

RESEARCH

Open Access



# Clinicopathological and prognostic values of fibronectin and integrin $\alpha\beta 3$ expression in primary osteosarcoma

Kai Shi<sup>1†</sup>, Sheng-lin Wang<sup>1†</sup>, Bin Shen<sup>2</sup>, Feng-qiang Yu<sup>1</sup>, Dan-feng Weng<sup>3</sup> and Jian-hua Lin<sup>1,4\*</sup>

## Abstract

**Background:** Osteosarcoma is a malignant bone tumor with a high potential for lung metastasis, and the prognosis for patients with metastatic disease is very poor. The interaction between fibronectin (FN) and integrin  $\alpha\beta 3$  in soft-tissue sarcoma promotes cell migration, invasion, and lung metastasis. This study aimed to investigate the prognostic significance of FN and  $\alpha\beta 3$  in osteosarcoma.

**Methods:** Immunohistochemistry and western blotting were used to detect the expression of FN and  $\alpha\beta 3$  in 60 osteosarcoma specimens and in 30 osteochondroma specimens. Furthermore, correlations of FN and  $\alpha\beta 3$  with the clinicopathological features of osteosarcoma patients were analyzed using the  $\chi^2$  test and Fisher's exact test. Disease-free survival and overall survival of osteosarcoma patients were assessed using the Kaplan-Meier method and Cox proportional hazards model. The predictive accuracy of the model was determined by the Harrell concordance index.

**Results:** FN ( $P < 0.05$ ) and  $\alpha\beta 3$  ( $P < 0.05$ ) were overexpressed in osteosarcoma specimens compared with osteochondroma specimens. High FN expression was associated with a poor response to chemotherapy ( $P = 0.001$ ) and poor disease-free ( $P < 0.001$ ) and overall ( $P < 0.001$ ) survival. High expression of  $\alpha\beta 3$  was linked to an advanced surgical stage ( $P = 0.028$ ), a poor response to chemotherapy ( $P = 0.002$ ), and both poor disease-free survival ( $P < 0.001$ ) and overall survival ( $P < 0.001$ ). FN and  $\alpha\beta 3$  co-expression were associated with sex ( $P = 0.011$ ), an advanced surgical stage ( $P = 0.013$ ), and a poor response to chemotherapy ( $P = 0.002$ ). Moreover, high expression of both proteins can serve as an independent prognostic value for reduced survival time in osteosarcoma patients.

**Conclusions:** The results of this study suggest that FN and  $\alpha\beta 3$  expression is associated with an unfavorable clinical outcome of osteosarcoma, and these molecules may constitute attractive therapeutic targets for osteosarcoma treatment. To improve the survival of osteosarcoma patients, further investigations are required to clarify their prognostic values in a larger population.

**Keywords:** Osteosarcoma, Fibronectin,  $\alpha\beta 3$ , Co-expression, Prognosis

\* Correspondence: [jianhual@126.com](mailto:jianhual@126.com)

<sup>†</sup>Kai Shi and Sheng-lin Wang contributed equally to this work.

<sup>1</sup>Department of Orthopaedics, the First Affiliated Hospital of Fujian Medical University, Fuzhou 350005, Fujian, People's Republic of China

<sup>4</sup>Department of Central Laboratory, the First Affiliated Hospital of Fujian Medical University, Fuzhou 350005, Fujian, People's Republic of China

Full list of author information is available at the end of the article



## Introduction

As the most frequently observed primary aggressive bone tumor, osteosarcoma occurs most often in childhood and adolescence, with a second incidence peak among individuals over 50 years of age [1]. Aggressive therapeutic modalities, including surgical resection and combinational chemotherapy, can cure 70% of patients with localized disease. However, the prognosis of patients with metastatic or relapsed osteosarcoma remains unfavorable, with no improvement over the past 30 years [1]. Thus, further investigation of the biomarkers for the prognosis of osteosarcoma is needed to develop effective agents for treatment.

In cancer development and progression, the extracellular matrix (ECM) undergoes compositional and organizational remodeling and facilitates tumor angiogenesis by regulating the dynamic behaviors of endothelial cells through various cell adhesion receptors. Accordingly, ECM proteins are potentially promising therapeutic targets [2].

Fibronectin (FN) is a multifunctional glycoprotein of the ECM that plays a crucial role in cell adhesion and angiogenesis. In the process of metastatic progression, FN acts as a potent guidance and motility cue for cancer cells via ECM remodeling and ECM-guided directional migration [3]; FN independently indicates unfavorable clinical outcomes in nasopharyngeal carcinoma [4] and head and neck squamous cell carcinomas [5]. As a bone matrix protein synthesized by osteoblasts, FN also regulates the differentiation and survival of osteoblasts [6]. Overall, high FN levels of expression are observed in osteosarcoma cell lines [7].

Integrins are cell adhesion receptors mediating tumor cell migration, proliferation, and invasion through recognition of diverse matrix ligands, including FN, collagen, and laminin [8]. Among members of the integrin family, integrin  $\alpha\beta3$  specifically binds to FN with high affinity [9]. Integrin  $\alpha\beta3$  expression is strongly increased in tumor cells, with a prominent role as a pro-angiogenic factor in the progression of various tumor types [10]. Upregulation of  $\alpha\beta3$  integrin is also involved in the exogenously induced cell migration, invasion, and anti-apoptotic activity of osteosarcoma cells [11]. Targeted imaging of integrin  $\alpha\beta3$  can be employed to specifically detect tumor location and size in osteosarcoma and may provide a potential tool in pre-operative assistance or therapy monitoring [12].

Interaction between FN and  $\alpha\beta3$  contributes to osteoblast adhesion and proliferation [13]. Furthermore, depletion of  $\alpha\beta3$  in osteosarcoma cells reduces cell adhesion and spread on FN [14]. However, the prognostic impact of FN and  $\alpha\beta3$  on osteosarcoma has yet to be explored. In this study, we analyzed FN and  $\alpha\beta3$  expression levels via immunohistochemistry and western

blotting and examined correlation of the individual expression as well as co-expression with the clinicopathological features, disease-free survival (DFS), and overall survival (OS) of patients with osteosarcoma to identify the potential clinicopathological and prognostic values of these factors in osteosarcoma.

## Materials and methods

### Patients and tissue specimens

We carried out a retrospective study of 60 patients with primary osteosarcoma who had undergone complete surgical resection at the First Affiliated Hospital of Fujian Medical University between 2009 and 2014. Histopathological diagnosis of all specimens was confirmed by a senior doctor of pathology. All patients underwent standardized neoadjuvant and postoperative chemotherapy with ifosfamide, cisplatin, and doxorubicin. Relevant clinical data were retrieved from medical records, including sex, age at diagnosis, tumor size, tumor location, histologic subtype, Enneking staging, and response to chemotherapy. Formalin-fixed and paraffin-embedded surgical tumor specimens for immunohistochemical staining were obtained from the archives of the Department of Pathology.

Follow-up of osteosarcoma patients was terminated on 31 August 2017 either by phone call or outpatient visit. To assess the development of local recurrence and distant metastasis, all patients with osteosarcoma were monitored by X-ray or lung computed tomography (CT) scans after surgical excision every 3 months during the first 3 years and every 6 months thereafter. DFS time was calculated from the date of diagnosis until the date of first tumor progression. OS time was calculated from the date of diagnosis until the date of death. Patients were censored at the date of the last follow-up if tumor progression or death had not occurred.

Written informed consent was provided by all participants involved in this study. The study protocol conformed to the ethical guidelines of the Declaration of Helsinki and was approved by the Ethics Committee of the First Affiliated Hospital of Fujian Medical University.

