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Cognitive and emotional alterations in *App* knock-in mouse models of A β amyloidosis

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Abstract

Background: Alzheimer's disease (AD), the most common cause of dementia, is characterized by the progressive deposition of amyloid- β (A β) peptides and neurofibrillary tangles. Mouse models of A β amyloidosis generated by knock-in (KI) of a humanized A β sequence provide distinct advantages over traditional transgenic models that rely on overexpression of amyloid precursor protein (APP). In *App*-KI mice, three familial AD-associated mutations were introduced into the endogenous mouse *App* locus to recapitulate A β pathology observed in AD: the Swedish (NL) mutation, which elevates total A β production; the Beyreuther/Iberian (F) mutation, which increases the A β 42/A β 40 ratio; and the Arctic (G) mutation, which promotes A β aggregation. *App*^{NL-G-F} mice harbor all three mutations and develop progressive A β amyloidosis and neuroinflammatory response in broader brain areas, whereas *App*^{NL} mice carrying only the Swedish mutation exhibit no overt AD-related pathological changes. To identify behavioral alterations associated with A β pathology, we assessed emotional and cognitive domains of *App*^{NL-G-F} and *App*^{NL} mice at different time points, using the elevated plus maze, contextual fear conditioning, and Barnes maze tasks.

Results: Assessments of emotional domains revealed that, in comparison with wild-type (WT) C57BL/6J mice, *App*^{NL-G-F/NL-G-F} mice exhibited anxiolytic-like behavior that was detectable from 6 months of age. By contrast, *App*^{NL/NL} mice exhibited anxiogenic-like behavior from 15 months of age. In the contextual fear conditioning task, both *App*^{NL/NL} and *App*^{NL-G-F/NL-G-F} mice exhibited intact learning and memory up to 15–18 months of age, whereas *App*^{NL-G-F/NL-G-F} mice exhibited hyper-reactivity to painful stimuli. In the Barnes maze task, *App*^{NL-G-F/NL-G-F} mice exhibited a subtle decline in spatial learning ability at 8 months of age, but retained normal memory functions.

Conclusion: *App*^{NL/NL} and *App*^{NL-G-F/NL-G-F} mice exhibit behavioral changes associated with non-cognitive, emotional domains before the onset of definitive cognitive deficits. Our observations consistently indicate that *App*^{NL-G-F/NL-G-F} mice represent a model for preclinical AD. These mice are useful for the study of AD prevention rather than treatment after neurodegeneration.

Keywords: Alzheimer's disease, Amyloid precursor protein, Knock-in mouse model, Emotional behavior, Spatial learning

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Background

Alzheimer's disease (AD) is characterized by a progressive decline in cognitive functions, usually starting with memory complaints, and eventually leading to multiple cognitive, neuropsychological, and behavioral deficits [1, 2]. The neuropathology of AD begins before overt cognitive symptoms, including the accumulation of amyloid- β peptide (A β) as extracellular plaques, aggregation of hyperphosphorylated tau as intracellular neurofibrillary tangles (NFTs), and activation of multiple neuroinflammatory pathways [3–5]. These brain pathologies are thought to induce neuronal cell loss in the hippocampus and cerebral cortex [3–5]. However, the etiology of AD is still not fully clarified, and many fundamental questions remain unanswered.

Mouse models of AD pathology are critical research tools for testing potential therapeutic approaches to AD and investigating the molecular mechanisms underlying AD pathogenesis [6, 7]. Several transgenic mouse lines overexpressing amyloid precursor protein (APP) have been developed as experimental models for A β amyloidosis [8, 9]. However, non-physiological overexpression of APP results in overproduction of various APP fragments in addition to A β [8], making it technically difficult to distinguish the pathophysiological effects caused by A β from those caused by other APP fragments [8, 10, 11]. Moreover, overexpression of APP causes memory impairment without A β deposition in some *App* transgenic mice [8, 11], suggesting that the brains of these transgenic mouse models may have pathophysiological properties that are not relevant to AD pathogenesis.

To produce A β pathology without non-physiological overexpression of APP in the mouse brain, alternative mouse models have been generated utilizing an *App* knock-in (KI) strategy [12] in which the murine A β sequence was humanized by changing three amino acids that differ between the mouse and human proteins. In addition, three familial AD-associated mutations were introduced into the endogenous mouse *App* locus: the Swedish (NL) mutation, which elevates total A β production [13]; the Beyreuther/Iberian (F) mutation, which increases the A β 42/A β 40 ratio [14, 15]; and the Arctic (G) mutation, which promotes A β aggregation [16, 17]. In *App*^{NL-G-F} mice, which harbor all three mutations within the A β sequence, A β amyloidosis is aggressively accelerated and neuroinflammation is observed in subcortical structures and cortical regions [12, 18]. By contrast, *App*^{NL} mice that carry only the Swedish mutation produce significantly higher levels of A β 40 and A β 42 but exhibit no overt AD-related brain pathology such as extracellular A β plaques or neuroinflammation [12, 18]. None of these *App*-KI mice exhibit tau pathology

or severe neuron loss, suggesting that they are models of preclinical AD [11].

Recent reports demonstrated that *App*-KI mice exhibit a reduction in the number of hippocampal mushroom spines [19, 20] and disruption of neural circuit activities organized by gamma oscillations in the medial entorhinal cortex [21]. They also revealed new mechanisms underlying A β pathology: genetic deletion of the orphan G protein GPR3, which regulates γ -secretase activity and A β generation, attenuates A β pathology [22], whereas ablation of kallikrein-related peptidase 7 (KLK7), an astrocyte-derived A β -degrading enzyme, accelerates A β pathologies in the brains of *App*-KI mice [23]. To further understand the utility of the *App*-KI mouse models for basic and translational research, it is crucial to obtain detailed information on their behavioral phenotypes, including cognitive and non-cognitive comorbidity related to AD [7, 12, 18, 24–26].

To investigate the behavioral changes associated with A β pathology, we searched for alterations in cognitive and emotional domains specifically present in *App*^{NL-G-F/NL-G-F} mice. As our experimental paradigms, we used the elevated plus maze (EPM), contextual fear conditioning (CFC), and Barnes maze (BM) tasks. Analysis with EPM revealed that *App*^{NL-G-F/NL-G-F} mice exhibited robust anxiolytic-like behaviors, whereas *App*^{NL/NL} mice exhibited anxiogenic-like behaviors, in comparison with wild-type (WT) C57BL/6J mice. In CFC and BM, no significant learning and memory deficits were observed in *App*^{NL-G-F/NL-G-F} or *App*^{NL/NL} mice, whereas *App*^{NL-G-F/NL-G-F} mice exhibited a subtle decline in spatial learning ability in the BM. These results suggest that *App*^{NL-G-F/NL-G-F} and *App*^{NL/NL} mice exhibit significant changes in anxiety-related behaviors, with minimal alterations in learning ability and memory. Our results provide information about behavioral readouts in *App*-KI mice that will be useful for future basic and translational research.

Results

In *App*^{NL-G-F/NL-G-F} mice, age-dependent cortical A β amyloidosis began by 2 months and saturated around 7 months of age (Additional file 1: Fig. S1) [12]. These mice also developed A β amyloidosis in the hippocampal and subcortical regions [12]. Despite aggressive A β amyloidosis in *App*^{NL-G-F/NL-G-F} mice, neuroinflammatory responses such as astrogliosis and microgliosis were not intense at the age of 6–9 months, whereas greater reactive gliosis was observed in cortical and hippocampal regions, as well as in subcortical regions, at the age of 15–18 months (Additional file 1: Fig. S1) [12, 18]. By contrast, A β plaques and neuroinflammatory responses were negligible even at 18 months of age in *App*^{NL/NL} mice, despite elevation of the A β level in the brain [12, 18].

Based on this neuropathological information, we carried out behavioral assays to capture cognitive (BM and CFC tasks) and emotional (EPM task) alterations in *App*-KI mice over the course of aging (Additional file 1: Fig. S1). In the experimental design, we noted that the same group of mice (Group 4) was repeatedly tested at 4, 6, and 8 months of age in the BM task (Additional file 1: Fig. S1).

***App*^{NL-G-F/NL-G-F} mice exhibit anxiolytic-like behavior, whereas *App*^{NL/NL} mice exhibit anxiogenic-like behavior, in comparison with control WT mice**

Anxiety-related behaviors were assessed using the EPM task, in which increased exploration of open arms indicates anxiolytic-like behavior [27, 28]. In 6–9-month-old *App*^{NL-G-F/NL-G-F} mice, the amount of time on (Fig. 1a; $F[2, 21] = 4.35, p = 0.026$, post hoc, WT vs. *App*^{NL-G-F/NL-G-F}, $p = 0.565$) and entries into (Fig. 1b; $F[2, 21] = 2.22, p = 0.133$) open arms during the 10-min test were slightly increased in comparison with WT mice, although these differences were not statistically significant with our sample size. The average total number of arm entries (Fig. 1c; $F[2, 21] = 1.95, p = 0.167$) and the distance travelled during the 10-min test (Additional file 2: Fig. S2a and b; $F[2, 21] = 0.27, p = 0.766$) were also slightly increased in *App*^{NL-G-F/NL-G-F} mice, although these differences were not statistically significant. By contrast, *App*^{NL/NL} mice exhibited similar levels of the amount of time on (Fig. 1a; post hoc, WT vs. *App*^{NL/NL}, $p = 0.170$) and entries into (Fig. 1b) open arms to those of WT mice, with no alterations in general exploratory activity (Fig. 1c, Additional file 2: Fig. S2b).

However, the patterns of exploration in *App*^{NL-G-F/NL-G-F} mice differed from those observed in WT mice. When we analyzed the time course of open arm exploration by scoring the percentage of time spent on the arms in each 2-min interval (Fig. 1d), WT mice exhibited significant reductions in the time spent on the open arms as the test

progressed ($F[4, 28] = 3.75, p = 0.014$), consistent with a previous report [28–30]. By contrast, *App*^{NL-G-F/NL-G-F} mice persistently explored the open arms throughout the test (Fig. 1d; $F[4, 28] = 0.68, p = 0.610$). At a later time point, *App*^{NL-G-F/NL-G-F} mice spent significantly more time on the open arms than WT mice (Fig. 1d; Time 8–10, $F[2, 21] = 6.11, p = 0.008$, post hoc, WT vs. *App*^{NL-G-F/NL-G-F}, $p = 0.022$). In contrast, similar to WT mice, *App*^{NL/NL} mice exhibited a decrease in open arm exploration as the test progressed (Fig. 1d; $F[4, 28] = 2.69, p = 0.051$). These results suggest that *App*^{NL-G-F/NL-G-F} mice have altered responses to aversive situations, such as open spaces.

