

Research Article

Metabolism and Metabolic Inhibition of Xanthotoxol in Human Liver Microsomes

Zhongnv Ma,¹ Xianbao Shi,² Gang Zhang,³ Feng Guo,¹ Lina Shan,² and Jiqun Cai¹

¹Department of Pharmaceutical Toxicology, School of Pharmacy, China Medical University, Shenyang 110001, China

²The First Affiliated Hospital of Liaoning Medical University, Jinzhou 121001, China

³Department of Medicinal Chemistry, Virginia Commonwealth University, Richmond, VA 23219, USA

Correspondence should be addressed to Jiqun Cai; caijiqun123@163.com

Received 23 November 2015; Revised 8 February 2016; Accepted 9 February 2016

Academic Editor: Karen Nieber

Copyright © 2016 Zhongnv Ma et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Cytochrome p450 (CYP450) enzymes are predominantly involved in Phase I metabolism of xenobiotics. In this study, the CYP450 isoforms involved in xanthotoxol metabolism were identified using recombinant CYP450s. In addition, the inhibitory effects of xanthotoxol on eight CYP450 isoforms and its pharmacokinetic parameters were determined using human liver microsomes. CYP1A2, one of CYP450s, played a key role in the metabolism of xanthotoxol compared to other CYP450s. Xanthotoxol showed stronger inhibition on CYP3A4 and CYP1A2 compared to other isoenzymes with the IC_{50} of 7.43 μ M for CYP3A4 and 27.82 μ M for CYP1A2. The values of inhibition kinetic parameters (K_i) were 21.15 μ M and 2.22 μ M for CYP1A2 and CYP3A4, respectively. The metabolism of xanthotoxol obeyed the typical monophasic Michaelis-Menten kinetics and V_{max} , K_m , and CL_{int} values were calculated as 0.55 $\text{nmol}\cdot\text{min}^{-1}\cdot\text{mg}^{-1}$, 8.46 μ M, and 0.06 $\text{mL}\cdot\text{min}^{-1}\cdot\text{mg}^{-1}$. In addition, the results of molecular docking showed that xanthotoxol was bound to CYP1A2 with hydrophobic and π - π bond and CYP3A4 with hydrogen and hydrophobic bond. We predicted the hepatic clearance (CL_H) and the CL_H value was 15.91 $\text{mL}\cdot\text{min}^{-1}\cdot\text{kg}^{-1}$ body weight. These data were significant for the application of xanthotoxol and xanthotoxol-containing herbs.

1. Introduction

Xanthotoxol (Figure 1), a biologically active linear furcoumarin, occurs in a large number of plants and is mainly extracted from the fruit of *Fructus Cnidii* [1]. Xanthotoxol shows strong pharmacological activities as anti-inflammatory, antioxidant, 5-HT antagonistic, and neuroprotective effects [2–4]. Respecting so many pharmacological activities of xanthotoxol, xanthotoxol is likely used with other drugs, and the possibility of herb-induced toxicity should be paid more attention. Among all of the metabolic processes, metabolisms catalyzed by the cytochrome P450 (CYP450) enzymes are the most important because almost 70%–80% of the known Phase I metabolisms are attributed to them [5]. Some of herbal medicines may result in CYP-mediated herb-drug interactions (HDIs) with prescribed other drugs [6]. For example, lovastatin was a substrate of cytochrome P450 3A4 (CYP3A4), and clarithromycin was an inhibitor of CYP3A4. When clarithromycin and lovastatin

were coadministered, clarithromycin inhibited the activity of CYP3A4 and increased the serum concentrations of lovastatin and subsequent elevated the risk of myopathy [7]. In recent years, using humanized enzyme to study the drug-drug interactions in vitro has avoided the species differences of enzyme's isoforms, expression, and activities [8, 9]. The US FDA has already confirmed the validity of in vitro enzymes to assess the in vivo interaction between medications [10, 11].

In this study, we examined the inhibitory potential of xanthotoxol on CYP450s and kinetic parameters using in vitro human liver microsomes (HLMs), which will provide the basis for further in vivo studies in future.

2. Materials and Methods

2.1. Materials. Xanthotoxol (purity > 98%) was purchased from Sichuan Weikeqi Biotechnology Co. Ltd. (Sichuan, China). Paclitaxel, 1-aminobenzotriazole (ABT), phenacetin, sulfaphenazole, chlorzoxazone, quinidine, clomethiazole,

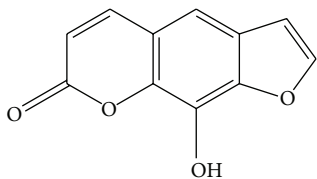


FIGURE 1: The structure of xanthotoxol.

furafylline, 8-methoxypsoralen, coumarin, diclofenac, quercetin, dextromethorphan, ketoconazole, testosterone, S-mephenytoin, omeprazole, glucose-6-phosphate dehydrogenase, NADP⁺, and D-glucose-6-phosphate were obtained from Sigma-Aldrich (St. Louis, MO, USA). All other reagents were the highest purity commercially available or HPLC grade.

2.2. Preparation and Characterization of Liver Microsomes. Liver microsomes from human (HLMs) used in this study were provided by the Research Institute for Liver Disease Co. (Shanghai, China). The HLMs were prepared from eleven individual human donor livers. Protein concentration and microsomes activities of CYP2C19, CYP2A6, CYP2C8, CYP2D6, CYP1A2, CYP2C9, CYP2E1, and CYP3A4 had been previously characterized by the Research Institute for Liver Disease Co.

