



# Generation and Characterization of Site-Specifically Mono-Ubiquitylated p53

Alexandra Julier,<sup>[a]</sup> Vanessa Radtke,<sup>[a]</sup> Andreas Marx,<sup>\*,[a]</sup> and Martin Scheffner<sup>\*,[a]</sup>

The tumor suppressor p53 is regulated by various posttranslational modifications including different types of ubiquitylation, which exert distinct effects on p53. While modification by ubiquitin chains targets p53 for degradation, attachment of single ubiquitin moieties (mono-ubiquitylation) affects the intracellular location of p53 and/or its interaction with chromatin. However, how this is achieved at the molecular level remains largely unknown. Similarly, since p53 can be ubiquitylated at different lysine residues, it remains unclear if the

eventual effect depends on the position of the lysine modified. Here, we combined genetic code expansion with oxime ligation to generate p53 site-specifically mono-ubiquitylated at position 120. We found that mono-ubiquitylation at this position neither interferes with p53 ubiquitylation by the E3 ligases HDM2 and E6AP in complex with the viral E6 oncoprotein nor affects p53 binding to a cognate DNA sequence. Thus, ubiquitylation per se does not affect physiologically relevant properties of p53.

The growth-suppressive properties of the tumor suppressor p53 are negatively affected in the majority of human cancers by different means including mutations in the TP53 gene<sup>[1]</sup> or overexpression of the proto-oncoprotein HDM2, a member of the RING family of E3 ubiquitin ligases.<sup>[2]</sup> p53 is best known for its ability to act as a sequence-specific transcriptional modulator that upon various stress stimuli, is activated and in consequence modulates the expression of various genes.<sup>[3]</sup> Thereby, p53 triggers different cellular responses including cell cycle arrest, apoptosis, senescence, alteration of metabolic pathways, and autophagy.<sup>[3]</sup> To control p53 function properly under normal growth conditions and upon stress, p53 is subject to various posttranslational modifications (PTMs) including phosphorylation, acetylation, and ubiquitylation, to name but a few.<sup>[4]</sup> Ubiquitylation is a highly complex and versatile PTM that ranges from the attachment of a single ubiquitin (Ub) moiety to a substrate protein (mono-ubiquitylation) to the attachment of a sheer indefinite number of differently linked Ub chains (poly-ubiquitylation).<sup>[5]</sup> p53 is subject to both, mono- and poly-ubiquitylation. HDM2-mediated poly-ubiquitylation, for instance, targets p53 for degradation,<sup>[6]</sup> while mono-ubiquitylation of p53 has been associated with nuclear-cytoplasmic


shuttling,<sup>[7]</sup> mitochondrial translocation,<sup>[8]</sup> and chromatin association.<sup>[9]</sup> At first glance, it may seem surprising that mono-ubiquitylation has different effects on p53. However, p53 can be ubiquitylated at different lysine residues and, thus, the eventual outcome likely depends on at which lysine p53 is mono-ubiquitylated.


A prerequisite to prove the general hypothesis that the effect of mono-ubiquitylation on the biochemical/biological properties of p53 depends on the actual site of modification is the availability of homogeneous populations of site-specifically ubiquitylated p53.<sup>[9]</sup> The generation of defined Ub-protein conjugates by enzymatic means is rather challenging, since most, if not all, E3 ligases are promiscuous insofar as they frequently ubiquitylate substrate proteins at several lysine residues and mediate both mono- and poly-ubiquitylation. Therefore, a number of chemical biological approaches to generate defined Ub-protein conjugates have been developed, such as native chemical ligation,<sup>[10]</sup> thiol-based ligation,<sup>[10b,11]</sup> Cu(I)-catalyzed alkyne-azide cycloaddition (CuAAC),<sup>[12]</sup> oxime ligation,<sup>[13]</sup> and sortylation.<sup>[14]</sup> However, none of these methods has been applied to p53 so far.

