



# Immune Checkpoint Inhibitors Induced Hepatotoxicity; Gastroenterologists' Perspectives

Maham Farshidpour<sup>1\*</sup>, William Hutson<sup>2</sup>

<sup>1</sup> Loma Linda University Transplantation Institute

<sup>2</sup> West Virginia University - Department of Medicine Section of Gastroenterology & Hepatology

**\* Corresponding Author:**

Maham Farshidpour, MD

Loma Linda University Transplant Institute  
197 E Caroline St, Suite 1400, San Bernardino, CA 92408  
Tel: + 909 558 3636  
Fax: + 909 337 2222  
Email: Mfarshidpour@llu.edu

Received : 01 Jul. 2021  
Accepted : 10 Jan. 2022  
Published: 30 Apr. 2022

## ABSTRACT

### BACKGROUND:

Immune checkpoint inhibitors (ICIs) have promising clinical activity and are essential medications for patients with several malignancies. However, by deranging the immune system, these novel agents could lead to immune-related adverse events (IRAEs). Hepatotoxicity with checkpoint inhibitors usually results in acute hepatitis or drug-induced liver injury.

### METHODS:

This review article discusses the recent clinical evidence available regarding checkpoint inhibitor-induced hepatitis and reviews an approach to their diagnosis and management.

### CONCLUSION:

ICIs have improved patients' outcomes with different forms of malignancy; however, ICIs-related liver damage is a clinically significant entity in these patients. All patients should be monitored carefully for IRAEs while undergoing treatment with ICIs.

### KEYWORDS:

Liver injury, Checkpoint inhibitors, Immune-related adverse events

### Please cite this paper as:

Farshidpour M, Hutson W. Immune Checkpoint Inhibitors Induced Hepatotoxicity; Gastroenterologists' Perspectives. *Middle East J Dig Dis* 2022;14(2):244-253. doi: 10.34172/mejdd.2022.279.

## INTRODUCTION

Some of the immunotherapy drugs called immune checkpoint inhibitors (ICIs) are novel and important medications for patients with a number of different malignancies such as breast cancer, renal cell carcinoma, Hodgkin's lymphoma, and hepatocellular carcinoma.<sup>1,2</sup>

These agents are monoclonal antibodies that specifically target down-regulators of the anti-cancer immune responses.<sup>3</sup> Immune checkpoints are a normal part of the immune system, and they protect healthy cells in the body from being attacked and destroyed by a strong immune response.<sup>4</sup> Most ICIs target three key checkpoints: cytotoxic T-lymphocyte associated protein 4 (CTLA-4; ipilimumab, and tremelimumab), programmed cell death receptor 1 (PD-1; pembrolizumab and nivolumab), and programmed death-ligand 1 (PD-L1; atezolizumab, avelumab, and durvalumab).<sup>5</sup> Despite impressive survival benefits and noticeable improvements in disease outcomes following ICI therapy, its use can be associated with serious adverse events related to excessive immune activation.<sup>6</sup> Importantly, immune checkpoint inhibition can stimulate



© 2022 The Author(s). This work is published by Middle East Journal of Digestive Diseases as an open access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by-nc/4.0/>). Non-commercial uses of the work are permitted, provided the original work is properly cited.

autoreactive T-cells, and this activation can lead to a unique range of toxicities identified as immune-related adverse events (IRAEs).<sup>7</sup> Adverse events include skin rashes, colitis, pancreatitis, nephritis, and hepatitis.<sup>8,9</sup> The gastrointestinal tract appears to be the most commonly affected organ, and IRAEs seem to be characterized by predominant neutrophilic and lymphocytic inflammation.<sup>10</sup> Particularly, the liver is prone to be affected by IRAEs, and the prevalence of ICIs-induced liver injury has been reported to be between 4% and 9% in patients treated with CTLA-4 monoclonal antibodies, and 18% of those treated with a combination of anti-PD-1 and anti-CTLA-4 inhibitors.<sup>11,12</sup> Therefore, addressing hepatic IRAEs has become a significant clinical issue for patients. The primary objectives of this article include reviewing currently available literature regarding ICI-associated acute hepatitis and liver injury while also guiding on novel management and therapeutic interventions that are currently available to manage such complications.

## MATERIALS AND METHODS

Medline/PubMed databases were searched thoroughly with search strategies using search keywords “hepatitis”, “checkpoint inhibitors”, “immune-related adverse events”, “Nivolumab”, “Ipilimumab”, “Pembrolizumab”, “Atezolizumab”, “Avelumab”, “Tremelimumab” and “Durvalumab” to classify studies published between the years 2000 and 2020. All types of related studies, including retrospective, cross-sectional, case reports, and cohorts, were selected. Adults with acute hepatitis or liver injury as a result of checkpoint inhibitor use were included. Studies related to non-humans were excluded from our review. The authors reviewed all selected articles for relevance to checkpoint inhibitor use and hepatotoxicity.

### Epidemiology of Hepatotoxicity

Hepatotoxicity with ICIs is relatively common as the major mechanism of action of ICIs involves the infiltration of immune cells into normal and neoplastic tissues.<sup>13,14</sup>

The rate of liver injury with ICIs varies between different checkpoint inhibitors. It has been reported that the incidence of various grades of autoimmune hepatotoxicity with CTLA-4 inhibitors is between 3%-

9%,<sup>15-17</sup> however, the hepatotoxicity with PD-1 inhibitors was noted to be between 1%-3%.<sup>18</sup> Additionally, the incidence of hepatotoxicity is much higher with combination therapy, with an incidence rate of 13%-30% for all grades and 6%-19% for  $\geq$  grade 3.<sup>1,19,20</sup> De Martin and colleagues observed acute hepatitis grade  $\geq 3$  in 16 patients with an incidence rate of 3.5% when treated with anti-PD-1/PD-L1 and anti-CTLA-4.<sup>21</sup> In a meta-analysis, Wang and colleagues showed that the risk of all and high-grade hepatotoxicity with CTLA-4 inhibitors are higher compared with control regimes. The odds ratio in this study for all-grade hepatotoxicity was 1.24 (95% confidence interval 0.75, 2.05;  $P < 0.39$ ) and the odds ratio for high-grade hepatotoxicity (grade  $\geq 3$ ) was 1.93 (95% confidence interval 0.84, 4.44;  $P < 0.12$ ). In the same study, the risk of all-grade hepatotoxicity and high-grade hepatotoxicity with the use of PD-1 inhibitors appears to be lower, with the odds ratio for all-grade hepatotoxicity noted to be 1.52 (95% confidence interval 1.24, 1.86;  $P < 0.0001$ ) and for high-grade hepatotoxicity was 0.48 (95% confidence interval 0.29, 0.80;  $P = 0.005$ ) (Table 1).<sup>22</sup>

