

# INTERNATIONAL UNION OF BASIC AND CLINICAL PHARMACOLOGY REVIEW

## Endothelin@25 – new agonists, antagonists, inhibitors and emerging research frontiers: IUPHAR Review 12

J J Maguire and A P Davenport

*Clinical Pharmacology Unit, University of Cambridge, Addenbrooke's Hospital, Cambridge, UK*

Since the discovery of endothelin (ET)-1 in 1988, the main components of the signalling pathway have become established, comprising three structurally similar endogenous 21-amino acid peptides, ET-1, ET-2 and ET-3, that activate two GPCRs, ET<sub>A</sub> and ET<sub>B</sub>. Our aim in this review is to highlight the recent progress in ET research. The ET-like domain peptide, corresponding to prepro-ET-1<sub>93-166</sub>, has been proposed to be co-synthesized and released with ET-1, to modulate the actions of the peptide. ET-1 remains the most potent vasoconstrictor in the human cardiovascular system with a particularly long-lasting action. To date, the major therapeutic strategy to block the unwanted actions of ET in disease, principally in pulmonary arterial hypertension, has been to use antagonists that are selective for the ET<sub>A</sub> receptor (ambrisentan) or that block both receptor subtypes (bosentan). Macitentan represents the next generation of antagonists, being more potent than bosentan, with longer receptor occupancy and it is converted to an active metabolite; properties contributing to greater pharmacodynamic and pharmacokinetic efficacy. A second strategy is now being more widely tested in clinical trials and uses combined inhibitors of ET-converting enzyme and neutral endopeptidase such as SLV306 (daglutril). A third strategy based on activating the ET<sub>B</sub> receptor, has led to the renaissance of the modified peptide agonist IRL1620 as a clinical candidate in delivering anti-tumour drugs and as a pharmacological tool to investigate experimental pathophysiological conditions. Finally, we discuss biased signalling, epigenetic regulation and targeting with monoclonal antibodies as prospective new areas for ET research.

### Correspondence

Anthony P Davenport, Clinical Pharmacology Unit, University of Cambridge, Addenbrooke's Hospital, Cambridge, CB2 0QQ, UK. E-mail: apd10@medschl.cam.ac.uk

### Received

28 March 2014

### Revised

22 July 2014

### Accepted

25 July 2014

This is the 12th in a series of reviews written by committees of experts of the Nomenclature Committee of the International Union of Basic and Clinical Pharmacology (NC-IUPHAR). A listing of all articles in the series and the Nomenclature Reports from IUPHAR published in Pharmacological Reviews can be found at <http://www.GuideToPharmacology.org>. This website, created in a collaboration between the British Pharmacological Society (BPS) and the International Union of Basic and Clinical Pharmacology (IUPHAR), is intended to become a 'one-stop shop' source of quantitative information on drug targets and the prescription medicines and experimental drugs that act on them. We hope that the Guide to Pharmacology will be useful for researchers and students in pharmacology and drug discovery and provide the general public with accurate information on the basic science underlying drug action.

*How to cite:* Maguire JJ and Davenport AP. (2014). Endothelin@25 – new agonists, antagonists, inhibitors and emerging research frontiers: IUPHAR Review 12. *Br J Pharmacol* 171: 5555–5572.

## Abbreviations

ECE-1, endothelin-converting enzyme-1; ECE-2, endothelin-converting enzyme-2; ELDP, endothelin-like domain peptide; ET, endothelin; HIF-1 $\alpha$ , hypoxia inducible factor-1 $\alpha$ ; NEP, neutral endopeptidase; PAH, pulmonary arterial hypertension

## Tables of Links

TARGETS
<b>GPCRs<sup>a</sup></b>
AT <sub>1</sub> receptors
ET <sub>A</sub> receptors
ET <sub>B</sub> receptors
GPR37
GPR37L
Hydroxycarboxylic acid receptors
$\mu$ opioid receptors
Parathyroid hormone receptors
<b>Enzymes<sup>b</sup></b>
Cathepsin A
Chymase
CYP3A4
CYP2C19
Endothelin-converting enzyme 1
Endothelin-converting enzyme 2
Neutral endopeptidase

LIGANDS	
5-Fluorouracil	Endothelin-3
Ambrisentan	IRL1620
Amyloid $\beta$ -peptide	KC-12615
Atrial natriuretic peptide (ANP)	Losartan
$\beta$ -Catenin	Macitentan
Bosentan	NO
BQ123	Prosaptide
BQ788	PGI <sub>2</sub>
BQ3020	Sarafotoxin S6b
Captopril	Sarafotoxin S6c
Dagliutril	Sitaxentan
Docetaxel	TGF $\beta$ 1
Doxorubicin	
Endothelin-1	
Endothelin-2	

These Tables list protein targets and ligands that are hyperlinked to corresponding entries in <http://www.guidetopharmacology.org>, the common portal for data from the IUPHAR/BPS Guide to PHARMACOLOGY (Pawson *et al.*, 2014) and the Concise Guide to PHARMACOLOGY 2013/14 (<sup>a,b</sup>Alexander *et al.*, 2013a,b).

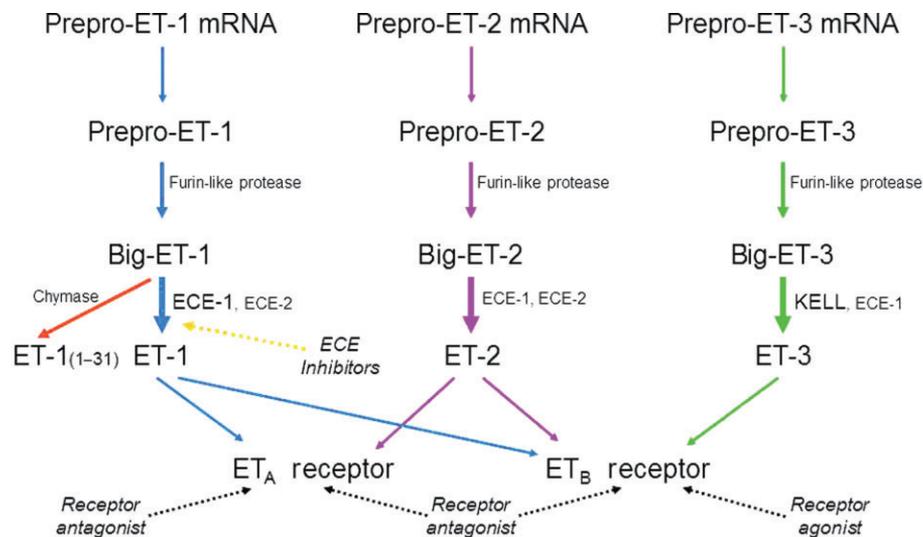
This article, written by members of the International Union of Basic and Clinical Pharmacology Committee on Receptor Nomenclature and Drug Classification (NC-IUPHAR) subcommittee for the endothelin receptors, confirms the existing nomenclature for these receptors and reviews our current understanding of their structure, pharmacology and functions, and their likely physiological roles in health and disease. More information on these receptor families can be found in the Concise Guide to PHARMACOLOGY (<http://onlinelibrary.wiley.com/doi/10.1111/bph.12445/abstract>) and for each member of the family in the corresponding database <http://www.guidetopharmacology.org/GRAC/FamilyDisplayForward?familyId=21&familyType=GPCR>.

## Introduction

Since the discovery of endothelin (ET)-1 in 1988 (Yanagisawa *et al.*, 1988; Inoue *et al.*, 1989) the components of the ET signalling pathway have become established, comprising three structurally similar endogenous 21-amino acid peptides, ET-1, ET-2 and ET-3, that activate two GPCRs, ET<sub>A</sub> (Arai *et al.*, 1990) and ET<sub>B</sub> (Sakurai *et al.*, 1990). In humans, ET-2 differs from ET-1 by only two amino acids, whereas ET-3 differs by six amino acids representing more substantial changes. ET-3 is the only isoform that can distinguish between the two receptor subtypes, having a similar potency at the ET<sub>A</sub> receptor as ET-1 and ET-2, but much lower affinity than these isoforms for the ET<sub>B</sub> receptor (Figure 1). Structur-

ally, ETs are unusual among the mammalian peptides in possessing two disulphide bridges. This feature is shared by the sarafotoxins, a family of peptides that were isolated from snake venom in the same year as the discovery of ET-1 (Takasaki *et al.*, 1988), and that provided the first selective agonist at the ET<sub>B</sub> receptor, sarafotoxin S6C (William *et al.*, 1991).

A number of features of the ET signalling pathway are unusual compared with other peptidergic systems and these continue to intrigue investigators, with over a thousand ET-related papers still published each year. ET-1 is the most abundant isoform in the human cardiovascular system, predominantly released from endothelial cells to cause potent and unusually long-lasting vasoconstriction that may persist for many hours. ET-1 is a key mediator in regulating vascular



**Figure 1**

Scheme of the biosynthesis of ET peptides and their interaction with receptors. Based on information from the literature including Barton and Yanagisawa (2008), Turner and Tanzawa (1997) and Lee *et al.* (1999).

function in the majority of organs systems balanced by opposing vasodilators, particularly NO, prostacyclin and endothelium-derived hyperpolarizing factor. Endothelial cell dysfunction occurs in pathophysiological conditions such as pulmonary arterial hypertension (PAH) and is associated with loss of these dilators and increased synthesis of ET. The consequence of this is vasoconstriction, proliferation of many different cell types particularly vascular smooth muscle, fibrosis and inflammation; processes associated with vascular remodelling. In disease, the deleterious actions of ET in the vasculature are mainly mediated by the  $ET_A$  receptor, whereas activation of  $ET_B$  receptors results in many of the beneficial effects of the peptide that frequently act as a regulatory counterbalance (Davenport and Maguire, 2006). The formation of the disulphide bridge in the ET peptides blocks the N-terminal amino acid conferring resistance to enzymic degradation in plasma. Internalization by  $ET_B$  scavenging receptors is therefore particularly important for termination of the ET signal in health and disease.

The major therapeutic strategy (Figure 1) to block the unwanted actions of ET in disease has been to use antagonists of  $ET_A$  receptors or both receptor subtypes (Palmer, 2009) with the first clinical application being bosentan in PAH (Rubin *et al.*, 2002). More recently, a second strategy has started to be more widely tested in clinical trials using inhibitors of ET-converting enzymes 1 (ECE-1; Xu *et al.*, 1994) and 2 (ECE-2; Emoto and Yanagisawa, 1995), the major biosynthetic pathway of ET (Figure 1) at least in the human vasculature (Russell and Davenport, 1999a,b). A third emerging strategy based on biosimilar agonists at the  $ET_B$  receptor (molecules similar, but not identical to the endogenous ligand) has led to the renaissance of IRL1620 as a clinical candidate in delivering anti-tumour drugs and in other pathophysiological conditions such as cerebral ischaemia.

The aim of this focused IUPHAR review is to highlight recent progress and some surprising new discoveries in the pharmacology of the ET system. The following references are

recommended for more detailed information on specific ET research areas including the heart (Kelland and Webb, 2006; Kirkby *et al.*, 2008; Kohan *et al.*, 2012; Ohkita *et al.*, 2012; Vignon-Zellweger *et al.*, 2012; Drawnel *et al.*, 2013), renal (Dhaun *et al.*, 2011; 2012; Kohan *et al.*, 2011a,b; Hyndman and Pollock, 2013), hypertension (Rautureau and Schiffrin, 2012; Speed and Pollock, 2013), PAH (Pernow *et al.*, 2012; Rubin, 2012; Liu *et al.*, 2013), cancer (Bagnato *et al.*, 2011; Said and Theodorescu, 2012; Rosanò *et al.*, 2013b), atherosclerosis and diabetes (Pernow *et al.*, 2012).

## ET peptides

### *Evidence for a new ET peptide: the ET-like domain peptide (ELDP)*

The ELDP has been recently identified as a peptide corresponding to prepro-ET-1<sub>93-166</sub> (Yuzgulen *et al.*, 2013) immediately adjacent to the gene sequence encoding big ET-1. The 74-amino acid peptide has been detected by HPLC and specific double recognition site immunoassays in conditioned media from two cell lines, endothelial (EA.hy 926) and epithelial (A549), as well as from primary cell cultures of human aortic endothelial cells that are known to secrete ET-1. In the aortic endothelial cells, the peptide was co-synthesized and co-released with ET-1. Plasma levels in untreated patients were 6.5 pmol·L<sup>-1</sup>, which compares with typical basal levels of immunoreactive ET-1 of 5 pmol·L<sup>-1</sup> (Davenport *et al.*, 1990). Levels of ELDP were significantly elevated in patients with heart failure suggesting a potential use as a bio-marker. While no effect was observed on BP in the anaesthetized rat, intriguingly ELDP significantly increased the duration of the pressor response of ET-1 (0.3 nmol·kg<sup>-1</sup>, likely to be a submaximal dose). Pretreatment of rat mesenteric arteries with 10 nM ELDP also potentiated a submaximal response of ET-1 by fivefold (Yuzgulen *et al.*, 2013). It is not unexpected that a

second peptide sharing a cleavage site with ET-1 would also be co-released, but it is intriguing that the peptide was able to potentiate ET-1 responses *in vitro* and *in vivo*. It is not yet reported, using saturation or competition-binding experiments, whether ELDP binds directly to ET receptors, binds to an allosteric site or whether the peptide modulates ET responses by other mechanisms. Intriguingly, ELDP encompasses the sequence of the putative 'endothelin-like peptide' corresponding to prepro-ET-1<sub>109-123</sub> proposed in the original *Nature* paper by Yanagisawa *et al.* (1988). Eight out of 15 residues in the corresponding sequence in ET-1 are identical and the four Cys residues are perfectly conserved and flanked by dibasic pairs that are recognized by endopeptidase processing enzymes, to yield a cleaved peptide. However, a synthetic peptide corresponding to this sequence was devoid of agonist or antagonist activity against ET-1, in vascular preparations (Cade *et al.*, 1990).

