Autologous costal chondral transplantation and costa-derived chondrocyte implantation: emerging surgical techniques

Youshui Gao^(D), Junjie Gao, Hengyuan Li, Dajiang Du, Dongxu Jin, Minghao Zheng and Changqing Zhang

Abstract: It is a great challenge to cure symptomatic lesions and considerable defects of hyaline cartilage due to its complex structure and poor self-repair capacity. If left untreated, unmatured degeneration will cause significant complications. Surgical intervention to repair cartilage may prevent progressive joint degeneration. A series of surgical techniques, including biological augmentation, microfracture and bone marrow stimulation, autologous chondrocyte implantation (ACI), and allogenic and autogenic chondral/osteochondral transplantation, have been used for various indications. However, the limited repairing capacity and the potential pitfalls of these techniques cannot be ignored. Increasing evidence has shown promising outcomes from ACI and cartilage transplantation. Nevertheless, the morbidity of autologous donor sites and limited resource of allogeneic bone have considerably restricted the wide application of these surgical techniques. Costal cartilage, which preserves permanent chondrocytes and the natural osteochondral junction, is an ideal candidate for the restoration of cartilage defects. Several in vitro and in vivo studies have shown good performance of costal cartilage transplantation. Although costal cartilage is a classic donor in plastic and cosmetic surgery, it is rarely used in skeletal cartilage restoration. In this review, we introduce the fundamental properties of costal cartilage and summarize costaderived chondrocyte implantation and costal chondral/osteochondral transplantation. We will also discuss the pitfalls and pearls of costal cartilage transplantation. Costal chondral/ osteochondral transplantation and costa-based chondrocytotherapy might be up-and-coming surgical techniques for recalcitrant cartilage lesions.

Keywords: cartilage defect, chondrocyte, costal cartilage, cytotherapy, tissue engineering

Received: 18 April 2019; revised manuscript accepted: 29 August 2019.

Introduction

Symptomatic osteochondral lesions of the diarthrosis, such as the hip, tibiofemoral, patellofemoral and tibiotalar joints, are clinically challenging due to the poor self-repair capacity of cartilage and the accelerated deterioration under a weightbearing environment.^{1,2} If a considerable cartilage defect is left untreated, degenerative changes and unmatured osteoarthritis inevitably arise.^{3,4} Advancing diagnostic techniques⁵ revealed that recalcitrant arthropathies in young people become notable diseases. Surgical interventions to repair cartilage could prevent the progression of joint degeneration.⁶

A series of surgical techniques are used for cartilage repair, with varied indications and surgical outcomes.^{7,8} Biological augmentation, including platelet-rich plasma (PRP),⁹ bone marrow aspirate concentrate (BMAC) and mesenchymal stem cells are used solely or synergistically for cartilage repair.² Microfracture is a traditional method for obtaining functional recovery and pain relief at short- and mid-term follow up.^{10–12} However, Ther Adv Musculoskel Dis

2019, Vol. 11: 1–16 DOI: 10.1177/ 1759720X19877131

© The Author(s), 2019. Article reuse guidelines: sagepub.com/journalspermissions

Correspondence to: Youshui Gao

Department of Orthopaedic Surgery, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, 600 Yishan Road, Shanghai 200233, China

Centre for Orthopaedic Translational Research, Medical School, University of Western Australia, Nedlands 6009, WA, Australia

Perron Institute for Neurological and Translational Science, Nedlands 6009, WA, Australia

gaoyoushui@sjtu.edu.cn Changging Zhang

Department of Orthopaedic Surgery, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, Shanghai 200233, China

Institute of Microsurgery on Extremities, Shanghai 200233, China **zhangcq@sjtu.edu.cn**

Junjie Gao

Department of Orthopaedic Surgery, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, Shanghai, China

Centre for Orthopaedic Translational Research, University of Western Australia, Nedlands, WA, Australia

Perron Institute for Neurological and Translational Science, Nedlands, WA, Australia

Hengyuan Li

Department of Orthopaedics, Second Affiliated Hospital of Zhejiang University School of Medicine, Zhejiang, China

Centre for Orthopaedic Translational Research, Medical School, University of Western Australia, Nedlands, WA, Australia

```
journals.sagepub.com/home/tab
```



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (http://www.creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

Perron Institute for Neurological and Translational Science, Nedlands, WA, Australia

Dajiang Du Dongxu Jin

Department of Orthopaedic Surgery, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, Shanghai, China

Minghao Zheng

Centre for Orthopaedic Translational Research, Medical School, University of Western Australia, Nedlands, WA, Australia Perron Institute for Neurological and Translational Science, Nedlands, WA, Australia many factors affect bone marrow stimulation techniques, which yield heterogeneous outcomes.^{10,13,14} Allografting immediately restores the contour of a large osteochondral defect and shows promising functional and radiographic outcomes.15,16 Mosaicplasty, in combination with fresh osteochondral allografting, regains frictionless articular cartilage with the superiority of a one-stage process and fast rehabilitation.¹⁷ Fresh and juvenile allograft cartilage is beneficial for osteochondral defects of large loading articulations and the small joints of the hands and feet.¹⁸⁻²¹ Autologous chondrocyte implantation (ACI) consists of a cartilage biopsy from a nonweightbearing area, in vitro chondrocyte culture and expansion, and surgical implantation of the collected chondrocytes.22 Long-term follow up of ACI has confirmed clinical improvement for osteochondritis dissecans.²³ Although there is no randomized controlled study comparing the abovementioned surgical techniques, increasing evidence shows more promising outcomes from allograft transplantation. Osteochondral allograft might be the most suitable protocol for prior failed cartilage repair.24-26 Histological methods are helpful in evaluating the microscopic appearance of the regenerated tissue following various techniques.²⁷ Microscopic observation of failed repair following abrasion, periosteal flap grafting and ACI revealed a mixed composition of collagen (Col) I, II and X. Fibrous proliferation was also obvious, but hyaline cartilage regeneration was limited.²⁸ A more recent study found that specimens from failed ACI, microfracture and periosteal transplantation were mainly composed of fibrous connective tissue and fibrocartilage. However, no Col X was found, and hyaline cartilage formation was scarce.²⁹ The distinct shortcomings of allogeneic osteochondral grafting stem from the imbalance between the large clinical demand and limited resources. Although autologous osteochondral plugs and chondrocytes are obtained from nonweight-bearing areas of diarthrosis, the total volume is restricted, and the potential long-term side effects should be noted.

Costal cartilage, which preserves permanent chondrocytes, is an ideal candidate for the repair of cartilage defects. The harvesting of costal cartilage does not interfere with the articular structure, and potential complications are avoidable and minimal. Autologous costal cartilage transplantation is widely used in the plastic and cosmetic surgery due to craniofacial microsomia,³⁰ auricles of congenital microtia,³¹ tracheal reconstruction³² and congenital tracheal stenosis.³³ Meanwhile, overgrowth or undergrowth of the costal cartilage might lead to deformity of the thoracic cavity.^{34–36} Therefore, the use of costal cartilage as the attracting donor and underlying pathology was examined in clinical and translational studies.

In this review, we first introduce the fundamental properties of costal cartilage, which contribute to the basis of its experimental and clinical applications (Figure 1). Second, the pre-clinical and clinical uses of costal cartilage and costa-derived chondrocyte implantation are summarized. Third, we also discuss the pitfalls and pearls of costal cartilage transplantation and lessons from its use in plastic surgery. Finally, the existing challenges and potential strategies are highlighted.

The properties of costal cartilage

Human articular cartilage is a multilayered structure containing chondrocytes in different stages of maturation and an ossification microenvironment (Figure 2). The extracellular matrix (ECM), which is mainly composed of Col II for hyaline cartilage, also contains Col III/IX/XI/ VI/X, glycosaminoglycans (GAGs), proteoglycans and glycoproteins.⁴ The ECM is the residential microenvironment of chondrocytes, and it plays an important role in the regulation of chondrogenesis and pathological processes.

