



The Atherosclerotic Plaque Microenvironment as a Therapeutic Target

Rajan Pandit¹ · Arif Yurdagul^{1,2}

Accepted: 18 March 2025
© The Author(s) 2025

Abstract

Purpose of Review Atherosclerosis is traditionally viewed as a disease triggered by lipid accumulation, but growing evidence underscores the crucial role of the plaque microenvironment in disease progression. This review explores recent advances in understanding how cellular and extracellular components of the plaque milieu drive atherosclerosis, with a focus on leveraging these microenvironmental factors for therapeutic intervention. This review highlights recent advances in cell-cell crosstalk and matrix remodeling, offering insights into innovative therapeutic strategies for atherosclerotic cardiovascular disease.

Recent Findings While atherosclerosis begins with the subendothelial retention of apolipoprotein B (ApoB)-containing lipoproteins, its progression is increasingly recognized as a consequence of complex cellular and extracellular dynamics within the plaque microenvironment. Soluble factors and extracellular matrix proteins shape mechanical properties and the biochemical landscape, directly influencing cell behavior and inflammatory signaling. For instance, the deposition of transitional matrix proteins, such as fibronectin, in regions of disturbed flow primes endothelial cells for inflammation. Likewise, impaired clearance of dead cells and chronic extracellular matrix remodeling contribute to lesion expansion and instability, further exacerbating disease severity.

Summary Targeting the plaque microenvironment presents a promising avenue for stabilizing atherosclerotic lesions. Approaches that enhance beneficial cellular interactions, such as boosting macrophage efferocytosis to resolve inflammation while mitigating proatherogenic signals like integrin-mediated endothelial activation, may promote fibrous cap formation and reduce plaque vulnerability. Harnessing these mechanisms may lead to novel therapeutic approaches aimed at modifying the plaque microenvironment to combat atherosclerotic cardiovascular disease.

Keywords Atherosclerosis · Atherosclerotic plaque microenvironment · Cardiovascular disease · Extracellular matrix remodeling · Inflammation resolution

Introduction

Atherosclerosis Initiation and Progression

Atherosclerosis is a progressive inflammatory disease that begins with the subendothelial retention of cholesterol-rich ApoB-containing lipoproteins in medium- and large-sized

arteries [1–3]. According to the response-to-retention model, this trapping of lipoproteins in the arterial intima represents the initial key event in atherosclerosis development [4, 5]. The retained low-density lipoprotein (LDL) particles become modified (e.g., by oxidation, glycation, or aggregation) and trigger endothelial cell activation and dysfunction. Endothelial dysfunction, characterized by increased permeability, reduced nitric oxide bioavailability, and upregulation in adhesion molecules, establishes a proinflammatory microenvironment that promotes immune cell infiltration [6–8]. Monocytes adhere to activated endothelium and transmigrate into the arterial wall, where they differentiate into macrophages and uptake lipids to form foam cells [9]. The persistent accumulation of lipids and proinflammatory mediators also impairs the clearance of dead cells by macrophages, a process termed ‘efferocytosis’, leading

✉ Arif Yurdagul
arif.yurdagul@lsuhs.edu

¹ Department of Molecular and Cellular Physiology, LSU Health Sciences Center at Shreveport, Shreveport, LA, USA

² Department of Pathology and Translational Pathobiology, LSU Health Sciences Center at Shreveport, Shreveport, LA, USA

to post-apoptotic necrosis and necrotic core formation in plaques [10]. This sequence produces unstable atherosclerotic plaques with large necrotic cores and thin fibrous caps, which may either rupture or undergo superficial plaque erosion, increasing the risk of thromboembolic cardiovascular events, such as myocardial infarction and stroke [11–13].

Clinical trials targeting inflammation have confirmed the link between lesional inflammation and cardiovascular events. The Canakinumab Anti-Inflammatory Thrombosis Outcome Study (CANTOS) showed that IL-1 β inhibition reduced recurrent events independent of lipid-lowering but was associated with a higher incidence of fatal infections [14, 15]. Similarly, the Colchicine Cardiovascular Outcomes Trial (COLCOT) demonstrated reduced ischemic events with low-dose colchicine, albeit with a slight increase in pneumonia in those receiving treatment [15]. These landmark trials underscore that while reducing inflammation can mitigate atherosclerosis, preserving host defenses remains essential. Thus, therapies that resolve inflammation without broadly suppressing immunity are a promising approach to combat atherosclerotic cardiovascular disease.

The Atherosclerotic Plaque Microenvironment

The plaque microenvironment— comprising extracellular matrix (ECM) proteins, biomechanical forces, and soluble factors— plays a pivotal role in atherogenesis. In healthy arteries, the intimal ECM is primarily composed of basement membrane proteins such as collagen IV, laminin, perlecan, and nidogen, which provide structural support and maintain cellular homeostasis [16, 17]. In atherosclerosis, this ECM undergoes extensive remodeling [18]. One of the most significant changes is the deposition of transitional matrix proteins in the subendothelial space, replacing the normal basement membrane with a proinflammatory provisional matrix. For instance, fibronectin accumulates at atheroprone sites exposed to disturbed flow even before the appearance of plaque formation [19]. This fibronectin-rich matrix “primes” endothelial cells for inflammatory activation by stimuli such as oxidized LDL (oxLDL), in contrast to the atheroprotective signaling elicited by laminin and collagen IV in healthy arteries [2, 18, 20]. Beyond fibronectin, other matrix components also contribute to the lesional microenvironment. Negatively charged glycosaminoglycans (e.g. hyaluronan, heparan sulfate, chondroitin sulfate) within proteoglycans bind and sequester growth factors such as platelet-derived growth factor (PDGF), transforming growth factor β (TGF β), hepatocyte growth factor (HGF), fibroblast growth factor (FGF), and insulin-like growth factor (IGF), modulating the proliferation and migration of cells [21–23]. As atherosclerosis progresses, lesional cells alter the microenvironment by releasing

cytokines, chemokines, and metabolites that propagate a self-amplifying cycle of chronic inflammation and fibrotic responses that drive more inflammation and ECM remodeling [23, 24]. However, important aspects of ECM remodeling in atherosclerosis remain poorly understood. The precise mechanisms governing the transition from a healthy basement membrane to a proatherogenic matrix, as well as how altered ECM composition sustains chronic inflammation and disease progression, still require further investigation. Because the plaque microenvironment plays a central role in regulating cell behavior, targeting this niche is therapeutically attractive. The following sections examine key pathways among lesional cells and discuss how modulating these interactions or the ECM could stabilize plaques.

Endothelial-Leukocyte Crosstalk

Endothelial Cell Activation

Atherosclerotic lesions preferentially form at arterial branch points and curvatures, where disturbed blood flow creates low and oscillatory shear stress. In contrast, high, unidirectional laminar flow is atheroprotective by driving nuclear localization of KLF2 and KLF4, which in turn enhances endothelial nitric oxide synthase (eNOS) expression and nitric oxide production [25, 26]. Recent studies indicate that atheroprotective shear stress induces HEG1-mediated KLF2 and KLF4 expression, and endothelial HEG1 is downregulated in individuals with advanced atherosclerosis. In contrast, disturbed flow suppresses the atheroprotective transcription factors KLF2 and KLF4 and activates NF- κ B and other inflammatory pathways in endothelial cells [27, 28]. Endothelial cells also secrete chemokines such as CCL2 (MCP-1), CCL5, and CX3CL1 that diffuse into the bloodstream. Once captured by endothelial selectins, circulating monocytes and other leukocytes firmly adhere through integrins, such as VLA-4 and LFA-1, which bind to endothelial VCAM-1 and ICAM-1. This multistep adhesion cascade (summarized in Table 1) guides leukocytes across the endothelium into the intima. Once transmigrated, monocytes become activated by macrophage colony-stimulating factor (M-CSF), CCL2, and other factors that drive their differentiation into macrophages. The local microenvironment will then shape the macrophage phenotype, as will be discussed later.