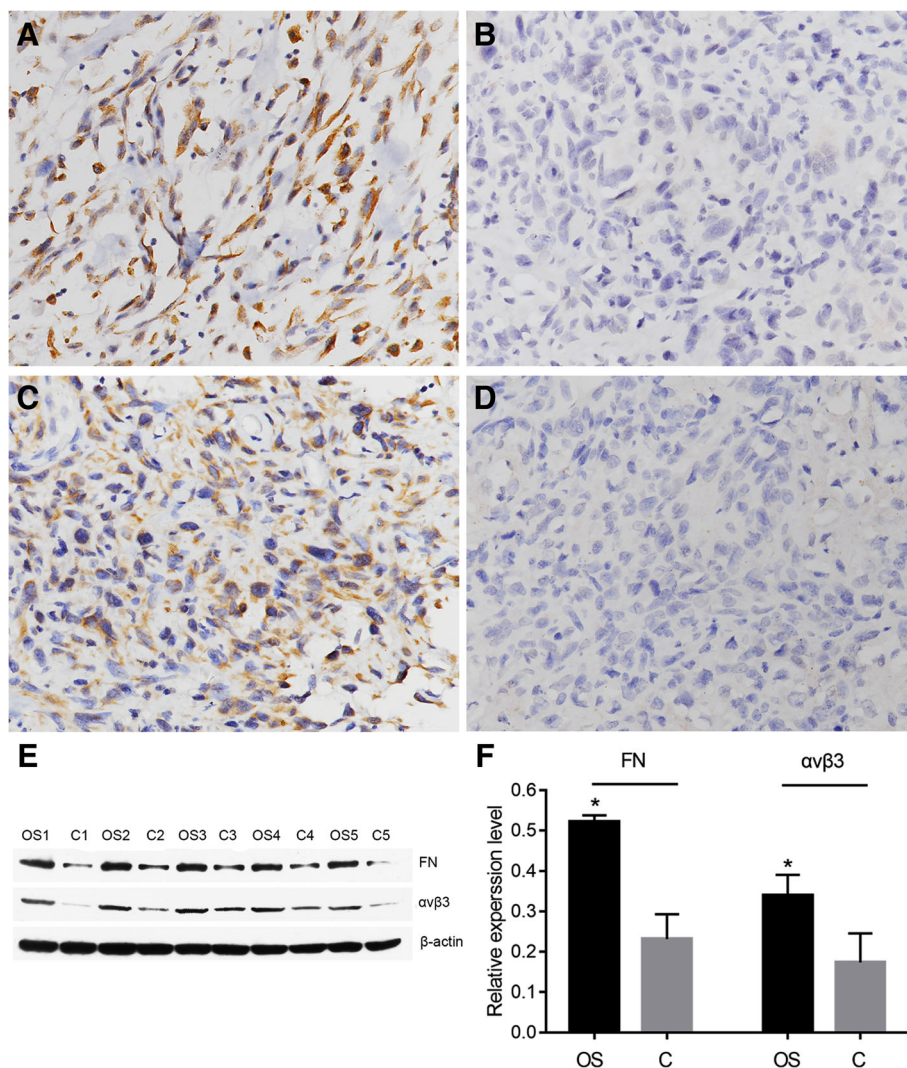
### Immunohistochemistry and scoring

Immunohistochemical expression of FN and  $\alpha\beta3$  in archival osteosarcoma specimens were examined by applying a PV-9000 Polymer Detection System (Zhongshan Goldenbridge Inc., Beijing, China), with 30 corresponding osteochondroma tissues which were resected at the First Affiliated Hospital of Fujian Medical University between 2009 and 2014 used as controls. Paraffin-embedded specimens were serially sectioned (4  $\mu\text{m}$ ) and incubated for 1 h at 60 °C. The tissue sections were then deparaffinized, hydrated, and incubated with 3% hydrogen peroxide for 10 min at room temperature to block endogenous peroxidase

activity. During the antigen retrieval process, the sections were placed in citrate buffer (pH 6.0) in an electromagnetic oven for 2 min and then allowed to cool to room temperature. The sections were incubated overnight at 4 °C with antibodies against FN (mouse monoclonal 2755-8; 1:50; Santa Cruz, USA) and  $\alpha\text{v}\beta 3$  (rabbit polyclonal orb10927; 1:50; Biorbyt, UK). Next, the sections were incubated with a polymer helper reagent for 20 min at room temperature and then poly-peroxidase-anti-mouse/rabbit IgG for 30 min at room temperature according to the manufacturer's instructions. After staining with diaminobenzidine (Zhongshan Goldenbridge Inc.), the sections were counterstained with hematoxylin, dehydrated, and

mounted. Negative (PBS (0.01 M, pH 7.2) rather than primary antibodies) and known positive (human esophagus tissue for FN and human lung cancer tissue for  $\alpha\text{v}\beta 3$ ) controls were stained in parallel with each set of sections studied.

The staining results were evaluated by two independent observers (CYP and ZZZ). Cytoplasmic staining for FN or  $\alpha\text{v}\beta 3$  in tumor cells was interpreted as a positive result. The average labelling index of FN from five random high-power fields was semi-quantitatively recorded as follows: 0, no positive staining;  $\pm$ , only a few scattered positive cells accounting for less than 20% of tumor cells; +, cluster(s) of positively stained cells accounting



**Fig. 1** Expression of FN and  $\alpha\text{v}\beta 3$  in osteosarcoma and osteochondroma specimens. Representative images of immunohistochemistry show high cytoplasmic FN expression in osteosarcoma (a) and low FN expression in osteochondroma (b) as well as high cytoplasmic  $\alpha\text{v}\beta 3$  expression in osteosarcoma (c) and low  $\alpha\text{v}\beta 3$  expression in osteochondroma (d). Original magnification,  $\times 200$ . e Representative images of western blotting show the expressions of FN and  $\alpha\text{v}\beta 3$  in the lysed osteosarcoma and osteochondroma. f Quantification of expression levels of FN and  $\alpha\text{v}\beta 3$  in osteosarcoma and osteochondroma tissues.  $\beta$ -actin was used as an internal loading control. OS, osteosarcoma; C, osteochondroma. Columns, mean from 60 or 30 tissues; bars, square deviation (\* $P < 0.05$  by an independent-sample *t* test)



**Table 1** Expression of FN and  $\alpha v\beta 3$  in osteosarcoma and corresponding osteochondroma

	Osteosarcoma, n (%)	Osteochondroma, n (%)	P value
FN			
High expression	19 (31.7)	0 (0)	0.002
Low expression	22 (36.6)	14 (46.7)	
Negative expression	19 (31.7)	16 (53.3)	
$\alpha v\beta 3$			
High expression	16 (26.7)	0 (0)	< 0.001
Low expression	24 (40.0)	8 (26.7)	
Negative expression	20 (33.3)	22 (73.3)	
FN <sup>+</sup> / $\alpha v\beta 3$ <sup>+</sup>	10 (16.7)	0 (0)	< 0.001
Others <sup>a</sup>	15 (25.0)	0 (0)	
FN <sup>-</sup> / $\alpha v\beta 3$ <sup>-</sup>	35 (58.3)	30 (100.0)	

