukemia in the absence of graft vs. host disease (GVHD) might be achieved with stem cell grafts enriched for  $\gamma\delta$  T cells (Fig. 1). To this end, we and others are nowadays conducting clinical trials using stem cell grafts depleted of  $\alpha\beta$  T cells

and B cells (NTR2463 and NTR3079).<sup>6</sup> Intriguingly,  $\alpha\beta$  TCR/CD19-depleted, but usually not CD3/CD19-depleted, grafts reconstitute very rapidly a broad  $\alpha\beta$  T-cell repertoire (J Kuball, unpublished observations), suggesting a very broad immunoregulatory role for  $\gamma\delta$  T cells that has recently also been proposed by others.<sup>7</sup> As an alternative, umbilical cord blood grafts can be used as a third party source of stem cells. These grafts typically contain high percentages of  $\gamma\delta$  T cells, and we have demonstrated that CMV- and leukemia-reactive  $\gamma\delta$  T cells can also be obtained from such an antigen-naïve repertoire. Importantly, all good manufacturing practice-grade clinical tools for the preparation of enriched stem cells grafts are available.

Finally, as our results suggest a central role for CD8 $\alpha\alpha$   $\gamma\delta$  T cells in anti-CMV immune responses, the isolation of these cells could be envisioned, although their precise function would have first to be deeply investigated.

Complementary to this “innate allo-SCT” approach,  $\gamma\delta$  TCRs with broad tumor-reactivity could be characterized and used to reprogram patient-derived conventional  $\alpha\beta$  T cells<sup>8</sup> (Fig. 1). Given the non-MHC-restricted antigen recognition pattern of  $\gamma\delta$  TCRs, defined  $\gamma\delta$  TCRs could—in contrast to  $\alpha\beta$  TCRs—be applied to a broad patient population in the absence of matched HLA types. Also, exogenous  $\gamma\delta$  TCR chains do not pair with their endogenous  $\alpha\beta$  counterparts, preventing the creation of novel TCRs with unpredictable (auto-) reactivity. As we have previously shown, introducing defined  $\gamma\delta$  TCRs effectively reprograms  $\alpha\beta$  T cells to kill a broad collection of tumor cells in vitro and in vivo.<sup>9</sup> In this setting, we also established a technique called combinatorial- $\gamma\delta$ TCR-chain exchange (CTE), allowing for the design of  $\gamma\delta$  TCRs with enhanced functional avidity toward malignant but not healthy tissues.<sup>10</sup> By exploiting the abundance, potent cytotoxic machinery and proliferative competence of  $\alpha\beta$  T cells even in advanced stages of disease, the engineering of autologous immune cells with such receptors would allow for the generation of large numbers of tumor-reactive T cells while tackling the major limitations of current approaches based on engineered  $\alpha\beta$  TCRs. Thus,  $\gamma\delta$  T cells and their receptors stand out as a promising avenue toward the development of new antitumor immunotherapies.

#### Disclosure of Potential Conflicts of Interest

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