We previously reported on the generation of defined Ub conjugates including Ub dimers and site-specifically mono-ubiquitylated forms of PCNA and the linker histone H1.2 by combining CuAAC with the genetic code expansion technology to incorporate unnatural amino acids at distinct positions of a protein of interest.<sup>[12a,f,i]</sup> To test whether this approach can also be applied to p53, we used the amber codon suppression (ACS) method to generate a p53 variant containing the pyrrolysine analog Plk at a distinct position and conjugated it via CuAAC to a Ub variant harboring azidohomoalanine at its C terminus. This resulted in reasonable amounts of p53-Ub conjugates (Figure S1 in the Supporting Information). We then subjected p53-Ub to ubiquitylation assays in the presence of HDM2 or the E3 ligase E6AP in complex with E6 oncoprotein of human papillomavirus type 16 to determine whether ubiquitylation at the desired position alters the property of p53 to serve as substrate for the

[a] Dr. A. Julier, Dr. V. Radtke, Prof. Dr. A. Marx, Prof. Dr. M. Scheffner  
Departments of Biology and Chemistry  
Konstanz Research School Chemical Biology  
University of Konstanz  
Universitätsstr. 10, 78457 Konstanz (Germany)  
E-mail: andreas.marx@uni-konstanz.de  
martin.scheffner@uni-konstanz.de

 Supporting information for this article is available on the WWW under <https://doi.org/10.1002/cbic.202100659>

 This article is part of a joint Special Collection with the Journal of Peptide Science on SPP 1623: Chemoselective reactions for the synthesis and application of functional proteins. Please see our homepage for more articles in the collection.

 © 2022 The Authors. ChemBioChem published by Wiley-VCH GmbH. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

two E3 ligases.<sup>[15]</sup> However, respective control reactions revealed that the reaction conditions of CuAAC, i.e. the presence of copper, drive p53 into a conformation that is neither recognized by HDM2 nor by the E6-E6AP complex (note that the structural requirements of p53 to serve as substrate for HDM2 and for the E6-E6AP complex are clearly different)<sup>[16]</sup> (Figure S1).

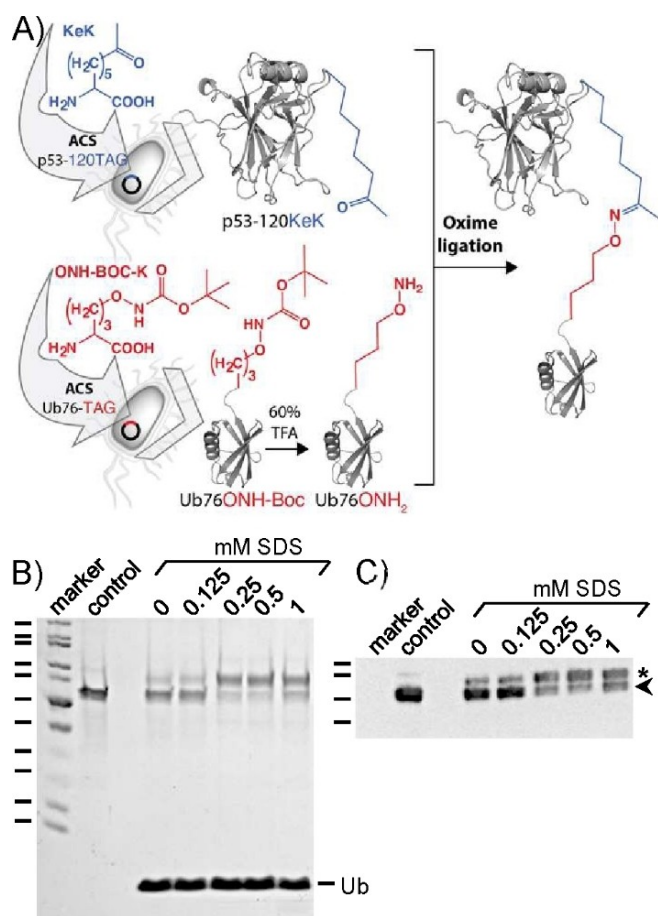
p53 is a zinc-binding protein, and since copper does not interfere with ubiquitylation reactions per se (Figure S1C), we assumed that copper somehow affects the structural integrity of p53 as previously shown for other zinc-binding proteins.<sup>[17]</sup> We therefore switched to oxime ligation, a bioorthogonal conjugation strategy that does not require metal ions and can be performed under non-denaturing conditions.<sup>[18]</sup> In this reaction, an aminoxy group reacts with a carbonyl group to form an oxime linkage (Figure 1A). Furthermore, oxime ligation