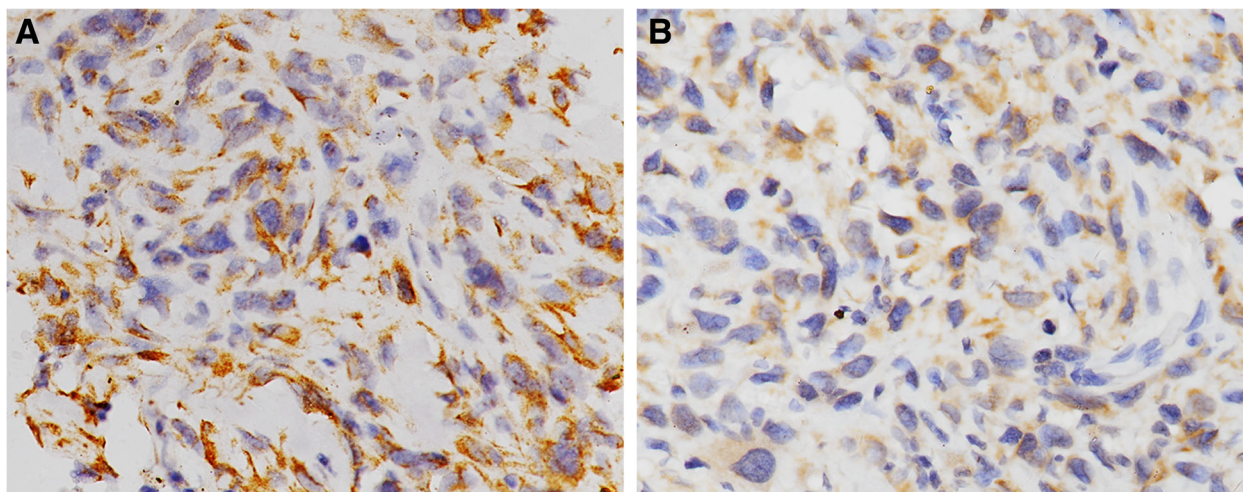
<sup>a</sup>High expression; <sup>-</sup>low/negative expression; <sup>+</sup>FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>+</sup> plus FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>-</sup>

for 20–30% of tumor cells; ++, cluster(s) of positively stained cells accounting for greater than 30% of tumor cells [15]. The average labelling index of  $\alpha v\beta 3$  from five random high-power fields was recorded as follows: 0, absent;  $\pm$ , weak expression, accounting for greater than 20% of tumor cells; +, moderate expression, accounting for greater than 20% of tumor cells; ++, strong expression, cells accounting for greater than 20% of tumor cells [16]. Specimens showing immunostaining of ++ were defined as high expression of FN or  $\alpha v\beta 3$ ; expression levels of  $\pm$  or + were defined as low expression, and 0 as negative expression.

#### Western blotting

Tissues (100 mg) of 60 osteosarcoma and 30 corresponding osteochondroma cases were ground into powder in liquid nitrogen and lysed in lysis buffer (cat. no. G2002;

Servicebio Technology, Wuhan, China). Protein concentrations in the lysates were then quantitated using a Bicinchoninic Acid Protein Assay kit (cat. no. G2026; Servicebio Technology) and preserved at  $-80^{\circ}\text{C}$ . Proteins (40  $\mu\text{g}$ ) were separated by 10% SDS-PAGE and transferred to PVDF membranes. The membranes were incubated with primary anti-FN (1:1000; Santa Cruz) and anti- $\alpha v\beta 3$  (1:1000; Biorbyt) antibodies overnight at  $4^{\circ}\text{C}$ , followed by incubation with a horseradish peroxidase-conjugated secondary antibody (cat. no. GB23404; 1:3000; Servicebio Technology) for 1 h at  $37^{\circ}\text{C}$ . The signal was visualized using an Enhanced Chemiluminescence detection system (Amersham Biosciences, UK) and Image Lab software version 3.0 (Bio-Rad Laboratories, Inc., USA).  $\beta$ -actin was simultaneously detected using mouse anti- $\beta$ -actin antibody (1:5000; Servicebio Technology) as a loading control.



**Fig. 2** Co-expression of FN and  $\alpha v\beta 3$  in one osteosarcoma specimen. Immunohistochemical staining showed high expression of FN (a) and  $\alpha v\beta 3$  (b) in one osteosarcoma specimen. Original magnification,  $\times 200$

**Table 2** Correlation between FN and  $\alpha v\beta 3$  expression in osteosarcoma

FN	$\alpha v\beta 3$			<i>r</i>	<i>P</i> value
	High expression	Low expression	Negative expression		
High expression	10	5	4	0.379	0.003
Low expression	3	14	5		
Negative expression	3	5	11		

*r* correlation coefficient

### Statistical analysis

The chi-square test, Fisher's exact test, or Student's *t* test (independent-sample) was used to compare FN and  $\alpha v\beta 3$  expression between osteosarcoma and osteochondroma and to determine whether their expression was correlated with the clinicopathological data of the osteosarcoma patients. Spearman's rank coefficient was applied to determine the correlation between FN and  $\alpha v\beta 3$  expression. Kaplan-Meier survival plots were employed for univariate analysis and the log-rank test was utilized to compare differences in survival distributions. The Cox proportional hazards model was used to perform multivariate analysis for all parameters significant in the univariate analysis. The Harrell concordance index (C-index) was calculated to measure the

performance of the model. All statistical analyses were performed using SPSS software version 19.0 (SPSS Inc., Chicago, USA).  $P < 0.05$  was considered statistically significant.

### Results

#### Expression of FN and $\alpha v\beta 3$ in osteosarcoma and osteochondroma specimens

FN and  $\alpha v\beta 3$  protein distribution were primarily observed in the cytoplasm of tumor cells (Fig. 1). The statistical results of immunohistochemistry are summarized in Table 1. FN and integrin  $\alpha v\beta 3$  were highly expressed in 19 (31.7%) and 16 (26.7%) of 60 osteosarcoma cases, which were not observed in the 30 osteochondroma cases. FN ( $P = 0.002$ ) and  $\alpha v\beta 3$  ( $P < 0.001$ ) showed higher

**Table 3** Association between individual FN and  $\alpha v\beta 3$  expression and clinicopathological characteristics in osteosarcoma

Clinicopathologic data	Case number	FN			$\alpha v\beta 3$		
		High ( <i>n</i> = 19)	Low/Neg ( <i>n</i> = 41)	<i>P</i> value	High ( <i>n</i> = 16)	Low/Neg ( <i>n</i> = 44)	<i>P</i> value
Sex							
Male	38	11	27	NS	9	29	NS
Female	22	8	14		7	15	
Age (years)							
< 18	26	6	20	NS	8	18	NS
≥ 18	34	13	21		8	26	
Tumor size (cm)							
< 5	23	6	17	NS	4	19	NS
≥ 5	37	13	24		12	25	
Tumor location							
Tibia or femur	40	11	25	NS	11	29	NS
Other location	20	4	16		5	15	
Histologic subtype							
Conventional	55	17	38	NS	14	41	NS
Special	5	2	3		2	3	
Enneking staging							
I-IIA	21	4	17	NS	2	19	0.028
IIB	39	15	24		14	25	
Response to chemotherapy <sup>a</sup>							
Good	28	3	25	0.001	2	26	0.002
Poor	32	16	16		14	19	

Neg negative, NS no significance

<sup>a</sup>Good: tumor necrosis ≥ 90%, poor: tumor necrosis < 90%

**Table 4** Association between co-expression of FN and αvβ3 and clinicopathological characteristics in osteosarcoma

Clinicopathologic data	Case number	FN <sup>+</sup> /αvβ3 <sup>+</sup> (n = 10)	Others <sup>a</sup> (n = 39)	FN <sup>-</sup> /αvβ3 <sup>-</sup> (n = 11)	P value
Sex					
Male	38	7	20	11	0.011
Female	22	3	19	0	
Age (years)					
< 18	26	5	15	6	NS
≥ 18	34	5	24	5	
Tumor size (cm)					
< 5	23	3	16	4	NS
≥ 5	37	7	23	7	
Tumor location					
Tibia or femur	40	7	25	8	NS
Other location	20	3	14	3	
Histologic subtype					
Conventional	55	9	36	10	NS
Special	5	1	3	1	
Enneking staging					
I-IIA	21	2	11	8	0.013
IIB	39	8	28	3	
Response to chemotherapy*					
Good	28	0	20	8	0.002
Poor	32	10	19	3	

