

Vaccine response following anti-CD20 therapy: a systematic review and meta-analysis of 905 patients

Abi Vijenthira, 1,2 Inna Gong, 2 Stephen D. Betschel, 2,3 Matthew Cheung, 2,4 and Lisa K. Hicks 2,5

¹Division of Medical Oncology and Hematology, Princess Margaret Cancer Centre, Toronto, ON, Canada; ²Department of Medicine, University of Toronto, Toronto, ON, Canada; ³Division of Allergy and Immunology, St. Michael's Hospital, Toronto, ON, Canada; ⁴Division of Medical Oncology and Hematology, Sunnybrook Health Sciences Centre, Toronto, ON, Canada; and ⁵Division of Hematology/Oncology, St. Michael's Hospital, Toronto, ON, Canada

The objective of this study was to perform a systematic review of the literature on vaccine responsiveness in patients who have received anti-CD20 therapy. PubMed and EMBASE were searched up to 4 January 2021 to identify studies of vaccine immunogenicity in patients treated with anti-CD20 therapy, including patients with hematologic malignancy or autoimmune disease. The primary outcomes were seroprotection (SP), seroconversion (SC), and/or seroresponse rates for each type of vaccine reported. As the pandemic influenza vaccine (2009 H1N1) has standardized definitions for SP and SC, and represented a novel primary antigen similar to the COVID-19 vaccine, meta-analysis was conducted for SC of studies of this vaccine. Pooled estimates, relative benefit ratios (RBs), and 95% confidence intervals (CIs) were calculated using a random-effects model. Thirtyeight studies (905 patients treated with anti-CD20 therapy) were included (19 studies of patients with hematologic malignancies). Patients on active (<3 months since last dose) anti-CD20 therapy had poor responses to all types of vaccines. The pooled estimate for SC after 1 pandemic influenza vaccine dose in these patients was 3% (95% CI, 0% to 9%), with an RB of 0.05 (95% CI, 0-0.73) compared with healthy controls and 0.22 (95% CI, 0.09-0.56) compared with disease controls. SC compared with controls seems abrogated for at least 6 months following treatment (3-6 months post anti-CD20 therapy with an RB of 0.50 [95% CI, 0.24-1.06] compared with healthy and of 0.44 [95% CI, 0.23-0.84] compared with disease controls). For all vaccine types, response to vaccination improves incrementally over time, but may not reach the level of healthy controls even 12 months after therapy.

Introduction

Immunocompromised patients including those with hematologic malignancy are at increased risk of infections, including those that are vaccine preventable. COVID-19 is a viral infection with high mortality in immunocompromised patients. In healthy vaccine recipients, severe COVID-19 can be prevented; however, patients with immunocompromising conditions and/or those on immunosuppression were excluded from initial vaccine trials. With the advent of the COVID-19 vaccination rollout, clinicians are faced with questions regarding the effectiveness of vaccination in immunocompromised patients, particularly in those receiving anti-CD20 therapy.

Submitted 1 March 2021; accepted 23 April 2021; published online 21 June 2021. DOI 10.1182/bloodadvances.2021004629.

All data are reported in the paper in tables or figures. Data tables are available from the corresponding author at lisak.hicks@unityhealth.to.

The full-text version of this article contains a data supplement. © 2021 by The American Society of Hematology

Although anti-CD20 therapy is known to efficiently deplete peripheral B cells, these constitute only 2% of the total B-cell population. The effect of anti-CD20 treatment on B cells located in peripheral lymphoid tissues is less clear. Additionally, anti-CD20 therapy has less of an effect on memory B cells and long-lived plasma cells, which do not express the CD20 antigen. Furthermore, the peripheral blood B-cell compartment begins to recover 76 to 9 months after therapy. There is limited and conflicting evidence on the ability of anti-CD20—treated patients to mount a vaccine response to both primary and recall vaccinations. Previous reports reviewing vaccine response following anti-CD20 therapy have been limited by their scope.

As the COVID-19 pandemic continues and vaccinations begin, to help inform counseling, public health policies, and future research on vaccination in this vulnerable patient population, reviewing the literature regarding vaccine responsiveness of patients receiving anti-CD20 therapy is critical. The objective of this study was to perform a systematic review of the literature on vaccine responsiveness in patients receiving anti-CD20 therapy.

Methods

This study is reported according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses.

Definitions

Definitions used in this article are in accordance with the World Health Organization (WHO) Report on Biological Standardization. Immunological correlates of protection (ICP) have been established for a number of different vaccines. When an ICP has been established, the seroprotection (SP) rate is the percentage of subjects who achieve an antibody level at or above the ICP. The seroconversion (SC) rate is a predefined increase in serum antibody concentration or titer. In subjects without a detectable or quantifiable antibody (below the lower limit of detection or quantification) prior to vaccination, SC is usually defined as achieving a quantifiable antibody level postvaccination. In subjects with a quantifiable antibody prior to vaccination (due to previous exposure or vaccination), SC is commonly defined by a predefined fold increase from pre- to postvaccination.

SP and SC have been standardized for influenza vaccines.²³ For other vaccines (eg, tetanus, diphtheria, pertussis, pneumovax conjugate and pneumovax polysaccharide, *Haemophilus influenzae* B, hepatitis B, hepatitis A, polio, varicella zoster), SP and SC definitions are not uniformly standardized. In our review, if an article used response criteria that did not meet the WHO definition for either SP or SC as indicated in this section, then the vaccination response rate was described as seroresponse (SR).

Eligibility criteria

All studies published on response to clinically available vaccinations in patients treated with anti-CD20 therapy were considered for inclusion, including noncancer and cancer populations. Only studies reporting SP, SC, and SR rates, and/or measurement of cellular (T-cell) immune response, were included. Case reports, conference proceedings, and studies with <10 patients who had received anti-CD20 therapy were excluded. Only English language reports were included. Full inclusion criteria are available in supplemental Table 1.

Information sources

PubMed and EMBASE databases were searched up to the week of 4 January 2021.

Search and study selection

The full search strategy is available in supplemental Table 2. Two authors (A.V. and I.G.) independently conducted the search strategy, and results were compared with ensure concordance. Differences in classification were discussed and resolved, with a third author (L.K.H.) available for resolution of disagreements. Titles and abstracts of articles were reviewed and any that were clearly irrelevant were excluded. Full texts of remaining articles were reviewed to find studies that met the inclusion criteria. Additionally, systematic reviews discovered in the search that focused on vaccine response in patients receiving immunosuppressive therapy were screened to identify additional references.

Data collection process and data items

A data extraction form was used to extract relevant information from the articles. Information was extracted on study design; geographic location; type of immunogenicity studies conducted (humoral vs cellular); the anti-CD20 drug used by patients; patient population; sample size; median/mean age; distribution by sex; type of vaccine administered; how response was defined; SP, SC, and SR rates; cellular response data if available; and safety data for patients with hematologic malignancies. Patients were considered to be on active treatment if the average interval between anti-CD20 therapy and vaccination was <12 weeks; otherwise, treatment durations were divided into average interval 3 to 6 months, 6 to 12 months, and >12 months. Data on controls (healthy and disease controls not receiving anti-CD20 therapy) in the study and their outcomes were also extracted.

Risk of bias in individual studies

To assess the quality of cohort studies and substudies of trials with a comparison group, the Newcastle-Ottawa assessment tool was used. 24,25 For 1-arm cohort studies, the Joanna Briggs Institute Critical Appraisal Checklist for Cohort Studies was used. For randomized studies, the Cochrane Risk of Bias Scale was used. See supplemental Table 3 for details on each tool. Assessment was conducted by 2 authors (A.V. and I.G.), while a third author (L.K.H.) was available for differences of opinion.

Summary measures

The primary outcomes were the SP, SC, and/or SR rates to each vaccine. As the pandemic influenza vaccination has standardized definitions for SP and SC, ²³ this enabled meta-analysis of point estimates of SC to vaccination in patients treated with anti-CD20 therapy, as well as relative benefit ratios (RBs) comparing SC in anti-CD20-treated patients to healthy or disease controls. Secondary outcomes were cellular response to vaccinations, and safety data on vaccinations in patients with hematologic malignancies. Due to limitations in data, subgroup and sensitivity analyses were not performed.

Synthesis of results

The majority of the data synthesis was descriptive. For metaanalysis, the principal summary measures used were pooled

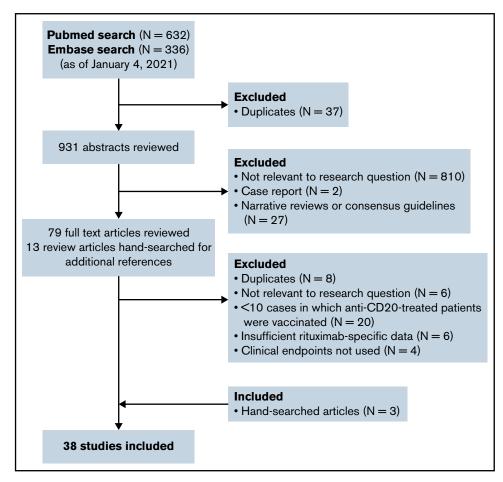


Figure 1. Flow diagram of studies assessed for inclusion.

estimates and RBs with 95% confidence intervals (Cls). Heterogeneity between estimates was assessed using the I2 statistic, and interpreted per the Cochrane Handbook recommendations: I² 0% to 40%, heterogeneity likely not substantial; 30% to 60%, moderate heterogeneity; 50% to 90%, substantial heterogeneity; and 75% to 100%, considerable heterogeneity.²⁸ Pooled estimates were transformed using the Freeman-Tukey double arcsine method, 29 and the final pooled results were back-transformed with a 95% CI for ease of interpretation. Meta-analysis was performed using a randomeffects model (DerSimonian and Laird) using the MetaXL (www. epigear.com) add-in for Microsoft Excel, as well as Review Manager 5.4 (Cochrane Collaboration, 2020).

Results

Study selection

Figure 1 shows the flow diagram for study selection. A total of 38 peer-reviewed studies (31 cohort, 3 phase 3, 2 phase 3 substudies, 1 phase 1, 1 phase 1/2 substudy) comprising 905 anti-CD20-treated patients from North America, Europe, and Asia were included. Nineteen studies assessed vaccine responses in patients receiving anti-CD20 for hematologic malignancy; 19 studies assessed vaccine responses in patients receiving anti-CD20 for autoimmune diseases such as rheumatoid arthritis or multiple sclerosis.

Study characteristics

Tables 1 and 2 list the summary characteristics of the included studies. Supplemental Table 4 lists study definitions of vaccine response assessment.

Risk of bias within studies

Supplemental Table 5 lists the results of the risk-of-bias assessment for individual studies. Twenty-eight studies had a low risk of bias.