2.3. CYP450 Probe Substrate Assays. HLMs phenacetin o-deethylation, coumarin 7-hydroxylation, paclitaxel 6 α -hydroxylation, diclofenac 4'-hydroxylation, dextromethorphan o-demethylation, chlorzoxazone 6-hydroxylation, testosterone 6 β -hydroxylation, and S-mephenytoin 4'-hydroxylation activities were utilized as selective markers for CYP1A2, CYP2A6, CYP2C8, CYP2C9, CYP2D6, CYP2E1, CYP3A4, and CYP2C19.

2.4. Incubation System for Liver Microsomes or Recombinant CYP450 Supersomes. The optimal conditions for microsomal incubation were described previously [12]. The incubation mixture, with a total volume of 200 μ L, consisted of 100 mM potassium phosphate buffer (pH 7.4), an NADPH-generating system (10 mM glucose 6-phosphate, 1 unit/mL of glucose 6-phosphate dehydrogenase, and 4 mM MgCl₂), liver microsomes (0.3 mg/mL), or recombinant CYP450s (15 nM). In all experiments, xanthotoxol was serially diluted to the required concentrations and the final methanol concentration did not exceed 1% (v/v) in the mixture. After a 5 min preincubation at 37°C, the NADP⁺ (1 mM, 20 μ L) was added into the mixture to initiate the reaction. The reaction was terminated by the addition of methanol (200 μ L) with internal standard. The mixture was kept on ice until it was centrifuged at 20 000 \times g for 10 min at 4°C. Aliquots of supernatants were stored at -40°C until analysis. Control incubations without NADPH or without substrate or without microsomes were carried out to ensure that the formation of metabolites was microsomes and NADPH dependent. All incubations throughout the study were carried out in at least three independent

experiments with standard deviations (SD) generally below 10%.

2.5. CYP450s Inhibition Experiments. Inhibition study was conducted using the above incubation system consisting of liver microsomes, NADPH-generating system, probe substrate at the concentration of about K_m value, and xanthotoxol (or the control inhibitor). The concentrations of positive inhibitors used were as follows: 10 μ M furafylline for CYP1A2, 2.5 μ M 8-methoxypsoralen for CYP2A6, 10 μ M quercetin for CYP2C8, 10 μ M sulfaphenazole for CYP2C9, 10 μ M quinidine for CYP2D6, 50 μ M clomethiazole for CYP2E1, 20 μ M omeprazole for CYP2C19, 1 μ M ketoconazole for CYP3A4, and 500 μ M ABT for broad CYP450s. Kinetic analysis was performed where activity has been inhibited by more than 90%. Half inhibition concentration (IC₅₀) value was obtained by incubating various xanthotoxol concentrations (0–100 μ M for CYP1A2 and CYP3A4). The K_i value was obtained by incubating various xanthotoxol and probe substrates concentration.

2.6. Kinetic Study. To estimate kinetic parameters of xanthotoxol and make sure that the formations of metabolites were in the linear range of both reaction time and the concentration of microsomes, xanthotoxol (0.25–200 μ M) was incubated with pooled HLMs for 30 min. Reaction velocities and substrate concentrations were used to calculate the apparent K_m and V_{max} values according to nonlinear regression from the Michaelis-Menten equation (see (1)). CL_{int} was calculated as V_{max}/K_m . In addition, the result was graphically represented on Eadie-Hofstee plots (velocities versus ratios of velocities to substrate concentrations) to determine whether the metabolism was monophasic or biphasic. Consider

$$V = \frac{V_{max} \cdot [S]}{K_m + [S]}, \quad (1)$$

where V_{max} was the maximum reaction velocity, K_m was the Michaelis constant that represented the substrate concentration at which the velocity was half of V_{max} , V was the reaction velocity, and $[S]$ was the substrate concentration.

2.7. Prediction of In Vivo Hepatic Clearance. The following equations were used to predict the xanthotoxol clearance in human [13]:

$$CL_{int \text{ in vitro}} = \frac{V_{max}}{K_m},$$

$$CL_{int \text{ in vivo}} = CL_{int \text{ in vitro}} \cdot SF, \quad (2)$$

$$CL_H = \frac{Q_H \cdot f_u \cdot CL_{int \text{ in vivo}}}{Q_H + f_u \cdot CL_{int \text{ in vivo}}},$$

where SF (scaling factor) represents the milligrams of microsomal protein per gram of liver multiplied by the grams of liver weight; CL_{int} is the intrinsic metabolic clearance; CL_H