NS no significance

\*Good: tumor necrosis ≥ 90%; poor: tumor necrosis < 90%; <sup>+</sup>high expression; <sup>-</sup>low/negative expression; <sup>a</sup>FN<sup>+</sup>/αvβ3<sup>-</sup> plus FN<sup>-</sup>/αvβ3<sup>+</sup>

rates of expression in osteosarcoma than in osteochondroma. The expressional levels of the two proteins were further verified by western blotting analysis. Similarly, the expression of FN ( $P < 0.001$ ) and αvβ3 ( $P = 0.003$ ) was also found to be upregulated in osteosarcoma tissues compared with osteochondroma tissues (Fig. 1).

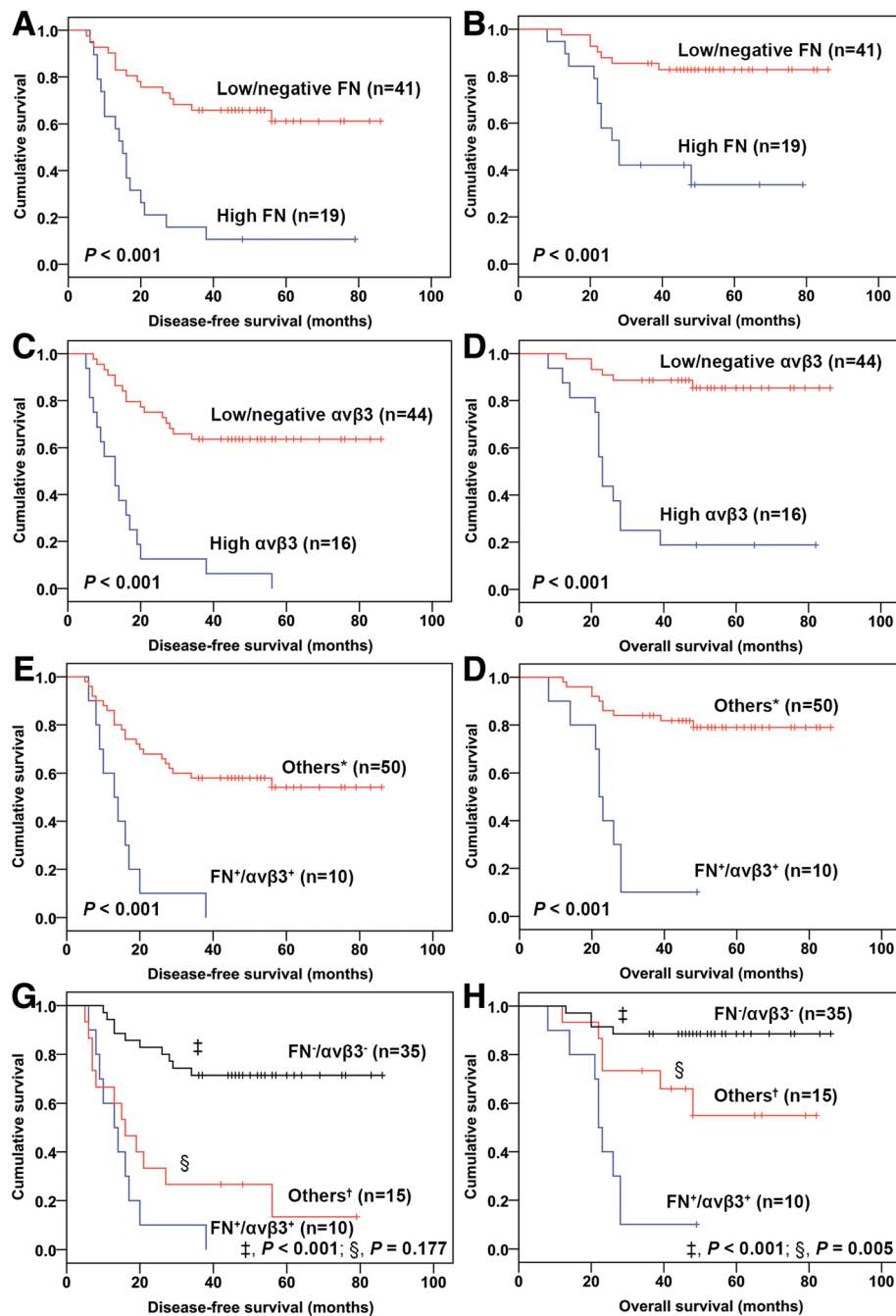
These results demonstrated that the expressional levels of FN and αvβ3 proteins were markedly increased in osteosarcoma tissues compared with the corresponding osteochondroma tissues. Furthermore, high expression (FN<sup>+</sup>/αvβ3<sup>+</sup>; Table 1; Fig. 2) and low/negative expression (FN<sup>-</sup>/αvβ3<sup>-</sup>; Table 1) of both FN and αvβ3 were

**Table 5** Univariate analysis of FN and αvβ3 expression and osteosarcoma patient survival based on the log-rank test

Characteristics	Case number	Disease-free survival (months)				Overall survival (months)				
		Mean	SD	95%CI	P value	Mean	SD	95%CI	P value	
FN										
High	19	21.74	4.83	12.28–31.19	< 0.001	42.65	6.43	30.04–55.26	< 0.001	
Low/negative	41	61.08	5.19	50.91–71.25		75.16	3.75	67.81–82.52		
αvβ3										
High	16	16.06	3.34	9.51–22.62	< 0.001	33.38	6.08	21.45–45.30	< 0.001	
Low/negative	44	61.41	4.96	51.69–71.13		77.30	3.31	70.80–83.79		
FN <sup>+</sup> /αvβ3 <sup>+</sup>	10	15.10	2.90	9.42–20.78	< 0.001	24.10	3.22	17.80–30.41	< 0.001	
vs. others <sup>a</sup>	50	55.60	4.91	45.98–65.22		73.31	3.61	66.23–80.39		
FN <sup>+</sup> /αvβ3 <sup>+</sup>	10	15.10	2.90	9.42–20.78	< 0.001 <sup>b</sup>	0.177 <sup>c</sup>	24.10	3.22	17.80–30.41	< 0.001 <sup>b</sup>
FN <sup>+</sup> /αvβ3 <sup>-</sup> plus FN <sup>-</sup> /αvβ3 <sup>+</sup>	15	27.60	6.75	14.36–40.84	< 0.001 <sup>b</sup>	58.57	7.41	44.06–73.09	0.022 <sup>b</sup>	
FN <sup>-</sup> /αvβ3 <sup>-</sup>	35	67.14	5.09	57.16–77.13		78.43	3.57	71.43–85.43		

SD standard deviation, CI confidence interval

<sup>a</sup>High expression; <sup>-</sup>low/negative expression; <sup>a</sup>FN<sup>+</sup>/αvβ3<sup>-</sup> plus FN<sup>-</sup>/αvβ3<sup>+</sup> plus FN<sup>-</sup>/αvβ3<sup>-</sup>; <sup>b</sup>vs. FN<sup>-</sup>/αvβ3<sup>-</sup>; <sup>c</sup>vs. FN<sup>+</sup>/αvβ3<sup>-</sup> plus FN<sup>-</sup>/αvβ3<sup>+</sup>