was used before to produce linkage-specific Ub conjugates,<sup>[13]</sup> as the oxime linkage is a reasonable mimic of the natural isopeptide bond (Figure S2). To equip p53 and Ub with the required functional groups at the desired positions, we used ACS again.<sup>[19]</sup> In brief (for details, see Supporting Information), the lysine codon at position 120 of p53 was replaced by the amber stop codon enabling the introduction of a carbonyl group at this position via site-specific incorporation of ketolysine (KeK).<sup>[20]</sup> We chose this position, as K120 contributes to the sequence-specific DNA binding capacity of p53<sup>[21]</sup> and is known to be ubiquitylated,<sup>[15]</sup> but whether K120 ubiquitylation affects DNA binding or other properties of p53 is unknown. *E. coli* cells harboring the respective expression plasmid and the required Pyrrolysine (Pyl)-tRNA-synthetase/tRNAPyl pair were cultured in presence of 10 mM KeK<sup>[22]</sup> resulting in the expression of p53-120KeK (Figure S3). Upon purification, the identity of p53-120KeK was verified by LC-MS/MS (Figure S6A).

To equip Ub with a C-terminal aminoxy group (Ub76ONH<sub>2</sub>) by ACS, the C-terminal glycine codon at position 76 was replaced by the amber stop codon. As unnatural amino acid, we chose the lysine derivative N $\epsilon$ -aminoxy-L-lysine, the aminoxy group of which had to be protected by an N $\epsilon$ -latent protection group to prevent unwanted reactions with aldehyde or keto groups of endogenous bacterial molecules.<sup>[13a]</sup> As N $\epsilon$ -latent protection group, we chose the acid-labile *tert*-butoxycarbonyl (Boc) group that also served as recognition motif for the respective Pyl-tRNA-synthetase/tRNAPyl pair for incorporation of N $\epsilon$ -aminoxy-(Boc)-L-lysine (U1).<sup>[13a]</sup> Upon expression in *E. coli* and subsequent purification (Figure S4), Ub76ONH-Boc was deprotected by using 60% TFA, and the identity of Ub76ONH<sub>2</sub> was verified by LC-MS (Figure S4C). This also showed that incorporation of U1 did not proceed to completion (Figure S4C), resulting in a mixture of truncated Ub (Ub75) and full-length Ub (Ub76ONH-Boc) (Figure S4C). Although the truncated form cannot be separated from full-length Ub, this is not a concern since it cannot react with p53-120KeK. Thus, upon oxime ligation, Ub75 can easily be removed from the p53-Ub conjugate by anion exchange chromatography (for details, see Supporting Information).

To ensure that the functionality of p53 in the context of the envisioned p53-Ub conjugate is affected solely by the presence of Ub and not by other factors, the oxime ligation reaction should be performed under near-physiological conditions. However, oxime reactions proceed rather slowly at neutral pH.<sup>[10]</sup> Since we previously showed that anionic surfactants enhance CuAAC bioconjugation reactions,<sup>[12f]</sup> we tested whether addition of non-denaturing concentrations of SDS also enhances oxime ligation. Indeed, increasing amounts of SDS resulted in increased yields of p53-120-Ub (Figure 1; Figure S4D, E) of up to 70% of the p53 input. That the ligation product indeed represents p53-120-Ub was verified by LC-MS/MS analysis (Figure S6B).

Upon further purification (Figure S5), we investigated the structural integrity of p53-120-Ub. Due to the fact that E6 recognizes p53 only when it is in a wild-type (wt)-like conformation,<sup>[23]</sup> we determined whether the incorporation of KeK or the covalent attachment of Ub to position 120 affects