Similarly, but differently, the growth plates of foetal bovine ribs are divisible into five different zones: the hypertrophic, lower proliferative, upper proliferative, intermediate and resting zones.³⁷ The deposition of perlecan increases during proliferation and might be sulphated. Costal cartilage undergoes a natural mineralization, which is not considered a degenerative change. For example, mineralization of the first rib begins at the end of puberty, and large cartilage canals with vascular invasion and perivascular connective tissue present in the middle of the second decade. This kind of osteogenesis should not be simply categorized as intramembranous or endochondral ossification.38 Human ribs have two separate cartilaginous regions: one region forms the joint with the sternum, and the other region is for longitudinal growth. The cartilage of the rib becomes vascularized immediately after birth, but mineralization is arrested only after differentiation of the costal chondrocyte has advanced to the late



Figure 1. Schematic of costal chondral/osteochondral transplantation and costa-derived chondrocytotherapy. The costal junction may indicate a deep osteochondral lesion, and the costal cartilage may be used for full-thickness cartilage defects. Alternatively, costal cartilage may be cut into small pieces, and digested and expanded *in vitro* to obtain sufficient chondrocytes. The costa-derived chondrocytes are functional seed cells to cure cartilage defects and growth plate injury.

stage. In vitro findings indicated that parathyroid hormone (PTH) stimulates costal chondrocytes to exhibit the phenotype of fully differentiated hypertrophy.³⁹ Therefore, PTH is essential for the late differentiation of costal cartilage. The costal cartilage is surrounded by a perichondrium, which is a kind of dense connective tissue that contains a special niche of housing chondrogenic pioneering cells, and it critically contributes to the costal cartilage regeneration.⁴⁰ Samples of ribs six to eight were collected from patients with pectus carinatum for microscopic morphological study. Scanning electron microscopy and atomic force microscopy showed collagen fibres approximately 600 nm in diameter that assembled into a large complex running parallel to the cartilage, with a unique straw-like structure. Chondrocytes likely occur singly or as doublets, and centrally located chondrocytes produce more aggrecan. The expression of decorin, Col2A1, aggrecan and tissue inhibitor of metalloproteinase 1 (TIMP1) is high, which indicates the underdifferentiated nature of costal cartilage.⁴¹

Biological properties

Investigators are enthusiastic about comparing the biological properties of chondrocytes from different origins. Chondrocytes derived from rib, ear and nose were isolated separately in one study and expanded in a cocktail composed of transforming growth factor beta-1 (TGF- β 1), fibroblast growth factor 2 (FGF-2) and alpha- and beta-receptor ligand of platelet-derived growth factor (PDGF-bb),⁴² altogether, TFP. The capacity and quality of the induced chondrogenesis were compared thereafter. A significant increase in glycosaminoglycan and messenger ribonucleic acid (mRNA) expression of Col II was found in costa-derived chondrocytes but the quality of



Figure 2. Multilayered structure of the cartilage from the femoral head.

Chondrocytes in different stages of maturation are buried in the extracellular matrix with varied ossification (Goldner Trichome staining).

generated cartilage pellets was inferior.⁴² A recent similar study also used chondrocytes from ear, nasoseptal and costal cartilage to compare the cell yield, proliferation, chondrogenesis and capacity of cartilage formation.⁴³ Three different kinds of cartilage were sensitive to basic fibroblast growth factor (bFGF) within eight passages and showed the strongest capacity of chondrogenesis at the fifth passage. Although minor matrix changes existed, the formed cartilaginous product had mature cartilage features, like the original native

tissue.⁴³ The authors of another study compared cell yield, expansion, and chondrocyte dedifferentiation and redifferentiation capacity between costal and articular cartilage of rabbit.44 These authors found that costal chondrocytes were advantageous in cell yield and expansion. Chondrocytes from both origins became dedifferentiated with serial cultures, but the capacity of redifferentiation improved when these chondrocytes were cocultured with a high density of collagen gel.44 The contraction of collagen gel can reflect the degree of fibroblastic dedifferentiation during chondrocyte expansion. They observed the surface area of the reconstructed cartilage being reduced to 11.11% for articular chondrocytes and 4.34% for the costal chondrocytes at 14 days. When costal chondrocytes from passage four were subjected to high-density three-dimensional (3D) culture, they could regain the hyaline cartilage phenotypes, similar to chondrocytes from passage two.44

Kitaoka and colleagues established a chondrocyte cell line from the costal cartilage of SV40 large T-antigen transgenic mice.⁴⁵ Costa-derived chondrocytes had similar phenotypes as articular cartilage and embryonic limbs, which suggests that costal cartilage is a competent alternative as a donor site for the repair of articular cartilage defect. Taken together, although inconsistencies might exist due to heterogeneity in cell passages and culture media, increasing evidence shows that costal chondrocytes maintain an underlying biological property to exhibit the desired phenotypes of chondrocytes derived from articular cartilage.

Biochemical and biophysical properties

Numerous biochemical and biophysical factors affect the biological behaviour of cultured chondrocytes. Growth factors, chemokines and many other stimulators are used for the promotion of proliferation and prevention of chondrocyte dedifferentiation.^{46,47} Biophysical factors, including cell density, oxygen concentration and ECM, also impact the characteristics of chondrocytes cultured *in vitro*.

The components of the culture medium and seeding density affect the postexpansion chondrogenic potential of costa-derived chondrocytes.⁴⁸ Supplementation of 1 ng/ml TGF- β 1, 5 ng/ml bFGF and 10 ng/ml PDGF increased glycosaminoglycan, decreased the ratio of Col I/Col II and strengthened the compressive property.⁴⁸ A density of 2 million cells/5 mm diameter construct was beneficial in enhancing the matrix synthesis and tensile/compressive properties compared with 3, 4 or 5 million cells/construct,48 which is inconsistent with previous studies that demonstrated the priority of higher chondrocyte density. We believe that the most efficacious seeding density and cocktail of growth factors must be investigated further in self-assembled cartilage formation. The oxygen concentration regulates the expansion and differentiation properties of costa-derived chondrocytes. Low oxygen (5%) in combination with serum-free medium and FGF2 improved cell expansion, reduced dedifferentiation, enhanced redifferentiation capacity into cartilage and ameliorated the proteomic profile.49 The electrical cell membrane of costal chondrocytes is close to that of cardiomyoblast cells, and gadolinium (Gd) blocks ion channels, which results in a significant reduction in membrane capacitance and conductance.⁵⁰

The ECM constitutes the microenvironment of chondrocytes, and it dramatically regulates expansion and dedifferentiation in an *in vitro* culture system.⁵¹ The ECM derived from articular chondrocytes was superior to that from bone marrow and yielded a better ratio of Col 2A1/Col 1A1 and a more cartilage-like ECM.⁵¹ An appropriate artificial matrix also plays a beneficial role in the maintenance of chondrocytes. An artificial matrix of poly(L-lactic-acid) maintained chondrocyte phenotype during *in vitro* expansion, which might be associated with cellular adhesion *via* the $\alpha_{s}\beta_{1}$ integrin.⁵²

Biomechanical properties

One of the most important characteristics of hyaline cartilage lies in its superior biomechanical properties. Physiologically, articular cartilage transfers load between bones and facilitates the frictionless motion of diarthrosis.⁵³ Mechanical stimulation is also a critical biophysical factor that may be used to induce changes in the gene expression of chondrocytes and ECM deposition.⁵⁴ *In vitro* studies using cyclic tensile strain and compressive stimulation directed the ossification mode of mesenchymal stem cells.^{54,55}

Mechanical loading induces costa-derived chondrocytes towards articular cartilage regeneration. Compressive stimuli at the phase of matrix synthesis and maturation increase instantaneous moduli by 92% and 87%, respectively. However, combined compression at both phases did not further strengthen the property beyond a onetime stimulation. In terms of magnitude, passive axial compression of 3.3 kPa and 5.5 kPa increased neocartilage compressive properties by 42% and 48%, respectively. When a further bioactive regimen was supplemented, the tensile properties of formed cartilage was increased 2.6-fold compared with an untreated control.⁵⁶ Murphy and colleagues found that TGF-\u03b31, chondroitinase ABC (C-ABC) and hydrostatic pressure separately or synergistically improved the hyaline cartilage regeneration of costal chondrocytes.⁵⁷ The optimum combination vielded robust hvaline cartilage with a property of tensile modulus of 2 MPa and a compressive instantaneous modulus of 650 kPa.