Response to Oxidized LDL

Positively charged ApoB-containing lipoproteins traverse the endothelium via paracellular or transcytosis routes [8]. Once in the subendothelial space, LDL undergoes oxidative modification and becomes trapped within the negatively

Table 1 Major Leukocyte-Endothelial adhesion molecules [101–103]

Leukocytes Molecules/ Receptors	Endothelial or ECM ligands	Role
L-selectin, VLA-4 ($\alpha_4\beta_1$)	s-Le ^x , VCAM-1 or FN	tethering
L-selectin, VLA-4, ESL-1, PSGL-1	s-Le ^x , VCAM-1, E-selectin, P-selectin	rolling
Chemokine-R, PAF-R	Chemokines, PAF	activation
LFA-1($\alpha_1\beta_2$); Mac-1 ($\alpha_M\beta_2$); VLA-4 ($\alpha_4\beta_1$)	ICAM-1,2,3 and JAM-A; ICAM1,2, FN, VN; VCAM-1, FN	adhesion
Mac-1 (LFA-1)	ICAM-1 (ICAM-2)	migration
PECAM-1, CD99	PECAM-1 (CD31), CD99	transmigration
$\alpha_6\beta_1$	Laminin	migrating through basal lamina
Mac-1, $\alpha_5\beta_1$	FG, FN	interact with fibronectin
JAM A, B	JAM A, B, C	homophilic/heterophilic interaction for firm interaction

ESL-1, E-selectin ligand 1; s-Le^x sialyl-Lewis x antigen; VCAM-1, vascular cell adhesion molecule 1; PSGL-1, P-selectin glycoprotein ligand 1; LFA-1, lymphocyte function-associated antigen 1; Mac-1, macrophage-1 antigen; PAF, platelet-activating factor; PAF-R, PAF receptor; ICAM, intercellular adhesion molecule; PECAM-1, platelet/endothelial cell adhesion molecule 1; PECAM-1, $\alpha_6\beta_1$, VLA-6, the integrin responsible for binding laminin. FN, fibronectin; FG, fibrinogen; VN, vitronectin

charged, proteoglycan-rich subendothelial matrix [8, 29–31]. These modified LDLs disrupt endothelial cell function by activating NF- κ B, increasing permeability, reducing nitric oxide production, upregulating adhesion molecules, and enhancing leukocyte infiltration (Fig. 1A) [32, 33].

Endothelial Responses Modulated by the ECM

The composition of the subendothelial ECM not only regulates LDL retention but also provides signals to endothelial cells that modulate inflammation [34, 35]. Endothelial cells interact with the basement membrane proteins via the integrin family of cell-matrix receptors. Such interactions can transmit atheroprotective signals. As one example, endothelial interactions with basement membrane proteins via integrin $\alpha_2\beta_1$ signaling blunt disturbed flow and oxLDL-induced inflammation [7, 24]. However, in early atherosclerosis, as the basement membrane is being replaced by fibronectin, endothelial cells switch to using integrin $\alpha_5\beta_1$ for adhesion. Activation of integrin $\alpha_5\beta_1$ by fibronectin amplifies endothelial cell activation, creating a positive feedback loop in disturbed flow regions [35, 36]. Thus, the arterial microenvironment, from biomechanical forces to matrix composition, critically regulates endothelial behavior and, consequently, leukocyte recruitment in atherosclerosis.

Endothelial-Vascular Smooth Muscle Cell Crosstalk

Endothelial cells and vascular smooth muscle cells (vSMCs) are in close proximity in the arterial microenvironment and communicate through various mechanisms, including direct cell-cell contact, paracrine signaling, and extracellular vesicles [37]. In a healthy vessel, this endothelial–vSMC crosstalk regulates vascular tone and structure. For instance, the vasodilatory signal nitric oxide is released by endothelial cells that act on adjacent vSMCs to induce relaxation, thus maintaining appropriate vessel caliber and blood pressure. Under laminar flow conditions, endothelial production of nitric oxide is high and vSMCs remain in a contractile, quiescent state. Disturbed flow, however, not only activates endothelial cells (as described above) but also alters the signals sent to vSMCs. ECs exposed to low/oscillatory shear can exhibit an EndMT (endothelial-to-mesenchymal transition) phenotype, producing factors like TGF β 2 and IL-1 β that impact vSMCs. TGF β 2 from activated endothelial cells can drive vSMCs toward a more synthetic or matrix-producing vSMC phenotype (Fig. 1B). Alternatively, endothelial-derived IL-1 β has been shown to stimulate vSMC migration and proliferation [38, 39]. These effects illustrate how inflammatory crosstalk from the endothelium can drive vSMCs away from their contractile phenotype.

Another key mode of communication between endothelial cells and vSMCs involves microRNAs (miRNAs), which can be transferred directly or via extracellular vesicles. A notable example is the transfer of miR-143/145 from vSMCs to endothelial cells. These miRNAs, highly expressed in vSMCs, promote the contractile SMC phenotype. When endothelial cells produce TGF β during direct interactions with vSMCs, it induces the formation of membrane nanotubes that facilitate the transfer of miR-143/145 from vSMCs to endothelial cells. The uptake of these miRNAs suppresses endothelial angiogenic activity and stabilizes vessels [40]. This exchange creates a protective feedback loop, with vSMCs helping to establish an atheroprotective, quiescent endothelial state. However, endothelial–vSMC crosstalk can become harmful in disease settings. Disturbed flow and chronic inflammation can reduce the transfer of beneficial signals, such as miR-143/145 while amplifying detrimental ones. For instance, activated endothelial cells may release factors or vesicles that drive vSMC dedifferentiation, promoting a less stable phenotype. Simultaneously, inflamed vSMCs may secrete proteases or cytokines that further damage the endothelium, exacerbating vascular dysfunction [41]. Because of this dynamic interplay, strategies that disrupt maladaptive endothelial–vSMC crosstalk or restore its protective functions represent a promising therapeutic strategy to attenuate atherosclerosis.