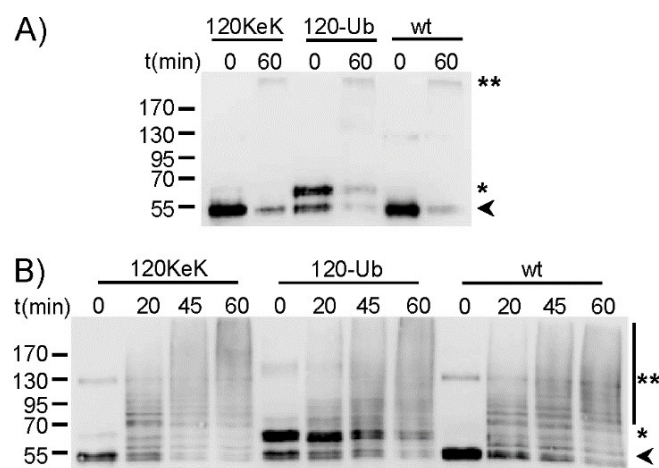


**Figure 1.** Generation of mono-ubiquitylated p53 via oxime ligation. A) Schematic of the approach. For details, see text. B), C) Oxime ligation is enhanced with increasing amounts of non-denaturing SDS concentrations. p53-120KeK (20 pmol) was mixed with a 50-molar excess of Ub76ONH<sub>2</sub> in presence of increasing concentrations of SDS as indicated and subjected to two freeze-thaw cycles<sup>[18a]</sup> or, alternatively, to incubation for 20 h at 25 °C (Figure S4E). Reaction products were subjected to SDS-PAGE followed by Coomassie blue staining (B) or by Western blot analysis using an anti-p53 antibody (C). Control, reaction in the absence of Ub76ONH<sub>2</sub>. Running positions of p53-120KeK and p53-120-Ub are indicated by an arrow and an asterisk, respectively. Running positions of molecular mass markers (kDa) are indicated from top to bottom in B): 120,100, 85, 70, 60, 50, 40, 30, 25, 20, 15, and in C): 70,60,50,40.

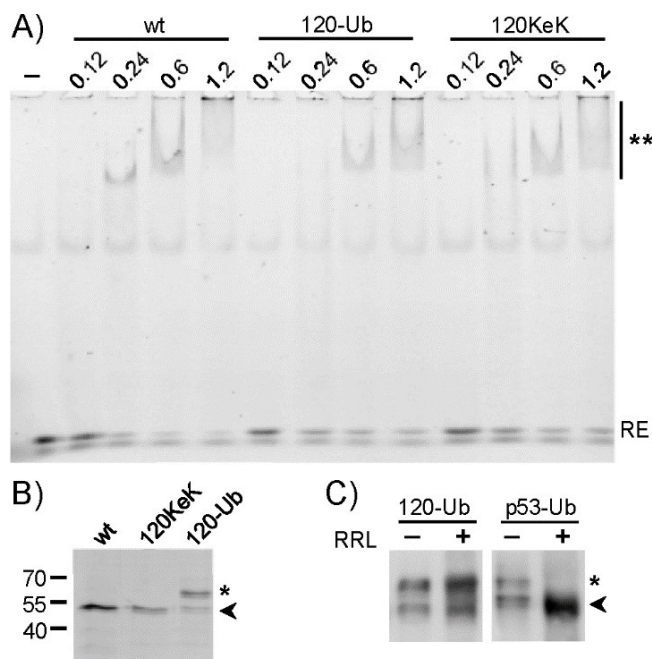
the ability of p53 to serve as a substrate in *in vitro* ubiquitylation assays using the E6-E6AP complex as E3 ubiquitin ligase (Figure 2A). Independent of the modification state, the p53 variants were recognized and ubiquitylated by the E6-E6AP complex to a similar extent. Similarly, like wt p53, p53-120KeK and p53-120-Ub were also ubiquitylated by HDM2 (Figure 2B). These data demonstrate that in contrast to CuAAC, the reaction conditions of oxime ligation do not affect the structural integrity of p53 and that mono-ubiquitylation at position 120 does not affect the property of p53 to be recognized by the E3 ligases tested.

Acetylation of p53 at K120 was reported to affect the sequence-specific DNA binding properties of p53.<sup>[21]</sup> Therefore, we assessed the ability of p53-120KeK and p53-120-Ub to bind to the p21 response element (p21-RE) in electromobility shift assays (EMSA) in comparison to wt p53. Yet, no significant difference in the binding ability of the different p53 variants was observed (Figure 3A, Figure S7). While this further supports the notion that the reaction conditions of oxime ligation and the incorporation of KeK do not detectably affect the structural integrity of p53, it may seem surprising that attachment of a rather large molecule such as Ub to K120 does not interfere with DNA binding. However, it was recently reported that upon binding of p53 to so-called high affinity RE such as p21-RE, the side chain of K120 is disordered and not involved in hydrogen bonding with nucleobases.<sup>[24]</sup> This provides a reasonable explanation for our observation that p53-120-Ub can still bind to the p21-RE and indicates that p53-120-Ub adopts a wt p53-like conformation.