### *Global knockout of ET-2 gene reveals distinct phenotype compared with ET-1/ET<sub>A</sub> and ET-3/ET<sub>B</sub> gene deletions*

ET-1-deficient homozygous mice die at birth of respiratory failure, which is secondary to severe craniofacial and cardiovascular abnormalities. ET<sub>A</sub> receptor and ECE-1 knockout mice have similar morphological abnormalities (Kurihara *et al.*, 1994; Clouthier *et al.*, 1998; Yanagisawa *et al.*, 1998). The phenotype is similar to a spectrum of human conditions, CATCH 22 (cardiac anomaly, abnormal face, thymic hypoplasia, cleft palate, hypocalcaemia, chromosome 22 deletions) and established the importance of the ET<sub>A</sub>/ET-1 signalling system for cardiovascular and craniofacial development. Gene deletions for ET-3 and ET<sub>B</sub> receptors exhibit a different and non-overlapping phenotype to ET-1/ET<sub>A</sub>-deficient animals. They are viable at birth and survive for up to 8 weeks, but display aganglionic megacolon, as a result of absence of ganglion neurones, together with a disorder of the pigment in their coats (Baynash *et al.*, 1994; Hosoda *et al.*, 1994). In these mice, enteric nervous system precursors and neural crest-derived epidermal melanoblasts fail to colonize the intestine and skin. This phenotype resembles Hirschsprung's disease in man.

Deleting genes encoding all the key molecules, ET-1, ET-3, ET<sub>A</sub>, ET<sub>B</sub>, ECE-1 and ECE-2 has been accomplished in mice generating important information about their effect on phenotype. The deletion of the gene for ET-2 has now been reported. The physiological role of ET-2 has been puzzling. It had been assumed that the actions of ET-2 would be similar, if released, to the more widely distributed and abundant ET-1. Current antagonists block both ET-2 and ET-1 with the same potency and are not yet able to distinguish the actions of these peptides.

A key advance in the field was the generation by Chang *et al.* (2013) of a global ET-2 gene knockout mouse, which surprisingly exhibited a distinct phenotype to global ET-1 or ET-3 gene deletions. These mice showed severe growth retardation, internal starvation characterized by hypoglycaemia, ketonaemia and increased levels of starvation-induced genes. Mice were profoundly hypothermic and the median lifespan could be significantly extended by housing in a warm environment. The intestine was morphologically and functionally normal, which was unexpected as murine ET-2 (see Ling

*et al.*, 2013), also known as vasoactive intestinal contractor, is present throughout the gastrointestinal tract suggesting, in this tissue at least, in the absence of ET-2, ET-1 continues to mediate signalling. In agreement, intestinal epithelium-specific ET-2 knockout mice showed no abnormalities in growth and survival. In marked contrast, dramatic changes were observed in lung morphology and function. Mice had breathing difficulties after the first week exhibiting enlarged air spaces with substantial simplification of lung alveolar structure, larger lung capacities leading to abnormally elevated carbon dioxide (hypercapnia) and deficiency of oxygen (hypoxaemia) in the blood. Hypothermia and lung dysfunction might not be specific, but may be due to a secondary effect of internal starvation because of ET-2 deficiency. However, it is possible that these studies identify an important function for ET-2 in the pulmonary system. The authors showed that mRNA encoding ET-2 was only present in epithelial cells whereas receptor mRNA was mainly present in mesenchyme, consistent with a paracrine function for ET-2 in the lung.

### *Pharmacological significance: is ET-2 the inducible isoform?*

The dramatic effects on the lung suggest a crucial role for ET-2 at birth, at least in mice. The lungs, and potentially the heart, remain major therapeutic targets for ET antagonists in humans in the treatment of PAH. In rodents, ET-2 was less widely distributed than ET-1, mainly found in heart, lung, ovary, stomach and all regions of the intestine (de la Monte *et al.*, 1995; Takizawa *et al.*, 2005). ET-2 expression in human tissue was similar, being present in the human heart (Plumpton *et al.*, 1996b), lung (Marciniak *et al.*, 1992), kidney (Karet and Davenport, 1996), vasculature, (Howard *et al.*, 1992) intestine and ovaries (Palanisamy *et al.*, 2006), but has not been investigated in pathophysiological tissue. In humans, alternatively spliced mRNA variants encoding ET-2 have been detected with a specific pattern of distribution in various tissues. Some of these variants contain sites for the post-transcriptional processing of preproET-2 into mature ET-2, which may be altered in a tissue-specific manner (O'Reilly *et al.*, 1993). The best established model of spatial and temporal ET-2 signalling is in the ovary, a highly vascular tissue, which undergoes cyclic changes as follicles grow, rupture and transform into corpora lutea and eggs are periodically released (Ko *et al.*, 2012). In rats, low levels of ET-1 are constitutively expressed throughout the ovulatory cycle, whereas ET-2 is induced transiently at much higher concentrations during the period of ovulation to luteal phases (Ko *et al.*, 2006). ET-2 is expressed in the granulosa cells of periovulatory follicles, but not during other stages of follicular development. In mice, induced superovulation results in a dramatic increase in ET-2 mRNA expression (Palanisamy *et al.*, 2006). ET-2 expression surged in response to gonadotropin and quickly declined by 13 h, which coincided with the time of follicular rupture. Crucially, both ET receptor subtypes are present and their ratio does not seem to change. Thus, the ET-2 gene appears to be switched on only when increased levels of ET are required, with ET-2-mediated contraction being the final signal facilitating ovulation (Ling *et al.*, 2013).

ET-1 signalling is well established in neural crest migration. In the developing mouse retina, constitutive over-expression of ET-2 affects vascular development by inhibiting endothelial cell migration across the retinal surface and subsequent endothelial cell invasion into the retina, an action mediated by ET<sub>A</sub> receptors. Interestingly, over-expression is spatially localized as it has no obvious action on vascular structures in brain or skin (Rattner *et al.*, 2013). Constitutive over-expression of ET-2 signalling also protected photoreceptors from light damage (Braunger *et al.*, 2013). Similarly, Bramall *et al.* (2013) found expression of ET-2 mRNA was greatly increased in the photoreceptors of mouse models of inherited photoreceptor degeneration and, using the global ET-2 knockout mice, showed increased ET-2 expression was protective of the mutant photoreceptors.

### Case for re-evaluating the role of ET-2

Targeting the ET-2 gene in mice provides compelling evidence that, while both ET-1 and ET-2 can coexist in the same tissue compartments, there is a critical, but distinct ET-2 pathway. A key role has now been established for ET-2 in ovarian physiology. This may be accomplished at the level of gene expression, but differences may also exist in peptide synthesis by ECEs and chymase, which may allow the two ET peptide pathways to be distinguished pharmacologically and become separate drug targets. Additionally, pharmacological differences have been identified, for example ET-2 dissociates from receptors much more rapidly than ET-1 and higher affinity has been reported, for example in the brain (Ling *et al.*, 2013). Detailed studies comparing rat mesenteric resistance and basilar arteries demonstrated that ET-1 and ET-2 initiate and maintain vasoconstriction by different downstream mechanisms raising the prospect of 'biased signalling' mediated by two structurally different agonists activating the same receptor (Compeer *et al.*, 2013).

### Potential new therapeutic strategies exploiting ET<sub>B</sub> receptor agonists

The pharmacological rationale for this strategy is that ET-1, tonically released from endothelial cells, also interacts with endothelial cell ET<sub>B</sub> receptors. The importance of this counter-regulatory pathway has been underestimated to date. Endothelial cells line the vasculature of every organ and tissue in the body that receives blood supply. Although the cells represent ~1% of the weight of the vessel wall, they have a combined mass comparable with some endocrine glands. Crucially, ET-1 feeding back onto endothelial receptors to release NO not only limits ET<sub>A</sub>-mediated vasoconstriction by stimulation of vascular cGMP, but also limits further ET-1 release. Thus in the vasculature, NO and other dilators are crucial in balancing the ET system, but these may be reduced or absent in pathophysiological conditions.

ET-1<sup>+/-</sup> heterozygous mice developed *elevated* BP and mild hypertension, rather than the fall in BP that might have been expected. Partial deletion of the gene allows survival and produced lower levels of ET-1 in plasma and lung tissue than wild type (Kurihara *et al.*, 1994). These results suggest that ET-1 has an essential physiological role in cardiovascular homeostasis. Low levels promote vasodilatation whereas

higher and pathophysiological concentrations of ET-1 increase BP and total peripheral vascular resistance. While ET<sub>A</sub> receptor selective antagonists such as BQ123 (Ihara *et al.*, 1992) cause the expected vasodilatation in humans (Haynes and Webb, 1994), the ET<sub>B</sub> receptor selective antagonist BQ788 (Ishikawa *et al.*, 1994) caused systemic *vasoconstriction* in healthy volunteers, showing that the main consequence of activation of endothelial ET<sub>B</sub> receptors by tonically secreted ET-1 was the physiological basal release of NO (Love *et al.*, 2000). In agreement, initial vasodilatation can be detected in the human forearm vascular bed following infusion of low concentrations of ET-1 whereas higher doses caused sustained vasoconstriction (Kiowski *et al.*, 1991). A contribution to vasoconstriction may also be the result of occupancy by ET-1 of the clearance ET<sub>B</sub> receptors causing an ET<sub>A</sub>-mediated vasoconstriction.

### ET<sub>B</sub> agonists in chemotherapy: IRL1620

ET-1 acting on ET<sub>A</sub> receptors has been proposed to stimulate cell proliferation, migration, invasion, osteogenesis and angiogenesis in several cancers. New vessels forming in tumours are characterized by high densities of ET<sub>A</sub> receptors in smooth muscle, for example in glioblastoma multiforme in the brain (Harland *et al.*, 1998). Conversely, ET<sub>B</sub> receptors may oppose tumour progression by promoting apoptosis and clearing ET-1 (Bagnato *et al.*, 2011; Rosanò *et al.*, 2013b). The strategy of stimulating ET<sub>B</sub> receptors to cause transient vasodilatation is being developed to increase the penetration of cytotoxic anti-tumour agents into tumours and to minimize the concentration in healthy tissue.

IRL1620 was originally developed as a tool compound (Takai *et al.*, 1992). The N-terminus has an N-succinyl modification, which is likely to reduce metabolism by non-specific peptidases, but it is not orally active and requires injection. Despite these unpromising pharmacokinetic features, it is being used *in vivo* and has emerged as a possible clinical candidate in improving the delivery of drugs to tumours. IRL1620 infused into rats improved the efficacy of doxorubicin and 5-fluorouracil by significantly increasing the amount of drug in tumours in rat models of prostate and breast cancer. In addition, radiation-induced reduction in tumour volume was enhanced, suggesting IRL1620 can significantly increase the efficacy of radiotherapy in the treatment of solid tumours. The results suggest that for a given dose of drug, the efficacy in reducing the tumour could be improved (Gulati and Rai, 2004; Rajeshkumar *et al.*, 2005a,b; Lenaz *et al.*, 2006; Rai *et al.*, 2006; Gulati *et al.*, 2012). A phase I trial to determine the safety, tolerability, pharmacokinetics and pharmacodynamics of IRL1620 (known as SPI-1620 licensed by Spectrum Pharmaceuticals, Technology Drive Irvine, CA, USA) in patients with recurrent progressive carcinoma has been successfully completed and shown to selectively and transiently increase tumour blood flow (Gulati *et al.*, 2012; <http://www.cancer.gov/clinicaltrials>). A phase II trial was initiated in 2013 to determine the effectiveness of SPI-1620 in combination with docetaxel in patients with advanced biliary cancer (<http://clinicaltrials.gov/ct2/show/NCT01773785>) and in combination with docetaxel compared with docetaxel alone for patients with non small-cell lung cancer after failure of platinum-based chemotherapy (<http://clinicaltrials.gov/show/NCT01741155>).

### *ET<sub>B</sub> agonists in neuroprotection*

The human brain contains the highest density of ET receptors, with the ET<sub>B</sub> receptor subtype comprising about 90%, in areas such as cerebral cortex (Harland *et al.*, 1998). Binding and functional studies have demonstrated glia mainly express ET<sub>B</sub> receptors whereas ET<sub>A</sub> receptors are localized mainly on neurones (Morton and Davenport, 1992). Smooth muscle cells from large arteries and small intracerebral vessels only express ET<sub>A</sub> receptors (Adner *et al.*, 1994; Harland *et al.*, 1995; Pierre and Davenport, 1999) with endothelial cell ET<sub>B</sub> receptors mediating relaxation (Lucas *et al.*, 1996). The small pial arteries and arterioles penetrating into the brain play a major role in the maintenance of cerebral blood flow (autoregulation). These vessels are particularly sensitive to ET-1 compared with peripheral vessels and the peptide has been a long-standing candidate in the genesis or maintenance of cerebrovascular disorders such as delayed vasospasm associated with subarachnoid haemorrhage or stroke. ET-1 does not cross the blood–brain barrier from the plasma, but may do so when compromised by subarachnoid haemorrhage, stroke or head injury. Strategies for targeting cerebrovascular disease have focused previously on the use of ET receptor antagonists, firstly to block vascular receptors mediating cerebrovasospasm that may be responsible for delayed cerebral ischaemia seen after aneurysmal subarachnoid haemorrhage and could contribute to ischaemic core volume in stroke. Secondly, to block neural receptors that mediate increases in intracellular free calcium (Morton and Davenport, 1992) and initiate the pathophysiological processes leading to neuronal death.