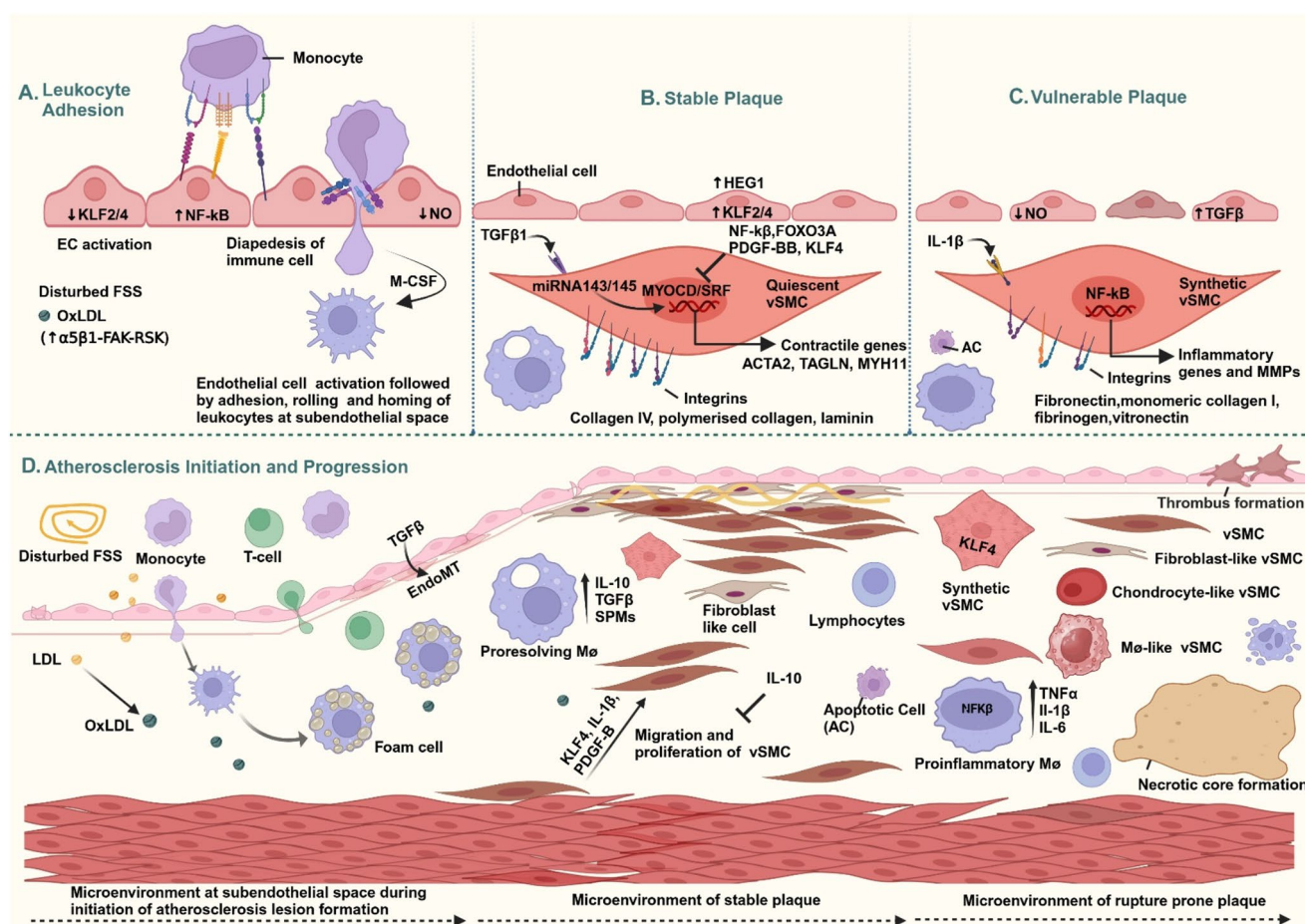


Fig. 1 Atherosclerosis Initiation and Progression

Vascular Smooth Muscle Cell Phenotypic Modulation in the Plaque Microenvironment

Vascular SMCs are traditionally regarded as the structural cells of the arterial wall, responsible for both vasoconstriction and dilation [42]. Under normal conditions, vSMCs exhibit a contractile phenotype, characterized by a high propensity to constrict when necessary and a low rate of proliferation and migration. These vSMCs express high levels of contractility-related genes, including alpha smooth muscle actin (*ACTA2*), calponin (*CNN1*), transgelin (*TAGLN*), smoothelin (*SMTN*), and SM-MHC (*MYH11*) (Fig. 1B). This contractile phenotype is primarily maintained by myocardin, which interacts with serum response factor (SRF) to drive the expression of contractile genes [43–46].

In atherosclerosis, vSMCs are exposed to inflammatory cytokines, growth factors, and lipid mediators that can trigger phenotypic modulation, a process in which contractile markers are downregulated and acquire features associated with other cell types. Drivers of this switch include KLF4, NF- κ B, and forkhead box O3a (FOXO3A), which become activated in vSMCs by atherogenic stimuli and inhibit

myocardin function. This results in a loss in the contractile phenotype and the emergence of a synthetic, migratory, and proliferative phenotype (Fig. 1B). Phenotypically modulated vSMCs carry the potential to transdifferentiate into cells resembling macrophages, mesenchymal stem cells, osteochondrogenic cells, or fibroblasts, and single-cell RNA sequencing coupled with lineage-tracing studies have revealed that a significant fraction of cells within advanced plaques that appear “macrophage-like” or “foam cell-like” originate from vSMCs rather than monocyte-derived macrophages [47, 48]. These vSMC-derived foam cells often have impaired cholesterol efflux capacity relative to *bona fide* macrophages, which may exacerbate lipid accumulation in atherosclerosis plaques.

Smooth muscle cells interact with ECM proteins via the integrin family of cell-matrix receptors, and integrin-mediated signaling is crucial in determining vSMC phenotype and alters atherosclerosis progression (Fig. 1B and C). For instance, disrupting vSMC interactions with matrix proteins, particularly fibronectin, using α 5 β 1 or α v β 3 inhibitors has been shown to reduce atherosclerotic plaque size. Notably, integrin α v β 3 inhibition decreases fibrous cap formation [7,

35]. Interestingly, while depleting fibronectin blocks endothelial cell activation, it simultaneously inhibits migration and proliferation of vSMCs, resulting in thin fibrous caps and poorly assembled ECM networks [19]. Vascular SMCs serve as a primary source of collagen within the lesional microenvironment, reinforcing the fibrous cap and enhancing plaque stability. However, in inflamed atheromas, macrophage-derived proteases, particularly MMPs, actively degrade collagen, weaken the protective fibrous cap, and compromise plaque stability, increasing the risk of plaque rupture [49, 50]. However, phenotypic switching of vSMCs is double-edged, whereby the synthetic state contributes to plaque expansion and instability by secreting inflammatory cytokines and the matrix-degrading enzymes matrix metalloproteases (MMPs) but can also produce ECM proteins that form the protective fibrous cap, an essential feature for stability. Interestingly, fibronectin in the lesional microenvironment promotes phenotypic switching that initially participates in fibrous cap formation, whereas a collagen-rich matrix can push vSMCs toward a more contractile, matrix-producing phenotype that stabilizes the cap. This dynamic underscores the complex role of vSMCs and highlights the importance of understanding the tissue microenvironment in atherosclerosis.

Indeed, approaches that return synthetic vSMCs towards a contractile phenotype can stabilize plaques. For example, microRNA-143/145, which promotes the contractile program in vSMCs, is protective, and deleting these miRNAs accelerates atherosclerosis. Similarly, TGF β 1 signaling sustains a differentiated state and increases collagen production [51, 52]. In contrast, KLF4 is a key driver of vSMC dedifferentiation during atherosclerosis (Fig. 1B). Conditional deletion of KLF4 in vSMCs of atheroprone mice fed a high-fat diet reduces lesion size, increases fibrous cap thickness, and decreases vSMC-derived macrophages and mesenchymal stem-like cells while increasing α SMA⁺ cells [53, 54]. Advancements in single-cell RNA sequencing (scRNA-seq) and lineage-tracing methods have uncovered distinct vSMC phenotypes during atherogenesis [55]. Recent findings have revealed at least four distinct phenotypes based on marker expression: contractile-like, fibroblast-like, chondrocyte-like, and macrophage-like. Further studies indicate that macrophages modulate vSMC function during atherosclerosis progression through IL-1 β , whereby macrophage-derived IL-1 β influences vSMC phenotype [55, 56]. Therapeutically targeting pathways such as KLF4 or NF- κ B in vSMCs or bolstering pro-contractile signals (myocardin/SRF, TGF β , miR-143/145) could tilt vSMCs toward a phenotype that stabilizes plaques rather than destabilizes them (Fig. 1B).

Efficient Efferocytosis by Macrophages in Inflammation Resolution

Efferocytosis is the process by which macrophages bind, engulf, and digest apoptotic cells, ensuring the efficient clearance of cellular debris and maintaining tissue homeostasis. Proresolving macrophages play a key role in resolving inflammation and preventing post-apoptotic necrosis by efficiently clearing apoptotic cells. When macrophages consume apoptotic cells, they respond by suppressing proinflammatory cytokine production and releasing anti-inflammatory and proresolving mediators [57–59]. For instance, efferocytosis stimulates the secretion of specialized proresolving lipid mediators (SPMs) and anti-inflammatory cytokines such as IL-10 and TGF β (Fig. 1D). IL-10 is particularly important for inflammation resolution, as it enhances cholesterol efflux via activating the PPAR γ -LXR pathway. This upregulates the cholesterol transporters ABCA1 and ABCG1 while simultaneously reducing TNF α , IL-6, and MCP-1 [60–65]. One mechanism by which IL-10 inhibits inflammation is through induction of the transcriptional regulator Bcl-3, which interferes with NF- κ B signaling to reduce IL-6 production in response to Toll-like receptor activation [66, 67]. Macrophages engulfing apoptotic cells also metabolize the contents of those cells. For example, the metabolism of apoptotic cell-derived arginine by macrophages leads to the production of polyamines (such as putrescine and spermidine) that further promote resolution, and spermidine and spermine can suppress inflammasome activation and IL-1 β release [60, 68, 69]. Additionally, the breakdown of fatty acids and cholesterol from ingested apoptotic cells generates SPMs and oxysterols that engage nuclear receptors, such as LXR, to increase the expression of the key apoptotic cell receptor MerTK [63].

