

CLINICAL AND TRANSLATIONAL RESEARCH IN *PNEUMOCYSTIS* AND *PNEUMOCYSTIS* PNEUMONIA¹

HUANG L.*

Summary:

Pneumocystis pneumonia (PcP) remains a significant cause of morbidity and mortality in immunocompromised persons, especially those with human immunodeficiency virus (HIV) infection.

Pneumocystis colonization is described increasingly in a wide range of immunocompromised and immunocompetent populations and associations between *Pneumocystis* colonization and significant pulmonary diseases such as chronic obstructive pulmonary disease (COPD) have emerged. This mini-review summarizes recent advances in our clinical understanding of *Pneumocystis* and PcP, describes ongoing areas of clinical and translational research, and offers recommendations for future clinical research from researchers participating in the "First centenary of the *Pneumocystis* discovery".

KEY WORDS: *Pneumocystis*, colonization, *Pneumocystis pneumonia* (PcP), human immunodeficiency virus (HIV), acquired immune deficiency syndrome (AIDS).

Résumé : LA RECHERCHE CLINIQUE SUR LE *PNEUMOCYSTIS* ET LA PNEUMOCYTOSE PULMONAIRE

La pneumocystose pulmonaire (PcP) est responsable d'une importante morbidité et mortalité chez les personnes immunodéprimées, particulièrement celles qui sont porteuses du virus de l'immunodéficience humaine (VIH). L'infection par le *Pneumocystis* s'observe de plus en plus, tant dans des populations immunodéprimées qu'immunocompétentes, et notamment chez des personnes atteintes de broncho-pneumopathies chroniques obstructives (BPCO). Cette mise au point recense les données récentes qui concernent la clinique de l'infection par le *Pneumocystis*, notamment la PcP, et les recommandations et les recherches en cours dans le domaine, telles qu'elles ont été présentées lors de la conférence commémorant le premier centenaire de la découverte de *Pneumocystis*.

MOTS-CLÉS : *Pneumocystis*, infection, pneumocystose pulmonaire (PcP), virus de l'immunodéficience humaine (VIH), syndrome de l'immunodéficience acquise (SIDA).

INTRODUCTION

The year 2009 marked the 100th anniversary of the first description of *Pneumocystis* by Carlos Chagas. Over the past 100 years, significant advances have been made in our understanding of both *Pneumocystis* and *Pneumocystis pneumonia* (PcP). The preeminence of PcP as a herald of the human immunodeficiency virus (HIV) / acquired immune deficiency syndrome (AIDS) epidemic and as a major cause of HIV-associated morbidity and mortality focused attention and resources on this previously uncommon opportunistic pneumonia. With the use of combination antiretroviral therapy to treat underlying HIV infection, the incidence of PcP has

declined, but an appreciation of *Pneumocystis* colonization in both immunocompromised and immunocompetent populations, and associations between *Pneumocystis* colonization and significant pulmonary diseases such as chronic obstructive pulmonary disease (COPD), have emerged. This mini-review summarizes recent advances in our clinical understanding of *Pneumocystis* and PcP, describes ongoing areas of clinical and translational research, and offers recommendations for future clinical research from researchers participating in the "First centenary of the *Pneumocystis* discovery" held in Brussels, Belgium on November 5-6, 2009.

EPIDEMIOLOGY OF PcP

PcP is a frequent AIDS-defining diagnosis. At its peak, PcP was an AIDS-defining diagnosis for greater than 20,000 new AIDS cases per year in the US (Centers for Diseases Control and Prevention, 1990-1993). Although the incidence of PcP has decreased in the current era of combination antiretroviral therapy, PcP remains a leading cause of AIDS in North American and European cohorts (Mocroft *et*

* Division of Pulmonary and Critical Care Medicine and HIV/AIDS Division, Department of Medicine, San Francisco General Hospital, University of California San Francisco.

Correspondence: Laurence Huang, M.D., HIV/AIDS Division, Ward 84, San Francisco General Hospital, 995 Potrero Avenue, San Francisco, CA 94110, USA.

Tel.: (415) 476-4082, extension 406 – Fax: (415) 476-6953.

E-mail: lhuang@php.ucsf.edu

¹ Stemming text to a communication at the "First centenary of the *Pneumocystis* discovery", Brussels, 5-6 November 2009.

al., 2009). In the multinational Antiretroviral Therapy Cohort Collaboration (ART-CC) established in 2000, PcP was the second most frequent AIDS-defining event after esophageal candidiasis (Mocroft *et al.*, 2009). Therefore, continued efforts to improve our understanding of both *Pneumocystis* and PcP are warranted (Table I).

PcP is an important cause of HIV-associated pneumonia but rates of PcP have decreased. At San Francisco General Hospital, the Division of Pulmonary and Critical Care Medicine has tracked confirmed cases of PcP – diagnosed by microscopic visualization of characteristic *Pneumocystis* cysts and/or trophic forms obtained from sputum induction or bronchoscopy – since 1990 (Fig. 1). At its peak in 1992, nearly 300 cases of HIV-associated PcP were diagnosed at this institution (Huang *et al.*, 1995). Today, this number has decreased to 20-30 cases per year. Most of these cases occurred in persons who were not receiving antiretroviral therapy or PcP prophylaxis and many were actually unaware of their HIV infection at the time of presentation (Fei *et al.*, 2009). This experience is similar at other institutions, where 23-31 % of reported PcP cases occurred in persons who were newly diagnosed with HIV infection at the time of PcP (Fei *et al.*, 2009a; Radhi *et al.*, 2008; Walzer *et al.*, 2008). Thus, increased efforts to test persons at risk of HIV, to engage HIV-infected persons in regular medical care, and to initiate and adhere to combination antiretroviral therapy and PcP prophylaxis are important strategies to decrease the incidence of the disease.

