





## Draft Genome Sequence of a Polyhydroxyalkanoate-Producing Bacillus cereus Strain Isolated from Nuevo Leon State, Mexico

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**ABSTRACT** Polyhydroxyalkanoates (PHA) are microbially produced biopolymers which are biodegradable and biocompatible. These compounds produced by microorganisms have been described as a potent alternative to synthetic plastics, which are often recalcitrant. Here, we report the draft genome sequence of a PHA-producing *Bacillus cereus* isolated in our laboratory.

acillus species are known to produce a wide variety of beneficial metabolites, among which are polyhydroxyalkanoates (PHA). We isolated a novel strain of *Bacillus cereus* from garden soil in San Nicolas de los Garza, Nuevo Leon, Mexico (25°43′38″N, 100°18′37″W), via serial dilution to a factor of 8. This dilution was plated onto a nutrient agar plate. The colonies which appeared on this plate were used for genomic DNA extraction for the identification of the colony. We observed that these colonies yielded a large amount of PHA under stress conditions (1, 2). Thus, we decided to sequence and analyze the genetic basis on which it produces biopolymers.

*B. cereus* 4N was grown to the logarithmic phase in lysogeny broth at 28°C; a portion of this culture was used for the extraction of genomic DNA. Genomic DNA was extracted using the Wizard genomic DNA extraction kit (Promega, USA) according to the manufacturer's instructions. To identify the bacterium, we amplified the 16S rRNA region of the *Bacillus cereus* genome using the primer set 8F (5'-AGAGTTTGATCCTGGCTCAG-3') and 1492R (5'-ACGGCTACCTTGTTACGACTT-3') as described by Mizuno et al. (3). The identity of *B. cereus* 4N was confirmed by a BLAST search analysis on the NCBI website, which yielded an identity of 99.88% with *Bacillus cereus* strain LL-1.

The genomic DNA was sequenced at the Laboratorio Nacional de Nutrigenómica y Microbiómica Digestiva Animal-IPN or National Laboratory of Nutrigenomics and Animal Digestive Microbiomics-IPN using 500 ng of the submitted genomic DNA. The DNA was quantified using the Qubit double-stranded DNA (dsDNA) high-sensitivity (HS) assay kit on the Qubit fluorometer (Thermo Fisher Scientific, MA, USA). The library preparation was conducted with the Nextera Flex library kit and individual indices for bar coding, using the Illumina reference (10000000254) as a guide. The library was sequenced using Illumina MiniSeq technology.

A total of 12,710,962 paired-end 145-bp reads were obtained, with a sequencing depth of >300X, after a quality check using FastQC v0.11.9 (4). Default parameters were used for all software unless otherwise stated. The raw reads were trimmed using Trimmomatic v0.32 to remove adaptors and improve the sequence quality (5). The trimmed reads were assembled using SPAdes v3.15.3 on the KBase online platform (6, 7). The obtained contigs from the assembled reads were reduced to 27 contigs using the Medusa combo online

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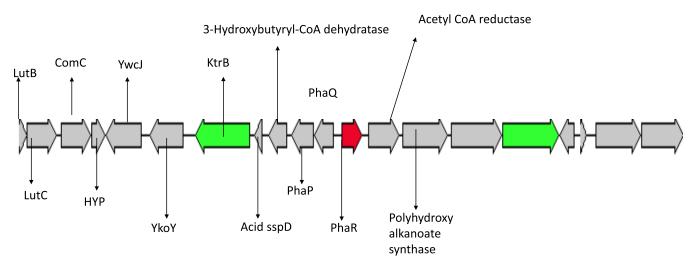


FIG 1 Image for the operon in Bacillus cereus 4N for the gene associated with the production of polyhydroxyalkanoate.

genome scaffolder (8) with a single-contig *Bacillus cereus* isolate as a reference. The reduced contigs were annotated using Prokka v1.14.5 (9), and the publicly available genome was annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (10). The genome comprised 6,213,840 bp in 27 contigs with an  $N_{50}$  value of 5,994,343 bp and a GC content of 34.69%. The genome contains 6,084 genes and 6,388 coding sequences (CDS), with 267 amino acids, 6,084 protein-coding sequences, and 67 RNAs (7 55, 1 165, and 1 235 rRNAs). *B. cereus* 4N has 53 tRNAs and 1 CRISPR array.

Five genes that are essential for the metabolism of PHA were found in an operon using the RAST annotation tool, as shown in Fig. 1. These genes include *PhaP*, *PhaR*, *PhaQ*, and polyhydroxyalkanoate synthase and 3-hydroxyl butyrate-coenzyme A (CoA) hydratase genes.

**Data availability.** The draft genome sequence of *B. cereus* 4N was deposited at GenBank under accession number JALBCM010000000; the project data are available under BioProject accession number PRJNA815817 and BioSample accession number SAMN26643023. The raw draft genome data were deposited in the Sequence Read Archive (SRA) under SRA accession number SRR18331562. The 16S rRNA sequence has been deposited at the NCBI website under accession number MH404097.1.

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## **REFERENCES**

- Martínez-Herrera RE, Alemán-Huerta ME, Almaguer-Cantú V, Rosas-Flores W, Martínez-Gómez VJ, Quintero-Zapata I, Rivera G, Rutiaga-Quiñones OM. 2020. Efficient recovery of thermostable polyhydroxybutyrate (PHB) by a rapid and solvent-free extraction protocol assisted by ultrasound. Int J Biol Macromol 164:771–782. https://doi.org/10.1016/j.ijbiomac .2020.07.101.
- Martínez-Herrera RE, Alemán-Huerta ME, Flores-Rodríguez P, Almaguer-Cantú V, Valencia-Vázquez R, Rosas-Flores W, Medrano-Roldán H, Ochoa-Martínez LA, Rutiaga-Quiñones OM. 2021. Utilization of Agave durangensis leaves by Bacillus cereus 4N for polyhydroxybutyrate (PHB) biosynthesis. Int J Biol Macromol 175:199–208. https://doi.org/10.1016/j.ijbiomac.2021 .01.167.
- Mizuno K, Ohta A, Hyakutake M, Ichinomiya Y, Tsuge T. 2010. Isolation of polyhydroxyalkanoate-producing bacteria from a polluted soil and characterization of the isolated strain Bacillus cereus YB-4. Polym Degrad Stab 95:1335–1339. https://doi.org/10.1016/j.polymdegradstab .2010.01.033.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. http://www.bioinformatics.babraham.ac.uk/projects/ fastqc/.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- 6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin

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- VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19: 455–477. https://doi.org/10.1089/cmb.2012.0021.
- Allen BH, Gupta N, Edirisinghe JN, Faria JP, Henry CS. 2022. Application of the metabolic modeling pipeline in KBase to categorize reactions, predict essential genes, and predict pathways in an isolate genome. Methods Mol Biol 2349:291–320. https://doi.org/10.1007/978-1-0716-1585-0\_13.
- 8. Bosi E, Donati B, Galardini M, Brunetti S, Sagot MF, Lió P, Crescenzi P, Fani R, Fondi M. 2015. MeDuSa: a multi-draft based scaffolder. Bioinformatics 31:2443–2451. https://doi.org/10.1093/bioinformatics/btv171.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/nar/gkw569.

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