In advanced atherosclerotic lesions, macrophage efferocytosis becomes defective. As macrophages are overwhelmed by excess apoptotic cells and exposed to an inflamed microenvironment, their capacity to phagocytose and process dead cells diminishes [70]. Impaired efferocytosis leads to the accumulation of uncleared apoptotic bodies, which undergo post-apoptotic necrosis—releasing cytotoxic contents and proinflammatory molecules into the plaque. The outcome is an expanding necrotic core and heightened inflammation, conditions strongly linked to plaque instability [71]. Consistently, genetic and pharmacological approaches that restore efferocytosis enhance features associated with plaque stability, including smaller necrotic cores and increased fibrous cap thickness [72]. For example, blockade of the “don’t eat me” signal CD47, which normally inhibits macrophage phagocytosis via SIRP α , with antagonistic antibodies can restore efferocytosis, leading to enhanced clearance of apoptotic cells and reduced necrotic

core size, and overall plaque stabilization [73]. Similarly, overexpressing a cleavage-resistant MerTK or targeting negative regulators of MerTK, such as CAMKII γ , improves features of plaque stability [74, 75].

A recently discovered feature of efferocytosis is that it can trigger a proliferative burst of macrophages with a pro-resolving phenotype. Apoptotic cell-derived nucleotides activate a DNA-PK–Akt–Myc pathway and drive the expansion of a macrophage subpopulation specialized in resolution through a process termed “efferocytosis-induced macrophage proliferation” (EIMP). These macrophages produce high levels of IL-10 and TGF β and can rapidly engulf additional apoptotic cells, thus forming a positive feedback loop for restoring homeostasis [58]. However, this beneficial cycle is easily disrupted in an inflamed microenvironment. TNF α and IL-1 β , which are abundant in advanced atherosclerosis, can promote ADAM17-dependent proteolytic cleavage of MerTK, substantially disabling efferocytosis [76]. Thus, restoring efferocytosis is an attractive therapeutic strategy to stabilize rupture-prone atheromas.

Crosstalk between Macrophages and Vascular Smooth Muscle Cells

Interactions between macrophages and vSMCs within the plaque significantly influence whether a lesion progresses towards instability or heals into a stable fibrous plaque—exerting both beneficial and detrimental effects. On the beneficial side, macrophages engaged in efferocytosis release mediators that support vSMC function. For example, IL-10 attenuates PDGF- or LPS-induced vSMC proliferation and migration, thereby potentially limiting intimal hyperplasia [66] [77]. As previously mentioned, TGF β promotes the contractile phenotype in vSMCs while also stimulating collagen synthesis. It also suppresses vSMC expression of inflammatory mediators, such as inducible nitric oxide synthase (iNOS) and IL-6, further promoting a stable plaque environment [78–80]. Additionally, macrophage-derived metabolites from efferocytosis serve as important signals for vSMCs. One notable example is lactate, whereby recent studies indicate that macrophage metabolizing apoptotic cells produce lactate, which in turn signals vSMCs to adopt a reparative, matrix-producing state [81]. Polyamines produced by macrophages following efferocytosis may have similar pro-fibrotic effects on vSMCs [23, 82]. Together, these interactions suggest that efferocytosis by macrophages promotes vSMC-mediated plaque stability.

Conversely, proinflammatory macrophages can drive vSMC dysfunction, contributing to plaque instability. For instance, in co-culture experiments, oxLDL-activated macrophages have been shown to trigger a phenotypic switch in nearby vSMCs, pushing them toward a macrophage-like

Table 2 Molecular and cellular interactions between lesional cells

Soluble Factors Source (vSMCs/Macrophages)	Signaling/Role	Ref
CCL2, CXCL1, BMPs from vSMCs	recruit macrophages (M ϕ s)	[104–106]
TNF α from macrophages	atherosclerotic plaque calcification formation via BMP-2 expression in VSMCs	[87]
MMP1/9 from both cells	reduce collagen synthesis (col I/III, elastin) and degrade ECM of the fibrous cap	[107]
IL-1 β , TLR-2, and VEGF-A from both cells	reduce plaque stability through inflammation, neovascularization, and calcification	[107]
IL-5 from macrophage	reduce inflammation and reduced Ang II-induced vSMC apoptosis within the aorta	[86]
oxLDL-activated monocytes co-cultured with vSMCs	nucleotide-binding oligomerization domain-like receptor protein 3 (NLRP3) inflammasome activation in VSMCs, transform vSMCs into a macrophage-like phenotype, gasdermin D-dependent pyroptosis of vSMCs	[83, 108]
Transfer of atherosclerotic factors via extracellular vesicles.	macrophages transfer miRNA, LDL, and protein cocktails to other macrophages and VSMCs.	[109, 110]

state through inflammasome signaling [83]. Additionally, macrophages release proteases, such as MMP-9 and MMP-13, which actively degrade collagen and elastin within the plaque microenvironment [84, 85]. Excessive macrophage activation can weaken the fibrous cap by driving collagen degradation faster than vSMCs can produce it, ultimately thinning the fibrous cap and increasing plaque instability. Moreover, proinflammatory mediators, such as TNF α , can induce vSMC apoptosis while simultaneously suppressing collagen synthesis, further weakening the fibrous cap [86]. Emerging evidence suggests that macrophages contribute to plaque calcification by secreting factors that stimulate an osteogenic program in vSMCs, potentially rendering plaques more susceptible to interfacial debonding [87] (Table 2). Ultimately, the impact of macrophage-SMC interactions on plaque stability hinges on the balance between resolving and inflammatory macrophage phenotypes.

Therapeutic Strategies Targeting the Plaque Microenvironment

Growing insights into the plaque microenvironment have driven the development of preclinical therapies that act directly on lesional cells rather than targeting systemic risk factors alone. One promising approach involves engineering nanoparticles to deliver drugs directly to atherosclerotic

lesions, enhancing local bioavailability while minimizing off-target effects. Similarly, nanoparticles decorated with macrophage-targeting ligands have been used to deliver anti-inflammatory agents, such as IL-10, colchicine, and rapamycin, effectively reducing vascular inflammation [88–90]. Others have delivered payloads to enhance efferocytosis, such as nanoparticles presenting MerTK on their surface, which promote apoptotic cell clearance and significantly attenuate atherosclerosis in mice [91]. A multi-targeted strategy using RNAi delivered by nanoparticles to silence the adhesion molecules ICAM-1, VCAM-1, and E-selectin simultaneously reduces leukocyte recruitment and plaque inflammation [92].

Recognizing the role of lipid and metabolic signals in lesional cells, nuclear receptor agonists have garnered substantial attention. Encapsulating the LXR agonist GW3965 in targeted nanoparticles enhanced cholesterol removal from plaque macrophages while avoiding hepatic steatosis, resulting in overall anti-atherogenic benefits [93]. Similarly, activating PPAR γ , which promotes IL-10 production and enhances efferocytosis, has been achieved using drugs such as pioglitazone, and pioglitazone-loaded nanoparticles were found to prevent features of plaque instability in mice by driving macrophage polarization towards a proresolving phenotype [94]. Notably, enhancing apoptotic cell clearance by macrophages has emerged as a promising strategy for plaque stability. One approach targets the inhibitory CD47-SIRP α pathway, where blocking the “don’t eat me” signal CD47 with antagonistic antibodies or peptides can reactivate macrophage efferocytosis, as demonstrated by reduced necrotic cores and smaller lesions in treated animals. Another approach is to supplement pro-efferocytic factors [95]. For instance, injecting recombinant annexin A1 or resolvin D1 boosts the engulfment capacity of phagocytes [96]. Additionally, the novel strategy of delivering the MerTK receptor itself to lesions via nanocarriers fused with MerTK ectodomains has been used to directly promote apoptotic cell uptake in plaques [91, 97].

Building on clinical trial successes, therapies that blunt specific cytokine signals in plaques are being refined. IL-1 β neutralization with canakinumab has already been proven to reduce cardiovascular events [14]. In addition to lowering inflammation, IL-1 β inhibition promotes the accumulation of fibrous cap-forming fibroblast-like cells originating from vSMCs and other stromal cells, which increases cap thickness [56]. This finding suggests IL-1 β blockade not only reduces inflammation but also favorably alters cell composition in the plaque. Meanwhile, low-dose colchicine (which broadly dampens inflammation, including IL-1 β and IL-18 production from macrophages) has emerged as a potential adjunct therapy for coronary disease [15]. Ongoing work

aims to maximize the anti-inflammatory benefit of these agents while minimizing immunosuppressive risks.