Comprehensive reviews document that HIV-associated PcP is reported throughout the world, in varying rates

(Davis *et al.*, 2007; Fisk *et al.*, 2003). However, data on the current rates of PcP in regions of the world that bear the greatest burden of HIV are limited. The scarcity of diagnostic and microbiologic tools to diagnose the disease is an important reason for the limited data on PcP rates. Thus, those institutions that possess these tools offer valuable windows into the epidemiology of PcP in low- and middle-income settings. At Mulago Hospital in Kampala, Uganda, the frequency of PcP among HIV-infected persons hospitalized with suspected pneumonia who have negative sputum acid fast bacilli (AFB) smears and undergo bronchoscopy has decreased from nearly 40 % of bronchoscopies to less than 10 % (Worodria *et al.*, 2003; Worodria *et al.*, 2010). Yet, the mortality associated with PcP remains high. Thus, efforts to improve both diagnostic and microbiologic capacity in low- and middle-income settings and to establish surveillance networks to track PcP cases are important clinical care and epidemiologic resources that should be developed.

Although the populations of non-HIV immunosuppressed are rising with the increased use of immunosuppressive or immunomodulating therapies to treat a wide spectrum of medical illnesses, data on the frequency of PcP among these populations are limited. As such, consensus on optimal diagnostic, therapeutic, and preventative strategies lag behind those for HIV-infected populations. Similar to recommendations for tracking PcP in low- and middle-income settings, collaborative networks of institutions that care for substantial numbers of these non-HIV individuals (similar to those established for HIV-infected persons) should be created.

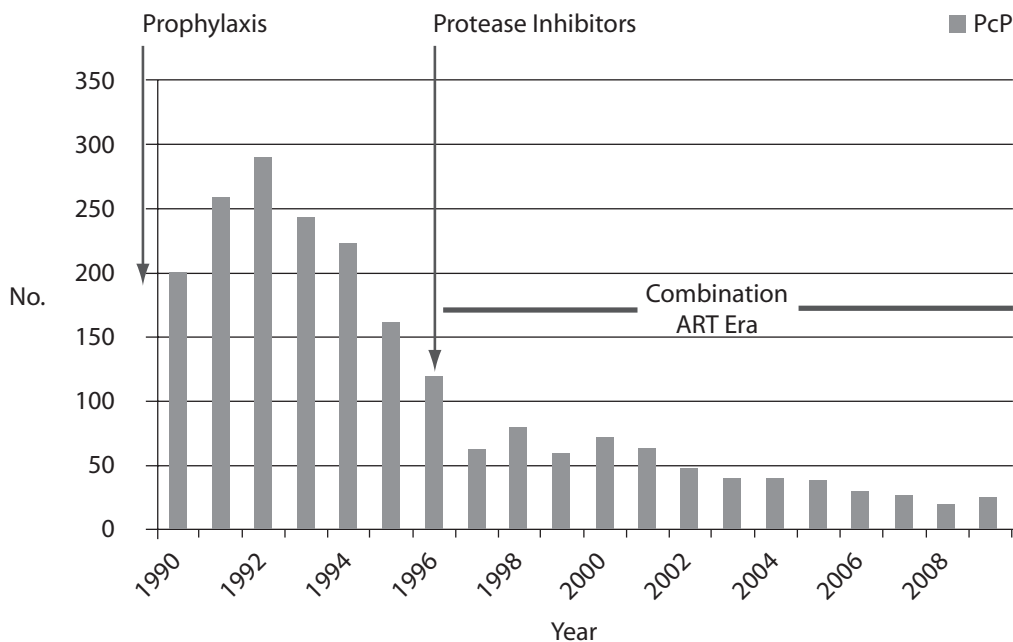


Fig. 1. – *Pneumocystis* pneumonia at San Francisco General Hospital.

Epidemiology

1. What is the current incidence of PcP in HIV-infected populations?
 - a. What is the incidence in high-income countries, including the US and Western Europe, where access to combination anti-retroviral therapy is generally widely available?
 - i. In these countries, which populations continue to develop PcP?
 - ii. Strategies to identify persons at risk for HIV and PcP need to be refined and preventative measures need to be implemented.
 - b. What is the incidence in low- and middle-income countries, where access to combination antiretroviral therapy is generally more limited?
 - i. In these countries, which populations develop PcP?
 - ii. Strategies to identify persons at risk for HIV and PcP need to be refined and preventative measures need to be implemented.
 - iii. Efforts to improve diagnostic and microbiologic capacity are needed.
 - iv. Surveillance networks to track PcP cases should be developed.
2. What is the current incidence of PcP in non-HIV, immunocompromised populations?
 - a. What is the incidence in populations immunocompromised from “traditional” immunosuppressive agents (*e.g.*, glucocorticoid medications) and disease therapies (*e.g.*, therapies for hematologic malignancy and cancer and after hematopoietic stem
 - b. What is the incidence in populations immunocompromised from “newer” biologic, immunomodulating agents (*e.g.*, tumor necrosis factor-alpha inhibitors)?
 - c. Surveillance networks to track PcP cases should be developed.

Diagnosis

3. What is the optimum approach to the diagnosis of PcP?
 - a. Bronchoscopy with bronchoalveolar lavage (BAL)?
 - b. Sputum induction (SI), followed by BAL, if SI is negative?
4. Is there a role for non-invasive tests in PcP diagnosis?
 - a. Oropharyngeal washing (OPW): Which specimen? Which assay? Which protocol?
 - b. Plasma s-adenosylmethionine?
 - c. (1-3)-beta-D-glucan?
 - d. Is the diagnostic accuracy different in non-HIV populations (lower *Pneumocystis* burden)?