Mono-ubiquitylation affects the intracellular localization of p53,<sup>[7-8]</sup> which is likely due to changes in the protein-protein interaction properties of mono-ubiquitylated p53. Since cells or



**Figure 2.** Ubiquitylation assays with p53 variants. Equal amounts of p53-120-Ub, p53-120KeK, and wild-type (wt) p53 were incubated with A) E6-E6AP or B) HDM2 under *in vitro* ubiquitylation conditions for the times indicated. Reaction products were analyzed by SDS-PAGE followed by Western blot analysis with an antibody directed against p53. Running positions of the non-modified form of wt p53 and p53-120KeK, p53-120-Ub, and (poly-)ubiquitylated forms are indicated by an arrow, asterisk, and double asterisk, respectively. Running positions of molecular mass standards (kDa) are indicated on the left.



**Figure 3.** A) Characterization of the DNA binding ability of p53 variants by EMSA. Increasing concentrations of p53-120-Ub, p53-120KeK, and wild-type (wt) p53 were incubated with 10 nM fluorescein-labeled p21 response element (RE) and analyzed on a 4% native TBE polyacrylamide gel followed by fluorescence read out at 473 nm. -, control reaction in the absence of p53; RE, running position of the unbound RE; \*\*, running position of the RE-p53 complex. B) Loading control for the proteins used in A). Running positions of non-modified p53/p53-120KeK and p53-120-Ub are indicated by an arrow and an asterisk, respectively. Running positions of molecular mass standards (kDa) are indicated on the left. C) The oxime-linkage of p53-120-Ub is not hydrolyzed. 150 ng oxime-linked p53-120-Ub or enzymatically mono-ubiquitylated p53 were incubated in the absence or presence of 5  $\mu$ L rabbit reticulocyte lysate (RRL) for 30 min at 37 °C. RRL served as a source of de-ubiquitylating enzymes. Reaction products were analyzed by SDS-PAGE followed by Western blot analysis with a p53-specific antibody. The running position of p53-120-Ub/enzymatically mono-ubiquitylated p53 and the non-modified form of p53 is denoted with an asterisk and an arrow, respectively.

cell lysates are rich in de-ubiquitylating enzymes (DUBs), Ub conjugates that are resistant to the action of DUBs would be a prerequisite for such interactome analyses. To test the stability of oxime-linked p53-120-Ub, we incubated p53-120-Ub with rabbit reticulocyte lysate for 30 min at 37 °C (Figure 3C). As previously shown for oxime-linked Ub-Ub conjugates<sup>[13a]</sup> and in contrast to enzymatically ubiquitylated p53, which served as control, the oxime-linked p53-120-Ub was not hydrolyzed. Finally, we showed that the oxime ligation strategy is not limited to the generation of mono-ubiquitylated forms of p53 but can also be applied to other proteins such as the linker histone H1.2 (Figure S8).

In conclusion, the combination of the genetic code expansion technology with oxime ligation enables the generation of mono-ubiquitylated forms of p53 and other proteins in a site-specific manner. Importantly, the reaction conditions of oxime ligation do not interfere per se with the structural integrity of p53 allowing to analyze known properties of p53 that may be affected by mono-ubiquitylation. Furthermore, oxime-linked p53-Ub conjugates are resistant to the action of



DUBs. Thus, they are well suited for affinity enrichment experiments to identify proteins that interact selectively with mono-ubiquitylated forms of p53 thereby modulating the growth-suppressive properties of p53.

## Acknowledgements

We thank Andreas Marquardt and Anna Sladewska-Marquardt of the Proteomics Center of the University of Konstanz for assistance with mass spectrometric experiments. We gratefully acknowledge funding by the Deutsche Forschungsgemeinschaft (SFB 969, project B3; SPP 1623) and financial support by the Konstanz Research School Chemical Biology. Open Access funding enabled and organized by Projekt DEAL.

## Conflict of Interest

The authors declare no conflict of interest.

