

RESEARCH ARTICLE

# The Idea Is Good, but. . .: Failure to Replicate Associations of Oxytocinergic Polymorphisms with Face-Inversion in the N170

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# Abstract

## **Background**

In event-related potentials, the N170 manifests itself especially in reaction to faces. In the healthy population, face-inversion leads to stronger negative amplitudes and prolonged latencies of the N170, effects not being present in patients with autism-spectrum-disorder (ASD). ASD has frequently been associated with differences in oxytocinergic neurotransmission. This ERP-study aimed to investigate the face-inversion effect in association with oxytocinergic candidate genes. It was expected that risk-allele-carriers of the oxytocin-receptor-gene-polymorphism (rs53576) and of CD38 (rs379863) responded similar to upright and inverted faces as persons with ASD. Additionally, reactions to different facial emotional expressions were studied. As there have been difficulties with replications of those molecular genetic association studies, we aimed to replicate our findings in a second study.

#### Method

Seventy-two male subjects in the first-, and seventy-eight young male subjects in the replication-study conducted a face-inversion-paradigm, while recording EEG. DNA was extracted from buccal cells.

#### Results

Results revealed stronger N170-amplitudes and longer latencies in reaction to inverted faces in comparison to upright ones. Furthermore, effects of emotion on N170 were evident. Those effects were present in the first and in the second study. Whereas we found molecular-genetic associations of oxytocinergic polymorphisms with the N170 in the first study, we failed to do so in the replication sample.



#### Conclusion

Results indicate that a deeper theoretical understanding of this research-field is needed, in order to generate possible explanations for these findings. Results, furthermore, support the hypotheses that success of reproducibility is correlated with strength of lower original p-values and larger effect sizes in the original study.

#### Introduction

Social interaction and communication is of special importance for human beings. During social interactions faces play an important role, as one can read many social relevant cues from a face. By perceiving a face, humans are very fast and efficient in deciding e.g. about sex, age or ethnicity of the watched person [1]. Furthermore, facial expression is an important social index of potential threat [2]. Identifying and interpreting this information correctly are important skills in order to react appropriately in social interactions [3]. Therefore, expertise in face-perception is one of the best developed functions of the visual system and seems to be mediated by distinct processes in the brain which respond especially to face-stimuli [4]. Molecular-genetic correlates underlying individual differences in face-perception- and processing are of special interest in relation to the science of prosocial behavior. The oxytocinergic system came more and more into the focus of attention in studying differences in prosocial behavior [5]. Different variants of the oxytocin-receptor-gene (OXTR; for a review see [6]), as well as another gene-polymorphism, CD38, which regulates oxytocin-secretion [7–9] play an important role in studying effects of differences in oxytocinergic functioning on individual differences in neurophysiological responsivity, personality and/or behavior. Relationships between differences in functioning of the oxytocinergic system and autism-related symptoms have been postulated, as well as neurophysiological differences in face-processing [5, 6, 9-12].

#### N170 and Face-Inversion Effect

One core-component in event-related potentials regarding face-perception- and processing is a negative deflection that occurs between 130-180ms and is largest over posterior-temporal electrodes (N170) [13]. In most electrophysiological studies, the effects of inversion have been found to be specific for faces, as compared to other complex stimuli [14–16], as well as to be stronger in the right-hemisphere [17-20]. Healthy adults exhibit a face-inversion effect (FIE), i.e. show an increase in amplitude and a delay in latency in the N170 in reaction to inverted faces compared to upright ones [4, 14, 21-27]. Through alteration of the familiar configuration of the facial features and spatial relations, inversion disrupts configural face processing, as especially second-order relations, such as the relative size of the features' spatial relations, are disturbed [28-33]. Therefore, latency and amplitude are sensitive to disruptions in early face processing stages, whereas facial familiarity or recognition does not influence the N170 [14, 34]. The face-inversion effect seen in healthy persons seems to be impaired in people with Autism-spectrum disorder (ASD) [10, 19, 23, 35], as they exhibit longer latencies in reaction to upright faces (compared to controls), but do not exhibit longer latencies in reaction to inverted faces, as controls do. These effects are not evident in reaction to objects. Furthermore, controls do show higher amplitudes in the N170 in the right hemisphere than in the left one, indicating a right-hemispheric specialization in early face processing that is weaker or absent in subjects with ASD [21].



Results regarding the question if emotional facial expression influences the N170 are heterogeneous: while several studies postulate that emotional content of stimuli is processed in later components and not as early as in the N170 [20, 36–41], other studies suggest that affect is associated with a stronger negativity in the N170 [2, 3, 38, 42, 43], which would imply early automatic encoding of emotional facial expressions [2]. A recent meta-analysis, however, came to the conclusion that emotional facial expression indeed influences N170-amplitudes [44].

# Molecular genetic associations of face-processing

Whereas meta-analyses regarding twin-studies suggest a sustainable heritability of different ERP-waveforms [45], there have only been a few studies regarding face processing in association with molecular genetics, which will be reported in the following sections.

**Serotonin** (5-HT). Serotonergic gene-polymorphisms, especially the serotonin transporter gene SLC6A4 with the serotonin-transporter length polymorphic region (5-HTTLPR) have frequently been associated with the responsivity to stimuli with emotional content (for a review see [46]). ERP-studies investigating associations with reactivity to emotional stimuli and 5-HTTLPR found that carriers of at least one short (s-) allele had higher amplitudes in the error-related negativity (ERN) [47]. In an emotional picture-processing task, s-allele carriers of the 5-HTTLPR reacted stronger in early stages of emotional processing, but not in late ones [48]. Battaglia and colleagues [49] reported associations of s-allele carriers and differences in reaction to emotional faces in the N400, but not in the N170. Similar results were reported from Labuschagne et al. [50] in a citalopram-challenge: Whereas an increase of serotonin led to an enhanced N250 as well as larger amplitudes in the late positive potential (LPP) in reaction to happy (vs. neutral) faces, no effect on the N170 was evident. Schmidt and colleagues [51] reported that the glutamatergic NMDA receptor antagonist ketamine, as well as the 5-HT agonist psilocybin led to reduced N170-amplitudes in reaction to fearful faces. Furthermore, ketamine also led to reduced N170-amplitudes in response to happy faces, whereas this was not the case under influence of psilocybin. As far as we know, no study investigated associations of serotonergic functionality with the FIE, yet.

**Dopamine (DA).** Regarding dopaminergic neurotransmission, associations of the catechol-O-methyltransferase (COMT) val158met polymorphism and time of onset of the N170 have been reported; hereby, the met-allele was associated with significantly reduced N170-latencies in children [52]. However, most studies that investigate dopaminergic polymorphisms in association with ERPs, report associations with the ERN, the conflict-N400 or the LPP [53–57]. Results regarding face-inversion in association with dopaminergic neurotransmission have not yet been published.

**Oxytocin (OX).** As reported above, results regarding face-processing and associations with dopaminergic, or serotonergic functioning have been sparse. Therefore, the most promising influencing factor on socially relevant cues, such as faces, might be oxytocinergic neurotransmission, because of various results regarding the influence of oxytocin on social behavior, trust, emotion- and face-recognition, as well as face processing [58]. Waller and colleagues [59] reported influences of oxytocin (intranasal administration) on face-processing of pictures of children's faces in the N250 and N300 in fathers: view of father's own child (in comparison to pictures of familiar or unfamiliar children) led to enhanced negativities in the N250 and N300, only in the placebo-condition, with this difference being absent after oxytocin-administration. Most promising were the results from Huffmeijer et al [60], where oxytocin-administration led to increases of amplitudes in reaction to emotional faces in the vertex positive potential (VPP), reflecting the same processes as the N170 [61]. The authors also found that the LPP showed higher amplitudes in general after oxytocin-administration in comparison to the control-group



[60]. Therefore, individual differences in oxytocin-secretion-/ and functioning seem to be worthwhile to study in association with processing of faces.