Parlati et al demonstrated in a retrospective study that 23.1% of the patients treated with ICIs developed a predominantly cholestatic pattern of liver injury with an incidence of 60.3%, while hepatocellular liver injury and mixed hepatocellular/cholestatic liver injury occurred in 20 (29.4%) and 7 (10.3%) patients, respectively.<sup>36</sup>

### Mechanisms of Action

Tumor-associated antigens (TAAs) will be expressed by transformed cells, which can be detected by the patient's immune system.<sup>37</sup> During the process of recognition and subsequent elimination of tumor cells, TAAs will be presented by antigen-presenting cells (APCs) alongside the major histocompatibility complex (MHC) I or II that bind with T-cell receptors (TCRs).<sup>38</sup>

The interactions of TCR with peptide/MHC and CD-28 (stimulatory checkpoint expressed on T cells) with B7 (CD-80) present on APCs lead to variations in gene expression, activation of T cell proliferation, and secretion of cytokines.<sup>39</sup> The CTLA-4 (CD152) and PD-1 (CD279) are two important immune checkpoint receptors, which tumor cells may use as a mechanism

**Table 1:** Incidence of immune checkpoint inhibitor-related hepatitis in randomized clinical trials

Author	Medication	Hepatitis Incidence %	Total patient	Cancer
Hodi et al <sup>23</sup>	Ipilimumab	3.8	131	Melanoma
Robert et al <sup>24</sup>	Ipilimumab plus dacarbazine	29.1	247	Melanoma
Kwon et al <sup>25</sup>	Ipilimumab plus radiotherapy	1	393	Prostate
Reck et al <sup>26</sup>	Ipilimumab plus paclitaxel and carboplatin	46.6	84	Small cell carcinoma of lung
Tarhini et al <sup>27</sup>	Tremelimumab plus Interferon Alfa-2b	21.6	37	Melanoma
Ribas et al <sup>28</sup>	Tremelimumab	1	325	Melanoma
Borghaei et al <sup>29</sup>	Nivolumab	9	287	Non squamous carcinoma of lung
Topalian et al <sup>30</sup>	Nivolumab	3.7	107	Melanoma
Postow et al <sup>31</sup>	Nivolumab plus ipilimumab	21	94	Melanoma
Hamanishi et al <sup>32</sup>	Nivolumab	40	20	Ovarian
Wolchok et al <sup>33</sup>	Nivolumab plus ipilimumab	53	21	Melanoma
Rosenberg et al <sup>34</sup>	Atezolizumab	3	310	Urothelial
Petrylak et al <sup>35</sup>	Atezolizumab	4	95	Urothelial

of immune resistance against malignant cells.<sup>40</sup> CTLA-4 is a downregulatory of T cell-mediated anti-tumor responses, and it prevents T cell activation and proliferation.<sup>41</sup> Similarly, the interaction between PD-1 and PD-L1/PD-L2 can lead to T cell inactivation.<sup>42</sup>

The mechanisms of IRAEs are not entirely understood; however, some studies are available to further understand the nature of these events. For example, CTLA-4 blockade eliminates CTLA-4-mediated protection from autoimmunity and may lead to serious and life-threatening clinical manifestations that resemble autoimmune conditions involving all organs and tissues, including the hepatobiliary system.<sup>43</sup> Due to activation of T cells, CD4<sup>+</sup> helper T cells secrete high levels of cytokines, and CD8<sup>+</sup> T cells infiltrate in tissue. Hepatotoxicity from ICI use most often resembles autoimmune hepatitis (AIH), and pathological review of liver biopsy tissue often reveals immune-mediated hepatic injury with focal or confluent necrosis and prominent lymphocytic infiltrate of activated T cells.<sup>9,44,45</sup> Furthermore, ICI-related hepatotoxicity appears to be dose-dependent. Wolchok and colleagues observed no grade 3 to 4 hepatotoxicity with ipilimumab at a dose of 0.3 mg/kg, while ICI-related hepatotoxicity increased to 30% with ipilimumab at a dose of 10 mg/kg in patients treated for

advanced melanoma.<sup>46</sup>

### Histological Features

Histologically, ICI use can induce various forms of pathological injury to hepatocytes, including panlobular hepatitis, perivenular infiltration with endothelialitis or a cholestatic pattern of injury with bile duct proliferation, as well as mixed portal inflammation with mild lobular necroinflammation.<sup>47,48</sup> In a case series study, Zen and colleagues showed the liver injury was largely characterized by lobular hepatitis with infiltration of CD3<sup>+</sup> or CD8<sup>+</sup> T cells in seven patients who were treated with nivolumab or ipilimumab. However, they reported that compared with classic AIH, centrilobular zonal necrosis and plasmacytosis were uncommon.<sup>9</sup> While there were some similarities between ICI-induced hepatotoxicity and AIH on review of the liver histology, there were also differences that indicate that ICIs-related liver toxicity might involve a separate idiosyncratic pattern of injury.<sup>45</sup> For instance, in ICIs-related hepatotoxicity, CD20<sup>+</sup> or CD4<sup>+</sup> lymphocytes are found to be significantly fewer in number, and plasmacytosis and eosinophilic infiltration are less frequently seen in ICIs-related liver injury.<sup>9</sup> Additionally, fibrin ring granulomas have also been reported and considered

as pathognomonic for ipilimumab-induced hepatic injury.<sup>49, 50</sup> Moreover, De Martin and colleagues identified two different histological patterns among 16 patients treated with anti-CTLA-4 or anti-PD-1/PD-L1 agents. They reported that granulomatous hepatitis with fibrin deposits was associated more with anti-CTLA-4 use, whereas lobular with non-granulomatous hepatitis development is related to anti-PD-1/anti-PD-L1 use (Table 2).<sup>21</sup>

Furthermore, Johncilla et al showed a steatosis pattern similar to non-alcoholic fatty liver disease in 2 of 11 patients with ICIs-induced hepatitis.<sup>31</sup> Given this evidence, it appears that performance of a liver biopsy and pathological evaluation of liver tissue is extremely beneficial in distinguishing between ICIs-induced liver injury, AIH, non-alcoholic fatty liver disease, and drug-induced hepatotoxicity; however additional studies are needed to further evaluate for subtle differences in the patterns of hepatic damage.<sup>44</sup>