## Data Availability Statement

The data that support the findings of this study are available in the supplementary material of this article.

**Keywords:** bioconjugation · oxime ligation · protein modifications · tumor suppressor p53 · ubiquitylation

- [1] V. J. Bykov, N. Issaeva, G. Selivanova, K. G. Wiman, *Carcinogenesis* **2002**, *23*, 2011–2018.
- [2] D. P. Xirodimas, M. Scheffner, *Subcell. Biochem.* **2010**, *54*, 116–135.
- [3] O. Laptchenko, C. Prives, *Cell Death Differ.* **2006**, *13*, 951–961.
- [4] Y. Liu, O. Tavana, W. Gu, *J. Mol. Cell Biol.* **2019**, *11*, 564–577.
- [5] a) A. Hershko, A. Ciechanover, *Annu. Rev. Biochem.* **1998**, *67*, 425–479; b) D. Komander, M. Rape, *Annu. Rev. Biochem.* **2012**, *81*, 203–229.
- [6] M. S. Rodriguez, J. M. Desterro, S. Lain, D. P. Lane, R. T. Hay, *Mol. Cell Biol.* **2000**, *20*, 8458–8467.
- [7] a) M. A. Lohrum, D. B. Woods, R. L. Ludwig, E. Balint, K. H. Vousden, *Mol. Cell Biol.* **2001**, *21*, 8521–8532; b) L. Nie, M. Sasaki, C. G. Maki, *J. Biol. Chem.* **2007**, *282*, 14616–14625.
- [8] N. D. Marchenko, S. Wolff, S. Erster, K. Becker, U. M. Moll, *EMBO J.* **2007**, *26*, 923–934.
- [9] L. Le Cam, L. K. Linares, C. Paul, E. Julien, M. Lacroix, E. Hatchi, R. Triboulet, G. Bossis, A. Shmueli, M. S. Rodriguez, O. Coux, C. Sardet, *Cell* **2006**, *127*, 775–788.
- [10] a) C. P. Hackenberger, D. Schwarzer, *Angew. Chem. Int. Ed.* **2008**, *47*, 10030–10074; *Angew. Chem.* **2008**, *120*, 10182–10228; b) P. E. Dawson, T. W. Muir, I. Clark-Lewis, S. B. Kent, *Science* **1994**, *266*, 776–779.
- [11] a) L. Yin, B. Krantz, N. S. Russell, S. Deshpande, K. D. Wilkinson, *Biochemistry* **2000**, *39*, 10001–10010; b) R. E. Morgan, V. Chudasama, P. Moody, M. E. Smith, S. Caddick, *Org. Biomol. Chem.* **2015**, *13*, 4165–4168; c) F. Meier, T. Abeywardana, A. Dhall, N. P. Marotta, J. Varkey, R. Langen, C. Chatterjee, M. R. Pratt, *J. Am. Chem. Soc.* **2012**, *134*, 5468–5471; d) L. Long, M. Furgason, T. Yao, *Methods* **2014**, *70*, 134–138; e) C. Chatterjee, R. K. McGinty, B. Fierz, T. W. Muir, *Nat. Chem. Biol.* **2010**, *6*, 267–269; f) T. Abeywardana, Y. H. Lin, R. Rott, S. Engelender, M. R. Pratt, *Chem. Biol.* **2013**, *20*, 1207–1213.
- [12] a) S. Eger, B. Castrec, U. Hübscher, M. Scheffner, M. Rubini, A. Marx, *ChemBioChem* **2011**, *12*, 2807–2812; b) S. Eger, M. Scheffner, A. Marx, M. Rubini, *Methods Mol. Biol.* **2012**, *832*, 589–596; c) N. D. Weikart, S. Sommer, H. D. Mootz, *Chem. Commun.* **2012**, *48*, 296–298; d) D. Schneider, T. Schneider, D. Rösner, M. Scheffner, A. Marx, *Bioorg. Med. Chem.* **2013**, *21*, 3430–3435; e) D. Rösner, T. Schneider, D. Schneider, M. Scheffner, A. Marx, *Nat. Protoc.* **2015**, *10*, 1594–1611; f) D. Schneider, T. Schneider, J. Aschenbrenner, F. Mortensen, M. Scheffner, A. Marx, *Bioorg. Med. Chem.