**Fig. 3** Kaplan-Meier analyses of disease-free survival (DFS) and overall survival (OS) time by FN and  $\alpha v\beta 3$  expression. **a, b** Significant differences in DFS ( $P < 0.001$ ) and OS ( $P < 0.001$ ) time were observed between high FN expression (FN<sup>+</sup>) and low/negative FN expression (FN<sup>-</sup>) groups. **c, d** Significant differences in DFS ( $P < 0.001$ ) and OS ( $P < 0.001$ ) time were observed between high  $\alpha v\beta 3$  expression ( $\alpha v\beta 3$ <sup>+</sup>) and low/negative  $\alpha v\beta 3$  expression ( $\alpha v\beta 3$ <sup>-</sup>) groups. **e, f** Significant differences in DFS ( $P < 0.001$ ) and OS ( $P < 0.001$ ) time were demonstrated between FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>+</sup> and "others" groups. **g** A significant difference in DFS ( $P < 0.001$ ) time was demonstrated between FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>+</sup> and FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>-</sup> groups. **h** Significant differences in OS time were demonstrated between FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>+</sup> and FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>-</sup> groups ( $P < 0.001$ ) and between FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>+</sup> and "others" groups ( $P = 0.005$ ). \*, "others" included the single high-expression and double low/negative-expression groups (FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>-</sup> plus FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>+</sup> plus FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>-</sup>); †, "others" included the single high-expression groups (FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>-</sup> plus FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>+</sup>)

identified in 10 and 50 osteosarcoma cases, respectively. The co-expression rates of the two proteins were significantly different in osteosarcoma and osteochondroma

specimens ( $P < 0.001$ ). A significant correlation for the level of expression was observed between FN and  $\alpha v\beta 3$  in osteosarcoma ( $r = 0.379$ ,  $P = 0.003$ , Table 2).

**Table 6** Univariate analysis of characteristics and osteosarcoma patient survival based on the log-rank test

Clinicopathologic data	Case number	Disease-free survival (months)				Overall survival (months)			
		Mean	SD	95%CI	<i>P</i> value	Mean	SD	95%CI	<i>P</i> value
Sex									
Male	38	53.81	5.59	42.85–64.76	NS	71.04	4.39	62.44–79.65	NS
Female	22	32.39	5.13	22.34–42.44		53.81	6.59	40.91–66.72	
Age (years)									
< 18	26	37.42	5.43	26.79–48.06	NS	51.51	4.42	42.84–60.17	NS
≥ 18	34	51.96	5.91	40.37–63.54		68.56	5.00	58.75–78.36	
Tumor location									
Tibia or femur	40	49.67	5.56	38.77–60.57	NS	67.47	4.52	58.60–76.33	NS
Other location	20	46.15	7.57	31.32–60.98		60.55	6.86	47.10–74.00	
Histologic subtype									
Conventional	55	49.20	4.77	39.86–58.54	NS	66.42	3.97	58.64–74.20	NS
Special	5	40.20	10.96	18.71–61.69		55.40	14.72	26.54–84.26	
Tumor size (cm)									
< 5	23	65.17	6.63	52.18–78.17	0.006	77.91	4.36	69.38–86.45	0.015
≥ 5	37	36.39	4.97	26.65–46.12		55.98	4.96	46.26–65.71	
Enneking staging									
I-IIA	21	70.05	6.27	57.75–82.34	0.001	79.71	4.25	71.39–88.04	0.010
IIB	39	36.42	5.08	26.46–46.38		56.54	4.87	47.00–66.07	
Response to chemotherapy <sup>a</sup>									
Good	28	63.86	6.14	51.83–75.89	0.002	79.11	3.76	71.74–86.48	0.001
Poor	32	32.84	4.91	23.22–42.47		51.56	5.42	40.94–62.18	

SD standard deviation, CI confidence interval, NS no significance

<sup>a</sup>Good: tumor necrosis ≥ 90%; poor: tumor necrosis < 90%

### Association between expression of FN and $\alpha v\beta 3$ in osteosarcoma and clinicopathological characteristics

The osteosarcoma patients' clinicopathological characteristics are summarized in the Additional file 1: Table S1. Association of FN and  $\alpha v\beta 3$  expression individually (Table 3) and together (Table 4) with clinicopathological parameters, including sex, age, tumor size, tumor location, histologic subtype, Enneking staging, and response to chemotherapy, were analyzed. Expression of FN and integrin  $\alpha v\beta 3$  was stratified according to conventional (osteoblastic, chondroblastic, and fibroblastic types) and special (small cell and telangiectatic types) osteosarcoma, rather than each histological subtype. In osteosarcoma, high FN expression was significantly associated with a poor response to chemotherapy ( $P = 0.001$ ; Table 3), whereas high expression of  $\alpha v\beta 3$  was significantly associated with advanced Enneking staging ( $P = 0.028$ ; Table 3) and a poor response to chemotherapy ( $P = 0.002$ ; Table 3). Tumors were more likely to develop to an advanced surgical stage ( $P = 0.013$ ; Table 4) and exhibit a worse response to chemotherapy ( $P = 0.002$ ; Table 4) as the combined expression levels progressed from FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>-</sup> to other groups (FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>-</sup> plus FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>+</sup>) and then an FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>+</sup> status. In addition, a sex difference

( $P = 0.011$ ) was found among the three expression levels of FN and  $\alpha v\beta 3$ .

### Association between expression of FN and $\alpha v\beta 3$ in osteosarcoma and clinical outcome

The mean patient follow-up time was 45.2 months (range 8 to 86 months). By the end of the follow-up period, 28 (46.6%) patients survived with no evidence of disease, 13 (21.7%) remained alive with disease, and 19 (31.7%) succumbed to osteosarcoma (Additional file 1: Table S1).

