



Complete Genome Sequences of Five Foot-and-Mouth Disease Viruses of Serotype A Isolated from Cattle in Nigeria between 2013 and 2015

Frank Vandebussche,^a Elisabeth Mathijs,^a Hussaini G. Ularanu,^b David O. Ehizibolo,^b Andy Haegeman,^c David Lefebvre,^c Annebel R. De Vleeschauer,^c Steven Van Borm,^a Kris De Clercq^c

^aMolecular Platform, Veterinary and Agrochemical Research Centre, Ukkel, Belgium

^bFMD Laboratory, Viral Research Division, National Veterinary Research Institute (NVRI), Vom, Nigeria

^cVesicular and Exotic Diseases Unit, Veterinary and Agrochemical Research Centre, Ukkel, Belgium

ABSTRACT The complete genome sequences of 5 foot-and-mouth disease viruses of serotype A are reported here. These viruses originate from outbreaks in northern Nigeria in 2013 to 2015 and belong to the A/AFRICA/G-IV lineage.

Foot-and-mouth disease virus (FMDV) is a positive-sense single-stranded RNA virus of the genus *Aphthovirus* (family *Picornaviridae*) that causes a highly contagious vesicular disease in cloven-hooved animals. The virus is classified into 7 immunologically distinct serotypes (O, A, C, Asia 1, South African territories [SAT] 1, SAT 2, and SAT 3), each containing numerous genetically and geographically distinct evolutionary lineages or topotypes (1). FMDV is endemic in Nigeria, with 4 of the 7 serotypes (i.e., A, O, SAT 1, and SAT 2) reported during the last two decades (2–4). A recent study described the isolation and characterization of 9 FMDV-A strains (lineage A/AFRICA/G-IV) from northern Nigeria during 2013 to 2015 (5). In 2017, a closely related FMDV-A strain crossed the Sahara and caused outbreaks in Algeria (6, 7) and Tunisia (8). Here, we report the complete genome sequences of 5 FMDV-A strains that were isolated from epithelial tissue samples from Nigerian cattle showing typical foot-and-mouth disease (FMD) lesions.

All samples were collected from cattle herds located in northern Nigeria between 2013 and 2015. Total RNA was extracted from cell culture supernatant with a NucleoSpin RNA virus kit (Macherey-Nagel) and treated with Baseline-ZERO DNase (Epicentre) to remove host DNA. cDNA was synthesized according to the manufacturer's instructions using SuperScript IV reverse transcriptase (Thermo Fisher Scientific), an anchored oligo(dT) primer, and an FMDV-specific internal primer. Sequencing libraries were prepared using a Nextera XT kit (Illumina) as described in the user's manual. The fragment length distributions of the libraries were verified on a Bioanalyzer system (Agilent Technologies), and libraries were quantified using a library quantification kit (Kapa Biosystems). Sequencing was performed at the Biotechnology and Molecular Biology Platform (WIV-ISP, Brussels, Belgium) using a MiSeq reagent kit version 3 (Illumina) with 2 × 300-bp paired-end sequencing on a MiSeq system (Illumina).

Raw reads were trimmed using Trimmomatic version 0.36 with a quality cutoff of 15, a sliding window of 4 nucleotides, and a minimum length cutoff of 50 nucleotides (9). *De novo* assembly of the L fragment was performed using Iterative Virus Assembler (IVA) version 1.0.8 (10) and SPAdes (version 3.9.0) (11). The S fragment and the beginning and end regions of the L fragment were verified with Sanger sequencing. The genomes were annotated using the Genome Annotation Transfer Utility (GATU)

Received 11 January 2018 Accepted 16 January 2018 Published 15 February 2018

Citation Vandebussche F, Mathijs E, Ularanu HG, Ehizibolo DO, Haegeman A, Lefebvre D, De Vleeschauer AR, Van Borm S, De Clercq K. 2018. Complete genome sequences of five foot-and-mouth disease viruses of serotype A isolated from cattle in Nigeria between 2013 and 2015. *Genome Announc* 6:e00039-18. <https://doi.org/10.1128/genomeA.00039-18>.

Copyright © 2018 Vandebussche et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Steven Van Borm, steven.vanborm@codacerva.be.

(12) with the FMDV strain FMDV/A/El-Fayoum/Egypt/2014 (GenBank accession number KP940474) serving as the reference genome.