Costal cartilage from mutant mice lacking $\beta 1$ integrins in their chondrocytes showed round chondrocytes and a loss of polarity. Although the levels of Col II and aggrecan were similar, the absence of $\beta 1$ integrins increased the stiffness and modified the diffusion properties of costal cartilage.⁵⁸ Therefore, $\beta 1$ integrins might play a critical role in the biomechanical function of costal cartilage.

Costal chondral/osteochondral grafting in animal models

Preclinical animal models are essential to reveal the cellular mechanisms and the histological and morphological results of costal cartilage transplantation.⁵⁹ Several studies reported convincing outcomes of costal chondral/osteochondral grafting for the treatment of cartilage and osteochondral defects. Histological and radiographical evaluations show that the transplanted costal chondral/osteochondral graft is capable of fusing with the receipt site, and it restores complex cartilage lesions in multiple animal models. A costal osteochondral plug was transplanted to a fullthickness articular defect of the knee in a rabbit model. Histological union was achieved in all animals. Chondrocyte viability was confirmed microscopically in the 48-week grafts. The general appearance of chondrocytes in the transplanted costa was similar to the host cartilage. The expression of Col II and aggrecan was consistent with normal articular cartilage.⁶⁰ For large osteochondral defects, multiple sliced costal cartilage could be synergistically used to repair the defect. Histological findings revealed that the

intergraft gap between separated costal cartilage was filled by fibrous tissue, and the cleft between transplanted cartilage and host bone was interlinked by newly formed woven bone.⁶¹ The junction and union between transplanted cartilage and recipient tissue is very important for functional recovery.⁶² Alternatively, a cancellous iliac bone block could be isolated and implanted onto the surface of costal cartilage to yield an osteochondral complex. The hybrid structure repaired the large full-thickness cartilage defect.⁶³ However, autologous sternal cartilage transplantation did not yield satisfactory outcomes for articular defects of the radial carpal bone in athletic horses.⁶⁴

With the accumulated data from animal studies, many striking issues have emerged that deepened our understanding of cartilage pathology and accelerated the translational research. First, the local microenvironment plays a crucial role in the regeneration following costal cartilage transplantation.65 To overcome the unfavourable microenvironment of a cartilage defect, Mao and colleagues used ECM derived from allogeneic human articular chondrocytes, which promoted the expansion of human primary chondrocytes in vitro with reduced dedifferentiation.⁵¹ Second, products from platelets promote cartilage repair following costal cartilage grafting. A hybrid technique involving the implantation of hyaline cartilage chips, platelet-poor plasma and intra-articular injection of growth-factor-rich plasma, enhanced chondrogenesis and yielded hyaline cartilage in a sheep model.⁶⁶ Next, the potential of self-repair is closely associated with the position of the cartilage defect and might be site specific. For instance, a 4×4 mm² defect at the load-bearing area of the femoral head achieved a better histological score compared with the head-neck junction area.⁶⁷ In situ tissue engineering for cartilage regeneration also showed an encouraging pilot outcome and potential clinical application. A kind of decellularized cartilage matrix-derived scaffold with a functionalized self-assembly peptide homed endogenous reparative cells and yielded superior hyaline-like cartilage.68

The anatomical characteristics of costal cartilage in many animals are well described.⁶⁹ Cartilage defects and reparative protocols in different animal models should not be simply compared, but the research data provide us a beneficial reference to initiate breakthrough studies in human beings.

Autologous cartilage/costa-derived chondrocyte implantation

Ribs have become a potential donor site of chondrocytes for ACI. ACI with seeding chondrocytes from nonarticular heterotopic cartilage has advantages of lesser donor-site morbidity with a comparable ECM synthesis profile to the articular cartilage.70 Chondrocytotherapy with seeding cells from the rib or nonweight-bearing areas of diarthrotic joints showed a prospective outcome for recalcitrant cartilage lesions. Cell-free scaffolds demonstrated several advantages; however, a recent study indicated an increased 5-year failure rate following implantation of a Col I scaffold, which suggests the clinical importance of seeding cells in cartilage repair.⁷¹ ACI may also be used as a salvage protocol for failed prior cartilage-repairing operation. Ellender and Minas used salvage ACI to treat a 10 cm² full-thickness chondral defect of the femoral head.⁷² The patient had a previous autologous osteochondral mosaicplasty. Chondrocytes were obtained from nonweightbearing areas of the knee, with a size of $0.5 \times 1 \,\mathrm{cm}^2$, which yielded approximately $2 \times 10^5 - 3 \times 10^5$ cells. A total of 48 million cells were produced after in vitro expansion. Notably, the authors used a special collagen to secure the implanted chondrocytes. The patient was free of pain at a 2-year follow up. Images of contrast-enhanced magnetic resonance imaging (MRI) demonstrated repair tissue fill and radiographs showed a normal joint space. The surgical technique was used in another, more difficult case of post-traumatic osteonecrosis of the femoral head.73 Histological staining of the biopsy tissue 15 months postoperatively showed viable chondrocytes, which were profoundly integrated with the subchondral bone. At 18 months postoperatively, computed tomography (CT) scanning demonstrated that the joint space exhibited good congruity. However, there are no case reports using autologous costa-derived chondrocytes for articular repair. On the one hand, a thin cartilage biopsy does not significantly influence the normal structure of the donor joint. On the other hand, in vitro studies show that the density of chondrocytes is higher in articular hyaline cartilage than in costal cartilage. Nevertheless, costa-derived chondrocytes are competitive candidates for chondrocytotherapy. For juvenile patients, long-term complications induced by the loss of articular cartilage must be clarified.

The adjuvant effects of platelet-derived products show inconsistent outcomes of chondrogenesis and cartilage repair during ACI. Platelet-derived fibrin demonstrated positive effects on cartilage repair, and the experimental outcome was better when autologous chondrocytes were used jointly.⁷⁴ PRP was also beneficial for cartilage regeneration at the donor site following costal cartilage harvesting.⁷⁵ A commercially available platelet lysate enhanced chondrocyte proliferation, but the chondrogenic capacity was inferior to the control.⁷⁶

Chondrocytes from the proliferating layer of the growth plate showed more chondrogenic phenotypes, a higher proliferation rate, increased expression of chondrogenic-related genes, higher content of glycosaminoglycan and more chondrogenic extracellular matrix.⁷⁷ Chondrocytes from the growth plate may be isolated, cultured and expanded *in vitro* and used to prevent bone bridge formation.^{78,79} However, no report has used autologous chondrocytes from the growth plate for cartilage repair, primarily due to potential severe complications.

Clinical autologous costal chondral/ osteochondral transplantation

Historically, costal chondral/osteochondral transplantation was largely used in plastic and cosmetic surgery. However, with the accumulated experience in the care of hand, wrist and elbow cartilage disease, costal chondral and osteochondral grafts broadened their indication for weightbearing joints, such as hip, knee and foot.