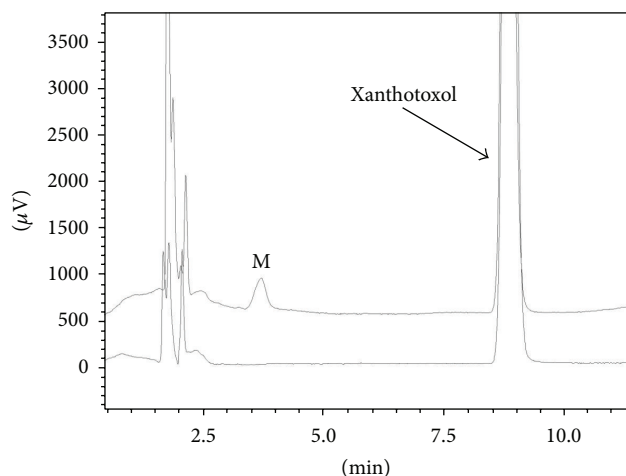


FIGURE 2: Chromatogram of incubation of xanthotoxol with human liver microsomes (HLMs). Xanthotoxol ($10\ \mu\text{M}$) was incubated with HLMs ($0.3\ \text{mg/mL}$) at 37°C for 60 min with (upper) or without (lower) a β -nicotinamide adenine dinucleotide phosphate-(NADPH-) generating system.

is hepatic clearance; f_u is the free fraction in blood (there are no data of xanthotoxol; here f_u was arbitrarily proposed to be 1); Q_H is the hepatic blood flow. The hepatic clearance of xanthotoxol was calculated using (2), and physiological parameters in human were described as follows: microsomal protein per gram of liver, liver weight per kilogram of body, and liver blood flow for human were $48.8\ \text{mg}$, $25.7\ \text{g}$, and $20.7\ \text{mL}\cdot\text{min}^{-1}\cdot\text{kg}^{-1}$, respectively [14].

2.8. Molecular Docking Analysis. The X-ray crystal structures of human CYP1A2 (pdb: 2H14) and CYP3A4 (pdb: 4K9W) were obtained from RCSB Protein Databank (<http://rcsb.org/>). The ligands for docking were prepared using SYBYL X2.1, and the energy was minimized using the external Tripos force field. The protonation state and energy minimization of the protein and the ligands were calculated using the default setting in SYBYL X2.1. The active sites were defined by a sphere of $6.0\ \text{\AA}$ from the native ligands in the crystal structures using Gold v5.2. The docked poses were scored using CHEMPLP scoring function. The best docked pose of the ligand was visualized using Pymol Molecular Graphics System v1.3.

2.9. HPLC Method. The HPLC system (Shimadzu, Kyoto, Japan) consisted of a CBM-20A lite system controller, two LC-20AB pumps, and an SPD-20A ultraviolet light (UV) detector. The chromatographic separation was achieved using a C18 column ($4.6\ \text{mm}:150\ \text{mm}, 5\ \text{mm}$ Kromasil). The mobile phases consisted of LC grade water containing 0.1% formic acid (A) and LC grade acetonitrile (B) with the following gradient profile: 0–12 min, 20% B; 12–13 min, 20–95% B; 13–19 min, 95% B; 19–20 min, 90–20% B; 20–25 min, 20% B. The flow rate was $1\ \text{mL}/\text{min}$. Detection wavelength was set at $310\ \text{nm}$ and the column temperature was set to 40°C .

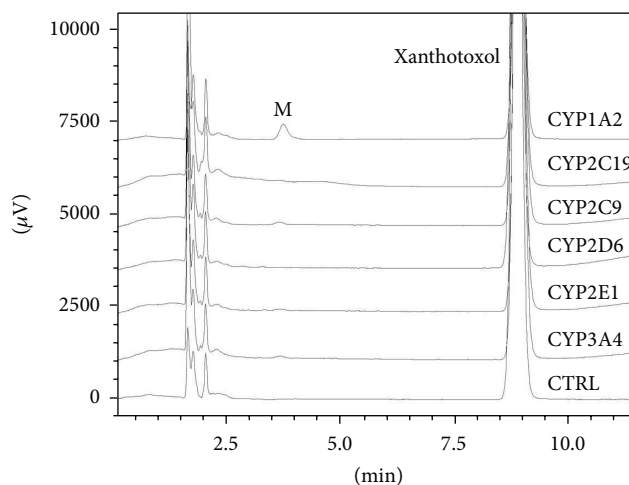


FIGURE 3: Representative HPLC profiles of xanthotoxol and its metabolite in recombinant CYP450 supersomes. Xanthotoxol ($10\ \text{mM}$) was incubated with recombinant CYP450 supersomes ($15\ \text{nM}$) and an NADPH-generating system at 37°C for 30 min.

3. Results

3.1. Analysis of Xanthotoxol Metabolites. After the incubations of xanthotoxol ($10\ \mu\text{M}$) in HLMs and NADPH-generating system, the new peak (M) was found at 3.7 min (Figure 2). The new peak was not observed in the negative controls without NADPH, or without substrate, or without microsomes.

3.2. Identifying CYP450s Involved in the Metabolism of Xanthotoxol. The enzymes involved in the xanthotoxol metabolism were investigated using cDNA-expressed human P450 isoforms including CYP3A4, CYP2D6, CYP2E1, CYP2C9, CYP2C19, and CYP1A2. The incubation for each isoenzyme was carried out as described for the liver microsomal study. As shown in Figure 3, the metabolite (M) was mainly generated in the presence of CYP1A2, and the amounts of M generated by CYP3A4, CYP2D6, CYP2E1, CYP2C9, and CYP2C19 were 4.5%, 0%, 7.3%, 13.5%, and 3.6% comparing to CYP1A2 level which was normalized to 100%.