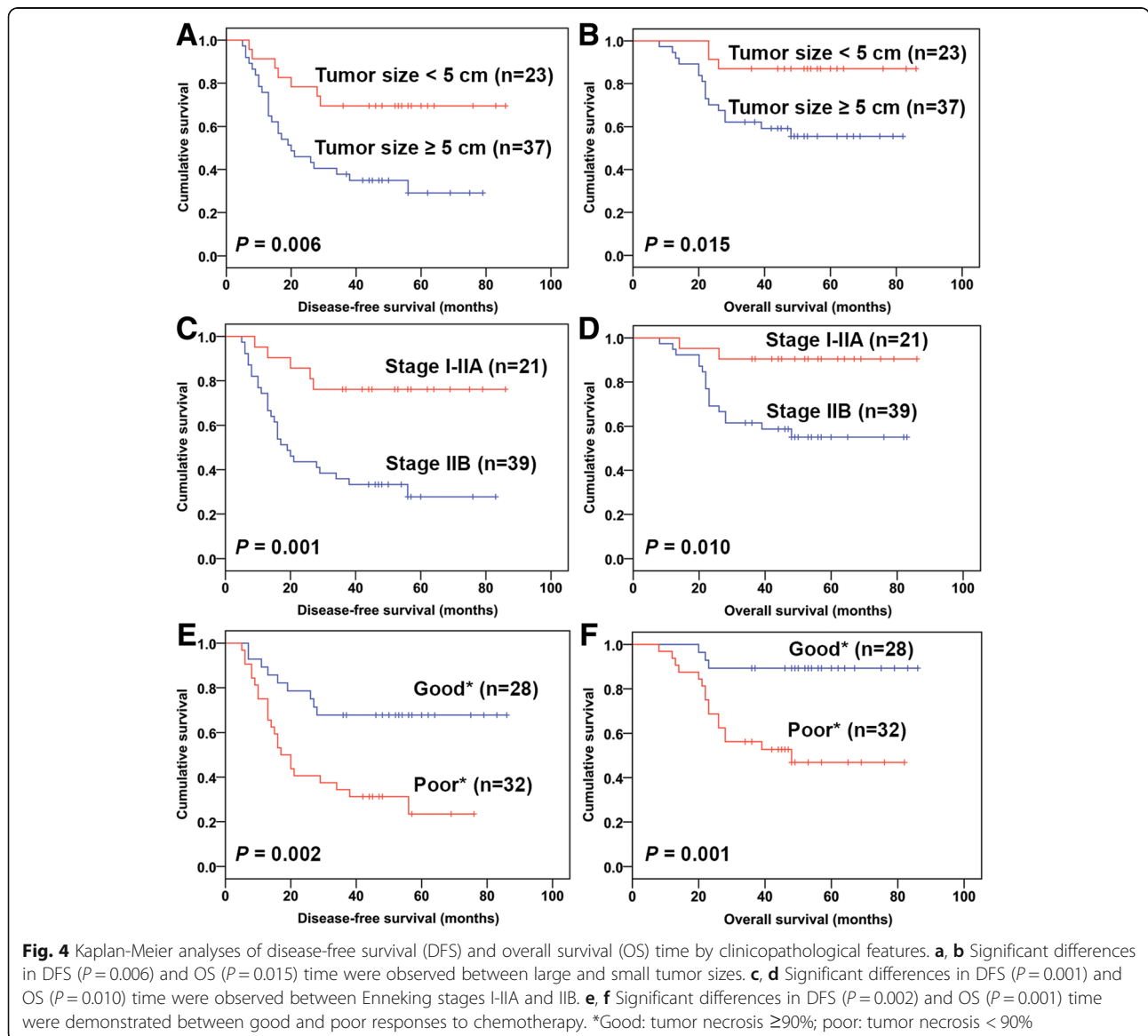
Univariate analysis of patient survival in relation to FN and  $\alpha v\beta 3$  expression levels is presented in Table 5, and survival curves are shown in Fig. 3. Mean DFS and OS times decreased with increasing expression of either FN (DFS,  $P < 0.001$ , Fig. 3a; OS,  $P < 0.001$ , Fig. 3b) or  $\alpha v\beta 3$  (DFS,  $P < 0.001$ , Fig. 3c; OS,  $P < 0.001$ , Fig. 3d). High expression of both FN and  $\alpha v\beta 3$  (FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>+</sup>) was associated with a shorter DFS time ( $P < 0.001$ , Fig. 3e) and OS time ( $P < 0.001$ , Fig. 3f) compared with the other groups (FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>-</sup> plus FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>+</sup> plus FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>-</sup>). Furthermore, the mean survival time decreased based on the extent of FN<sup>+</sup> or  $\alpha v\beta 3$ <sup>+</sup> expression, which was longest for the FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>-</sup> group (DFS, 67.14 months; OS, 78.43 months) followed by



the single high-expression groups (FN<sup>+</sup>/αvβ3<sup>-</sup> plus FN<sup>-</sup>/αvβ3<sup>+</sup>; DFS, 27.60 months; OS, 58.57 months) and then the FN<sup>+</sup>/αvβ3<sup>+</sup> group (DFS, 15.10 months; OS, 24.10 months). The DFS time difference between the FN<sup>+</sup>/αvβ3<sup>+</sup> group and FN<sup>-</sup>/αvβ3<sup>-</sup> group ( $P < 0.001$ , Fig. 3g) as well as between the single high-expression groups (FN<sup>+</sup>/αvβ3<sup>-</sup> plus FN<sup>-</sup>/αvβ3<sup>+</sup>) and the FN<sup>-</sup>/αvβ3<sup>-</sup> group ( $P < 0.001$ ) was statistically significant. A significant difference in OS time was also noted between the FN<sup>+</sup>/αvβ3<sup>+</sup> group and the FN<sup>-</sup>/αvβ3<sup>-</sup> group ( $P < 0.001$ , Fig. 3h), the single high-expression groups (FN<sup>+</sup>/αvβ3<sup>-</sup> plus FN<sup>-</sup>/αvβ3<sup>+</sup>) and the FN<sup>-</sup>/αvβ3<sup>-</sup> group ( $P = 0.022$ ), and between the FN<sup>+</sup>/αvβ3<sup>+</sup> group and the single high-expression groups ( $P = 0.005$ , Fig. 3h).

To explore the independent prognostic ability of FN and αvβ3 co-expression, three clinicopathological factors

(i.e., tumor size, Enneking staging, and response to chemotherapy) that were significant predictors of survival time in univariate analysis (Table 6, Fig. 4) were evaluated by multivariate analysis. As shown in Table 7, according to multivariate analysis, FN<sup>+</sup>/αvβ3<sup>+</sup> independently predicted worse DFS (hazard ratio (HR) = 2.66,  $P = 0.025$ , C-index = 0.75) and OS (HR = 3.75,  $P = 0.011$ , C-index = 0.84) of patients with osteosarcoma compared with other groups (FN<sup>+</sup>/αvβ3<sup>-</sup> plus FN<sup>-</sup>/αvβ3<sup>+</sup> plus FN<sup>-</sup>/αvβ3<sup>-</sup>). Compared with FN<sup>-</sup>/αvβ3<sup>-</sup>, FN<sup>+</sup>/αvβ3<sup>+</sup> exhibited a statistically more significant predictive value for shorter DFS (HR = 5.62,  $P = 0.002$ , C-index = 0.79) and OS (HR = 6.35,  $P = 0.010$ , C-index = 0.85). Similarly, high expression of FN and αvβ3 individually (FN<sup>+</sup>/αvβ3<sup>-</sup> plus FN<sup>-</sup>/αvβ3<sup>+</sup>) was significantly associated with worse DFS compared with FN<sup>-</sup>/αvβ3<sup>-</sup>. Therefore, co-expression of FN and αvβ3 was significantly



**Table 7** Co-expression of FN and  $\alpha v\beta 3$  and osteosarcoma patient survival based on multivariate analysis

Characteristics	Comparison	Disease-free survival (months)			Overall survival (months)		
		HR	95%CI	P value	HR	95%CI	P value
FN <sup>+</sup> / $\alpha v\beta 3$ <sup>+</sup>	vs. others <sup>a</sup>	2.66	1.13–6.23	0.025	3.75	1.36–10.36	0.011
Tumor size (cm)	< 5 vs. $\geq 5$	2.55	1.06–6.14	0.036	2.57	0.72–9.24	0.147
Enneking staging	I-IIA vs. IIB	3.34	1.20–9.28	0.021	2.90	0.60–13.87	0.184
Response to chemotherapy	Good vs. poor	0.75	0.30–1.84	0.526	0.48	0.12–1.99	0.309
C-index (95%CI)		0.75	0.65–0.86		0.84	0.70–0.97	
FN <sup>+</sup> / $\alpha v\beta 3$ <sup>+</sup>	vs. FN <sup>-</sup> / $\alpha v\beta 3$ <sup>-</sup>	5.62	1.91–16.52	0.002	6.35	1.54–26.14	0.010
FN <sup>+</sup> / $\alpha v\beta 3$ <sup>-</sup> plus FN <sup>-</sup> / $\alpha v\beta 3$ <sup>+</sup>	vs. FN <sup>-</sup> / $\alpha v\beta 3$ <sup>-</sup>	3.79	1.50–9.54	0.005	2.31	0.59–8.96	0.228
Tumor size (cm)	< 5 vs. $\geq 5$	2.04	0.84–4.93	0.115	2.39	0.67–8.56	0.182
Enneking staging	I-IIA vs. IIB	3.46	1.22–9.77	0.019	2.69	0.55–13.08	0.220
Response to chemotherapy	Good vs. poor	1.08	0.41–2.79	0.882	0.62	0.14–2.80	0.536
C-index (95%CI)		0.79	0.69–0.90		0.85	0.71–0.98	

HR hazard ratio, CI confidence interval

<sup>+</sup>High expression; <sup>-</sup>low/negative expression; <sup>a</sup>FN<sup>+</sup>/ $\alpha v\beta 3$ <sup>-</sup> plus FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>+</sup> plus FN<sup>-</sup>/ $\alpha v\beta 3$ <sup>-</sup>

correlated with poor DFS and OS in osteosarcoma patients.