# Oxytocinergic gene polymorphisms

Oxytocin-receptor-gene. Different genetic variants of the oxytocin-receptor-gene have been associated with impairments in social behavior [5, 62]. There is only one receptor for Oxytocin: OXTR [62]. OXTR has also been found in the amygdala, which points to the potential modulation of amygdala-activity through OX [63]. One single-nucleotide-polymorphism (SNP), found on intron 3 is SNP rs53576 (G>A), seems to have an inhibiting influence on transcription of the OXTR gene [64]. Its function is, however, not totally clear, yet [6], but it seems to have an influence on social behavior and interactions [6, 65]. In a sample with healthy students, subjects homozygous for the G-allele showed more dispositional and behavioral empathy, measured with the "Reading the Mind in the Eyes test", than students with at least one A-allele. Furthermore, associations of optimistic behavior [66], and of prosocial temperament and the GG-genotype in comparison to A-allele carriers have been reported. Besides, a significant reduction in amygdala-activity in reaction to negative emotional faces [67], as well as more positive emotions in G-allele-carriers [68] are being postulated. In a sample of 195 Chinese families, an association of SNP rs5357 G-allele-carriers and autism has been reported [69]. This effect could, however, not be replicated in a Caucasian sample [70].

CD38. Beside the OXTR-gene, CD38, which is known as a transmembrane glycoprotein with fundamental importance in neuropeptide release [7], is involved in the regulation of the oxytocinergic system as well [7, 71, 72]. Within the CD38-gene, various SNPs can be distinguished. Munesue and colleagues [73] reported that the SNP rs3796863 (C>A), located on intron seven, showed associations with high functioning autism (IQ > 70) in a Caucasian population (US), but not in a Japanese sample. Furthermore, Lerer and colleagues [74] reported associations between several SNPs across the CD38 gene and low functioning ASD, as well as a reduction in CD38 expression in lymphocytes in persons with ASD. Sauer and colleagues [75] showed associations between this SNP (rs3796863) and neural processing of social stimuli, with homozygous C-allele carriers expressing higher activation in the left fusiform gyrus (FG), as well as slower reaction times during processing of neutral and emotional faces. Intranasal application of OX led to reduction of reaction times in CC-homozygous subjects and, therefore, to improved processing of social relevant stimuli [75]. Results regarding genetic variants of the CD38 gene hint to an involvement in the regulation of social interactions and might have important implications for psychiatric disorders, which are characterized by social deficits [76]. Therefore, both polymorphisms, CD38 and OXTR seem to be worthwhile to be studied in associations with individual differences regarding the reaction to social stimuli—such as (emotional) faces.

# Research question

The goal of this study was to find out, if differences in oxytocin-related polymorphisms-namely oxytocin-receptor-gene-polymorphism (rs53576) and CD38-polymorphism (rs3796863) were influencing neurophysiological face-processing in healthy adults. In comparison to results found in association with subjects with ASD and the FIE, we conducted an EEG-study with a face-inversion-paradigm that is similar to the one used by McPartland et al. [10], in a sample of 72 healthy young men. Important questions in this context are, if alterations of the N170 in response to faces and objects are associated with genetic determinants of the oxytocinergic system, and, if these can be observed in healthy subjects. This would point to an influence of genetically determined oxytocinergic gene variants on processing of socially and emotionally relevant cues.



Due to the ongoing discussion about reproducibility problems in psychological- and especially neuroscience [77–79], and in order to see if found effects were stable and reliable, we conducted a replication of the original study with the same paradigm and a comparable sample of 78 healthy young males.

Thus, it was hypothesized:

- 1. Neutral upright vs. neutral inverted faces (face-inversion) and upright vs. inverted chairs (object-inversion as control stimulus):
  - a. Stronger amplitudes and longer latencies in reaction to inverted than to upright neutral faces in general are expected.
  - b. N170-amplitudes and latencies in reaction to faces are expected to be higher and faster than to chairs, respectively.
  - c. Furthermore, it is expected that risk-allele carriers of OXTR and/or CD38 will not show a FIE, and, therefore, exhibit no shorter N170-latencies and smaller N170-amplitudes in reaction to inverted faces, as well as no lateralization in the right hemisphere. Latencies in reaction to objects (chairs) are expected to be shorter in risk-allele carriers than in non-risk-allele-carriers.
- Exploratory analyzes would further reveal amplitude differences in reaction to emotional (happy, angry, neutral) facial expressions, with and without associations with OXTR-SNP and/or CD38-genotypes.

#### Method

#### **Participants**

**Study 1.** Seventy-two native German-speaking Caucasian healthy young males participated in this study. Subjects had normal or corrected-to-normal vision and were right-handed. EEG-testing was only one part of the study; fMRI-testing and a learning-paradigm took place on different days after EEG-testing. All subjects gave their written informed consent before testing and received a monetary compensation of  $\epsilon$ 10/h for their participation in the entire project. Before testing, participants provided genetic samples with buccal cells from saliva. Five participants had to be excluded from the EEG- and further analyses, as they had more than two standard deviations less trials than average trial number; leaving a final sample of sixty-seven male subjects between 18 and 33 years of age (M = 24.8; SD = 3.04).

Subjects were grouped on genotype-level in AA and AG vs. GG (A+ vs. GG) for rs53576 (OXTR SNP), whereas the risk-allele group is the one with at least one A-allele (A+); as well as grouping on genotype-level for rs3796863 (CD38 SNP) in CC vs. AA and AC, whereas the critical genotype-variant is CC in this case.

**Study 2.** Seventy-seven native German-speaking Caucasian healthy young males participated in this study. Subjects had normal or corrected-to-normal vision and were right-handed. EEG-recording consisted of the face-inversion paradigm and an emotional Stroop task, which was balanced, so that 50% of the subjects started emotional Stroop task and the other 50% started with the face-inversion paradigm. All subjects gave their written informed consent before testing and received a monetary compensation for their participation in the project. Before testing, participants provided genetic samples with buccal cells from saliva. Five participants had to be excluded from the EEG-analyzes, as they had more than two standard deviations less trials than average-trial number, leaving a final sample of seventy-two male subjects between 18 and 32 years of age (M = 24.30; SD = 3.20).



The face-inversion paradigm, ERP-recordings- and analyzes, genotyping, and statistical analyzes were conducted the same way as in study 1.

Both studies were approved by the ethics committee of the faculty of psychology of the Justus-Liebig-University Giessen and comply with the Declaration of Helsinki.

# Face-Inversion paradigm

Stimuli consisted of gray-scale digital images of neutral, happy and angry faces, 20 per category, 10 women and 10 men—upright and inverted (KDEF-database; [80]), as well as of a set of 20 chairs (with kind regards from the Laboratoire de Recherché Experimentale, Institute de Psychologie, Université de Lausanne, CH), upright and inverted. Pictures were standardized for size, contrast and luminosity. Pictures of faces, as well as the pictures of chairs were cut out in an oval form, so that hair did not influence face-perception. Additionally, subjects had to react to a grayscale image of a butterfly (appearing 60 times across all blocks) by pressing a button of a response pad (Cedrus RB-730, Cedrus Corporation, San Pedro, CA, USA), in order to keep attention stable during the task. All images had a resolution of 420x520 pixels. Stimuli were centrally presented on a 15" computer screen with a black background. The task consisted of six blocks with 90 trials each, summing up to 540 trials with randomized presentation of the stimuli. In the overall paradigm, each face and each chair was presented three times upright and three times inverted, leading to 60 trials per category.

Stimuli were presented for 750ms. A central fixation cross was presented during a randomized inter-trial-interval (1000-1500ms; M = 1250ms). There was a break of 20s after every block. Participants were seated in a comfortable chair at 60cm distance to the screen. Stimulus presentation and response recording was conducted with Presentation 16.3 software (Neurobehavioral Systems Inc., Albany, CA, USA) and a Pentium (Intel Corporation, Santa Clara, CA, USA) based personal computer. SPSS Statistics 21 was used for statistical analyses (IBM Corp., Somer, NY, USA).