3.3. Inhibition of Xanthotoxol against CYP3A4 and CYP1A2. All positive control inhibitors performed strong inhibition to the corresponding probe reactions with more than 80% of control activity inhibited. $100\ \mu\text{M}$ xanthotoxol inhibited the activities of CYP3A4, CYP2C9, CYP1A2, CYP2A6, CYP2D6, CYP2C8, CYP2C19, and CYP2E1 by 3.3, 35.9, 9.5, 82.1, 72.4, 52.1, 68.3, and 25.9%, respectively (Figure 4). Furthermore, kinetic analysis of CYP3A4 and CYP1A2 was performed. As shown in Figures 5 and 6, xanthotoxol inhibited diclofenac $4'$ -hydroxylation (CYP1A2) and testosterone 6β -hydroxylation (CYP3A4) in a concentration-dependent manner with the IC_{50} of $27.82\ \mu\text{M}$ for CYP1A2 and $7.43\ \mu\text{M}$ for CYP3A4. Lineweaver-Burk and Dixon plots showed that the inhibition of xanthotoxol to CYP1A2 and CYP3A4 was best fit to a

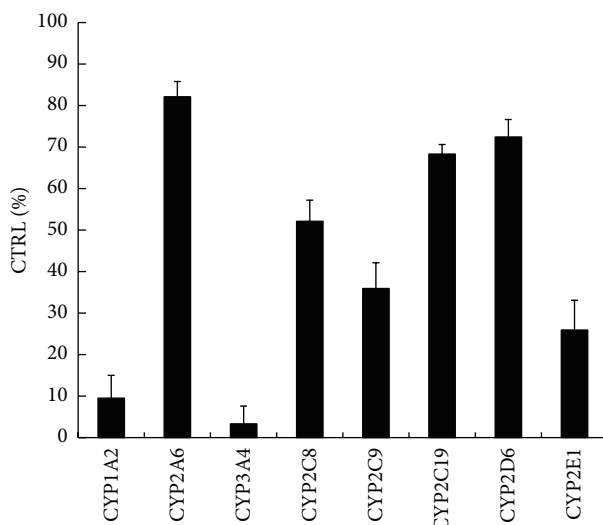


FIGURE 4: The inhibitory effects of xanthotoxol on CYP450s in human liver microsomes (HLMs).

noncompetitive way. The K_i value was calculated to be 21.15 and 2.22 μM for CYP1A2 and CYP3A4, respectively.

3.4. Kinetic Characteristics of Xanthotoxol Metabolism. Over the whole concentration range tested, the metabolism of xanthotoxol obeyed the typical monophasic Michaelis-Menten kinetics, as evidenced by the Eadie-Hofstee plot (Figure 7). Reaction velocity showed significant concentration-dependent characteristic. V_{\max} and K_m values were calculated as 0.55 $\text{nmol}\cdot\text{min}^{-1}\cdot\text{mg}^{-1}$ and 8.46 μM via nonlinear regression from the Michaelis-Menten equation using Origin software. Intrinsic clearance (CL_{int}) was 0.06 $\text{mL}\cdot\text{min}^{-1}\cdot\text{mg}^{-1}$.

3.5. Prediction of In Vivo Hepatic Clearance of Humans. Using the values of kinetic parameters generated in HLMs, CL_H was calculated to be 15.91 $\text{mL}\cdot\text{min}^{-1}\cdot\text{kg}^{-1}$ body weight. The percentage of CL_H versus hepatic blood flow (Q_H)% was 76.8%.

3.6. Molecular Docking Study. As shown in Figure 8(a), there was hydrophobic effect between xanthotoxol and CYP1A2 with ILE117. In addition, the benzene rings of xanthotoxol also showed π - π stacking interaction with PHE125, PHE226, and PHE260. For docking to CYP3A4 (Figure 8(b)), xanthotoxol was bound to CYP3A4 via hydrogen bond with ARG105 and hydrophobic interactions with ILE369 and LEU373.

4. Discussion

Cytochrome P450 enzymes (CYP450s) are heme-thiolate proteins that are responsible for the oxidative metabolism of numerous xenobiotics as well as endogenous substrates [15]. The inhibition against CYP450s has resulted in costly late failures of drug development and withdrawal of drugs on the market [16]. Thus, estimating HDIs is a priority

to predict inhibitory potential of major components on CYP450 isoforms and to clarify the inhibition mechanism and kinetics.

Using pooled HLMs, we determined the inhibitory effects of xanthotoxol on eight CYP450 isoforms and the results showed that xanthotoxol exhibited a stronger and noncompetitive inhibition to CYP1A2-mediated phenacetin o-deethylation and CYP3A4-mediated testosterone 6 β -hydroxylation with the IC_{50} values of 27.82 μM and 7.43 μM , respectively. The inhibition kinetic parameters (K_i) were calculated to be 21.15 μM and 2.22 μM for CYP1A2 and CYP3A4, respectively. The metabolite (M) of xanthotoxol was detected in HLMs and obeyed the typical monophasic Michaelis-Menten kinetics. The generation of the metabolite was NADPH dependent. Aminobenzotriazole (ABT), a nonspecific inhibitor of cytochrome P450s (CYPs), showed potent inhibition against metabolite production, which proves that metabolism was catalyzed by CYP450s. The kinetic characteristics of xanthotoxol metabolism were 0.55 $\text{nmol}\cdot\text{min}^{-1}\cdot\text{mg}^{-1}$ and 8.46 μM for V_{\max} and K_m . CL_{int} was 0.065 $\text{mL}\cdot\text{min}^{-1}\cdot\text{mg}^{-1}$. According to the Eadie-Hofstee plots, xanthotoxol metabolism showed a monophasic feature, suggesting that xanthotoxol was metabolized by only one isozyme or two isozymes with the same K_m value. CYP3A4 is the most important CYP isoform for human and metabolizes approximately 50% of marketed drugs [17]. Thus, potential metabolism-based herb-drug interactions might occur during coadministration of xanthotoxol with other CYP3A4 or CYP1A2 substrates.