## Discussion

Osteosarcoma is the second leading cause of cancer-related death in children and young adults due to its high metastatic potential [17]. Identification of tumor metastasis-associated biomarkers followed by development of promising therapies targeting molecular pathways will ultimately help to improve the prognosis of these patients. In this study, we found that high expression of FN or  $\alpha v\beta 3$  individually as well as their combined expression can serve as predictors for poor clinical survival among osteosarcoma patients.

The high level of FN expression in archived osteosarcoma tissues observed in our results was consistent with a previous study by Na et al. [18]. Additionally, osteosarcoma cells are better spread and have more actin stress fibers, when cultured with FN, compared with fetal bovine serum [19]. The correlation between FN and a poor response to chemotherapy of osteosarcoma found in the present study demonstrated that FN may support the aggressive potential of tumor cells. Overexpression of  $\alpha v\beta 3$  increases distant spread towards bone metastatic sites in various osteotropic tumors [20] and facilitates enhanced cell migration and metastatic potential in osteosarcoma [21]. In the present study, integrin  $\alpha v\beta 3$  was found to be upregulated in osteosarcoma and associated with advanced surgical stage and a worse response to chemotherapy, indicating the involvement of  $\alpha v\beta 3$  in treatment-resistant mechanisms.

Our results revealed that expression of FN or  $\alpha v\beta 3$  alone, as well as their co-expression, significantly contributes to the poor DFS and OS of osteosarcoma patients. Because it promotes tumor cell invasion and directional migration, FN acts as an independent

unfavorable prognostic indicator for malignant tumors [4, 5, 22], and interaction between FN and  $\alpha v\beta 3$  may serve as a regulatory point to activate osteoblast adhesion and differentiation [13]. A previous study demonstrated that the FN- $\alpha v\beta 3$  integrin axis promotes tumor cell migration, invasion, and metastasis by upregulating the activity of integrin-linked kinase [23], which is an independent prognostic factor for poor survival of osteosarcoma [24]. Thrombin-enhanced cell adhesion of osteosarcoma to FN can be inhibited by rhodostomin, which acts against integrin  $\alpha v\beta 3$  [11]. Therefore, multiple lines of evidence indicate that high FN and  $\alpha v\beta 3$  expression levels may contribute to the metastatic progression of osteosarcoma via various pathways. Antagonists targeting FN and  $\alpha v\beta 3$  are potentially able to increase the survival of patients with osteosarcoma. FN-targeted antibodies, such as L19-TNF [25] and F8-TNF [26], have demonstrated efficacy in inhibiting tumor growth and early pulmonary metastases of human osteosarcoma. Pending the results of ongoing studies, etaracizumab [27], a humanized version of LM609, which targets  $\alpha v\beta 3$ , may represent another agent for the treatment of osteosarcoma.

Some limitations in the present study should be noted. First, selection bias should be considered due to the retrospective nature of the study. In addition, the application of neoadjuvant chemotherapy before surgery has a potential to influence the fidelity of FN and  $\alpha v\beta 3$  results. Our findings warrant further investigation into the quantitative analyses of FN and  $\alpha v\beta 3$  using biopsy specimens of osteosarcoma. Third, our study cohort included a relatively small number of patients, and the follow-up time for evaluating patient survival was relatively short. Thus, further large-scale studies with a longer follow-up time should be performed to offer more convincing evidence.

## Conclusions

Our study demonstrates that the expression of FN and  $\alpha v \beta 3$  is increased in osteosarcoma specimens and associated with poor clinical outcomes. Moreover, FN and  $\alpha v \beta 3$  co-expression is independently correlated with short DFS and OS. Hence, FN and  $\alpha v \beta 3$  may represent attractive therapeutic targets for the treatment of osteosarcoma. To improve the survival of osteosarcoma patients, further investigations are required to identify the prognostic significance in a larger population.

## Additional file

**Additional file 1: Table S1.** Clinicopathological characteristics of osteosarcoma. (DOCX 13 kb)

### Abbreviations

DFS: Disease-free survival; FN: Fibronectin; OS: Overall survival

### Acknowledgements

Not applicable

### Funding

This study was supported from the National Nature Science Foundation of China (grant number: 31571292) and Startup Fund for Scientific Research, Fujian Medical University (grant number: 2017XQ1089). The funders had no role in the study design, data collection and analysis, or preparation of the manuscript.

### Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

### Authors' contributions

KS conducted the experiments. S-LW participated in collecting the data and drafted the manuscript. BS, F-QY, and D-FW contributed to the statistical analysis and manuscript writing. J-HL conceived the present study and helped revise the manuscript. All authors read and approved the final manuscript.

### Ethics approval and consent to participate

The study was conducted in accordance with the Ethics Committee of the First Affiliated Hospital of Fujian Medical University and the 1964 Helsinki Declaration. Informed consent was obtained from all participants included in the study.

### Consent for publication

Not applicable

### Competing interests

The authors declare that they have no competing interests.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

### Author details

<sup>1</sup>Department of Orthopaedics, the First Affiliated Hospital of Fujian Medical University, Fuzhou 350005, Fujian, People's Republic of China. <sup>2</sup>Faculty of Education, the Chinese University of Hong Kong, Shatin, N.T. 999077, Hong Kong, People's Republic of China. <sup>3</sup>Department of Pathology, the First Affiliated Hospital of Fujian Medical University, Fuzhou 350005, Fujian, People's Republic of China. <sup>4</sup>Department of Central Laboratory, the First Affiliated Hospital of Fujian Medical University, Fuzhou 350005, Fujian, People's Republic of China.