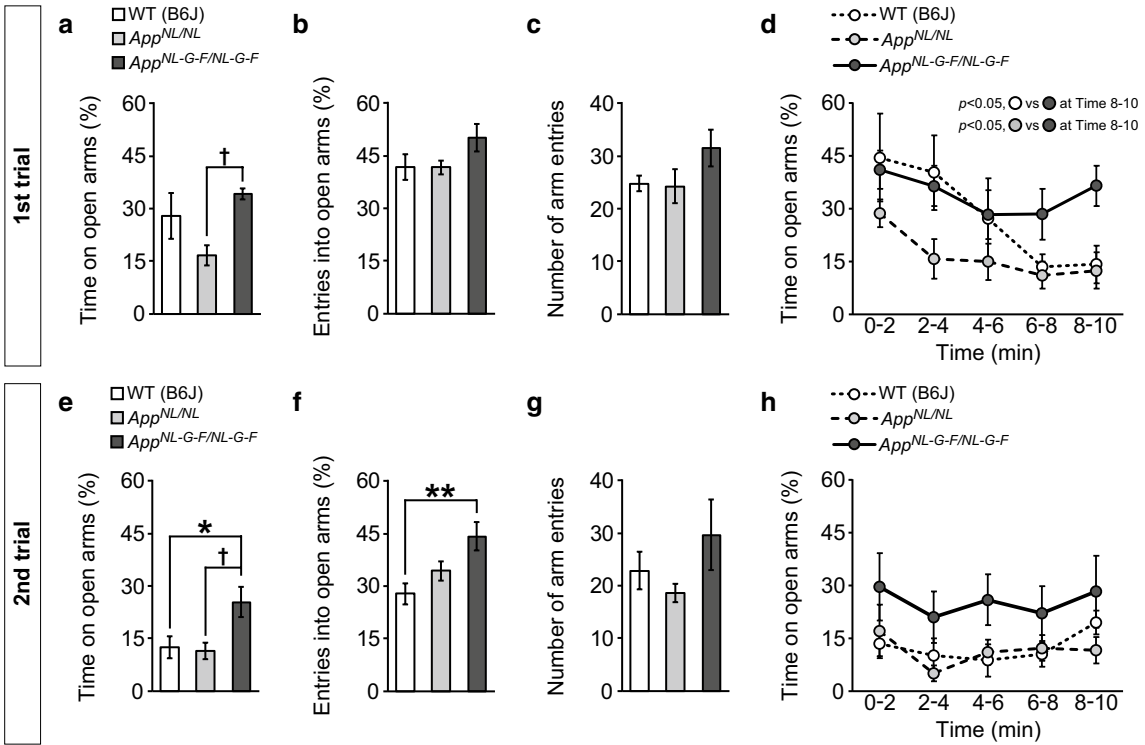
Previous studies demonstrated that laboratory rodents exhibited a significant reduction of open arm exploration when re-exposed to the EPM [28, 30, 31]. This suggests that prior test experience caused a qualitative shift in emotional state, and the acquisition of a phobic state rather than an unconditioned anxiety response. To investigate whether prior test experience could alter anxiety-related behavior, we re-tested the *App*-KI and WT mice in the same EPM paradigm.

As reported previously, WT mice exhibited robust avoidance responses to the open arms in the second trial of our EPM task, reflected by reduced percentages of time spent on and entries into the arms. By contrast, 6–9-month-old *App*^{NL-G-F/NL-G-F} mice spent significantly more time on the open arms (Fig. 1e; $F[2, 21] = 5.30, p = 0.014$, post hoc, WT vs. *App*^{NL-G-F/NL-G-F}, $p = 0.034$) and entered them more frequently (Fig. 1f; $F[2, 21] = 6.11, p = 0.008$, post hoc, WT vs. *App*^{NL-G-F/NL-G-F}, $p = 0.006$) during the 10-min test period than WT mice. The time course analysis also revealed a persistent and durable exploration of open arms in *App*^{NL-G-F/NL-G-F} mice (Fig. 1h; $F[4, 28] = 0.207, p = 0.932$). These mice showed slightly higher preference toward the open arms in comparison with WT mice at each time point, although the differences were not statistically significant (Fig. 1h; Time

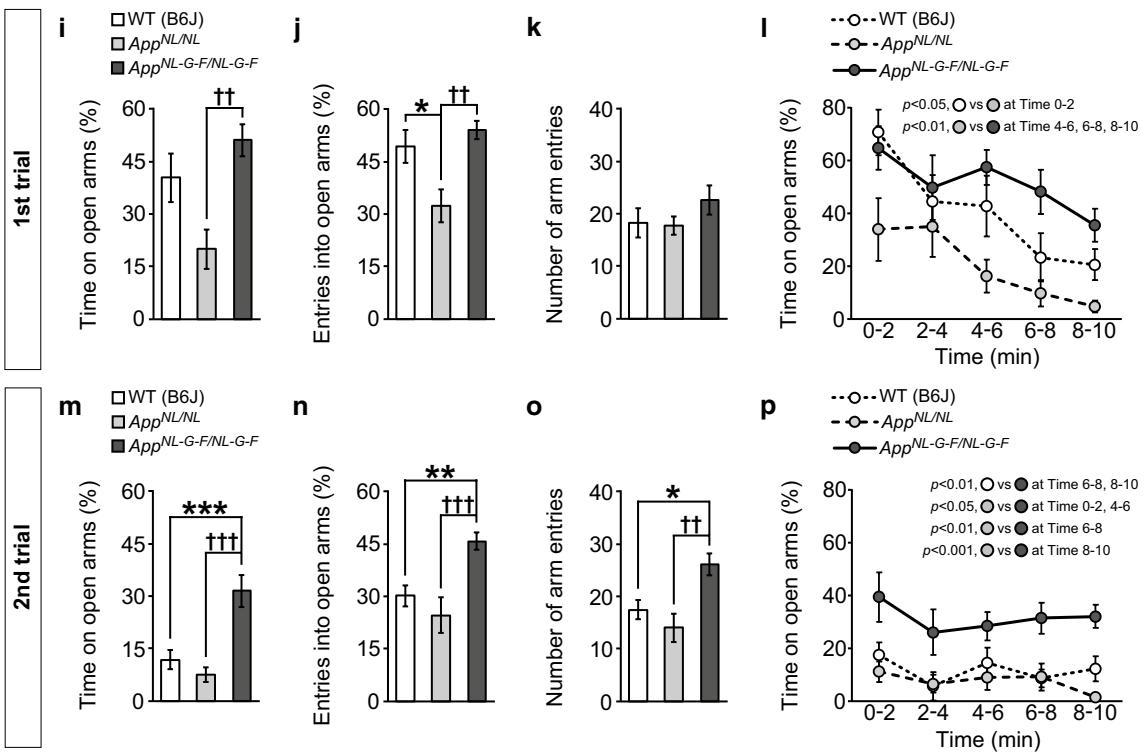
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Fig. 1 Anxiety-related behaviors in *App*^{NL-G-F/NL-G-F} and *App*^{NL/NL} mice assessed by the elevated plus maze task. Anxiety-related behaviors were assessed at both 6–9 (a–h) and 15–18 (i–p) months of age. At 6–9 months of age, *App*^{NL-G-F/NL-G-F} mice exhibited slightly increased levels of open arm exploration than WT mice, as indicated by the percentages of time spent on (a) and entries into (b) the open arms in the first trial. *App*^{NL/NL} and WT mice showed similar levels of open arm exploration. General exploratory activity in *App*^{NL-G-F/NL-G-F} mice was also slightly increased in comparison with WT mice (c). *App*^{NL-G-F/NL-G-F} mice persistently explored the open arms throughout the 10-min test, in contrast to the WT and *App*^{NL/NL} mice (d). In the second trial, *App*^{NL-G-F/NL-G-F} mice spent more time on (e) and entered more often into (f) the open arms than WT mice, with a slight increase in general activity (g). WT, *App*^{NL/NL} and *App*^{NL-G-F/NL-G-F} mice did not exhibit elevated avoidance responses to the open arms throughout the test in the second trial (h). At 15–18 months of age, *App*^{NL-G-F/NL-G-F} mice exhibited slight increases in open arm exploration in comparison with WT mice in the first trial, whereas *App*^{NL/NL} mice exhibited reduced levels of the exploration (i and j). General exploratory activity in *App*^{NL-G-F/NL-G-F} mice was slightly increased in comparison with WT mice (k). WT and *App*^{NL/NL} mice exhibited elevated open arm avoidance as the test progressed, whereas *App*^{NL-G-F/NL-G-F} mice did not (l). In the second trial, *App*^{NL-G-F/NL-G-F} mice spent more time on (m) and entered more often into (n) the open arms than WT mice. *App*^{NL-G-F/NL-G-F} mice also exhibited an elevation in general activity during the test (o). WT, *App*^{NL/NL} and *App*^{NL-G-F/NL-G-F} mice did not exhibit elevated avoidance responses to the open arms throughout the test in the second trial (p). 6–9 month-old; n = 8 WT (B6J), n = 8 *App*^{NL/NL}, n = 8 *App*^{NL-G-F/NL-G-F}; 15–18 month-old; n = 12 WT (B6J), n = 10 *App*^{NL/NL}, n = 11 *App*^{NL-G-F/NL-G-F}. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$ versus WT (B6J), † $p < 0.05$; †† $p < 0.01$; ††† $p < 0.001$ versus *App*^{NL/NL}

6–9 month-old



15–18 month-old



0–2, $F[2, 21] = 1.33$, $p = 0.286$; Time 2–4, $F[2, 21] = 2.44$, $p = 0.111$; Time 4–6, $F[2, 21] = 3.05$, $p = 0.069$; Time 6–8, $F[2, 21] = 1.35$, $p = 0.280$; Time 8–10, $F[2, 21] = 1.59$, $p = 0.228$). $App^{NL/NL}$ and WT mice engaged in similar levels of open arm exploration (Fig. 1e, f and h). As with the case of the first trial, $App^{NL-G-F/NL-G-F}$ mice exhibited slight increases in the average total number of arm entries (Fig. 1g; $F[2, 21] = 1.53$, $p = 0.240$) and the distance travelled during the 10-min test (Additional file 2: Fig. S2c and d; $F[2, 21] = 1.22$, $p = 0.316$), although these differences were not statistically significant with our sample size.

Taken together, these results suggest that 6–9-month-old $App^{NL-G-F/NL-G-F}$ mice exhibit robust anxiolytic-like behavior, even after they have habituated to a test environment.

To investigate whether the observed anxiolytic-like behavior in $App^{NL-G-F/NL-G-F}$ mice was maintained during aging, we performed the same EPM task at 15–18 months of age. In the first trial, $App^{NL-G-F/NL-G-F}$ mice showed a tendency to spend more time on (Fig. 1i; $F[2, 30] = 6.78$, $p = 0.004$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.401$) and a slightly higher frequency of entries into (Fig. 1j; $F[2, 30] = 7.01$, $p = 0.003$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.700$) the open arms during the 10-min test than WT mice, although these differences were not statistically significant. By contrast, $App^{NL/NL}$ mice tended to spend less time in the open arms (Fig. 1i; post hoc, WT vs. $App^{NL/NL}$, $p = 0.054$) and entered them significantly less frequently (Fig. 1j; post hoc, WT vs. $App^{NL/NL}$, $p = 0.021$) than WT mice, suggesting anxiogenic-like behavior in $App^{NL/NL}$ mice. General exploratory activity was slightly increased in $App^{NL-G-F/NL-G-F}$ mice, although the difference was not statistically significant with our sample size (Fig. 1k; $F[2, 30] = 1.07$, $p = 0.356$; Additional file 2: Fig. S2e and f; $F[2, 30] = 0.18$, $p = 0.836$).

As observed at 6–9 months of age, 15–18-month-old WT and $App^{NL/NL}$ mice exhibited clear avoidance of the open arms as the test progressed (Fig. 1l; WT, $F[4, 44] = 8.96$, $p < 0.001$; $App^{NL/NL}$, $F[4, 36] = 4.15$, $p = 0.007$), whereas $App^{NL-G-F/NL-G-F}$ mice did not exhibit a significant change in open arm exploration during the test ($F[4, 40] = 1.83$, $p = 0.141$). At an early time point, $App^{NL/NL}$ mice spent significantly less time on the open arms than WT mice (Fig. 1l; Time 0–2, $F[2, 30] = 4.13$, $p = 0.026$, post hoc, WT vs. $App^{NL/NL}$, $p = 0.021$). By contrast, $App^{NL-G-F/NL-G-F}$ mice exhibited slightly higher open arm exploration in the latter half of the test in comparison with WT mice, but the differences were not statistically significant (Fig. 1l; Time 4–6, $F[2, 30] = 5.30$, $p = 0.011$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.466$; Time 6–8, $F[2, 30] = 5.74$, $p = 0.008$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.079$; Time 8–10, $F[2, 30] = 7.94$, $p = 0.002$, post hoc,

WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.125$). These results suggest that 15–18-month-old $App^{NL-G-F/NL-G-F}$ mice exhibit alterations in the habituation process to aversive stimuli.