Results of individual studies

Humoral responses to pandemic influenza, seasonal influenza, tetanus, diphtheria, pertussis, Haemophilus influenzae B, hepatitis B, hepatitis A, polio, and pneumococcal vaccinations. Table 3 provides a high-level summary of all of the reports of humoral immune responses in patients receiving rituximab. Details of individual studies are available in Tables 1 and 2. As outlined in both tables, SP, SC, and/or SR rates were lower in anti-CD20-treated patients than in healthy controls or disease controls, regardless of the time that had elapsed from treatment. In patients on active anti-CD20 therapy, response to vaccination was poor regardless of the vaccine studied, with SC rates of 0% to 25%. 30-37 Response rates appeared to improve as more time elapsed after anti-CD20 therapy, with the best responses in

Table 1. Literature summary: humoral responses

Study	Design	Location	Anti-CD20 drug	Population/ Control population	Z	Median age, y*	Sex, % F	Interval between anti- CD20 and vaccination	% dS	% >S	SR %
T-cell-dependent vaccinations											
Pandemic influenza											
Bedognetti et al, 2012 ⁴⁴	Cohort	Italy	RTX	NHL in CR; Healthy controls	14; 14		Z Z	33 mo (range, 14-78 mo)	D1: 93, D2: 93; D1: 86, D2: 93	D1: 36, D2: 64; D1: 71; D2: 79	
Berglund et al, 2014 ³⁰	Cohort	Sweden	RTX	Lymphoma on treatment; Disease controls with cancer on active treatment other than RTX	13; 83	63†	1 86	All patients on active therapy	D1: 0, D2: 8; D1: 62, D2: 87	D1: 0, D2: 0; D1: 62, D2: 84	
de Lavallade et al, 2011 ⁴⁰	Cohort	ž	XF	B-cell malignancies	25	+29	404	19 mo (range, 2-83 mo)	D1: 33, D2: 42; 5 patients, 6 mo: D1: 0, D2: 0; 4 patients, 6-12 mo: D1: 50, D2: 50; 3 patients, >12 mo: D1: 66, D2: 100	D1: 33, D2: 42; 5 patients, <6 mo: D1: 0, D2: 0; 4 patients, 6-12 mo: D1: 50, D2: 50; 3 patients, >12 mo: D1: 66, D2: 100	
de Lavallade et al, 2011 ⁴⁰	Cohort	ž	RTX	Disease controls of B-cell malignancy not previously treated	o	57+	40+	19 mo (range, 2-83 mo)	D1: 56, D2: 89	D1: 78, D2: 78	
de Lavallade et al, 2011 ⁴⁰	Cohort	¥	RTX	Disease controls postallotransplant	22	57†	40+	19 mo (range, 2-83 mo)	D1: 46, D2: 73	D1: 46, D2: 73	
de Lavallade et al, 2011 ⁴⁰	Cohort	Ϋ́	RTX	Disease controls with chronic myeloid leukemia	20	57†	40+	19 mo (range, 2-83 mo)	D1: 85, D2: 90	D1: 80, D2: 95	
Hottinger et al, 2012 ³¹	Cohort	Switzerland	RTX	Lymphoma on treatment; Healthy controls; Lymphoma on active treatment without RTX; Lymphoma not on active treatment	11; 138; 51; 119	61+	47+	On active therapy	D1: 0, D2: 27; 87; D1: 75, D2: 92; D1: 79, D2: 90	D1: 0, D2: 18; 87; D1: 75, D2: 92; D1: 79, D2: 87	
lde et al, 2014 ⁴⁵	Cohort	Japan	RTX	Lymphoma and acute Ieukemia	1	59+	60+	NR	D1: 0, D2: 0	10	D1: 0, D2: 9
lssa et al, 2011 ⁵⁹	Cohort	USA	RTX	Postallotransplant with RTX exposure in previous 1 y; Disease controls with no previous RTX exposure	71	55+	51+	<12 mo	88 		
Kapetanovic et al, 2014 ³²	Cohort	Sweden	RTX	RA; Disease controls RA and spondyloarthropathy on non-RTX treatments	281	62	80	On active therapy	6 patients: 1D, 17, 4 patients: 2D, 25; 1D: 50, 2D: 65	6 patients: 1D, 0, 4 patients: 2D, 25; 1D: 45, 2D: 56	

response; D1, dose 1 of a 2-dose series; D2, dose 2 of a 2-dose series; DM, diabetes mellitus; DMARD, disease-modifying anti-rheumatic drug; F, female; HBV, hepatitis B virus; IBD, inflammatory bowel disease; IFN, interferon; IQR, interferon; OCR, core-interquartile range; ITP, immune thrombocytopenia; MMF, mycophenolate mofetil; MS, multiple sclerosis; MTX, methotrexate; NHL, non-Hodgkin lymphoma; NMOSD, neuromyelitis optica spectrum disorder; NR, not reported; OCR, core-lizumab; OCR, core-lizumab; OCR, seroconversion; SR, seroresponse; T1DM, type 1 diabe-lizumab; OCR, seroconversion; SR, seroresponse; T1DM, type 1 diabe-1D, response at the end of 1 dose of a 1-dose series; 2D, response at the end of 2 doses of a 2-dose series; AIRD, autoimmune rheumatic disease; ASCT, autologous stem cell transplant; CBT, cord blood transplant; CR, complete mellitus; UK, United Kingdom; USA, United States of America.

*Median age; where not reported, mean age reported.

Rituximab-specific age and percentage female not reported, thus age and percentage female of entire study population reported.

Table 1. (continued)

. April 40	Decision	Costion	Anti-CD20	Population/	2	Median	% yes	Interval between anti-	ő Q	% ၂ ၂	% Q
Kim et al, 2013 ⁶⁰	Cohort		RTX	7			18	20 wk (range, 1-45 wk)	19; 100; 20; 86	38; 75; 40; 93	
Mackay et al, 2011 ³³	Cohort	Canada	XIX	Hematologic malignancy; Disease controls with hematologic malignancy on active therapy without RTX; Disease controls with solid tumor on active therapy	11; 20 20	53+	35+	On active therapy	.9. 40; 50	0; 33; 45	
Monkman et al, 2011 ³⁴	Cohort	Canada	RIX	Hematologic malignancy; Disease controls who had never received RTX	16; 46	67	47	12 patients: on active therapy; 4 patients: previously received (unknown length of time since last treatment)	Active RTX: 33	Active RTX: 17 and previous RTX: 13;	
Muller et al, 2013 ⁴³	Cohort	Switzerland	RTX	Autoimmune rheumatic diseases	16	45	88	Median, 6 mo (range, 1-36 mo)			D1: 38, D2: 44
Villa et al, 2013 ⁹⁷	Phase 3	Canada	RTX	Lymphoid malignancies on active chemotherapy or within 1 y following ASCT; Disease controls who had not received RTX	14; 26	49-57†	09	On active therapy, therapy within last 3 mo, or ASCT within past year	D1: 14, D2: 21; D1: 35, D2: 46	D1: 0, D2: 14; D1: 27, D2: 39	
Yri et al, 2011 ⁶¹	Cohort	Norway	RTX	Lymphoma; 6 Healthy controls	67; 51	63	43	Median, 21 d (range, 0-171 d)	0; 82		
Seasonal influenza											
Arad et al, 2011 ³⁹	Cohort	Israel	RTX	RA; Healthy controls; Disease controls on 1 DMARDs	29; 16; 17	62	79	16 patients:	58-97; 88-100; 88-100	14-35 (avg. 26); 38-44 (avg. 42); 61-78 (avg. 68)	
Bar-Or et al, 2020 ⁶²	Phase 3	USA and Canada	OCR	Relapsing MS; Disease controls on IFN 3 or no treatment	68; 34	40	99	10-18 wk	56-80; 75-97	10-60; 53-89	
Bedognetti et al, 2011 ⁵²	Cohort	Italy	RTX	NHL in CR; Healthy controls	31; 34	99	61	29 mo (range, 7-65 mo)	23-74; 44-94	3-29; 29-53	
Bedognetti et al, 2012 ⁴⁴ (conjugated seasonal influenza)	Cohort	Italy	RTX	NHL in CR; Healthy controls	14; 14	65+	χ Z	33 mo (range, 14-78 mo)	57-79; 86-100	29-43; 43-79	
Berglund et al, 2014 ³⁰	Cohort	Sweden	RTX	Lymphoma on treatment; 1 Disease controls of patients with cancer on active treatment	13; 83	63†	59†	All patients on active treatment	8-17; 59-70	0; 42-50	

1D, response at the end of 1 dose of a 1-dose series; 2D, response at the end of 2 doses of a 2-dose series; ARD, autoimmune rheumatic disease; ASCT, autologous stem cell transplant; CBT, cord blood transplant; CR, complete response; D1, dose 1 of a 2-dose series; D2, dose 2 of a 2-dose series; DM, diabetes mellitus; DMARD, disease-modifying anti-rheumatic drug; F, female; HBV, hepatitis B virus; IBD, inflammatory bowel disease; IPI, interferon; IQR, interferon; IQR, interferon; IQR, multiple sclerosis; MTX, methotrexate; NHL, non-Hodgkin lymphoma; NMOSD, neuromyelitis optica spectrum disorder; NR, not reported; OCR, corelizumab; OCR, corelizumab; RA, rheumatoid arthritis; RAR BCL, relapsed/refractory B-cell lymphoma; RTX, rituximab; SLE, systemic lupus erythematosus; SC, serocronversion; SR, seroprotection; SR, seroresponse; T1DM, type 1 diabetes mellitus; UK, United Kingdom; USA, United States of America.

*Median age; where not reported, mean age reported. †Rituximab-specific age and percentage female not reported, thus age and percentage female of entire study population reported.

Table 1. (continued)

