



## Original Article

# Phenotypic profile of pulmonary aspergillosis and associated cellular immunity among people living with human immunodeficiency virus in Maiduguri, Nigeria

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## ABSTRACT

**Objective:** *Aspergillus* causes many forms of pulmonary infectious diseases ranging from colonization (noninvasive) to invasive aspergillosis. This largely depends on the underlying host's lung health and immune status. Pulmonary aspergillosis (PA), especially the invasive form, occurs as opportunistic to human immunodeficiency virus (HIV) as a result of cluster of differentiation (CD)4+ lymphopenia. The majority of patients with comorbid HIV and aspergillosis go undiagnosed. This study aimed to isolate, identify the etiologies, and determine the prevalence of PA among HIV-infected persons with a productive cough (at least <2 weeks) at the HIV Clinics of the University of Maiduguri Teaching Hospital, Nigeria. **Materials and Methods:** After ethical approval, three consecutive early morning sputum samples were collected from patients with negative tuberculosis results. The samples were individually inoculated onto Sabouraud dextrose agar supplemented with chloramphenicol and cycloheximide in duplicate for 7 days at 37°C and 25°C, respectively. The fungal isolates were examined morphologically and microscopically and identified using the standard biochemical reagents. CD4+ cell counts were performed using flow cytometry. Self-administered questionnaires were used to assess the patients data. All patients were antiretroviral naïve. **Results:** The prevalence of PA was 12.7% in these 150 patients. Of the 19 fungal culture-positive individuals, *Aspergillus fumigatus* accounted for the highest proportion of the isolates (8, 42.1%) followed by *Aspergillus niger* (5, 26.3%), *Aspergillus flavus* (4, 21.1%), and *Aspergillus terreus* (2, 10.5%). Based on the assessment of functionality of cellular immunity, HIV participants who were negative for PA (131/150) had significantly higher mean  $\pm$  standard deviation CD4 T-cell counts (245.65  $\pm$  178.32 cells/mL) than those with aspergillosis (126.13  $\pm$  105.27 cells/mL) ( $P = 0.0051$ ). PA was relatively highest among patients with CD4+ cell counts <200 cells/mL (12, 34.3%) followed by those with CD4+ cell counts between 200 and 350 cells/mL (5, 9.6%) and least among those with CD4+ cell counts >350 cells/mL (2, 3.2%). The Chi-square test showed a significant association between the prevalence of PA and the CD4+ cell count, age, and gender ( $P < 0.05$ ) but not with occupation or education level ( $P > 0.05$ ). **Conclusion:** The findings from this study indicate that *Aspergillus* spp. is a significant etiology of acute productive cough in people living with HIV and this is related to the CD4+ cell count of coinfecting persons.

**KEYWORDS:** *Aspergillus*, Chest diseases, Coinfection, Human immunodeficiency virus

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## INTRODUCTION

Pulmonary aspergillosis (PA) has become a vital cause of pulmonary fungal infections [1]. The disease is mostly caused by *Aspergillus fumigatus* and can be invasive, semi-invasive, or noninvasive [2]. The incidence and prevalence of PA are on the rise due to an increase in the number of immunocompromised patients, but the disease is largely

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unexplored in developing countries. Because of this, it has emerged as a worldwide healthcare problem [1,3,4].

PA occurs mostly in immunosuppressed individuals and, if not treated early, eventually leads to invasive PA as a result of hematogenous dissemination from the lungs [3,5]. It rarely occurs in immunocompetent individuals but does occur in the presence of other pulmonary or systemic abnormalities such as fibrotic lung disease or treatment with corticosteroids, parenteral nutrition, use of multiple antibiotics, and prolonged hospitalization [1,2,6-8]. PA can also coexist or act as a sequel to a previous pulmonary infection, and its symptoms such as nonproductive cough, pleuritic chest pain, fever, and dyspnea are nonpathognomonic. This results in a diagnostic challenge for clinicians who either misdiagnose it or diagnose the disease after the death of the patient [9-11]. Regardless of severe immunosuppression that accompanies advanced human immunodeficiency virus (HIV) infection, there are relatively few cases of PA in HIV patients [12]. This might be due to either highly active antiretroviral therapy (HAART) or the phagocytic cell function that accompanies T-cell dysfunction. However, PA develops in HIV patients with underlying risk factors such as neutropenia, corticosteroid treatment, or even a low cluster of differentiation (CD) 4 cell count (<50 cells/mL) in advanced HIV infection [12].