Costal osteochondral transplantation is an effective method to restore metacarpophalangeal and proximal interphalangeal joints to avoid arthrodesis and arthroplasty.80,81 Costal osteochondral grafting was also used to reconstruct proximal scaphoid due to fracture or necrosis.⁸² For finger joints reconstructed with costal osteochondral graft, Sato and colleagues harvested the fifth or sixth costo-osteochondral junction, which was shaped to fit the defect in metacarpophalangeal joint in three cases, proximal interphalangeal joint in nine, distal interphalangeal joint in three and thumb interphalangeal joint in two. The average time to bone union of the graft was 58 days. The biopsy histology showed that scattered chondrocytes were viable within the ECM and exhibited the same appearance as normal hyaline cartilage.⁸³ Costal cartilage transplantation is indicated to cure trapeziometacarpal osteoarthritis. In a retrospective study including 100 cases, the surgical technique restored thumb stability and strength.84

Costal cartilage transplantation shows encouraging effects for the treatment of intra-articular malunion following distal radius fractures. In a series of seven patients, costal cartilage from the eighth rib was harvested and implanted in a trough created at the epiphysis metaphysis junction of the distal radius. Union was achieved in all cases without complications, and the functional and radiographic outcomes were satisfactory.85 For advanced traumatic arthritis of the radiocarpal joint, costal osteochondral transplantation is a convincing alternative to preserve wrist motion. In a study enrolling 29 cases (7 due to malunited intra-articular distal radius fractures; 18 due to scaphoid osteopathy; and 4 due to Kienböck disease), costal cartilage from the ninth rib showed satisfactory radiographic and histological results. The graft was vital, and no signs of resorption or necrosis were observed.86

The clinical transplantation of autologous costal osteochondral graft was used for osteochondritis dissecans of the *capitulum humeri* secondary to elbow trauma and yielded satisfactory results in two patients.87 In a more recent cohort study of 26 patients, cylindrical costal osteochondral graft was used to cure large defects ($\geq 15 \text{ mm}$ in diameter) of the capitellum due to osteochondritis dissecans. All patients were satisfied with the functional improvement. Osseous union and revascularization of the graft were achieved successfully.88 Sato and colleagues demonstrated that the favourable indication was advanced osteochondritis dissecans of the capitulum in teenagers with significant symptoms.⁸⁹ Costal osteochondral graft was also successfully used to cure fracture-induced osteonecrosis of the radial head in an adolescent boy.90

Although costal chondral/osteochondral transplantation demonstrated remarkable benefits for cartilage defects of nonweight-bearing joints, it is rarely used in the lower limbs. We reported a 20-year-old man with traumatic osteonecrosis of the femoral head.⁹¹ Through the Smith-Petersen approach, the affected hip was dislocated to expose the femoral head, and another surgical team harvested the costal osteochondral graft. The degenerated cartilage and subchondral bone were removed using an 8 mm trephine. The costal chondral graft was trimmed into pieces to match the cylindrical defect. Essential internal fixation with absorbable screws was adopted to enhance the stability of costal grafts. The short-term follow up showed that pain was completely relieved, and ambulation was dramatically improved. We

accumulated more successful cases of costal chondral grafting for cartilage defects of the hip, knee, ankle and the first metatarsophalangeal joint.

Pitfalls and pearls

Costal cartilage transplantation is widely used in clinical practice,^{30,31,92–98} but multiple factors might influence the prognosis. With the increasing evidence of costal cartilage, pitfalls of this emerging surgical technique must also be highlighted. Pitfalls and pearls from autologous costal cartilage grafting in plastic surgery and allogeneic cartilage transplantation are instructive to improve the quality of skeletal reconstruction with costal cartilage.

Warping and overgrowth of the cartilage

Warping of the costal cartilage is a serious complication in plastic surgery.93,99 For costal cartilage transplantation, preoperative evaluation is very important, and the size of harvested cartilage should be sufficient.¹⁰⁰ The surgical technique of transverse slicing of the sixth-seventh costal cartilaginous junction may yield adequate cartilage and was reported as an effective method to prevent warping. Warping of the cartilage may cause the loss of press-fit stability, which suggests that cartilage fixation is an essential procedure. Growth of the transplanted free costochondral grafts was observed in children with hemifacial microsomia. Overgrowth might occur, but the underlying mechanism of costochondral growth has not been definitely elucidated.98

Morphological matching

The demand of the frictionless motion of nonweight-bearing joints demonstrated that morphological matching was not a critical issue for upper limbs. However, the role of contour matching must be addressed in biological restorations of weight-bearing joints. An absolute match never exists, but it must be approximated. Autologous osteochondral graft from the ipsilateral nonweight-bearing area of the femoral head could be harvested to repair the cartilage defects in the weight-bearing surface, and very similar curvatures showed satisfactory clinical and radiographic outcomes at mid-term follow up.¹⁰¹ Although the sphericity of femoral head was not restored to normal, care should be taken to harvest osteochondral plugs with a similar spherity.¹⁰² In rare conditions, autologous osteochondral grafts from the femoral head due to hemiarthroplasty are used to repair cartilage defects of the lateral femoral condyle in multiple osteonecrosis.¹⁰³ We hypothesize that a remodelling mechanism likely includes the plasticity and elasticity of the costal cartilage.

Age and defect size

Cartilage repair is critical for adolescents to avoid unmatured degenerative joint. However, we do not know the exact effect of increasing age on the outcome of cartilage transplantation. One study revealed that arthroscopic matrix-assisted autologous chondrocytotherapy was beneficial for patients older than 40 years with International Cartilage Regeneration and Joint Preservation Society (ICRS) grade 3–4 lesions. Age is not a strict contraindication of cartilage reparative surgery.¹⁰⁴ In our experience, costal chondral grafting relieved pain and improved range of motion at short-term follow up.

ICRS grade, defect size, and symptom duration did not affect the baseline Knee Injury and Osteoarthritis Outcome Score, but the size of the defect was a considerable factor of surgical protocol.¹⁰⁵ The absolute and relative size of the cartilage lesion did not influence postoperative outcomes after osteochondral allograft transplantation for isolated femoral condyle lesions.¹⁰⁶ Large defect size is not a contraindication of costal cartilage transplantation, especially for young patients.

Complications

Postoperative complications are minimal, and cosmetic concerns are acceptable using the advanced techniques of costal harvesting.^{107,108} For patients younger than 10-years old, the risks of the chest-wall deformities and thoracic scoliosis should be noted following harvesting of costal cartilage. The incidence of deformity might be higher for upper ribs. An intact costochondral junction was critical in minimizing these severe complications.¹⁰⁹

Calcification of the costal cartilage is a natural progress that should be carefully evaluated preoperatively using ultrasonography¹¹⁰ or dual-energy CT imaging.¹¹¹ Generally, ossification initiates from the first rib cartilage and progresses up to

the twelfth rib.¹⁰⁰ Costal cartilage calcification is closely associated with ageing.^{112–114} Severe calcification of the costal cartilage might increase the incidence of pneumothorax due to perichondrium adhesion. It is difficult to trim calcified cartilage, which might also become a new predisposition to induce cartilage erosion.

The microenvironment of the cartilage defect is also vital to the healing of transplanted cartilage. Inflammatory responses following cartilage damage might involve neutrophil infiltration and cytokine release, which are closely associated with the consequent repair process.¹¹⁵ Therefore, an exact understanding of the spatiotemporal distribution of the inflammation cascade plays a fundamental role in reconstructive surgery.

Challenges and strategies

The challenges of cartilage degeneration and surgical restoration are great. Although we have a considerable understanding of the biology of chondrocytes and physiology of cartilage, little is known about the natural history of self-repair modulation and specific function of this multilayered structure (Figure 2). The concept of the osteochondral unit must be highlighted in the of cartilage pathogenesis degeneration.¹¹⁶ Cartilage damage is rarely an independent event of chondrocytes, and it intimately involves the whole joint. The uncertainty of the relationship between intra-articular joint disease on imaging and complaints of pain must be further investigated to deepen our understanding of the mechanism of articular pathology.¹¹⁷ Surgical techniques of arthroscopy, although less invasive in facilitating enhanced recovery,118 cannot resolve fundamental problems of the arthropathy.