Choice et al. 2017 ^a Choice LiSA RTN Automace Stateshop 23 S1 Choice Choice LiSA RTN Automace Stateshop 24 S2 S2 S2 Choice Choice LiSA RTN Handly controls Choice C	Study	Design	Location	Anti-CD20 drug	Population/ Control population	z	Median age, y*	Sex, % F	Interval between anti- CD20 and vaccination	% dS	% > S	SR %
Color List List Red Red List	ho et al, 2017 ⁶³	Cohort	USA	RTX	Autoimmune blistering skin diseases; Healthy controls	23; 28	51	65	11 mo (range, 5-24 mo)	69-77; 64-100		
1 1 1 1 1 1 1 1 1 1	Eisenberg et al, 2013 ⁴²	Cohort	USA	RTX	Rheumatologic disease; Healthy controls	17; 15	49	94	7-9 mo			17; 67
Cohor Cohor The Netherlander RTX Additional refinance from the file and a controls on MIX Cohor The Netherlander RTX RAM, controls on MIX Cohor)ren et al, 2008 ⁵¹	Cohort	Israel	XTX	RA; Healthy controls; Disease controls treated with DMARDs	14; 21; 29	53	76	7 patients: <6 mo; All patients: <18 mo; no difference in interval between responders and non-responders		21-36; 40-45; 30-67	
Cohort The Netherlands RTX RAting controls on MTX Page Cohort The Netherlands RTX Rating controls on MTX Page Cohort	iichi et al, 2019 ⁶⁴	Cohort	Spain	RTX	Autoimmune inflammatory diseases (AIRD, psoriasis, or IBD)	20	49†	59+	N N	40-55; ≥12 wk: 80; <12 wk: 25		
Cohort USA and Comada Comada Cohort Co	an Assen et al, 2010 ³⁵		The Netherlands	RTX	RA; Healthy controls; Disease controls on MTX	23; 29; 20	26	20	11 patients: 4-8 wk; 12 patients: 6-10 mo	17-26; 48-86; 25-65	4-8 wk: 0; 6-10 mo: 25	
Cohort USA and OCR Relapsing MS; Se Se Se Thook To treatment of the control on IFN Se Se Se Thook To treatment of the control on IFN Se Se Se Se To the control on IFN Se Se Se To the control on IFN Se Se Se To the control on IFN Se Se Se Se To the control on IFN Se Se Se To the control on IFN Se Se Se Se Se Se To the control on IFN Se Se Se Se Se Se Se S	tanus											
Phase 3 USA and Canada CoRA Relepsing MS; Contact Cont	Nbert et al, 2008 ⁶⁵	Cohort	NSA	RTX	SLE	14	43	86	7 mo			36
Phase 3 USA RTX Active RA; 68; 50 78 22-26 wk Cohort Switzerland RTX Healthy controls on MTX 11; 52+ 57+ 4.9 mo (GR, 3.7-5.6 mo) 73; Cohort Italy RTX and OFA Frequently relapsingly attencine dependent pediatric idiopathic method is good and control syndrome 11 19† 34 6-9 mo 9 Cohort USA RTX RRZ CL post-ASCT and consolidation RTX 15 71 48 6-9 mo 9 Phase 3 USA RTX B-cell NHL 15 71 48 Median, 9 mo (range, 10-82 mo) 93 mobitodic subcool RTX Type 1 DM; placebo 16; 20 37 44 wk 44 wk mobitodic spease controls who places con	3ar-Or et al, 2020 ⁶²	Phase 3	USA and Canada	OCR	Relapsing MS; Disease controls on IFN or no treatment	68; 34	40	99	10 wk	100; 100	24; 55	
Cohort Italy RTX and OFA Frequently relapsing/ steroicd dependent pediatric diopathic pediatric diopathic and of substitudy 11	Singhamet al, 2010 ⁶⁶	Phase 3	USA	RTX	Active RA; Disease controls on MTX	68; 32	20	78	22-26 wk		39; 42	
Cohort List RTX and OFA Frequently relapsing/steroid-dependent pediatric idiopathic nephrotic syndrome 11 19th 38th 36 mo (range, 10-82 mo) 9 Cohort USA RTX RVR BCL post-ASCT and consolidation RTX 22 51th 34th 6-9 mo Cohort USA RTX B-cell NHL 15 71 48 Median, 9 mo (range, 9.13 mo) 93 Inchese 3 substudy USA RTX RTX RRIX peral LDM; reated by with placebo 46; 20 20 37 44 wk 44 wk Incheses controls who in the controls who in the page of the	3ühler et al, 2019 ⁶⁷	Cohort	Switzerland	RTX		11; 253	52†	57†	4.9 mo (IQR, 3.7-5.6 mo)	73; 100	9;	
Cohort USA RTX R/R BCL post-ASCT and consolidation RTX 15 71 48 Median, 9 mo (range, post of ange) and range and range) and range and	colucci et al, 2019 ⁵³	Cohort	Italy	RTX and OFA	Frequently relapsing/ steroid- dependent pediatric idiopathic nephrotic syndrome	Ξ	194	33+	36 mo (range, 10-82 mo)	o	o o	
Cohort USA RTX B-cell NHL 15 71 48 Median, 9 mo (range, p. 13 mo) 93 Phase 3 substudy USA RTX Type 1 DM; Disease controls treated with placebo 46; 29 20 37 44 wk o Cohort France RTX Renal transplant; Prace outrols who places controls who places that the place of the place	lorwitz et al, 2004 ⁶⁸	Cohort	USA	RTX	R/R BCL post-ASCT and consolidation RTX	22	51+	34†	6-9 mo			64-68
Phase 3 bush but by substudy RTX Disease controls treated substudy 46; 29 20 37 44 wk o Cohort France RTX Renal transplant; Disease controls who pad not received RTX 13; 55 23 Median, 9 mo (IQR, 11.5 mo) RTX RTX RTX RTX RTX A-11.5 mo)	lustafa et al, 2020 ⁶⁹	Cohort	USA	RIX	B-cell NHL	15	71	48	Median, 9 mo (range, 1-24 mo; IQR, 9-13 mo)	86	7	
Cohort France RTX Renal transplant; 13; 55 23 Median, 9 mo (IQR, Disease controls who 26 26 4-11.5 mo) had not received RTX	escovitz et al, 2011 ⁷⁰	Phase 3 substudy	USA	RTX	Type 1 DM; Disease controls treated with placebo	46; 29	20	37	44 wk			67; 83
	uissant-Lubrano et al, 2010 ⁵⁰	Cohort	France	RTX	Renal transplant; Disease controls who had not received RTX	13; 26	වා	23	Median, 9 mo (IQR, 4-11.5 mo)	92 96		£ 6

1D, response at the end of 1 dose of a 1-dose series; 2D, response at the end of 2 doses of a 2-dose series; ARD, autoimmune rheumatic disease; ASCT, autologous stem cell transplant; CBT, cord blood transplant; CR, complete response; D1, dose 1 of a 2-dose series; D2, dose 2 of a 2-dose series; DM, diabetes mellitus; DMARD, disease-modifying anti-rheumatic drug; F, female; HBV, hepatitis B virus; IBD, inflammatory bowel disease; IFN, interferon; IDR, interferon; IDR, interferon; IDR, interferon; IDR, more reported; OCR, ocieliterange; ITP, immune thrombocytopenia; MMF, mycophenolate mofetil; MS, multiple sclerosis; MTX, methotrexate; NHL, non-Hodgkin lymphoma; NMOSD, neuromyelitis optica spectrum disorder; NR, not reported; OCR, ocielizamab; OCR, ocielizamab; RA, rheumatoid arthritis; R/R BCL, relapsed/refractory B-cell lymphoma; RTX, rituximab; OLE, systemic lupus erythematosus; SC, seroconversion; SR, seroresponse; T1DM, type 1 diabetes mellitus; UK, United Kingdom; USA, United States of America.

*Median age; where not reported, mean age reported. TRituximab-specific age and percentage female not reported, thus age and percentage female of entire study population reported.

Table 1. (continued)

ly	Design	Location	Anti-CD20 drug	Population/ Control population	z	Median age, y*	Sex, % F	Interval between anti- CD20 and vaccination	SP % dS	s % os	SR %
Cohort		USA	RTX	Postallogeneic CBT for heme malignancy, disease free 6 mo posttransplant; Disease controls who had not received RTX	13; 48	94	포	Median, 15 mo (range, 3-35 mo)		65 65	
Phase 1/2 substudy	⁴ /2	The Netherlands	RTX	Relapsed low-grade lymphoma	Ξ	53	N N	4 wk		25	
Cohort	Ti.	Switzerland	RTX	RA and vasculitis; Healthy controls	11; 253	52†	57+	Median, 4.9 mo (IQR, 3.7-5.6 mo)	64; 84-88	43	
Cohort	iort	USA	RTX	B-cell NHL	15	71	48	Median, 9 mo (range, 1-24 mo; IQR, 9-13 mo)	29	20	
Pha	Phase 3 substudy	USA	XIX	Type 1 DM; Type 1 DM treated with placebo	46; 29	20	37	44 wk			69; 76
Ŏ	Cohort	NSA	XTX	Postallogeneic CBT for heme malignancy, disease free 6 mo posttransplant; Disease controls who had not received RTX	13;	8	Z Z	Median, 15 mo (range, 3-35 mo)		75; 70	
•	Cohort	NSA	XTX	Postallogenetic CBT for heme malignancy, disease free 6 mo posttransplant; Disease controls who had not received RTX	13;	8	X X	Median, 15 mo (range, 3-35 mo)		60; 66	
Ü	Cohort	NSA	XIX	Post-ASCT RTX for NHL	17	45†	Z Z	Median, 31 mo in all (pre-transplant RTX and post)	0		0
O	Cohort	USA	RTX	R/R BCL post-ASCT and consolidation RTX	22	51+	34†	e-9 mo		70	D1: 73, D2: 77
Su	Phase 3 substudy	Canada	XTX	ITP; Disease controls treated with placebo	17; 7	40	7.1	9 шо		29; 83	
O	Cohort	USA	RTX	Postallogeneic CBT for heme malignancy, disease free 6 mo posttransplant; Disease controls who had not received RTX	13; 48	34	Z Z	Median, 15 mo (range, 3-35 mo)		85; 89	

response; D1, dose 1 of a 2-dose series; D2, dose 2 of a 2-dose series; DM, diabetes mellitus; DMARD, disease-modifying anti-rheumatic drug; F, female; HBV, hepatitis B virus; IBD, inflammatory bowel disease; IFN, interferon; IQR, interferon; IQR, multiple sclerosis; MTX, methotrexate; NHL, non-Hodgkin lymphoma; NMOSD, neuromyelitis optica spectrum disorder; NR, not reported; OCR, corelizumab; OR, encompetitis, R/R BCL, relapsed/refractory B-cell lymphoma; RTX, rituximab; SL, setroconversion; SP, seroprotection; SR, seroresponse; T1DM, type 1 diabe-1D, response at the end of 1 dose of a 1-dose series; 2D, response at the end of 2 doses of a 2-dose series; AIRD, autoimmune rheumatic disease; ASCT, autologous stem cell transplant; CBT, cord blood transplant; CR, complete

tes mellitus; UK, United Kingdom; USA, United States of America. *Median age; where not reported, mean age reported. †Rituximab-specific age and percentage female not reported, thus age and percentage female of entire study population reported.

Table 1. (continued)

Study	Design	Location	Anti-CD20 drug	Population/ Control population	z	Median age, y*	Sex, % F	Interval between anti- CD20 and vaccination)S % dS	SC % SR %	. 0
Hepatitis B											
Avivi et al, 2019 ⁷¹	Cohort	Israel	RTX	NHL who met criteria for "minimal B-cell recovery"; Healthy controls, ≤35 y; Healthy controls, ≥55 y	22; 8; 17	0	œ Z	38 mo (range, 14-56 mo)	64; 100; 59		
Jaffe et al, 2006 ⁵⁴	Cohort	USA	RTX	Postallotransplant; Postallotransplant who had not received RTX	25; 244	24†	38†	Median, 17 mo	56 5	56; 65	
Richi et al, 2020 ⁷²	Cohort	Spain	ZI X	Autoinmune inflammatory diseases (AIRD, psoriasis, or IBD); Healthy controls; Autoimmune inflammatory diseases on other biological DMARDs; Autoimmune inflammatory diseases on synthetic DMARDs	14; 49; 48	Φ	+19	^앞	9 9 9 9 9 9 9 9 9 9 9 9 9 9 9 9 9 9 9	29; 98; 94	
Hepatitis A virus											
Pescovitz et al, 2011 ⁷⁰	Phase 3 substudy	NSA	RTX	Type 1 DM; T1DM treated with placebo	46; 29	20	37	44 wk	47; 67		
van der Kolk et al, 2002 ³⁶	Phase 1/2 substudy	The Netherlands	RTX	Relapsed low-grade lymphoma	Ξ	53	N N	4 wk	0		
Polio virus											
Jaffe et al, 2006 ⁵⁴	Cohort	USA	RTX	Postallotransplant; Postallotransplant who had not received RTX	25; 244	24+	38+	NR; HBV in trial given after median 16.6 mo		91;	
Shah et al, 2015 ⁴⁶	Cohort	USA	RTX	Postallogeneic CBT for heme malignancy, disease free 6 mo posttransplant; Disease controls who had not received RTX	13;	34	œ Z	Median, 15 mo (range, 3-35 mo)		62; 80	
van der Kolk et al, 2002 ³⁶	Phase 1/2 substudy	The Netherlands	RTX	Relapsed low-grade lymphoma	Ξ	23	Z Z	4 wk		20	
Pneumococcal conjugate (PCV-7)											
Kapetanovic et al, 2013 ⁷³	Cohort	Sweden	XTX	RA; Disease controls on abatacept; Disease controls on tocilizumab; Disease controls on	55; 17; 16; 85;	65	67	86 d (range, 0-894 d)		5; 18; 50; 21; 48	1
1D. response at the end of 1 dose of a 1-dose series: 9D, response at the	of 1 dose of a	1-dose series: 2D.	response at the	e end of 2 doses of a 2-dose s	eries: AIRD autoir	nmine rheimat	ic dispase.	e end of 2 doses of a 2-dose series: AIRD autoimmune rheumatic disease: ASCT autologous stem cell transplant: CBT cord blood transplant: CR. complete	plant: CBT, cord blood trail	neplant: CR com	nlete

1D, response at the end of 1 dose of a 1-dose series; 2D, response at the end of 2 doses of a 2-dose series; ABC, autoimmune rheumatic disease; ASCT, autologous stem cell transplant; CBT, cord blood transplant; CR, complete response; D1, dose 1 of a 2-dose series; D2, dose 2 of a 2-dose series; DM, diabetes mellitus; DMARD, disease-modifying anti-rheumatic drug; F, female; HBV, hepatitis B virus; IBD, inflammatory bowel disease; IFN, interferon; IGR, interquartile range; IFP, immune thrombocytopenia; MMF, mycophenolate mofetil; MS, meltiple sclerosis; MTX, methotrexate; NHL, non-Hodgkin lymphoma; NMOSD, neuromyelitis optica spectrum disorder; NR, not reported; OCR, correlizands) CA, of ottaminab; RR BCL, relapsed/refractory B-cell lymphoma; RTX, rituximab; SLE, systemic lupus erythematosus; SC, seroconversion; SR, seroresponse; T1DM, type 1 diabemane and percentage female not reported.

