



Complete Genome Sequence of *Lactobacillus reuteri* SKKU-OGDONS-01, Isolated from a Chicken's Small Intestine

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ABSTRACT *Lactobacillus reuteri* SKKU-OGDONS-01 is a potentially indigenous probiotic strain isolated from the small intestine of a 27-week-old chicken. The complete genome of *L. reuteri* SKKU-OGDONS-01 comprises a single circular chromosome. Its length is 2,259,968 bp, with a G+C content of 38.9%.

Probiotics are live microorganisms, which when administered in an adequate amount can confer benefits to the host (1, 2). In the past, the use of antibiotics (antibiotic growth promoters) in feed additives was prohibited due to concerns about antimicrobial resistance in animal microbiota. As such, there was a need to find alternatives, such as probiotics, that could enhance the immunity of farm animals to fight against various pathogens (3–5). To find candidates for feed additives, indigenous *Lactobacillus* spp., which are potential probiotics, were isolated from the small intestine of a 27-week-old female chicken. The study protocol and standard operating procedures were reviewed and approved by the Institutional Animal Care and Use Committee of the National Institute of Animal Science in South Korea (approval 2018-273). To isolate intestinal bacteria, the small intestine of a chicken was harvested and the sample homogenized using 1.6-mm stainless steel beads. After spreading the homogenized tissue on de Man-Rogosa-Sharpe (MRS) agar, a number of colonies appeared after 2 days. Of the numerous colonies, only one colony was picked from the MRS agar and incubated in MRS medium at 37°C for 2 days. Cells were cultured and later maintained at –85°C using 80% glycerol. To extract the genomic DNA of *Lactobacillus reuteri* SKKU-OGDONS-01, the bacterial cell wall was lysed using lysozyme and mutanolysin, and genomic DNA isolation was performed using the G-spin genomic DNA extraction kit (iNtRON, Republic of Korea) (6). In order to perform strain identification, we conducted a sequence comparison using the 16S rRNA sequence. Two primers (7), 9F (5'-GAGTTTGATCCTGGCTCAG-3') and 1542R (5'-AGAAAGGAGGTGATCCAGCC-3'), were used for amplification of the 16S rRNA gene, followed by sequencing via the Sanger/capillary sequencing method. We found that the 16S rRNA gene sequence of *L. reuteri* SKKU-OGDONS-01 was 99% similar to those of other *L. reuteri* strains reported in the NCBI database. Construction of the complete genome sequence of *L. reuteri* SKKU-OGDONS-01 was based on two sequencing methods, using the PacBio RS II platform (Pacific Biosciences, USA) and the HiSeq 2000 platform (Illumina, USA) at MacroGen (Seoul, Republic of Korea). Of the total 151,868 reads obtained after PacBio sequencing, the average read length was 9,424 bp (451× coverage), and these raw sequence data were assembled using the Hierarchical Genome Assembly Process (HGAP3) protocol in the SMRT Analysis software version 2.3.0 (8). Additionally, Illumina sequencing was conducted using the HiSeq 2000 platform. The paired-end reads from a 325-bp insert library were sequenced for *L. reuteri* SKKU-OGDONS-01. A total of 57,137,834 reads, with an average read length of 100 bp, were generated from Illumina sequencing. After error

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correction using Pilon (version 1.21) (9) with Illumina short reads, a single contig was generated, at 2,259,968 bp in length, with a G+C content of 38.9%. Using comparative genome analysis, the whole genome of *L. reuteri* SKKU-OGDONS-01 was about 97% similar to those of other *L. reuteri* strains, including *L. reuteri* strain I49 (GenBank accession number [NZ_CP015408](https://doi.org/10.1093/jn/138.6.1250S)). Genome annotation was carried out using the NCBI Prokaryote Genome Annotation Pipeline (PGAP) version 4.6 (10). Based on the PGAP result, the chromosome included 2,165 genes and 1,941 coding sequences (CDS). The whole genome of *L. reuteri* SKKU-OGDONS-01 consists of 70 tRNA genes and 18 rRNA genes.

Data availability. The chromosomal sequence of *L. reuteri* SKKU-OGDONS-01 has been deposited at GenBank under the accession number [CP029615](https://doi.org/10.1093/jn/138.6.1250S). The sequencing reads (under Sequence Read Archive number [SRP162209](https://doi.org/10.1093/jn/138.6.1250S)) can be accessed through BioProject number [PRJNA473291](https://doi.org/10.1093/jn/138.6.1250S) and BioSample number [SAMN09270376](https://doi.org/10.1093/jn/138.6.1250S).

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REFERENCES

1. Farnworth ER. 2008. The evidence to support health claims for probiotics. *J Nutr* 138:1250S–1254S. <https://doi.org/10.1093/jn/138.6.1250S>.
2. World Health Organization, Food and Agriculture Organization of the United Nations. 2010. Evaluation of certain food additives: seventy-first report of the Joint FAO/WHO Expert Committee on Food Additives. *World Health Organ Tech Rep Ser* 956:1–80.
3. Versteegen MW, Williams BA. 2002. Alternatives to the use of antibiotics as growth promoters for monogastric animals. *Anim Biotechnol* 13: 113–127. <https://doi.org/10.1081/ABIO-120005774>.
4. Dibner J, Richards J. 2005. Antibiotic growth promoters in agriculture: history and mode of action. *Poult Sci* 84:634–643. <https://doi.org/10.1093/ps/84.4.634>.
5. Mathur S, Singh R. 2005. Antibiotic resistance in food lactic acid bacteria—a review. *Int J Food Microbiol* 105:281–295. <https://doi.org/10.1016/j.ijfoodmicro.2005.03.008>.
6. Macklaim JM, Fernandes AD, Di Bella JM, Hammond J-A, Reid G, Gloor GB. 2013. Comparative meta-RNA-seq of the vaginal microbiota and differential expression by *Lactobacillus iners* in health and dysbiosis. *Microbiome* 1:12. <https://doi.org/10.1186/2049-2618-1-12>.
7. Yoon J-H, Lee ST, Park Y-H. 1998. Inter- and intraspecific phylogenetic analysis of the genus *Nocardioides* and related taxa based on 16S rDNA sequences. *Int J Syst Evol Microbiol* 48:187–194. <https://doi.org/10.1099/00207713-48-1-187>.
8. Zhang Z, Schwartz S, Wagner L, Miller W. 2000. A greedy algorithm for aligning DNA sequences. *J Comput Biol* 7:203–214. <https://doi.org/10.1089/10665270050081478>.
9. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
10. Fernández-González AJ, Lasa AV, Fernández-López M. 2018. Whole-genome sequences of two *Arthrobacter* strains isolated from a holm oak rhizosphere affected by wildfire. *Genome Announc* 6:e00071-18. <https://doi.org/10.1128/genomeA.00071-18>.