An innovative area of research involves reprogramming immune cells to an atheroprotective state *ex vivo* before reintroduction. Monocytes can be metabolically “trained” to adopt a proresolving phenotype. For example, treating human monocytes with 4-phenylbutyric acid (4-PBA) *ex vivo* induces a sustained anti-inflammatory phenotype. When reintroduced into mice, these 4-PBA-trained monocytes home to lesions and ameliorate atherosclerosis. Mechanistically, trained monocytes show reduced expression of adhesion molecules (ICAM-1) and chemoattractants (CCL2) due to the dampening of TLR signaling adaptors, along with the restoration of cellular housekeeping functions like pexophagy. These trained monocytes also upregulate the surface protein CD24, which can engage Siglec-10 on other immune cells to transmit inhibitory signals. Through this mechanism, CD24^{hi}-trained monocytes curb the activity of neutrophils, T cells, and B cells in the plaque microenvironment, collectively reducing inflammation [98]. Although still in the preclinical phase, these cell-based therapies highlight the potential for modulating cell function within the plaque microenvironment.

Conclusions

Our understanding of atherogenesis has evolved beyond a strictly lipid-centric view to encompass the intricate interplay of cellular and extracellular components within the plaque microenvironment. Advances in single-cell transcriptomics, fate-mapping models, and metabolomics have uncovered an unprecedented heterogeneity of cell phenotypes within atherosclerotic lesions, with some driving chronic inflammation while others resolution [56, 99, 100]. Translating these insights into new therapies will require the continued integration of cutting-edge technologies. Multi-omics approaches provide a comprehensive molecular landscape of atherosclerosis, while high-resolution imaging techniques, such as MALDI mass-spectrometry imaging, enable spatial mapping of key metabolic signatures. Furthermore, artificial intelligence and systems biology are becoming indispensable tools for deciphering the complex regulatory networks that govern atherosclerosis progression. These emerging insights could guide the development of precision therapies, such as those that enhance efferocytosis and promote collagen assembly while suppressing pathways that contribute to plaque expansion and rupture. Ultimately, atherosclerotic plaques are highly dynamic microenvironments where continuous interactions between immune cells, vSMCs, and ECM components shape disease outcomes. Targeting the plaque microenvironment through modulation

of cell phenotypes, matrix remodeling, or precision drug delivery represents a promising strategy to mitigate plaque instability and prevent acute cardiovascular events.

Panel A: Endothelial cell activation augments the expression of endothelial adhesion molecules that bind cognate molecules/receptors on the leukocytes, resulting in tethering, rolling, adhesion, and diapedesis toward the subendothelial space.

Panel B: Stable Plaque: Efferocytosis by macrophages release pro-resolving mediators such as TGF β and IL10 that promote vSMC quiescence. The presence of a basement membrane including collagen IV and laminin mitigate inflammation via integrin α 2 β 1 and α 6 β 1. The accumulation of non-smooth muscle fibroblast-like cells within the fibrous cap has recently been found in a stable plaque microenvironment.

Panel C: Unstable Plaque: Defective efferocytosis and an inflamed microenvironment favors plaque instability. The inefficient removal of apoptotic cells and activation of macrophages by oxLDL triggers the release of the proinflammatory mediators TNF α and IL-1 β , which promotes the synthetic phenotype in vSMCs. The abnormal deposition of transitional ECM proteins such as fibronectin and vitronectin augment the inflammatory response in vSMCs and endothelial cells mediated by integrins α 5 β 1 and α v β 3 signaling. The MMPs released by dedifferentiated vSMCs promotes ECM degradation and thins the fibrous cap.

Panel D: Atherosclerosis Initiation and Progression: At the initial stage of the atherosclerosis, the persistent retention of LDL, followed by its oxidation, activates endothelial cells and drives leukocyte recruitment. Along with the appearance of foam cells, vSMCs migrate and proliferate towards the intimal layer, expanding lesion size. However, significant death of macrophages, defective efferocytosis of remaining macrophages, vSMC phenotypic switching, and ECM remodeling at the atherosclerotic microenvironment drive the thinning of the fibrous cap, making plaque unstable and prone to rupture.

Key References

- Rohwedder I, Montanez E, Beckmann K, Bengtsson E, Dunér P, Nilsson J, et al. Plasma fibronectin deficiency impedes atherosclerosis progression and fibrous cap formation. *EMBO molecular medicine*. 2012;4(7):564–76.

This study demonstrates that fibronectin has a dual role in atherosclerosis: initially, it drives atherosclerotic lesion formation, but also drives plaque

stabilization by augmenting the recruitment of smooth vascular muscle cells.

- Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *New England journal of medicine*. 2017;377(12):1119–31.

This is a randomized, double-blind trial that tested the inflammatory hypothesis in atherosclerosis using a therapeutic monoclonal antibody targeting interleukin-1 β . It provided invaluable evidence on antiinflammatory therapies as a potential treatment strategy to lower this risk of cardiovascular disease, independent of lipid-level lowering.

- Fidler TP, Dunbar A, Kim E, Hardaway B, Pauli J, Xue C, et al. Suppression of IL-1 β promotes beneficial accumulation of fibroblast-like cells in atherosclerotic plaques in clonal hematopoiesis. *Nature cardiovascular research*. 2024;3(1):60–75.

This study provides a potential mechanistic insight into the beneficial action of anti-inflammatory therapy, interleukin-1 β inhibition with canakinumab, as seen in the CANTOS trial. The study showed IL-1 β -mediated crosstalk between myeloid cell and stromal cells including endothelial, fibroblast, and smooth muscle cells in the setting of clonal hematopoiesis of indeterminate potential. Interestingly, researchers discovered the aggregation of non-smooth muscle fibroblast-like cells at the fibrous cap to stabilize the plaque following anti-IL-1 β .

Acknowledgements The figure was illustrated using BioRender.

Author Contributions R.P and A.Y.J wrote the manuscript and prepared the figure.

Funding This work was supported by a Center for Cardiometabolic Diseases and Sciences Predoctoral Fellowship (RP) and the following National Institutes of Health awards to AYJ: R00HL145131 and R01HL167758.

Data Availability No datasets were generated or analysed during the current study.

Declarations

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects.