Treatment

5. New drugs to treat and prevent PcP are needed.
6. What is the optimum second-line treatment for PcP?
 - a. Prospective, randomized clinical trials are needed.
7. Does TMP-SMX drug resistance exist?
 - a. What is (are) the mechanisms?
 - i. Dihydropteroate synthase (DHPS) gene mutations?
 - ii. Dihydrofolate reductase (DHFR) gene mutations?
 - iii. Sub-therapeutic trimethoprim or sulfamethoxazole drug levels?
 - iv. Host factors?

Outcome and Intensive care

8. What is the optimal timing for initiation of ART in HIV-associated PcP?
 - a. In Intensive care unit (ICU)?
 - b. Outside of the ICU?
9. What is the incidence of PcP-IRIS?
 - a. What are the risk factors?
 - b. Can PcP-IRIS be prevented (prophylaxis)?

Pneumocystis colonization

10. What are the implications of *Pneumocystis* colonization for the colonized individual?
 - a. COPD?
 - b. Other pulmonary diseases?
11. Can persons colonized with *Pneumocystis* transmit infection to others?
 - a. Risk factors for transmission – source, environment, contact.

Table I. – Conference Recommendations for future directions in clinical and translational research in *Pneumocystis* and *Pneumocystis* pneumonia.

PcP DIAGNOSIS

Classically, PcP presents with fevers, non-productive cough, and progressive shortness of breath. Chest radiograph demonstrates bilateral, symmetric reticular (interstitial) or granular opacities. Traditionally, bronchoscopy with bronchoalveolar lavage (BAL) is regarded as the gold standard procedure to diagnose PcP in HIV-infected persons with diagnostic sensitivity $\geq 98\%$ reported (Huang *et al.*, 1995). An early study reported a lower number of *Pneumocystis* organisms and a lower sensitivity of BAL for PcP in non-HIV immunocompromised persons compared to HIV-infected persons (Limper *et al.*, 1989). More recently, BAL combined with sensitive laboratory techniques (e.g., immunofluorescence testing, polymerase chain reaction, PCR) has been reported as sensitive to diagnose PcP in these non-HIV immunocompromised populations (Azoulay *et al.*, 2009). Since *Pneumocystis* DNA can be detected by PCR assay in the absence of clinical or microbiological pneumonia (Davis *et al.*, 2007), the increased sensitivity of PCR-based assays may be offset by a decreased specificity for PcP.

Pneumocystis cannot be cultured. Historically, the diagnosis of PcP has involved an invasive pulmonary procedure (i.e., bronchoscopy) to obtain specimens combined with a basic laboratory test (i.e., microscopic examination of stained respiratory specimens) to visualize the cysts and/or trophic forms. However, bronchoscopy requires specialized personnel, rooms, and equipment, and it is also expensive and carries an associated risk of complications. Thus, bronchoscopy is limited in its availability throughout many areas of the world that are burdened with HIV/AIDS. Laboratory advances (e.g., PCR), however, have revolutionized the diagnosis of many infectious diseases and PCR assays for *P. jirovecii* have been developed. These factors led researchers to examine whether the use of an advanced laboratory test (e.g., PCR) could be combined with a non-invasive pulmonary procedure to effectively diagnose PcP.

Oral or oropharyngeal (i.e., gargle) wash specimens combined with PCR assays have been examined as non-invasive tests to diagnose PcP. Two studies from San Francisco General Hospital that tested three different PCR-based assays found a diagnostic sensitivity of OPW up to 88% and a specificity up to 90% (de Oliveira *et al.*, 2007; Larsen *et al.*, 2004). Procedural factors such as collecting the OPW specimen within one day of PcP treatment initiation and having the patient cough vigorously prior to specimen collection may increase the sensitivity of the procedure. Studies are being conducted to validate these findings in both high- and low-income settings.

Blood-based assays have also been studied for diagnosis of PcP. A series of studies from New York indicate that plasma S-adenosylmethionine (SAM) levels could be used to distinguish between persons with PcP and those with non-PcP pneumonia and healthy controls (Skelly *et al.*, 2003; Skelly *et al.*, 2008). More recently, serum (1-3)-beta-D-glucan has shown promise as a test for PcP. Studies from 2009 reported that serum (1-3)-beta-D-glucan had a sensitivity of 100% and a specificity of 96.4% (using a cutoff of 100 pg/mL), that the assay may be more useful for PcP diagnosis than for monitoring response to treatment, that levels differ between patients with PcP and those who are PcP-negative but colonized with *Pneumocystis jirovecii*, and that the detection rate is lower in non-HIV PcP patients than in HIV-associated PcP patients (Desmet *et al.*, 2009; Nakamura *et al.*, 2009; Shimizu *et al.*, 2009; Watanabe *et al.*, 2009). Studies to determine which non-invasive test has the best performance characteristics in different immunocompromised populations at risk for PcP should be done.

PcP TREATMENT AND PREVENTION AND PUTATIVE TRIMETHOPRIM-SULFAMETHOXAZOLE DRUG RESISTANCE

Trimethoprim-sulfamethoxazole is the recommended first-line treatment for PcP in HIV-infected patients with mild, moderate, and severe PcP and also in non-HIV patients. Alternative regimens include intravenous pentamidine, clindamycin plus primaquine, trimethoprim plus dapsone, and atovaquone suspension. Adjunctive corticosteroids are recommended for patients with moderate to severe PcP as demonstrated by an arterial oxygen tension (PaO₂) less than 70 mm Hg or an alveolar-arterial oxygen gradient (A-a O₂) greater than 35 mm Hg. In HIV-infected patients, the recommended duration of treatment is 21 days while it is usually 14 days in non-HIV patients. However, a substantial proportion of individuals cannot complete a full course of trimethoprim-sulfamethoxazole due to treatment-limiting toxicity or are switched to an alternate treatment regimen due to perceived treatment failure (Fisk *et al.*, 2009). Although there are only limited data from prospective, randomized clinical trials comparing second-line PcP treatments, a tri-center observational study and a systematic review suggest that the combination of clindamycin plus primaquine is an effective alternative to intravenous pentamidine as second-line PcP treatment (Benfield *et al.*, 2008; Helweg-Larsen *et al.*, 2009). The development of new PcP treatment options and prospective, randomized controlled trials

comparing second-line PcP treatments are both important needs.