*Median age; where not reported, mean age reported.

#Rituximab-specific age and percentage female of entire study population reported.

Table 1. (continued)

(505)											
Study	Design	Location	Anti-CD20 drug	Population/ Control population	z	Median age, y*	Sex, % F	Interval between anti- CD20 and vaccination	% dS	% 2S	SR %
				MTX; Alternate disease controls with spondyloarthropathy on anti-inflammatories and analgesics							
T-cell-independent vaccinations											
Pneumococcal polysaccharide (PCV-23)											
Albert et al, 2008 ⁶⁵	Cohort	USA	RTX	SLE	4	43	63	7 mo			29
Bar-Or et al, 2020 ⁶²	Phase 3	USA and Canada	OCR	Relapsing MS; Disease controls: relapsing MS on IFN or no treatment	68; 34	40	99	14 wk			37% to ≥ 12 serotypes; 97% to ≥ 12 serotypes
Berglund et al, 2014 ³⁰	Cohort	Sweden	RTX	Lymphoma on treatment; Disease controls with cancer on active treatment other than RTX	13; 83	63+	29+	All patients on active treatment	25; 68	0;	
Bingham et al, 2010 ⁶⁶	Phase 3	USA	XTX	Active RA; Disease controls: active RA on MTX alone	68; 32	50	78	22-26 wk			19% to ≥6 serotypes; 61% to ≥6 serotypes
Horwitz et al, 2004 ⁶⁸	Cohort	USA	RTX	R/R BCL post-ASCT and consolidation RTX	22	51+	34†	6-9 mo	41		
Mustafa et al, 2020 ⁶⁹	Cohort	USA	RTX	B-cell NHL	15	71	48	Median, 9 mo (range, 1-24 mo; IQR, 9-13 mo)	33% to ≥7 serotypes	20	
Nazi et al, 2013 ³⁸	Phase 3 substudy	Canada	RTX	ITP ITP treated with placebo	17; 7	40	7.1	6 mo		21; 67	

1D, response at the end of 1 dose of a 1-dose series; 2D, response at the end of 2 doses of a 2-dose series; ARD, autoimmune rheumatic disease; ASCT, autologous stem cell transplant; CBT, cord blood transplant; CR, complete response; D1, dose 1 of a 2-dose series; D2, dose 2 of a 2-dose series; DM, diabetes mellitus; DMARD, disease-modifying anti-rheumatic drug; F, female; HBV, hepatitis B virus; IBD, inflammatory bowel disease; IFN, interferon; IDR, interperon; IDR, interperon; IDR, mycophenolate mofetil; MS, multiple scherosis; MTX, methotrexate; NHL, non-Hodgkin lymphoma; NMOSD, neuromyelitis optica spectrum disorder; NR, not reported; OCR, correlizance of America.

Median age; When most reported; Assert of America.

Median age; where not reported, mean age reported.

**Redian age; where not reported, mean age and percentage female of entire study population reported.

Table 2. Literature summary: cellular responses

, ,				
Measurement of T-cell response and outcomes	Measurement: Flow cytometry for activated CD4 T cells, influenza-specific cells were defined as the percentage of CD69'/IFN-γ-producing cells within the total CD4* T-cell population in the antigen-stimulated cultures subtracted by those in the negative control cultures. Outcome: No difference in percentage increase in influenza-specific CD4* subsets in patients treated with anti-CD20 therapy compared with DMARD-treated patients (median, 0.1% to 0.3% and 0.1% to 0.2%, respectively); healthy controls had figher baseline influenza-specific CD4* T cells, but their CD4* subsets decreased by half postvaccination (0.6% to 0.3%)	Measurement: Effector function of antigenspecific CD8 ⁺ and CD4 ⁺ T cells assessed by intracellular-cytokine staining for IFN-γ and TNF-α; response considered positive if combined percentage of H1N1-specific TNF-α plus IFN-γ-producing CD4 ⁺ or CD8 ⁺ T cells was twofold or higher compared with background level (nonstinulated PBMCs) and if there was a minimum of 0.05% H1N1-specific TNF-α plus IFN-γ-producing CD4 ⁺ or CD8 ⁺ T cells (after subtracting background) Outcome: Cellular data only available for all B-cell malignancy patients together (not RTX specific); 7% pre- and 36% postvaccine had H1N1-specific T-cell response; no effect of prior chemotherapy, halthy controls had 44% pre and 48% post, no effect of prior chemotherapy on induction of H1N1-specific T cells after 2 doses of vaccine (data NR)	Measurement: T-cell ELISPOTs were performed using a standard γ-IFN T-cell ELISPOT assay Outcome: T-cell responses similar in anti-CD20-treated patients at baseline compared with healthy controls, but no significant increases after vaccination, no increase in proliferation (data NR); no changes in the T-cell repertoire as detected by spectratyping over time (data NR)	Measurement: Presence of IFN-y-producing 2009 H1N1 influenza virus-specific CD4 ⁺ and CD8 ⁺ T cells Outcomes: After first vaccination, virus-specific CD4 ⁺ and CD8 ⁺ T-cell responses were significantly lower in patients with low B cells than those with normal B cells; booster vaccination stimulated the antiviral T-cell response only in patients with low B cells
Vaccine type	Seasonal	Pandemic influenza	Seasonal	Pandemic influenza
Interval between anti-CD20 and vaccination	16 patients: <5 mo; 13 patients: >5 mo	19 mo (range, 2-83 mo)	7-9 mo	Median, 6 mo (range, 1-36 mo)
Sex, % F	79	404	0 4	88
Median age, y*	62	+ 449	6	45
z	59	20	7	9
Population	RA A	B-cell malignancies	Rheumatologic disease	Autoimmune rheumatic diseases
Anti-CD20 drug	XTA	አ አ	X,	XTX
Location	Israel	United Kingdom	United States	Switzerland
Design	Cohort	Cohort	Cohort	Cohort
Study, year	Arad et al, 2011 ³⁹	de Lavallade et al, 2011 ⁴⁰	Eisenberg et al, 2013 ⁴²	Muller et al, 2013 ⁴³

DMARD, disease-modifying antirheumatic drug; ELISPOT, enzyme-linked immune absorbent spot; GMFR, geometric mean fold rise; PBMC, peripheral blood mononuclear cell; TNF, tumor necrosis factor; VZV, varicella zoster virus. See Table 1 for expansion of other abbreviations.

*Median age; where not reported, mean age reported.

*Rituximab-specific age and percentage female not reported, thus age and percentage female of entire study population reported.

٠	ζ	
	Ó	Ĺ
	1	
	c	
:	:	
Ī	Ċ	
	7	
	Č	
•	٠	
		į
	(l
Ċ	Ć	

dy, year	Design	Location	Anti-CD20 drug	Population	z	Median age, y*	Sex, % F	Interval between anti-CD20 and vaccination	Vaccine type	Measurement of T-cell response and outcomes
azi et al, 2013 ³⁸	Phase 3 substudy	Canada	RTX	₫.	17	04	12	e m	Tetanus toxoid	Measurement: IFN-y ELISPOT assay to measure frequency of tetanus toxoid-specific T cells Outcomes: Mean number of IFN-y-producing T cells was reduced in patients who had received rituximab compared with placebo at 1 wk (38 cells vs 93 cells per 5 × 10° total cells) and 1 mo (14 cells vs 43 cells per 5 × 10° total cells) total cells)
arino et al, 2017 ⁴¹	Phase 1	Worldwide	RTX	B-cell lymphoma	80	19	54	On active treatment	Inactivated VZV	Measurement: IFN-y ELISPOT assay was used to detect the presence of IFN-y-secreting VZV-specific cells from PBMCs before and after immunization Outcome: GMFR met primary outcome at 4.34 (95% Cl, 3.01-6.24)

DMARD, disease-modifying antirheumatic drug; ELISPOT, enzyme-linked immune absorbent spot; GMFR, geometric mean fold rise; PBMC, peripheral blood mononuclear cell; TNF, tumor necrosis factor; VZV, varicella zoster virus. reported, thus age and percentage female of entire study population reported reported, mean age regand percentage female See Table 1 for expansion of other abbreviations. *Median age; where not

patients >12 months from therapy, though this did not always translate to an equivalent response to controls.

Based on the descriptive nature of the data, ascertaining whether patients had better responses to T-cell-dependent vaccinations than T-cell-independent vaccinations is difficult.

Meta-analysis of humoral response to pandemic influenza vaccine. Thirteen studies (222 anti-CD20 treated patients and 993 control patients) reported on response rates to the pandemic influenza vaccine. Ten of the 13 studies included patients with hematologic malignancies (rather than autoimmune disease) treated with anti-CD20 therapy. The pooled estimates of SC for patients treated with anti-CD20 therapy are listed in Table 4. Patients on active (<3 months since last dose) anti-CD20 therapy had poor response to vaccination, with a pooled estimate of SC after 1 vaccine dose of 3% (95% CI, 0% to 9%; 6 studies, 75 patients). This improved incrementally over time with pooled estimates of 21% (95% CI, 0% to 65%), 50% (95% CI, 4% to 96%), and 41% (95% Cl, 19% to 65%) within 3 to 6 months, 6 to 12 months, or >12 months from last dose of anti-CD20 therapy, respectively, though the sample size was limited (see Table 4).

The pooled RBs for SC of patients treated with anti-CD20 therapy compared with healthy controls and compared with disease controls are listed in Table 5 and visualized in Figures 2 and 3. Patients on active anti-CD20 therapy had significantly lower SC rates after 1 dose of the pandemic influenza vaccine compared with healthy controls (RB, 0.05 [95% CI, 0-0.73]) and compared with disease controls (RB, 0.22 [95% Cl, 0.09-0.56]). The results did not change significantly even when limiting only to studies of patients with hematologic malignancy (Table 5).