#### ERP recordings and analyzes

EEG signals were amplified using a 0.1-80 Hz bandpass filter (BrainAmpDC amplifier; Brain Products GmbH, Gilching, Germany) and a sampling rate of 250Hz at 32 scalp sites according to the 10-20-system. Active Ag/AgCl electrodes were used (actiCAP 32Ch, Brain Products GmbH, Herrsching, Germany). Impedances were kept below  $7k\Omega$ . Vertex electrode was used as onlinereference. The data were processed digitally offline, using Brain Vision Analyzer software (Brain Products GmbH, Gilching, Germany). Data were filtered using a 30 Hz low pass filter, manually inspected for artifacts that were not caused by eye movements and, subsequently, excluded. To remove eye-movement artifacts, an Independent Component Analyses (ICA-) based correction algorithm was applied as implemented in the Brain Vision Analyzer. Thereafter, data were re-referenced to an average reference, filtered using a 0.5 Hz high pass- and a 50 Hz notch filter and segmented (stimulus-locked; -200ms to 1000ms). Segments were averaged for each experimental condition and individual; consisting of 54 trials on average (SD = 4.9) in the first study, and 54 trials in the second study (SD = 5.2). For statistical analyzes of the N170, amplitudes and latencies in the time window between 120 and 220ms were extracted at temporo-parietal-occipital electrodes P7, P8, PO9, PO10, TP9 and TP10. Electrode choice was based on visual inspection of the Grand Average waves and in accordance with the typical topography of the N170.

# Genotyping

DNA was extracted from buccal cells and purification of genomic DNA was performed with a standard commercial extraction kit (MagNA Pure LC DNA Isolation Kit I; Roche Diagnostics,



Mannheim, Germany) in a MagNA Pure LC System (Roche, Mannheim, Germany). Genotyping of the rs3796863 polymorphism was conducted with a commercial TaqMan<sup>®</sup> SNP Genotyping Assay (C\_\_\_1216944\_10, Applied Biosystems, Carlsbad, USA) on a Mastercycler<sup>®</sup> eprealplex (Eppendorf, Hamburg, Germany) according to manufacturers' standard protocol. Genotyping of SNP rs53576 was conducted using the commercial TaqMan<sup>®</sup> SNP Genotyping Assay (C\_\_\_3290335\_10, Applied Biosystems, Carlsbad, USA) on a Mastercycler<sup>®</sup> eprealplex (Eppendorf, Hamburg, Germany) according to manufacturers' standard protocol.

# Statistical Analyzes

Amplitudes/latencies of the N170 were entered into two separate ANOVAs for repeated measurements:

- 1. The within-subject factors electrode (P7, P8, PO9, PO10, TP9 and TP10), perspective (upright vs. inverted), category (neutral face vs. chair), hemisphere (left vs. right), and the between-subject factors genotypes (GG vs. A<sup>+</sup> in OXTR; CC vs. A<sup>+</sup> in CD38).
- 2. In the exploratory analyses of emotional effects on N170, ANOVAs for repeated measurements with within-subject factors electrode (P7, P8, PO9, PO10, TP9 and TP10), emotion (happy, angry, and neutral), hemisphere (left vs. right) and the between-subject-factors genotypes (GG vs. A<sup>+</sup> in rs53576; CC vs. A<sup>+</sup> in rs3796863) were calculated.

Post-hoc tests were calculated with Bonferroni-correction for multiple comparisons. Green-house-Geisser correction was applied in case of violation of sphericity assumption.

#### Results

# Distribution of genotypes

**Study 1: CD38 rs3796863.** Thirty-five male subjects had at least one A-allele and were therefore grouped as A<sup>+</sup>; thirty-six were homozygous C-allele-carriers. One subject could not be genotyped for rs3796863 and was therefore excluded from further analyses. Genotype distribution did not deviate from Hardy-Weinberg-Equilibrium ( $\chi^2 = 0.20$ , p = .65).

**Study 1: OXTRS rs53576.** Thirty-six male subjects were homozygous G-allele-carriers; thirty-six had at least one A-allele and therefore grouped as A1<sup>+</sup>. Genotype distribution did not deviate from Hardy-Weinberg-Equilibrium ( $\chi^2 = 3.18$ , p = .07).

**Study 2: CD38 rs3796863.** Thirty-one male subjects had at least one A-allele and were therefore grouped as A<sup>+</sup>; forty-six were homozygous C-allele-carriers. Genotype distribution did not deviate from Hardy-Weinberg-Equilibrium ( $\chi^2 = 0.25$ , p = .61).

**Study 2: OXTR rs53576.** Forty-one male subjects had at least one A-allele and were therefore grouped as A<sup>+</sup>; thirty-six were homozygous G-allele-carriers. Genotype distribution did not deviate from Hardy-Weinberg-Equilibrium ( $\chi^2 = 0.21$ , p = .64).

# General Face-Inversion (Neutral faces—inverted neutral faces & chairs—inverted chairs)

**Amplitudes Study 1: General effects.** Results show a main effect for perspective (upright vs. inverted), with higher negative amplitudes in reaction to inverted images, a main effect of stimulus category with stronger negative amplitudes in reaction to faces than to chairs, as well as stronger amplitudes in the right hemisphere. Perspective and stimulus-category (faces vs. chairs) interacted significantly, with higher negativities in reaction to inverted faces than to inverted chairs. Furthermore, results show an interaction of perspective and hemisphere, with a stronger right-hemispheric lateralization in the inverted position (see <u>Table 1</u> below).



Table 1. Mean amplitude (μVolt) and Standard Error of the Mean (SEM) in response to position of neutral faces (upright vs. inverted) and chairs (upright vs. inverted); Hemisphere (HEM), Oxytocin-receptor-gene-polymorphism rs53576 (OXTR).

	Study 1	Study 1			
	Mean amplitude (μVolt)	SEM	Mean amplitude (μVolt)	SEM	
Face upright	-3.60	0.30	-4.85	0.33	
Face inverted	-4.46	0.38	-6.50	0.40	
Chair upright	-1.63		-1.29	0.23	
Chair inverted	-1.51		-1.12	0.24	
Face upright HEM right	-3.95	0.39	-5.43	0.39	
Face upright HEM left	-3.25	0.27	-4.15	0.32	
Face inverted HEM right	-5.13	0.47	-7.29	0.47	
Face inverted HEM left	-3.78	0.36	-5.55	0.38	
OXTR GG right hemisphere	-2.72	0.41	-3.76	0.41	
OXTR A+ right hemisphere	-3.40	0.45	-3.72	0.43	

**Amplitudes Study 1: Genotype-interactions.** There was no main effect for genotype (GT) in both polymorphisms in any condition, but an interaction of hemisphere and OXTR-genotype, with right-hemispheric lateralization in the A-allele-carriers, but not in the GG-group. No additive effects or interactions of OXTR and CD38 were evident.

Amplitudes Study 2: General effects. Results show a main effect for perspective (upright vs. inverted), with higher negative amplitudes in reaction to inverted images, a main effect of stimulus category with stronger negative amplitudes in reaction to faces than to chairs, as well as stronger amplitudes in the right hemisphere (Table 1). Perspective and stimulus category (faces vs. chairs) interacted significantly, with higher negativities in reaction to inverted faces than to inverted chairs. The interaction of perspective and hemisphere, with a stronger right-hemispheric lateralization in the inverted perspective, was significant. General effects are comparable to the results in study 1.

**Amplitudes Study 2: Genotype-interactions.** There was no main effect for genotype in both polymorphisms, and no interaction of hemisphere and OXTR-genotype, as it has been found in study 1. Furthermore, no additive effects of OXTR and CD38 were evident. Results of both studies are summarized in Tables 1 and 2.

**Latencies Study 1: General effects.** Concerning latencies, results indicate a main effect for perspective, with longer latencies in reaction to inverted faces (see <u>Table 3</u> below), as well as a main effect for stimulus category with shorter latencies in reaction to faces than to chairs.

Table 2. Results of ANOVAs with repeated measurements regarding N170-amplitudes in reaction to upright and inverted neutral faces, and objects (chairs); Hemisphere (HEM), perspective (upright vs. inverted picture), Oxytocin-receptor-gene-polymorphism rs53576 (OXTR).