Complete genome sequences were obtained for all 5 isolates, with lengths ranging from 8,126 to 8,135 nucleotides. The first 3 to 4 nucleotides of the genomes and the composition or length of the poly(C) tract could not be determined. The genomes were predicted to contain a single open reading frame (ORF) of 6,990 to 6,996 nucleotides encoding a polyprotein of 2,329 to 2,331 amino acids.

Accession number(s). The complete genome sequences for FMDV A/NIG/1/13, A/NIG/1/15, A/NIG/3/15, A/NIG/5/15, and A/NIG/6/15 have been deposited in GenBank under the accession numbers [MG725872](#) to [MG725876](#).

ACKNOWLEDGMENTS

Special thanks go to the technical staff of the Vesicular and Exotic Diseases Unit of CODA-CERVA. The support of Pierre Kerkhofs, Director General of CODA-CERVA, and David Shamaki, Acting Executive Director of NVRI, is highly appreciated.

The study that yielded these results was funded by the Veterinary and Agrochemical Research Centre (CODA-CERVA), Ukkel, Belgium, and the OIE Laboratory Twinning project between CODA-CERVA (Belgium) and NVRI (Nigeria) as part of a capacity-building program.

REFERENCES

- Samuel AR, Knowles NJ. 2001. Foot-and-mouth disease type O viruses exhibit genetically and geographically distinct evolutionary lineages (topotypes). *J Gen Virol* 82:609–621. <https://doi.org/10.1099/0022-1317-82-3-609>.
- Fasina FO, Connell DR, Talabi OA, Lazarus DD, Adeleke GA, Olusanya TP, Hernandez JA. 2013. Foot-and-mouth disease virus strains and examination of exposure factors associated with seropositivity of cattle herds in Nigeria during 2007–2009. *Prev Vet Med* 109:334–342. <https://doi.org/10.1016/j.prevetmed.2012.10.004>.
- Ularanu HG, Ibu JO, Wood BA, Abenga JN, Lazarus DD, Wungak YS, Knowles NJ, Wadsworth J, Mioulet V, King DP, Shamaki D, Adah MI. 2017. Characterization of foot-and-mouth disease viruses collected in Nigeria between 2007 and 2014: evidence for epidemiological links between West and East Africa. *Transbound Emerg Dis* 64:1867–1876. <https://doi.org/10.1111/tbed.12584>.
- Vosloo W, Bastos AD, Sangare O, Hargreaves SK, Thomson GR. 2002. Review of the status and control of foot and mouth disease in sub-Saharan Africa. *Rev Sci Tech* 21:437–449. <https://doi.org/10.20506/rst.21.3.1349>.
- Ehizibolo DO, Haegeman A, De Vleeschauwer AR, Umoh JU, Kazeem HM, Okolocha EC, Van Borm S, De Clercq K. 2017. Detection and molecular characterization of foot and mouth disease viruses from outbreaks in some states of Northern Nigeria 2013–2015. *Transbound Emerg Dis* 64:1979–1990. <https://doi.org/10.1111/tbed.12602>.
- ProMED-mail. 31 March 2016. Foot & mouth disease—Algeria (02): (RE) bovine, st A, new strain, OIE. ProMED-mail archive no. 20170331.4939480. <http://www.promedmail.org>.
- FAO World Reference Laboratory for Foot-and-Mouth Disease. 2017. Genotyping report: FMDV serotype A. Country: Algeria. FAO World Reference Laboratory for Foot-and-Mouth Disease (WRLFMD), The Pirbright Institute, Surrey, United Kingdom.
- ProMED-mail. 28 April 2017. Foot & mouth disease—Tunisia: (BZ) bovine, serotype A, OIE. ProMED-mail archive no. 20170428.5002447. <http://www.promedmail.org>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Hunt M, Gall A, Ong SH, Brener J, Ferns B, Goulder P, Nastouli E, Keane JA, Kellam P, Otto TD. 2015. IVA: accurate *de novo* assembly of RNA virus genomes. *Bioinformatics* 31:2374–2376. <https://doi.org/10.1093/bioinformatics/btv120>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Tcherepanov V, Ehlers A, Upton C. 2006. Genome Annotation Transfer Utility (GATU): rapid annotation of viral genomes using a closely related reference genome. *BMC Genomics* 7:150. <https://doi.org/10.1186/1471-2164-7-150>.