A new emerging strategy is to use ET<sub>B</sub> receptor agonists such as IRL1620 to provide vasodilatation and neuroprotection. The peptide reduced neurological damage following permanent middle cerebral artery occlusion in rats, a model of focal ischaemic stroke. Animals received i.v. injections of IRL1620 after the occlusion, which dramatically reduced infarct volume (by more than 80% in the acute and 70% in the chronic study), prevented cerebral oedema, reduced oxidative stress markers and improved all neurological and motor function for up to 7 days (Leonard *et al.*, 2011; 2012; Leonard and Gulati, 2013). Rats treated with the amyloid peptide A $\beta$ <sub>1–40</sub> administered into the intracerebral vessels display increased markers of oxidative stress in the brain. IRL1620 significantly reduced oxidative stress and importantly the cognitive impairment (Briyal *et al.*, 2014). As discussed later, a reduction in ECE activity is associated with accumulation of amyloid  $\beta$ -peptide and neurotoxicity early in progression of Alzheimer's disease (Eckman *et al.*, 2001; 2003; 2006; Pacheco-Quinto and Eckman, 2013). These results are limited to disease models in a single species, and it is unclear whether the molecular mechanisms would translate to humans, but taken together, they suggest that an ET<sub>B</sub> receptor agonist might offer a new therapeutic strategy in Alzheimer's disease and provide neuroprotection following cerebral ischaemia in conditions such as stroke.

### *No evidence for further ET<sub>B</sub> receptor subtypes*

Previous studies have suggested that ET<sub>B</sub> receptors could be further subdivided into ET<sub>B1</sub> present on endothelial cells and ET<sub>B2</sub> on smooth muscle cells. Studies continue to be published with this misleading nomenclature, but current evi-

dence only supports the existence of two subtypes, ET<sub>A</sub> and ET<sub>B</sub>, according to NC-IUPHAR nomenclature (Davenport, 2002; Alexander *et al.*, 2013a,b). Firstly, Mizuguchi *et al.* (1997) demonstrated unequivocally that in ET<sub>B</sub> receptor knockout mice, both the direct constrictor and indirect vasodilator responses to the ET<sub>B</sub> agonist sarafotoxin S6C were abolished. Selective deletion of endothelial ET<sub>B</sub> receptors in mice (demonstrated by autoradiography to leave unaltered ET<sub>B</sub> receptors expressed by other cell types) impaired, as expected, the clearance of an i.v. bolus of labelled ET-1 compared with controls (Bagnall *et al.*, 2006; Kelland *et al.*, 2010). Secondly, Flynn *et al.* (1998) were unable to distinguish pharmacologically, in extensive competition-binding experiments, between ET<sub>B</sub> receptors expressed by human isolated endothelial and smooth muscle cells in culture. In concordance, saturation-binding assays in human tissue always found ET<sub>B</sub> radiolabelled ligands bound with a single affinity and Hill slopes close to unity with no suggestion of further subtypes (Molenaar *et al.*, 1992; 1993; Nambi *et al.*, 1994) or in competition-binding versus radiolabelled ET-1 in human native (Peter and Davenport, 1995; 1996; Russell and Davenport, 1996) or recombinant ET<sub>B</sub> receptors (Nambi *et al.*, 1994; Reynolds *et al.*, 1995). Clozel and Gray (1995) showed that endothelial and smooth muscle ET<sub>B</sub> receptors cannot be distinguished functionally.

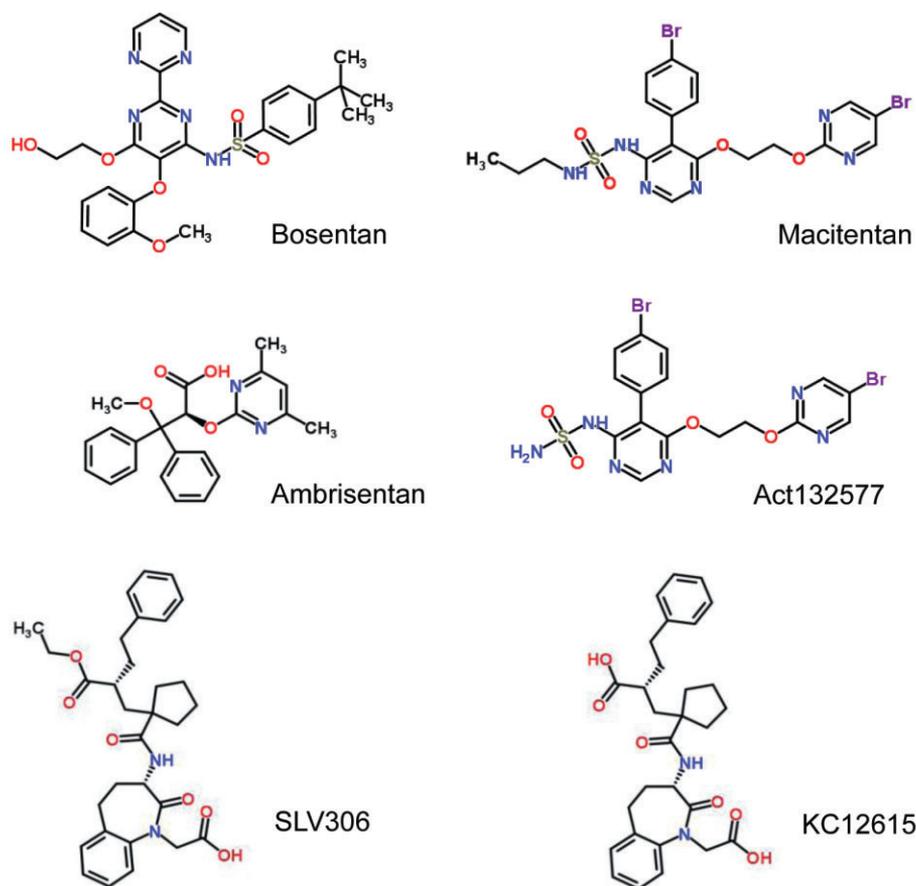
### *Do ETs interact with any other GPCRs?*

The virtually complete sequencing of the human genome has allowed the identification of all of the human gene sequences that could potentially encode GPCRs that are currently classified as 'orphan' to indicate that their endogenous ligand is not yet known (Foord *et al.*, 2005; Davenport *et al.*, 2013). In this catalogue, the most closely related to the ET<sub>A</sub> and ET<sub>B</sub> receptor subtypes are the orphan receptors GPR37 (also known as ET receptor type B-like or Parkin-associated ET receptor-like receptor) and its related receptor, GPR37L1. A recent high-throughput screen tested ~10 000 biologically active compounds for binding to 82 remaining orphan GPCRs. None of the ~20 ET peptides tested at high concentration (including all three mature isoform and their corresponding big ET precursors, C-terminal metabolites, BQ123 and the ET<sub>B</sub> receptor agonist BQ3020) activated any of the expressed receptors, including GPR37 or GPR37L, supporting the established concept of ETs binding to only two receptor subtypes. Two orphan neuropeptides, prosaptide and prosaposin, have recently been proposed as cognate ligands for GPR37 and GPR37L (Meyer *et al.*, 2013).

## Clinical application of ET antagonists

### *Bosentan, ambrisentan and withdrawal of sitaxentan*

PAH is a progressive condition with no cure and has a major impact on the ability to lead a normal life. It is an orphan disease (~100 000 patients in US and Europe). PAH involves constriction of pulmonary arteries and is characterized by high BP in the lungs, ultimately leading to right heart failure and death. A number of pathways have been implicated in the development of PAH including bone morphogenetic



**Figure 2**

Structures of ET receptor antagonists in clinical use bosentan, ambrisentan and macitentan. The structures of the NEP/ECE inhibitor pro-drug SLV306 and its active metabolite are also shown.

proteins, prostacyclin and ET-1. Restoring the imbalance between constriction and vasodilatation of blood vessels is the basis for current medical therapies, although the cause of death is right heart failure. Although ET<sub>A</sub> receptors are significantly increased in the right ventricle of patients with PAH (Kuc *et al.*, 2014) and in the left ventricle of patients with heart failure (Zolk *et al.*, 1999), surprisingly, ET receptor antagonists have clinical efficacy in the former, but not the latter group (Kohan *et al.*, 2012).

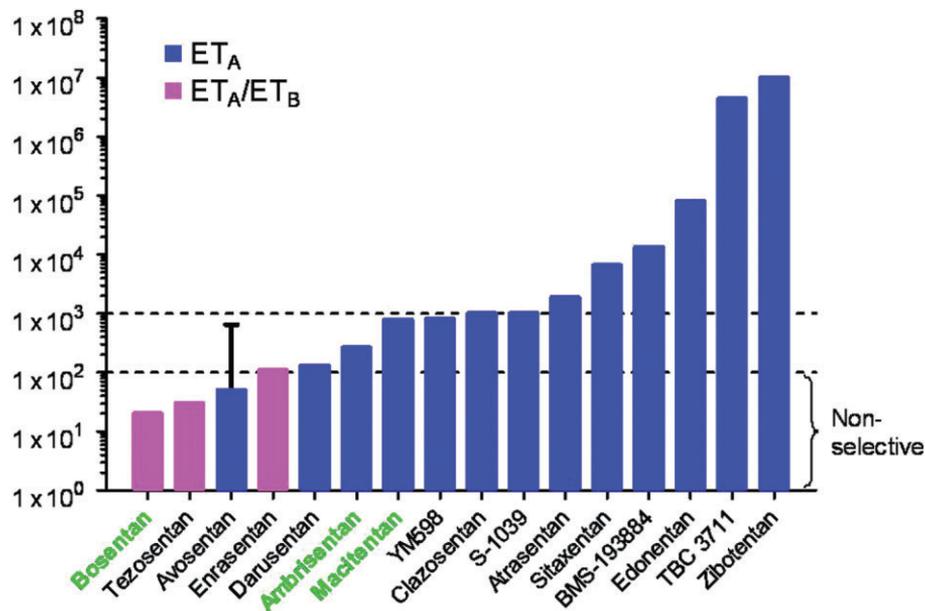
Bosentan (Tracleer, Ro47-0203) was the first ET receptor antagonist to be introduced into the clinic for the treatment of PAH (Rubin *et al.*, 2002) and, as an orally acting agent, at the time represented a major advance over existing therapies such as prostacyclin analogues. Bosentan is classified as a mixed ET<sub>A</sub>/ET<sub>B</sub> receptor antagonist blocking both receptors (Figures 2 and 3). The second antagonist to enter the clinic in 2007 was ambrisentan (Letairis, Volibris, LU208075, Figures 2 and 3), which was reported to display some ET<sub>A</sub> receptor selectivity (Vatter and Seifert, 2006) followed by the most highly selective ET<sub>A</sub> receptor antagonist sitaxentan (Thelin, TBC11251) (Barst *et al.*, 2004). While hepatotoxicity is a known side effect of ET antagonists, it is usually reversible and related to dose. Unfortunately, cases of idiosyncratic hepatitis resulting in acute liver failure leading to death have

been reported with sitaxentan and the compound was withdrawn in 2010 (Don *et al.*, 2012).

### Next generation of ET antagonists: macitentan

Despite the current use of ET receptor antagonists and drugs targeting the two other principal pathways- that of NO with PDE5 inhibitors and that involving prostacyclin (PGI<sub>2</sub>) – meta-analysis of PAH trials shows existing therapies only moderately increased the most widely used objective evaluation of functional exercise capacity (6 min walk distance) by 11%. The prognosis for patients with PAH remains poor with ~15% mortality within 1 year. There remains an urgent need for new efficacious treatments that has led to the development of macitentan.

Macitentan (Opsumit, ACT-064992, Figures 2 and 3) represents the next generation of orally active ET receptor antagonists and was developed by modifying the structure of bosentan to improve efficacy and tolerability (Bolli *et al.*, 2012). Macitentan is described as a dual antagonist that blocks both ET<sub>A</sub> and ET<sub>B</sub> receptors and it inhibited [<sup>125</sup>I]-ET-1 binding to human recombinant ET<sub>A</sub> receptors with an IC<sub>50</sub> of 0.2nM and to ET<sub>B</sub> receptors with an IC<sub>50</sub> of 391 nM. On the basis of these results, macitentan displays about 800-fold



**Figure 3**

Selectivity of ET receptor antagonists for ET<sub>A</sub> versus ET<sub>B</sub> receptors shown on the vertical axis as reported by the companies that discovered the compounds. Selectivity was mainly determined by measuring affinity constants in separate competition assays against [<sup>125</sup>I]-ET-1 using human recombinant ET<sub>A</sub> versus ET<sub>B</sub> receptors and may not reflect selectivity measured in clinically relevant native tissues. Bosentan, ambrisentan and macitentan are currently approved for clinical use and are highlighted.

selectivity. A phase III clinical trial was successfully completed in 2012 (Pulido *et al.*, 2013), and the compound gained approval from the US FDA in 2013 for the treatment of PAH. Macitentan is metabolized by the cytochrome P450 system, predominantly CYP3A4 and to a lesser extent the CYP2C19 iso-enzyme. Unlike other antagonists currently in use, one of the metabolites of macitentan, ACT-132577 (Figure 2), is pharmacologically active. Although it has a lower potency than the parent compound ACT-132577 reaches higher plasma concentrations, with a longer half-life of about 48 h (Iglarz *et al.*, 2008; Sidharta *et al.*, 2011; 2013a,b). These factors are likely to contribute to improved activity of macitentan compared with bosentan. While *in vitro* studies suggested macitentan was likely to interact with other drugs (Weiss *et al.*, 2013), other observed pharmacokinetic benefits included fewer interactions with other drugs at clinically used concentrations, no requirement to alter doses in patients with renal or hepatic impairment, improved hepatic safety and reduced oedema/fluid retention compared with bosentan. Key differences were also identified in the pharmacodynamic parameters. For example, in calcium release assays macitentan was more potent ( $K_B = 0.1$  nM) than bosentan ( $K_B = 1.1$  nM) and had a significantly longer receptor occupancy (17 min compared with 70 s) (Iglarz *et al.*, 2008; Bruderer *et al.*, 2012a,b; 2013; Gatfield *et al.*, 2012). The authors suggested that the macitentan binding site differed slightly from the bosentan binding site and that this difference in interaction with amino acids in the receptor contributed to the slow dissociation of macitentan from the receptor, particularly leading to insurmountable antagonism. A number of clinical trials are actively recruiting (Patel and McKeage, 2014) including the use of macitentan for the treat-

ment of digital ulcers in patients with systemic sclerosis, Eisenmenger's syndrome and, perhaps the most challenging, in patients with brain tumours (glioblastoma).