A healthy, functional innate immune system plays a crucial role in preventing *Aspergillus* infection. This pivotal role for the innate immune system makes it a main research focus in studying the pathogenesis of aspergillosis [11,12]. Although sometimes overshadowed by innate immune response, the adaptive immune response, and in particular T-helper (Th) responses, is also a key player in host defense against *Aspergillus*. Virtually, all Th subsets have been described to play a role during aspergillosis, with the Th1 response being crucial for fungal clearance. Dendritic cells are the key players in bridging the innate and adaptive immune response against *Aspergillus* spp. by specifically activating naïve CD4+ T-cells and triggering their differentiation into disparate lineages of effector cells. Clinically, depletion of CD4 T-cell count is a major risk factor for contacting aspergillosis [11,12]. The diagnosis of invasive aspergillosis rests upon a sputum culture and histopathology, antigen detection serology, and polymerase chain reaction study [12].

This study was carried out to isolate, identify the etiologies, and determine the prevalence of PA and associated CD4+ T-cell counts among HIV-infected persons with a productive cough (at least <2 weeks) at the HIV Clinic of the University of Maiduguri Teaching Hospital (UMTH), Nigeria.

## MATERIALS AND METHODS

### Study design

This was a cross-sectional study conducted using blood and sputum samples from 150 antiretroviral therapy naïve HIV-infected individuals. All participants aged between 9 and 55 years were screened and their HIV status was confirmed using Uni-Gold Recombigen® HIV-1/2 (Trinity Biotech, Wicklow, Ireland) and Determine™ (Alere, Auckland City, New Zealand) proprietary reagents. These samples were collected at the HIV Clinic of the UMTH, Maiduguri, Nigeria.

### Study area

This hospital-based research was conducted at UMTH in Borno State, Nigeria. New cases of HIV infections are diagnosed in this tertiary hospital, and those undergoing therapy are also monitored. Sputum samples were cultured for fungal isolates at the Medical Microbiology Laboratory, while the CD4 T-cell counts in the blood samples were analyzed at the Immunology Laboratory at UMTH.

### Informed consent and ethical approval

The study was explained to the enrolled participants, and they gave their written informed consent. Parents/guardians gave approval for the participants who were children. Participants were all confirmed seropositive for HIV but without commencement of antiretroviral therapy. A structured questionnaire was used to obtain bio-data from these participants in accordance with the Declaration of Helsinki. Parents/guardians filled questionnaires on behalf of their children. Those who were seronegative for HIV, had not given consent, or had other etiologies with similar pathologies, including active tuberculosis and nontuberculosis mycobacterial infection, as well as other bacterial infections were excluded from this study. Ethical approval was obtained from the Ethical Research Committee of the UMTH, Maiduguri, Borno State. Data generated were anonymously analyzed throughout the study.

### Sample size calculation

The sample size for this study was derived using data from a PA cross-sectional study conducted in Maiduguri, Nigeria, by Adisa *et al.* [13]. Thus, the minimum sample size required for this study was 116 using a 5% error margin and 95% confidence interval. However, statistical credence was given to this study by increasing the sample size to 150. Therefore, a total of 150 volunteers were recruited as participants for this study.

### Sample collection and preparation

Whole-blood samples of 4 mL were collected aseptically in ethylenediaminetetraacetic acid containers and used for CD4+ cell counts. Furthermore, three consecutive early morning sputum samples were obtained from the participants in a sterile plain container for fungal analysis.

Samples were collected between April 2016 and August 2016. Blood and sputum were analyzed within 1 h of collection.

### Laboratory analytical procedures

#### Culture and identification of fungal isolates

The sputum samples were inoculated and aerobically incubated in duplicate for 7 days using Sabouraud dextrose agar supplemented separately with chloramphenicol and cycloheximide at 37°C and 25°C, respectively. The fungal isolates were examined morphologically and microscopically and identified using standard biochemical reagents for mycology such as starch and cellulose hydrolysis tests.