Competing Interests The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Williams KJ, Tabas I. The response-to-retention hypothesis of early atherogenesis. *Arterioscler Thromb Vasc Biol.* 1995;15(5):551–61.
- Bäck M, Yurdagul A Jr, Tabas I, Öörni K, Kovanen PT. Inflammation and its resolution in atherosclerosis: mediators and therapeutic opportunities. *Nat Reviews Cardiol.* 2019;16(7):389–406.
- Stroope C, Nettersheim FS, Coon B, Finney AC, Schwartz MA, Ley K, et al. Dysregulated cellular metabolism in atherosclerosis: mediators and therapeutic opportunities. *Nat Metabolism.* 2024;6(4):617–38.
- Tabas I, Williams KJ, Borén J. Subendothelial lipoprotein retention as the initiating process in atherosclerosis: update and therapeutic implications. *Circulation.* 2007;116(16):1832–44.
- Boren J, Chapman MJ, Krauss RM, Packard CJ, Bentzon JF, Binder CJ, et al. Low-density lipoproteins cause atherosclerotic cardiovascular disease: pathophysiological, genetic, and therapeutic insights: a consensus statement from the European atherosclerosis society consensus panel. *Eur Heart J.* 2020;41(24):2313–30.
- Galkina E, Ley K. Immune and inflammatory mechanisms of atherosclerosis. *Annu Rev Immunol.* 2009;27(1):165–97.
- Yurdagul A Jr, Finney AC, Woolard MD, Orr AW. The arterial microenvironment: the where and why of atherosclerosis. *Biochem J.* 2016;473(10):1281–95.
- Zhang X, Sessa WC, Fernández-Hernando C. Endothelial transcytosis of lipoproteins in atherosclerosis. *Front Cardiovasc Med.* 2018;5:130.
- Gerrity RG. The role of the monocyte in atherogenesis: I. Transition of blood-borne monocytes into foam cells in fatty lesions. *Am J Pathol.* 1981;103(2):181.
- Schrijvers DM, De Meyer GR, Kockx MM, Herman AG, Martinet W. Phagocytosis of apoptotic cells by macrophages is impaired in atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2005;25(6):1256–61.
- Falk E, Nakano M, Bentzon JF, Finn AV, Virmani R. Update on acute coronary syndromes: the pathologists' view. *Eur Heart J.* 2013;34(10):719–28.
- Finn AV, Nakano M, Narula J, Kolodgie FD, Virmani R. Concept of vulnerable/unstable plaque. *Arterioscler Thromb Vasc Biol.* 2010;30(7):1282–92.
- Davies MJ, Thomas A. Thrombosis and acute coronary-artery lesions in sudden cardiac ischemic death. *N Engl J Med.* 1984;310(18):1137–40.
- Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory therapy with Canakinumab for atherosclerotic disease. *N Engl J Med.* 2017;377(12):1119–31.
- Fiolet AT, Opstal TS, Mosterd A, Eikelboom JW, Jolly SS, Keech AC, et al. Efficacy and safety of low-dose Colchicine in patients with coronary disease: a systematic review and meta-analysis of randomized trials. *Eur Heart J.* 2021;42(28):2765–75.
- Chistiakov DA, Sobenin IA, Orekhov AN. Vascular extracellular matrix in atherosclerosis. *Cardiol Rev.* 2013;21(6):270–88.
- Hynes RO, Naba A. Overview of the matrisome—an inventory of extracellular matrix constituents and functions. *Cold Spring Harb Perspect Biol.* 2012;4(1):a004903.
- Lu P, Takai K, Weaver VM, Werb Z. Extracellular matrix degradation and remodeling in development and disease. *Cold Spring Harb Perspect Biol.* 2011;3(12):a005058.
- Rohwedder I, Montanez E, Beckmann K, Bengtsson E, Dunér P, Nilsson J, et al. Plasma fibronectin deficiency impedes atherosclerosis progression and fibrous cap formation. *EMBO Mol Med.* 2012;4(7):564–76.
- Lin PK, Davis GE. Extracellular matrix remodeling in vascular disease: defining its regulators and pathological influence. *Arterioscler Thromb Vasc Biol.* 2023;43(9):1599–616.
- Schneller M, Vuori K, Ruoslahti E. $\alpha\beta 3$ integrin associates with activated insulin and PDGF β receptors and potentiates the biological activity of PDGF. *EMBO J.* 1997.
- Sarrazin S, Lamanna WC, Esko JD. Heparan sulfate proteoglycans. *Cold Spring Harb Perspect Biol.* 2011;3(7):a004952.
- Yurdagul A Jr. Crosstalk between macrophages and vascular smooth muscle cells in atherosclerotic plaque stability. *Arteriosclerosis, thrombosis, and vascular biology.* 2022;42(4):372–80.
- Yurdagul A Jr, Orr AW. Blood brothers: hemodynamics and cell–matrix interactions in endothelial function. *Antioxid Redox Signal.* 2016;25(7):415–34.
- Dekker RJ, Van Soest S, Fontijn RD, Salamanca S, De Groot PG, VanBavel E, et al. Prolonged fluid shear stress induces a distinct set of endothelial cell genes, most specifically lung Kruppel-like factor (KLF2). *Blood J Am Soc Hematol.* 2002;100(5):1689–98.
- He M, Huang T-S, Li S, Hong H-C, Chen Z, Martin M, et al. Atheroprotective flow upregulates ITPR3 (inositol 1, 4, 5-trisphosphate receptor 3) in vascular endothelium via KLF4 (Kruppel-like factor 4)-mediated histone modifications. *Arterioscler Thromb Vasc Biol.* 2019;39(5):902–14.
- Tamargo IA, Baek KI, Xu C, Kang DW, Kim Y, Andueza A, et al. HEG1 protects against atherosclerosis by regulating stable flow-induced KLF2/4 expression in endothelial cells. *Circulation.* 2024;149(15):1183–201.
- Sangwung P, Zhou G, Nayak L, Chan ER, Kumar S, Kang D-W, et al. KLF2 and KLF4 control endothelial identity and vascular integrity. *JCI Insight.* 2017;2(4):e91700.
- Kraehling JR, Chidlow JH, Rajagopal C, Sugiyama MG, Fowler JW, Lee MY, et al. Genome-wide RNAi screen reveals ALK1 mediates LDL uptake and transcytosis in endothelial cells. *Nat Commun.* 2016;7(1):13516.
- Wight TN. A role for proteoglycans in vascular disease. *Matrix Biol.* 2018;71:396–420.
- CAMEJO G. The interaction of lipids and lipoproteins with the intercellular matrix of arterial tissue: its possible role in atherogenesis. *Adv Lipid Res.* 1982;19:1–53.
- Khan BV, Harrison DG, Olbrych MT, Alexander RW, Medford RM. Nitric oxide regulates vascular cell adhesion molecule 1 gene expression and redox-sensitive transcriptional events in human vascular endothelial cells. *Proc Natl Acad Sci.* 1996;93(17):9114–9.
- Habas K, Shang L. Alterations in intercellular adhesion molecule 1 (ICAM-1) and vascular cell adhesion molecule 1 (VCAM-1) in human endothelial cells. *Tissue Cell.* 2018;54:139–43.
- Hahn C, Schwartz MA. Mechanotransduction in vascular physiology and atherogenesis. *Nat Rev Mol Cell Biol.* 2009;10(1):53–62.
- Yurdagul A Jr, Green J, Albert P, McInnis MC, Mazar AP, Orr AW. $\alpha 5 \beta 1$ integrin signaling mediates oxidized low-density