Cellular responses to vaccination. Six studies reporting cellular response data on 171 patients treated with anti-CD20 therapy were included. Descriptive data are available in Table 2. Notably, all but 1 study³⁸ included patients who were receiving concomitant immunosuppressive therapy that may have impaired T-cell function (eg, mycophenolate, cyclosporine). Three of the included studies found no significant difference in T-cell vaccine response between anti-CD20-treated patients vs healthy or disease controls. 39-41 In these studies, the average interval between anti-CD20 and vaccination ranged from being on active treatment to 19 months. The other 3 studies found that patients treated with anti-CD20 had lower T-cell responses compared with healthy or disease controls, or to patients with normal B-cell subsets. 38,42,43 The average interval between anti-CD20 therapy and vaccination in these studies was 6 to 9 months.

Safety of vaccination in patients with hematologic malignancy. Of the 19 studies reporting data on patients with hematologic malignancies, 9 reported data on adverse events related to vaccination. ^{30,31,33,37,40,41,44-47} The majority of studies found that adverse reactions were mild, and 1 study noted that there were no differences in side effect profiles in patients compared with healthy controls.40 Only 1 study reported serious adverse events (SAEs): a phase 1 study of a novel inactivated varicella zoster vaccine schedule reported 10 SAEs related to vaccination among the 80 patients studied,41 with the most common SAEs being febrile neutropenia and pneumonia.

Table 3. Summarized data from all studies listed in Table 1, by average duration of time since last anti-CD20 therapy

Duration of time since anti-CD20 therapy	Summarized SP rate, %	Summarized SC rate, %	Summarized SR rate, %
Pandemic influenza (N = 13 studies, 222 patients treated with anti-CD20 therapy)*			
Active treatment, <3 mo	0-33 ^{30-34,40,61}	0-25 ^{30-34,40}	
3-6 mo	19 ⁶⁰	38 ⁶⁰	38-44 ⁴³
6-12 mo	50 ⁴⁰	50 ⁴⁰	
>12 mo	66-100 ^{40,44}	66-100 ^{40,44}	
Seasonal influenza (N = 12 studies, 252 patients treated with anti-CD20 therapy)†			
Active treatment, <3 mo	8-25 ^{30,64}	030,35	
3-6 mo	56-80 ⁶²	10-60 ⁶²	
6-12 mo	69-77 ⁶³	25 ³⁵	17 ⁴²
>12 mo	23-79 ^{44,52}	3-43 ^{44,52}	
Tetanus, diphtheria, and pertussis (12 studies, 309 patients treated with anti-CD20 therapy)‡			
Active treatment, <3 mo	100 ⁶²	24-25 ^{36,62}	
3-6 mo	64-73 ⁶⁷	9-39 ^{66,67}	
6-12 mo	66-93 ^{50,69}	7-20 ⁶⁹	31-69 ^{50,65,68,70}
>12 mo	0-9 ^{47,53}	9-75 ^{46,53}	0 ⁴⁷
Haemophilus influenzae B (3 studies, 52 patients treated with anti-CD20 therapy)			
Active treatment, <3 mo			
3-6 mo			
6-12 mo		29 ³⁸	73-77 after 1-2 doses ⁶⁸
>12 mo		85 ⁴⁶	
Hepatitis B (3 studies, 61 patients treated with anti-CD20 therapy)§			
Active treatment, <3 mo			
3-6 mo			
6-12 mo			
>12 mo	56-64 ^{54,71}	56 ⁵⁴	
Hepatitis A (2 studies, 57 patients treated with anti-CD20 therapy)			
Active therapy, <3 mo	0 ⁴⁷		
3-6 mo			
6-12 mo	47 (slightly lower than disease controls of 67) ⁷⁰		
>12 mo			
Polio virus (3 studies, 49 patients treated with anti-CD20 therapy)			
Active treatment, <3 mo			20 ³⁶
3-6 mo			
6-12 mo			
>12 mo			62, slightly lower the in disease controls (80) ⁴⁶

^{*}See Tables 4 and 5 for meta-analyzed data. In all studies, SP, SC, and/or SR rates were similar or lower in anti-CD20-treated patients than healthy controls or disease controls. One study did not report the average duration since last dose of anti-CD20 therapy but had SP and SR rates of 0% and 0% to 9%, respectively. 45 One study indicated that patients had received anti-CD20 therapy <12 months ago but did not specify the average time interval, and reported a SP rate of 9%. ⁵⁶

The all studies, SP, SC, and/or SR rates were lower in anti-CD20-treated patients than in healthy controls or disease controls.

[‡]In all studies, SP, SC, and/or SR rates were similar or lower in anti-CD20-treated patients than healthy controls or disease controls. Other than in 2 studies^{47,53} (a study of patients post-autologous stem cell transplant plus rituximab for non-Hodgkin lymphoma, and a study of pediatric patients treated with rituximab for relapsing/steroid-dependent idiopathic

nephrotic syndrome) where the SP rates were only 0% to 9%, SP rates were moderate to high regardless of the duration that had elapsed from anti-CD20 therapy. §One study did not report the interval after anti-CD20 therapy during which patients were vaccinated.⁷² Overall values for anti-CD20-treated patients were similar to healthy controls ≥55 years and disease controls, but worse than healthy controls ≤35 years.

^{||}One study did not report the interval after anti-CD20 therapy during which patients received vaccination but had a SR rate of 91%.54

[¶]All studies showed that SP, SC, and/or SR rates were lower in anti-CD20-treated patients compared with disease controls.

Duration of time since anti-CD20 therapy	Summarized SP rate, %	Summarized SC rate, %	Summarized SR rate, %
Pneumococcal conjugate (PCV-7) (1 study, 55 patients treated with anti-CD20 therapy)			
Active treatment, <3 mo			5, compared with 21- 50 in disease controls ⁷³
3-6 mo			
6-12 mo			
>12 mo			
Pneumococcal polysaccharide (PCV-23) (7 studies, 217 patients treated with anti-CD20 therapy)¶			
Active treatment, <3 mo	25 ³⁰	030	
3-6 mo			19 to ≥6 serotypes ⁶⁶
6-12 mo	33 (to \geq 7 serotypes) to 41 68	20-21 ^{38,69}	29 ⁶⁵
>12 mo			

^{*}See Tables 4 and 5 for meta-analyzed data. In all studies, SP, SC, and/or SR rates were similar or lower in anti-CD20-treated patients than healthy controls or disease controls. One study did not report the average duration since last dose of anti-CD20 therapy but had SP and SR rates of 0% and 0% to 9%, respectively. 45 One study indicated that patients had received anti-CD20 therapy <12 months ago but did not specify the average time interval, and reported a SP rate of 9%.

Discussion

There are over 900 000 people living with lymphoma in the United States alone, 48 and anti-CD20 therapies have been a component of first-line therapy for many lymphoproliferative disorders for over 20 years. Furthermore, anti-CD20 therapies are an important treatment modality for numerous autoimmune conditions, including multiple sclerosis and rheumatoid arthritis, where their use is widespread. Understanding vaccine responsiveness in the context of B-cell-depleting therapies, especially given the number of patients that will need vaccination against COVID-19, is imperative.

To interpret vaccine responses (SP, SC, and/or SR rates), it is important to note that there are 2 types of vaccine antigens. A primary antigen is an antigen that the patient has not been previously exposed to, which elicits an extrafollicular response that causes B-cell proliferation and differentiation into plasma cells, and generally reaches a peak value 4 weeks after immunization. ⁴⁹ A recall antigen is one in which an anamnestic immune response is anticipated (eg, tetanus booster vaccination), in as little as 7 days.49 For recall

antigens, SP and SC rates postvaccination are expected to be high. From that perspective, we chose to focus our meta-analysis on the pandemic influenza vaccine from the 2009 H1N1 pandemic, as this vaccine was a primary antigen, analogous to the COVID-19 vaccine. As patients would have no preexisting memory B cells to this antigen, this allows the best assessment of B-cell-depleted patients' immune response to vaccination. Additionally, the pandemic influenza vaccine has standardized definitions of SP and SC. The SC rate was chosen as the primary outcome to meta-analyze as SC demonstrates the ability to mount a response to vaccination (by either a several-fold increase in antibody titers, or conversion from negative titers to protective titers). SP rates can be adequate in patients who have had previous exposure to an antigen, even if they do not respond to the vaccination given.

Although the literature was heterogenous in terms of patient population, vaccine types, and interval since anti-CD20 therapy, we documented 4 main findings: (1) vaccination appears safe in patients on anti-CD20 therapy; (2) the response to vaccination in patients on active anti-CD20 therapy is low and approaches 0%; (3) anti-CD20

Table 4. Pooled estimates of SC to 1 or 2 doses of the pandemic influenza vaccine in patients treated with anti-CD20 therapy

			Pooled estimate of S	iC, % (95% CI, I ² %)
Duration of time since last anti-CD20 treatment, mo	No. of studies	Total no. of patients	For 1 vaccine dose	For 2 vaccine doses
Active, <3	6	75	3 (0-9, 0)	12 (2-27, 37)
3-6	2	21	21 (0-65, 71)	4 (0-32, 0)
6-12	1	12	50 (4-96, N/A)	50 (4-96, N/A)
>12	2	26	41 (19-65, 0)	75 (39-100, 41)

N/A, not applicable.

tln all studies, SP, SC, and/or SR rates were lower in anti-CD20-treated patients than in healthy controls or disease controls.

[#]In all studies, SP, SC, and/or SR rates were similar or lower in anti-CD20-treated patients than healthy controls or disease controls. Other than in 2 studies^{47,53} (a study of patients post-autologous stem cell transplant plus rituximab for non-Hodgkin lymphoma, and a study of pediatric patients treated with rituximab for relapsing/steroid-dependent idiopathic nephrotic syndrome) where the SP rates were only 0% to 9%, SP rates were moderate to high regardless of the duration that had elapsed from anti-CD20 therapy.

[§]One study did not report the interval after anti-CD20 therapy during which patients were vaccinated.72 Overall values for anti-CD20-treated patients were similar to healthy controls ≥55 years and disease controls, but worse than healthy controls ≤35 years.

^{||}One study did not report the interval after anti-CD20 therapy during which patients received vaccination but had a SR rate of 91%.54

[¶]All studies showed that SP, SC, and/or SR rates were lower in anti-CD20-treated patients compared with disease controls.