In the second trial, 15–18-month-old $App^{NL-G-F/NL-G-F}$ mice spent significantly more time on the open arms (Fig. 1m; $F[2, 30] = 13.87$, $p < 0.001$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p < 0.001$) and entered them more often (Fig. 1n; $F[2, 30] = 9.37$, $p < 0.001$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.009$) than WT mice. The time course analysis also revealed a persistent and durable exploration of open arms in $App^{NL-G-F/NL-G-F}$ mice (Fig. 1p; $F[4, 40] = 0.74$, $p = 0.570$). $App^{NL-G-F/NL-G-F}$ mice spent more time on the open arms from the beginning of the test (Fig. 1p; Time 0–2, $F[2, 30] = 5.13$, $p = 0.012$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.053$; Time 2–4, $F[2, 30] = 3.31$, $p = 0.050$; Time 4–6, $F[2, 30] = 3.51$, $p = 0.043$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.157$) and particularly at later time points (Time 6–8, $F[2, 30] = 7.26$, $p = 0.003$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.005$; Time 8–10, $F[2, 30] = 15.14$, $p < 0.001$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.003$). In addition, $App^{NL-G-F/NL-G-F}$ mice exhibited a significant increase in the total number of arm entries in comparison with WT mice (Fig. 1o; $F[2, 30] = 7.85$, $p = 0.002$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.021$). We also measured the distance travelled during the test (Additional file 2: Fig. S2g and h) and noticed that $App^{NL-G-F/NL-G-F}$ mice moved longer than WT mice, though the difference was not statistically significant with our sample size ($F[2, 30] = 3.76$, $p = 0.035$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.154$). In contrast to the first trial, $App^{NL/NL}$ and WT mice exhibited similar levels of open arm exploration (Fig. 1m and n), presumably due to habituation of WT mice to the test environment.

Taken together, these results suggest that 15–18-month-old $App^{NL-G-F/NL-G-F}$ mice exhibit robust anxiolytic-like behaviors, with increases in general exploratory activity, whereas $App^{NL/NL}$ mice displayed unconditioned anxious phenotypes in comparison with WT mice.

$App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice exhibit normal learning and memory of contextual fear up to 15–18 months of age in comparison with WT mice

The CFC task is a commonly used procedure for inducing learned fear, which is believed to be hippocampal-dependent [32, 33]. In this paradigm, a particular context as a conditioned stimulus evokes fear through association with an aversive event, such as a footshock [34]. Conditioned fear responses are impaired in both human patients and mouse models of AD [35–38].

At 6–9 months of age, the velocities of both $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice during administration of each footshock were comparable to those of WT mice (Fig. 2a; first, $F[2,$

18] = 0.32, $p = 0.732$; second, $F[2, 18] = 0.71$, $p = 0.506$; third, $F[2, 18] = 1.30$, $p = 0.297$). In addition, $App^{NL-G-F/NL-G-F}$, $App^{NL/NL}$, and WT mice exhibited the same levels of the freezing response upon subsequent presentation of footshocks during conditioning (Fig. 2b; genotype, $F[2, 18] = 0.19$, $p = 0.830$; time, $F[1.6, 28.7] = 18.02$, $p < 0.001$). To determine whether there were any locomotor deficits that could have confounded the outcome, we compared the distance travelled during the pre-shock period (the 3-min period prior to the first footshock) among genotypes (Additional file 3: Fig. S3a and b). At these ages, $App^{NL-G-F/NL-G-F}$ mice seemed to be less active than WT mice during the pre-shock period, although the difference was not statistically significant with our sample size (Additional file 3: Fig. S3b; $F[2, 18] = 2.99$, $p = 0.076$). These results suggest that all genotypes were capable of detecting and responding to footshock stimuli at similar levels.

In the context test, min-by-min scoring of the percentage of freezing behavior revealed that all genotypes exhibited similar increases in the response as the test progressed (Fig. 2c; genotype, $F[2, 18] = 10.25$, $p = 0.371$; time, $F[2.6, 46.4] = 21.08$, $p < 0.001$). Moreover, levels of the freezing response during the total 5-min period were comparable among all genotypes (Fig. 2d; $F[2, 18] = 1.05$, $p = 0.371$). These results suggest that both $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice can learn and memorize the association between cues in the experimental chamber and footshock as effectively as WT mice.

At 15–18 months of age, $App^{NL-G-F/NL-G-F}$ mice exhibited significantly higher shock reactivity than WT mice, as revealed by an increased velocity during the second and third footshocks (Fig. 2e; first, $F[2, 19] = 0.85$, $p = 0.444$; second, $F[2, 19] = 7.36$, $p = 0.004$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.007$; third, $F[2, 19] = 10.82$, $p < 0.001$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.008$). This result suggests that 15–18-month-old $App^{NL-G-F/NL-G-F}$ mice have heightened sensitivity to painful stimuli. During conditioning, both $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice exhibited levels of freezing upon subsequent presentation of footshocks similar to those of WT mice (Fig. 2f; genotype,

$F[2, 19] = 0.0$, $p = 0.994$; time, $F[1.9, 36.5] = 84.15$, $p < 0.001$). We also found that $App^{NL/NL}$ mice moved significantly less than WT mice during the pre-shock period (Additional file 3: Fig. S3c and d; $F[2, 19] = 5.13$, $p = 0.017$, post hoc, WT vs. $App^{NL/NL}$, $p = 0.016$). However, a slight reduction in locomotor activity in $App^{NL/NL}$ mice does not significantly affect the behavioral outcomes of the CFC task in $App^{NL/NL}$ mice, since these mice can exhibit similar levels of shock reactivity and freezing behavior with WT mice (Fig. 2e and f). Locomotor activity during the pre-shock period was also slightly decreased in $App^{NL-G-F/NL-G-F}$ mice, but the difference was not statistically significant with our sample size (Additional file 3: Fig. S3d; post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.092$).

In the context test, the min-by-min data for freezing behavior revealed that the time course of the freezing response was similar among all genotypes (Fig. 2g; genotype, $F[2, 19] = 0.23$, $p = 0.799$; time, $F[4, 76] = 5.06$, $p = 0.001$). During the total 5-min period of the test, both $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice exhibited levels of freezing behavior similar to those of WT mice (Fig. 2h; $F[2, 19] = 0.23$, $p = 0.800$).

Taken together, these results suggest that both $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice have intact learning and memory of contextual fear, even at 15–18 months of age.

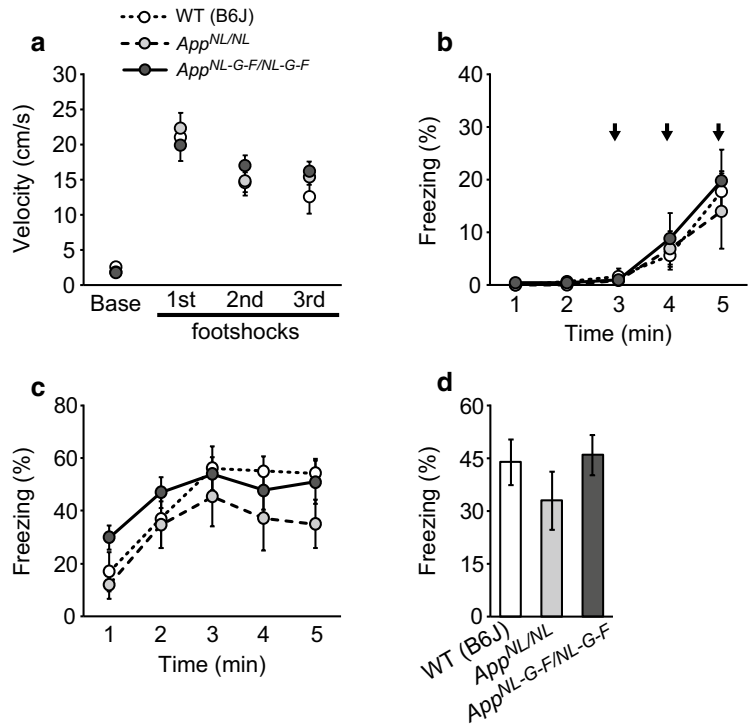
$App^{NL-G-F/NL-G-F}$ mice exhibit alterations in spatial learning ability, with intact memory, in the BM task at 8 months of age

The BM task is a spatial memory task that requires animals to learn the location of an escape hole using spatial cues, and is therefore thought to be hippocampal-dependent [39, 40]. This task is commonly used for assessment of memory deficits in animal models of AD [41–43]. In our experiments, mice were asked to acquire the spatial location of a target hole that was connected to a dark escape box during the acquisition phase (Fig. 3a [left]). One day after the fifth session of the acquisition phase, a probe test was conducted without an escape box to investigate whether mice had learned the location of

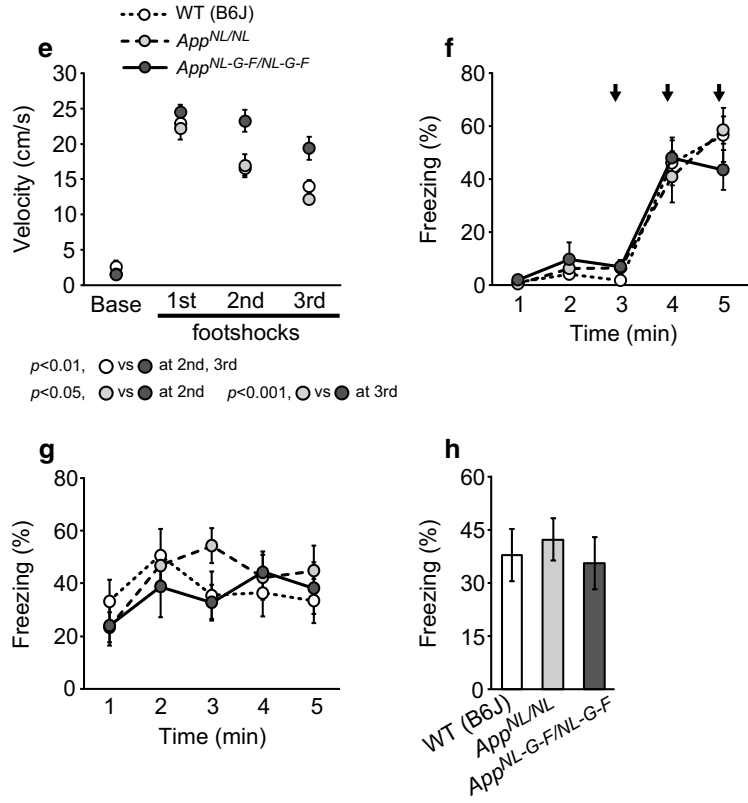
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Fig. 2 Emotional learning and fear memory in $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice, assessed by the contextual fear conditioning task. Learning and memory of contextual fear were assessed at both 6–9 (a–d) and 15–18 (e–h) months of age. At 6–9 months of age, $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice exhibited similar levels of shock reactivity, as indicated by velocity, during each presentation of footshock (a). During conditioning, all genotypes exhibited the same levels of freezing response to subsequent presentation of footshock (indicated by black arrows) (b). In the context test, all genotypes exhibited similar increases in the freezing response during the test, as revealed by the min-by-min data (c). Total levels of freezing response during the 5-min test period were comparable among all genotypes (d). At 15–18 months of age, $App^{NL-G-F/NL-G-F}$ mice exhibited higher shock reactivity than WT mice, as revealed by an increase in velocity during the second and third footshocks (e). During conditioning, all genotypes exhibited the same levels of freezing response to subsequent presentation of footshock (indicated by black arrows) (f). The time course of the freezing response in the context test was not different among genotypes (g). During the 5-min test period, the percentages of time spent in the frozen state by $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice were similar to that in WT mice (h). 6–9 month-old; n = 6 WT (B6J), n = 6 $App^{NL/NL}$, n = 9 $App^{NL-G-F/NL-G-F}$. 15–18 month-old; n = 8 WT (B6J), n = 7 $App^{NL/NL}$, n = 7 $App^{NL-G-F/NL-G-F}$

6–9 month-old



15–18 month-old



the target hole by extra-maze cues (Fig. 3a [middle]). To further assess cognitive flexibility, mice were subjected to the reversal learning task (five sessions) 1 day after the probe test (Fig. 3a [right]). And as mentioned in the experimental design above, the same group of mice was repeatedly tested at 4, 6, and 8 months of age in this BM task (Additional file 1: Fig. S1).