### Clinical Presentation

The clinical characteristics of ICIs-induced hepatotoxicity are quite heterogeneous but they are usually in line with an autoimmune induced liver injury.<sup>36</sup> Hepatic injury with checkpoint inhibitors typically results in asymptomatic elevations of aspartate aminotransferase (AST) and alanine aminotransferase (ALT) and, less frequently, bilirubin. Some patients may have pain in the right upper quadrant, fever, fatigue, or jaundice; however, many patients may also present without symptoms.<sup>12</sup> The *National Cancer Institute* (NCI) and Common Terminology Criteria for Adverse Events (CTCAE) are typically utilized to categorize the severity of immune-related toxicity grading. It is very important to recognize the toxicity grade of immune-related hepatitis as it assists with the treatment options and also provides guidance regarding the optimal time to resume ICI therapy (Table 3).<sup>15</sup>

The median onset of transaminase elevation depends

**Table 2:** Comparison of the clinicopathologic features of hepatic injury due to ICIs, DILI, and AIH

Feature	ICIs	DILI	AIH
Autoimmune antibody	Absent	ANA, SMA, pANCA	ANA, Anti LKM1, SMA
Histology	PD-1/L1: Lobular, non-granulomatous hepatitis	Cholestasis and bile duct injury	Lymphoplasmacytic interface hepatitis, emperipolesis, and hepatocyte rosettes <sup>52</sup>
	CTLA-4: Central vein endothelialitis, granulomatous hepatitis with fibrin ring deposits <sup>21</sup>	non-caseating granulomas, mild lobular and portal inflammation <sup>51</sup>	
Type of immune cells	Eosinophilic infiltration and plasmacytosis less frequently with significantly fewer CD20 <sup>+</sup> or CD4 <sup>+</sup> lymphocytes	Prominent intra-acinar lymphocyte Prominent port neutrophils <sup>53</sup>	Prominent intra-acinar plasma cell and eosinophils <sup>53</sup>

Abbreviations: ICI, Immune checkpoint inhibitors; DILI, Drug-induced liver injury; AIH, Autoimmune hepatitis; ANA, Antinuclear antibodies; SMA, Smooth muscle antibodies; pANCA, Perinuclear antineutrophil cytoplasmic antibodies; LKM1, Liver kidney microsomal type 1.

**Table 3:** Common Terminology Criteria for Adverse Events (CTCAE), version 5<sup>15</sup>

Grading	Grade 1	Grade 2	Grade 3	Grade 4	Grade 5
Hepatitis	AST/ALT < 3x ULN and/or total bilirubin < 1.5x ULN	AST/ALT 3-5x ULN and/or total bilirubin 1.5-3x ULN	AST/ALT 5-20x ULN and/or total bilirubin 3-10x ULN	Decompensated liver function, AST/ALT > 20x ULN and/or total bilirubin > 10x ULN	Death

Abbreviations: ULN, Upper limit of normal; AST, aspartate aminotransferase; ALT, alanine aminotransferase.

on the type of ICI used and the primary malignancy being treated, but it has been reported that the elevation typically occurs 6-14 weeks following the initiation of the ICI treatment.<sup>54,55</sup> For example, the median onset of hepatitis following nivolumab treatment in lung cancer was reported to be around 25 weeks, but was 4 weeks for melanoma.<sup>56</sup> Hepatitis B virus (HBV) reactivation, which is defined as a reappearance of HBV DNA in patients with previously inactive HBV, is a serious complication in cancer patients who are undergoing immunosuppressive treatment or chemotherapy.<sup>57,58</sup>

### ICI Treatment in Chronic Viral Hepatitis

Patients with cancer who have chronic hepatitis B (hepatitis B surface antigen-positive) or hepatitis C (hepatitis C virus RNA positive) infections are always at risk for exacerbation of viral hepatitis in the setting of immunosuppression, and the impact of checkpoint inhibitors on these chronic viral infections is poorly understood.<sup>59</sup> Zhang and colleagues showed that in 114 patients with HBsAg-positive status who were treated with anti-PD-1/PD-L1 agents, the incidence of reactivation of HBV was 5.3%. The inadequacy of prophylactic antiviral therapy was the most significant risk factor, with an odds ratio of 17.50. They recommend screening of all patients for HBV before initiation of anti-PD-1/PD-L1 therapy and initiation of prophylactic antiviral treatment for those who are seropositive (HBsAg positive), regardless of baseline HBV DNA level.<sup>60</sup> Additional considerations regarding HBsAg seroconversion do not appear to be discussed in great detail in the currently available literature; however, they may warrant future investigations in the management of patients with HBsAg positivity receiving ICI treatment.

Conversely, Shah and colleagues noted in a retrospective analysis of patients who were infected with human immunodeficiency virus (HIV), HBV, or HCV treated with ICI therapy that there was no evidence of viral reactivation.<sup>61</sup> Additionally, previous trials proved no cases of HCV-related flares in cancer patients with positive serology for HCV who had undergone treatment with ICIs.<sup>62,63</sup>

### Prognosis and Mortality of ICIs-Induced Hepatitis

ICIs-induced hepatitis typically improves after

treatment with corticosteroids. The time to resolution is about 8 weeks, and relapses are frequent as corticosteroids are tapered.<sup>64</sup> In the VigiLyza database of 333 anti-PD1/PD-L1 fatalities, 74 deaths (22%) were due to hepatitis. Ipilimumab use in 193 patients resulted in fatality secondary to hepatitis in roughly 16% of the treated patients.<sup>65</sup> Wang and colleagues discovered a fatality rate of 0.04% due to hepatitis among 19,127 patients who were treated with ICIs.<sup>66</sup>

### Management of ICIs-Induced Hepatitis

It was recommended to check baseline liver enzymes prior to ICI infusion, followed by monitoring with serial serological surveillance.<sup>67-69</sup> Importantly, despite known ICIs-induced hepatitis, patients should be assessed for other etiologies of hepatitis to exclude viral hepatitis, autoimmune, and drug-related etiologies as well as rhabdomyolysis. Tsung and others showed that of 491 patients treated with pembrolizumab, only a minority of the liver injury cases were related to pembrolizumab-induced hepatotoxicity. The majority of hepatitis was attributed to malignancy-related biliary strictures or cholestasis.<sup>70</sup> Most cases of hepatitis-related to IRAEs may respond well to supportive care and temporary interruption of ICI use.<sup>71</sup> Generally, the management of ICIs-induced liver injury depends on the grade of liver injury.<sup>72</sup> Grade 1 hepatitis can be managed with close monitoring of AST, ALT, and bilirubin levels while the patient continues treatment with the ICI. The ICI should be discontinued if a patient develops grade 2 hepatitis and held until hepatitis improves to a grade 1 level or resolves completely. It is advised to initiate prednisone or an equivalent corticosteroid at 0.5–1 mg/kg/day if the liver enzymes fail to improve or rise upon repeat testing with temporary cessation of the agent. Grade 3 hepatitis or greater can be managed with permanent discontinuation of ICIs, and in those patients who do not respond adequately to corticosteroids and develop worsening hepatitis or progression of their hepatitis to liver injury, the use of an immunosuppressant including azathioprine (1–2 mg/kg), mycophenolate mofetil (500–1,000 mg twice per day), or tacrolimus with serum trough levels targeting 8–10 ng/mL, should be considered (Table 4).<sup>59,69,73</sup>