Table 5. Relative benefit ratios for SC to the pandemic influenza vaccine in healthy controls and disease controls compared with patients on anti-CD20 therapy

Average duration of time since anti-CD20 therapy, mo No. of studies analyzed	No. of studies analyzed	1 dose (95% CI)	l² %, P	2 doses (95% CI)	l² %, P	$I^2 \ \%$, P No. of studies analyzed	1 dose (95% CI)	l² %, P	2 doses (95% CI)	l² %, P
	All patients	All patients on anti-CD20 therapy vs healthy controls	erapy vs hea	Ithy controls		All patie	All patients on anti-CD20 therapy vs disease controls	erapy vs dise	ase controls	
On active treatment, <3	-	0.05 (0-0.73)	N/A, 0.03	ı	ı	9	0.22 (0.09-0.56)	3, .001	0.25 (0.12-0.55)	0, .0006
3-6	-	0.50 (0.24-1.06) N/A, 0.07	N/A, 0.07	1	1	2	0.44 (0.23-0.84)	0, .01	0.10 (0.01-1.49)	N/A, .10
6-12	I	I	I	I	I	-	0.77 (0.28-2.10)	N/A, .61	0.62 (0.23-1.67)	N/A, .35
>12	-	0.50 (0.23-1.09)	N/A, 0.08	0.50 (0.23-1.09) N/A, 0.08 0.82 (0.51-1.32) N/A, 0.41	N/A, 0.41	-	1.03 (0.45-2.35)	N/A, .94	1.10 (0.74-1.63)	N/A, .65
						Only patients with hematologic malignancy on anti-CD20 therapy vs disease controls	atologic malignancy	on anti-CD20	therapy vs disease	controls
On active treatment, <3						വ	0.19 (0.06-0.61)	21, .005	0.22 (0.09-0.52)	0, .0007
3-6						-	0.13 (0.01-1.85)	N/A, .13	0.10 (0.01-1.49)	N/A, .10
6-12						-	0.77 (0.28-2.10)	N/A, .61	0.62 (0.23-1.67)	N/A, .35
>12						-	1.03 (0.45-2.35)	N/A, .94	1.10 (0.74-1.63)	N/A, .65

RB is the ratio of SC rate in patients treated with anti-CD20 therapy to the SC rate in healthy or disease controls. Bold values represent statistically significant relative benefit ratios not applicable. therapy lowers patients' vaccine response beyond the impact of their disease or other treatments, given the higher response to vaccination in disease controls; and (4) response to vaccination improves incrementally over time but may not reach the level of healthy controls even 12 months after therapy. Although not the primary aim of our study, we also noted that patients with lymphoproliferative disorders not exposed to anti-CD20 therapy (on active cancer therapy, previous non-anti-CD20 treatment, or never treated) had reasonable responses to vaccination (eg, SC rates 27% to 80% to pandemic influenza vaccine, see Tables 1 and 2).

It is notable that SC rates in studies of patients on active anti-CD20 therapy were 0% to 25%, 30-37 regardless of the vaccine type assessed. A small number of studies reported that vaccine responses did not vary over time. However, these studies may have been underpowered to detect a time trend, as our meta-analysis for pandemic influenza vaccine found that SC rates did improve as time between last anti-CD20 treatment and vaccination increased.

Whether 2 vaccine doses enhance immunogenicity in patients receiving anti-CD20 therapy remains unclear. When assessing the response to pandemic influenza vaccination, the point estimate of SC in patients on active therapy was just 3% (95% Cl, 0% to 9%) after 1 dose, and 12% (95% Cl, 2% to 27%) after 2 doses. Five studies assessed SC after 1 and 2 doses of pandemic influenza vaccine in patients receiving anti-CD20 vs disease controls. As illustrated in Tables 4 and 5, the point estimate of SC does not appear appreciably different in those receiving 1 or 2 doses of vaccine, nor do patients on anti-CD20 seem to improve in their SC rates compared with disease controls. This finding is unchanged when limiting the analysis only to patients with hematologic malignancies. However, due to the small sample sizes of included studies, the relative impact or lack of impact of booster vaccinations remains essentially unknown.

Our third finding was that the reduction in response to vaccination seen in patients on anti-CD20 therapy is likely above and beyond the immunocompromising effects of the disease alone and the other treatments used in the disease. The SC rates to the pandemic influenza vaccination in patients on active anti-CD20 therapy or even 3 to 6 months from therapy were significantly lower than in disease controls (RBs of 0.22 [95% Cl, 0.09-0.56] and 0.44 [95% Cl, 0.23-0.84], respectively). However, by an average of 6 to 12 months from anti-CD20 therapy, the difference between groups was less pronounced, and by >12 months the difference seemed trivial, though we are limited by the number of studies available for analysis.

Given the finding that anti-CD20 therapy abrogates vaccine responses, one strategy may be to delay anti-CD20 therapy (when possible) until after vaccination. However, currently we do not know how long of a gap is required between vaccination and anti-CD20 treatment of vaccination to be effective. One small study of patients with relapsed low-grade lymphoma³⁶ assessed this strategy: 5 patients were given a primary antigen vaccine (hepatitis A) 2 weeks prior to starting on rituximab treatment. None of patients developed a humoral response. On the other hand, 5 of 10 patients immunized 2 weeks before rituximab had a response to a recall antigen (polio virus or tetanus toxoid). These results highlight the need for further research to determine the optimal gap between vaccination, particularly of a novel primary vaccine, and anti-CD20 therapy. In the absence of data, clinicians and patients need to weigh the expected benefits of initiating anti-CD20 therapy against its anticipated impact

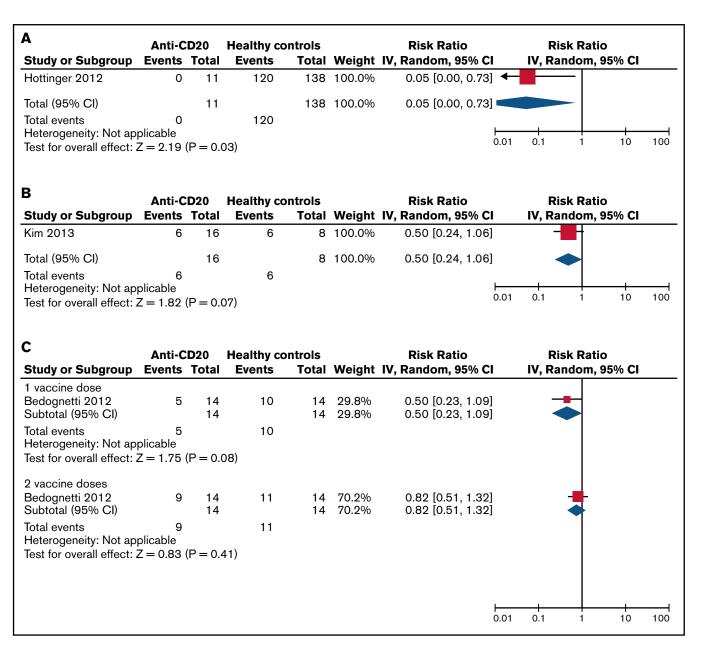


Figure 2. Risk benefit of SC to the pandemic influenza vaccine in anti-CD20-treated patients vs healthy controls. (A) On active therapy (<3 months from therapy). (B) Three to 6 months from therapy. (C) More than 12 months from therapy. Events, the number of patients with SC response to vaccination; Total, the total number of patients assessed for SC response to vaccination; Weight, the weighted contribution of each study to the meta-analysis.

on vaccine response. In rare instances, it may be appropriate to delay anti-CD20 therapy to allow for COVID-19 vaccination.

Finally, the response to vaccination does seem to improve incrementally over time as the B-cell compartment recovers following rituximab treatment. Generally, the effect of anti-CD20 therapy was least pronounced >1 year after treatment, although even these patients had impaired responses compared with healthy controls, with SC rates of 33% to 100% for pandemic infuenza, 40,44 3% to 43% for seasonal influenza, 44,52 9% to 75% for tetanus, diphtheria, and pertussis, 46,53 85% for haemophilus influenzae type b, 46 and 56% for hepatitis B.54 The finding was also noted in correlative analyses in a study by de Lavallade, 40 which found that the interval between therapy and vaccination was significantly longer in patients who were seroprotected compared with those who were not (4.7 vs 17.5 months; P = .001). Whether the degree of B-cell recovery is associated with response remains unclear, as this is not generally measured in clinical practice, and thus was not a focus of this review.

Notably, we should be cautious applying immunological correlates of protection (which have only been validated in young healthy adults⁵⁵) to immunocompromised patients.⁵⁶ Whether an incremental rise in titers not meeting criteria for SP or SC might still be valuable in immunocompromised patients to guard against clinical infection remains unclear.

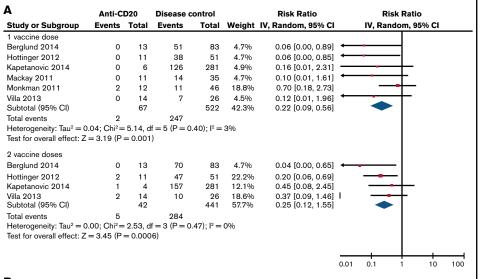
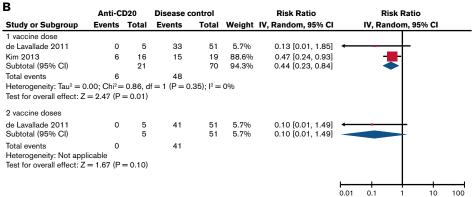


Figure 3. Risk ratios of SC to the pandemic influenza vaccine when treated with either 1 or 2 doses when comparing anti-CD20-treated patients to disease controls. (A) On active therapy (<3 months from therapy). (B) Three to 6 months from therapy. (C) Six to 12 months from therapy. (D) More than 12 months from therapy. Events, the number of patients with SC response to vaccination; Total, the total number of patients assessed for SC response to vaccination; Weight, the weighted contribution of each study to the meta-analysis.



Study or Subgroup Events Total Events Total Weight IV, Random, 95% Cl IV, Random, 95% Cl	С	Anti-C	D20	Disease o	ontrol		Risk Ratio	Risk Ratio
de Lavallade 2011 2 4 33 51 49.4% 0.77 [0.28, 2.10] Subtotal (95% CI) 4 51 49.4% 0.77 [0.28, 2.10] Total events 2 33 Heterogeneity: Not applicable Test for overall effect: Z = 0.50 (P = 0.61) 2 vaccine doses de Lavallade 2011 2 4 41 51 50.6% 0.62 [0.23, 1.67] Subtotal (95% CI) 4 51 50.6% 0.62 [0.23, 1.67] Total events 2 41 Heterogeneity: Not applicable	Study or Subgroup	Events	Total	Events	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
Subtotal (95% CI) 4 51 49.4% 0.77 [0.28, 2.10] Total events 2 33 Heterogeneity: Not applicable Test for overall effect: Z = 0.50 (P = 0.61) 2 vaccine doses de Lavallade 2011 2 4 41 51 50.6% 0.62 [0.23, 1.67] Subtotal (95% CI) 4 51 50.6% 0.62 [0.23, 1.67] Total events 2 41 Heterogeneity: Not applicable	1 vaccine dose							
Total events 2 33 Heterogeneity: Not applicable Test for overall effect: Z = 0.50 (P = 0.61) 2 vaccine doses de Lavallade 2011 2 4 41 51 50.6% 0.62 [0.23, 1.67] Subtotal (95% Cl) 4 51 50.6% 0.62 [0.23, 1.67] Total events 2 41 Heterogeneity: Not applicable	de Lavallade 2011	2	4	33	51	49.4%	0.77 [0.28, 2.10]	
Heterogeneity: Not applicable Test for overall effect: Z = 0.50 (P = 0.61) 2 vaccine doses de Lavallade 2011	Subtotal (95% CI)		4		51	49.4%	0.77 [0.28, 2.10]	-
Test for overall effect: Z = 0.50 (P = 0.61) 2 vaccine doses de Lavallade 2011	Total events	2		33				
2 vaccine doses de Lavallade 2011 2 4 41 51 50.6% 0.62 [0.23, 1.67] Subtotal (95% CI) 4 51 50.6% 0.62 [0.23, 1.67] Total events 2 41 Heterogeneity: Not applicable	Heterogeneity: Not appli	icable						
de Lavallade 2011 2 4 41 51 50.6% 0.62 [0.23, 1.67] Subtotal (95% CI) 4 51 50.6% 0.62 [0.23, 1.67] Total events 2 41 Heterogeneity: Not applicable	Test for overall effect: Z	= 0.50 (P	= 0.61)					
Subtotal (95% CI) 4 51 50.6% 0.62 [0.23, 1.67] Total events 2 41 Heterogeneity: Not applicable	2 vaccine doses							
Total events 2 41 Heterogeneity: Not applicable	de Lavallade 2011	2	4	41	51	50.6%	0.62 [0.23, 1.67]	
Heterogeneity: Not applicable	Subtotal (95% CI)		4		51	50.6%	0.62 [0.23, 1.67]	
	Total events	2		41				
	Heterogeneity: Not appli	icable						
			= 0.35)					
							Ļ	01 01 1 10