#### Procedure for fungal identification

##### Potassium hydroxide wet mount

A large drop of potassium hydroxide solution was placed on a clean glass slide, and a small quantity of the specimen was transferred onto it. The preparation was covered with a clean cover slip, avoiding air bubbles. The slide was kept at

room temperature in moisture for 15 min and was observed under  $\times 10$  and  $\times 40$  objectives [14].

**Lactophenol cotton blue staining (needle mount preparation)**

A drop of lactophenol cotton blue stain was placed on a clean glass slide and a small fragment of a colony (cottony, woolly, or powdery) picked from the midpoint of the culture was placed on it and covered with a clean coverslip. After excess stain was removed by blotting, the slide was examined under  $\times 10$  and  $\times 40$  objectives. Features seen in the stained slide were compared with the established characteristic fungal features such as vegetative and reproductive structures using mycology [15,16].

**Starch hydrolysis test**

Starch agar medium (starch 20.0 g/L, peptone 5.0 g/L, yeast extract 3.0 g/L, agar 15.0 g/L; pH 7.0) was inoculated with isolated fungal cultures and incubated at 25°C for 5–7 days in an inverted position. The surface was flooded with iodine solution for 30 s, and zones around the fungal growth were observed.

**Cellulose hydrolysis test**

Czapek-mineral salt agar medium (KCl 0.5 g/L,  $K_2HPO_4$  1.0 g/L,  $NaNO_3$  2.0 g/L,  $MgSO_4 \cdot 7H_2O$  0.5 g/L, peptone 2.0 g/L, carboxymethyl cellulose 5.0 g/L) was complemented with agar 2% and autoclaved at 15 psi for 15 min and was poured in sterile Petri plates (25 mL/plate) by laminar flow and allowed to solidify. Fungal cultures were inoculated and incubated for 5 days at 35°C in an inverted position. The plate surface was then flooded with a 1% aqueous solution of hexadecyltrimethylammonium bromide for 30 s and observed for clear zone formation around the fungal growth.

**Determination of cluster of differentiation 4+ cell count**

Based on the manufacturer’s instructions, the CD4+ cell counts in the whole blood were analyzed using a Partec™ CyFlow Analyzer (Sysmex, Norderstedt, Germany) Model SL3. This device used the principle of light scattering property (based on dissimilarity in cell size or granularity) and the fluorescence of cells following staining with monoclonal antibodies to markers on the cell surface bound to fluorescent dyes. Cell populations of interest were then gated after identification. Absolute CD4+ cell counts were subsequently analyzed using a single-platform technique.

**Statistical analysis**

Data obtained were analyzed using SPSS software version 24 (IBM Corporation, Armonk, NY, USA) and were presented as percentages and mean  $\pm$  standard deviation (SD). Student’s *t*-test was used to compare continuous variables, while the Chi-square was used to compare categorical variables. A  $P < 0.05$  at a confidence interval of 95% was considered statistically significant.

**RESULTS**

Of the 150 samples studied, 19 (12.7%) were culture positive for PA. Two (11%) of these 19 specimens were positive for *Aspergillus terreus*, 4 (21%) were positive for *Aspergillus flavus*, 5 (26%) were positive for *Aspergillus niger*, and 8 (42%) were positive for *A. fumigatus* [Figure 1].

Based on the assessment of functionality of cellular immunity, HIV participants who were negative for PA (131/150) had significantly higher mean  $\pm$  SD CD4 T-cell counts (245.65  $\pm$  178.32 cells/mL) than those with PA (126.13  $\pm$  105.27 cells/mL) ( $P = 0.0051$ ) [Table 1]. PA was diagnosed in 12 of 35 participants (34.3%) with CD4+ cell counts  $< 200$  cells/mL, 5 of 52 (9.6%) with counts of 200–350 cells/mL, and 2 of 63 (3.2%) with counts  $> 350$  cells/mL. Statistically significant differences were observed between CD4+ cell counts and the occurrence of pulmonary aspergillosis ( $\chi^2 = 20.356$ ,  $P = 0.00004$ ) [Table 2].