Trimethoprim-sulfamethoxazole is also the recommended first-line prevention for primary and secondary prophylaxis against PcP. However, the widespread use of this medication for PcP prophylaxis has been associated with increases in trimethoprim-sulfamethoxazole-resistant bacteria and has raised concerns over potential trimethoprim-sulfamethoxazole drug resistance in *P. jirovecii*. Trimethoprim-sulfamethoxazole drug resistance might also result in resistance to trimethoprim plus dapsone (a sulfone), thereby further limiting the therapeutic options available to treat PcP. The inability to culture *P. jirovecii* has hindered efforts to document drug-resistance in *Pneumocystis* but researchers have explored this question by examining genetic mutations within the dihydrofolate reductase (DHFR) and dihydropteroate synthase (DHPS) genes, the enzymatic targets of trimethoprim and sulfa (sulfamethoxazole and dapsone) medications, respectively (Huang *et al.*, 2004). In other micro-organisms, DHFR and DHPS mutations have been shown to cause drug resistance. Although DHFR mutations are infrequently found, non-synonymous DHPS mutations are found in up to 81 % of HIV-infected patients with PcP and the use of sulfa medications for PcP prophylaxis is strongly associated with the presence of these mutations (Crothers *et al.*, 2005; Huang *et al.*, 2000). DHPS mutations continue to be reported worldwide and there are geographic differences in the observed frequency of mutations (Alvarez-Martinez *et al.*, 2008; Beard *et al.*, 2000; Crothers *et al.*, 2005; Esteves *et al.*, 2008; Huang *et al.*, 2000; Magne *et al.*, 1989; Rabodonirina *et al.*, 2006; Wissmann *et al.*, 2006). Furthermore, the presence of DHPS mutations has been associated with increased mortality in one study and increased risk of trimethoprim-sulfamethoxazole PcP treatment failure in a second study, although other studies have failed to find these associations and instead have reported that risk factors such as low serum albumin and early ICU admission were stronger predictors of PcP mortality (Crothers *et al.*, 2005; Helweg-Larsen *et al.*, 1999; Kazanjian *et al.*, 2000).

At present, there exists a seeming paradox regarding the clinical significance of DHPS mutations. Studies consistently report that the majority of patients with PcP and DHPS mutations who are treated with trimethoprim-sulfamethoxazole respond to this treatment (Crothers *et al.*, 2005; Helweg-Larsen *et al.*, 1999; Kazanjian *et al.*, 2000; Navin *et al.*, 2001). However, patients with DHPS mutations who are treated with trimethoprim-sulfamethoxazole tend to have worse outcomes compared to those with wild-type DHPS who are treated with trimethoprim-sulfamethoxazole and those with DHPS mutations who are treated with

a non-sulfa-based regimen (Crothers *et al.*, 2005). The precise explanation for these observations remains a focus of current investigation.

PcP MORTALITY AND INTENSIVE CARE

Despite differences in geography and demographic characteristics, in-hospital mortality among HIV-infected patients with PcP is similar (ranging from 10.3 to 13.5 %) in different cohort studies from Los Angeles, London, and San Francisco (Fei *et al.*, 2009b; Radhi *et al.*, 2008; Walzer *et al.*, 2008). Each of these studies also reported on predictors of mortality. In the Los Angeles study of 262 HIV-infected patients diagnosed with PcP from January 2000 through December 2003, need for mechanical ventilation, development of a pneumothorax, and low serum albumin were independent predictors of increased mortality (Radhi *et al.*, 2008). In the London study of 494 consecutive HIV-infected patients with 547 episodes of laboratory-confirmed PcP from June 1985 through June 2006, increasing patient age, subsequent episode of PcP, low hemoglobin level, low partial pressure of oxygen breathing room air, presence of medical comorbidity, and pulmonary Kaposi sarcoma were independent predictors associated with increased mortality (Walzer *et al.*, 2008). Mortality was comparable during the periods from June 1985 through December 1989, January 1990 through June 1996, and July 1996 through June 2006 ($p = 0.14$). Finally, in the San Francisco study of 451 consecutive HIV-infected patients diagnosed with 524 episodes of microscopically-confirmed PcP from January 1997 through December 2006, increasing patient age, recent injection drug use, increased total bilirubin, decreased serum albumin, and increased alveolar-arterial oxygen gradient were independent predictors of increased mortality (Fei *et al.*, 2009b). Using these five predictors, a six-point PcP mortality prediction rule was derived that stratified patients according to increasing risk of mortality: score 0-1, 4 % mortality; score 2-3, 12 % mortality; and score 4-5, 48 % mortality. Studies are being conducted to validate these single-institution findings in other cohorts.