In this study, we identified the CYP450 isoforms involved in xanthotoxol metabolism by screening assays with recombinant CYP450 supersomes including CYP1A2, CYP3A4, CYP2E1, CYP2C9, CYP2D6, and CYP2C19. CYP1A2 contributed main effect to catalyze the formation of metabolite of xanthotoxol, while other enzymes displayed a very limited ability to metabolize xanthotoxol. Thus, xanthotoxol was selectively metabolized by CYP1A2.

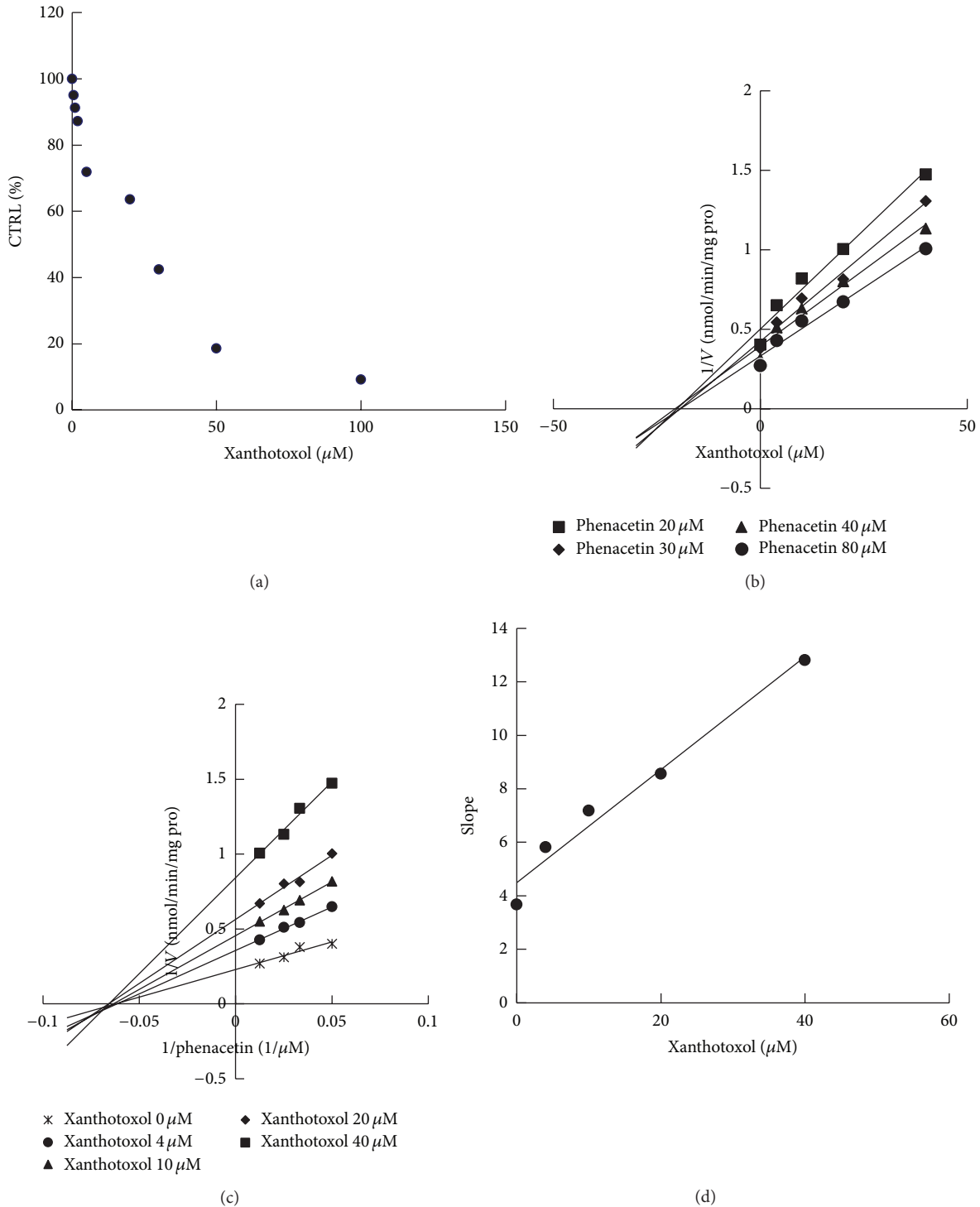


FIGURE 5: (a) Inhibitory effects of xanthotoxol on phenacetin o-deethylation activity (CYP1A2). (b) Dixon plot of inhibition effect of xanthotoxol on phenacetin o-deethylation (CYP1A2). (c) Lineweaver-Burk plot of inhibitory effect of xanthotoxol on phenacetin o-deethylation (CYP1A2). (d) Secondary plot of slopes from Lineweaver-Burk plot versus xanthotoxol concentrations. Each data point represented mean of triplicate incubations.