Discontinuation of ICIs in grade 3 hepatitis might



**Table 4:** Management algorithms for checkpoint inhibitors hepatitis

ASCO Recommendations <sup>74</sup>	
Grade 1 (G1)	<ul style="list-style-type: none"> <li>• Continue ICIs with close monitor</li> <li>• Check LFT two times weekly</li> <li>• Supportive care</li> </ul>
Grade 2 (G2)	<ul style="list-style-type: none"> <li>• Hold ICIs temporarily and resume if recover to G1 or less</li> <li>• Start corticosteroid 0.5-1 mg/kg/day prednisone or equivalent if the abnormal LFT elevation persists</li> <li>• Monitoring LFT to every 3 days</li> <li>• May resume ICIs treatment followed by steroid taper over 4 weeks once LFT improves to G1 on corticosteroid 10 mg/day</li> </ul>
Grade 3 (G3)	<ul style="list-style-type: none"> <li>• Discontinue ICIs permanently</li> <li>• Start intravenous corticosteroid 1-2 mg/kg methylprednisolone or equivalent</li> <li>• Start mycophenolate mofetil or azathioprine, If no improvement after 3 days</li> <li>• Daily or every other day LFT</li> <li>• Consider inpatient treatment</li> <li>• Corticosteroid taper around 4-6 weeks once LFT improves</li> </ul>
Grade 4 (G4)	<ul style="list-style-type: none"> <li>• Discontinue ICIs permanently</li> <li>• Administer IV 2 mg/kg/day methylprednisolone equivalents</li> <li>• Start mycophenolate mofetil, If corticosteroid refractory or no improvement after 3 days</li> <li>• Monitor LFT daily</li> <li>• Consider inpatient monitoring</li> <li>• Corticosteroid taper over 4-6 weeks when symptoms improve to G1 or less</li> <li>• Consider transfer to a tertiary care facility with hepatology consult if necessary</li> </ul>

Abbreviations: ASCO, American Society of Clinical Oncology; LFT, liver function test; ICIs, Immune checkpoint inhibitors.

be challenging for patients who may benefit strongly from an oncological perspective from ICI therapy, particularly in those patients who may have limited chemotherapeutic options.<sup>74</sup> On the other hand, the safety and benefit of retreatment with ICIs after recovery from an IRAE is unknown.<sup>75</sup> Santini and others showed that 19 out of 39 patients with advanced non-small cell lung cancer who were managed with anti-PD-L1 and their course of treatment was complicated with IRAE, developed recurrent IRAE following administration of ICI therapy. Therefore, they recommended those patients who needed to be admitted in the hospital for an initial IRAE, and those who had already accomplished a complete or partial response before an initial IRAE, not be retreated.<sup>76</sup> Based on which ICI was used, the response of the primary malignant cells to the ICI, and the response of liver enzymes following ICI discontinuation, the physician would be able to weigh the risks versus benefits of restarting ICI therapy.<sup>77</sup> Other treatments for ICIs-induced hepatitis have been explored. For example, the tumor necrosis factor-

alpha (TNF- $\alpha$ ) blocker infliximab is not recommended for the treatment of ICIs-induced hepatitis, given the concern of liver toxicity.<sup>69</sup> Additionally, Chmiel and colleagues reported a case of severe steroid-resistant fulminant hepatitis induced by ipilimumab that resolved with anti-thymocyte globulin. They showed that the elevation of hepatic transaminases improved significantly within 24 hours after 1.5 mg/kg of anti-thymocyte globulin for 2 consecutive days.<sup>78</sup>

Riveiro-Barciela et al described the effective use of plasma exchange (PE) (1500 mL of 5% albumin plus four units of plasma as replacement fluid) to treat fulminant hepatitis related to ipilimumab use in patients with melanoma. They showed that PE could help to remove ipilimumab since this molecule had some ideal target characteristics, such as a low volume of distribution (0.1 L/kg) and high molecular weight (148 000 Da).<sup>79</sup>

## CONCLUSION

ICIs have improved patients' outcomes with different

forms of malignancy; however, ICIs-related liver damage is a clinically significant entity in these groups of patients.<sup>80</sup> All patients should be monitored carefully for IRAEs while they are undergoing treatment with ICIs. Prompt recognition of hepatitis is important to ensure that proper treatment is started promptly. The incidence of hepatotoxicity depends on the type and dosage of agents and appears to be more severe with CTLA-4 and PD-1 combination therapy. Management of severe ICIs-related hepatitis should consist of termination of the ICI and treatment with corticosteroids. The management should be escalated to other immunosuppressive agents for those patients who do not demonstrate a significant response to corticosteroids. Patients with severe or corticosteroid-refractory hepatitis will benefit from collaboration between hepatology and oncology care teams to determine appropriate courses of action on an individualized basis for each patient.

In summary, the rates of ICIs-induced hepatitis, according to several trials are appeared to be less frequent compared with the most common IRAEs; however, physicians need to uphold a low threshold for evaluation and treating suspected immune hepatitis as delays can lead to permanent discontinuation of cancer therapy.

#### ETHICAL APPROVAL

There is nothing to be declared.

#### CONFLICT OF INTEREST

The authors declare no conflict of interest related to this work.