A few institutions have tracked PcP in HIV-infected patients requiring critical care in the Intensive Care Unit (ICU). Among these institutions, a study from San Francisco General Hospital in the current combination antiretroviral therapy era reported that critically ill HIV-infected patients with PcP who received combination antiretroviral therapy had a significantly lower mortality compared to patients who did not receive antiretroviral therapy (Morris *et al.*, 2003). The use of combination antiretroviral therapy was an independent predictor

that was associated with lower mortality (odds ratio, OR = 0.14; 95 % confidence interval, CI = 0.02-0.84; $p = 0.03$). A study from London, however, failed to find a mortality difference associated with combination antiretroviral therapy (Miller *et al.*, 2006). In this study, mortality improved from 71 % before mid-1996 to 34 % after mid-1996 ($p = 0.008$). This improvement in mortality was ascribed to general improvements in care as no patients were started on combination antiretroviral therapy. In the absence of data from randomized clinical trials, the pros and cons of initiating antiretroviral therapy in critically ill HIV-infected ICU patients have been debated. Recently, the National Institutes of Health (NIH)-funded AIDS Clinical Trials Group (ACTG) published its results of a prospective, multicenter randomized clinical trial of HIV-infected non-ICU inpatients hospitalized with an acute opportunistic infection (Zolopa *et al.*, 2009). Overall, 282 subjects were enrolled and 63 % had PcP. The study found no difference in their primary endpoint. However, subjects randomized to the early antiretroviral therapy arm had fewer AIDS progressions/deaths (OR = 0.51, 95 % CI = 0.27-0.94, $p = 0.035$) and a longer time to AIDS progression/death (stratified Hazard Ratio, HR = 0.53, 95 % CI = 0.30-0.92). Importantly, there was no increase in adverse events in subjects randomized to early antiretroviral therapy in this study but cases of severe immune reconstitution inflammatory syndrome (IRIS, also called immune reconstitution syndrome, IRS, and immune reconstitution disease, IRD) resulting in respiratory failure and requiring invasive mechanical ventilation have been reported and serve as a cautionary note (Jagannathan *et al.*, 2009). Whether the ACTG results from hospitalized but non-critically ill patients can be extrapolated into the ICU, where patients are often receiving mechanical ventilation and have renal and/or hepatic insufficiency, is an important but largely unanswered question (Huang *et al.*, 2006).

PNEUMOCYSTIS COLONIZATION AND POTENTIAL PERSON-TO-PERSON TRANSMISSION

The presence of *Pneumocystis* organisms or *P. jirovecii* DNA detected in the absence of PcP has been termed *Pneumocystis* colonization (also called carriage and sub-clinical infection). *Pneumocystis* colonization has been increasingly reported and documented to occur in infants and children, pregnant women, immunocompetent adults with underlying pulmonary disease, non-HIV immunocompromised individuals, and HIV-infected persons (Morris *et*

al., 2008). For example, in one study of 58 infants < 1 year of age who died, *Pneumocystis* colonization was detected in 100 % (Beard *et al.*, 2005). HIV-infected inpatients hospitalized with non-PcP pneumonia also appear to have a high prevalence of *Pneumocystis* colonization, with one study reporting a prevalence of 68 % (Davis *et al.*, 2008).

Colonization with *Pneumocystis* has also been associated with airways obstruction and chronic obstructive pulmonary disease (COPD) and possibly with other pulmonary diseases as well (Vidal *et al.*, 2006). In one study, *Pneumocystis* colonization was detected in 36.7 % of HIV-negative patients with very severe COPD (Global Health Initiative on Obstructive Lung Disease [GOLD] Stage IV) compared with 5.3 % of smokers with normal lung function or less severe COPD (GOLD Stages 0, I, II, and III) ($p = 0.004$) and with 9.1 % of control subjects ($p = 0.007$) (Morris *et al.*, 2004). *Pneumocystis* colonized subjects exhibited more severe airway obstruction (median FEV1 = 21 % predicted vs 62 % predicted in non-colonized subjects, $p = 0.006$). In a second study, patients colonized with *Pneumocystis* had higher proinflammatory cytokine levels than did those patients without evidence of *Pneumocystis* colonization (Calderon *et al.*, 2007). Finally, HIV-infected outpatients who were colonized with *Pneumocystis* had worse airway obstruction and higher sputum matrix metalloprotease-12 levels, suggesting that *Pneumocystis* colonization may be important in HIV-associated COPD (Morris *et al.*, 2009). Studies to further evaluate the role of *Pneumocystis* colonization in the development and the progression of HIV-associated COPD are ongoing.

Although the clinical significance of *Pneumocystis* colonization remains to be elucidated completely, important insights can be gained from studies in laboratory animals. An animal model has been developed to study *Pneumocystis* colonization in immunocompetent and simian immunodeficiency virus (SIV)-infected cynomolgus macaques and the potential role of *Pneumocystis* colonization on COPD (Kling *et al.*, 2009). Animal studies have also been used to study disease transmission. These studies demonstrate that animals are a reservoir for the specific *Pneumocystis* species that affects them, that animals carrying *Pneumocystis* develop PcP after being immunosuppressed (reactivation), and that immunocompromised *Pneumocystis*-free animals develop PcP after exposure to not only immunocompromised animals infected with *Pneumocystis* but also immunocompetent animals that are colonized with *Pneumocystis*. Animal studies also demonstrate that animal-to-animal transmission occurs and occurs via an airborne route.

Person-to-person transmission of *Pneumocystis* via an airborne (aerosol) route has been suggested. However,

a precise understanding of the factors involved in disease transmission remains unclear and there are no universal recommendations for the respiratory isolation of persons with active PCP. Even if such recommendations became standard practice, disease transmission could hypothetically occur via persons who are colonized with *Pneumocystis* as is the case in animal studies. Molecular epidemiology and serology studies to examine potential person-to-person transmission are ongoing.

CONCLUSIONS

Over the past 100 years, significant advances have been made in our understanding of both *Pneumocystis* and *Pneumocystis* pneumonia (PCP). This mini-review described recent advances in our understanding of *Pneumocystis* and PCP, ongoing areas of clinical and translational research, and offered recommendations for future clinical research from researchers participating in the First centenary commemorative Conference on the discovery of *Pneumocystis*. The attendees of the conference sincerely hope that the next 100 years bring continued advances in our understanding of the organism and the disease.