## Compounds interacting with ET-1 synthesis and metabolism

Members of the neprilysin (NEP)-like family of zinc metalloendopeptidases play key roles in the ET pathway (Turner and Murphy, 1996; Turner *et al.*, 2001). Neutral endopeptidase (NEP) is a membrane-bound thermolysin-like zinc metalloendopeptidase, which is particularly abundant in human kidneys. The enzyme metabolizes a number of peptides including enkephalins, tachykinins, natriuretic peptides as well as the ETs (Turner and Tanzawa, 1997). Inactivation of ET-1 is via a two-stage process, opening of the Ser<sup>5</sup>-Leu<sup>6</sup> bond, followed by cleavage at the amino side of Ile<sup>19</sup> resulting in an inactive peptide, which is inhibited by phosphoramidon (Skolovsky *et al.*, 1990). Pharmacological intervention in the pathway is challenging because NEP-like enzymes also include the synthetic enzymes ECE-1, ECE-2 and KELL. The ECEs are also inhibited by phosphoramidon and ECE inhibitors currently in clinical trials have significant NEP inhibitory activity and it seems counter-intuitive to inhibit the degradative pathway. However, in practice, inactivation of ET-1 is thought to be mainly via binding and internalization of the ET<sub>B</sub> receptor and ET-1 is essentially stable in plasma. Binding to ET<sub>B</sub> receptors, particularly in those organs such as the lung expressing high densities of the subtype, are critical for inactivation of the peptide. After internalization of the ligand-

receptor complex to the lysosome, ET-1 is thought to be degraded, like other peptides, by cathepsin A. In support, cathepsin A knockout mice showed reduced ET-1 degradation and significantly increased arterial BP. In humans, genetic defects of cathepsin A include hypertension and cardiomyopathies (Seyrantepe *et al.*, 2008).

### ECE-1

It is now well established that ET is synthesized in a three-step process, with pre-pro-ET-1 initially cleaved by a signal peptidase to proET-1, which is in turn cleaved by a furin enzyme to an inactive precursor big ET-1 (Figure 1). Although low MW inhibitors of furins have been reported, furins cleave a number of other proteins to mature or active forms and therefore are not an easy tractable drug target for selectively reducing ET-1, without altering other pathways. Targeting the ECE enzymes responsible for transformation of big ET-1 to the mature, biologically active ET-1 has been more promising (Xu *et al.*, 1994; Turner and Murphy, 1996). In humans there are four isoforms, ECE-1a–d, derived from a single gene by the action of different promoters. Structurally, they differ only in the amino acid sequence of the extreme N-terminus. ECE-1 localizes to the small secretory vesicles of the constitutive pathway from where ET-1 is continuously released to maintain normal vascular tone. Unusually for vasoactive peptides, ET-1 is also synthesized by ECE-1 and stored in specialized Weibel–Palade bodies within endothelial cells until its release following an external physiological or pathophysiological stimulus (the regulated pathway) to produce further vasoconstriction (Russell *et al.*, 1998a,b; Russell and Davenport, 1999b).

In addition to intracellular endothelial cell ECE, the enzyme is also present on vascular smooth muscle, efficiently converts big ET-1 in human vessels *in vitro* and is up-regulated in atherosclerosis (Maguire *et al.*, 1997; Maguire and Davenport, 1998). Given the larger volume of the smooth muscle compared with the single layer of endothelium, smooth muscle ECE may be a more important source of ET-1 in pathophysiological conditions.

### ECE-2

ET-1 is also synthesized by a second membrane-bound metalloprotease, ECE-2 (Emoto and Yanagisawa, 1995; Yanagisawa *et al.*, 2000; Lorenzo *et al.*, 2001) with ~60% sequence similarity to ECE-1. It is distinguishable from ECE-1 by having an optimum pH of 5.5 for activity. In human endothelial cells, ECE-2 was localized to the acidified environment of vesicles of the secretory pathway, but unlike ECE-1 it is not found in storage granules (Russell and Davenport, 1999b). Four isoforms exist, differing in their N-terminus: ECE-2a-1 and ECE-2a-2 are expressed predominantly in peripheral tissues and ECE-2b-1 and ECE-2b-2 in the brain, possibly representing the neuronal isoforms (Ikeda *et al.*, 2002). The physiological importance of this pathway for ET-1 synthesis remains to be determined, as ECE-2 also metabolizes other peptides such as bradykinin. However, the requirement for an acidic pH suggests a role in pathophysiological conditions associated with low pH such as ischaemia. ECE-1/ECE-2 knockout mice display increased developmental defects compared with deletion of ECE-1 or ECE-2.

### Alternative, non-ECE synthetic pathway: chymase

ET-1 can also be synthesized indirectly by chymase, a serine protease present in mast cells. Big ET-1 is converted to ET-1<sub>1–31</sub> by cleaving the Tyr<sup>31</sup>–Gly<sup>32</sup> bond (Figure 1), which in turn is converted to the mature peptide via Trp<sup>21</sup>–Val<sup>22</sup> bond (Fecteau *et al.*, 2005; D’Orleans-Juste *et al.*, 2008). The existence of an alternative pathway was originally predicted when ET-1 and ET-2 were detected in embryos of the ECE-1/ECE-2 double-knockout mouse (Yanagisawa *et al.*, 2000).

The importance of this alternative pathway remains unclear, but importantly ET-1<sub>1–31</sub> was equipotent with big ET-1 in causing vasoconstriction in human isolated vessels, including coronary arteries, and this was associated with the appearance of measurable levels of ET-1 in the bathing medium. ET-1<sub>1–31</sub> displayed no selectivity between ET<sub>A</sub> and ET<sub>B</sub> receptors in human heart and vasoconstriction was fully blocked by ET<sub>A</sub> receptor selective antagonists, reflecting the predominance of the ET<sub>A</sub> receptor on vascular smooth muscle (Maguire *et al.*, 2001; Maguire and Davenport, 2004). The precise physiological role of mast cells within human blood vessels is unclear, but following degranulation, which may occur under pathophysiological conditions, the mast cell chymase is associated with interstitial spaces with the potential to convert circulating big ET-1 and provide a further source of ET-1. Mast cell expression is increased in cardiovascular disease, for example in atherosclerotic lesions. It is therefore possible that the contribution of this pathway within the vasculature, leading to over-expression of ET-1, may be underestimated particularly in conditions of endothelial malfunction where opposing levels of endogenous vasodilators may be reduced. It is unclear whether under conditions of NEP/ECE inhibition the rising levels of big ET-1 would favour increased conversion by the serine protease pathway, thus increasing the pressor effect via ET<sub>A</sub> receptors or whether excretion of unmetabolized big ET-1 by the kidney would be sufficient to remove the elevated levels of precursors (Johnström *et al.*, 2010).

### KELL and ET-3 synthesis

Although big ET-3 is converted by ECE-1 to ET-3, owing to difference in the C-terminus the efficiency is much less than for ET-1. In contrast, big ET-3 is reported to be efficiently converted by KELL (Lee *et al.*, 1999). KELL is a membrane-bound glycoprotein expressed in human erythrocytes and one of the major antigens; it is also related to mammalian NEP-like enzymes including ECE-1 and ECE-2 (Turner and Tanzawa, 1997). If KELL is the main synthetic pathway for ET-3, a possible benefit of inhibiting ECE would be to increase the ratio of ET-3 to ET-1, which could then differentially produce beneficial vasodilatation via the ET<sub>B</sub> receptor, but this speculative hypothesis has not been tested.

### Pharmacological inhibition of ECE by research compounds

A combination of phosphoramidon and thiorphan has been widely used to identify ECE activity. This is based on the finding that the conversion of big ET-1 to ET-1 is inhibited by phosphoramidon, but not by thiorphan, and has been shown both *in vitro* and *in vivo*. Importantly for evaluating the sig-

nificance of animal models, both compounds have also been used in clinical studies to characterize big ET-1 conversion (see Webb, 1995; Plumpton *et al.*, 1996a; Hand *et al.*, 1999). Low MW, non-peptide ECE inhibitors have been developed and one that has been widely used *in vitro* and *in vivo* animal models and is commercially available is CGS26303 (De Lombaert *et al.*, 1994). CGS26303 inhibited conversion of all three big ETs in human isolated blood vessels but, importantly, did not interfere with the interaction of mature peptides with ET receptors (Yap *et al.*, 2000). Although primarily an NEP inhibitor, SOL1, a more recent combined NEP/ECE non-peptide inhibitor with modest inhibition of ECE-1 *in vitro*, was remarkably potent *in vivo*, fully blocking the big ET-1-induced rise in BP at a dose of 10  $\mu\text{mol}\cdot\text{kg}^{-1}$  (Nelissen *et al.*, 2012).

A disadvantage of using phosphoramidon is that it is not selective for ECE. An alternative tool compound is PD159790, which inhibits ECE-1 with an  $\text{IC}_{50}$  value of 3  $\mu\text{M}$ ; at this concentration the compound is selective for ECE-1 over NEP (Ahn *et al.*, 1998). PD159790 has been shown experimentally in HUVECs to inhibit conversion of big ET-1 at pH 6.9, optimum for ECE-1, but did not affect big ET-1 conversion to the mature peptide at pH 5.4, optimum for ECE-2 (Russell and Davenport, 1999a). The compound did not inhibit the further metabolism of ET-1<sub>1-31</sub>, the chymase product of big ET-1 (Maguire *et al.*, 2001) and can be used to distinguish between the three different pathways for ET synthesis. While the mature peptide is located in intracellular Weibel–Palade bodies or secretory vesicles within endothelial cells and a proportion of big ET-1 is converted to ET-1 intracellularly, it is not reported whether ECE inhibitors can cross the plasma membrane to access these intracellular sites. The main effects of these inhibitors may be on external ECE. In agreement with this proposal, the SLV306 metabolite KC-12615 (see later) effectively prevented conversion of exogenous big ET-1 in human vasculature (Seed *et al.*, 2012).

### Emerging NEP/ECE inhibitors

Selective inhibitors of ECE have not progressed into clinical applications. SLV306 (daglutril, Figure 2) is an orally active, mixed enzyme inhibitor of both ECE and NEP. It is a pro-drug being converted *in vivo* to the active metabolite, KC-12615. This latter molecule has a pharmacological profile similar to phosphoramidon, inhibiting NEP in the nanomolar range, but with more modest inhibition in the micromolar range for ECE (Meil *et al.*, 1998; Jeng *et al.*, 2002). The therapeutic basis is that while inhibition of NEP alone increased plasma concentrations of atrial natriuretic factor (ANP) to cause vasodilatation, NEP inhibitors are ineffective as anti-hypertensives, probably because NEP also degrades vasoconstrictor peptides including ET. A combined ECE/NEP inhibitor would be predicted to reduce the systemic conversion of big ET-1 to the mature peptide and increase dilator peptides such as ANP. SLV306 is well tolerated with few or none of the side effects such as increases in liver function, oedema, observed with ET receptor antagonists (Dickstein *et al.*, 2004; Parvanova *et al.*, 2013). A potential disadvantage is big ET-1 might still be converted to ET-1 by an alternative pathway such as chymase. However, in animal models with normal renal function, this did not occur and big ET-1 labelled with the positron emitter  $^{18}\text{F}$  was rapidly accumulated unchanged in the kidney follow-

ing inhibition of NEP/ECE, with no evidence of conversion by another pathway (Johnström *et al.*, 2010).

The effect of a combined NEP/ECE inhibitor has been tested in volunteers in a randomized, double-blind trial. Following oral administration of three increasing doses of SLV306 (to reach an average target concentration of 75, 300, 1200  $\text{ng}\cdot\text{mL}^{-1}$  of the active metabolite KC-12615), big ET-1 was infused into 13 male volunteers at a rate of 8 and 12  $\text{pmol}\cdot\text{kg}^{-1}\cdot\text{min}^{-1}$  (20 min each). At the two highest concentrations tested, SLV306 dose-dependently attenuated the rise in BP after big ET-1 infusion. There was a significant increase in circulating big ET-1 levels compared with placebo, indicating that SLV306 was inhibiting an increasing proportion of endogenous ECE activity. Importantly, plasma ANP concentrations also significantly increased, consistent with systemic NEP inhibition (Seed *et al.*, 2012).

### SLV306 in animal models and patients with type 2 diabetes and nephropathy

Diabetes causes activation of the renal ET system, which leads to progressive renal damage by cell proliferation and interstitial inflammation. Inhibitors of the renin–angiotensin system are widely used in treatment for hypertensive patients with type 2 diabetes, but are less effective in the advanced stages of diabetic renal disease. Studies in an animal model suggested that SLV306 had a similar efficacy to the angiotensin converting enzyme (ACE) inhibitor captopril in reducing proteinuria and preventing nephrosclerosis (Thone-Reinke *et al.*, 2004). In this study, rats were treated with streptozotocin for twenty weeks and the effects of SLV306 (30  $\text{mg}\cdot\text{kg}^{-1}$  per day) compared with those of captopril (10  $\text{mg}\cdot\text{kg}^{-1}$  per day). SLV306 significantly decreased renal interstitial matrix content as well as protein and albumin excretion in diabetic rats, independent of BP and was as effective as captopril. These results suggested SLV306 treatment on top of blocking the renin–angiotensin system might have an additional benefit in reducing BP and improving renal function.