#### REFERENCES

- Grover S, Rahma OE, Hashemi N, Lim RM. Gastrointestinal and hepatic toxicities of checkpoint inhibitors: algorithms for management. *Am Soc Clin Oncol Educ Book* 2018;38:13-9. doi: [10.1200/edbk\\_100013](https://doi.org/10.1200/edbk_100013)
- Wu X, Gu Z, Chen Y, Chen B, Chen W, Weng L, et al. Application of PD-1 blockade in cancer immunotherapy. *Comput Struct Biotechnol J* 2019;17:661-74. doi: [10.1016/j.csbj.2019.03.006](https://doi.org/10.1016/j.csbj.2019.03.006)
- Som A, Mandalia R, Alsaadi D, Farshidpour M, Charabaty A, Malhotra N, et al. Immune checkpoint inhibitor-induced colitis: a comprehensive review. *World J Clin Cases* 2019;7(4):405-18. doi: [10.12998/wjcc.v7.i4.405](https://doi.org/10.12998/wjcc.v7.i4.405)
- Azoury SC, Straughan DM, Shukla V. Immune checkpoint inhibitors for cancer therapy: clinical efficacy and safety. *Curr Cancer Drug Targets* 2015;15(6):452-62. doi: [10.2174/156800961506150805145120](https://doi.org/10.2174/156800961506150805145120)
- Seidel JA, Otsuka A, Kabashima K. Anti-PD-1 and anti-CTLA-4 therapies in cancer: mechanisms of action, efficacy, and limitations. *Front Oncol* 2018;8:86. doi: [10.3389/fonc.2018.00086](https://doi.org/10.3389/fonc.2018.00086)
- Rocha M, Correia de Sousa J, Salgado M, Araújo A, Pedroto I. Management of gastrointestinal toxicity from immune checkpoint inhibitor. *GE Port J Gastroenterol* 2019;26(4):268-74. doi: [10.1159/000494569](https://doi.org/10.1159/000494569)
- Trinh S, Le A, Gowani S, La-Beck NM. Management of immune-related adverse events associated with immune checkpoint inhibitor therapy: a minireview of current clinical guidelines. *Asia Pac J Oncol Nurs* 2019;6(2):154-60. doi: [10.4103/apjon.apjon\\_3\\_19](https://doi.org/10.4103/apjon.apjon_3_19)
- Abdel-Wahab N, Shah M, Suarez-Almazor ME. Adverse events associated with immune checkpoint blockade in patients with cancer: a systematic review of case reports. *PLoS One* 2016;11(7):e0160221. doi: [10.1371/journal.pone.0160221](https://doi.org/10.1371/journal.pone.0160221)
- Zen Y, Yeh MM. Hepatotoxicity of immune checkpoint inhibitors: a histology study of seven cases in comparison with autoimmune hepatitis and idiosyncratic drug-induced liver injury. *Mod Pathol* 2018;31(6):965-73. doi: [10.1038/s41379-018-0013-y](https://doi.org/10.1038/s41379-018-0013-y)
- Everett J, Srivastava A, Misdraji J. Fibrin ring granulomas in checkpoint inhibitor-induced hepatitis. *Am J Surg Pathol* 2017;41(1):134-7. doi: [10.1097/pas.0000000000000759](https://doi.org/10.1097/pas.0000000000000759)
- Michot JM, Bigenwald C, Champiat S, Collins M, Carbonnel F, Postel-Vinay S, et al. Immune-related adverse events with immune checkpoint blockade: a comprehensive review. *Eur J Cancer* 2016;54:139-48. doi: [10.1016/j.ejca.2015.11.016](https://doi.org/10.1016/j.ejca.2015.11.016)
- Lombardi A, Mondelli MU. Review article: immune checkpoint inhibitors and the liver, from therapeutic efficacy to side effects. *Aliment Pharmacol Ther* 2019;50(8):872-84. doi: [10.1111/apt.15449](https://doi.org/10.1111/apt.15449)
- Kawakami H, Tanizaki J, Tanaka K, Haratani K, Hayashi H, Takeda M, et al. Imaging and clinicopathological features of nivolumab-related cholangitis in patients with non-small cell lung cancer. *Invest New Drugs* 2017;35(4):529-36. doi: [10.1007/s10637-017-0453-0](https://doi.org/10.1007/s10637-017-0453-0)
- Mathew Thomas V, Bindal P, Ann Alexander S, McDonald K. Nivolumab-induced hepatitis: a rare side effect of an immune check point inhibitor. *J Oncol Pharm Pract* 2020;26(2):459-61. doi: [10.1177/1078155219837342](https://doi.org/10.1177/1078155219837342)
- Reddy HG, Schneider BJ, Tai AW. Immune checkpoint inhibitor-associated colitis and hepatitis. *Clin Transl Gastroenterol* 2018;9(9):180. doi: [10.1038/s41424-018-0049-9](https://doi.org/10.1038/s41424-018-0049-9)
- Weber JS, Kähler KC, Hauschild A. Management of immune-related adverse events and kinetics of response with ipilimumab. *J Clin Oncol* 2012;30(21):2691-7. doi: [10.1200/jco.2012.41.6750](https://doi.org/10.1200/jco.2012.41.6750)
- Sanjeevaiah A, Kerr T, Beg MS. Approach and management of checkpoint inhibitor-related immune hepatitis. *J Gastrointest Oncol* 2018;9(1):220-4. doi: [10.2174/156800961506150805145120](https://doi.org/10.2174/156800961506150805145120)