Hepatic clearance is considered to be one of the most important pharmacokinetic parameters as it directly relates to drug elimination and bioavailability. So a prediction of hepatic metabolic clearance is of primary importance during

the drug discovery and development process. In this study, we investigated the quantitative prediction of human hepatic metabolic clearance from in vitro experiments using human liver microsomes focusing on CYP450s metabolism. CL_H

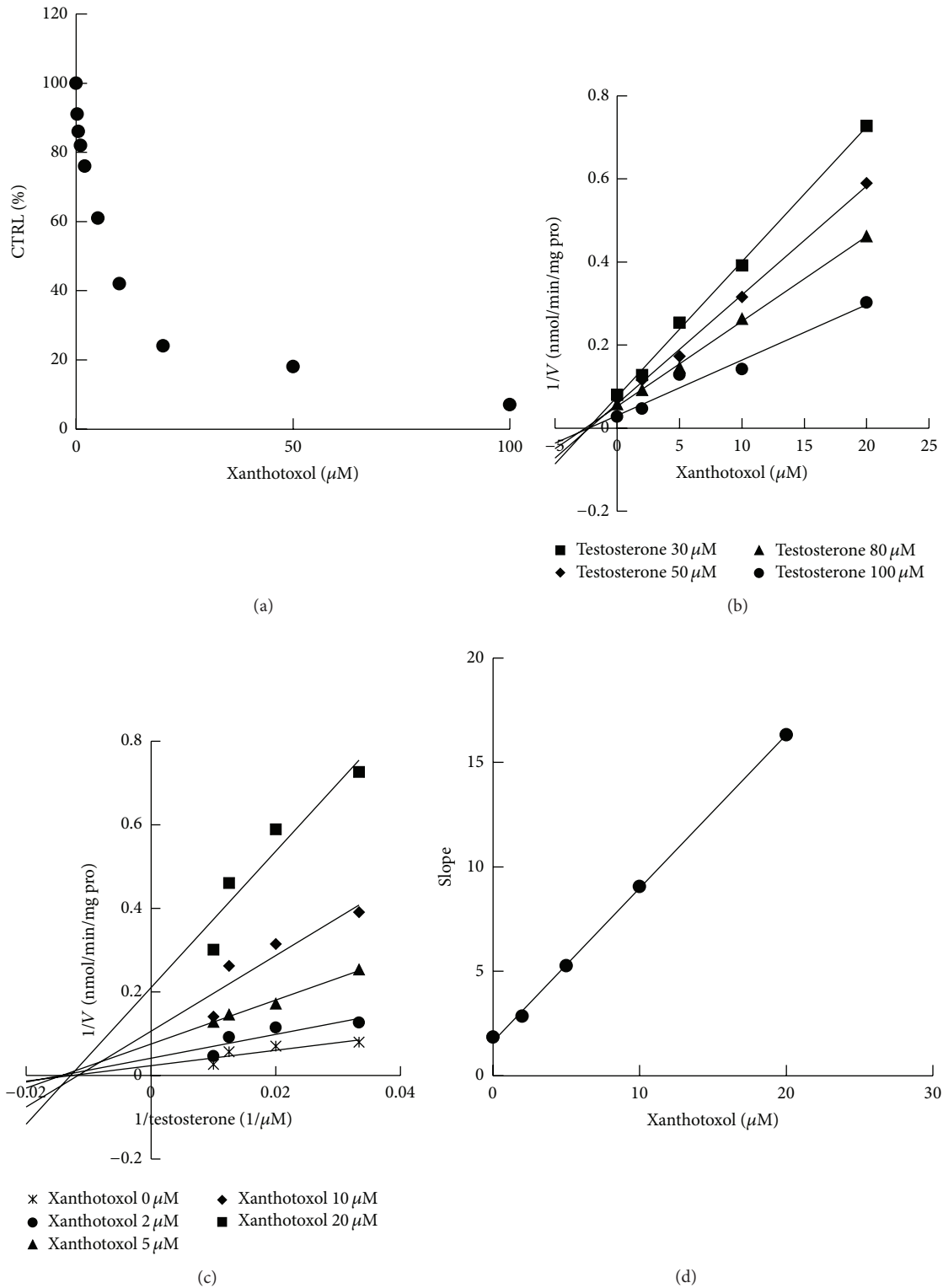


FIGURE 6: (a) Inhibitory effects of xanthotoxol on testosterone 6 β -hydroxylation activity (CYP3A4). (b) Dixon plot of inhibition effect of xanthotoxol on testosterone 6 β -hydroxylation (CYP3A4). (c) Lineweaver-Burk plot of inhibitory effect of xanthotoxol on testosterone 6 β -hydroxylation (CYP3A4). (d) Secondary plot of slopes from Lineweaver-Burk plot versus xanthotoxol concentrations. Each data point represented mean of triplicate incubations.