	Study 1 df: 1, 62			Study 2			
				df: 1, 68			
	F	р	$\eta_p^2$	F	р	$\eta_p^2$	
Face vs. Object	104.55	<. 01	.628	223.23	< .01	.767	
Face-inversion	23.99	< .01	.279	43.98	< .01	.393	
Face-inversion vs. Object-inversion	33.82	< .01	.353	106.51	< .01	.610	
Perspective x HEM	7.65	< .01	.110	6.46	.011	.087	
OXTR x HEM	4.83	.032	.072	.644	.425	.009	



	Study 1		Study 2		
	Mean latency (ms)	SEM	Mean latency (ms)	SEM	
Face upright	170.18	1.67	171.11	1.32	
Face inverted	174.15	1.48	176.63	1.18	
Chair upright	176.41	2.32	184.20	1.93	
Chair inverted	182.30	2.11	189.51	1.77	
Face upright HEM right	169.25	2.02	170.08	1.41	
Face upright HEM left	171.11	2.20	172.30	1.53	
Face inverted HEM right	172.87	1.68	176.70	1.32	
Face inverted HEM left	175.41	1.89	176.72	1.27	

Table 3. Mean latencies (ms) and standard error of the mean (SEM) in response to perspective of neutral faces (upright vs. inverted) and chairs (upright vs. inverted); Hemisphere (HEM).

There was no main effect of hemisphere, but a stronger right-hemispheric lateralization in reaction to inverted images.

**Latencies Study 1: Genotype-interactions.** A three-way-interaction of CD38 GT x stimulus category x position was shown, with longer latencies in reaction to inverted chairs in risk-carriers (CC), as well as longer latencies in reaction to inverted faces in the non-risk-carriers (A+) (Fig 1). No interaction effects of OXTR or additive/interaction effects of OXTR with CD38 were evident.

**Latencies Study 2: General effects.** Concerning latencies, results indicate a main effect for perspective, with longer latencies in reaction to inverted faces (see <u>Table 3</u>), as well as a main effect for stimulus category with longer latencies in reaction to faces than to chairs. There was no main effect of hemisphere, and no right-hemispheric lateralization in reaction to inverted images.

**Latencies Study 2: Genotype-interactions.** Neither an effect of genotype, nor an interaction of CD38-genotype, stimulus category and perspective, as in study 1 was evident. Furthermore, no interaction-effects of OXTR or additive effects of OXTR with CD38 could be shown. Results regarding N170-latencies of both studies are summarized in Tables  $\underline{3}$  and  $\underline{4}$ .

## Emotional face-inversion (positive, negative and neutral faces)

**Amplitudes Study 1: General effects.** A general FIE was evident, with stronger negative amplitudes in reaction to inverted faces, as well as a main effect of hemisphere with higher negative amplitudes in the right hemisphere. Concerning different facial expressions, amplitudes in reaction to happy faces were significantly higher than towards neutral, but not to angry ones, irrespective if upright or inverted (see <u>Table 5</u> below). Furthermore, results indicate an interaction of perspective and hemisphere, with higher amplitudes in reaction to inverted faces in the right hemisphere (which was evident in reaction to neutral faces as well).

Amplitudes Study 1: Genotype-Interactions. An interaction of OXTR with hemisphere indicates that risk-allele-carriers show a stronger lateralization in the right hemisphere—as seen in reaction to neutral faces as well. Furthermore, a three-way-interaction of CD38-GT x perspective x hemisphere indicates that subjects carrying risk-alleles (CC) of the CD38-SNP show a stronger lateralization in the right hemisphere in reaction to upright and inverted faces, whereas the non-risk-group expresses this lateralization only in reaction to inverted faces (Fig 2). No interaction or additive effects of OXTR with CD38 could be shown (see Table 6 below).

**Amplitudes Study 2: General effects.** Amplitudes were higher in reaction to inverted faces in general. A main effect of hemisphere indicates higher negative amplitudes in the right



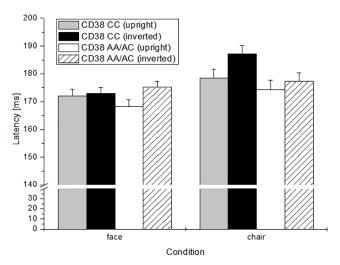


Fig 1. N170-latencies in reaction to upright and inverted faces and chairs in dependence of CD38-allele-frequency. Error bars reflect the standard error of the mean (SEM).

hemisphere. Concerning different facial expressions, amplitudes in reaction to happy faces were significantly higher than to neutral-, but not to angry ones, irrespective if upright or inverted (<u>Table 5</u>). Furthermore, results indicate an interaction of perspective and hemisphere, with stronger amplitudes in reaction to inverted faces in the right hemisphere. Effects are comparable to those in study 1.

**Amplitudes Study 2: Genotype-interactions.** No interaction of OXTR and hemisphere, and no interaction of CD38-GT, perspective and hemisphere, as in study 1, were evident. Furthermore, interactions or additive effects of OXTR with CD38 could not be found. Results regarding N170-amplitudes in reaction to emotional faces are summarized in tables <u>5</u> and <u>6</u>.

**Latencies Study 1: General effects.** Latency differed significantly in dependence of perspective, with longer latencies in reaction to inverted faces. Latencies were longer in reaction to angry than to happy faces, but not to neutral ones (<u>Table 7</u>). Latencies did not differ in dependence of hemisphere.

**Latencies Study 1: Genotype-interactions.** CD38-GT interacted with perspective and facial expression, indicating longer latencies in reaction to angry inverted faces in risk-allele-carriers (CC) than to neutral or happy inverted faces, indicating a present FIE for risk-carriers of CD38, and an absent FIE for CD38-non-risk-carriers (Fig 3). Besides, OXTR-GT interacted significantly with hemisphere and emotion, with shorter latencies in the right hemisphere in

Table 4. Results of ANOVAs with repeated measurements regarding N170-latencies in reaction to upright and inverted neutral faces, and objects (chairs); Hemisphere (HEM), perspective (upright vs. inverted picture), CD38-SNP rs379863 (CD38); category (faces/chairs).

	Study 1  df: 1, 62			Study 2 df: 1, 68			
	F	р	$\eta_{\rho}^{2}$	F	р	$\eta_p^2$	
Face vs. Object	14.33	<. 01	.188	90.59	< .01	.571	
Face-inversion	37.01	< .01	.374	44.31	< .01	.395	
Face-inversion vs. Object-inversion	.697	.407	.011	.029	.866	.000	
Perspective x HEM	4.59	.036	.069	.005	.942	.000	
CD38 x category x perspective	6.44	.014	.094	.895	.348	.013	



Table 5. Mean difference (µVolt) between facial expressions, standard error of the mean (SEM);

Study 1			Study 2			
Emotion	Mean difference (μVolt)	SEM	p <sup>b</sup>	Mean difference (μVolt)	SEM	<b>p</b> <sup>b</sup>
happy vs. angry	20	0.15	0.55	21	.050	.064
happy vs. neutral	17	0.05	< .01**	30	.067	< .01**
angry vs. neural	.03	0.14	1	08	.076	.763

<sup>\*\*</sup> The mean difference is significant at the .01 level;

reaction to upright angry faces in the risk-allele-group. No interaction or additive effects of OXTR with CD38 were evident.

**Latencies Study 2: General effects.** Latency differed significantly in dependence of perspective, with longer latencies in reaction to inverted faces. Latencies were longer in reaction to angry and to neutral faces compared to happy ones. Latencies did not differ in dependence of hemisphere. Therefore, general results are similar to those of study 1.

**Latencies Study 2: Genotype-interactions.** CD38-GT did not interact with perspective and facial expression, as in study 1. Besides, OXTR-GT did not interact significantly with hemisphere and emotion, as found in study 1. Furthermore, no additive effects of OXTR with CD38 were evident. Results of both studies regarding N170-latency-effects in reaction to emotional faces are summarized in tables 7, 8 and 9.

#### **Discussion**

General effects of face-inversion, with higher amplitudes in reaction to inverted than to upright faces and higher amplitudes in reaction to faces than to chairs (manipulation check) were evident in the first, as well as in the replication study. Furthermore, effects of emotional faces, with higher amplitudes and shorter latencies in reaction to happy than to neutral faces were also present in the first, as well as in the replication study.

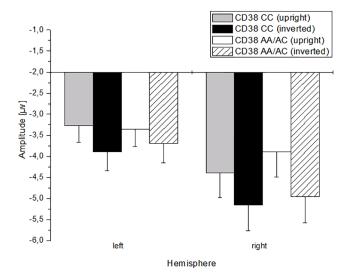


Fig 2. N170-amplitudes in reaction to upright and inverted neutral faces in dependence on allelefrequency of CD38 and hemisphere. Error bars reflect the standard error of the mean (SEM).