ACKNOWLEDGEMENTS

Dr. Huang is supported by grants from the National Institutes of Health: NIH K24 HL087713, NIH R01 HL090335, NIH R01 HL090335-02S109, and NIH U01 HL098964.

REFERENCES

- ALVAREZ-MARTINEZ M.J., MORENO A., MIRO J.M., VALLS M.E., RIVAS P.V., DE LAZZARI E., SUED O., BENITO N., DOMINGO P., RIBERA E., SANTIN M., SIRERA G., SEGURA F., VIDAL F., RODRIGUEZ F., RIERA M., CORDERO M.E., ARRIBAS J.R., JIMENEZ DE ANTA M.T., GATELL J.M., WILSON P.E. & MESHNIK S.R. *Pneumocystis jirovecii* pneumonia in Spanish HIV-infected patients in the combined antiretroviral therapy era: prevalence of dihydropteroate synthase mutations and prognostic factors of mortality. *Diagn Microbiol Infect Dis*, 2008, 62, 34-43.
- AZOULAY E., BERGERON A., CHEVRET S., BELE N., SCHLEMMER B. & MENOTTI J. Polymerase chain reaction for diagnosing *Pneumocystis* pneumonia in non-HIV immunocompromised patients with pulmonary infiltrates. *Chest*, 2009, 135, 655-661.
- BEARD C.B., CARTER J.L., KEELY S.P., HUANG L., PIENIAZEK N.J., MOURA I.N., ROBERTS J.M., HIGHTOWER A.W., BENS M.S., FREEMAN A.R., LEE S., STRINGER J.R., DUCHIN J.S., DEL RIO C., RIMLAND D., BAUGHMAN R.P., LEVY D.A., DIETZ V.J., SIMON P. & NAVIN T.R. Genetic variation in *Pneumocystis carinii* isolates from different geographic regions: implications for transmission. *Emerg Infect Dis*, 2000, 6, 265-272.
- BEARD C.B., FOX M.R., LAWRENCE G.G., GUARNER J., HANZLICK R.L., HUANG L., DEL RIO C., RIMLAND D., DUCHIN J.S. & COLLEY D.G. Genetic differences in *Pneumocystis* isolates recovered from immunocompetent infants and from adults with AIDS: epidemiological implications. *J Infect Dis*, 2005, 192, 1815-1818.
- BENFIELD T., ATZORI C., MILLER R.F. & HELWEG-LARSEN J. Second-line salvage treatment of AIDS-associated *Pneumocystis jirovecii* pneumonia: a case series and systematic review. *J Acquir Immune Defic Syndr*, 2008, 48, 63-67.
- CALDERON E.J., RIVERO L., RESPALDIZA N., MORILLA R., MONTES-CANO M.A., FRIAZA V., MUNOZ-LOBATO F., VARELA J.M., MEDRANO F.J. & DE LA HORRA C. Systemic inflammation in patients with chronic obstructive pulmonary disease who are colonized with *Pneumocystis jirovecii*. *Clin Infect Dis*, 2007, 45, e17-19.
- CENTERS FOR DISEASE CONTROL AND PREVENTION. HIV/AIDS Surveillance Report. 1990.
- CENTERS FOR DISEASE CONTROL AND PREVENTION. HIV/AIDS Surveillance Report. 1991.
- CENTERS FOR DISEASE CONTROL AND PREVENTION. HIV/AIDS Surveillance Report. 1992.
- CENTERS FOR DISEASE CONTROL AND PREVENTION. HIV/AIDS Surveillance Report. 1993.
- CROTHERS K., BEARD C.B., TURNER J., GRONER G., FOX M., MORRIS A., EISER S. & HUANG L. Severity and outcome of HIV-associated *Pneumocystis* pneumonia containing *Pneumocystis jirovecii* dihydropteroate synthase gene mutations. *AIDS*, 2005, 19, 801-805.
- DAVIS J.L. & HUANG L. *Pneumocystis* pneumonia, in: Global HIV/AIDS medicine. Volberding P.A., Sande M.A., Lange J., Greene W.C. (eds), First ed., Saunders Elsevier, Inc., Philadelphia, 2007, 309-321.
- DAVIS J.L., WELSH D.A., BEARD C.B., JONES J.L., LAWRENCE G.G., FOX M.R., CROTHERS K., MORRIS A., CHARBONNET D., SWARTZMAN A. & HUANG L. *Pneumocystis* colonisation is common among hospitalised HIV infected patients with non-*Pneumocystis* pneumonia. *Thorax*, 2008, 63, 329-334.
- DE OLIVEIRA A., UNNASCH T.R., CROTHERS K., EISER S., ZUCCHI P., MOIR J., BEARD C.B., LAWRENCE G.G. & HUANG L. Performance of a molecular viability assay for the diagnosis of *Pneumocystis* pneumonia in HIV-infected patients. *Diagn Microbiol Infect Dis*, 2007, 57, 169-176.
- DESMET S., VAN WIJNGAERDEN E., MAERTENS J., VERHAEGEN J., VERBEKEN E., DE MUNTER P., MEERSSEMAN W., VAN MEENSEL B., VAN ELDERE J. & LAGROU K. Serum (1-3)-beta-D-glucan as a tool for diagnosis of *Pneumocystis jirovecii* pneumonia in patients with human immunodeficiency virus infection or hematological malignancy. *J Clin Microbiol*, 2009, 47, 3871-3874.
- ESTEVEZ F., MONTES-CANO M.A., DE LA HORRA C., COSTA M.C., CALDERON E.J., ANTUNES F. & MATOS O. *Pneumocystis jirovecii* multilocus genotyping profiles in patients from Portugal and Spain. *Clin Microbiol Infect*, 2008, 14, 356-362.