We found that $App^{NL/NL}$, $App^{NL-G-F/NL-G-F}$, and WT mice performed equally well in acquisition of the target hole in the BM at the ages of 4 months (Fig. 3b; $F[2, 20] = 3.12$, $p = 0.066$, Fig. 3c; $F[2, 20] = 0.48$, $p = 0.625$, Fig. 3d; $F[2, 20] = 1.10$, $p = 0.353$) and 6 months (Fig. 3g; $F[2, 20] = 1.27$, $p = 0.303$, Fig. 3h; $F[2, 20] = 2.80$, $p = 0.085$, Fig. 3i; $F[2, 20] = 0.24$, $p = 0.788$). The number of errors (Fig. 3b; $F[4, 80] = 23.21$, $p < 0.001$, Fig. 3g; $F[2.7, 54.8] = 10.07$, $p < 0.001$), latency (Fig. 3c; $F[1.9, 37.6] = 29.26$, $p < 0.001$, Fig. 3h; $F[1.7, 33.1] = 8.06$, $p = 0.002$), and distance (Fig. 3d; $F[2.3, 45.3] = 23.18$, $p < 0.001$, Fig. 3i; $F[2.8, 56.5] = 7.11$, $p = 0.001$) to reach the target hole significantly decreased as the session progressed, suggesting that all genotypes had similar learning ability.

In the probe test, all genotypes exhibited similar levels of preference toward the target quadrant that contained the target hole and the two adjacent holes at both 4 months (Fig. 3e; genotype, $F[2, 20] = 1.06$, $p = 0.365$; quadrant, $F[2.0, 40.3] = 49.06$, $p < 0.001$) and 6 months of age (Fig. 3j; genotype, $F[2, 20] = 1.56$, $p = 0.235$; quadrant, $F[1.6, 32.0] = 30.84$, $p < 0.001$). A percentage of time spent in the target quadrant for each genotype was significantly higher than chance level (25%) at both 4 months (Fig. 3e; WT, $t(14) = 6.59$, $p < 0.001$; $App^{NL/NL}$, $t(14) = 5.88$, $p < 0.001$; $App^{NL-G-F/NL-G-F}$, $t(12) = 3.28$, $p = 0.007$) and 6 months of age (Fig. 3j; WT, $t(14) = 3.91$, $p = 0.002$; $App^{NL/NL}$, $t(14) = 4.07$, $p = 0.001$; $App^{NL-G-F/NL-G-F}$, $t(12) = 3.22$, $p = 0.007$). Moreover, all genotypes exhibited similar levels of exploration of the holes in the target quadrant, with

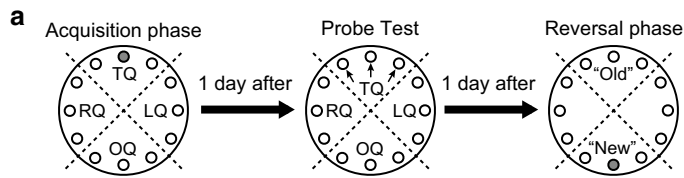
no differences in general exploratory activity, at both 4 months (Fig. 3f; (left) $F[2, 20] = 0.05$, $p = 0.949$; (right) $F[2, 20] = 1.03$, $p = 0.375$) and 6 months of age (Fig. 3k; (left) $F[2, 20] = 0.67$, $p = 0.524$; (right) $F[2, 20] = 0.75$, $p = 0.487$). These results suggest that both $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice had intact spatial learning and memory at 4 and 6 months of age.

At 8 months of age, $App^{NL-G-F/NL-G-F}$ mice exhibited a significant increase in the number of errors (Fig. 3l; $F[2, 18] = 5.34$, $p = 0.015$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.015$), latency (Fig. 3m; $F[2, 18] = 10.28$, $p = 0.001$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p < 0.001$), and distance (Fig. 3n; $F[2, 18] = 6.24$, $p = 0.009$, post hoc, WT vs. $App^{NL-G-F/NL-G-F}$, $p = 0.016$) in comparison with WT mice. However, $App^{NL-G-F/NL-G-F}$ mice still exhibited a significant decrease in the number of errors (Fig. 3l; $F[2.5, 45.2] = 11.47$, $p < 0.001$), latency (Fig. 3m; $F[1.4, 25.2] = 6.91$, $p = 0.008$), and distance (Fig. 3n; $F[2.1, 38.1] = 8.35$, $p = 0.001$), and were able to solve the task proficiently (at levels comparable to those of WT mice) by the fifth training session. These results suggest that 8-month-old $App^{NL-G-F/NL-G-F}$ mice have subtle alterations in their ability to learn the spatial location of the target hole.

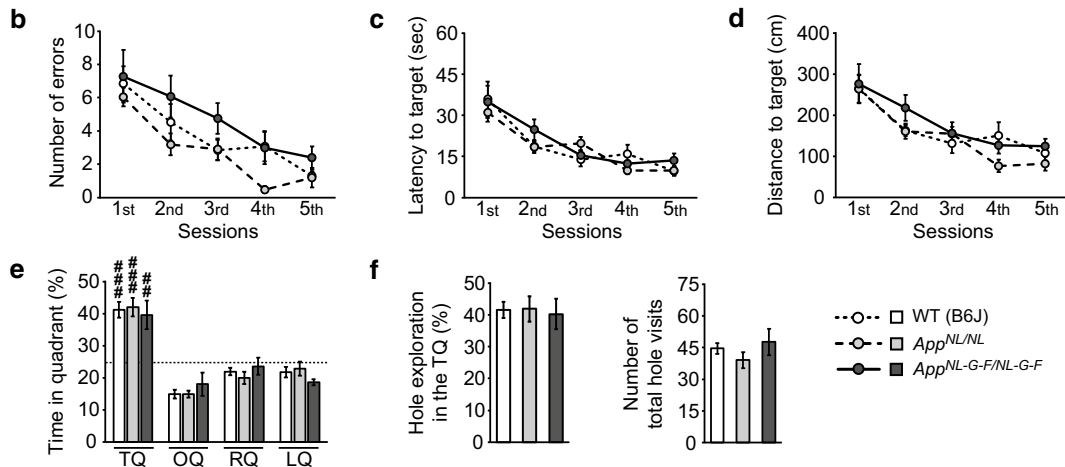
In the probe test, all genotypes exhibited similar levels of preference toward the target quadrant (Fig. 3o; genotype, $F[2, 18] = 1.36$, $p = 0.283$; quadrant, $F[1.8, 31.9] = 38.63$, $p < 0.001$). The percentages of time spent in the target quadrant were significantly higher above chance level for WT and $App^{NL/NL}$ mice (Fig. 3o; WT, $t(14) = 6.00$, $p < 0.001$; $App^{NL/NL}$, $t(12) = 5.07$, $p < 0.001$), but not for $App^{NL-G-F/NL-G-F}$ mice ($t(10) = 2.11$, $p = 0.062$), with our sample size. The percentage of hole exploration in the target quadrant (Fig. 3p; (left) $F[2, 18] = 3.35$, $p = 0.058$) and the total number of hole visits (Fig. 3p; (right) $F[2, 18] = 0.35$, $p = 0.712$) were similar among all genotypes.

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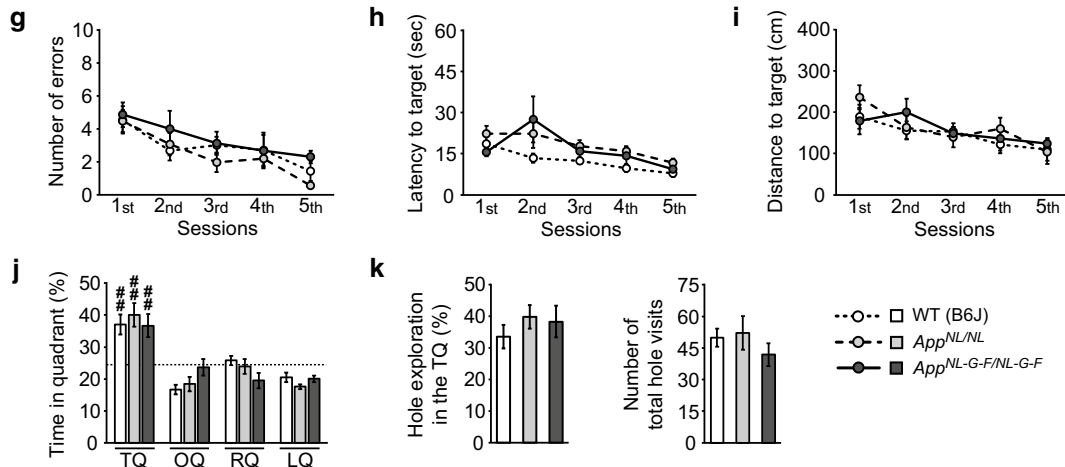
Fig. 3 Spatial learning and memory in $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice assessed by the Barnes maze task. Spatial learning and memory were assessed at 4 (b–f), 6 (g–k), and 8 (i–p) months of age. In the acquisition phase (a [left]), one hole (indicated by a gray hole) was designated as the target hole with an escape box. A probe test was performed 1 day after the last training session, in which the escape box was removed (a [middle]). The three black arrows indicate the target hole and adjacent holes, respectively. In the reversal phase (a [right]), the target hole was moved to the position opposite the original 1 day after the probe test. TQ: target quadrant; OQ: opposite quadrant; RQ: right quadrant; LQ: left quadrant. At 4 and 6 months of age, $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice performed as well as WT mice in acquisition of the target hole, as revealed by similar decreases in the number of errors (b and g), latency (c and h), and distance (d and i) across the acquisition phase. In the probe test, all genotypes exhibited similar levels of preference toward the target quadrant (TQ) above chance level (25%, as indicated by dotted lines) (e and j) and similar levels of exploration of the holes in the target quadrant (f [left] and k [left]) with no differences in exploratory activity (f [right] and k [right]). At 8 months of age, $App^{NL-G-F/NL-G-F}$ mice made more errors (l), took more time (m), and travelled farther (n) to reach the target hole than WT mice throughout the acquisition phase. In the probe test, WT and $App^{NL/NL}$ mice exhibited significant preference toward the target quadrant (TQ) above chance level (25%, as indicated by dotted lines) (o). All genotypes exhibited similar levels of exploration of the holes in the target quadrant (p [left]), with no alterations in general activity (p [right]). 4 month-old; n = 8 WT (B6J), n = 8 $App^{NL/NL}$, n = 7 $App^{NL-G-F/NL-G-F}$. 6 month-old; n = 8 WT (B6J), n = 8 $App^{NL/NL}$, n = 7 $App^{NL-G-F/NL-G-F}$. 8 month-old; n = 8 WT (B6J), n = 7 $App^{NL/NL}$, n = 6 $App^{NL-G-F/NL-G-F}$. ## $p < 0.01$, ### $p < 0.001$ versus chance level