<sup>&</sup>lt;sup>b</sup> Adjustment for multiple comparisons: Bonferroni.



Table 6. Results of ANOVAs with repeated measurements regarding N170-amplitudes in reaction to upright and inverted happy, and neutral faces; Hemisphere (HEM), perspective (upright vs. inverted picture), CD38-SNP rs379863 (CD38), Oxytocin-receptor-gene-polymorphism rs53576 (OXTR).

		Study 1			Study 2			
		df: 2, 61			df: 2, 67			
	F	р	$\eta_{\rho}^{2}$	F	р	$\eta_{\rho}^{2}$		
Face-inversion	54.15	< .01	.466	77.21	< .01	.532		
Happy vs. Neutral Faces	5.23	< .01	.146	9.80	< .01	.226		
Hem	11.41	< .01	.155	18.12	< .01	.210		
Face-inversion x HEM	8.56	< .01	.121	4.67	< .05	.064		
OXTR x HEM	7.91	< .01	.113	.625	.432	.009		
CD38 x FI x HEM	4.04	< .05	.061	.717	.400	.010		

Results of the first study indeed indicate associations of different variants of oxytocinergic gene-polymorphisms with amplitude and latency of the N170, with a stronger right-hemispheric lateralization in reaction to faces in OXTR risk-group (A+), an absent FIE in risk-carriers of CD38 (CC), as well as longer latencies in reaction to angry inverted faces in CD38-risk-allele carriers. These genotype-interactions could not be replicated in the second study, though.

#### **Face-Inversion Effect**

Reactions to upright and inverted faces vs. objects (in this case: chairs) were as expected: amplitudes in response to inverted pictures were higher, and amplitudes in reaction to faces were stronger than to chairs. Lateralization showed—as expected—higher negative amplitudes in the right hemisphere, being stronger in reaction to faces than to chairs. Furthermore, genotype of the OXTR-gene polymorphism rs53576 interacted significantly with hemisphere, indicating that the so-called risk-allele-carriers showed—different as expected—a stronger right-hemisphere-lateralization than GG-homozygous. As the other results regarding inversion effects are replicating former findings, results regarding GT-hemisphere-interaction direct in the other direction than expected. The expectation was that risk-allele carriers would react similar as persons with ASD or their relatives, where a lack of lateralization was found—but the opposite was the case: risk-allele carriers showed a stronger lateralization than the so-called non-risk-group, independent of face-inversion or category. Several reasons can account for this effect: results regarding SNP rs53576 have been heterogeneous, with differences in ethnic groups such as Europeans and Asians, so that the "risk-allele" varied in dependence of sample and/or ethnicity [6, 62].

Table 7. Mean latency difference (ms) between facial expressions and standard error of the mean (SEM);

	Study 1			Study 2			
Emotion	Mean difference (ms)	SEM	Significance <sup>b</sup>	Mean difference (ms)	SEM	Significance <sup>b</sup>	
happy vs. angry	-3.32	1.10	< .05*	-2.58	0.40	< .01**	
happy vs. neutral	-1.3	0.54	.06	-1.08	0.32	< .01**	
angry vs. neutral	2.01	0.97	.127	1.49	0.42	< .01**	

<sup>\*</sup> The mean difference is significant at the .05 level;

<sup>\*\*</sup> The mean difference is significant at the .01 level;

<sup>&</sup>lt;sup>b</sup> Adjustment for multiple comparisons: Bonferroni.



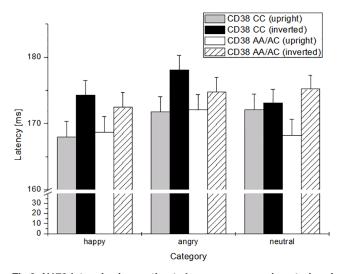


Fig 3. N170-latencies in reaction to happy, angry, and neutral upright and inverted faces in dependence of CD38-allele-frequency. Error bars reflect the standard error of the mean (SEM).

General reactions in latency of N170 were as expected, with longer latencies to inverted pictures and longer latencies in response to chairs than to faces. Furthermore, the CD38-risk group showed longer latencies in reaction to inverted chairs than to faces. This is an effect which was postulated—as subjects with ASD show the same reaction-pattern. It might indicate that attention to objects is more evident in risk-allele-carriers of this CD38-SNP, as the inversion effect shows up in reaction to objects and not to faces.

Results in study 2 replicated the main effects regarding face-inversion, with higher amplitudes in reaction to upright faces than to upright chairs and with higher amplitudes of inverted faces in comparison to upright ones, as well as with a lateralization in the right hemisphere. Neither the right hemispheric-lateralization of OXTR-risk-allele-carriers could be identified, nor longer latencies in reaction to inverted faces in risk-allele carriers of CD38—as found in study 1—could be reproduced.

# Emotional modulation of the Face-Inversion effect

Comparison of happy, angry and neutral upright and inverted faces revealed that N170-amplitudes showed the inversion-effect as well—with the highest negativity in reaction to inverted faces (independent of emotional category), being stronger in the right hemisphere. Concerning effects of distinct emotions, happy faces revealed higher amplitudes than neutral ones, irrespective if upright or inverted. Former results regarding effects of emotional expression on N170

Table 8. Mean latency (ms) and standard error of the mean (SEM) for different facial expressions (happy vs. angry vs. neutral) presented upright and inverted.

	Study 1		Study 2		
Facial expression	Mean latency (ms)	SEM	Mean latency (ms)	SEM	
Нарру	170.86	1.54	169.78	1.32	
Angry	174.18	1.28	172.21	1.29	
Neutral	172.16	1.46	171.25	1.35	



Table 9. Results of ANOVAs with repeated measurements regarding N170-latencies in reaction to upright and inverted happy and neutral faces (emotion); Hemisphere (HEM), perspective (upright vs. inverted picture), CD38-SNP rs379863 (CD38), Oxytocin-receptor-gene-polymorphism rs53576 (OXTR).

	Study 1 df: 2, 61			Study 2 df: 2, 67			
	F	р	$\eta_p^2$	F	р	$\eta_{\rho}^{2}$	
FI	27.89	< .01	.310	89.57	< .01	.568	
Angry vs. Happy Faces	6.82	< .01	.099	22.54	< .01	.249	
OXTR x HEM x emotion	3.54	< .05	.054	.287	.751	.004	
CD38 x perspective x emotion	3.53	< .05	.054	.737	.481	.011	

reported no effects of happy emotional expression, but more negative amplitudes in reaction to fearful [2], as well as sad [81] faces.

Results regarding latency revealed longer latencies in reaction to inverted faces, irrespective of facial expression. In contrast to the differences regarding happy faces in amplitude, latency differed in reaction to angry faces compared to happy ones: subjects showed longer latencies in reaction to angry facial expressions than to happy ones. Furthermore, there was an interaction of reaction to angry faces and CD38, so that risk-allele-carriers reacted later to angry inverted faces than the non-risk group. This effect might point to a greater interference in risk-allele carriers in reaction to angry (inverted) faces, whereas this inversion-effect is not evident in reaction to neutral or happy faces. It has been hypothesized that recognition of inverted faces is better in persons with ASD [82, 83]: it is possible that recognition of angry inverted faces is also better in risk-allele carriers of rs3796863 –and therefore, the reaction to angry inverted faces is interfered by the angry expression. As this SNP has also been associated with other social deficits—such as social—phobia [84], interference in reaction to angry inverted faces might, therefore, be possible. Moreover, reaction to upright angry faces was faster in the right hemisphere for the non-risk group of the OXTR-SNP. This effect might indicate that emotion-recognition of angry faces is faster in the non-risk-group than in the risk-group.

One has to state that these effects are only correlational, so nothing can be said about causal effects of the analyzed polymorphisms. Nevertheless, effects of the first study support the hypotheses that CD38 SNP rs3796863 is involved in face-processing similar to that of subjects with ASD or their relatives. Effects of OXTR-gene SNP 53576 rather indicate that the "risk-allele-carriers" process faces as healthy subjects do, whereas G-allele carriers do not show that right-hemispheric lateralization.