- lipoprotein-induced inflammation and early atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2014;34(7):1362–73.
36. Al-Yafeai Z, Yurdagul A Jr, Peretik JM, Alfaidi M, Murphy PA, Orr AW. Endothelial FN (Fibronectin) deposition by A5 β 1 integrins drives atherogenic inflammation. *Arteriosclerosis, thrombosis, and vascular biology.* 2018;38(11):2601–14.
 37. Méndez-Barbero N, Gutiérrez-Muñoz C, Blanco-Colio LM. Cellular crosstalk between endothelial and smooth muscle cells in vascular wall remodeling. *Int J Mol Sci.* 2021;22(14):7284.
 38. Maleszewska M, Moonen J-RA, Huijckman N, van de Sluis B, Krenning G, Harmsen MC. IL-1 β and TGF β 2 synergistically induce endothelial to mesenchymal transition in an NF κ B-dependent manner. *Immunobiology.* 2013;218(4):443–54.
 39. Liang G, Wang S, Shao J, Jin Y-J, Xu L, Yan Y, et al. Tenascin-X mediates flow-induced suppression of EndMT and atherosclerosis. *Circul Res.* 2022;130(11):1647–59.
 40. Climent M, Quintavalle M, Miragoli M, Chen J, Condorelli G, Elia L. TGF β triggers miR-143/145 transfer from smooth muscle cells to endothelial cells, thereby modulating vessel stabilization. *Circul Res.* 2015;116(11):1753–64.
 41. Li M, Qian M, Kyler K, Xu J. Endothelial–vascular smooth muscle cells interactions in atherosclerosis. *Front Cardiovasc Med.* 2018;5:151.
 42. Wall VZ, Bornfeldt KE. Arterial smooth muscle. *Arteriosclerosis, thrombosis, and vascular biology.* 2014;34(10):2175–9.
 43. Hedin U, Bottger BA, Forsberg E, Johansson S, Thyberg J. Diverse effects of fibronectin and laminin on phenotypic properties of cultured arterial smooth muscle cells. *J Cell Biol.* 1988;107(1):307–19.
 44. Kingsley K, Huff J, Rust W, Carroll K, Martinez A, Fitchmun M, et al. ERK1/2 mediates PDGF-BB stimulated vascular smooth muscle cell proliferation and migration on laminin-5. *Biochem Biophys Res Commun.* 2002;293(3):1000–6.
 45. Chahine MN, Blackwood DP, Dibrov E, Richard MN, Pierce GN. Oxidized LDL affects smooth muscle cell growth through MAPK-mediated actions on nuclear protein import. *J Mol Cell Cardiol.* 2009;46(3):431–41.
 46. Orr AW, Lee MY, Lemmon JA, Yurdagul A Jr, Gomez MF, Schoppee Bortz PD, et al. Molecular mechanisms of collagen isotype-specific modulation of smooth muscle cell phenotype. *Arterioscler Thromb Vasc Biol.* 2009;29(2):225–31.
 47. Feil S, Fehrenbacher B, Lukowski R, Essmann F, Schulze-Osthoff K, Schaller M, et al. Transdifferentiation of vascular smooth muscle cells to macrophage-like cells during atherogenesis. *Circul Res.* 2014;115(7):662–7.
 48. Xue Y, Luo M, Hu X, Li X, Shen J, Zhu W, et al. Macrophages regulate vascular smooth muscle cell function during atherosclerosis progression through IL-1 β /STAT3 signaling. *Commun Biol.* 2022;5(1):1316.
 49. Beck-Joseph J, Tabrizian M, Lehoux S, Corrigendum. Molecular interactions between vascular smooth muscle cells and macrophages in atherosclerosis. *Front Cardiovasc Med.* 2024;11:1462284.
 50. Filippov S, Koenig GC, Chun T-H, Hotary KB, Ota I, Bugge TH, et al. MT1-matrix metalloproteinase directs arterial wall invasion and Neointima formation by vascular smooth muscle cells. *J Exp Med.* 2005;202(5):663–71.
 51. Grootaert MO, Bennett MR. Vascular smooth muscle cells in atherosclerosis: time for a re-assessment. *Cardiovascular Res.* 2021;117(11):2326–39.
 52. Long X, Miano JM. Transforming growth factor- β 1 (TGF- β 1) utilizes distinct pathways for the transcriptional activation of MicroRNA 143/145 in human coronary artery smooth muscle cells. *J Biol Chem.* 2011;286(34):30119–29.
 53. Yoshida T, Sinha S, Dandré F, Wamhoff BR, Hoofnagle MH, Kremer BE, et al. Myocardin is a key regulator of CArG-dependent transcription of multiple smooth muscle marker genes. *Circul Res.* 2003;92(8):856–64.
 54. Shankman LS, Gomez D, Cherepanova OA, Salmon M, Alencar GF, Haskins RM, et al. KLF4-dependent phenotypic modulation of smooth muscle cells has a key role in atherosclerotic plaque pathogenesis. *Nat Med.* 2015;21(6):628–37.
 55. Bennett MR, Sinha S, Owens GK. Vascular smooth muscle cells in atherosclerosis. *Circul Res.* 2016;118(4):692–702.
 56. Fidler TP, Dunbar A, Kim E, Hardaway B, Pauli J, Xue C, et al. Suppression of IL-1 β promotes beneficial accumulation of fibroblast-like cells in atherosclerotic plaques in clonal hematopoiesis. *Nat Cardiovasc Res.* 2024;3(1):60–75.
 57. Kumar D, Pandit R, Yurdagul A Jr. Mechanisms of continual efferocytosis by macrophages and its role in mitigating atherosclerosis. *Immunometabolism.* 2023;5(1):e00017.
 58. Gerlach BD, Ampomah PB, Yurdagul A, Liu C, Laurant MC, Wang X, et al. Efferocytosis induces macrophage proliferation to help resolve tissue injury. *Cell Metabol.* 2021;33(12):2445–63. e8.
 59. Fadok VA, Bratton DL, Konowal A, Freed PW, Westcott JY, Henson PM. Macrophages that have ingested apoptotic cells in vitro inhibit Proinflammatory cytokine production through autocrine/paracrine mechanisms involving TGF-beta, PGE2, and PAF. *J Clin Invest.* 1998;101(4):890–8.
 60. Yurdagul A, Subramanian M, Wang X, Crown SB, Ilkayeva OR, Darville L, et al. Macrophage metabolism of apoptotic cell-derived arginine promotes continual efferocytosis and resolution of injury. *Cell Metabol.* 2020;31(3):518–33. e10.
 61. Han X, Kitamoto S, Wang H, Boisvert WA. Interleukin-10 overexpression in macrophages suppresses atherosclerosis in hyperlipidemic mice. *FASEB J.* 2010;24(8):2869.
 62. Halvorsen B, Holm S, Yndestad A, Scholz H, Sagen EL, Nebb H, et al. Interleukin-10 increases reverse cholesterol transport in macrophages through its bidirectional interaction with liver X receptor α . *Biochem Biophys Res Commun.* 2014;450(4):1525–30.
 63. Yurdagul A Jr, Kong N, Gerlach BD, Wang X, Ampomah P, Kuriakose G, et al. ODC (ornithine decarboxylase)-dependent Putrescine synthesis maintains MerTK (MER tyrosine-protein kinase) expression to drive resolution. *Arterioscler Thromb Vasc Biol.* 2021;41(3):e144–59.
 64. Noelia A, Bensinger SJ, Hong C, Beceiro S, Bradley MN, Zelcer N, et al. Apoptotic cells promote their own clearance and immune tolerance through activation of the nuclear receptor LXR. *Immunity.* 2009;31(2):245–58.
 65. Ngai D, Schilperoort M, Tabas I. Efferocytosis-induced lactate enables the proliferation of pro-resolving macrophages to mediate tissue repair. *Nat Metabolism.* 2023;5(12):2206–19.
 66. Dagvadorj J, Naiki Y, Tumurkhuu G, Noman ASM, Iftekar-E-Khuda I, Koide N, et al. Interleukin (IL)-10 attenuates lipopolysaccharide-induced IL-6 production via Inhibition of I κ B- ζ activity by Bcl-3. *Innate Immun.* 2009;15(4):217–24.
 67. Anti-Inflammatory S-R. Cutting edge: A transcriptional repressor. *J Immunol.* 2007;179:7215–9.
 68. Yang Q, Zheng C, Cao J, Cao G, Shou P, Lin L, et al. Spermidine alleviates experimental autoimmune encephalomyelitis through inducing inhibitory macrophages. *Cell Death Differ.* 2016;23(11):1850–61.
 69. Zhang M, Caragine T, Wang H, Cohen PS, Botchkina G, Soda K, et al. Spermine inhibits Proinflammatory cytokine synthesis in human mononuclear cells: a counterregulatory mechanism that restrains the immune response. *J Exp Med.* 1997;185(10):1759–68.
 70. Cai B, Thorp EB, Doran AC, Subramanian M, Sansbury BE, Lin C-S et al. MerTK cleavage limits proresolving mediator biosynthesis and exacerbates tissue inflammation. *Proceedings of the National Academy of Sciences.* 2016;113(23):6526–31.