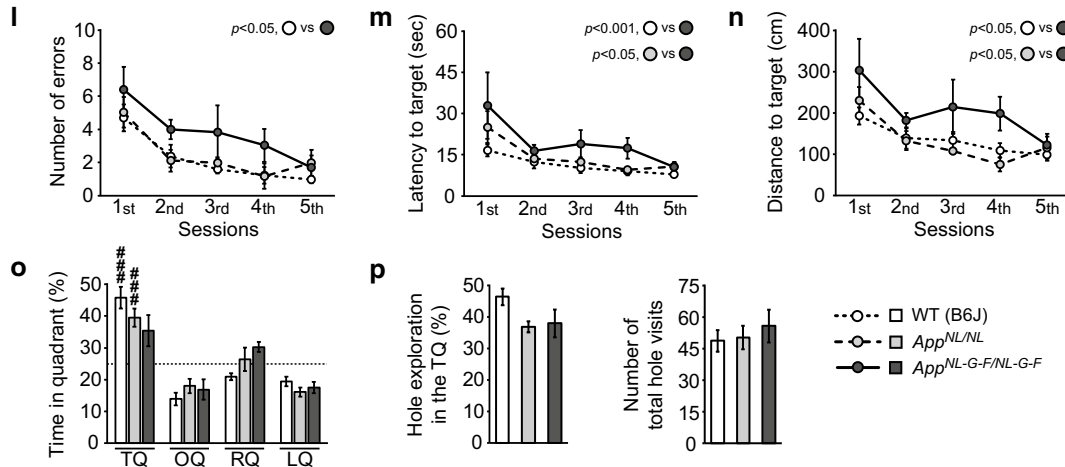
4 month-old

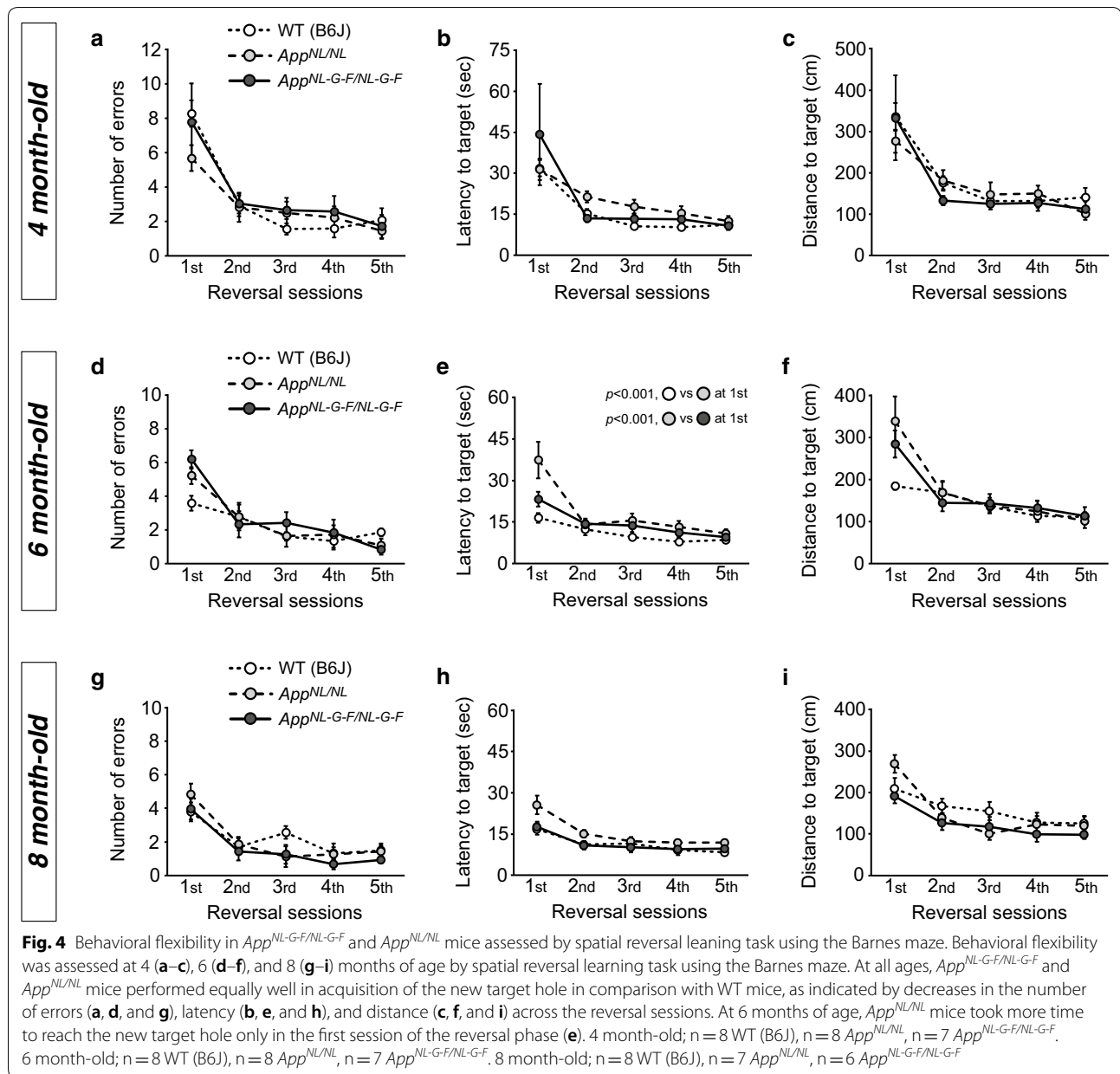


6 month-old



8 month-old





Taken together, these results suggest that, at 8 months of age, $App^{NL-G-F/NL-G-F}$ mice exhibit reduced spatial learning ability in comparison with WT mice, but still retain normal spatial memory.

Both $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice exhibit normal flexibility in a reversal learning task up to 8 months of age

Reversal learning, a way to model some aspects of higher-order cognitive functions in rodents [44, 45], requires cognitive flexibility and impulse control, and thus taps into components of human executive function [46, 47]. Previous studies demonstrated that transgenic mouse

models of A β amyloidosis are impaired in reversal learning [48–50]. To assess reversal learning using the BM, we moved the target hole to the opposite position 1 day after the probe test (Fig. 3a [right]).

We found that both $App^{NL-G-F/NL-G-F}$ and $App^{NL/NL}$ mice exhibited similar levels of performance in the reversal learning task in comparison with WT mice at 4 months (Fig. 4a; $F[2, 20] = 0.35$, $p = 0.711$, Fig. 4b; $F[2, 20] = 0.87$, $p = 0.434$, Fig. 4c; $F[2, 20] = 0.32$, $p = 0.733$), 6 months (Fig. 4d; $F[2, 20] = 0.38$, $p = 0.690$, Fig. 4e; $F[2, 20] = 6.31$, $p = 0.008$, Fig. 4f; $F[2, 20] = 1.73$, $p = 0.202$), and 8 months

of age (Fig. 4g; $F[2, 18]=0.66$, $p=0.530$, Fig. 4h; $F[2, 18]=3.00$, $p=0.075$, Fig. 4i; $F[2, 18]=1.41$, $p=0.269$).

The number of errors (Fig. 4a; $F[1.8, 36.4]=28.55$, $p<0.001$, Fig. 4d; $F[2.5, 49.8]=30.91$, $p<0.001$, Fig. 4g; $F[2.2, 40.0]=28.20$, $p<0.001$), latency (Fig. 4b; $F[1.1, 22.9]=13.28$, $p=0.001$, Fig. 4e; $F[1.3, 25.6]=24.78$, $p<0.001$, Fig. 4h; $F[2.1, 38.6]=29.80$, $p<0.001$) and distance (Fig. 4c; $F[1.5, 29.1]=20.47$, $p<0.001$, Fig. 4f; $F[1.4, 28.6]=21.32$, $p<0.001$, Fig. 4i; $F[2.4, 42.7]=21.93$, $p<0.001$) to the new target hole were progressively reduced in all genotypes and at all ages, suggesting that both *App*^{NL-G-F/NL-G-F} and *App*^{NL/NL} mice could adjust their response to find the new location as effectively as WT mice.

We noticed that 6-month-old *App*^{NL/NL} mice spent more time to reach the new target hole than WT mice at the first session of the reversal phase (Fig. 4e; genotype \times session, $F[2.6, 25.6]=3.47$, $p=0.037$, simple main effect on first session, $p<0.001$, post hoc, WT vs. *App*^{NL/NL}, $p<0.001$). However, no significant change in the latency was detected at the second session (simple main effect on second session, $p=0.833$). Moreover, no such difference was observed in the 8-month-old *App*^{NL/NL} mice (Fig. 4h).

Taken together, these results suggest that both *App*^{NL-G-F/NL-G-F} and *App*^{NL/NL} mice exhibit normal cognitive flexibility in the reversal learning task up to 8 months of age.

Discussion

A β amyloidosis, tau aggregation, neuroinflammation, neurodegeneration, and cognitive deficits are defining features of AD. To date, multiple transgenic mouse models overexpressing human APP with familial AD mutations have been shown to develop age-dependent A β amyloidosis, neuroinflammation, and cognitive impairments in spatial memory and contextual fear memory [6, 7, 51]. However, these APP-overexpressing mouse models may have pathophysiological properties caused by non-physiological overexpression of APP, in addition to A β pathology [11].

To overcome this issue, several *App*-KI mouse models have been generated that recapitulate A β pathology without APP overexpression. The first reported *App*-KI line, *App*^{NLh} mice, harbor the Swedish mutation with humanized A β sequences in the murine *App* gene locus. Although *App*^{NLh} mice do not develop A β pathology, the double KI line interbred with mutant *Psen1*^{P264L/P264L} KI line (*App*^{NLh/NLh} \times *Psen1*^{P264L/P264L} mice) progressively develop A β pathology and cognitive impairments with no alterations in locomotor activity and anxiety-related behavior [52, 53]. More recently developed *App*^{DSL} mice harbor three familial

AD-associated mutations (Swedish, London, and Dutch) with a humanized A β sequence in the murine *App* locus [54]. Similar to the *App*^{NLh} mice, *App*^{DSL} mice do not develop A β pathology independently, but do so when interbred with *Psen1*^{M146V} KI mice. The double KI line *App*^{DSL/DSL} \times *Psen1*^{M146V/M146V} also exhibits elevated levels of anxiety, followed by deficits in spatial learning and memory [54]. However, these earlier *App*-KI mouse models required homozygous expression of familial AD mutant *Psen1* alleles to obtain A β deposition [52, 54, 55], and the potential effects of homozygous mutation in *Psen1* on observed phenotypes must be considered.

By contrast, *App*^{NL-G-F/NL-G-F} mice exhibit progressive amyloid pathology, including microglial and astrocytic activation and loss of synaptic markers, in the absence of *Psen1* mutation [12, 18]. Reports of cognitive deficits in *App*^{NL-G-F/NL-G-F} mice have varied between laboratories [12, 25, 26], although mild impairment in spatial reversal learning and enhanced impulsivity have been detected using an automated IntelliCage apparatus [18]. These results suggest that, despite aggressive A β pathology and neuroinflammation, cognitive alterations in *App*^{NL-G-F/NL-G-F} mice are modest.

Consistent with these previous studies, our results demonstrated that neither *App*^{NL-G-F/NL-G-F} nor *App*^{NL/NL} mice exhibited severe memory deficits in the CFC (Fig. 2) or BM tasks (Fig. 3). However, we detected a subtle decline in spatial learning ability in *App*^{NL-G-F/NL-G-F} mice at the age of 8 months (Fig. 3). Because the learning deficits were not evident at younger ages (4 and 6 months of age), this may represent an aspect of age-dependent cognitive impairment in *App*^{NL-G-F/NL-G-F} mice. Moreover, alterations in acquisition of spatial information may occur before the onset of memory deficit in *App*^{NL-G-F/NL-G-F} mice. In our experimental design, the BM task was not run at a similar age range to the EPM and CFC tasks (Additional file 1: Fig. S1). To clarify whether *App*^{NL-G-F/NL-G-F} mice show more dramatic deficits in spatial learning and memory during aging, it would be important to examine a spatial task after 8 months of age.

Several previous studies have shown that commonly used transgenic models such as the Tg2576 and *App*^{Swe}/*Psen1*^{dE9} mice exhibit deficits in spatial learning in the BM [41–43]. Of particular interest, the TgCRND8 mouse model exhibit poor spatial learning in the BM task [56], while they also have deficits in attentional control [57]. Given that attentional deficits are likely to influence performance on memory tasks, it is conceivable that reduced attentional control of *App*^{NL-G-F/NL-G-F} mice [18] might have contributed to alterations in spatial learning ability in our BM task.