* **2016**, *24*, 995–1001; g) X. Zhao, J. Lutz, E. Höllmüller, M. Scheffner, A. Marx, F. Stengel, *Angew. Chem. Int. Ed.* **2017**, *56*, 15764–15768; *Angew. Chem.* **2017**, *129*, 15972–15976; h) V. Hagmann, S. Sommer, P. Fabian, J. Bierlmeier, N. van Treel, H. D. Mootz, D. Schwarzer, J. E. Azevedo, G. Dodt, *Sci. Rep.* **2018**, *8*, 16014; i) E. Höllmüller, S. Geiggas, M. L. Niedermeier, K. M. Kammer, S. M. Kienle, D. Rosner, M. Scheffner, A. Marx, F. Stengel, *Nat. Commun.* **2021**, *12*, 3497.
- [13] a) M. Stanley, S. Virdee, *ChemBioChem* **2016**, *17*, 1472–1480; b) A. Shanmugham, A. Fish, M. P. Luna-Vargas, A. C. Faesen, F. El Oualid, T. K. Sixma, H. Ovaas, *J. Am. Chem. Soc.* **2010**, *132*, 8834–8835; c) S. K. Singh, I. Sahu, S. M. Mali, H. P. Hemantha, O. Kleinfeld, M. H. Glickman, A. Brik, *J. Am. Chem. Soc.* **2016**, *138*, 16004–16015.
- [14] M. Fottner, A. D. Brunner, V. Bittl, D. Horn-Ghetko, A. Jussupow, V. R. I. Kaila, A. Bremm, K. Lang, *Nat. Chem. Biol.* **2019**, *15*, 276–284.
- [15] W. M. Chan, M. C. Mak, T. K. Fung, A. Lau, W. Y. Siu, R. Y. Poon, *Mol. Cancer Res.* **2006**, *4*, 15–25.
- [16] E. A. Medcalf, J. Milner, *Oncogene* **1993**, *8*, 2847–2851.
- [17] Y. Kim, S. H. Kim, D. Ferracane, J. A. Katzenellenbogen, C. M. Schroeder, *Bioconjugate Chem.* **2012**, *23*, 1891–1901.
- [18] a) S. M. Agten, D. P. Suylen, T. M. Hackeng, *Bioconjugate Chem.* **2016**, *27*, 42–46; b) S. M. Agten, P. E. Dawson, T. M. Hackeng, *J. Pept. Sci.* **2016**, *22*, 271–279; c) D. K. Kolmel, E. T. Kool, *Chem. Rev.* **2017**, *117*, 10358–10376; d) S. Wang, G. N. Nawale, S. Kadekar, O. P. Oommen, N. K. Jena, S. Chakraborty, J. Hilborn, O. P. Varghese, *Sci. Rep.* **2018**, *8*, 2193.
- [19] J. W. Chin, *Annu. Rev. Biochem.* **2014**, *83*, 379–408.
- [20] Y. Huang, W. Wan, W. K. Russell, P. J. Pai, Z. Wang, D. H. Russell, W. Liu, *Bioorg. Med. Chem. Lett.* **2010**, *20*, 878–880.
- [21] a) R. Vainer, S. Cohen, A. Shahar, R. Zarivach, E. Arbely, *J. Mol. Biol.* **2016**, *428*, 3013–3025; b) E. Arbely, E. Natan, T. Brandt, M. D. Allen, D. B. Veprincev, C. V. Robinson, J. W. Chin, A. C. Joerger, A. R. Fersht, *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 8251–8256.
- [22] H. Neumann, S. Y. Peak-Chew, J. W. Chin, *Nat. Chem. Biol.* **2008**, *4*, 232–234.
- [23] M. Scheffner, T. Takahashi, J. M. Huibregtse, J. D. Minna, P. M. Howley, *J. Virol.* **1992**, *66*, 5100–5105.
- [24] M. Farkas, H. Hashimoto, Y. Bi, R. V. Davuluri, L. Resnick-Silverman, J. J. Manfredi, E. W. Debler, S. B. McMahon, *Nat. Commun.* **2021**, *12*, 484.

Manuscript received: December 1, 2021

Revised manuscript received: January 12, 2022

Accepted manuscript online: January 13, 2022

Version of record online: February 3, 2022