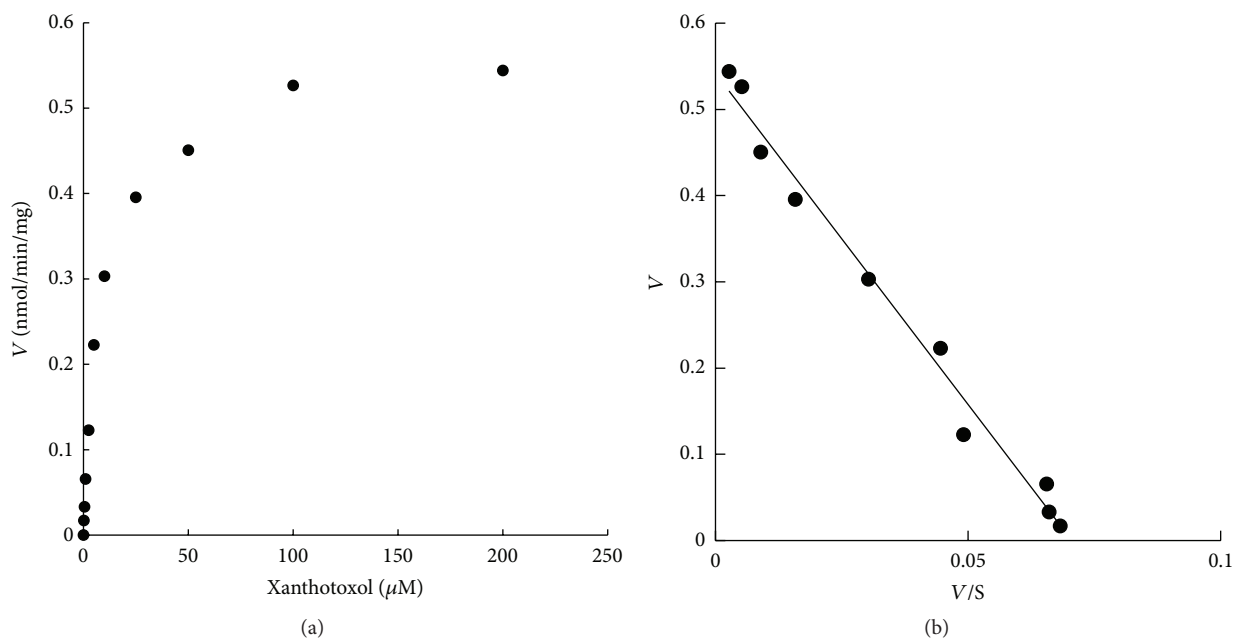


FIGURE 7: Michael-Menten plots (a) and Eadie-Hofstee plots (b) of xanthotoxol metabolism in human liver microsomes (HLMs). Incubation conditions were carried out as described in Section 2. Each point represents the mean of three independent experiments in triplicate determinations.

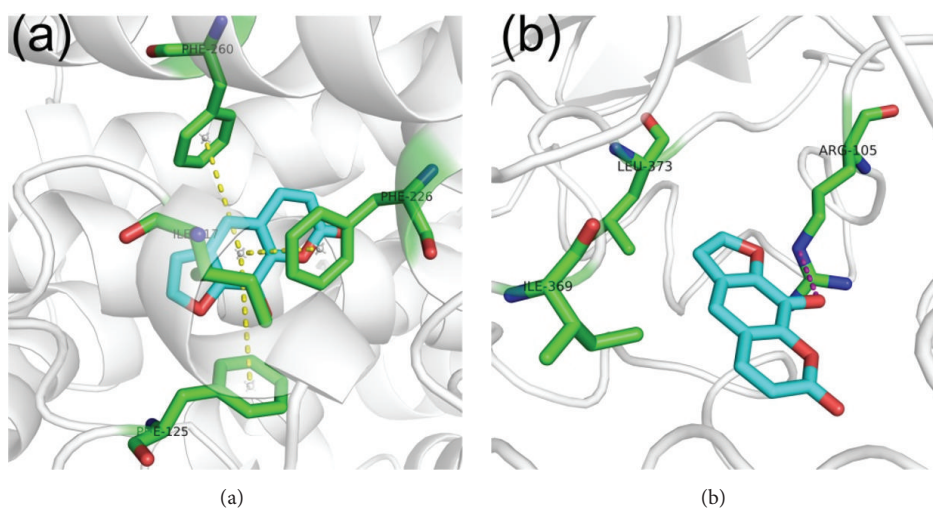


FIGURE 8: Binding modes of compound with CYP 1A2 (a) and 3A4 (b). The compound was displayed in cyan stick, residues were displayed in green sticks, hydrogen bond was displayed in purple dotted line, and π - π stacking was displayed in yellow dotted line.

was calculated to be $15.91 \text{ mL}\cdot\text{min}^{-1}\cdot\text{kg}^{-1}$ body weight. In general, drugs that have CL_H above 70% Q_H are classified as high-clearance drugs and those below 30% Q_H are classified as low-clearance drugs [18]. So we predicted from in vitro data that xanthotoxol was high-clearance drug in human body.

Analyzing molecular docking, we found that xanthotoxol was bound to CYP1A2 with hydrophobic interaction and π - π stacking and CYP3A4 with hydrogen bonds and hydrophobic interaction, which implied that xanthotoxol exhibited stronger interaction with CYP3A4 than CYP1A2. The enzyme kinetic studies confirmed the molecular docking study that

xanthotoxol had lower IC_{50} and K_i values against CYP3A4 than CYP1A2.

It is important to understand the pharmacokinetic profiles of xanthotoxol and its drug interaction with CYP450s. Our results indicate that xanthotoxol inhibited the activity of CYP1A2 and CYP3A4 and therefore may interact with drugs that were metabolized by these two isozymes.