The effect of emotional facial expression on N170 amplitude and latency was again observed in the second study: Amplitudes in reaction to upright happy faces were stronger than to neutral ones, indicating that emotional expression indeed has an influence on the N170. Furthermore, latencies were longer in reaction to inverted angry than to inverted happy faces, indicating that, firstly, emotional facial expression influences N170-latency, and that angry inverted faces disturb the processing of faces more readily than happy faces do. Associations of CD38-risk-allele-carriers with later latencies in reaction to inverted angry faces, as well as associations of hemisphere with OXTR could, however, not be reproduced.

#### General discussion

Whereas we found molecular-genetic associations of oxytocinergic polymorphisms with reactivity to faces in the N170 in the first study, we failed to do so in the replication sample. What



could be replicated, however, are the general effects of face-inversion, independent of oxytocinergic polymorphisms. We investigated the effects of emotional facial expression on N170 on an exploratory basis, as results regarding those effects on the N170 have been heterogeneous [2, 20, 23, 36, 37, 40, 40, 85]. Both studies came to the same result that emotional facial expression directly had an effect on the N170. These effects show that N170 amplitudes-, and latencies are not independent of facial emotional expression, and, therefore, can be influenced by perceiving different facially expressed emotions.

Study 1 yielded significant findings regarding interactions of OXTR and CD38 and face-perception in N170. The replication study, however, failed to do so. It would be too easy to say that results in the first study are clearly due to an Type I error, as it is as well possible that results in the replication study are due to a Type II-error [78]. Although we made sure that the replication study was conducted under the same experimental conditions as the first study, different confounding variables, such as personality, or selective sampling, could have influenced the found results—in both ways, with false-positive findings in the first study, as well as falsenegative findings in the second study. This might indicate an incomplete theoretical understanding of the investigated research field, pointing to other possible explanations for the finding, and leading to other research ideas, which is the only way that science can push forward. Another issue which undermines the hypothesis of a Type I error in the first study is the fact that success of reproducibility is correlated with the strength of lower original p-values and larger effect sizes in the original study, as well as with the finding that more surprising original findings seem to be less likely to be reproduced than less surprising ones [79]. Surprising effects in this study were for example the right-hemispheric-lateralization effects of OXTR-risk-allelecarriers, because an effect in the opposite direction had been expected. The replication study failed to find any effect of OXTR, however.

The question is, though, if both studies were just underpowered, why are the face-inversion-effects, the lateralization effect, and most importantly, the effects of facial emotional expression consistently found in both the original sample as well as in the replication study? It rather seems to be the case that those effects are the stable, reliable ones, whereas the genotype-interactions could not reliably be found. Those results support the hypothesis that reproducibility is correlated with a lower *p*-value and larger effect size [79].

#### Limitations of the studies

The current samples (study 1: N = 72; study 2: N = 78) were comparatively small for molecular-genetic association studies, although, in a comparable range with other combined EEG and molecular genetic studies. Furthermore, subjects were young males—and mainly students—so the results cannot be generalized to all ages or both sexes. However, as subclinical autistic traits are more pronounced in males, they are, therefore, the sample with a higher priority to analyze [86].

The sample of the replication study (N = 78) was only marginally bigger than the first study. Therefore, power-problems occur, as a higher number of subjects would have been necessary in order to maintain the (already small) power of the first study [78, 87]. The procedure of the replication study was the same as in study one, with the exception of another task (emotional Stroop) that subjects additionally conducted in the second study. It is possible that the task influenced the results of the face-inversion-paradigm. However, general face-inversion-effects were replicated from the first study, only the genotype-interactions were not evident in the second study. However, present results rather support the hypothesis that small effect sizes that are close to p = .05 (present genotype-interactions in the first study were often close to p = .05) are often not possible to replicate, whereas strong effects—such as the general face-inversion-



effects (which had strong effect sizes in both studies reported here)—can easily be replicated [79]. This contradicts the hypothesis that the emotional Stroop task interfered with the replication of the results in the second study.

# Conclusion & perspectives

We tried to replicate our findings regarding face-inversion and associations with the oxytocinergic polymorphisms OXTR (rs53576) and CD38 (rs3796863), but could, however, not find these effects in the replication sample. This can be due to many reasons and it does not mean (as already mentioned above) that these effects were definitely accidental. It does, however, show that found effects cannot always be replicated in another study that directly replicates the same paradigm, even in the same laboratory, and with the same stimuli. These results directly correlate with the findings of Nosek and colleagues [79] who criticize the fact that many published results in psychological science cannot be replicated. Present results just emphasize the great necessity to conduct replication studies, especially in the field of psychological neuroscience, in order to come closer to the "true" result. Replications are not a waste of time but an urgent imperative in order to stay a credible and authentic science, which does not only publish effects, independent of the fact if they can be reliably found or not.

This experiment is the first to study individual differences regarding the face-inversion-effect combined with molecular-genetics, and to conduct a direct replication in order to find out if found effects were reliable and stable.

Results are, however, not only highly interesting regarding molecular genetic associations of face-processing, but also regarding general information processing of emotional faces, as an effect of emotional facial expression of the N170 could reliably be found in both the original as well as in the replication sample.

# **Supporting Information**

S1 Dataset. Data regarding N170-amplitudes-/latencies and genotypes of the first study. (SAV)

S2 Dataset. Data regarding N170-amplitudes-/latencies and genotypes of the second study.  $(\mathrm{SAV})$ 

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#### **Author Contributions**

Conceived and designed the experiments: AJLM AH JH. Performed the experiments: JE. Analyzed the data: AJLM. Contributed reagents/materials/analysis tools: AJLM AH PG. Wrote the paper: AJLM AH JH JE.

#### References

- 1. Bruce V, Young A. Understanding face recognition. British Journal of Psychology 1986; 77(3):305–27.
- 2. Batty M, Taylor MJ. Early processing of the six basic facial emotional expressions. Cognitive Brain Research 2003; 17(3):613–20. PMID: 14561449
- Blau VC, Maurer U, Tottenham N, McCandliss BD. The face-specific N170 component is modulated by emotional facial expression. Behavioral and Brain Functions 2007; 3(7):1–13.