Costal chondral transplantation and costaderived chondrocytotherapy are promising surgical techniques in caring for challenging cartilage lesions. For patients with inherent or secondary joint disease, healthy hyaline cartilage is inherently limited. In these scenarios, costa should be considered as the origin of chondrocytes. However, the costa is just a donor site of chondrocytes and hyaline cartilage, which might yield optimal outcomes in combination with other advanced surgical techniques or biotechniques. In terms of basic research, the optimal protocol to preserve phenotypes of chondrocytes and increase extracellular matrix production is still at the stage of exploration.^{119,120} Although chondrocytes are more resistant to hypoxia and ischaemia, the effect of microenvironment with irritated inflammation on cellular senescence and death is not fully elucidated currently.121,122 In terms of clinical investigations, observational studies are indispensable in revealing the natural history of arthropathies in young adults, and high-standard clinical trials are desired to reflect the therapeutic effect of costal chondral transplantation and costa-derived chondrocytotherapy.123 In vitro expansion is currently used to yield sufficient chondrocytes;124 however, we do not know whether preculture is necessary for a small lesion. The quantity and quality of transplanted chondrocytes are everlasting topics in cytotherapy.¹¹⁹ The density (number of chondrocytes per defect size) of chondrocyte implantation is multifactorial and might be personalized.²² Tissue engineering might shed light on future methods to strengthen costal chondral transplantation and costa-derived ACI.¹²⁵ Tissue-engineering approaches are based on a strategic liaison between cells, scaffolds and signalling molecules to inspire intrinsic repair capacity or rely on extrinsic materials for regeneration.^{59,126,127} The spatial complexity of cell types and tissue organization demonstrate that routine biomaterials for bone engineering cannot restore a totally biomimetic structure. It is possible to use novel 3D bioprinting technology to biofabricate skeletal architecture, layer by layer, which is especially significant for cartilage engineering due to its complex nature.128,129 Pilot studies indicated that a 3D matrix might yield beneficial effects on the phenotypes and vitality of chondrocytes.^{51,130,131} However, the optimum formulation of synthetic matrix is to be determined.^{52,132} Biomaterial-guided gene therapy is also an attractive strategy for augmenting the intrinsic mechanisms of cartilage repair and suppressing the detrimental inflammatory response, simultaneously.133,134 Stem cells play a fundamental role in cartilage repair.119,127 In situ biofabrication using adipose-derived mesenchymal stem cells is reportedly feasible for cartilage repair.¹²⁴ It is interesting to discern the shift from cartilageand bone marrow- to adipose-derived cells in clinical translational research, but more phase III trials were based on seed cells from cartilage.¹³⁵ Strategies for in situ cartilage repair may be initiated through endogenous reparative cell-homing techniques⁶⁸ to yield so-called in vivo tissue engineering to repair the cartilage defect.¹³⁶ Cytokines and chemokines, including bone morphogenetic

proteins, are beneficial for articular cartilage regeneration.¹³⁷ Detrimental factors suppressing chondrogenesis might be targeted to repair cartilage.¹³⁸ CRISPR-Cas9 may be used to target matrix metallopeptidase 13 in human chondrocytes, resulting in decreased metalloproteinase and increased Col II deposition.¹³⁹ Finally, products of tissue engineering cartilage derived from autologous chondrocytes were used in human trials.^{140,141} Although tissue engineering is currently used for alar lobule restoration and articular defects, we believe more sophisticated commercially available products could challenge defects of cartilage.¹⁴⁰⁻¹⁴²

Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: this study was supported in part by the National Natural Science Foundation of China (no. 81820108020).

Conflict of interest statement

The authors declare that there is no conflict of interest.

ORCID iD

Youshui Gao (D) https://orcid.org/0000-0001-9242-2486

References

- Brophy RH, Wojahn RD and Lamplot JD. Cartilage restoration techniques for the patellofemoral joint. J Am Acad Orthop Surg 2017; 25: 321–329.
- Southworth TM, Naveen NB, Nwachukwu BU, et al. Orthobiologics for focal articular cartilage defects. *Clin Sports Med* 2019; 38: 109–122.
- Gomoll AH and Minas T. The quality of healing: articular cartilage. *Wound Repair Regen* 2014; 22(Suppl. 1): 30–38.
- Carballo CB, Nakagawa Y, Sekiya I, et al. Basic science of articular cartilage. *Clin Sports Med* 2017; 36: 413–425.
- Svard T, Lakovaara M, Pakarinen H, et al. Quantitative MRI of human cartilage in vivo: relationships with arthroscopic indentation stiffness and defect severity. *Cartilage* 2018; 9: 46–54.
- 6. Jungmann PM, Gersing AS, Baumann F, *et al.* Cartilage repair surgery prevents progression of knee degeneration. *Knee Surg Sports Traumatol*

Arthrosc. Epub ahead of print 12 December 2018. DOI: 10.1007/s00167-018-5321-8.

- 7. Hotham WE and Malviya A. A systematic review of surgical methods to restore articular cartilage in the hip. *Bone Joint Res* 2018; 7: 336–342.
- 8. Jordan MA, Van Thiel GS, Chahal J, *et al.* Operative treatment of chondral defects in the hip joint: a systematic review. *Curr Rev Musculoskelet Med* 2012; 5: 244–253.
- Kennedy MI, Whitney K, Evans T, et al. Platelet-rich plasma and cartilage repair. Curr Rev Musculoskelet Med 2018; 11: 573–582.
- Orth P, Gao L and Madry H. Microfracture for cartilage repair in the knee. A systematic review of the contemporary literature. *Knee Surg Sports Traumatol Arthrosc*. Epub ahead of print 18 January 2019. DOI: 10.1007/s00167-019-05359-9.
- Philippon MJ, Schenker ML, Briggs KK, et al. Can microfracture produce repair tissue in acetabular chondral defects? *Arthroscopy* 2008; 24: 46–50.
- 12. Steadman J, Rodkey W and Rodrigo J. Microfracture: surgical technique and rehabilitation to treat chondral defects. *Clin Orthop Relat Res* 2001: S362–S369.
- Chen H, Hoemann CD, Sun J, et al. Depth of subchondral perforation influences the outcome of bone marrow stimulation cartilage repair. *J Orthop Res* 2011; 29: 1178–1184.
- Frisbie D, Oxford J, Southwood L, *et al.* Early events in cartilage repair after subchondral bone microfracture. *Clin Orthop Relat Res* 2003; 407: 215–227.
- Cottom JM and Maker JM. Cartilage allograft techniques and materials. *Clin Podiatr Med Surg* 2015; 32: 93–98.
- Nikolaou VS and Giannoudis PV. History of osteochondral allograft transplantation. *Injury* 2017; 48: 1283–1286.
- 17. Wang D, Chang B, Coxe FR, et al. Clinically meaningful improvement after treatment of cartilage defects of the knee with osteochondral grafts. Am J Sports Med. Epub ahead of print 27 November 2018. DOI: 10.1177/0363546518808030.
- Van Dyke B, Berlet GC, Daigre JL, et al. First metatarsal head osteochondral defect treatment with particulated juvenile cartilage allograft transplantation: a case series. *Foot Ankle Int* 2018; 39: 236–241.

- Gracitelli GC, Tirico LE, McCauley JC, et al. Fresh osteochondral allograft transplantation for fractures of the knee. *Cartilage* 2017; 8: 155–161.
- Tirico LEP, Early SA, McCauley JC, et al. Fresh osteochondral allograft transplantation for spontaneous osteonecrosis of the knee: a case series. Orthop J Sports Med 2017; 5: 2325967117730540.
- Mei XY, Alshaygy IS, Safir OA, *et al.* Fresh osteochondral allograft transplantation for treatment of large cartilage defects of the femoral head: a minimum two-year follow-up study of twenty-two patients. *J Arthroplasty* 2018; 33: 2050–2056.
- 22. Lopez-Alcorocho JM, Aboli L, Guillen-Vicente I, *et al.* Cartilage defect treatment using high-density autologous chondrocyte implantation: two-year follow-up. *Cartilage* 2018; 9: 363–369.
- 23. Beck JJ, Sugimoto D and Micheli L. Sustained results in long-term follow-up of autologous chondrocyte implantation (ACI) for distal femur juvenile osteochondritis dissecans (JOCD). Adv Orthop 2018; 2018: 7912975.
- 24. Rosa D, Di Donato SL, Balato G, *et al.* How to manage a failed cartilage repair: a systematic literature review. *Joints* 2017; 5: 93–106.
- Oladeji LO, Cook JL, Stannard JP, et al. Large fresh osteochondral allografts for the hip: growing the evidence. *Hip Int* 2018; 28: 284–290.
- Gaul F, Tirico LEP, McCauley JC, et al. Long-term follow-up of revision osteochondral allograft transplantation of the ankle. Foot Ankle Int 2018; 39: 522–529.
- Hayashi S, Nakasa T, Ishikawa M, *et al.* Histological evaluation of early-phase changes in the osteochondral unit after microfracture in a full-thickness cartilage defect rat model. *Am J Sports Med* 2018; 46: 3032–3039.
- Nehrer S, Spector M and Minas T. Histologic analysis of tissue after failed cartilage repair procedures. *Clin Orthop Relat Res* 1999; 365: 149–162.
- LaPrade RF, Bursch LS, Olson EJ, et al. Histologic and immunohistochemical characteristics of failed articular cartilage resurfacing procedures for osteochondritis of the knee: a case series. Am J Sports Med 2008; 36: 360–368.
- Tahiri Y, Chang CS, Tuin J, et al. Costochondral grafting in craniofacial microsomia. Plast Reconstr Surg 2015; 135: 530–541.