- 10.21037/jgo.2017.08.14
18. Robert C, Ribas A, Wolchok JD, Hodi FS, Hamid O, Kefford R, et al. Anti-programmed-death-receptor-1 treatment with pembrolizumab in ipilimumab-refractory advanced melanoma: a randomised dose-comparison cohort of a phase 1 trial. *Lancet* 2014;384(9948):1109-17. doi: [10.1016/s0140-6736\(14\)60958-2](https://doi.org/10.1016/s0140-6736(14)60958-2)
  19. Maughan BL, Bailey E, Gill DM, Agarwal N. Incidence of immune-related adverse events with program death receptor-1- and program death receptor-1 ligand-directed therapies in genitourinary cancers. *Front Oncol* 2017;7:56. doi: [10.3389/fonc.2017.00056](https://doi.org/10.3389/fonc.2017.00056)
  20. Robert C, Schachter J, Long GV, Arance A, Grob JJ, Mortier L, et al. Pembrolizumab versus ipilimumab in advanced melanoma. *N Engl J Med* 2015;372(26):2521-32. doi: [10.1056/NEJMoa1503093](https://doi.org/10.1056/NEJMoa1503093)
  21. De Martin E, Michot JM, Papouin B, Champiat S, Mateus C, Lambotte O, et al. Characterization of liver injury induced by cancer immunotherapy using immune checkpoint inhibitors. *J Hepatol* 2018;68(6):1181-90. doi: [10.1016/j.jhep.2018.01.033](https://doi.org/10.1016/j.jhep.2018.01.033)
  22. Wang W, Lie P, Guo M, He J. Risk of hepatotoxicity in cancer patients treated with immune checkpoint inhibitors: a systematic review and meta-analysis of published data. *Int J Cancer* 2017;141(5):1018-28. doi: [10.1002/ijc.30678](https://doi.org/10.1002/ijc.30678)
  23. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 2010;363(8):711-23. doi: [10.1056/NEJMoa1003466](https://doi.org/10.1056/NEJMoa1003466)
  24. Robert C, Thomas L, Bondarenko I, O'Day S, Weber J, Garbe C, et al. Ipilimumab plus dacarbazine for previously untreated metastatic melanoma. *N Engl J Med* 2011;364(26):2517-26. doi: [10.1056/NEJMoa1104621](https://doi.org/10.1056/NEJMoa1104621)
  25. Kwon ED, Drake CG, Scher HI, Fizazi K, Bossi A, van den Eertwegh AJ, et al. Ipilimumab versus placebo after radiotherapy in patients with metastatic castration-resistant prostate cancer that had progressed after docetaxel chemotherapy (CA184-043): a multicentre, randomised, double-blind, phase 3 trial. *Lancet Oncol* 2014;15(7):700-12. doi: [10.1016/s1470-2045\(14\)70189-5](https://doi.org/10.1016/s1470-2045(14)70189-5)
  26. Reck M, Bondarenko I, Luft A, Serwatowski P, Barlesi F, Chacko R, et al. Ipilimumab in combination with paclitaxel and carboplatin as first-line therapy in extensive-disease-small-cell lung cancer: results from a randomized, double-blind, multicenter phase 2 trial. *Ann Oncol* 2013;24(1):75-83. doi: [10.1093/annonc/mds213](https://doi.org/10.1093/annonc/mds213)
  27. Tarhini AA, Cheria J, Moschos SJ, Tawbi HA, Shuai Y, Gooding WE, et al. Safety and efficacy of combination immunotherapy with interferon alfa-2b and tremelimumab in patients with stage IV melanoma. *J Clin Oncol* 2012;30(3):322-8. doi: [10.1200/jco.2011.37.5394](https://doi.org/10.1200/jco.2011.37.5394)
  28. Ribas A, Kefford R, Marshall MA, Punt CJ, Haanen JB, Marmol M, et al. Phase III randomized clinical trial comparing tremelimumab with standard-of-care chemotherapy in patients with advanced melanoma. *J Clin Oncol* 2013;31(5):616-22. doi: [10.1200/jco.2012.44.6112](https://doi.org/10.1200/jco.2012.44.6112)
  29. Borghaei H, Paz-Ares L, Horn L, Spigel DR, Steins M, Ready NE, et al. Nivolumab versus docetaxel in advanced nonsquamous non-small-cell lung cancer. *N Engl J Med* 2015;373(17):1627-39. doi: [10.1056/NEJMoa1507643](https://doi.org/10.1056/NEJMoa1507643)
  30. Topalian SL, Sznol M, McDermott DF, Kluger HM, Carvajal RD, Sharfman WH, et al. Survival, durable tumor remission, and long-term safety in patients with advanced melanoma receiving nivolumab. *J Clin Oncol* 2014;32(10):1020-30. doi: [10.1200/jco.2013.53.0105](https://doi.org/10.1200/jco.2013.53.0105)
  31. Postow MA, Chesney J, Pavlick AC, Robert C, Grossmann K, McDermott D, et al. Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. *N Engl J Med* 2015;372(21):2006-17. doi: [10.1056/NEJMoa1414428](https://doi.org/10.1056/NEJMoa1414428)
  32. Hamanishi J, Mandai M, Ikeda T, Minami M, Kawaguchi A, Murayama T, et al. Safety and antitumor activity of anti-PD-1 antibody, nivolumab, in patients with platinum-resistant ovarian cancer. *J Clin Oncol* 2015;33(34):4015-22. doi: [10.1200/jco.2015.62.3397](https://doi.org/10.1200/jco.2015.62.3397)
  33. Wolchok JD, Kluger H, Callahan MK, Postow MA, Rizvi NA, Lesokhin AM, et al. Nivolumab plus ipilimumab in advanced melanoma. *N Engl J Med* 2013;369(2):122-33. doi: [10.1056/NEJMoa1302369](https://doi.org/10.1056/NEJMoa1302369)
  34. Rosenberg JE, Hoffman-Censits J, Powles T, van der Heijden MS, Balar AV, Necchi A, et al. Atezolizumab in patients with locally advanced and metastatic urothelial carcinoma who have progressed following treatment with platinum-based chemotherapy: a single-arm, multicentre, phase 2 trial. *Lancet* 2016;387(10031):1909-20. doi: [10.1016/s0140-6736\(16\)00561-4](https://doi.org/10.1016/s0140-6736(16)00561-4)
  35. Petrylak DP, Powles T, Bellmunt J, Braith F, Loriot Y, Morales-Barrera R, et al. Atezolizumab (MPDL3280A) monotherapy for patients with metastatic urothelial cancer: long-term outcomes from a phase 1 study. *JAMA Oncol* 2018;4(4):537-44. doi: [10.1001/jamaoncol.2017.5440](https://doi.org/10.1001/jamaoncol.2017.5440)
  36. Parlati L, Vallet-Pichard A, Batista R, Hernvann A, Sogni P, Pol S, et al. Incidence of grade 3-4 liver injury under immune checkpoints inhibitors: a retrospective study. *J Hepatol* 2018;69(6):1396-7. doi: [10.1016/j.jhep.2018.08.014](https://doi.org/10.1016/j.jhep.2018.08.014)
  37. Rivoltini L, Carrabba M, Huber V, Castelli C, Novellino L, Dalerba P, et al. Immunity to cancer: attack and escape in T lymphocyte-tumor cell interaction. *Immunol Rev* 2002;188:97-113. doi: [10.1034/j.1600-065x.2002.18809.x](https://doi.org/10.1034/j.1600-065x.2002.18809.x)
  38. Melero I, Hervas-Stubbs S, Glennie M, Pardoll DM, Chen L. Immunostimulatory monoclonal antibodies for cancer therapy. *Nat Rev Cancer* 2007;7(2):95-106. doi: [10.1038/nrc2051](https://doi.org/10.1038/nrc2051)
  39. Nagorsen D, Scheibenbogen C, Marincola FM, Letsch A, Keilholz U. Natural T cell immunity against cancer. *Clin Cancer Res* 2003;9(12):4296-303.
  40. Tarhini A. Immune-mediated adverse events associated with ipilimumab ctla-4 blockade therapy: the underlying