Parvanova *et al.* (2013) tested the efficacy of SLV306 in 45 patients with type 2 diabetes mellitus who had albuminuria and were already receiving the angiotensin receptor antagonist losartan, together with up to two additional anti-hypertensive drugs, in a randomized, crossover, double-blind, placebo-controlled trial. Although 8 weeks of treatment with SLV306 together with losartan did not significantly alter urinary albumin excretion or renal haemodynamic measures, the authors showed for the first time that the combination decreased ambulatory BP (particularly for systolic hypertension) in this patient group that are often resistant to treatment. There was a small, but significant increase in plasma big ET-1, consistent with ECE inhibition, but surprisingly not in pro-ANP. Increases in the natriuretic peptides was measured in healthy volunteers by Seed *et al.* (2012). Interestingly, the effect of SLV306 in this study on BP was higher at night (10 versus 12 mmHg). This is of potential importance as increased hypertension at night is a strong cardiovascular risk factor in this patient population. The molecular mechanism is not yet known, as plasma levels of big ET-1 were not reported separately for daytime versus night. The study was comparatively short and did not reveal significant changes in albumin excretion as predicted from animal studies. Long-

term trials are required to determine whether the observed lowering of BP by SLV306 will translate into longer-term renal and cardio-protection.

### *SLV306 and congestive heart failure*

The effect of three single oral doses of SLV306 was tested in patients with congestive heart failure who underwent right-sided heart catheterization in a randomized, double-blind, placebo-controlled design (Dickstein *et al.*, 2004). Pulmonary pressures and right atrial pressure decreased significantly in all SLV306 dose groups with the maximum decrease occurring at 6–8 h. Despite plasma levels of the drug increasing with dose, there was no clear dose–response relationship, which may have been the result of the comparatively small numbers (18–20) in the study.

### *Insight in NEP/ECE inhibition from animal models*

The efficacy of inhibiting NEP/ECE in animal models associated with increases in the ET signalling pathway has provided clues to future clinical applications. The development of nephropathy in diabetes is associated with a poor outcome, eventually leading to end-stage renal disease. In patients with diabetes, urinary excretion of protein and albumin rises and is associated with increased risk of cardiovascular disease. In diabetic rats, SLV306 decreased renal matrix protein content, protein and albumin excretion. The magnitude of these effects was comparable to those of ACE inhibition and independent of BP (Thone-Reinke *et al.*, 2004). Currently, there are few drugs for the treatment of chronic renal failure. SLV338, a NEP/ECE inhibitor, abolished renal tissue damage (interstitial fibrosis, glomerulosclerosis, renal arterial remodelling) in rat models of both acute kidney failure as well as chronic renal failure. The compound preserved kidney function and reduced mortality (Sharkovska *et al.*, 2011). In spontaneously hypertensive stroke-prone rats, SLV338 significantly improved survival in comparison with the vehicle-treated group in a BP-independent manner and could offer a new therapeutic approach for primary stroke prevention and improvement of mortality (Wengenmayer *et al.*, 2011). SLV338 was also tested for cardiac protection in rat model of experimental renovascular hypertension (two-kidney, one-clip). SLV338 prevented cardiac remodelling to the same extent as losartan, but in a BP-independent manner. This effect was at least partly mediated via suppression of cardiac TGF- $\beta$ 1 expression (Kalk *et al.*, 2011).

ET has been proposed to be a mediator in toxic liver injury. However, while SLV338 largely prevented the activation of the ET system it did not prevent D-galactosamine-induced acute liver injury in rats. The authors speculated that SLV338 should be tested in a less severe model of liver injury, as very severe intoxication might not be relevantly amenable to pharmacological interventions (Hoher *et al.*, 2011).

### *ECE-1 and amyloid deposition*

The strategy in the cardiovascular and renal systems has been to *inhibit* ECE-1 activity. However, evidence is emerging that ECE-1 may function in the brain as a novel enzyme degrading amyloid  $\beta$ -peptides at several sites. Deposition of amyloid in the brain in Alzheimer's disease is determined not only by its

production, but also by its catabolism. ECE-1 inhibition produces, in addition to extracellular accumulation, accumulation of intracellular amyloid  $\beta$ -peptides within endosomal/lysosomal and autophagic vesicles and an intracellular pool, is partly regulated by ECE activity at the sites of production. Reduction in ECE activity leads to accumulation of amyloid  $\beta$ -peptide, which is associated with neurotoxicity early in progression of Alzheimer's disease (Eckman *et al.*, 2001; 2003; 2006; Pacheco-Quinto and Eckman, 2013). The clearance of  $A\beta_{1-40}$  in mice was almost completely inhibited by phosphoramidon as well as insulin indicating that human  $A\beta_{1-40}$  was degraded, at least in part, by a phosphoramidon-sensitive pathway, implicating both ECE and NEP (Ito *et al.*, 2013).

To date, these investigations have comprised *in vitro* or *in vivo* rodent studies. It is not yet clear whether *enhancing* ECE-1 activity is a potential drug target in Alzheimer's disease rather than *inhibiting* ECE-1, as in the periphery. ECE-like immunoreactivity has been localized to afferent and efferent fibres of neurones and neuronal cell bodies of mixed morphology in human brain (Giaid *et al.*, 1991). Drugs increasing ECE activity such as enzyme enhancers or recombinant ECE would have to cross the blood–brain barrier, and it is not clear what effect this would have on ET signalling in the periphery.

## What are the new ET drug targets in the future?

### *Epigenetics*

Epigenetics can be defined as heritable changes in phenotype through mechanisms other than changes in DNA sequence. Epigenetic changes will therefore be preserved when cells divide and affect normal development and disease progression. Processes mediating epigenetic regulation include DNA methylation and histone modification, which involves post-translational covalent modification of histone proteins by a range of writers, erasers and readers. This in turn modulates the ability of associated DNA to be transcribed. The histone code is read by specific families of proteins such as the bromodomains. These are of pharmacological significance because of the recent discovery of low MW inhibitors, which selectively modulate gene expression (Printha *et al.*, 2012).

Epigenetic regulation is of particular importance in the ET pathway with transcription being the primary level of ET-1 regulation of the gene *EDN1* by histone modifications and DNA methylation (Welch *et al.*, 2013). Silencing of the *EDNRB* gene by DNA methylation during development of tumours results in the down-regulation of the receptor. As a result, promotion of apoptosis via the  $ET_B$  receptor is reduced or lost, suggesting the  $ET_B$  receptor could be a target for epigenetic drugs or  $ET_B$  agonists where ET may be the cause of some tumour types, including melanomas and oligodendrogliomas (Bagnato *et al.*, 2011). Intriguingly, epigenetic inactivation of ET-2 and ET-3 mRNA and protein was found in rat and human colon tumours and cancer cell lines, as a result of hypermethylation of *EDN2* and *EDN3* genes. Restoring expression of ET-2 and ET-3 in human cells significantly attenuated the migration and invasion of human colon cancer cells (Wang *et al.*, 2013). As ET-3 displays high affinity for the  $ET_B$  receptor, forced expression of ET-3 might

antagonize the actions of ET-1 mediated through ET<sub>A</sub> receptors. Such a mechanism would be consistent with proposed beneficial effects of the ET<sub>B</sub> receptor agonist, IRL-1620, in cancer.

### *Life before birth – is ET a critical pathway?*

Maternal malnutrition and uteroplacental vascular insufficiency causes foetal growth restriction or intrauterine growth retardation. Low birthweight is linked to the later development of cardiovascular disease and hypertension. Maternal treatment with dexamethasone increased ET-1 constrictor responses and ET<sub>A</sub> receptor expression in placental arteries from the foetus (Docherty *et al.*, 2001; Kutzler *et al.*, 2003). Maternal nutrient restriction increased the histone acetylation and hypoxia inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ) binding levels in the ET-1 gene promoter of pulmonary vein endothelial cells (PVECs) in intrauterine growth restriction (IUGR) newborn rats, and continued up to 6 weeks after birth. These epigenetic changes could result in an IUGR rat being highly sensitive to hypoxia later in life, causing more significant PAH or pulmonary vascular remodelling. Recently, Xu *et al.* (2013) have shown that restricting the diets of pregnant rats so that they were undernourished increased the histone acetylation and HIF-1 $\alpha$  binding levels in the proximal promoter region of ET-1, up-regulating the expression of ET-1, and this continued for 6 weeks after birth of the offspring. The authors speculate that that this intrauterine growth retardation could cause varying degrees of PAH later in life.

Generally, increased levels of histone acetylation are associated with increased transcriptional activity, whereas decreased levels of acetylation are correlated with suppressed gene expression. These data show that the open chromatin domains marked by histone H3 and H3K9/18 acetylation at the proximal promoter of ET-1 in IUGR rats are essential for transcription. Up-regulated ET-1 protein expression in PVEC from IUGR hypoxia rats is closely associated with the presence of increased acetylated H3 histones.

### *Biased signalling in the ET pathway*

Pharmacology is undergoing a revolution in understanding the mechanism of 'biased signalling' via GPCRs. It was originally thought that ligands binding to a receptor would equally activate the G-protein pathway to produce a physiological response such as vasoconstriction (such as ET-1 acting on an ET<sub>A</sub> receptor) as well as activating the  $\beta$ -arrestin pathway, which eventually leads to desensitization, receptor internalization and 'silencing' of the pathway. It is now clear that some ligands are biased to one pathway over the other and secondly, rather than silencing,  $\beta$ -arrestin can activate alternative signalling pathways, some of which may be pathophysiological leading to longer-term signalling responses such as migration and proliferation.

Both ET receptor subtypes follow an  $\beta$ -arrestin and dynamin/clathrin-dependent mechanism of internalization, but it has been established that ET<sub>A</sub> receptors are recycled to the plasma membrane for further signalling while ET<sub>B</sub> receptors are targeted to lysosomes and degraded (Bremnes *et al.*, 2000). In epithelial ovarian cancer, activation of ET-1/ET<sub>A</sub> receptor signalling is linked to many tumour-promoting processes including proliferation, angiogenesis, invasion and

metastasis. NF- $\kappa$ B is an important signalling molecule in immunity, inflammation and cancer and  $\beta$ -arrestin is required for ET-1-induced NF- $\kappa$ B activation (Cianfrocca *et al.*, 2014). ET-1 promoted podocyte migration via ET<sub>A</sub> receptors and increased  $\beta$ -arrestin-1, sustaining renal injury, a pathogenic pathway that can affect podocyte phenotype in proliferative glomerular disorders (Buell *et al.*, 2014).  $\beta$ -Arrestin-1 has also been found to be a nuclear transcriptional regulator of ET-1-induced  $\beta$ -catenin signalling, an important mechanism for controlling cell division and progression of epithelial ovarian cancer and necessary for epigenetic modification, such as histone acetylation, and gene expression (Rosanò *et al.*, 2009; 2013a). In addition, these effects are blocked by ET receptor antagonists and support a role for ET<sub>A</sub>-mediated/ $\beta$ -arrestin-1 facilitating inter-protein interaction in invasive and metastatic behaviour of ovarian cancer.

### *Biased ET ligands?*

Agonists that are biased towards  $\beta$ -arrestin signalling for parathyroid hormone and angiotensin AT<sub>1</sub> receptors have been identified. G-protein pathway-selective agonists have been identified for nicotinic acid (nomenclature revised by NC-IUPHAR to hydroxycarboxylic acid) and  $\mu$ -opioid receptors (Luttrell, 2014). The race is now to determine whether such strategies can be exploited therapeutically.

Do biased ligands (ligands that binding to the same receptor but activate different signalling pathways) exist for ET receptors? The study by Compeer *et al.* (2013) already mentioned suggested ET-1 and ET-2 initiate and maintain vasoconstriction by different downstream mechanisms. Biased signalling can be identified by comparing the affinities of ligands in  $\beta$ -arrestin recruitment assays with a G-protein-mediated response such as vasoconstriction (Maguire *et al.*, 2012). In this study the rank order of potency for  $\beta$ -arrestin recruitment at the ET<sub>A</sub> (ET-1  $\geq$  ET-2  $>$  ET-3) and ET<sub>B</sub> (ET-1 = ET-2 = ET-3) receptors was as expected and there was no obvious major differences in potency of ETs when comparing with G-protein-mediated constrictor assays in human vessels. However, at the ET<sub>A</sub> receptor sarafotoxin S6b was a partial agonist in  $\beta$ -arrestin recruitment, but a full agonist in causing constriction, suggesting the possibility of biased ligands. Such a bias could have been selected for during evolution by prolonging the effects of envenomation of the mammalian prey. While bosentan displays no selectivity for ET<sub>A</sub> over ET<sub>B</sub> receptors in radioligand binding and G-protein functional assays, unexpectedly, it was significantly more effective an inhibitor of  $\beta$ -arrestin recruitment mediated by ET<sub>A</sub>, compared with the ET<sub>B</sub> receptors (Maguire *et al.*, 2012). The result for bosentan is intriguing as many of the detrimental actions of ET-1, particularly in cancer, may use the  $\beta$ -arrestin pathway, and this suggests the potential to block a deleterious pathway while preserving activation of a beneficial pathway.

### *The next 25 years – quo vadis?*

Since the discovery of ET, intense medicinal chemistry programmes have identified receptor antagonists, ET<sub>B</sub> receptor agonists and inhibitors of the key synthetic enzyme, ECE-1.