Twelve of the 19 patients with PA were male with a prevalence of 22.6% (14/62), which was higher than that of their female counterparts with a seroprevalence of 5.7% (5/88). Seropositivity was decreased in participants aged 25–34 years (25.7%, 9/35) compared with those

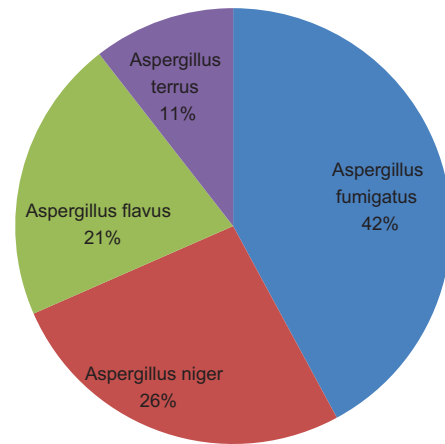


Figure 1: Phenotypic profile of *Aspergillus* spp. isolated from participants

Table 1: Comparison of CD4+ T-cell counts in human immunodeficiency virus-infected participants with and without pulmonary aspergillosis

Group	CD4+ T-cell counts (cells/mL)	
	Range	Mean $\pm$ SD
Participants with pulmonary aspergillosis (n=19)	53-374	126.13 $\pm$ 105.27
Participants without pulmonary aspergillosis (n=131)	125-789	245.65 $\pm$ 178.32
<i>t</i> -test value	-	2.8453
<i>P</i>	-	0.0051*

SD: Standard deviation, \*Significant

Table 2: Prevalence of aspergillosis based on CD4+ T-cell count range

CD4+ T-cell count (cells/mL)	Number of participants with that particular CD4+ T-cell count	Number with culture positive (%)	$\chi^2$	<i>P</i> value for linear trend
$< 200$	35	12 (34.3)	20.356	0.00004*
200-350	52	5 (9.6)		
$> 350$	63	2 (3.2)		
Total	150	19 (12.7)		

\*Significant

aged >44 years (12.1%, 4/33). According to educational status, the highest prevalence of PA was 28.6% (2/7) in those who had attended college, while the lowest prevalence of 6.8% (5/73) was recorded in those with a high-school education. In terms of occupation, the highest prevalence of 25.0% (2/8) was found in unemployed participants, while the lowest prevalence of 6.2% (4/65) was reported among civil servants. Statistically significant differences were observed between age ( $P = 0.00459$ ) and gender ( $P = 0.0022$ ) and the occurrence of PA [Table 3].

**DISCUSSION**

*Aspergillus* spp. ubiquitously occur as saprobes and are commonly present in the soil. Recently, they have been associated with disease in humans, thus making them a parasite as well [1,10]. PA caused by *Aspergillus* spp. is devastating in immunocompromised patients, and HIV infection is one of the major risk factors due to CD4 lymphopenia [12]. The symptoms of the disease, which include acute productive cough, pleuritic chest pain, hemoptysis, dyspnea, and fever, are nonspecific as they resemble the symptoms similar diseases; this makes the disease insidious and it is a diagnostic challenge [2].

In this study, a persistent acute productive cough, which is one of the common symptoms of the disease, was studied among HIV-infected persons. Recently, the incidence of pulmonary disease in HIV-infected persons with or without underlying risk factors has been reported and associated with mortality [11,17-19].

All patients tested were experiencing a persistent acute productive cough, similar to the reports of Denning *et al.* [20] and Blot *et al.* [21], confirming acute productive cough as one of the nonspecific symptoms of the disease. The 12.7% prevalence of PA in this study was not the same as the infection rates reported in two other studies from Southeastern Nigeria by Ogba *et al.* [22] and Ochiabuto *et al.* [23], who independently reported the prevalence rates of 6.3% and 27.9%, respectively, in HIV patients with a productive cough. These discrepancies could be a result of the positive influence of HAART among these participants as all were volunteers enrolled from treatment centers.