71. Doran AC, Yurdagul A Jr, Tabas I. Efferocytosis in health and disease. *Nat Rev Immunol*. 2020;20(4):254–67.
72. Shah PK. Molecular mechanisms of plaque instability. *Curr Opin Lipidol*. 2007;18(5):492–9.
73. Kojima Y, Volkmer J-P, McKenna K, Civelek M, Lusis AJ, Miller CL, et al. CD47-blocking antibodies restore phagocytosis and prevent atherosclerosis. *Nature*. 2016;536(7614):86–90.
74. Doran AC, Ozcan L, Cai B, Zheng Z, Fredman G, Rymond CC, et al. CAMKII γ suppresses an efferocytosis pathway in macrophages and promotes atherosclerotic plaque necrosis. *J Clin Invest*. 2017;127(11):4075–89.
75. Cai B, Thorp EB, Doran AC, Sansbury BE, Daemen MJ, Dörweiler B, et al. MerTK receptor cleavage promotes plaque necrosis and defective resolution in atherosclerosis. *J Clin Invest*. 2017;127(2):564–8.
76. Thorp E, Vaisar T, Subramanian M, Mautner L, Blobel C, Tabas I. Shedding of the Mer tyrosine kinase receptor is mediated by ADAM17 protein through a pathway involving reactive oxygen species, protein kinase C δ , and p38 mitogen-activated protein kinase (MAPK). *J Biol Chem*. 2011;286(38):33335–44.
77. He C, Medley SC, Hu T, Hinsdale ME, Lupu F, Virmani R, et al. PDGFR β signalling regulates local inflammation and synergizes with hypercholesterolaemia to promote atherosclerosis. *Nat Commun*. 2015;6(1):7770.
78. Owens GK. Regulation of differentiation of vascular smooth muscle cells. *Physiol Rev*. 1995;75(3):487–517.
79. Grainger DJ. Transforming growth factor B and atherosclerosis: so far, so good for the protective cytokine hypothesis. *Arteriosclerosis, thrombosis, and vascular biology*. 2004;24(3):399–404.
80. Feinberg MW, Watanabe M, Lebedeva MA, Depina AS, Hanai J-i, Mammoto T, et al. Transforming growth factor- β 1 inhibition of vascular smooth muscle cell activation is mediated via Smad3. *J Biol Chem*. 2004;279(16):16388–93.
81. Yang L, Gao L, Nickel T, Yang J, Zhou J, Gilbertsen A, et al. Lactate promotes synthetic phenotype in vascular smooth muscle cells. *Circul Res*. 2017;121(11):1251–62.
82. Grossi M, Rippe C, Sathanoori R, Swärd K, Forte A, Erlinge D, et al. Vascular smooth muscle cell proliferation depends on caveolin-1-regulated polyamine uptake. *Biosci Rep*. 2014;34(6):e00153.
83. Miteva K, Burger F, Baptista D, Roth A, Da Silva R, Mach F, et al. Effect of monocytes on NLRP3 inflammasome activation in vascular smooth muscle cells phenotypic switch and foam cells formation in atherosclerosis. *Atherosclerosis*. 2020;315:e20.
84. Gough PJ, Gomez IG, Wille PT, Raines EW. Macrophage expression of active MMP-9 induces acute plaque disruption in apoE-deficient mice. *J Clin Invest*. 2006;116(1):59–69.
85. Quillard T, Tesmenitsky Y, Croce K, Travers R, Shvartz E, Koskinas KC, et al. Selective Inhibition of matrix metalloproteinase-13 increases collagen content of established mouse atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2011;31(11):2464–72.
86. Ren W, Wang Z, Wang J, Wu Z, Ren Q, Yu A, et al. IL-5 overexpression attenuates aortic dissection by reducing inflammation and smooth muscle cell apoptosis. *Life Sci*. 2020;241:117144.
87. Ikeda K, Souma Y, Akakabe Y, Kitamura Y, Matsuo K, Shimoda Y, et al. Macrophages play a unique role in the plaque calcification by enhancing the osteogenic signals exerted by vascular smooth muscle cells. *Biochem Biophys Res Commun*. 2012;425(1):39–44.
88. Tang J, Li T, Xiong X, Yang Q, Su Z, Zheng M, et al. Colchicine delivered by a novel nanoparticle platform alleviates atherosclerosis by targeted inhibition of NF- κ B/NLRP3 pathways in inflammatory endothelial cells. *J Nanobiotechnol*. 2023;21(1):460.
89. Boada C, Zinger A, Tsao C, Zhao P, Martinez JO, Hartman K, et al. Rapamycin-loaded biomimetic nanoparticles reverse vascular inflammation. *Circul Res*. 2020;126(1):25–37.
90. Kamaly N, Fredman G, Fojas JJR, Subramanian M, Choi WI, Zepeda K, et al. Targeted interleukin-10 nanotherapeutics developed with a microfluidic chip enhance resolution of inflammation in advanced atherosclerosis. *ACS Nano*. 2016;10(5):5280–92.
91. Qiu S, Liu J, Chen J, Li Y, Bu T, Li Z, et al. Targeted delivery of MerTK protein via cell membrane engineered nanoparticle enhances efferocytosis and attenuates atherosclerosis in diabetic ApoE $^{-/-}$ mice. *J Nanobiotechnol*. 2024;22(1):178.
92. Sager HB, Dutta P, Dahlman JE, Hulsmans M, Courties G, Sun Y, et al. RNAi targeting multiple cell adhesion molecules reduces immune cell recruitment and vascular inflammation after myocardial infarction. *Sci Transl Med*. 2016;8(342):ra34280–80.
93. Yu M, Amengual J, Menon A, Kamaly N, Zhou F, Xu X, et al. Targeted nanotherapeutics encapsulating liver X receptor agonist GW3965 enhance antiatherogenic effects without adverse effects on hepatic lipid metabolism in Ldlr $^{-/-}$ mice. *Adv Healthc Mater*. 2017;6(20):1700313.
94. Nakashiro S, Matoba T, Umez U, Koga J-i, Tokutome M, Katsuki S, et al. Pioglitazone-incorporated nanoparticles prevent plaque destabilization and rupture by regulating monocyte/macrophage differentiation in ApoE $^{-/-}$ mice. *Arterioscler Thromb Vasc Biol*. 2016;36(3):491–500.
95. Flores AM, Hosseini-Nassab N, Jarr K-U, Ye J, Zhu X, Wirka R, et al. Pro-efferocytic nanoparticles are specifically taken up by lesional macrophages and prevent atherosclerosis. *Nat Nanotechnol*. 2020;15(2):154–61.
96. Dalli J, Consalvo AP, Ray V, Di Filippo C, D'Amico M, Mehta N, et al. Proresolving and tissue-protective actions of Annexin A1-based cleavage-resistant peptides are mediated by formyl peptide receptor 2/lipoxin A4 receptor. *J Immunol*. 2013;190(12):6478–87.
97. Chuang ST, Stein JB, Nevins S, Kilic Bektas C, Choi HK, Ko WK, et al. Enhancing CAR macrophage efferocytosis via surface engineered lipid nanoparticles targeting LXR signaling. *Adv Mater*. 2024;36(19):2308377.
98. Geng S, Lu R, Zhang Y, Wu Y, Xie L, Caldwell BA, et al. Monocytes reprogrammed by 4-PBA potently contribute to the resolution of inflammation and atherosclerosis. *Circul Res*. 2024;135(8):856–72.
99. Williams JW, Winkels H, Durant CP, Zaitsev K, Ghosheh Y, Ley K. Single cell RNA sequencing in atherosclerosis research. *Circul Res*. 2020;126(9):1112–26.
100. Seeley EH, Liu Z, Yuan S, Stroope C, Cockerham E, Rashdan NA, et al. Spatially resolved metabolites in stable and unstable human atherosclerotic plaques identified by mass spectrometry imaging. *Arterioscler Thromb Vasc Biol*. 2023;43(9):1626–35.
101. Muller WA. Getting leukocytes to the site of inflammation. *Vet Pathol*. 2013;50(1):7–22.
102. Langer HF, Chavakis T. Leukocyte–endothelial interactions in inflammation. *J Cell Mol Med*. 2009;13(7):1211–20.
103. Ebnet K, Suzuki A, Ohno S, Vestweber D. Junctional adhesion molecules (JAMs): more molecules with dual functions? *J Cell Sci*. 2004;117(1):19–29.
104. Bhardwaj S, Roy H, Babu M, Shibuya M, Yla-Herttuala S. Adenoviral gene transfer of VEGFR-2 specific VEGF-E chimera induces MCP-1 expression in vascular smooth muscle cells and enhances neointimal formation. *Atherosclerosis*. 2011;219(1):84–91.
105. Hayakawa E, Yoshimoto T, Sekizawa N, Sugiyama T, Hirata Y. Overexpression of receptor for advanced glycation end products induces monocyte chemoattractant protein-1 expression in rat vascular smooth muscle cell line. *J Atheroscler Thromb*. 2012;19(1):13–22.
106. Hara T, Phuon PT, Fukuda D, Yamaguchi K, Murata C, Nishimoto S, et al. Protease-activated receptor-2 plays a critical role in vascular inflammation and atherosclerosis in Apolipoprotein E-deficient mice. *Circulation*. 2018;138(16):1706–19.

107. Butoi E, Gan A, Tucureanu M, Stan D, Macarie R, Constantinescu C, et al. Cross-talk between macrophages and smooth muscle cells impairs collagen and metalloprotease synthesis and promotes angiogenesis. *Biochim Et Biophys Acta (BBA)-Molecular Cell Res.* 2016;1863(7):1568–78.
108. Cheng CK, Yi M, Wang L, Huang Y. Role of gasdermin D in inflammatory diseases: from mechanism to therapeutics. *Front Immunol.* 2024;15:1456244.
109. Nguyen M-A, Karunakaran D, Geoffrion M, Cheng HS, Tandoc K, Perisic Matic L, et al. Extracellular vesicles secreted by atherogenic macrophages transfer MicroRNA to inhibit cell migration. *Arterioscler Thromb Vasc Biol.* 2018;38(1):49–63.
110. Weinert S, Poitz DM, Auffermann-Gretzinger S, Eger L, Herold J, Medunjanin S, et al. The lysosomal transfer of LDL/cholesterol from macrophages into vascular smooth muscle cells induces their phenotypic alteration. *Cardiovascular Res.* 2013;97(3):544–52.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.