In this study, we repeatedly subjected the same group of mice to the BM task until the age of 8 months (Additional

file 1: Fig. S1), suggesting that mice might become familiar with the rules and environment of the maze. In fact, habituation of the testing environment and rule learning by repeated exposure to the maze resulted in the fewer errors at the age of 8 months in comparison with younger ages (Figs. 3 and 4). Thus, as suggested by a previous study in *App^{Swe}/Psen1^{dE9}* mice using the BM task [42], these processes might be compromised in 8-month-old *App^{NL-G-F/NL-G-F}* mice, which may lead to observed spatial learning impairment. These results also suggest that the experimental strategy testing *App*-KI mouse models in the BM task without prior test experience is likely to yield different results observed in this study.

In addition to cognitive deficits, 60–80% of AD cases are associated with non-cognitive neuropsychiatric symptoms [58, 59], including anxiety disturbances, depressive symptoms, activity disturbances, and aggression [60–62]. For example, some AD patients are subjected to anxiety, whereas the opposite tendency (disinhibition) has also been reported [60, 61]. Intriguingly, several APP-overexpressing mouse models exhibit anxiety disturbances [7, 24] and an increase in open arm exploration has been observed in several APP-overexpressing mouse models with parenchymal A β plaques, including APP23, Tg2576, and *App^{Swe}/Psen1^{dE9}* mice [24].

A very recent study reported anxiolytic-like behavior in *App^{NL-G-F/NL-G-F}* mice in comparison with *App^{NL/NL}* mice, detectable from 3 months of age [25]. Similarly, we found that *App^{NL-G-F/NL-G-F}* mice exhibited anxiolytic-like behavior in comparison with WT mice (Fig. 1). These data from *App*-KI mice suggest that anxiolytic-like behaviors observed in mouse models of A β amyloidosis are associated with A β -mediated pathologies [24, 53] rather than overexpression of APP. Interestingly, some mouse models with traumatic brain injury exhibit increases in open arm exploration in the EPM task, followed by elevated levels of reactive gliosis and cerebrovascular dysfunction [63–66]. Another study demonstrated that local neuroinflammation within the dorsal raphe nucleus, resulting in serotonergic hypofunction, caused the same behavioral consequences in the EPM task [67]. Because elevated levels of reactive gliosis are associated with A β pathology in *App^{NL-G-F/NL-G-F}* mice [12], these pathological changes (including neuroinflammatory responses and vascular dysfunction) may play a role in the expression of anxiolytic-like behavior.

We also noticed that activity of *App^{NL-G-F/NL-G-F}* mice was slightly higher than that of WT mice in the EPM task (Fig. 1 and Additional file 2: Fig. S2), raising a possibility that there may be a general increase in locomotor activity in *App^{NL-G-F/NL-G-F}* mice. However, in the CFC task, *App^{NL-G-F/NL-G-F}* mice were rather less active than WT mice during the pre-shock period (Additional file 3:

Fig. S3). These results suggest that the hyperactive phenotypes observed in the EPM task may be elicited by an aversive situation, rather than an innate behavioral trait in *App^{NL-G-F/NL-G-F}* mice.

We also found that *App^{NL/NL}* mice, which do not develop A β pathology, exhibited lower open arm durations at 15–18 months of age, suggesting elevated anxiety levels in these mice. A previous study also reported that transgenic mice overexpressing APP^{Swe} without A β pathology exhibited elevated anxiety levels in the same EPM paradigm [24, 68]. Increased anxiety levels in these mice are not due to overproduction of N- or C-terminal fragment- β (NTF- β or CTF- β) of APP, as demonstrated by the observation that *App^{NL-G-F/NL-G-F}* mice exhibited anxiolytic-like behavior, whereas Tg13592 mice overexpressing CTF- β did not exhibit altered anxiety levels in comparison with their non-transgenic controls [69]. Although *App^{NL/NL}* mice develop neither A β pathology nor neuroinflammatory response, they have dramatically increased levels of soluble A β in comparison with WT mice. Thus, higher levels of soluble A β may induce changes in synaptic functions, which may be responsible for emotional control in these KI mice. Hyperanxious behavior in the EPM task is often associated with alterations in several neurotransmitter systems, including γ -aminobutyric acid (GABA)ergic and serotonergic neurotransmission [70, 71]. Thus, altered neurotransmission caused by high levels of soluble A β may contribute to the expression of anxiogenic-like behavior in *App^{NL/NL}* mice.

Conclusion

Our results demonstrate that *App^{NL-G-F/NL-G-F}* and *App^{NL/NL}* mice exhibit behavioral changes associated with non-cognitive, emotional domains before the onset of definitive cognitive deficits. These observations consistently indicate that *App^{NL-G-F/NL-G-F}* mice represent a model for preclinical AD and that they are useful for the study of AD prevention rather than treatment after neurodegeneration. This study provides information that will be critical for both translational and basic research for AD using *App^{NL-G-F/NL-G-F}* and *App^{NL/NL}* mice.

Methods

Animals

The original lines of *App*-KI (*App^{NL-G-F/NL-G-F}* and *App^{NL/NL}*) mice were established as C57BL/6J congenic line (a genetic background strain) by repeated backcrosses as described previously [12] and obtained from RIKEN Center for Brain Science (Wako, Japan). All experiments were performed with male *App^{NL-G-F/NL-G-F}*, *App^{NL/NL}* and WT (C57BL/6J) mice at the Institute for Animal Experimentation in National Center for Geriatrics and Gerontology. After weaning at postnatal day

(PND) 28–35, all mice were housed socially in same-sex groups in a temperature-controlled environment under a 12-h light/dark cycle (lights on at 7:00, lights off at 19:00), with food and water available ad libitum. We prepared four independent groups of male mice with mixed genotypes (*App*^{NL-G-F/NL-G-F}, *App*^{NL/NL}, and WT) and assessed cognitive and emotional domains using three behavioral paradigms at different ages (Additional file 1: Fig. S1). All handling and experimental procedures were performed in accordance with the Guidelines for the Care of Laboratory Animals of National Center for Geriatrics and Gerontology (Obu, Japan). All animals were euthanized with intraperitoneal barbiturate overdose (sodium pentobarbital, 120 mg/kg body weight) after each behavioral experiment.

Elevated plus maze task

The apparatus consisted of two opposing open arms (25 × 5 cm) and two opposing closed arms (25 × 5 cm, surrounded by 15 cm-high transparent walls) that extended from a center platform (5 × 5 cm) forming a cross shape (O'hara & Co., Tokyo, Japan) [72]. The maze was elevated 50 cm above the floor with a light intensity on the center platform of approximately 100 lx. To avoid falls, the open arms were surrounded by a 0.3 cm-high rim. On the test day, 6–9- (*n* = 8/genotype) and 15–18- (*n* = 10–12/genotype) month-old mice were placed individually in the center platform facing an open arm and allowed to freely explore the apparatus for 10 min. At the end of the first trial, the mice were returned to their homecage and socially housed until the beginning of the second trial. The apparatus was cleaned with distilled water and then ethanol to remove any odor cues between subject mice. All animals were retested after 10–27 days from the first trial. Time spent on open arms (s), total distance travelled (cm) and numbers of open and closed arm entries were automatically measured by the ANY-Maze video tracking software (Stoelting Co., IL, USA) [73]. The number of open and closed arm entries was combined to yield a measure of total arm entries, which reflected general exploratory activity during the test. Open arm entries were analyzed as a percentage score by dividing the number of open arm entries by the total number of arm entries (% Entries into open arms = [Number of open arm entries/Number of total arm entries] × 100).

Contextual fear conditioning task

Mice were handled for 3 days prior to the commencement of contextual fear conditioning. The mice were trained and tested in conditioning chambers (17 × 10 × 10 cm) with a stainless-steel grid floor (0.2 cm diameter, spaced 0.5 cm apart; O'hara & Co., Tokyo, Japan) [72] surrounded by a sound-attenuating

white chest (approximately 200 lx in the chest) with a background noise (55 dB). On the conditioning day, 6–9- (*n* = 6–9/genotype) and 15–18- (*n* = 7–8/genotype) month-old mice were individually placed in the conditioning chamber and allowed to explore freely for 3 min. At the end of this 3-min period, a mild footshock (0.5 mA, 2 s) was presented. Two more footshocks were presented with a 1-min inter-stimulus interval, and then the mice were returned to their home cage at 30 s after the last footshock. One day after the conditioning, mice were placed in the same chamber and allowed to explore freely for 5 min without footshock administration. The chambers were cleaned with distilled water and then ethanol to remove any odor cues between subject mice. In each test, percentage of freezing time and distance travelled (cm) were calculated automatically using ImageFZ software (O'hara & Co., Tokyo, Japan). To assess sensitivity or reactivity to footshock, we also calculated the velocity during each footshock presentation and an equivalent baseline period (actual 2-s period just prior to the first footshock), based on distance travelled during a given time period, since it has been suggested to be the most sensitive aspect of shock reactivity [34, 74, 75].

Barnes maze task

The Barnes circular maze task was conducted on a white circular surface (1.0 m in diameter, with 12 holes equally spaced around the perimeter; O'hara & Co., Tokyo, Japan) [72]. The circular open field was elevated 75 cm above the floor with a light intensity on the center of the circular open field of approximately 1000 lx. A black acrylic escape box (17 × 13 × 7 cm) was located under one of the holes, and the hole above the escape box represented the target hole. The location of the target hole was consistent for a given mouse, and mice within a squad were assigned to the same target hole location across the sessions. Trials were administered in a spaced fashion so that all mice within a squad completed a given trial before subsequent trials were run. The maze was rotated 90° between trials, with the spatial location of the target hole unchanged with respect to the distal visual room cues, to prevent a bias based on olfactory or proximal cues within the maze. After each trial, the apparatus including the cylinder and escape box were cleaned carefully with distilled water and then ethanol to eliminate any potential odor cues.

One day after the habituation to familiarize mice with the maze and the escape box, they were subjected to 5 days training sessions (four trials per session). The same group of mice was repeatedly tested at 4, 6 and 8 months of age (Additional file 1: Fig. S1). During the acquisition phase, 4- (*n* = 7–8/genotype), 6- (*n* = 7–8/genotype) and 8- (*n* = 6–8/genotype) month-old mice were individually placed in a white acrylic cylinder

(17 cm-high, 11 cm diameter) before the start of each trial, and after approximately 30 s the cylinder was removed to start the trial. Each trial ended when the mouse entered the escape box or after 5 min had elapsed. The mice that could not find the target hole were guided to the hole manually and allowed to enter the escape box to remain there for 1 min. For each trial, the number of errors, latency (s) and distance travelled (cm) to reach the target hole were automatically measured by custom-written software in MATLAB (The MathWorks, Inc., MA, USA). In our software, target zones were defined to include each separate hole and 1 cm around them. We recorded an error when a mouse touched a target zone that did not have an escape box beneath it.

One day after the last training session, a probe test was conducted without the escape box for 3 min, to confirm that this spatial task was acquired based on navigation by distal environmental cues. The time spent in each quadrant (TQ; target quadrant, OQ; opposite quadrant, RQ; right quadrant, LQ; left quadrant) (s) and numbers of the visits to the target hole and two adjacent holes (indicated by black arrows in Fig. 3a) and total hole visits during the test were measured by the software. Hole exploration in the target quadrant was defined by percentage of the visits to three holes in the target quadrant for total hole visits during the test.

For reversal leaning task (reversal phase), the target was moved to a new position opposite to the original 1 day after the probe test, and mice were retrained in 5 days reversal sessions to find the new location of the escape box. During the reversal phase, the number of errors, latency (s), and distance travelled (cm) to reach the new target were also calculated by the software.

Statistical analysis

Statistical differences between genotypes against behavioral parameters with one dependent variable were determined by repeated-measures analysis of variance (ANOVA). When necessary, Greenhouse–Geisser estimates of sphericity were used to correct for degrees of freedom. Bonferroni post hoc comparisons were used to evaluate group differences. For the comparisons of multiple means with genotypes as one independent variable, one-way ANOVA followed by the Tukey's post hoc tests was used. One-sample *t* test was used to compare performance on the probe test of the Barnes maze task against chance level (25%). Data are presented as means \pm SEM. All alpha levels were set at 0.05.