- FEI M.W., SANT C.A., KIM E.J., SWARTZMAN A., DAVIS J.L., JARLSBERG L.G. & HUANG L. Severity and outcomes of *Pneumocystis* pneumonia in patients newly diagnosed with HIV infection: an observational cohort study. *Scand J Infect Dis*, 2009a, 41, 672-678.
- FEI M.W., KIM E.J., SANT C.A., JARLSBERG L.G., DAVIS J.L., SWARTZMAN A. & HUANG L. Predicting mortality from HIV-associated *Pneumocystis* pneumonia at illness presentation: an observational cohort study. *Thorax*, 2009b, 64, 1070-1076.
- FISK D.T., MESHNICK S. & KAZANJIAN P.H. *Pneumocystis carinii* pneumonia in patients in the developing world who have acquired immunodeficiency syndrome. *Clin Infect Dis*, 2003, 36, 70-78.
- FISK M., SAGE E.K., EDWARDS S.G., CARTLEDGE J.D. & MILLER R.F. Outcome from treatment of *Pneumocystis jirovecii* pneumonia with co-trimoxazole. *Int J STD AIDS*, 2009, 20, 652-653.
- HELWEG-LARSEN J., BENFIELD T., ATZORI C. & MILLER R.F. Clinical efficacy of first- and second-line treatments for HIV-associated *Pneumocystis jirovecii* pneumonia: a tri-centre cohort study. *J Antimicrob Chemother*, 2009, 64, 1282-1290.
- HELWEG-LARSEN J., BENFIELD T.L., EUGEN-OLSEN J., LUNDGREN J.D. & LUNDGREN B. Effects of mutations in *Pneumocystis carinii* dihydropteroate synthase gene on outcome of AIDS-associated *P. carinii* pneumonia. *Lancet*, 1999, 354, 1347-1351.
- HUANG L., HECHT F.M., STANSELL J.D., MONTANTI R., HADLEY W.K. & HOPEWELL P.C. Suspected *Pneumocystis carinii* pneumonia with a negative induced sputum examination. Is early bronchoscopy useful? *Am J Respir Crit Care Med*, 1995, 151, 1866-1871.
- HUANG L., BEARD C.B., CREASMAN J., LEVY D., DUCHIN J.S., LEE S., PIENIAZEK N., CARTER J.L., DEL RIO C., RIMLAND D. & NAVIN T.R. Sulfa or sulfone prophylaxis and geographic region predict mutations in the *Pneumocystis carinii* dihydropteroate synthase gene. *J Infect Dis*, 2000, 182, 1192-1198.
- HUANG L., CROTHERS K., ATZORI C., BENFIELD T., MILLER R., RABODONIRINA M. & HELWEG-LARSEN J. Dihydropteroate synthase gene mutations in *Pneumocystis* and sulfa resistance. *Emerg Infect Dis*, 2004, 10, 1721-1728.
- HUANG L., QUARTIN A., JONES D. & HAVLIR D.V. Intensive care of patients with HIV infection. *N Engl J Med*, 2006, 355, 173-181.
- JAGANNATHAN P., DAVIS E., JACOBSON M. & HUANG L. Life-threatening immune reconstitution inflammatory syndrome after *Pneumocystis* pneumonia: a cautionary case series. *AIDS*, 2009, 23, 1794-1796.
- KAZANJIAN P., ARMSTRONG W., HOSSLER P.A., BURMAN W., RICHARDSON J., LEE C.H., CRANE L., KATZ J. & MESHNICK S.R. *Pneumocystis carinii* mutations are associated with duration of sulfa or sulfone prophylaxis exposure in AIDS patients. *J Infect Dis*, 2000, 182, 551-557.
- KLING H.M., SHIPLEY T.W., PATIL S., MORRIS A. & NORRIS K.A. *Pneumocystis* colonization in immunocompetent and simian immunodeficiency virus-infected cynomolgus macaques. *J Infect Dis*, 2009, 199, 89-96.
- LARSEN H.H., HUANG L., KOVACS J.A., CROTHERS K., SILCOTT V.A., MORRIS A., TURNER J.R., BEARD C.B., MASUR H. & FISCHER S.H. A prospective, blinded study of quantitative touch-down polymerase chain reaction using oral-wash samples for diagnosis of *Pneumocystis* pneumonia in HIV-infected patients. *J Infect Dis*, 2004, 189, 1679-1683.
- LIMPER A.H., OFFORD K.P., SMITH T.F. & MARTIN W.J. 2ND. *Pneumocystis carinii* pneumonia. Differences in lung parasite number and inflammation in patients with and without AIDS. *Am Rev Respir Dis*, 1989, 140, 1204-1209.
- MAGNE D., LACUBE P., ANGOULVANT A., MELIANI L., BOTTEREL F., BOUGNOUX M.E., CHOCHILLON C., CORNET M., DANNAOUI E., DATRY A., DUNAND J., GALEAZZI G., BOUGES-MICHEL C., YERA H., SARFATI C. & ROUX P. *Pneumocystosis*: survey and DHPS genotype analysis in 14 Parisian hospitals in 2003 and 2004. *J Eukaryot Microbiol*, 2006, 53 (Suppl 1), S106-107.
- MILLER R.F., ALLEN E., COPAS A., SINGER M. & EDWARDS S.G. Improved survival for HIV infected patients with severe *Pneumocystis jirovecii* pneumonia is independent of highly active antiretroviral therapy. *Thorax*, 2006, 61, 716-721.
- MOCROFT A., STERNE J.A., EGGER M., MAY M., GRABAR S., FURRER H., SABIN C., FATKENHEUER G., JUSTICE A., REISS P., D'ARMINIO MONFORTE A., GILL J., HOGG R., BONNET F., KITAHATA M., STASZEWSKI S., CASABONA J., HARRIS R. & SAAG M. Variable impact on mortality of AIDS-defining events diagnosed during combination antiretroviral therapy: not all AIDS-defining conditions are created equal. *Clin Infect Dis*, 2009, 48, 1138-1151.
- MORRIS A., WACHTER R.M., LUCE J., TURNER J. & HUANG L. Improved survival with highly active antiretroviral therapy in HIV-infected patients with severe *Pneumocystis carinii* pneumonia. *AIDS*, 2003, 17, 73-80.
- MORRIS A., SCIURBA F.C., LEBEDEVA I.P., GITHAIGA A., ELLIOTT W.M., HOGG J.C., HUANG L. & NORRIS K.A. Association of chronic obstructive pulmonary disease severity and *Pneumocystis* colonization. *Am J Respir Crit Care Med*, 2004, 170, 408-413.
- MORRIS A., WEI K., AFSHAR K. & HUANG L. Epidemiology and clinical significance of *Pneumocystis* colonization. *J Infect Dis*, 2008, 197, 10-17.
- MORRIS A., ALEXANDER T., RADHI S., LUCHT L., SCIURBA F.C., STEELE C. & NORRIS K.A. Airway obstruction is increased in *Pneumocystis*-colonized HIV-infected outpatients. *J Clin Microbiol*, 2009, 47, 3773-3776.
- NAKAMURA H., TATEYAMA M., TASATO D., HARANAGA S., YARA S., HIGA F., OHTSUKI Y. & FUJITA J. Clinical utility of serum beta-D-glucan and KL-6 levels in *Pneumocystis jirovecii* pneumonia. *Intern Med*, 2009, 48, 195-202.
- NAVIN T.R., BEARD C.B., HUANG L., DEL RIO C., LEE S., PIENIAZEK N.J., CARTER J.L., LE T., HIGHTOWER A. & RIMLAND D. Effect of mutations in *Pneumocystis carinii* dihydropteroate synthase gene on outcome of *P. carinii* pneumonia in patients with HIV-1: a prospective study. *Lancet*, 2001, 358, 545-549.
- RABODONIRINA M., NAHIMANA A., WEBER R., FRANCIOLI P., BILLE J. & HAUSER P.M. Geographical variation in the prevalence of *Pneumocystis jirovecii* dihydropteroate synthase muta-