Anti-Cl	D20	Disease c	ontrol		Risk Ratio	Risk Ratio
Events	Total	Events	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
2	3	33	51	18.6%	1.03 [0.45, 2.35]	
	3		51	18.6%	1.03 [0.45, 2.35]	•
2		33				
icable						
	= 0.94)					
3	3	41	51	81.4%	1.10 [0.74, 1.63]	•
	3		51	81.4%	1.10 [0.74, 1.63]	▼
3		41				
icable						
	= 0.65)					
	,					
						0.01 0.1 1 10 100
	2 icable = 0.07 (P = 3 icable	3 2 icable = 0.07 (P = 0.94) 3 3 3 3	2 3 33 33 33 33 34 3 3	Events Total Events Total 2 3 33 51 2 33 51 2 icable 33 41 51 3 3 41 51 3 41 51 51 3 icable 41 51 51	Events Total Events Total Weight 2 3 33 51 18.6% 2 33 51 18.6% 3 33 41 51 81.4% 3 3 41 51 81.4% 3 41 51 81.4% 3 41 51 81.4%	Events Total Events Total Weight IV, Random, 95% CI 2 3 33 51 18.6% 1.03 [0.45, 2.35] 2 33 51 18.6% 1.03 [0.45, 2.35] icable 0.07 (P = 0.94) 0.07 (P = 0.94) 0.07 (P = 0.94) 3 3 41 51 81.4% 1.10 [0.74, 1.63] 3 41 51 81.4% 1.10 [0.74, 1.63] 3 41 51 81.4% 1.10 [0.74, 1.63]

Additionally, cellular responses are also important in vaccinationmediated protection,⁵⁷ and although these are also affected by anti-CD20 therapy, the degree to which cellular immunity is impacted is less clear.35 Studies were small and contradictory in their findings with respect to whether T-cell immunity was preserved or diminished following anti-CD20 therapy. Some of the studies were also confounded by the inclusion of patients who were on concomitant immunosuppressive therapies that impair T-cell function (eg, mycophenolate).

There are several limitations to our work. First, the pool of studies included in this systematic review was heterogenous with respect to patient population, vaccine studied, definitions of SP and SC, and interval between anti-CD20 treatment and vaccination. Although precise estimates of interval between anti-CD20 and vaccination were available for some studies, others only provided median/mean intervals and ranges. Definitions of SP and SC varied between studies, which was expected as there are no widely used definitions for SP and SC other than for pandemic influenza vaccinations. We sought to limit the heterogeneity encountered as much as possible by categorizing studies both by vaccine studied and by average interval between anti-CD20 and vaccination. We also only metaanalyzed data related to the pandemic influenza vaccine, which has standard definitions for SP and SC. Nonetheless, heterogeneity in interval from anti-CD20 therapy does remain, and the most accurate interpretation of amalgamated data only applies to patients on active treatment as only these studies had homogenous populations. Additionally, the sample sizes of each study were small, so direct extrapolation of SP. SC. and/or SR from any 1 study should be avoided. Finally, we did not have data on baseline hypogammaglobulinemia for all studies, and whether concomitant immunoglobulin replacement therapy was used for patients, as this can provide an exogenous source of SP detected in laboratory studies. We also did not collect data on other concomitant immunomodulatory medications used for patients and the timing of their use with respect to vaccination, which may also have impacted their immune responses.

This systematic review highlights the available literature on both humoral and cellular vaccination response in patients treated with anti-CD20 therapy. We found limited and heterogenous data, suggesting the need for a robust prospective study in a large sample of patients. The COVID-19 vaccination rollout represents an opportunity to study primary vaccine responses in immunocompromised patients, including those receiving anti-CD20 therapy.

We report that SC after vaccination in patients on active anti-CD20 therapy is low and approaches 0%. The low response rate of patients on anti-CD20 therapy appears to be largely attributable to the anti-CD20 therapy rather than the disease or other treatments, and response to vaccination seems to improve incrementally over time but may not reach the level of healthy controls even 12 months after therapy. During the current pandemic, this argues for the need for rapid, real-world studies to guide clinical practice. We need to guickly determine the SC rate following COVID-19 vaccination, whether it can be improved upon with booster shots, and/or influenced via the timing of vaccination relative to anti-CD20 therapy. Additionally, these data serve as a reminder to continue stringent evidence-based infectionprevention strategies such as infection control measures, physical distancing, and appropriate shielding for patients on anti-CD20 therapy. 58 Additionally, in an effort to decrease the household risk of exposure, advocating for vaccination of the caregivers and household contacts of patients receiving anti-CD20 therapy may be appropriate. Finally, while we await future prospective data addressing this issue, we do recommend COVID-19 vaccination for patients receiving anti-CD20 therapy as even a muted response to vaccination is likely better than no response.

Acknowledgment

The authors thank the authors and patients of all studies included in this review.

Authorship

Contribution: A.V. performed literature search, article selection, analysis, and manuscript writing; I.G. performed article selection and manuscript review and revision; S.D.B. and M.C. reviewed and revised the manuscript; and L.K.H. conceived of the study, assisted with analysis, and reviewed and revised the manuscript.

Conflict-of-interest disclosure: L.K.H. was one of the principal investigators on a study partially funded by Gilead Sciences. The remaining authors declare no competing financial interests.

ORCID profiles: A.V., 0000-0001-5369-1501; I.G., 0000-0001-9306-2945.

Correspondence: Lisa K. Hicks, St. Michael's Hospital, Room 2-084, Donnelly Wing, 30 Bond St, Toronto, ON M5B 1W8, Canada: e-mail lisak.hicks@unityhealth.to.

References

- Cooksley CD, Avritscher EB, Bekele BN, Rolston KV, Geraci JM, Elting LS. Epidemiology and outcomes of serious influenza-related infections in the cancer population. Cancer. 2005;104(3):618-628.
- Wong A, Marrie TJ, Garg S, Kellner JD, Tyrrell GJ; SPAT Group. Increased risk of invasive pneumococcal disease in haematological and solidorgan malignancies. Epidemiol Infect. 2010;138(12):1804-1810.
- Ramamurthy MKR, Abraham AM, Sridharan G. Viral infections in immunocompromised hosts. Proc Natl Acad Sci India Sect B Biol Sci. 2012;82(1):95-109.
- Vijenthira A, Gong IY, Fox TA, et al. Outcomes of patients with hematologic malignancies and COVID-19: a systematic review and meta-analysis of 3377 patients. Blood. 2020;136(25):2881-2892.
- Williamson EJ, Walker AJ, Bhaskaran K, et al. Factors associated with COVID-19-related death using OpenSAFELY. Nature. 2020;584(7821):430-436. 5.
- Baden LR, El Sahly HM, Essink B, et al; COVE Study Group. Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine. N Engl J Med. 2021;384(5):403-416.

- Polack FP, Thomas SJ, Kitchin N, et al; C4591001 Clinical Trial Group. Safety and efficacy of the BNT162b2 mRNA Covid-19 vaccine. N Engl J Med. 2020;383(27):2603-2615.
- 8. Voysey M, Clemens SAC, Madhi SA, et al; Oxford COVID Vaccine Trial Group. Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. Lancet. 2021;397(10269):99-111.
- Sadoff J, Le Gars M, Shukarev G, et al. Interim results of a phase 1-2a Trial of Ad26.COV2.S Covid-19 vaccine [published online ahead of print 13 January 2021]. N Engl J Med. doi:10.1056/NEJMoa2034201.
- 10. St Clair EW. Good and bad memories following rituximab therapy [editorial]. Arthritis Rheum. 2010;62(1):1-5.
- 11. Audia S, Samson M, Guy J, et al. Immunologic effects of rituximab on the human spleen in immune thrombocytopenia. *Blood.* 2011;118(16):4394-4400.
- 12. Thurlings RM, Vos K, Wijbrandts CA, Zwinderman AH, Gerlag DM, Tak PP. Synovial tissue response to rituximab: mechanism of action and identification of biomarkers of response. *Ann Rheum Dis.* 2008;67(7):917-925.
- 13. Genberg H, Hansson A, Wernerson A, Wennberg L, Tydén G. Pharmacodynamics of rituximab in kidney allotransplantation. *Am J Transplant*. 2006;6(10):2418-2428.
- 14. Nakou M, Katsikas G, Sidiropoulos P, et al. Rituximab therapy reduces activated B cells in both the peripheral blood and bone marrow of patients with rheumatoid arthritis: depletion of memory B cells correlates with clinical response. *Arthritis Res Ther.* 2009;11(4):R131.
- Thaunat O, Patey N, Gautreau C, et al. B cell survival in intragraft tertiary lymphoid organs after rituximab therapy. Transplantation. 2008;85(11):1648-1653.
- Kamburova EG, Koenen HJ, Borgman KJ, ten Berge IJ, Joosten I, Hilbrands LB. A single dose of rituximab does not deplete B cells in secondary lymphoid organs but alters phenotype and function. Am J Transplant. 2013;13(6):1503-1511.
- 17. Mamani-Matsuda M, Cosma A, Weller S, et al. The human spleen is a major reservoir for long-lived vaccinia virus-specific memory B cells. *Blood*. 2008;111(9):4653-4659.
- 18. Ahuja A, Anderson SM, Khalil A, Shlomchik MJ. Maintenance of the plasma cell pool is independent of memory B cells. *Proc Natl Acad Sci USA*. 2008;105(12):4802-4807.
- 19. Cheng DR, Barton R, Greenway A, Crawford NW. Rituximab and protection from vaccine preventable diseases: applying the evidence to pediatric patients. Expert Rev Vaccines. 2016;15(12):1567-1574.
- Cooper N, Arnold DM. The effect of rituximab on humoral and cell mediated immunity and infection in the treatment of autoimmune diseases. Br J Haematol. 2010;149(1):3-13.
- 21. Hua C, Barnetche T, Combe B, Morel J. Effect of methotrexate, anti-tumor necrosis factor α, and rituximab on the immune response to influenza and pneumococcal vaccines in patients with rheumatoid arthritis: a systematic review and meta-analysis. *Arthritis Care Res (Hoboken)*. 2014:66(7):1016-1026.
- 22. World Health Organization. WHO Expert Committee on Biological Standardization: seventieth report. https://apps.who.int/iris/handle/10665/332102. Accessed 10 January 2021.
- 23. US Food and Drug Administration. Clinical Data Needed to Support the Licensure of Pandemic Influenza Vaccines: Guidance for Industry. May 2007. https://www.fda.gov/regulatory-information/search-fda-guidance-documents/clinical-data-needed-upport-licensure-pandemic-influenza-vaccines. Accessed 10 January 2021.
- 24. Wells GA, Shea B, O'Connell D, et al. The Newcastle-Ottawa Scale (NOS) for assessing the quality of nonrandomised studies in meta-analyses. 2011. http://www.ohri.ca/programs/clinical_epidemiology/oxford.asp. Accessed 10 January 2021.
- 25. Zeng X, Zhang Y, Kwong JS, et al. The methodological quality assessment tools for preclinical and clinical studies, systematic review and metaanalysis, and clinical practice guideline: a systematic review. *J Evid Based Med.* 2015;8(1):2-10.
- 26. Munn Z, Moola S, Lisy K, Riitano D, Tufanaru C. Methodological guidance for systematic reviews of observational epidemiological studies reporting prevalence and cumulative incidence data. *Int J Evid-Based Healthc.* 2015;13(3):147-153.
- 27. Sterne JAC, Savović J, Page MJ, et al. RoB 2: a revised tool for assessing risk of bias in randomised trials. BMJ. 2019;366:l4898.
- 28. Higgins JPT, Thomas J, Chandler J, et al, eds. Cochrane Handbook for Systematic Reviews of Interventions. 2nd ed. Chichester, United Kingdom: John Wiley & Sons; 2019.
- 29. Barendregt JJ, Doi SA, Lee YY, Norman RE, Vos T. Meta-analysis of prevalence. J Epidemiol Community Health. 2013;67(11):974-978.
- 30. Berglund A, Willén L, Grödeberg L, Skattum L, Hagberg H, Pauksens K. The response to vaccination against influenza A(H1N1) 2009, seasonal influenza and Streptococcus pneumoniae in adult outpatients with ongoing treatment for cancer with and without rituximab. *Acta Oncol.* 2014;53(9):1212-1220.
- 31. Hottinger AF, George AC, Bel M, et al; H1N1 Study Group. A prospective study of the factors shaping antibody responses to the AS03-adjuvanted influenza A/H1N1 vaccine in cancer outpatients. *Oncologist.* 2012;17(3):436-445.
- 32. Kapetanovic MC, Kristensen L-E, Saxne T, Aktas T, Mörner A, Geborek P. Impact of anti-rheumatic treatment on immunogenicity of pandemic H1N1 influenza vaccine in patients with arthritis. *Arthritis Res Ther.* 2014;16(1):R2.
- 33. Mackay HJ, McGee J, Villa D, et al. Evaluation of pandemic H1N1 (2009) influenza vaccine in adults with solid tumor and hematological malignancies on active systemic treatment. *J Clin Virol.* 2011;50(3):212-216.
- 34. Monkman K, Mahony J, Lazo-Langner A, Chin-Yee BH, Minuk LA. The pandemic H1N1 influenza vaccine results in low rates of seroconversion for patients with hematological malignancies. *Leuk Lymphoma*. 2011;52(9):1736-1741.