Most studies report that *A. fumigatus* is the major cause of PA among *Aspergillus* species [10]. In this study, *A. fumigatus* accounted for 42.1% of isolates, followed by *A. niger*, *A. flavus*, and *A. terreus*. These findings are similar to the reports of Ved *et al.* [1] and Ochiabuto *et al.* [23]. However, Chandwani *et al.* [24] reported that *A. niger* was the most common species isolated in their study. These variations could be attributed to variations in geographical locations and ecological niches. Second, *A. fumigatus* has been regarded by researchers [22,25] as the most invasive *Aspergillus* species due to the high frequency of clinical manifestations (cough, chest pain, fever, hemoptysis, weight loss, and dyspnea) associated with this pathogen as observed in the participants enrolled in this study.

In this study, the mean CD4+ cell count of the participants with PA was  $126.13 \pm 105.7$  cells/mL of the blood. Participants positive for PA generally had lower CD4 counts (<200 cells/mL) than their noninfected counterparts. This finding is in conformity with previous studies [22,24,26,27] that reported aspergillosis in HIV patients with mean CD4+ cell counts <200 cells/mL. Cases of immunosuppression in HIV patients with evidence of low CD4+ cell counts (<200 cells/mL) are associated with the dysfunction or depletion in numbers of macrophages and neutrophils which are considered as sentinels in the eradication of opportunistic infections, including that of fungi. Due to the depletion of these cells, the immune system fails to efficiently eradicate these *Aspergillus* species, and consequently, this encourages their survival. HIV-*Aspergillus* coinfection has been observed in patients with CD4+ cell counts much lower than 100 cells/mL [28], patients with a history of other HIV-opportunistic coinfections, and untreated (HAART naïve) HIV patients [24,27].

The mean age of the participants in the study was  $22.4 \pm 13.0$  years, and majority of the participants positive for PA were between 25 and 34 years. This finding is in agreement with Adisa *et al.* [13]. According to Ogba *et al.* [22], who reported a similar observation, individuals in this age range have high sexual activity which could be responsible for HIV-*Aspergillus* coinfection.

In this study, more males than females were infected. This is in line with the findings of Njunda *et al.* [4], who also reported a higher prevalence of *Aspergillus* in males than females (15.4% vs. 14.7%), but this was not statistically significant ( $P = 1.0000$ ). The relatively higher prevalence ( $P = 0.0022$ ) in males might be due to the fact that men often work in areas where *Aspergillus* is present, such as in the soil.

**Table 3: Sociodemographic risk factors for pulmonary aspergillosis among study participants**

Variables	Observation	Number of participants	Number of positive (%)	P
Age (years), mean±SD	22.4±13.0	NA	NA	
Gender	Male	62	14 (22.6)	0.0022*
	Female	88	5 (5.7)	
	Total	150	19 (12.7)	
Age (years), range	<15	8	3 (37.5)	0.00459*
	15-24	12	1 (8.3)	
	25-34	35	9 (25.7)	
	35-44	62	2 (3.2)	
	>44	33	4 (12.1)	
	Total	150		
Educational level	No formal education	42	8 (19.0)	0.142
	Primary	28	4 (14.3)	
	High school/secondary	73	5 (6.8)	
	College	7	2 (28.6)	
	Total	150		
Occupation	Farmer	18	3 (16.7)	0.2913
	Sex worker	21	4 (19.0)	
	Student	38	6 (15.8)	
	Civil servant	65	4 (6.2)	
	Unemployed	8	2 (25.0)	
	Total	150		

SD: Standard deviation, NA: Not available, \*Significant



More of the participants enrolled in this study had no formal education, and this could have had an influence on their knowledge of the risk factors related to PA and the need to screen for this pathogen as observed by Jhun *et al.* [29]. As with occupation, there was no statistical difference in the level of education ( $P = 0.2913$ ) in infection with *Aspergillus* species in this study.

## CONCLUSION

This study revealed the various etiologies of PA, their rate of occurrence, and association with immune status in HIV-seropositive individuals. CD4+ cell counts could be involved in mycological infection in HIV-infected persons. Even though PA cannot be diagnosed solely using cultural isolation of fungi, findings from this study warrant further studies, especially histological, imaging, and serum *Aspergillus* galactomannan testing to determine the association with fungal cultural results and the CD4 T-cell count in the diagnosis of PA.

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Nil.

## Conflicts of interest

There are no conflicts of interest.

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