- mechanisms and clinical management. *Scientifica (Cairo)* 2013;2013:857519. doi: [10.1155/2013/857519](https://doi.org/10.1155/2013/857519)
41. Engelhardt JJ, Sullivan TJ, Allison JP. CTLA-4 overexpression inhibits T cell responses through a CD28-B7-dependent mechanism. *J Immunol* 2006;177(2):1052-61. doi: [10.4049/jimmunol.177.2.1052](https://doi.org/10.4049/jimmunol.177.2.1052)
  42. Wang W, Lau R, Yu D, Zhu W, Korman A, Weber J. PD1 blockade reverses the suppression of melanoma antigen-specific CTL by CD4+ CD25(Hi) regulatory T cells. *Int Immunol* 2009;21(9):1065-77. doi: [10.1093/intimm/dxp072](https://doi.org/10.1093/intimm/dxp072)
  43. Della Vittoria Scarpati G, Fuscicello C, Perri F, Sabbatino F, Ferrone S, Carlomagno C, et al. Ipilimumab in the treatment of metastatic melanoma: management of adverse events. *Onco Targets Ther* 2014;7:203-9. doi: [10.2147/ott.s57335](https://doi.org/10.2147/ott.s57335)
  44. Johncilla M, Misraji J, Pratt DS, Agoston AT, Lauwers GY, Srivastava A, et al. Ipilimumab-associated hepatitis: clinicopathologic characterization in a series of 11 cases. *Am J Surg Pathol* 2015;39(8):1075-84. doi: [10.1097/pas.0000000000000453](https://doi.org/10.1097/pas.0000000000000453)
  45. Shah P, Sundaram V, Björnsson E. Biologic and checkpoint inhibitor-induced liver injury: a systematic literature review. *Hepatol Commun* 2020;4(2):172-84. doi: [10.1002/hep4.1465](https://doi.org/10.1002/hep4.1465)
  46. Wolchok JD, Neyns B, Linette G, Negrier S, Lutzky J, Thomas L, et al. Ipilimumab monotherapy in patients with pretreated advanced melanoma: a randomised, double-blind, multicentre, phase 2, dose-ranging study. *Lancet Oncol* 2010;11(2):155-64. doi: [10.1016/s1470-2045\(09\)70334-1](https://doi.org/10.1016/s1470-2045(09)70334-1)
  47. Kim KW, Ramaiya NH, Krajewski KM, Jagannathan JP, Tirumani SH, Srivastava A, et al. Ipilimumab associated hepatitis: imaging and clinicopathologic findings. *Invest New Drugs* 2013;31(4):1071-7. doi: [10.1007/s10637-013-9939-6](https://doi.org/10.1007/s10637-013-9939-6)
  48. Ibraheim H, Perucha E, Powell N. Pathology of immune-mediated tissue lesions following treatment with immune checkpoint inhibitors. *Rheumatology (Oxford)* 2019;58(Suppl 7):vii17-vii28. doi: [10.1093/rheumatology/kez465](https://doi.org/10.1093/rheumatology/kez465)
  49. Brahmer JR, Govindan R, Anders RA, Antonia SJ, Sagorsky S, Davies MJ, et al. The Society for Immunotherapy of Cancer consensus statement on immunotherapy for the treatment of non-small cell lung cancer (NSCLC). *J Immunother Cancer* 2018;6(1):75. doi: [10.1186/s40425-018-0382-2](https://doi.org/10.1186/s40425-018-0382-2)
  50. Kleiner DE, Berman D. Pathologic changes in ipilimumab-related hepatitis in patients with metastatic melanoma. *Dig Dis Sci* 2012;57(8):2233-40. doi: [10.1007/s10620-012-2140-5](https://doi.org/10.1007/s10620-012-2140-5)
  51. Da BL, Ben-Yakov G, Kleiner D, Koh C. Drug-induced liver injury: understanding the different immune-mediated phenotypes and clinical management. *Curr Hepatol Rep* 2018;17(3):235-44. doi: [10.1007/s11901-018-0407-9](https://doi.org/10.1007/s11901-018-0407-9)
  52. Gurung A, Assis DN, McCarty TR, Mitchell KA, Boyer JL, Jain D. Histologic features of autoimmune hepatitis: a critical appraisal. *Hum Pathol* 2018;82:51-60. doi: [10.1016/j.humpath.2018.07.014](https://doi.org/10.1016/j.humpath.2018.07.014)
  53. Suzuki A, Brunt EM, Kleiner DE, Miquel R, Smyrk TC, Andrade RJ, et al. The use of liver biopsy evaluation in discrimination of idiopathic autoimmune hepatitis versus drug-induced liver injury. *Hepatology* 2011;54(3):931-9. doi: [10.1002/hep.24481](https://doi.org/10.1002/hep.24481)
  54. Spain L, Diem S, Larkin J. Management of toxicities of immune checkpoint inhibitors. *Cancer Treat Rev* 2016;44:51-60. doi: [10.1016/j.ctrv.2016.02.001](https://doi.org/10.1016/j.ctrv.2016.02.001)
  55. Horvat TZ, Adel NG, Dang TO, Momtaz P, Postow MA, Callahan MK, et al. Immune-related adverse events, need for systemic immunosuppression, and effects on survival and time to treatment failure in patients with melanoma treated with ipilimumab at Memorial Sloan Kettering Cancer Center. *J Clin Oncol* 2015;33(28):3193-8. doi: [10.1200/jco.2015.60.8448](https://doi.org/10.1200/jco.2015.60.8448)
  56. Pollack MH, Betof A, Dearden H, Rapazzo K, Valentine I, Brohl AS, et al. Safety of resuming anti-PD-1 in patients with immune-related adverse events (irAEs) during combined anti-CTLA-4 and anti-PD1 in metastatic melanoma. *Ann Oncol* 2018;29(1):250-5. doi: [10.1093/annonc/mdx642](https://doi.org/10.1093/annonc/mdx642)
  57. Kawsar HI, Shahnewaz J, Gopalakrishna KV, Spiro TP, Daw HA. Hepatitis B reactivation in cancer patients: role of prechemotherapy screening and antiviral prophylaxis. *Clin Adv Hematol Oncol* 2012;10(6):370-8.
  58. Hoofnagle JH. Reactivation of hepatitis B. *Hepatology* 2009;49(5 Suppl):S156-65. doi: [10.1002/hep.22945](https://doi.org/10.1002/hep.22945)
  59. Reynolds K, Thomas M, Dougan M. Diagnosis and management of hepatitis in patients on checkpoint blockade. *Oncologist* 2018;23(9):991-7. doi: [10.1634/theoncologist.2018-0174](https://doi.org/10.1634/theoncologist.2018-0174)
  60. Zhang X, Zhou Y, Chen C, Fang W, Cai X, Zhang X, et al. Hepatitis B virus reactivation in cancer patients with positive hepatitis B surface antigen undergoing PD-1 inhibition. *J Immunother Cancer* 2019;7(1):322. doi: [10.1186/s40425-019-0808-5](https://doi.org/10.1186/s40425-019-0808-5)
  61. Shah NJ, Ghassan AS, Blackburn M, Cook M, Kelly WJ, Belouali A, et al. Abstract 3230: Safety and efficacy of immune checkpoint inhibitors (ICIs) in patients with HIV, hepatitis B, or hepatitis C viral infections. 2019. doi: [10.1158/1538-7445.am2019-3230](https://doi.org/10.1158/1538-7445.am2019-3230)
  62. El-Khoueiry AB, Sangro B, Yau T, Crocenzi TS, Kudo M, Hsu C, et al. Nivolumab in patients with advanced hepatocellular carcinoma (CheckMate 040): an open-label, non-comparative, phase 1/2 dose escalation and expansion trial. *Lancet* 2017;389(10088):2492-502. doi: [10.1016/s0140-6736\(17\)31046-2](https://doi.org/10.1016/s0140-6736(17)31046-2)
  63. Zhu AX, Finn RS, Edeline J, Cattani S, Ogasawara S, Palmer D, et al. Pembrolizumab in patients with advanced hepatocellular carcinoma previously treated with sorafenib (KEYNOTE-224): a non-randomised, open-label phase 2 trial. *Lancet Oncol* 2018;19(7):940-52. doi: [10.1016/s1470-2045\(18\)30351-6](https://doi.org/10.1016/s1470-2045(18)30351-6)
  64. Connolly C, Bambhania K, Naidoo J. Immune-related adverse events: a case-based approach. *Front Oncol* 2019;9:530. doi: [10.3389/fonc.2019.00530](https://doi.org/10.3389/fonc.2019.00530)