- Haxby JV, Hoffman EA, Gobbini MI. Human neural systems for face recognition and social communication. Biological Psychiatry 2002; 51(1):59–67. PMID: 11801231
- Ebstein RP, Israel S, Lerer E, Uzefovsky F, Shalev I, Gritsenko I et al. Arginine vasopressin and oxytocin modulate human social behavior. Annals of the New York Academy of Sciences 2009; 1167(1):87– 102.
- Meyer-Lindenberg A, Domes G, Kirsch P, Heinrichs M. Oxytocin and vasopressin in the human brain: social neuropeptides for translational medicine. Nature Reviews Neuroscience 2011; 12(9):524–38. doi: 10.1038/nrn3044 PMID: 21852800
- Jin D, Liu H, Hirai H, Torashima T, Nagai T, Lopatina O et al. CD38 is critical for social behaviour by regulating oxytocin secretion. Nature 2007; 446(7131):41–5. PMID: 17287729
- Bartz JA, McInnes LA. CD38 regulates oxytocin secretion and complex social behavior. Bioessays 2007; 29(9):837–41. PMID: 17688286
- Higashida H, Yokoyama S, Kikuchi M, Munesue T. CD38 and its role in oxytocin secretion and social behavior. Oxytocin, Vasopressin and Social Behavior 2012; 61(3):351–8. Available from: URL: <a href="http://www.sciencedirect.com/science/article/pii/S0018506X11002881">http://www.sciencedirect.com/science/article/pii/S0018506X11002881</a>.
- McPartland J, Dawson G, Webb SJ, Panagiotides H, Carver LJ. Event-related brain potentials reveal anomalies in temporal processing of faces in autism spectrum disorder. Journal of Child Psychology and Psychiatry 2004; 45(7):1235–45. PMID: <u>15335344</u>
- Wu N, Li Z, Su Y. The association between oxytocin receptor gene polymorphism (OXTR) and trait empathy. Journal of Affective Disorders 2012; 138(3):468–72. doi: 10.1016/j.jad.2012.01.009
   PMID: 22357335
- Hollander E, Bartz J, Chaplin W, Phillips A, Sumner J, Soorya L et al. Oxytocin increases retention of social cognition in autism. Biological Psychiatry 2007; 61(4):498–503. PMID: 16904652
- Luck SJ, Kappenman ES. The Oxford handbook of event-related potential components: Oxford university press; 2011.
- Eimer M. Effects of face inversion on the structural encoding and recognition of faces: Evidence from event-related brain potentials. Cognitive Brain Research 2000; 10(1):145–58.
- Rossion B, Caldara R, Seghier M, Schuller A, Lazeyras F, Mayer E. A network of occipito-temporal face-sensitive areas besides the right middle fusiform gyrus is necessary for normal face processing. Brain 2003; 126(11):2381–95.
- Rossion B, Jacques C. Does physical interstimulus variance account for early electrophysiological face sensitive responses in the human brain? Ten lessons on the N170. Neuroimage 2008; 39(4):1959–79.
   PMID: 18055223
- Dawson G, Carver L, Meltzoff AN, Panagiotides H, McPartland J, Webb SJ. Neural Correlates of Face and Object Recognition in Young Children with Autism Spectrum Disorder, Developmental Delay, and Typical Development. Child Development 2002; 73(3):700–17. PMID: 12038546
- Dawson G, Webb SJ, McPartland J. Understanding the nature of face processing impairment in autism: insights from behavioral and electrophysiological studies. Developmental Neuropsychology 2005; 27 (3):403–24. PMID: 15843104
- Webb SJ, Dawson G, Bernier R, Panagiotides H. ERP evidence of atypical face processing in young children with autism. Journal of Autism and Developmental Disorders 2006; 36(7):881–90. PMID: 16897400
- Apicella F, Sicca F, Federico RR, Campatelli G, Muratori F. Fusiform Gyrus responses to neutral and emotional faces in children with Autism Spectrum Disorders: a High Density ERP study. Behavioural Brain Research 2013; 251:155–62. doi: 10.1016/j.bbr.2012.10.040 PMID: 23124137
- 21. Bentin S, Allison T, Puce A, Perez E, McCarthy G. Electrophysiological studies of face perception in humans. Journal of Cognitive Neuroscience 1996; 8(6):551–65. PMID: 20740065
- Itier RJ, Taylor MJ. N170 or N1? Spatiotemporal differences between object and face processing using ERPs. Cerebral Cortex 2004; 14(2):132–42. PMID: 14704210
- Dawson G, Webb SJ, Carver L, Panagiotides H, McPartland J. Young children with autism show atypical brain responses to fearful versus neutral facial expressions of emotion. Developmental Science 2004; 7(3):340–59. PMID: 15595374
- **24.** Luo W, He W, Yang S, Feng W, Chen T, Wang L et al. Electrophysiological evidence of facial inversion with rapid serial visual presentation. Biological Psychology 2013; 92(2):395–402. doi: 10.1016/j. biopsycho.2012.11.019 PMID: 23254280
- Itier RJ, Taylor MJ. Inversion and contrast polarity reversal affect both encoding and recognition processes of unfamiliar faces: a repetition study using ERPs. Neuroimage 2002; 15(2):353–72. PMID: 11798271



- Rossion B. Spatio-temporal localization of the face inversion effect: an event-related potentials study. Biological Psychology 1999; 50(3):173–89. PMID: 10461804
- Eimer M. Does the face-specific N170 component reflect the activity of a specialized eye processor? Neuroreport 1998; 9(13):2945–8. PMID: 9804295
- 28. Barton JJS, Keenan JP, Bass T. Discrimination of spatial relations and features in faces: Effects of inversion and viewing duration. British Journal of Psychology 2001: 92(3):527–49.
- Farah MJ, Wilson KD, Drain HM, Tanaka JR. The inverted face inversion effect in prosopagnosia: Evidence for mandatory, face-specific perceptual mechanisms. Vision Research 1995; 35(14):2089–93. PMID: 7660612
- Searcy JH, Bartlett JC. Inversion and processing of component and spatial-relational information in faces. Journal of Experimental Psychology: Human Perception and Performance 1996; 22(4):904. PMID: 8756958
- 31. Yin RK. Looking at upside-down faces. Journal of Experimental Psychology 1969; 81(1):141.
- Leder H, Bruce V. When inverted faces are recognized: The role of configural information in face recognition. The Quarterly Journal of Experimental Psychology: Section A 2000; 53(2):513–36.
- Liu J, Harris A, Kanwisher N. Perception of face parts and face configurations: an fMRI study. Journal
  of Cognitive Neuroscience 2010; 22(1):203–11. doi: 10.1162/jocn.2009.21203 PMID: 19302006
- Bentin S, Deouell LY. Structural encoding and identification in face processing: erp evidence for separate mechanisms. Cognitive Neuropsychology 2000; 17(1):35–55. doi: 10.1080/026432900380472
   PMID: 20945170
- **35.** Hileman CM, Henderson H, Mundy P, Newell L, Jaime M. Developmental and individual differences on the P1 and N170 ERP components in children with and without autism. Developmental Neuropsychology 2011; 36(2):214–36. doi: 10.1080/87565641.2010.549870 PMID: 21347922
- **36.** Eimer M, Holmes A. An ERP study on the time course of emotional face processing. Neuroreport 2002; 13(4):427–31. PMID: 11930154
- Ashley V, Vuilleumier P, Swick D. Time course and specificity of event-related potentials to emotional expressions. Neuroreport 2004; 15(1):211–6. PMID: 15106860
- Eimer M, Holmes A. Event-related brain potential correlates of emotional face processing. Neuropsychologia 2007; 45(1):15–31. PMID: 16797614
- 39. Holmes A, Nielsen MK, Green S. Effects of anxiety on the processing of fearful and happy faces: An event-related potential study. Biological Psychology 2008; 77(2):159–73. Available from: URL: <a href="http://www.sciencedirect.com/science/article/pii/S0301051107001627">http://www.sciencedirect.com/science/article/pii/S0301051107001627</a>. PMID: 18022310
- 40. Batty M, Meaux E, Wittemeyer K, Rogé B, Taylor MJ. Early processing of emotional faces in children with autism: an event-related potential study. Journal of Experimental Child Psychology 2011; 109 (4):430–44. doi: 10.1016/j.jecp.2011.02.001 PMID: 21458825
- 41. Wong TKW, Fung PCW, Chua SE, McAlonan GM. Abnormal spatiotemporal processing of emotional facial expressions in childhood autism: dipole source analysis of event-related potentials. European Journal of Neuroscience 2008; 28(2):407–16. doi: 10.1111/j.1460-9568.2008.06328.x PMID: 18702712
- Pizzagalli DA, Lehmann D, Hendrick AM, Regard M, Pascual-Marqui RD, Davidson RJ. Affective judgments of faces modulate early activity (160 ms) within the fusiform gyri. Neuroimage 2002; 16(3):663–77. PMID: 12169251
- Eger E, Jedynak A, Iwaki T, Skrandies W. Rapid extraction of emotional expression: evidence from evoked potential fields during brief presentation of face stimuli. Neuropsychologia 2003; 41(7):808–17. PMID: 12631531
- **44.** Hinojosa JA, Mercado F, Carretié L. N170 sensitivity to facial expression: A meta-analysis. Neuroscience & Biobehavioral Reviews 2015; 55:498–509.
- 45. van Beijsterveldt C, van Baal G. Twin and family studies of the human electroencephalogram: a review and a meta-analysis. Biological Psychology 2002; 61(1–2):111–38. PMID: 12385672
- Jonassen R, Landrø NI. Serotonin transporter polymorphisms (5-HTTLPR) in emotion processing: implications from current neurobiology. Progress in Neurobiology 2014; 117:41–53. doi: 10.1016/j. pneurobio.2014.02.003 PMID: 24548605
- Fallgatter AJ, Herrmann MJ, Roemmler J, Ehlis A, Wagener A, Heidrich A et al. Allelic variation of serotonin transporter function modulates the brain electrical response for error processing. Neuropsychopharmacology 2004; 29(8):1506–11. PMID: 15187981
- **48.** Herrmann MJ, Huter T, Müller F, Mühlberger A, Pauli P, Reif A et al. Additive effects of serotonin transporter and tryptophan hydroxylase-2 gene variation on emotional processing. Cerebral cortex (New York, N.Y.: 1991) 2007; 17(5):1160–3.