Both mixed ET<sub>A</sub>/ET<sub>B</sub> and ET<sub>A</sub> selective receptor antagonists have become established in the treatment of PAH, while NEP/ECE inhibitors such as SLV306 have promise as an alternative to receptor blockade and IRL1620 and other ET<sub>B</sub> receptor ligands have potential in improving cancer therapy.

All of these are approaches that have exploited low MW compounds. Over 50 therapeutic monoclonal antibodies have been approved for clinical use, but none yet against a GPCR target, emphasizing the technical challenge. Endomab-B1, a monoclonal antibody has recently been reported to bind with subnanomolar affinity for the ET<sub>B</sub> receptor, competed with ET-1 binding with greater efficacy than BQ788, and functions as an antagonist to block the ET-1-induced IP<sub>3</sub>-calcium signalling pathway (Allard *et al.*, 2013). Whether this antibody has clinical applications remains to be discovered.

## Acknowledgements

We thank the British Heart Foundation (PS/02/001, PG/05/127/19872, FS/12/64/130001), the Wellcome Trust Programme in Metabolic and Cardiovascular Disease (096822/Z/11/Z), the National Institute for Health Research Cambridge Biomedical Research Centre and the Pulmonary Hypertension Association UK.

## Conflict of interest

None.

## References

- Adner M, Jansen I, Edvinsson L (1994). Endothelin-A receptors mediate contraction in human cerebral, meningeal and temporal arteries. *J Auton Nerv Syst* 49 (Suppl.): S117–S121.
- Ahn K, Sisneros AM, Herman SB, Pan SM, Hupe D, Lee C *et al.* (1998). Novel selective quinazoline inhibitors of endothelin converting enzyme-1. *Biochem Biophys Res Commun* 243: 184–190.
- Alexander SPH, Benson HE, Faccenda E, Pawson AJ, Sharman JL, Spedding M *et al.* (2013a). The concise guide to PHARMACOLOGY 2013/14: G protein-coupled receptors. *Br J Pharmacol* 170: 1459–1581.
- Alexander SPH, Benson HE, Faccenda E, Pawson AJ, Sharman JL, Spedding M *et al.* (2013b). The concise Guide to PHARMACOLOGY 2013/14: Enzymes. *Br J Pharmacol* 170: 1797–1867.
- Allard B, Wijkhuisen A, Borrull A, Deshayes F, Priam F, Lamourette P *et al.* (2013). Generation and characterization of rendomab-B1, a monoclonal antibody displaying potent and specific antagonism of the human endothelin B receptor. *MAbs* 5: 56–69.
- Arai H, Hori S, Aramori I, Ohkubo H, Nakanishi S (1990). Cloning and expression of a cDNA encoding an endothelin receptor. *Nature* 348: 730–732.
- Bagnall AJ, Kelland NF, Gulliver-Sloan F, Davenport AP, Gray GA, Yanagisawa M *et al.* (2006). Deletion of endothelial cell endothelin B receptors does not affect blood pressure or sensitivity to salt. *Hypertension* 48: 286–293.
- Bagnato A, Loizidou M, Pflug BR, Curwen J, Growcott J (2011). Role of the endothelin axis and its antagonists in the treatment of cancer. *Br J Pharmacol* 163: 220–233.
- Barst RJ, Langleben D, Frost A, Horn EM, Oudiz R, Shapiro S *et al.* (2004). Sitaxsentan therapy for pulmonary arterial hypertension. *Am J Respir Crit Care Med* 169: 441–447.
- Barton M, Yanagisawa M (2008). Endothelin: 20 years from discovery to therapy. *Can J Physiol Pharmacol* 86: 485–498.
- Baynash AG, Hosoda K, Giaid A, Richardson JA, Emoto N, Hammer RE *et al.* (1994). Interaction of endothelin-3 with endothelin-B receptor is essential for development of epidermal melanocytes and enteric neurons. *Cell* 79: 1277–1285.
- Bolli MH, Boss C, Binkert C, Buchmann S, Bur D, Hess P *et al.* (2012). The discovery of N-[5-(4-bromophenyl)-6-[2-[(5-bromo-2-pyrimidinyl)oxy]ethoxy]-4-pyrimidinyl]-N'-p ropylsulfamide (Macitentan), an orally active, potent dual endothelin receptor antagonist. *J Med Chem* 55: 7849–7861.
- Bramall AN, Szego MJ, Pacione LR, Chang I, Diez E, D'Orleans-Juste P *et al.* (2013). Endothelin-2-mediated protection of mutant photoreceptors in inherited photoreceptor degeneration. *PLoS ONE* 8: e58023.
- Braunger BM, Ohlmann A, Koch M, Tanimoto N, Volz C, Yang Y *et al.* (2013). Constitutive overexpression of Norrin activates Wnt/beta-catenin and endothelin-2 signaling to protect photoreceptors from light damage. *Neurobiol Dis* 50: 1–12.
- Bremnes T, Paasche JD, Mehlum A, Sandberg C, Bremnes B, Attramadal H (2000). Regulation and intracellular trafficking pathways of the endothelin receptors. *J Biol Chem* 275: 17596–17604.
- Briyal S, Shepard C, Gulati A (2014). Endothelin receptor type B agonist, IRL-1620, prevents beta amyloid (Aβ) induced oxidative stress and cognitive impairment in normal and diabetic rats. *Pharmacol Biochem Behav* 120C: 65–72.
- Bruderer S, Aanismaa P, Homery MC, Hausler S, Landskroner K, Sidharta PN *et al.* (2012a). Effect of cyclosporine and rifampin on the pharmacokinetics of macitentan, a tissue-targeting dual endothelin receptor antagonist. *AAPS J* 14: 68–78.
- Bruderer S, Hopfgartner G, Seiberling M, Wank J, Sidharta PN, Treiber A *et al.* (2012b). Absorption, distribution, metabolism, and excretion of macitentan, a dual endothelin receptor antagonist, in humans. *Xenobiotica* 42: 901–910.
- Bruderer S, Marjason J, Sidharta PN, Dingemans J (2013). Pharmacokinetics of macitentan in Caucasian and Japanese subjects: the influence of ethnicity and sex. *Pharmacology* 91: 331–338.
- Buelli S, Rosano L, Gagliardini E, Corna D, Longaretti L, Pezzotta A *et al.* (2014). Beta-arrestin-1 drives endothelin-1-mediated podocyte activation and sustains renal injury. *J Am Soc Nephrol* 25: 523–533.
- Cade C, Lumma WC Jr, Mohan R, Rubanyi GM, Parker-Botelho LH (1990). Lack of biological activity of preproendothelin [110–130] in several endothelin assays. *Life Sci* 47: 2097–2103.
- Chang I, Bramall AN, Baynash AG, Rattner A, Rakheja D, Post M *et al.* (2013). Endothelin-2 deficiency causes growth retardation, hypothermia, and emphysema in mice. *J Clin Invest* 123: 2643–2653.
- Cianfrocca R, Tocci P, Semprucci E, Spinella F, Di Castro V, Bagnato A *et al.* (2014). Beta-arrestin 1 is required for endothelin-1-induced NF-kappaB activation in ovarian cancer cells. *Life Sci*. doi: 10.1016/j.lfs.2014.01.078
- Clouthier DE, Hosoda K, Richardson JA, Williams SC, Yanagisawa H, Kuwaki T *et al.* (1998). Cranial and cardiac neural crest defects in endothelin-A receptor-deficient mice. *Development* 125: 813–824.

- Clozel M, Gray GA (1995). Are there different ETB receptors mediating constriction and relaxation? *J Cardiovasc Pharmacol* 26 (Suppl. 3): S262–S264.
- Compeer MG, Janssen GM, De Mey JG (2013). Endothelin-1 and endothelin-2 initiate and maintain contractile responses by different mechanisms in rat mesenteric and cerebral arteries. *Br J Pharmacol* 170: 1199–1209.
- Davenport AP (2002). International Union of Pharmacology. XXIX. Update on endothelin receptor nomenclature. *Pharmacol Rev* 54: 219–226.
- Davenport AP, Maguire JJ (2006). Endothelin. *Handb Exp Pharmacol* 152: 295–329.
- Davenport AP, Ashby MJ, Easton P, Ella S, Bedford J, Dickerson C *et al.* (1990). A sensitive radioimmunoassay measuring endothelin-like immunoreactivity in human plasma: comparison of levels in patients with essential hypertension and normotensive control subjects. *Clin Sci (Lond)* 78: 261–264.
- Davenport AP, Alexander SP, Sharman JL, Pawson AJ, Benson HE, Monaghan AE *et al.* (2013). International Union of Basic and Clinical Pharmacology. LXXXVIII. G protein-coupled receptor list: recommendations for new pairings with cognate ligands. *Pharmacol Rev* 65: 967–986.
- De Lombaert S, Ghai RD, Jeng AY, Trapani AJ, Webb RL (1994). Pharmacological profile of a non-peptidic dual inhibitor of neutral endopeptidase 24.11 and endothelin-converting enzyme. *Biochem Biophys Res Commun* 204: 407–412.
- Dhaun N, Goddard J, Webb DJ (2011). Endothelin antagonism in patients with nondiabetic chronic kidney disease. *Contrib Nephrol* 172: 243–254.
- Dhaun N, Webb DJ, Kluth DC (2012). Endothelin-1 and the kidney-beyond BP. *Br J Pharmacol* 167: 720–731.
- Dickstein K, De Voogd HJ, Miric MP, Willenbrock R, Mitrovic V, Pacher R *et al.* (2004). Effect of single doses of SLV306, an inhibitor of both neutral endopeptidase and endothelin-converting enzyme, on pulmonary pressures in congestive heart failure. *Am J Cardiol* 94: 237–239.
- Docherty CC, Kalmar-Nagy J, Engelen M, Koenen SV, Nijland M, Kuc RE *et al.* (2001). Effect of *in vivo* fetal infusion of dexamethasone at 0.75 GA on fetal ovine resistance artery responses to ET-1. *Am J Physiol Regul Integr Comp Physiol* 281: R261–R268.
- Don GW, Joseph F, Celermajer DS, Corte TJ (2012). Ironic case of hepatic dysfunction following the global withdrawal of sitaxentan. *Intern Med J* 42: 1351–1354.
- D’Orleans-Juste P, Houde M, Rae GA, Bkaily G, Carrier E, Simard E (2008). Endothelin-1 (1–31): from chymase-dependent synthesis to cardiovascular pathologies. *Vascul Pharmacol* 49: 51–62.
- Drawnel FM, Archer CR, Roderick HL (2013). The role of the paracrine/autocrine mediator endothelin-1 in regulation of cardiac contractility and growth. *Br J Pharmacol* 168: 296–317.
- Eckman EA, Reed DK, Eckman CB (2001). Degradation of the Alzheimer’s amyloid beta peptide by endothelin-converting enzyme. *J Biol Chem* 276: 24540–24548.
- Eckman EA, Watson M, Marlow L, Sambamurti K, Eckman CB (2003). Alzheimer’s disease beta-amyloid peptide is increased in mice deficient in endothelin-converting enzyme. *J Biol Chem* 278: 2081–2084.
- Eckman EA, Adams SK, Troendle FJ, Stodola BA, Kahn MA, Fauq AH *et al.* (2006). Regulation of steady-state beta-amyloid levels in the brain by neprilysin and endothelin-converting enzyme but not angiotensin-converting enzyme. *J Biol Chem* 281: 30471–30478.
- Emoto N, Yanagisawa M (1995). Endothelin-converting enzyme-2 is a membrane-bound, phosphoramidon-sensitive metalloprotease with acidic pH optimum. *J Biol Chem* 270: 15262–15268.
- Fecteau MH, Honore JC, Plante M, Labonte J, Rae GA, D’Orleans-Juste P (2005). Endothelin-1 (1–31) is an intermediate in the production of endothelin-1 after big endothelin-1 administration *in vivo*. *Hypertension* 46: 87–92.
- Flynn MA, Haleen SJ, Welch KM, Cheng XM, Reynolds EE (1998). Endothelin B receptors on human endothelial and smooth-muscle cells show equivalent binding pharmacology. *J Cardiovasc Pharmacol* 32: 106–116.
- Foord SM, Bonner TI, Neubig RR, Rosser EM, Pin JP, Davenport AP *et al.* (2005). International Union of Pharmacology. XLVI. G protein-coupled receptor list. *Pharmacol Rev* 57: 279–288.
- Gatfield J, Mueller Grandjean C, Sasse T, Clozel M, Nayler O (2012). Slow receptor dissociation kinetics differentiate macitentan from other endothelin receptor antagonists in pulmonary arterial smooth muscle cells. *PLoS ONE* 7: e47662.
- Giaid A, Gibson SJ, Herrero MT, Gentleman S, Legon S, Yanagisawa M *et al.* (1991). Topographical localisation of endothelin mRNA and peptide immunoreactivity in neurones of the human brain. *Histochemistry* 95: 303–314.
- Gulati A, Rai A (2004). Endothelin-1-induced vasodilatation in rat Breast tumor is mediated through endothelin-B receptors. *J Cardiovasc Pharmacol* 44: S483–S486.
- Gulati A, Sunila ES, Kuttan G (2012). IRL-1620, an endothelin-B receptor agonist, enhanced radiation induced reduction in tumor volume in Dalton’s Lymphoma Ascites tumor model. *Arzneimittelforschung* 62: 14–17.
- Hand MF, Haynes WG, Webb DJ (1999). Reduced endogenous endothelin-1-mediated vascular tone in chronic renal failure. *Kidney Int* 55: 613–620.
- Harland SP, Kuc RE, Pickard JD, Davenport AP (1995). Characterization of endothelin receptors in human brain cortex, gliomas, and meningiomas. *J Cardiovasc Pharmacol* 26 (Suppl. 3): S408–S411.
- Harland SP, Kuc RE, Pickard JD, Davenport AP (1998). Expression of endothelin(A) receptors in human gliomas and meningiomas, with high affinity for the selective antagonist PD156707. *Neurosurgery* 43: 890–898.
- Haynes WG, Webb DJ (1994). Contribution of endogenous generation of endothelin-1 to basal vascular tone. *Lancet* 344: 852–854.
- Hoher B, Heiden S, von Websky K, Rahnenfuhrer J, Kalk P, Pfab T (2011). Dual endothelin-converting enzyme/neutral endopeptidase blockade in rats with D-galactosamine-induced liver failure. *Eur J Med Res* 16: 275–279.
- Hosoda K, Hammer RE, Richardson JA, Baynash AG, Cheung JC, Giaid A *et al.* (1994). Targeted and natural (piebald-lethal) mutations of endothelin-B receptor gene produce megacolon associated with spotted coat color in mice. *Cell* 79: 1267–1276.
- Howard PG, Plumpton C, Davenport AP (1992). Anatomical localization and pharmacological activity of mature endothelins and their precursors in human vascular tissue. *J Hypertens* 10: 1379–1386.
- Hyndman KA, Pollock JS (2013). Nitric oxide and the A and B of endothelin of sodium homeostasis. *Curr Opin Nephrol Hypertens* 22: 26–31.