65. Lindquist M, Edwards IR. The WHO Programme for International Drug Monitoring, its database, and the technical support of the Uppsala Monitoring Center. *J Rheumatol* 2001;28(5):1180-7.
66. Wang DY, Salem JE, Cohen JV, Chandra S, Menzer C, Ye F, et al. Fatal toxic effects associated with immune checkpoint inhibitors: a systematic review and meta-analysis. *JAMA Oncol* 2018;4(12):1721-8. doi: [10.1001/jamaoncol.2018.3923](https://doi.org/10.1001/jamaoncol.2018.3923)
67. Jennings JJ, Mandaliya R, Nakshabandi A, Lewis JH. Hepatotoxicity induced by immune checkpoint inhibitors: a comprehensive review including current and alternative management strategies. *Expert Opin Drug Metab Toxicol* 2019;15(3):231-44. doi: [10.1080/17425255.2019.1574744](https://doi.org/10.1080/17425255.2019.1574744)
68. Haanen J, Carbone F, Robert C, Kerr KM, Peters S, Larkin J, et al. Management of toxicities from immunotherapy: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 2017;28(Suppl\_4):iv119-iv42. doi: [10.1093/annonc/mdx225](https://doi.org/10.1093/annonc/mdx225)
69. Brahmer JR, Lacchetti C, Schneider BJ, Atkins MB, Brassil KJ, Caterino JM, et al. Management of immune-related adverse events in patients treated with immune checkpoint inhibitor therapy: American Society of Clinical Oncology Clinical Practice Guideline. *J Clin Oncol* 2018;36(17):1714-68. doi: [10.1200/jco.2017.77.6385](https://doi.org/10.1200/jco.2017.77.6385)
70. Tsung I, Dolan R, Lao CD, Fecher L, Riggenbach K, Yeboah-Korang A, et al. Liver injury is most commonly due to hepatic metastases rather than drug hepatotoxicity during pembrolizumab immunotherapy. *Aliment Pharmacol Ther* 2019;50(7):800-8. doi: [10.1111/apt.15413](https://doi.org/10.1111/apt.15413)
71. Rudzki JD. Management of adverse events related to checkpoint inhibition therapy. *Memo* 2018;11(2):132-7. doi: [10.1007/s12254-018-0416-y](https://doi.org/10.1007/s12254-018-0416-y)
72. Suzman DL, Pelosof L, Rosenberg A, Avigan MI. Hepatotoxicity of immune checkpoint inhibitors: an evolving picture of risk associated with a vital class of immunotherapy agents. *Liver Int* 2018;38(6):976-87. doi: [10.1111/liv.13746](https://doi.org/10.1111/liv.13746)
73. Mancini S, Amorotti E, Vecchio S, Ponz de Leon M, Roncucci L. Infliximab-related hepatitis: discussion of a case and review of the literature. *Intern Emerg Med* 2010;5(3):193-200. doi: [10.1007/s11739-009-0342-4](https://doi.org/10.1007/s11739-009-0342-4)
74. Wood LS, Moldawer NP, Lewis C. Immune checkpoint inhibitor therapy: key principles when educating patients. *Clin J Oncol Nurs* 2019;23(3):271-80. doi: [10.1188/19.cjon.271-280](https://doi.org/10.1188/19.cjon.271-280)
75. Remon J, Mezquita L, Corral J, Vilariño N, Reguart N. Immune-related adverse events with immune checkpoint inhibitors in thoracic malignancies: focusing on non-small cell lung cancer patients. *J Thorac Dis* 2018;10(Suppl 13):S1516-S33. doi: [10.21037/jtd.2017.12.52](https://doi.org/10.21037/jtd.2017.12.52)
76. Santini FC, Rizvi H, Plodkowski AJ, Ni A, Lacouture ME, Gambarin-Gelwan M, et al. Safety and efficacy of re-treating with immunotherapy after immune-related adverse events in patients with NSCLC. *Cancer Immunol Res* 2018;6(9):1093-9. doi: [10.1158/2326-6066.cir-17-0755](https://doi.org/10.1158/2326-6066.cir-17-0755)
77. Puzanov I, Diab A, Abdallah K, Bingham CO, 3rd, Brogdon C, Dadu R, et al. Managing toxicities associated with immune checkpoint inhibitors: consensus recommendations from the Society for Immunotherapy of Cancer (SITC) Toxicity Management Working Group. *J Immunother Cancer* 2017;5(1):95. doi: [10.1186/s40425-017-0300-z](https://doi.org/10.1186/s40425-017-0300-z)
78. Chmiel KD, Suan D, Liddle C, Nankivell B, Ibrahim R, Bautista C, et al. Resolution of severe ipilimumab-induced hepatitis after antithymocyte globulin therapy. *J Clin Oncol* 2011;29(9):e237-40. doi: [10.1200/jco.2010.32.2206](https://doi.org/10.1200/jco.2010.32.2206)
79. Riveiro-Barciela M, Muñoz-Couselo E, Fernandez-Sojo J, Diaz-Mejia N, Parra-López R, Buti M. Acute liver failure due to immune-mediated hepatitis successfully managed with plasma exchange: new settings call for new treatment strategies? *J Hepatol* 2019;70(3):564-6. doi: [10.1016/j.jhep.2018.10.020](https://doi.org/10.1016/j.jhep.2018.10.020)
80. Marin-Acevedo JA, Dholaria B, Soyano AE, Knutson KL, Chumsri S, Lou Y. Next generation of immune checkpoint therapy in cancer: new developments and challenges. *J Hematol Oncol* 2018;11(1):39. doi: [10.1186/s13045-018-0582-8](https://doi.org/10.1186/s13045-018-0582-8)