- tions within Western Europe. *J Eukaryot Microbiol*, 2006, 53 (Suppl 1), S112-113.
- RADHI S., ALEXANDER T., UKWU M., SALEH S. & MORRIS A. Outcome of HIV-associated *Pneumocystis* pneumonia in hospitalized patients from 2000 through 2003. *BMC Infect Dis*, 2008, 8, 118.
- SHIMIZU Y., SUNAGA N., DOBASHI K., FUEKI M., FUEKI N., MAKINO S. & MORI M. Serum markers in interstitial pneumonia with and without *Pneumocystis jirovecii* colonization: a prospective study. *BMC Infect Dis*, 2009, 9, 47.
- SKELLY M., HOFFMAN J., FABBRI M., HOLZMAN R.S., CLARKSON A.B. JR. & MERALI S. S-adenosylmethionine concentrations in diagnosis of *Pneumocystis carinii* pneumonia. *Lancet*, 2003, 361, 1267-1268.
- SKELLY M.J., HOLZMAN R.S. & MERALI S. S-adenosylmethionine levels in the diagnosis of *Pneumocystis carinii* pneumonia in patients with HIV infection. *Clin Infect Dis*, 2008, 46, 467-471.
- VIDAL S., DE LA HORRA C., MARTIN J., MONTES-CANO M.A., RODRIGUEZ E., RESPALDIZA N., RODRIGUEZ F., VARELA J.M., MEDRANO F.J. & CALDERON E.J. *Pneumocystis jirovecii* colonisation in patients with interstitial lung disease. *Clin Microbiol Infect*, 2006, 12, 231-235.
- WALZER P.D., EVANS H.E., COPAS A.J., EDWARDS S.G., GRANT A.D. & MILLER R.F. Early predictors of mortality from *Pneumocystis jirovecii* pneumonia in HIV-infected patients: 1985-2006. *Clin Infect Dis*, 2008, 46, 625-633.
- WATANABE T., YASUOKA A., TANUMA J., YAZAKI H., HONDA H., TSUKADA K., HONDA M., GATANAGA H., TERUYA K., KIKUCHI Y. & OKA S. Serum (1-3)-beta-D-glucan as a noninvasive adjunct marker for the diagnosis of *Pneumocystis* pneumonia in patients with AIDS. *Clin Infect Dis*, 2009, 49, 1128-1131.
- WISSMANN G., ALVAREZ-MARTINEZ M.J., MESHNICK S.R., DIHEL A.R. & PROLLA J.C. Absence of dihydropteroate synthase mutations in *Pneumocystis jirovecii* from Brazilian AIDS patients. *J Eukaryot Microbiol*, 2006, 53, 305-307.
- WORODRIA W., DAVIS J.L., CATTAMANCHI A., ANDAMA A., DEN BOON S., YOO S.D., HOPEWELL P.C. & HUANG L. Bronchoscopy is useful for diagnosing smear-negative tuberculosis in HIV-infected patients. *Eur Respir J*, 2010, 36, 446-448.
- WORODRIA W., OKOT-NWANG M., YOO S.D. & AISU T. Causes of lower respiratory infection in HIV-infected Ugandan adults who are sputum AFB smear-negative. *Int J Tuberc Lung Dis*, 2003, 7, 117-123.
- ZOLOPA A., ANDERSEN J., POWDERLY W., SANCHEZ A., SANNE I., SUCKOW C., HOGG E. & KOMAROW L. Early antiretroviral therapy reduces AIDS progression/death in individuals with acute opportunistic infections: a multicenter randomized strategy trial. *PLoS One*, 2009, 4, e5575.

Received on August 24th, 2010
Accepted on October 11th, 2010