- Iglarz M, Binkert C, Morrison K, Fischli W, Gatfield J, Treiber A *et al.* (2008). Pharmacology of macitentan, an orally active tissue-targeting dual endothelin receptor antagonist. *J Pharmacol Exp Ther* 327: 736–745.
- Ihara M, Noguchi K, Saeki T, Fukuroda T, Tsuchida S, Kimura S *et al.* (1992). Biological profiles of highly potent novel endothelin antagonists selective for the ET<sub>A</sub> receptor. *Life Sci* 50: 247–255.
- Ikeda S, Emto N, Alimsardjono H, Yokoyama M, Matsuo M (2002). Molecular isolation and characterization of novel four subisoforms of ECE-2. *Biochem Biophys Res Commun* 293: 421–426.
- Inoue A, Yanagisawa M, Kimura S, Kasuya Y, Miyuchi T, Goto K *et al.* (1989). The human endothelin family: three structurally and pharmacologically distinct isopeptides predicted by three separate genes. *Proc Natl Acad Sci U S A* 86: 2863–2867.
- Ishikawa K, Ihara M, Noguchi K, Mase T, Mino N, Saeki T *et al.* (1994). Biochemical and pharmacological profile of a potent and selective endothelinB-receptor antagonist, BQ-788. *Proc Natl Acad Sci U S A* 91: 4892–4896.
- Ito S, Matsumiya K, Ohtsuki S, Kamiie J, Terasaki T (2013). Contributions of degradation and brain-to-blood elimination across the blood–brain barrier to cerebral clearance of human amyloid-beta peptide(1–40) in mouse brain. *J Cereb Blood Flow Metab* 33: 1770–1777.
- Jeng AY, Mulder P, Kwan AL, Battistini B (2002). Nonpeptidic endothelin-converting enzyme inhibitors and their potential therapeutic applications. *Can J Physiol Pharmacol* 80: 440–449.
- Johnström P, Fryer TD, Richards HK, Maguire JJ, Clark JC, Pickard JD *et al.* (2010). Positron emission tomography of [18F]-big endothelin-1 reveals renal excretion but tissue-specific conversion to [18F]-endothelin-1 in lung and liver. *Br J Pharmacol* 159: 812–819.
- Kalk P, Sharkovska Y, Kashina E, von Websky K, Relle K, Pfab T *et al.* (2011). Endothelin-converting enzyme/neutral endopeptidase inhibitor SLV338 prevents hypertensive cardiac remodeling in a blood pressure-independent manner. *Hypertension* 57: 755–763.
- Karet FE, Davenport AP (1996). Localization of endothelin peptides in human kidney. *Kidney Int* 49: 382–387.
- Kelland NF, Kuc RE, McLean DL, Azfer A, Bagnall AJ, Gray GA *et al.* (2010). Endothelial cell-specific ETB receptor knockout: autoradiographic and histological characterisation and crucial role in the clearance of endothelin-1. *Can J Physiol Pharmacol* 88: 644–651.
- Kelland NF, Webb DJ (2006). Clinical trials of endothelin antagonists in heart failure: a question of dose? *Exp Biol Med* (Maywood) 231: 696–699.
- Kiowski W, Luscher TF, Linder L, Buhler FR (1991). Endothelin-1-induced vasoconstriction in humans. Reversal by calcium channel blockade but not by nitrovasodilators or endothelium-derived relaxing factor. *Circulation* 83: 469–475.
- Kirkby NS, Hadoke PW, Bagnall AJ, Webb DJ (2008). The endothelin system as a therapeutic target in cardiovascular disease: great expectations or bleak house? *J Pharmacol* 153: 1105–1119.
- Ko C, Gieske MC, Al-Alem L, Hahn Y, Su W, Gong MC *et al.* (2006). Endothelin-2 in ovarian follicle rupture. *Endocrinology* 147: 1770–1779.
- Ko C, Meidan R, Bridges PJ (2012). Why two endothelins and two receptors for ovulation and luteal regulation? *Life Sci* 91: 501–506.
- Kohan DE, Inscho EW, Wesson D, Pollock DM (2011a). Physiology of endothelin and the kidney. *Compr Physiol* 1: 883–919.
- Kohan DE, Rossi NF, Inscho EW, Pollock DM (2011b). Regulation of blood pressure and salt homeostasis by endothelin. *Physiol Rev* 91: 1–77.
- Kohan DE, Cleland JG, Rubin LJ, Theodorescu D, Barton M (2012). Clinical trials with endothelin receptor antagonists: what went wrong and where can we improve? *Life Sci* 91: 528–539.
- Kuc RE, Carlebur M, Maguire JJ, Yang P, Long L, Toshner M *et al.* (2014). Modulation of endothelin receptors in the failing right ventricle of the heart and vasculature of the lung in human pulmonary arterial hypertension. *Life Sci*. doi: 10.1016/j.lfs.2014.02.020
- Kurihara Y, Kurihara H, Suzuki H, Kodama T, Maemura K, Nagai R *et al.* (1994). Elevated blood pressure and craniofacial abnormalities in mice deficient in endothelin-1. *Nature* 368: 703–710.
- Kutzler MA, Molnar J, Schlafer DH, Kuc RE, Davenport AP, Nathanielsz PW (2003). Maternal dexamethasone increases endothelin-1 sensitivity and endothelin a receptor expression in ovine foetal placental arteries. *Placenta* 24: 392–402.
- Lee S, Lin M, Mele A, Cao Y, Farmer J, Russo D *et al.* (1999). Proteolytic processing of big endothelin-3 by the kallikrein blood group protein. *Blood* 94: 1440–1450.
- Lenaz L, Gulati A, Sunila E (2006). IRL-1620 increases the efficacy of radiation treatment in mice bearing lymphoma cell induced tumors. *Blood* 108: 269B.
- Leonard MG, Gulati A (2013). Endothelin B receptor agonist, IRL-1620, enhances angiogenesis and neurogenesis following cerebral ischemia in rats. *Brain Res* 1528: 28–41.
- Leonard MG, Briyal S, Gulati A (2011). Endothelin B receptor agonist, IRL-1620, reduces neurological damage following permanent middle cerebral artery occlusion in rats. *Brain Res* 1420: 48–58.
- Leonard MG, Briyal S, Gulati A (2012). Endothelin B receptor agonist, IRL-1620, provides long-term neuroprotection in cerebral ischemia in rats. *Brain Res* 1464: 14–23.
- Ling L, Maguire JJ, Davenport AP (2013). Endothelin-2, the forgotten isoform: emerging role in the cardiovascular system, ovarian development, immunology and cancer. *Br J Pharmacol* 168: 283–295.
- Liu C, Chen J, Gao Y, Deng B, Liu K (2013). Endothelin receptor antagonists for pulmonary arterial hypertension. *Cochrane Database Syst Rev* (2): CD004434.
- Lorenzo MN, Khan RY, Wang Y, Tai SC, Chan GC, Cheung AH *et al.* (2001). Human endothelin converting enzyme-2 (ECE2): characterization of mRNA species and chromosomal localization. *Biochim Biophys Acta* 1522: 46–52.
- Love MP, Ferro CJ, Haynes WG, Plumpton C, Davenport AP, Webb DJ *et al.* (2000). Endothelin receptor antagonism in patients with chronic heart failure. *Cardiovasc Res* 47: 166–172.
- Lucas GA, White LR, Juul R, Cappelen J, Aasly J, Edvinsson L (1996). Relaxation of human temporal artery by endothelin ETB receptors. *Peptides* 17: 1139–1144.
- Luttrell LM (2014). Minireview: more than just a hammer: ligand ‘bias’ and pharmaceutical discovery. *Mol Endocrinol* 28: 281–294.
- Maguire JJ, Davenport AP (1998). Increased response to big endothelin-1 in atherosclerotic human coronary artery: functional evidence for up-regulation of endothelin-converting enzyme activity in disease. *Br J Pharmacol* 125: 238–240.
- Maguire JJ, Davenport AP (2004). Alternative pathway to endothelin-converting enzyme for the synthesis of endothelin in human blood vessels. *J Cardiovasc Pharmacol* 44 (Suppl. 1): S27–S29.

- Maguire JJ, Johnson CM, Mockridge JW, Davenport AP (1997). Endothelin converting enzyme (ECE). Activity in human vascular smooth muscle. *Br J Pharmacol* 122: 1647–1654.
- Maguire JJ, Kuc RE, Davenport AP (2001). Vasoconstrictor activity of novel endothelin peptide, ET-1(1–31), in human mammary and coronary arteries *in vitro*. *Br J Pharmacol* 134: 1360–1366.
- Maguire JJ, Kuc RE, Pell VR, Green A, Brown M, Kumar S *et al.* (2012). Comparison of human ETA and ETB receptor signalling via G-protein and beta-arrestin pathways. *Life Sci* 91: 544–549.
- Marciniak SJ, Plumpton C, Barker PJ, Huskisson NS, Davenport AP (1992). Localization of immunoreactive endothelin and proendothelin in the human lung. *Pulm Pharmacol* 5: 175–182.
- Meil J, Wurl M, Thormahlen D, Rose H (1998). Pharmacology of the active metabolite of SLV 306 – a mixed inhibitor of NEP and ECE. *Naunyn Schmiedebergs Arch Pharmacol* 358 (Suppl. 1): R513.
- Meyer RC, Giddens MM, Schaefer SA, Hall RA (2013). GPR37 and GPR37L1 are receptors for the neuroprotective and glioprotective factors prosaptide and prosaposin. *Proc Natl Acad Sci U S A* 110: 9529–9534.
- Mizuguchi T, Nishiyama M, Moroi K, Tanaka H, Saito T, Masuda Y *et al.* (1997). Analysis of two pharmacologically predicted endothelin B receptor subtypes by using the endothelin B receptor gene knockout mouse. *Br J Pharmacol* 120: 1427–1430.
- Molenaar P, Kuc RE, Davenport AP (1992). Characterization of two new ETB selective radioligands, [125I]-BQ3020 and [125I]-[Ala1,3,11,15]ET-1 in human heart. *Br J Pharmacol* 107: 637–639.
- Molenaar P, O'Reilly G, Sharkey A, Kuc RE, Harding DP, Plumpton C *et al.* (1993). Characterization and localization of endothelin receptor subtypes in the human atrioventricular conducting system and myocardium. *Circ Res* 72: 526–538.
- de la Monte SM, Quertermous T, Hong CC, Bloch KD (1995). Regional and maturation-associated expression of endothelin 2 in rat gastrointestinal tract. *J Histochem Cytochem* 43: 203–209.
- Morton AJ, Davenport AP (1992). Cerebellar neurons and glia respond differentially to endothelins and sarafotoxin S6b. *Brain Res* 581: 299–306.
- Nambi P, Pullen M, Spielman W (1994). Species differences in the binding characteristics of [125I]IRL-1620, a potent agonist specific for endothelin-B receptors. *J Pharmacol Exp Ther* 268: 202–207.
- Nelissen J, Lemkens P, Sann H, Bindl M, Bassissi F, Jasserand D *et al.* (2012). Pharmacokinetic and pharmacodynamic properties of SOL1: a novel dual inhibitor of neutral endopeptidase and endothelin converting enzyme. *Life Sci* 91: 587–592.
- Ohkita M, Tawa M, Kitada K, Matsumura Y (2012). Pathophysiological roles of endothelin receptors in cardiovascular diseases. *J Pharmacol Sci* 119: 302–313.
- O'Reilly G, Charnock-Jones DS, Morrison JJ, Cameron IT, Davenport AP, Smith SK (1993). Alternatively spliced mRNAs for human endothelin-2 and their tissue distribution. *Biochem Biophys Res Commun* 193: 834–840.
- Pacheco-Quinto J, Eckman EA (2013). Endothelin-converting enzymes degrade intracellular beta-amyloid produced within the endosomal/lysosomal pathway and autophagosomes. *J Biol Chem* 288: 5606–5615.
- Palanisamy GS, Cheon YP, Kim J, Kannan A, Li Q, Sato M *et al.* (2006). A novel pathway involving progesterone receptor, endothelin-2, and endothelin receptor B controls ovulation in mice. *Mol Endocrinol* 20: 2784–2795.
- Palmer MJ (2009). Endothelin receptor antagonists: status and learning 20 years on. *Prog Med Chem* 47: 203–237.
- Parvanova A, van der Meer IM, Iliev I, Perna A, Gaspari F, Trevisan R *et al.* (2013). Effect on blood pressure of combined inhibition of endothelin-converting enzyme and neutral endopeptidase with daglutril in patients with type 2 diabetes who have albuminuria: a randomised, crossover, double-blind, placebo-controlled trial. *Lancet Diabetes Endocrinol* 1: 19–27.
- Patel T, McKeage K (2014). Macitentan: first global approval. *Drugs* 74: 127–133.
- Pawson AJ, Sharman JL, Benson HE, Faccenda E, Alexander SP, Buneman OP *et al.* (2014). The IUPHAR/BPS Guide to PHARMACOLOGY: an expert-driven knowledgebase of drug targets and their ligands. *Nucleic Acids Res* 42 (Database Issue): D1098–D1106.
- Pernow J, Shemyakin A, Böhm F (2012). New perspectives on endothelin-1 in atherosclerosis and diabetes mellitus. *Life Sci* 91: 507–516.
- Peter MG, Davenport AP (1995). Delineation of endothelin receptors in human left ventricular smooth-muscle cells. *J Cardiovasc Pharmacol* 26 (Suppl. 3): S355–S357.
- Peter MG, Davenport AP (1996). Characterization of the endothelin receptor selective agonist, BQ3020 and antagonists BQ123, FR139317, BQ788, 50235, Ro462005 and bosentan in the heart. *Br J Pharmacol* 117: 455–462.
- Pierre LN, Davenport AP (1999). Blockade and reversal of endothelin-induced constriction in pial arteries from human brain. *Stroke* 30: 638–643.
- Plumpton C, Ferro CJ, Haynes WG, Webb DJ, Davenport AP (1996a). The increase in human plasma immunoreactive endothelin but not big endothelin-1 or its C-terminal fragment induced by systemic administration of the endothelin antagonist TAK-044. *Br J Pharmacol* 119: 311–314.
- Plumpton C, Ashby MJ, Kuc RE, O'Reilly G, Davenport AP (1996b). Expression of endothelin peptides and mRNA in the human heart. *Clin Sci (Lond)* 90: 37–46.
- Prinjha RK, Witherington J, Lee K (2012). Place your BETs: the therapeutic potential of bromodomains. *Trends Pharmacol Sci* 33: 146–153.
- Pulido T, Adzerikho I, Channick RN, Delcroix M, Galie N, Ghofrani HA *et al.* (2013). Macitentan and morbidity and mortality in pulmonary arterial hypertension. *N Engl J Med* 369: 809–818.
- Rai A, Rajeshkumar NV, Gulati A (2006). Effect of the ET<sub>B</sub> receptor agonist, IRL-1620, on paclitaxel plasma pharmacokinetics of breast tumor rats. *Exp Biol Med* 231: 1120–1122.
- Rajeshkumar NV, Rai A, Gulati A (2005a). Endothelin B receptor agonist, IRL 1620, a novel adjuvant to enhance the delivery and efficacy of paclitaxel in a rat mammary tumor model. *Clin Cancer Res* 11: 9138S–9239S.
- Rajeshkumar NV, Rai A, Gulati A (2005b). Endothelin B receptor agonist, IRL 1620, enhances the anti-tumor efficacy of paclitaxel in breast tumor rats. *Breast Cancer Res Treat* 94: 237–247.
- Rattner A, Yu H, Williams J, Smallwood PM, Nathans J (2013). Endothelin-2 signaling in the neural retina promotes the endothelial tip cell state and inhibits angiogenesis. *Proc Natl Acad Sci U S A* 110: E3830–E3839.
- Rautureau Y, Schiffrin E (2012). Endothelin in hypertension: an update. *Curr Opin Nephrol Hypertens* 21: 128–136.