- Cho BC, Kim JY and Byun JS. Two-stage reconstruction of the auricle in congenital microtia using autogenous costal cartilage. *J Plast Reconstr Aesthet Surg* 2007; 60: 998–1006.
- 32. Thomet C, Modarressi A, Ruegg EM, *et al.* Long-segment tracheal reconstruction with free radial forearm flap reinforced by rib cartilage. *Ann Plast Surg* 2018; 80: 525–528.
- Oue T, Kamata S, Usui N, *et al.* Histopathologic changes after tracheobronchial reconstruction with costal cartilage graft for congenital tracheal stenosis. *J Pediatr Surg* 2001; 36: 329–333.
- Park CH, Kim TH, Haam SJ, *et al.* The etiology of pectus carinatum involves overgrowth of costal cartilage and undergrowth of ribs. *J Pediatr Surg* 2014; 49: 1252–1258.
- Nakaoka T, Uemura S, Yoshida T, *et al.* Overgrowth of costal cartilage is not the etiology of pectus excavatum. *J Pediatr Surg* 2010; 45: 2015–2018.
- Tocchioni F, Ghionzoli M, Calosi L, et al. Rib cartilage characterization in patients affected by pectus excavatum. Anat Rec (Hoboken) 2013; 296: 1813–1820.
- West L, Govindraj P, Koob TJ, et al. Changes in perlecan during chondrocyte differentiation in the fetal bovine rib growth plate. *J Orthop Res* 2006; 24: 1317–1326.
- Kampen WU, Ciaassen H and Kirsch T. Mineralization and osteogenesis in the human first rib cartilage. *Ann Anat* 1995; 177: 171–177.
- Bahrami S, Plate U, Dreier R, *et al.* Endochondral ossification of costal cartilage is arrested after chondrocytes have reached hypertrophic stage of late differentiation. *Matrix Biol* 2001; 19: 707–715.
- Srour MK, Fogel JL, Yamaguchi KT, et al. Natural large-scale regeneration of rib cartilage in a mouse model. *J Bone Miner Res* 2015; 30: 297–308.
- 41. Stacey MW, Grubbs J, Asmar A, *et al.* Decorin expression, straw-like structure, and differentiation of human costal cartilage. *Connect Tissue Res* 2012; 53: 415–421.
- 42. Tay A, Farhadi J, Suetterlin R, *et al.* Cell yield, proliferation, and postexpansion differentiation capacity of human ear, nasal, and rib chondrocytes. *Tissue Eng* 2004; 10: 762–770.
- He A, Xia H, Xiao K, *et al.* Cell yield, chondrogenic potential, and regenerated cartilage type of chondrocytes derived from ear, nasoseptal, and costal cartilage. *J Tissue Eng Regen Med* 2018; 12: 1123–1132.

- 44. Lee J, Lee E, Kim HY, *et al.* Comparison of articular cartilage with costal cartilage in initial cell yield, degree of dedifferentiation during expansion and redifferentiation capacity. *Biotechnol Appl Biochem* 2007; 48: 149–158.
- 45. Kitaoka E, Satomura K, Hayashi E, et al. Establishment and characterization of chondrocyte cell lines from the costal cartilage of SV40 large T antigen transgenic mice. J Cell Biochem 2001; 81: 571–582.
- Willers C, Partsalis T and Zheng MH. Articular cartilage repair: procedures versus products. *Expert Rev Med Devices* 2007; 4: 373–392.
- 47. Berendsen AD and Olsen BR. Bone development. *Bone* 2015; 80: 14–18.
- Murphy MK, Huey DJ, Reimer AJ, et al. Enhancing post-expansion chondrogenic potential of costochondral cells in self-assembled neocartilage. PLoS One 2013; 8: e56983.
- 49. Koh S, Purser M, Wysk R, *et al.* Improved chondrogenic potential and proteomic phenotype of porcine chondrocytes grown in optimized culture conditions. *Cell Reprogram* 2017; 19: 232–244.
- Stacey MW, Sabuncu AC and Beskok A. Dielectric characterization of costal cartilage chondrocytes. *Biochim Biophys Acta* 2014; 1840: 146–152.
- 51. Mao Y, Block T, Singh-Varma A, et al. Extracellular matrix derived from chondrocytes promotes rapid expansion of human primary chondrocytes in vitro with reduced dedifferentiation. Acta Biomater 2019; 85: 75–83.
- Costa E, Gonzalez-Garcia C, Gomez Ribelles JL, et al. Maintenance of chondrocyte phenotype during expansion on PLLA microtopographies. *J Tissue Eng* 2018; 9: 2041731418789829.
- Robinson DL, Kersh ME, Walsh NC, et al. Mechanical properties of normal and osteoarthritic human articular cartilage. J Mech Behav Biomed Mater 2016; 61: 96–109.
- 54. De Croos JN, Dhaliwal SS, Grynpas MD, et al. Cyclic compressive mechanical stimulation induces sequential catabolic and anabolic gene changes in chondrocytes resulting in increased extracellular matrix accumulation. *Matrix Biol* 2006; 25: 323–331.
- 55. Carroll SF, Buckley CT and Kelly DJ. Cyclic tensile strain can play a role in directing both intramembranous and endochondral ossification of mesenchymal stem cells. *Front Bioeng Biotechnol* 2017; 5: 73.

- 56. Huwe LW, Sullan GK, Hu JC, *et al.* Using costal chondrocytes to engineer articular cartilage with applications of passive axial compression and bioactive stimuli. *Tissue Eng Part A* 2018; 24: 516–526.
- 57. Murphy M, DuRaine G, Reddi A, *et al.* Inducing articular cartilage phenotype in costochondral cells. *Arthritis Res Ther* 2013; 15: R214.
- Bougault C, Cueru L, Bariller J, *et al.* Alteration of cartilage mechanical properties in absence of β1 integrins revealed by rheometry and FRAP analyses. *J Biomech* 2013; 46: 1633–1640.
- Tessaro I, Nguyen V, Di Giancamillo A, et al. Animal models for cartilage repair. J Biol Regul Homeost Agents 2018; 32: 105–116.
- 60. Sato K, Moy OJ, Peimer CA, et al. An experimental study on costal osteochondral graft. Osteoarthritis Cartilage 2012; 20: 172–183.
- 61. Du D, Sugita N, Liu Z, *et al.* Repairing osteochondral defects of critical size using multiple costal grafts: an experimental study. *Cartilage* 2015; 6: 241–251.
- 62. Komura M, Komura H, Otani Y, *et al.* The junction between hyaline cartilage and engineered cartilage in rabbits. *Laryngoscope* 2013; 123: 1547–1551.
- 63. Mori R, Kataoka H, Kuriwaka M, *et al.* Articular cartilage restoration with costal cartilage previously fused with bone. *Clin Orthop Relat Res* 2003; 406: 262–274.
- 64. Howard R, McIlwraith C, Trotter G, et al. Long-term fate and effects of exercise on sternal cartilage autografts used for repair of large osteochondral defects in horses. Am J Vet Res 1994; 55: 1158–1167.
- 65. Rikimaru-Nishi Y, Rikimaru H, Hashiguchi S, *et al.* Histological study of costal cartilage after transplantation and reasons for avoidance of postoperative resorption and retention of cartilage structure in rats. *J Plast Surg Hand Surg* 2018; 52: 352–358.
- 66. Dominguez Perez JM, Fernandez-Sarmiento JA, Aguilar Garcia D, et al. Cartilage regeneration using a novel autologous growth factors-based matrix for full-thickness defects in sheep. Knee Surg Sports Traumatol Arthrosc. Epub ahead of print 21 August 2018. DOI: 10.1007/s00167-018-5107-z.
- 67. Yamasaki T, Yasunaga Y, Oshima S, et al. Healing potential of the cartilage correlates with location on the femoral head: a basic research using a rabbit model. *Hip Int* 2016; 26: 31–35.