Competing Interests

The authors have declared that there are no competing interests.

Acknowledgments

The authors thank Shuman Yang for revising the English language.

References

- [1] W. He, B.-L. Zhang, S.-Y. Zhou, X.-L. Sun, and S.-Y. Zhang, "Facile total synthesis of xanthotoxol," *Synthetic Communications*, vol. 37, no. 3, pp. 361–367, 2007.
- [2] W. He, W. Chen, Y. Zhou, Y. Tian, and F. Liao, "Xanthotoxol exerts neuroprotective effects via suppression of the inflammatory response in a rat model of focal cerebral ischemia," *Cellular and Molecular Neurobiology*, vol. 33, no. 5, pp. 715–722, 2013.
- [3] T. B. Ng, F. Liu, and Z. T. Wang, "Antioxidative activity of natural products from plants," *Life Sciences*, vol. 66, no. 8, pp. 709–723, 2000.
- [4] O. P. Sethi and K. Naik, "Xanthotoxol (ST)—a potent 5-HT antagonist," *Indian Journal of Physiology and Pharmacology*, vol. 23, no. 2, pp. 142–143, 1979.
- [5] Y.-M. Han, I.-S. Kim, S.-U. Rehman, K. Choe, and H.-H. Yoo, "In vitro evaluation of the effects of *Eurycoma longifolia* extract on CYP-mediated drug metabolism," *Evidence-Based Complementary and Alternative Medicine*, vol. 2015, Article ID 631329, 6 pages, 2015.
- [6] I.-X. Goh, C.-H. How, and S. Tavintharan, "Cytochrome P450 drug interactions with statin therapy," *Singapore Medical Journal*, vol. 54, no. 3, pp. 131–135, 2013.
- [7] N. Çoruh, A.-G. S. Celep, and F. Özgökçe, "Antioxidant properties of *Prangos ferulacea* (L.) Lindl., *Chaerophyllum macropodium* Boiss. and *Heracleum persicum* Desf. from Apiaceae family used as food in Eastern Anatolia and their inhibitory effects on glutathione-S-transferase," *Food Chemistry*, vol. 100, no. 3, pp. 1237–1242, 2007.
- [8] L.-N. Shan, S.-M. Yang, G. Zhang et al., "Comparison of the inhibitory potential of bavachalcone and corylin against UDP-glucuronosyltransferases," *Evidence-Based Complementary and Alternative Medicine*, vol. 2014, Article ID 958937, 6 pages, 2014.
- [9] S. Liu, X. Zhang, F. Qiu et al., "Metabolic interaction of the active constituents of *Coptis chinensis* in human liver microsomes," *Evidence-Based Complementary and Alternative Medicine*, vol. 2015, Article ID 802903, 5 pages, 2015.
- [10] S. W. Grimm, H. J. Einolf, S. D. Hall et al., "The conduct of in vitro studies to address time-dependent inhibition of drug-metabolizing enzymes: a perspective of the Pharmaceutical Research and Manufacturers of America," *Drug Metabolism and Disposition*, vol. 37, no. 7, pp. 1355–1370, 2009.
- [11] T.-D. Bjornsson, J.-T. Callaghan, H.-J. Einolf et al., "The conduct of in vitro and in vivo drug-drug interaction studies: a Pharmaceutical Research and Manufacturers of America (PhRMA) perspective," *Drug Metabolism and Disposition*, vol. 31, no. 7, pp. 815–832, 2003.
- [12] Y.-Y. Zhang, Y. Liu, J.-W. Zhang et al., "C-7 configuration as one of determinants in taxanes metabolism by human cytochrome P450 enzymes," *Xenobiotica*, vol. 39, no. 12, pp. 903–914, 2009.
- [13] W. Li, Y. Liu, Y.-Q. He et al., "Characterization of triptolide hydroxylation by cytochrome P450 in human and rat liver microsomes," *Xenobiotica*, vol. 38, no. 12, pp. 1551–1565, 2008.
- [14] Y. Naritomi, S. Terashita, S. Kimura, A. Suzuki, A. Kagayama, and Y. Sugiyama, "Prediction of human hepatic clearance from in vivo animal experiments and in vitro metabolic studies with liver microsomes from animals and humans," *Drug Metabolism and Disposition*, vol. 29, no. 10, pp. 1316–1324, 2001.
- [15] X. Du, H. Ye, C. Zhang, L. Ye, and G. Lin, "Effect of kanglaite on rat cytochrome P450," *Pharmaceutical Biology*, vol. 53, no. 7, pp. 995–1001, 2015.
- [16] R. Arimoto, "Computational models for predicting interactions with cytochrome p450 enzyme," *Current Topics in Medicinal Chemistry*, vol. 6, no. 15, pp. 1609–1618, 2006.
- [17] S.-F. Zhou, "Drugs behave as substrates, inhibitors and inducers of human cytochrome P450 3A4," *Current Drug Metabolism*, vol. 9, no. 4, pp. 310–322, 2008.
- [18] X.-Q. Li, A. Björkman, T. B. Andersson, L. L. Gustafsson, and C. Masimirembwa, "Identification of human cytochrome P450s that metabolise anti-parasitic drugs and predictions of in vivo drug hepatic clearance from in vitro data," *European Journal of Clinical Pharmacology*, vol. 59, no. 5-6, pp. 429–442, 2003.