- Reynolds EE, Keiser JA, Haleen SJ, Walker DM, Olszewski B, Schroeder RL *et al.* (1995). Pharmacological characterization of PD 156707, an orally active ETA receptor antagonist. *J Pharmacol Exp Ther* 273: 1410–1417.
- Rosanò L, Cianfrocca R, Masi S, Spinella F, Di Castro V, Biroccio A *et al.* (2009).  $\beta$ -Arrestin links endothelin A receptor to beta-catenin signaling to induce ovarian cancer cell invasion and metastasis. *Proc Natl Acad Sci U S A* 106: 2806–2811.
- Rosanò L, Cianfrocca R, Tocci P, Spinella F, Di Castro V, Spadaro F *et al.* (2013a).  $\beta$ -Arrestin-1 is a nuclear transcriptional regulator of endothelin-1-induced beta-catenin signaling. *Oncogene* 32: 5066–5077.
- Rosanò L, Spinella F, Bagnato A (2013b). Endothelin 1 in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer* 13: 637–651.
- Rubin LJ (2012). Endothelin receptor antagonists for the treatment of pulmonary artery hypertension. *Life Sci* 91: 517–521.
- Rubin LJ, Badesch DB, Barst RJ *et al.* (2002). Bosentan therapy for pulmonary arterial hypertension. *N Engl J Med* 346: 896–903.
- Russell FD, Davenport AP (1996). Characterization of the binding of endothelin ETB selective ligands in human and rat heart. *Br J Pharmacol* 119: 631–636.
- Russell FD, Davenport AP (1999a). Evidence for intracellular endothelin-converting enzyme-2 expression in cultured human vascular endothelial cells. *Circ Res* 84: 891–896.
- Russell FD, Davenport AP (1999b). Secretory pathways in endothelin synthesis. *Br J Pharmacol* 26: 391–398.
- Russell FD, Skepper JN, Davenport AP (1998a). Human endothelial cell storage granules: a novel intracellular site for isoforms of the endothelin-converting enzyme. *Circ Res* 83: 314–321.
- Russell FD, Skepper JN, Davenport AP (1998b). Evidence using immunoelectron microscopy for regulated and constitutive pathways in the transport and release of endothelin. *J Cardiovasc Pharmacol* 31: 424–430.
- Said N, Theodorescu D (2012). Permissive role of endothelin receptors in tumor metastasis. *Life Sci* 91: 522–527.
- Sakurai T, Yanagisawa M, Takawa Y, Miyazaki H, Kimura S, Goto K *et al.* (1990). Cloning of a cDNA encoding a non-isopeptide-selective subtype of the endothelin receptor. *Nature* 348: 732–735.
- Seed A, Kuc RE, Maguire JJ, Hillier C, Johnston F, Essers H *et al.* (2012). The dual endothelin converting enzyme/neutral endopeptidase inhibitor SLV-306 (daglutril), inhibits systemic conversion of big endothelin-1 in humans. *Life Sci* 91: 743–748.
- Seyrantepe V, Hinek A, Peng J, Fedjaev M, Ernest S, Kadota Y *et al.* (2008). Enzymatic activity of lysosomal carboxypeptidase (cathepsin) A is required for proper elastic fiber formation and inactivation of endothelin-1. *Circulation* 117: 1973–1981.
- Sharkovska Y, Kalk P, von Websky K, Relle K, Pfab T, Alter M *et al.* (2011). Renoprotective effects of combined endothelin-converting enzyme/neutral endopeptidase inhibitor SLV338 in acute and chronic experimental renal damage. *Clin Lab* 57: 507–515.
- Sidharta PN, van Giersbergen PL, Halabi A, Dingemans J (2011). Macitentan: entry-into-humans study with a new endothelin receptor antagonist. *Eur J Clin Pharmacol* 67: 977–984.
- Sidharta PN, Lindegger N, Ulc I, Dingemans J (2013a). Pharmacokinetics of the novel dual endothelin receptor antagonist macitentan in subjects with hepatic or renal impairment. *J Clin Pharmacol* 55: 291–300.
- Sidharta PN, van Giersbergen PL, Dingemans J (2013b). Safety, tolerability, pharmacokinetics, and pharmacodynamics of macitentan, an endothelin receptor antagonist, in an ascending multiple-dose study in healthy subjects. *J Clin Pharmacol* 54: 291–300.
- Skolovsky M, Galron R, Kloog Y, Bdolah A, Indig FE, Blumberg S *et al.* (1990). Endothelins are more sensitive than sarafotoxins to neutral endopeptidase: possible physiological significance. *Proc Natl Acad Sci U S A* 87: 4702–4706.
- Speed JS, Pollock DM (2013). Endothelin, kidney disease, and hypertension. *Hypertension* 61: 1142–1145.
- Takasaki C, Tamiya N, Bdolah A, Wollberg Z, Kochva E (1988). Sarafotoxins S6: several isotoxins from *Atractaspis engaddensis* (burrowing asp) venom that affect the heart. *Toxicon* 26: 543–548.
- Takai M, Umemura I, Yamasaki K, Watakabe T, Fujitani Y, Oda K *et al.* (1992). A potent and specific agonist, Suc-[Glu<sup>9</sup>,Ala<sup>11</sup>,15]-endothelin-1(8–21), IRL 1620, for the ETB receptor. *Biochem Biophys Res Commun* 184: 953–959.
- Takizawa S, Uchida T, Adur J, Kozakai T, Kotake-Nara E, Quan J *et al.* (2005). Differential expression of endothelin-2 along the mouse intestinal tract. *J Mol Endocrinol* 35: 201–209.
- Thone-Reinke C, Simon K, Richter CM, Godes M, Neumayer HH, Thormahlen D *et al.* (2004). Inhibition of both neutral endopeptidase and endothelin-converting enzyme by SLV306 reduces proteinuria and urinary albumin excretion in diabetic rats. *J Cardiovasc Pharmacol* 44: S76–S79.
- Turner AJ, Murphy LJ (1996). Molecular pharmacology of endothelin converting enzymes. *Biochem Pharmacol* 26: 91–102.
- Turner AJ, Tanzawa K (1997). Mammalian membrane metalloproteinases: NEP, ECE, KELL, and PEX. *FASEB J* 11: 355–364.
- Turner AJ, Isaac RE, Coates D (2001). The neprilysin (NEP). family of zinc metalloendopeptidases: genomics and function. *Bioessays* 23: 261–269.
- Vatter H, Seifert V (2006). Ambrisentan, a non-peptide endothelin receptor antagonist. *Cardiovasc Drug Rev* 24: 63–76.
- Vignon-Zellweger N, Heiden S, Miyauchi T, Emoto N (2012). Endothelin and endothelin receptors in the renal and cardiovascular systems. *Life Sci* 91: 490–500.
- Wang R, Lohr CV, Fischer K, Dashwood WM, Greenwood JA, Ho E *et al.* (2013). Epigenetic inactivation of endothelin-2 and endothelin-3 in colon cancer. *Int J Cancer* 132: 1004–1012.
- Webb DJ (1995). Endogenous endothelin generation maintains vascular tone in humans. *J Hum Hypertens* 9: 459–463.
- Weiss J, Theile D, Ruppell MA, Speck T, Spalwicz A, Haefeli WE (2013). Interaction profile of macitentan, a new non-selective endothelin-1 receptor antagonist, *in vitro*. *Eur J Pharmacol* 701: 168–175.
- Welch AK, Jacobs ME, Wingo CS, Cain BD (2013). Early progress in epigenetic regulation of endothelin pathway genes. *Br J Pharmacol* 168: 327–334.
- Wengenmayer C, Krikov M, Mueller S, Lucht K, Villringer A, Hoher B *et al.* (2011). Novel therapy approach in primary stroke prevention: simultaneous inhibition of endothelin converting enzyme and neutral endopeptidase in spontaneously hypertensive, stroke-prone rats improves survival. *Neurol Res* 33: 201–207.
- William DL, Jones KL, Pettibone DJ, Lis EV, Clineschmidt BV (1991). Sarafotoxin S6c, an agonist which distinguishes between endothelin receptor subtypes. *Biochem Biophys Res Commun* 175: 556–561.

Xu D, Emoto N, Giaid A, Slaughter C, Kaw S, deWit D *et al.* (1994). ECE-1: a membrane-bound metalloprotease that catalyzes the proteolytic activation of big endothelin-1. *Cell* 78: 473–485.

Xu XF, Lv Y, Gu WZ, Tang LL, Wei JK, Zhang LY *et al.* (2013). Epigenetics of hypoxic pulmonary arterial hypertension following intrauterine growth retardation rat: epigenetics in PAH following IUGR. *Respir Res* 14: 20.

Yanagisawa H, Yanagisawa M, Kapur RP, Richardson JA, Williams SC, Clouthier DE *et al.* (1998). Dual genetic pathways of endothelin-mediated intercellular signaling revealed by targeted disruption of endothelin converting enzyme-1 gene. *Development* 125: 825–836.

Yanagisawa H, Hammer RE, Richardson JA, Emoto N, Williams SC, Takeda SI *et al.* (2000). Disruption of ECE-1 and ECE-2 reveals a role for endothelin-converting enzyme-2 in murine cardiac development. *J Clin Invest* 105: 1373–1382.

Yanagisawa M, Inoue A, Ishikawa T, Kasuya Y, Kimura S, Kumagaye S *et al.* (1988). Primary structure, synthesis, and biological activity of rat endothelin, an endothelium-derived vasoconstrictor peptide. *Proc Natl Acad Sci U S A* 85: 6964–6967.

Yap EY, Battistini B, McKay KO (2000). Contraction to big endothelin-1, big endothelin-2 and big endothelin-3, and endothelin-converting enzyme inhibition in human isolated bronchi. *Br J Pharmacol* 129: 170–176.

Yuzgulen J, Wood EG, Villar IC, Douthwaite JA, Patel NSA, Jegard J *et al.* (2013). Characterisation of the 'endothelin-like domain peptide' (ELDP) co-synthesised with endothelin-1 from the *EDN1* gene. *Life Sciences* 93: DOI: 10.1016/j.lfs.2013.12.037.

Zolk O, Quatteck J, Sitzler G, Schrader T, Nickenig G, Schnabel P *et al.* (1999). Expression of endothelin-1, endothelin-converting enzyme, and endothelin receptors in chronic heart failure. *Circulation* 99: 2118–2123.