- 68. Sun X, Yin H, Wang Y, *et al.* In situ articular cartilage regeneration through endogenous reparative cell homing using a functional bone marrow-specific scaffolding system. *ACS Appl Mater Interfaces* 2018; 10: 38715–38728.
- Badran KW, Waki C, Hamamoto A, et al. The rabbit costal cartilage reconstructive surgical model. *Facial Plast Surg* 2014; 30: 76–80.
- 70. El Sayed K, Haisch A, John T, et al. Heterotopic autologous chondrocyte transplantation-a realistic approach to support articular cartilage repair? *Tissue Eng Part B Rev* 2010; 16: 603–616.
- 71. Schüttler K, Götschenberg A, Klasan A, et al. Cell-free cartilage repair in large defects of the knee: increased failure rate 5 years after implantation of a collagen type I scaffold. Arch Orthop Trauma Surg 2019; 139: 99–106.
- Ellender P and Minas T. Autologous chondrocyte implantation in the hip: case report and technique. *Oper Tech Sports Med* 2008; 16: 201–206.
- 73. Akimau P, Bhosale A, Harrison PE, *et al.* Autologous chondrocyte implantation with bone grafting for osteochondral defect due to posttraumatic osteonecrosis of the hip–a case report. *Acta Orthop* 2006; 77: 333–336.
- 74. Sheu SY, Wang CH, Pao YH, et al. The effect of platelet-rich fibrin on autologous osteochondral transplantation: an in vivo porcine model. *Knee*, 2017; 24: 1392–1401.
- 75. Sengul AT, Buyukkkarabacak YB, Altunkaynak BZ, *et al.* Effects of platelet-rich plasma on cartilage regeneration after costal cartilage resection: a stereological and histopathological study. *Acta Chir Belg* 2017; 117: 21–28.
- 76. Sykes JG, Kuiper JH, Richardson JB, et al. Impact of human platelet lysate on the expansion and chondrogenic capacity of cultured human chondrocytes for cartilage cell therapy. *Eur Cell Mater* 2018; 35: 255–267.
- 77. Liu ZM, Shen PC, Lu CC, et al. Characterization of the proliferating layer chondrocytes of growth plate for cartilage regeneration. *Tissue Eng Part A*. Epub ahead of print 2 November 2018. DOI: 10.1089/ten. TEA.2018.0110.
- Foster B, Hansen A, Gibson G, et al. Reimplantation of growth plate chondrocytes into growth plate defects in sheep. *J Orthop Res* 1990; 8: 555–564.
- 79. Tomaszewski R, Bohosiewicz J, Gap A, *et al.* Autogenous cultured growth plate chondrocyte

transplantation in the treatment of physeal injury in rabbits. *Bone Joint Res* 2014; 3: 310–316.

- Hasegawa T and Yamano Y. Arthroplasty of the proximal interphalangeal joint using costal cartilage grafts. *J Hand Surg Br* 1992; 17: 583–585.
- Okuyama N, Sato K, Nakamura T, et al. Re: Total joint reconstruction for MP joint ankylosis using costal osteochondral graft: a case report. J Hand Surg Eur Vol 2009; 34: 132–133.
- Sandow M. Proximal scaphoid costoosteochondral replacement arthroplasty. J Hand Surg Br 1998; 23: 201–208.
- Sato K, Sasaki T, Nakamura T, *et al.* Clinical outcome and histologic findings of costal osteochondral grafts for cartilage defects in finger joints. *J Hand Surg Am* 2008; 33: 511–515.
- 84. Tropet Y, Gallinet D, Lepage D, et al. Treatment of trapeziometacarpal osteoarthritis by partial trapeziectomy and costal cartilage autograft. A review of 100 cases. *Chir Main* 2012; 31: 145–151.
- 85. Obert L, Lepage D, Sergent P, *et al.* Posttraumatic malunion of the distal radius treated with autologous costal cartilage graft: a technical note on seven cases. *Orthop Traumatol Surg Res* 2011; 97: 430–437.
- Obert L, Lepage D, Ferrier M, et al. Rib cartilage graft for posttraumatic or degenerative arthritis at wrist level: 10-year results. *J Wrist* Surg 2013; 2: 234–238.
- Sato K, Mio F, Hosoya T, *et al.* Two cases with osteochondritis dissecans of the capitulum humeri treated with costal osteochondral graft transplantation. *J Shoulder Elbow Surg* 2003; 12: 403–407.
- Shimada K, Tanaka H, Matsumoto T, et al. Cylindrical costal osteochondral autograft for reconstruction of large defects of the capitellum due to osteochondritis dissecans. J Bone Joint Surg Am 2012; 94: 992–1002.
- Sato K, Nakamura T, Toyama Y, et al. Costal osteochondral grafts for osteochondritis dissecans of the capitulum humeri. *Tech Hand Up Extrem Surg* 2008; 12: 85–91.
- 90. Iwai S, Sato K, Nakamura T, et al. Costoosteochondral graft for post-traumatic osteonecrosis of the radial head in an adolescent boy. J Bone Joint Surg Br 2011; 93: 111–114.
- Zhang C, Du D, Xu P, *et al.* Costal chondral transplantation to reconstruct cartilage of the femoral head: a case report. *Chin J Orthop* 2018; 38: 813–817.

- 92. Wallace CG, Mao HY, Wang CJ, et al. Threedimensional computed tomography reveals different donor-site deformities in adult and growing microtia patients despite total subperichondrial costal cartilage harvest and donor-site reconstruction. *Plast Reconstr Surg* 2014; 133: 640–651.
- Wee JH, Park MH, Oh S, et al. Complications associated with autologous rib cartilage use in rhinoplasty: a meta-analysis. *JAMA Facial Plast* Surg 2015; 17: 49–55.
- 94. Cerci Ozkan A, Bilgili AM and Guven E. Monobloc reconstruction of dome, medial crura, and columella with gamma-shaped costal cartilage graft. *Plast Reconstr Surg Glob Open* 2017; 5: e1629.
- 95. Liu L, Bu Z, Fan J, et al. Augmentation of the nasal dorsum using the multistrip autologous cartilage technique. *Plast Reconstr Surg* 2017; 140: 1163–1166.
- Nelson M and Gaball C. Technique to reduce time, pain, and risk in costal cartilage harvest. *JAMA Facial Plast Surg* 2017; 19: 333–334.
- Sun Z, Yu X, Chen W, et al. Costal cartilage assessment in surgical timing of microtia reconstruction. *J Craniofac Surg* 2017; 28: 1521–1525.
- Munro I, Phillips J and Griffin G. Growth after construction of the temporomandibular joint in children with hemifacial microsomia. *Cleft Palate J* 1989; 26: 303–311.
- 99. Teshima TL, Cheng H, Pakdel A, et al. Transverse slicing of the sixth-seventh costal cartilaginous junction: a novel technique to prevent warping in nasal surgery. J Craniofac Surg 2016; 27: e50–e55.
- Oki M and Asato H. Development of rib cartilage. Adv Otorhinolaryngol 2014; 75: 36–38.
- 101. Johnson JD, Desy NM and Sierra RJ. Ipsilateral femoral head osteochondral transfers for osteochondral defects of the femoral head. J Hip Preserv Surg 2017; 4: 231–239.
- 102. Girard J, Roumazeille T, Sakr M, et al. Osteochondral mosaicplasty of the femoral head. *Hip Int* 2011; 21: 542–548.
- 103. Tanaka Y, Nakamura S, Mukai S, et al. Domino osteochondral autograft transplantation for osteonecrosis of the knee and femoral head: a case based review. *J Orthop Sci*. Epub ahead of print 30 July 2018. DOI: 10.1016/j.jos.2018.06.017.
- 104. Filardo G, Andriolo L, Sessa A, *et al.* Age is not a contraindication for cartilage surgery: a critical

analysis of standardized outcomes at long-term follow-up. Am J Sports Med 2017; 45: 1822–1828.