Additional files

Additional file 1: Fig. S1. Time course of experimental procedures for assessing cognitive and emotional domains in *App*-KI mice. Based on pathological information about the brains of *App*^{NL-G-F/NL-G-F} mice, cognitive and emotional domains in *App*-KI mice were assessed at different ages using three behavioral assays. The same group of mice (Group 4) was assessed at 4, 6 and 8 months of age for spatial learning and memory and behavioral flexibility using the Barnes maze (BM) task, and at 15–18 months of age for contextual fear memory using the contextual fear conditioning (CFC) task. Time courses of brain pathology in *App*^{NL-G-F/NL-G-F} mice are shown based on previous studies.

Additional file 2: Fig. S2. Locomotor activity of *App*^{NL-G-F/NL-G-F} and *App*^{NL/NL} mice during the first and second trials in the elevated plus maze task. The distance travelled during the 10-min test of the first and second trials in the elevated plus maze task was compared among genotypes at both 6–9 (a–d) and 15–18 (e–h) months of age. Representative images of movement tracks during the first and second trials for each genotype at 6–9 (a and c) and 15–18 (e and g) months of age were shown (closed arms are indicated by shaded areas). At 6–9 months of age, *App*^{NL-G-F/NL-G-F} mice exhibited slight increases in distance travelled during the first (b) and second (d) trials in comparison with WT mice. By contrast, locomotor activity in *App*^{NL/NL} mice was comparable with WT mice in the two trials. At 15–18 months of age, *App*^{NL-G-F/NL-G-F} mice exhibited a slight increase in movement compared to WT mice during the first (f) and second (g) trials. *App*^{NL/NL} mice moved at similar levels compared with WT mice in the two trials. 6–9 month-old; n = 8 WT (B6J), n = 8 *App*^{NL/NL}, n = 8 *App*^{NL-G-F/NL-G-F}. 15–18 month-old; n = 12 WT (B6J), n = 10 *App*^{NL/NL}, n = 11 *App*^{NL-G-F/NL-G-F}. **p* < 0.05 versus *App*^{NL/NL}.

Additional file 3: Fig. S3. Locomotor activity in *App*^{NL-G-F/NL-G-F} and *App*^{NL/NL} mice during the pre-shock period in the contextual fear conditioning task. The distance travelled during the pre-shock period (3-min period just prior to the first footshock) in conditioning was compared among genotypes at both 6–9 (a and b) and 15–18 (c and d) months of age. Representative images of movement tracks during the pre-shock period in each genotype at 6–9 (a) and 15–18 (c) months of age were shown. At 6–9 months of age, *App*^{NL-G-F/NL-G-F} mice exhibited a slight decrease in distance travelled during the pre-shock period in comparison with WT mice (b). At 15–18 months of age, *App*^{NL/NL} mice exhibited a significant decrease in distance travelled during the pre-shock period in comparison with WT mice (d). Locomotor activity in *App*^{NL-G-F/NL-G-F} mice was also slightly decreased in comparison with WT mice. 6–9 month-old; n = 6 WT (B6 J), n = 6 *App*^{NL/NL}, n = 9 *App*^{NL-G-F/NL-G-F}. 15–18 month-old; n = 8 WT (B6 J), n = 7 *App*^{NL/NL}, n = 7 *App*^{NL-G-F/NL-G-F}. **p* < 0.05 versus WT (B6J).

Abbreviations

AD: Alzheimer's disease; APP: amyloid precursor protein; A β : amyloid- β ; NFT: neurofibrillary tangle; KI: knock-in; WT: wild-type; EPM: elevated plus maze; CFC: contextual fear conditioning; BM: Barnes maze; ANOVA: analysis of variance; CTF- β : C-terminal fragment- β ; NTF- β : N-terminal fragment- β ; GABA: γ -aminobutyric acid.

Authors' contributions

YS, MS and KMI conceived the experiments, YS conducted the experiment, YS, MS and KMI analyzed and interpret the data. TS and TCS provided the *App*-KI mice and interpret the data. YS, MS and KMI wrote the paper. All authors critically reviewed the manuscript. All authors read and approved the final manuscript.

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Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding authors on reasonable request.

Consent for publication

Not applicable.

Ethics approval and consent to participate

All animal experimental procedures were performed according to the NIH Guide for the Care and Use of Laboratory Animals and other national regulations and policies with the approval of the Animal Care and Use Committee at National Center for Geriatrics and Gerontology, Japan (Approval number: 29-3).

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References

- Scheltens P, Blennow K, Breteler MM, de Strooper B, Frisoni GB, Salloway S, Van der Flier WM. Alzheimer's disease. *Lancet*. 2016;388(10043):505–17.
- Winblad B, Amouyel P, Andrieu S, Ballard C, Brayne C, Brodaty H, Cedazo-Minguez A, Dubois B, Edvardsson D, Feldman H, et al. Defeating Alzheimer's disease and other dementias: a priority for European science and society. *Lancet Neurol*. 2016;15(5):455–532.
- Braak H, Braak E. Demonstration of amyloid deposits and neurofibrillary changes in whole brain sections. *Brain Pathol*. 1991;1(3):213–6.
- Hyman BT, Phelps CH, Beach TG, Bigio EH, Cairns NJ, Carrillo MC, Dickson DW, Duyckaerts C, Frosch MP, Masliah E, et al. National Institute on Aging–Alzheimer's Association guidelines for the neuropathological assessment of Alzheimer's disease. *Alzheimers Dement*. 2012;8(1):1–13.
- Serrano-Pozo A, Frosch MP, Masliah E, Hyman BT. Neuropathological alterations in Alzheimer disease. *Cold Spring Harb Perspect Med*. 2011;1(1):a006189.
- Puzzo D, Gulisano W, Palmeri A, Arancio O. Rodent models for Alzheimer's disease drug discovery. *Expert Opin Drug Discov*. 2015;10(7):703–11.
- Webster SJ, Bachstetter AD, Nelson PT, Schmitt FA, Van Eldik LJ. Using mice to model Alzheimer's dementia: an overview of the clinical disease and the preclinical behavioral changes in 10 mouse models. *Front Genet*. 2014;5:88.
- Balducci C, Forloni G. APP transgenic mice: their use and limitations. *Neuromolecular Med*. 2011;13(2):117–37.
- Kitazawa M, Medeiros R, Laferla FM. Transgenic mouse models of Alzheimer disease: developing a better model as a tool for therapeutic interventions. *Curr Pharm Des*. 2012;18(8):1131–47.
- Gidycz DC, Deibel SH, Hong NS, McDonald RJ. Barriers to developing a valid rodent model of Alzheimer's disease: from behavioral analysis to etiological mechanisms. *Front Neurosci*. 2015;9:245.
- Sasaguri H, Nilsson P, Hashimoto S, Nagata K, Saito T, De Strooper B, Hardy J, Vassar R, Winblad B, Saido TC. APP mouse models for Alzheimer's disease preclinical studies. *EMBO J*. 2017;36(17):2473–87.
- Saito T, Matsuba Y, Mihira N, Takano J, Nilsson P, Itohara S, Iwata N, Saido TC. Single App knock-in mouse models of Alzheimer's disease. *Nat Neurosci*. 2014;17(5):661–3.
- Citron M, Oltersdorf T, Haass C, McConlogue L, Hung AY, Seubert P, Vigo-Pelfrey C, Lieberburg I, Selkoe DJ. Mutation of the beta-amyloid precursor protein in familial Alzheimer's disease increases beta-protein production. *Nature*. 1992;360(6405):672–4.
- Guardia-Laguarta C, Pera M, Clarimon J, Molinuevo JL, Sanchez-Valle R, Llado A, Coma M, Gomez-Isla T, Blesa R, Ferrer I, et al. Clinical, neuropathologic, and biochemical profile of the amyloid precursor protein I716F mutation. *J Neuropathol Exp Neurol*. 2010;69(1):53–9.
- Lichtenthaler SF, Wang R, Grimm H, Uljon SN, Masters CL, Beyreuther K. Mechanism of the cleavage specificity of Alzheimer's disease gamma-secretase identified by phenylalanine-scanning mutagenesis of the transmembrane domain of the amyloid precursor protein. *Proc Natl Acad Sci USA*. 1999;96(6):3053–8.
- Hashimoto T, Adams KW, Fan Z, McLean PJ, Hyman BT. Characterization of oligomer formation of amyloid-beta peptide using a split-luciferase complementation assay. *J Biol Chem*. 2011;286(31):27081–91.
- Tsubuki S, Takaki Y, Saido TC. Dutch, Flemish, Italian, and Arctic mutations of APP and resistance of Aβ to physiologically relevant proteolytic degradation. *Lancet*. 2003;361(9373):1957–8.
- Masuda A, Kobayashi Y, Kogo N, Saito T, Saido TC, Itohara S. Cognitive deficits in single App knock-in mouse models. *Neurobiol Learn Mem*. 2016;135:73–82.
- Zhang H, Sun S, Wu L, Pchitskaya E, Zakharova O, Fon Tacer K, Bezprozvanny I. Store-operated calcium channel complex in postsynaptic spines: a new therapeutic target for Alzheimer's disease treatment. *J Neurosci*. 2016;36(47):11837–50.
- Zhang H, Wu L, Pchitskaya E, Zakharova O, Saito T, Saido T, Bezprozvanny I. Neuronal store-operated calcium entry and mushroom spine loss in amyloid precursor protein knock-in mouse model of Alzheimer's disease. *J Neurosci*. 2015;35(39):13275–86.
- Nakazono T, Lam TN, Patel AY, Kitazawa M, Saito T, Saido TC, Igarashi KM. Impaired in vivo gamma oscillations in the medial entorhinal cortex of knock-in Alzheimer model. *Front Syst Neurosci*. 2017;11:48.
- Huang Y, Skwarek-Maruszewska A, Horre K, Vandeweyer E, Wolfs L, Snellinx A, Saito T, Radaelli E, Corthout N, Colombelli J, et al. Loss of GPR3 reduces the amyloid plaque burden and improves memory in Alzheimer's disease mouse models. *Sci Transl Med*. 2015;7(309):309ra164.
- Kidana K, Tatebe T, Ito K, Hara N, Kakita A, Saito T, Takatori S, Ouchi Y, Ikeuchi T, Makino M, et al. Loss of kallikrein-related peptidase 7 exacerbates amyloid pathology in Alzheimer's disease model mice. *EMBO Mol Med*. 2018;10:e8184.
- Lalonde R, Fukuchi K, Strazielle C. APP transgenic mice for modelling behavioural and psychological symptoms of dementia (BPSD). *Neurosci Biobehav Rev*. 2012;36(5):1357–75.
- Latif-Hernandez A, Shah D, Craessaerts K, Saido T, Saito T, De Strooper B, Van der Linden A, D'Hooge R. Subtle behavioral changes and increased prefrontal-hippocampal network synchronicity in APP(NL-G-F) mice before prominent plaque deposition. *Behav Brain Res*. 2017. <https://doi.org/10.1016/j.bbr.2017.11.017>.
- Whyte LS, Hemsley KM, Lau AA, Hassiotis S, Saito T, Saido TC, Hopwood JJ, Sargeant TJ. Reduction in open field activity in the absence of memory deficits in the AppNL-G-F knock-in mouse model of Alzheimer's disease. *Behav Brain Res*. 2018;336:177–81.
- Bourin M. Animal models for screening anxiolytic-like drugs: a perspective. *Dialogues Clin Neurosci*. 2015;17(3):295–303.
- Carobrez AP, Bertoglio LJ. Ethological and temporal analyses of anxiety-like behavior: the elevated plus-maze model 20 years on. *Neurosci Biobehav Rev*. 2005;29(8):1193–205.
- Calzavara MB, Patti CL, Lopez GB, Abilio VC, Silva RH, Frussa-Filho R. Role of learning of open arm avoidance in the phenomenon of one-trial tolerance to the anxiolytic effect of chlordiazepoxide in mice. *Life Sci*. 2005;76(19):2235–46.
- Jurgenson M, Aonurm-Helm A, Zharkovsky A. Behavioral profile of mice with impaired cognition in the elevated plus-maze due to a