- 35. van Assen S, Holvast A, Benne CA, et al. Humoral responses after influenza vaccination are severely reduced in patients with rheumatoid arthritis treated with rituximab. Arthritis Rheum. 2010;62(1):75-81.
- van der Kolk LE, Baars JW, Prins MH, van Oers MH. Rituximab treatment results in impaired secondary humoral immune responsiveness. Blood. 2002;100(6):2257-2259.
- 37. Villa D, Gubbay J, Sutherland DR, et al. Evaluation of 2009 pandemic H1N1 influenza vaccination in adults with lymphoid malignancies receiving chemotherapy or following autologous stem cell transplant. Leuk Lymphoma. 2013;54(7):1387-1395.
- 38. Nazi I, Kelton JG, Larché M, et al. The effect of rituximab on vaccine responses in patients with immune thrombocytopenia. Blood. 2013;122(11):1946-1953.
- 39. Arad U, Tzadok S, Amir S, et al. The cellular immune response to influenza vaccination is preserved in rheumatoid arthritis patients treated with rituximab. Vaccine. 2011;29(8):1643-1648.
- de Lavallade H, Garland P, Sekine T, et al. Repeated vaccination is required to optimize seroprotection against H1N1 in the immunocompromised host. Haematologica. 2011;96(2):307-314.
- 41. Parrino J, McNeil SA, Lawrence SJ, et al. Safety and immunogenicity of inactivated varicella-zoster virus vaccine in adults with hematologic malignancies receiving treatment with anti-CD20 monoclonal antibodies. Vaccine. 2017;35(14):1764-1769.
- 42. Eisenberg RA, Jawad AF, Boyer J, et al. Rituximab-treated patients have a poor response to influenza vaccination. J Clin Immunol. 2013;33(2):388-396.
- 43. Muller RB, Maier R, Hoschler K, et al. Efficient boosting of the antiviral T cell response in B cell-depleted patients with autoimmune rheumatic diseases following influenza vaccination. Clin Exp Rheumatol. 2013;31(5):723-730.
- Bedognetti D, Ansaldi F, Zanardi E, et al. Seasonal and pandemic (A/H1N1 2009) MF-59-adjuvanted influenza vaccines in complete remission non-Hodgkin lymphoma patients previously treated with rituximab containing regimens. Blood. 2012;120(9):1954-1957.
- 45. Ide Y, Imamura Y, Ohfuji S, et al. Immunogenicity of a monovalent influenza A(H1N1)pdm09 vaccine in patients with hematological malignancies. Hum Vaccin Immunother. 2014;10(8):2387-2394.
- 46. Shah GL, Shune L, Purtill D, et al. Robust vaccine responses in adult and pediatric cord blood transplantation recipients treated for hematologic malignancies. Biol Blood Marrow Transplant. 2015;21(12):2160-2166.
- 47. Small TN, Zelenetz AD, Noy A, et al. Pertussis immunity and response to tetanus-reduced diphtheria-reduced pertussis vaccine (Tdap) after autologous peripheral blood stem cell transplantation. Biol Blood Marrow Transplant. 2009;15(12):1538-1542.
- Surveillance, Epidemiology, and End Results Program. Cancer Stat Facts. 2017. https://seer.cancer.gov/statfacts/. Accessed February 2021.
- 49. Plotkin SA, Orenstein WA, Offit PA, eds. Vaccines. 6th ed. Philadelphia, PA: Elsevier Saunders; 2013.
- 50. Puissant-Lubrano B, Rostaing L, Kamar N, Abbal M, Fort M, Blancher A. Impact of rituximab therapy on response to tetanus toxoid vaccination in kidney-transplant patients. Exp Clin Transplant. 2010;8(1):19-28.
- 51. Oren S, Mandelboim M, Braun-Moscovici Y, et al. Vaccination against influenza in patients with rheumatoid arthritis: the effect of rituximab on the humoral response. Ann Rheum Dis. 2008;67(7):937-941.
- Bedognetti D, Zoppoli G, Massucco C, et al. Impaired response to influenza vaccine associated with persistent memory B cell depletion in non-Hodgkin's lymphoma patients treated with rituximab-containing regimens. J Immunol. 2011;186(10):6044-6055.
- 53. Colucci M, Carsetti R, Serafinelli J, et al. Prolonged impairment of immunological memory after anti-CD20 treatment in pediatric idiopathic nephrotic syndrome. Front Immunol. 2019;10:1653.
- 54. Jaffe D, Papadopoulos EB, Young JW, et al. Immunogenicity of recombinant hepatitis B vaccine (rHBV) in recipients of unrelated or related allogeneic hematopoietic cell (HC) transplants. Blood. 2006;108(7):2470-2475.
- 55. de Jong JC, Palache AM, Beyer WE, Rimmelzwaan GF, Boon AC, Osterhaus AD. Haemagglutination-inhibiting antibody to influenza virus. Dev Biol (Basel), 2003:115:63-73.
- Orange JS, Ballow M, Stiehm ER, et al. Use and interpretation of diagnostic vaccination in primary immunodeficiency: a working group report of the Basic and Clinical Immunology Interest Section of the American Academy of Allergy, Asthma & Immunology. J Allergy Clin Immunol. 2012;130(suppl 3):S1-S24.
- 57. McElhaney JE, Xie D, Hager WD, et al. T cell responses are better correlates of vaccine protection in the elderly. J Immunol. 2006;176(10):6333-6339.
- Chu DK, Akl EA, Duda S, Solo K, Yaacoub S, Schünemann HJ; COVID-19 Systematic Urgent Review Group Effort (SURGE) study authors. Physical distancing, face masks, and eye protection to prevent person-to-person transmission of SARS-CoV-2 and COVID-19: a systematic review and meta-analysis. Lancet. 2020;395(10242):1973-1987.
- 59. Issa NC, Marty FM, Gagne LS, et al. Seroprotective titers against 2009 H1N1 influenza A virus after vaccination in allogeneic hematopoietic stem cell transplantation recipients. Biol Blood Marrow Transplant. 2011;17(3):434-438.
- Kim W, Kim SH, Huh SY, et al. Reduced antibody formation after influenza vaccination in patients with neuromyelitis optica spectrum disorder treated with rituximab. Eur J Neurol. 2013;20(6):975-980.
- 61. Yri OE, Torfoss D, Hungnes O, et al. Rituximab blocks protective serologic response to influenza A (H1N1) 2009 vaccination in lymphoma patients during or within 6 months after treatment. Blood. 2011;118(26):6769-6771.
- 62. Bar-Or A, Calkwood JC, Chognot C, et al. Effect of ocrelizumab on vaccine responses in patients with multiple sclerosis: the VELOCE study. Neurology. 2020;95(14):e1999-e2008.

- 63. Cho A, Bradley B, Kauffman R, et al. Robust memory responses against influenza vaccination in pemphigus patients previously treated with rituximab. JCI Insight. 2017;2(12):e93222.
- Richi P, Martín MD, Navío MT, et al. Antibody responses to influenza vaccine in patients on biological therapy: results of RIER cohort study. Med Clin (Barc), 2019:153(10):380-386.
- 65. Albert D, Dunham J, Khan S, et al. Variability in the biological response to anti-CD20 B cell depletion in systemic lupus erythaematosus. Ann Rheum Dis. 2008;67(12):1724-1731.
- 66. Bingham CO III, Looney RJ, Deodhar A, et al. Immunization responses in rheumatoid arthritis patients treated with rituximab: results from a controlled clinical trial. Arthritis Rheum. 2010;62(1):64-74.
- 67. Bühler S, Jaeger VK, Adler S, et al. Safety and immunogenicity of tetanus/diphtheria vaccination in patients with rheumatic diseases-a prospective multi-centre cohort study. Rheumatology (Oxford). 2019;58(9):1585-1596.
- 68. Horwitz SM, Negrin RS, Blume KG, et al. Rituximab as adjuvant to high-dose therapy and autologous hematopoietic cell transplantation for aggressive non-Hodgkin lymphoma. Blood. 2004;103(3):777-783.
- Mustafa SS, Jamshed S, Vadamalai K, Ramsey A. The use of 20% subcutaneous immunoglobulin replacement therapy in patients with B cell non-Hodgkin lymphoma with humoral immune dysfunction after treatment with rituximab. Clin Lymphoma Myeloma Leuk. 2020;20(9):e590-e596.
- 70. Pescovitz MD, Torgerson TR, Ochs HD, et al; Type 1 Diabetes TrialNet Study Group. Effect of rituximab on human in vivo antibody immune responses. J Allergy Clin Immunol. 2011;128(6):1295-1302.e5.
- 71. Avivi I, Zisman-Rozen S, Naor S, et al. Depletion of B cells rejuvenates the peripheral B-cell compartment but is insufficient to restore immune competence in aging. Aging Cell. 2019;18(4):e12959.
- 72. Richi P, Alonso O, Martín MD, et al. Evaluation of the immune response to hepatitis B vaccine in patients on biological therapy: results of the RIER cohort study. Clin Rheumatol. 2020;39(9):2751-2756.
- 73. Kapetanovic MC, Saxne T, Jönsson G, Truedsson L, Geborek P. Rituximab and abatacept but not tocilizumab impair antibody response to pneumococcal conjugate vaccine in patients with rheumatoid arthritis. Arthritis Res Ther. 2013;15(5):R171.