Received: 21 September 2018 Accepted: 15 January 2019

Published online: 28 January 2019

## References

- Kansara M, Teng MW, Smyth MJ, Thomas DM. Translational biology of osteosarcoma. *Nat Rev Cancer*. 2014;14:722–35. <https://doi.org/10.1038/nrc3838>.
- Zhou Z, Lu ZR. Molecular imaging of the tumor microenvironment. *Adv Drug Deliv Rev*. 2017;113:24–48. <https://doi.org/10.1016/j.addr.2016.07.012>.
- Oudin MJ, Jonas O, Kosciuk T, Broyle LC, Guido BC, Wyckoff J, Riquelme D, Lamar JM, Asokan SB, Whittaker C, et al. Tumor cell-driven extracellular matrix remodeling drives haptotaxis during metastatic progression. *Cancer Discov*. 2016;6:516–31. <https://doi.org/10.1158/2159-8290.CD-15-1183>.
- Ma LJ, Lee SW, Lin LC, Chen TJ, Chang IW, Hsu HP, Chang KY, Huang HY, Li CF. Fibronectin overexpression is associated with latent membrane protein 1 expression and has independent prognostic value for nasopharyngeal carcinoma. *Tumour Biol*. 2014;35:1703–12. <https://doi.org/10.1007/s13277-013-1235-8>.
- Gopal S, Veracini L, Grall D, Butori C, Schaub S, Audebert S, Camoin L, Baudelet E, Radwanska A, Beghelli-de la Forest Divonne S, et al. Fibronectin-guided migration of carcinoma collectives. *Nat Commun*. 2017;8:14105. <https://doi.org/10.1038/ncomms14105>.
- Globus RK, Doty SB, Lull JC, Holmuhamedov E, Humphries MJ, Damsky CH. Fibronectin is a survival factor for differentiated osteoblasts. *J Cell Sci*. 1998; 111(Pt 10):1385–93.
- Kilian O, Dahse R, Alt V, Zardi L, Rosenhahn J, Exner U, Battmann A, Schnettler R, Kosmehl H. Expression of EDA+ and EDB+ fibronectin splice variants in bone. *Bone*. 2004;35:1334–45. <https://doi.org/10.1016/j.bone.2004.08.008>.
- Huttenlocher A, Horwitz AR. Integrins in cell migration. *Cold Spring Harb Perspect Biol*. 2011;3:a005074. <https://doi.org/10.1101/cshperspect.a005074>.
- Benito-Jardón M, Klapproth S, Gimeno-Lluch I, Petzold T, Bharadwaj M, Müller DJ, Zuchtriegel G, Reichel CA, Costell M. The fibronectin synergy site re-enforces cell adhesion and mediates a crosstalk between integrin classes. *eLife*. 2017;6:e22264. <https://doi.org/10.7554/eLife.22264>.
- Desgrosellier JS, Cheresch DA. Integrins in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer*. 2010;10:9–22. <https://doi.org/10.1038/nrc2748>.
- Yang RS, Chiang HS, Tang CH, Yeh CS, Huang TF. Rhodostomin inhibits thrombin-enhanced adhesion of ROS 17/2.8 cells through the blockade of  $\alpha v \beta 3$  integrin. *Toxicol*. 2005;46:387–93. <https://doi.org/10.1016/j.toxicol.2005.05.002>.
- Dutour A, Jossierand V, Jury D, Guillemet S, Decouvelaere AV, Chotel F, Pointecouteau T, Rizo P, Coll JL, Blay JY. Targeted imaging of  $\alpha v \beta 3$  expressing sarcoma tumor cells in vivo in pre-operative setting using near infrared: a potential tool to reduce incomplete surgical resection. *Bone*. 2014;62:71–8. <https://doi.org/10.1016/j.bone.2014.02.004>.
- Zhang Y, Li L, Zhu J, Kuang H, Dong S, Wang H, Zhang X, Zhou Y. In vitro observations of self-assembled ECM-mimetic bioceramic nanoreservoir delivering rFN/CDH to modulate osteogenesis. *Biomaterials*. 2012;33:7468–77. <https://doi.org/10.1016/j.biomaterials.2012.06.095>.
- Koistinen P, Pulli T, Uitto VJ, Nissinen L, Hyypia T, Heino J. Depletion of  $\alpha v$  integrins from osteosarcoma cells by intracellular antibody expression induces bone differentiation marker genes and suppresses gelatinase (MMP-2) synthesis. *Matrix Biol*. 1999;18:239–51.
- Zhang J, Zhi H, Zhou C, Ding F, Luo A, Zhang X, Sun Y, Wang X, Wu M, Liu Z. Up-regulation of fibronectin in oesophageal squamous cell carcinoma is associated with activation of the Erk pathway. *J Pathol*. 2005;207:402–9. <https://doi.org/10.1002/path.1846>.
- Lessey BA, Damjanovich L, Coutifaris C, Castelbaum A, Albelda SM, Buck CA. Integrin adhesion molecules in the human endometrium. Correlation with the normal and abnormal menstrual cycle. *J Clin Invest*. 1992;90:188–95. <https://doi.org/10.1172/JCI115835>.
- Ta HT, Dass CR, Choong PF, Dunstan DE. Osteosarcoma treatment: state of the art. *Cancer Metastasis Rev*. 2009;28:247–63. <https://doi.org/10.1007/s10555-009-9186-7>.
- Na KY, Bacchini P, Bertoni F, Kim YW, Park YK. Syndecan-4 and fibronectin in osteosarcoma. *Pathology*. 2012;44:325–30. <https://doi.org/10.1097/PAT.0b013e328353447b>.
- Kilpadi KL, Sawyer AA, Prince CW, Chang PL, Bellis SL. Primary human marrow stromal cells and Saos-2 osteosarcoma cells use different

- mechanisms to adhere to hydroxylapatite. *J Biomed Mater Res A*. 2004;68:273–85. <https://doi.org/10.1002/jbma.20043>.
20. Stucci S, Tucci M, Passarelli A, Silvestris F. Avbeta3 integrin: pathogenetic role in osteotropic tumors. *Crit Rev Oncol Hematol*. 2015;96:183–93. <https://doi.org/10.1016/j.critrevonc.2015.05.018>.
  21. Duan X, Jia S-F, Zhou Z, Langley RR, Bolontrade MF, Kleinerman ES. Association of  $\alpha v \beta 3$  integrin expression with the metastatic potential and migratory and chemotactic ability of human osteosarcoma cells. *Clin Exp Metastasis*. 2005;21:747–53. <https://doi.org/10.1007/s10585-005-0599-6>.
  22. Yi W, Xiao E, Ding R, Luo P, Yang Y. High expression of fibronectin is associated with poor prognosis, cell proliferation and malignancy via the NF-kappaB/p53-apoptosis signaling pathway in colorectal cancer. *Oncol Rep*. 2016;36:3145–53. <https://doi.org/10.3892/or.2016.5177>.
  23. Liang CH, Chiu SY, Hsu IL, Wu YY, Tsai YT, Ke JY, Pan SH, Hsu YC, Li KC, Yang PC, et al. alpha-Catulin drives metastasis by activating ILK and driving an alphavbeta3 integrin signaling axis. *Cancer Res*. 2013;73:428–38. <https://doi.org/10.1158/0008-5472.CAN-12-2095>.
  24. Rhee SH, Han I, Lee MR, Cho HS, Oh JH, Kim HS. Role of integrin-linked kinase in osteosarcoma progression. *J Orthop Res*. 2013;31:1668–75. <https://doi.org/10.1002/jor.22409>.
  25. Mortara L, Orecchia P, Castellani P, Borsi L, Carnemolla B, Balza E. Schedule-dependent therapeutic efficacy of L19mTNF-alpha and melphalan combined with gemcitabine. *Cancer Med*. 2013;2:478–87. <https://doi.org/10.1002/cam4.89>.
  26. Robl B, Botter SM, Boro A, Meier D, Neri D, Fuchs B. Evaluation of F8-TNF-alpha in models of early and progressive metastatic osteosarcoma. *Transl Oncol*. 2017;10:419–30. <https://doi.org/10.1016/j.tranon.2017.02.005>.
  27. Gramoun A, Shorey S, Bashutski JD, Dixon SJ, Sims SM, Heersche JN, Manolson MF. Effects of Vitaxin, a novel therapeutic in trial for metastatic bone tumors, on osteoclast functions in vitro. *J Cell Biochem*. 2007;102:341–52. <https://doi.org/10.1002/jcb.21296>.

**Ready to submit your research? Choose BMC and benefit from:**

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

**At BMC, research is always in progress.**

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