- deficiency in neural cell adhesion molecule. *Pharmacol Biochem Behav.* 2010;96(4):461–8.
31. Bertoglio LJ, Carobrez AP. Previous maze experience required to increase open arms avoidance in rats submitted to the elevated plus-maze model of anxiety. *Behav Brain Res.* 2000;108(2):197–203.
 32. Maren S, Phan KL, Liberzon I. The contextual brain: implications for fear conditioning, extinction and psychopathology. *Nat Rev Neurosci.* 2013;14(6):417–28.
 33. Tovote P, Fadok JP, Luthi A. Neuronal circuits for fear and anxiety. *Nat Rev Neurosci.* 2015;16(6):317–31.
 34. Shoji H, Takao K, Hattori S, Miyakawa T. Contextual and cued fear conditioning test using a video analyzing system in mice. *J Vis Exp.* 2014;85:e50871.
 35. Dineley KT, Xia X, Bui D, Sweatt JD, Zheng H. Accelerated plaque accumulation, associative learning deficits, and up-regulation of alpha 7 nicotinic receptor protein in transgenic mice co-expressing mutant human presenilin 1 and amyloid precursor proteins. *J Biol Chem.* 2002;277(25):22768–80.
 36. Hamann S, Monarch ES, Goldstein FC. Impaired fear conditioning in Alzheimer's disease. *Neuropsychologia.* 2002;40(8):1187–95.
 37. Hoefer M, Allison SC, Schauer GF, Neuhaus JM, Hall J, Dang JN, Weiner MW, Miller BL, Rosen HJ. Fear conditioning in frontotemporal lobar degeneration and Alzheimer's disease. *Brain.* 2008;131(Pt 6):1646–57.
 38. Knafo S, Venero C, Merino-Serrais P, Fernaud-Espinosa I, Gonzalez-Soriano J, Ferrer I, Santpere G, DeFelipe J. Morphological alterations to neurons of the amygdala and impaired fear conditioning in a transgenic mouse model of Alzheimer's disease. *J Pathol.* 2009;219(1):41–51.
 39. Harrison FE, Reiserer RS, Tomarken AJ, McDonald MP. Spatial and nonspatial escape strategies in the Barnes maze. *Learn Mem.* 2006;13(6):809–19.
 40. Sharma S, Rakoczy S, Brown-Borg H. Assessment of spatial memory in mice. *Life Sci.* 2010;87(17–18):521–36.
 41. O'Leary TP, Brown RE. Visuo-spatial learning and memory deficits on the Barnes maze in the 16-month-old APP^{swe}/PS1^{dE9} mouse model of Alzheimer's disease. *Behav Brain Res.* 2009;201(1):120–7.
 42. Reiserer RS, Harrison FE, Syverud DC, McDonald MP. Impaired spatial learning in the APP^{swe} + PSEN1^{DeltaE9} bigenic mouse model of Alzheimer's disease. *Genes Brain Behav.* 2007;6(1):54–65.
 43. Yassine N, Lazaris A, Dorner-Ciossek C, Despres O, Meyer L, Maitre M, Mensah-Nyagan AG, Cassel JC, Mathis C. Detecting spatial memory deficits beyond blindness in tg2576 Alzheimer mice. *Neurobiol Aging.* 2013;34(3):716–30.
 44. Floresco SB, Jentsch JD. Pharmacological enhancement of memory and executive functioning in laboratory animals. *Neuropsychopharmacology.* 2011;36(1):227–50.
 45. Humby T, Wilkinson LS. Assaying dissociable elements of behavioural inhibition and impulsivity: translational utility of animal models. *Curr Opin Pharmacol.* 2011;11(5):534–9.
 46. Chudasama Y. Animal models of prefrontal-executive function. *Behav Neurosci.* 2011;125(3):327–43.
 47. Stopford CL, Thompson JC, Neary D, Richardson AM, Snowden JS. Working memory, attention, and executive function in Alzheimer's disease and frontotemporal dementia. *Cortex.* 2012;48(4):429–46.
 48. Filali M, Lalonde R, Theriault P, Julien C, Calon F, Planel E. Cognitive and non-cognitive behaviors in the triple transgenic mouse model of Alzheimer's disease expressing mutated APP, PS1, and Mapt (3xTg-AD). *Behav Brain Res.* 2012;234(2):334–42.
 49. Papadopoulos P, Rosa-Neto P, Rochford J, Hamel E. Pioglitazone improves reversal learning and exerts mixed cerebrovascular effects in a mouse model of Alzheimer's disease with combined amyloid-beta and cerebrovascular pathology. *PLoS ONE.* 2013;8(7):e68612.
 50. Zhuo JM, Prakasam A, Murray ME, Zhang HY, Baxter MG, Sambamurti K, Nicolle MM. An increase in A β 42 in the prefrontal cortex is associated with a reversal-learning impairment in Alzheimer's disease model Tg2576 APP^{swe} mice. *Curr Alzheimer Res.* 2008;5(4):385–91.
 51. Ameen-Ali KE, Wharton SB, Simpson JE, Heath PR, Sharp P, Berwick J. Review: neuropathology and behavioural features of transgenic murine models of Alzheimer's disease. *Neuropathol Appl Neurobiol.* 2017;43(7):553–70.
 52. Flood DG, Reaume AG, Dorfman KS, Lin YG, Lang DM, Trusko SP, Savage MJ, Annaert WG, De Strooper B, Siman R, et al. FAD mutant PS-1 gene-targeted mice: increased A β 42 and A β deposition without APP overproduction. *Neurobiol Aging.* 2002;23(3):335–48.
 53. Webster SJ, Bachstetter AD, Van Eldik LJ. Comprehensive behavioral characterization of an APP/PS-1 double knock-in mouse model of Alzheimer's disease. *Alzheimers Res Ther.* 2013;5(3):28.
 54. Li H, Guo Q, Inoue T, Polito VA, Tabuchi K, Hammer RE, Pautler RG, Taffet GE, Zheng H. Vascular and parenchymal amyloid pathology in an Alzheimer disease knock-in mouse model: interplay with cerebral blood flow. *Mol Neurodegener.* 2014;9:28.
 55. Reaume AG, Howland DS, Trusko SP, Savage MJ, Lang DM, Greenberg BD, Siman R, Scott RW. Enhanced amyloidogenic processing of the beta-amyloid precursor protein in gene-targeted mice bearing the Swedish familial Alzheimer's disease mutations and a "humanized" A β sequence. *J Biol Chem.* 1996;271(38):23380–8.
 56. Walker JM, Fowler SW, Miller DK, Sun AY, Weisman GA, Wood WG, Sun GY, Simonyi A, Schachtman TR. Spatial learning and memory impairment and increased locomotion in a transgenic amyloid precursor protein mouse model of Alzheimer's disease. *Behav Brain Res.* 2011;222(1):169–75.
 57. Romberg C, Horner AE, Bussey TJ, Saksida LM. A touch screen-automated cognitive test battery reveals impaired attention, memory abnormalities, and increased response inhibition in the TgCRND8 mouse model of Alzheimer's disease. *Neurobiol Aging.* 2013;34(3):731–44.
 58. Craig D, Mirakhor A, Hart DJ, McLroy SP, Passmore AP. A cross-sectional study of neuropsychiatric symptoms in 435 patients with Alzheimer's disease. *Am J Geriatr Psychiatry.* 2005;13(6):460–8.
 59. Lyketsos CG, Sheppard JM, Steinberg M, Tschanz JA, Norton MC, Steffens DC, Breitner JC. Neuropsychiatric disturbance in Alzheimer's disease clusters into three groups: the Cache County study. *Int J Geriatr Psychiatry.* 2001;16(11):1043–53.
 60. Chung JA, Cummings JL. Neurobehavioral and neuropsychiatric symptoms in Alzheimer's disease: characteristics and treatment. *Neurol Clin.* 2000;18(4):829–46.
 61. Hart DJ, Craig D, Compton SA, Critchlow S, Kerrigan BM, McLroy SP, Passmore AP. A retrospective study of the behavioural and psychological symptoms of mid and late phase Alzheimer's disease. *Int J Geriatr Psychiatry.* 2003;18(11):1037–42.
 62. Shin IS, Carter M, Masterman D, Fairbanks L, Cummings JL. Neuropsychiatric symptoms and quality of life in Alzheimer disease. *Am J Geriatr Psychiatry.* 2005;13(6):469–74.
 63. Logsdon AF, Lucke-Wold BP, Turner RC, Li X, Adkins CE, Mohammad AS, Huber JD, Rosen CL, Lockman PR. A mouse model of focal vascular injury induces astrocyte reactivity, tau oligomers, and aberrant behavior. *Arch Neurosci.* 2017;4(2):e44254.
 64. Mannix R, Berglass J, Berkner J, Moleus P, Qiu J, Andrews N, Gunner G, Berglass L, Jantzie LL, Robinson S, et al. Chronic gliosis and behavioral deficits in mice following repetitive mild traumatic brain injury. *J Neurosurg.* 2014;121(6):1342–50.
 65. Mouzon BC, Bachmeier C, Ojo JO, Acker CM, Ferguson S, Paris D, Ait-Ghezala G, Crynen G, Davies P, Mullan M, et al. Lifelong behavioral and neuropathological consequences of repetitive mild traumatic brain injury. *Ann Clin Transl Neurol.* 2018;5(1):64–80.
 66. Ojo JO, Mouzon B, Algamil M, Leary P, Lynch C, Abdullah L, Evans J, Mullan M, Bachmeier C, Stewart W, et al. Chronic repetitive mild traumatic brain injury results in reduced cerebral blood flow, axonal injury, gliosis, and increased t-tau and tau oligomers. *J Neuropathol Exp Neurol.* 2016;75(7):636–55.
 67. Howerton AR, Roland AV, Bale TL. Dorsal raphe neuroinflammation promotes dramatic behavioral stress dysregulation. *J Neurosci.* 2014;34(21):7113–23.
 68. Savonenko AV, Xu GM, Price DL, Borchelt DR, Markowska AL. Normal cognitive behavior in two distinct congenic lines of transgenic mice hyperexpressing mutant APP SWE. *Neurobiol Dis.* 2003;12(3):194–211.
 69. Lalonde R, Dumont M, Fukuchi K, Strazielle C. Transgenic mice expressing the human C99 terminal fragment of betaAPP: effects on spatial learning, exploration, anxiety, and motor coordination. *Exp Gerontol.* 2002;37(12):1401–12.
 70. Cryan JF, Sweeney FF. The age of anxiety: role of animal models of anxiety in drug discovery. *Br J Pharmacol.* 2011;164(4):1129–61.
 71. Olivier JD, Vinkers CH, Olivier B. The role of the serotonergic and GABA system in translational approaches in drug discovery for anxiety disorders. *Front Pharmacol.* 2013;4:74.

72. Shoji H, Takao K, Hattori S, Miyakawa T. Age-related changes in behavior in C57BL/6 J mice from young adulthood to middle age. *Mol Brain*. 2016;9:11.
73. Sakakibara Y, Kasahara Y, Hall FS, Lesch KP, Murphy DL, Uhl GR, Sora I. Developmental alterations in anxiety and cognitive behavior in serotonin transporter mutant mice. *Psychopharmacology (Berlin)*. 2014;231(21):4119–33.
74. Anagnostaras SG, Josselyn SA, Frankland PW, Silva AJ. Computer-assisted behavioral assessment of Pavlovian fear conditioning in mice. *Learn Mem*. 2000;7(1):58–72.
75. Anagnostaras SG, Wood SC, Shuman T, Cai DJ, Leduc AD, Zurn KR, Zurn JB, Sage JR, Herrera GM. Automated assessment of pavlovian conditioned freezing and shock reactivity in mice using the video freeze system. *Front Behav Neurosci*. 2010;4:158.

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