- 105. Hochrein A, Zinser W, Spahn G, *et al.* What parameters affect knee function in patients with untreated cartilage defects: baseline data from the German Cartilage Registry. *Int Orthop.* Epub ahead of print 30 August 2018. DOI: 10.1007/ s00264-018-4125-2.
- 106. Tirico LEP, McCauley JC, Pulido PA, et al. Lesion size does not predict outcomes in fresh osteochondral allograft transplantation. Am J Sports Med 2018; 46: 900–907.
- 107. Yang HC, Cho HH, Jo SY, *et al.* Donorsite morbidity following minimally invasive costal cartilage harvest technique. *Clin Exp Otorhinolaryngol* 2015; 8: 13–19.
- 108. Foulad A, Hamamoto A, Manuel C, et al. Precise and rapid costal cartilage graft sectioning using a novel device: clinical application. JAMA Facial Plast Surg 2014; 16: 107–112.
- Ohara K, Nakamura K and Ohta E. Chest wall deformities and thoracic scoliosis after costal cartilage graft harvesting. *Plast Reconstr Surg* 1997; 99: 1030–1036.
- 110. Bozzato A, Bumm K, Hertel V, et al. Ultrasonographic evaluation of calcification patterns in costal cartilage: implications for rib graft harvesting. *JAMA Facial Plast Surg* 2013; 15: 457–460.
- 111. Zhang S, Zhen J, Li H, *et al.* Characteristics of Chinese costal cartilage and costa calcification using dual-energy computed tomography imaging. *Sci Rep* 2017; 7: 2923.
- 112. Ikeda T. Estimating age at death based on costal cartilage calcification. *Tohoku J Exp Med* 2017; 243: 237–246.
- 113. Lau AG, Kindig MW, Salzar RS, et al. Micromechanical modeling of calcifying human costal cartilage using the generalized method of cells. Acta Biomater 2015; 18: 226–235.
- 114. Sandoz B, Badina A, Laporte S, *et al.* Quantitative geometric analysis of rib, costal cartilage and sternum from childhood to teenagehood. *Med Biol Eng Comput* 2013; 51: 971–979.
- 115. Chung R, Cool JC, Scherer MA, *et al.* Roles of neutrophil-mediated inflammatory response in the bony repair of injured growth plate cartilage in young rats. *J Leukoc Biol* 2006; 80: 1272–1280.

- 116. Lepage S, Robson N, Gilmore H, et al. Beyond cartilage repair: the role of the osteochondral unit in joint health and disease. *Tissue Eng Part B Rev.* Epub ahead of print 14 January 2019. DOI: 10.1089/ten.TEB.2018.0122.
- 117. Heerey JJ, Kemp JL, Mosler AB, et al. What is the prevalence of imaging-defined intra-articular hip pathologies in people with and without pain? A systematic review and meta-analysis. Br J Sports Med 2018; 52: 581–593.
- 118. Kubo T, Utsunomiya H, Watanuki M, et al. Hip Arthroscopic osteochondral autologous transplantation for treating osteochondritis dissecans of the femoral head. Arthrosc Tech 2015; 4: e675–e680.
- 119. Nam Y, Rim YA, Lee J, *et al.* Current therapeutic strategies for stem cell-based cartilage regeneration. *Stem Cells Int* 2018; 2018: 8490489.
- 120. Chen MJ, Whiteley JP, Please CP, et al. Identifying chondrogenesis strategies for tissue engineering of articular cartilage. J Tissue Eng 2019; 10: 2041731419842431.
- 121. Stegen S, Laperre K, Eelen G, *et al.* HIF-1 α metabolically controls collagen synthesis and modification in chondrocytes. *Nature* 2019; 565: 511–515.
- 122. Kang D, Shin J, Cho Y, et al. Stress-activated miR-204 governs senescent phenotypes of chondrocytes to promote osteoarthritis development. Sci Transl Med 2019; 11. pii: eaar6659.
- 123. Islam A, Fossum V, Hansen AK, *et al.* In vitro chondrogenic potency of surplus chondrocytes from autologous transplantation procedures does not predict short-term clinical outcomes. *BMC Musculoskelet Disord* 2019; 20: 19.
- 124. Onofrillo C, Duchi S, O'Connell CD, *et al.* Biofabrication of human articular cartilage: a path towards the development of a clinical treatment. *Biofabrication* 2018; 10: 045006.
- 125. Deng Y, Lei G, Lin Z, *et al.* Engineering hyaline cartilage from mesenchymal stem cells with low hypertrophy potential via modulation of culture conditions and Wnt/beta-catenin pathway. *Biomaterials* 2019; 192: 569–578.
- 126. Kreuz PC, Gentili C, Samans B, *et al.* Scaffoldassisted cartilage tissue engineering using infant chondrocytes from human hip cartilage. *Osteoarthritis Cartilage* 2013; 21: 1997–2005.
- 127. Zhang J, Dong S, Sivak WN, *et al.* Stem cells in cartilage diseases and repair 2018. *Stem Cells Int* 2018; 2018: 3672890.

- 128. Daly AC, Freeman FE, Gonzalez-Fernandez T, et al. 3D bioprinting for cartilage and osteochondral tissue engineering. Adv Healthc Mater 2017; 6: 1700298.
- 129. Li Z, Jia S, Xiong Z, et al. 3D-printed scaffolds with calcified layer for osteochondral tissue engineering. J Biosci Bioeng 2018; 126: 389–396.
- Li L, Yu F, Zheng L, *et al.* Natural hydrogels for cartilage regeneration: modification, preparation and application. *J Orthop Translat* 2018; 17: 26–41.
- 131. Peng L, Zhou Y, Lu W, et al. Characterization of a novel polyvinyl alcohol/chitosan porous hydrogel combined with bone marrow mesenchymal stem cells and its application in articular cartilage repair. BMC Musculoskelet Disord 2019; 20: 257.
- 132. Cho SA, Cha SR, Park SM, et al. Effects of hesperidin loaded poly(lactic-co-glycolic acid) scaffolds on growth behavior of costal cartilage cells in vitro and in vivo. J Biomater Sci Polym Ed 2014; 25: 625–640.
- Cucchiarini M and Madry H. Biomaterialguided delivery of gene vectors for targeted articular cartilage repair. *Nat Rev Rheumatol* 2019; 15: 18–29.
- 134. Armiento AR, Stoddart MJ, Alini M, *et al.* Biomaterials for articular cartilage tissue engineering: learning from biology. *Acta Biomater* 2018; 65: 1–20.
- 135. Negoro T, Takagaki Y, Okura H, et al. Trends in clinical trials for articular cartilage repair by cell therapy. NPJ Regen Med 2018; 3: 17.
- Gurer B, Cabuk S, Karakus O, et al. In vivo cartilage tissue engineering. J Orthop Surg Res 2018; 13: 107.
- Deng ZH, Li YS, Gao X, et al. Bone morphogenetic proteins for articular cartilage regeneration. Osteoarthritis Cartilage 2018; 26: 1153–1161.
- 138. Lolli A, Colella F, De Bari C, *et al.* Targeting anti-chondrogenic factors for the stimulation of chondrogenesis: a new paradigm in cartilage repair. *J Orthop Res* 2019; 37: 12–22.
- 139. Seidl C, Fulga T and Murphy C. CRISPR-Cas9 targeting of MMP13 in human chondrocytes leads to significantly reduced levels of the metalloproteinase and enhanced type II collagen accumulation. *Osteoarthritis Cartilage* 2019; 27: 140–147.

Visit SAGE journals online

journals.sagepub.com/

SAGE journals

, home/tab

- 140. Mumme M, Barbero A, Miot S, *et al.* Nasal chondrocyte-based engineered autologous cartilage tissue for repair of articular cartilage defects: an observational first-in-human trial. *Lancet* 2016; 388: 1985–1994.
- 141. Fulco I, Miot S, Haug MD, et al. Engineered autologous cartilage tissue for nasal reconstruction

after tumour resection: an observational first-inhuman trial. *Lancet* 2014; 384: 337–346.

142. Candrian C, Vonwil D, Barbero A, *et al.* Engineered cartilage generated by nasal chondrocytes is responsive to physical forces resembling joint loading. *Arthritis Rheum* 2008; 58: 197–208.