- Battaglia M, Ogliari A, Zanoni A, Citterio A, Pozzoli U, Giorda R et al. Influence of the serotonin transporter promoter gene and shyness on children's cerebral responses to facial expressions. Archives of General Psychiatry 2005; 62(1):85–94. PMID: <u>15630076</u>
- Labuschagne I, Croft RJ, Phan KL, Nathan PJ. Augmenting serotonin neurotransmission with citalopram modulates emotional expression decoding but not structural encoding of moderate intensity sad facial emotional stimuli: an event-related potential (ERP) investigation. Journal of Psychopharmacology (Oxford, England) 2010; 24(8):1153–64.
- Schmidt A, Kometer M, Bachmann R, Seifritz E, Vollenweider F. The NMDA antagonist ketamine and the 5-HT agonist psilocybin produce dissociable effects on structural encoding of emotional face expressions. Psychopharmacology 2013; 225(1):227–39. doi: 10.1007/s00213-012-2811-0 PMID: 22836372
- 52. Battaglia M, Zanoni A, Giorda R, Pozzoli U, Citterio A, Beri S et al. Effect of the catechol-O-methyltransferase val(158)met genotype on children's early phases of facial stimuli processing. Genes, Brain, and Behavior 2007; 6(4):364–74. PMID: 16939639
- 53. Althaus M, Groen Y, Wijers AA, Mulder Lambertus J M, Minderaa RB, Kema IP et al. Differential effects of 5-HTTLPR and DRD2/ANKK1 polymorphisms on electrocortical measures of error and feedback processing in children. Clinical neurophysiology: official journal of the International Federation of Clinical Neurophysiology 2009; 120(1):93–107.
- 54. Larson MJ, Clayson PE, Primosch M, Leyton M, Steffensen SC. The Effects of Acute Dopamine Precursor Depletion on the Cognitive Control Functions of Performance Monitoring and Conflict Processing: An Event-Related Potential (ERP) Study. PLoS one 2015; 10(10):e0140770. doi: 10.1371/journal.pone.0140770 PMID: 26492082
- 55. Osinsky R, Hewig J, Alexander N, Hennig J. COMT Val158Met genotype and the common basis of error and conflict monitoring. Brain Research 2012; 1452:108–18. doi: 10.1016/j.brainres.2012.02.054 PMID: 22425188
- 56. Rangel-Gomez M, Hickey C, van Amelsvoort T, Bet P, Meeter M. The detection of novelty relies on dopaminergic signaling: evidence from apomorphine's impact on the novelty N2. PLoS one 2013; 8(6): e66469. doi: 10.1371/journal.pone.0066469 PMID: 23840482
- 57. Meyer A, Klein DN, Torpey DC, Kujawa AJ, Hayden EP, Sheikh HI et al. Additive effects of the dopamine D2 receptor and dopamine transporter genes on the error-related negativity in young children. Genes, Brain, and Behavior 2012; 11(6):695–703. doi: 10.1111/j.1601-183X.2012.00812.x PMID: 22682503
- 58. Meyer-Lindenberg A. Impact of prosocial neuropeptides on human brain function. Progress in Brain Research 2008; 170:463–70. doi: 10.1016/S0079-6123(08)00436-6 PMID: 18655902
- Waller C, Wittfoth M, Fritzsche K, Timm L, Wittfoth-Schardt D, Rottler E et al. Attachment representation modulates oxytocin effects on the processing of own-child faces in fathers. Psychoneuroendocrinology 2015; 62:27–35. doi: 10.1016/j.psyneuen.2015.07.003 PMID: 26221767
- 60. Huffmeijer R, Alink Lenneke R A, Tops M, Grewen KM, Light KC, Bakermans-Kranenburg MJ et al. The impact of oxytocin administration and maternal love withdrawal on event-related potential (ERP) responses to emotional faces with performance feedback. Hormones and Behavior 2013; 63(3):399–410. doi: 10.1016/j.yhbeh.2012.11.008 PMID: 23159479
- 61. Joyce C, Rossion B. The face-sensitive N170 and VPP components manifest the same brain processes: the effect of reference electrode site. Clinical neurophysiology: official journal of the International Federation of Clinical Neurophysiology 2005; 116(11):2613–31.
- 62. Ebstein RP, Knafo A, Mankuta D, Chew SH, San Lai P. The contributions of oxytocin and vasopressin pathway genes to human behavior. Hormones and Behavior 2012; 61(3):359–79. doi: 10.1016/j. yhbeh.2011.12.014 PMID: 22245314
- Huber D, Veinante P, Stoop R. Vasopressin and oxytocin excite distinct neuronal populations in the central amygdala. Science (New York, N.Y.) 2005; 308(5719):245–8.
- Mizumoto Y, Kimura T, Ivell R. A genomic element within the third intron of the human oxytocin receptor gene may be involved in transcriptional suppression. Molecular and Cellular Endocrinology 1997; 135 (2):129–38. PMID: <u>9484908</u>
- 65. Kumsta R, Heinrichs M. Oxytocin, stress and social behavior: neurogenetics of the human oxytocin system. Current Opinion in Neurobiology 2013; 23(1):11–6. doi: 10.1016/j.conb.2012.09.004 PMID: 23040540
- 66. Saphire-Bernstein S, Way BM, Kim HS, Sherman DK, Taylor SE. Oxytocin receptor gene (OXTR) is related to psychological resources. Proceedings of the National Academy of Sciences of the United States of America 2011; 108(37):15118–22. doi: 10.1073/pnas.1113137108 PMID: 21896752
- 67. Tost H, Kolachana B, Hakimi S, Lemaitre H, Verchinski BA, Mattay VS et al. A common allele in the oxytocin receptor gene (OXTR) impacts prosocial temperament and human hypothalamic-limbic structure



- and function. Proceedings of the National Academy of Sciences of the United States of America 2010; 107(31):13936–41. doi: 10.1073/pnas.1003296107 PMID: 20647384
- Lucht MJ, Barnow S, Sonnenfeld C, Rosenberger A, Grabe HJ, Schroeder W et al. Associations between the oxytocin receptor gene (OXTR) and affect, Ioneliness and intelligence in normal subjects. Progress in Neuro-psychopharmacology & Biological Psychiatry 2009; 33(5):860–6.
- 69. Wu S, Jia M, Ruan Y, Liu J, Guo Y, Shuang M et al. Positive association of the oxytocin receptor gene (OXTR) with autism in the Chinese Han population. Biological Psychiatry 2005; 58(1):74–7. PMID: 15992526
- Jacob S, Brune CW, Carter CS, Leventhal BL, Lord C, Cook EH. Association of the oxytocin receptor gene (OXTR) in Caucasian children and adolescents with autism. Neuroscience Letters 2007; 417 (1):6–9. PMID: <u>17383819</u>
- Higashida H, Lopatina O, Yoshihara T, Pichugina YA, Soumarokov AA, Munesue T et al. Oxytocin signal and social behaviour: comparison among adult and infant oxytocin, oxytocin receptor and CD38 gene knockout mice. Journal of Neuroendocrinology 2010; 22(5):373–9. doi: 10.1111/j.1365-2826. 2010.01976.x PMID: 20141571
- Salmina AB, Lopatina O, Ekimova MV, Mikhutkina SV, Higashida H. CD38/cyclic ADP-ribose system: a new player for oxytocin secretion and regulation of social behaviour. Journal of Neuroendocrinology 2010; 22(5):380–92. doi: 10.1111/j.1365-2826.2010.01970.x PMID: 20141572
- 73. Munesue T, Yokoyama S, Nakamura K, Anitha A, Yamada K, Hayashi K et al. Two genetic variants of CD38 in subjects with autism spectrum disorder and controls. Neuroscience Research 2010; 67 (2):181–91. doi: 10.1016/j.neures.2010.03.004 PMID: 20435366
- 74. Lerer E, Levi S, Israel S, Yaari M, Nemanov L, Mankuta D et al. Low CD38 expression in lymphoblastoid cells and haplotypes are both associated with autism in a family-based study. Autism research: official journal of the International Society for Autism Research 2010; 3(6):293–302.
- **75.** Sauer C, Montag C, Wörner C, Kirsch P, Reuter M. Effects of a common variant in the CD38 gene on social processing in an oxytocin challenge study: possible links to autism. Neuropsychopharmacology: official publication of the American College of Neuropsychopharmacology 2012; 37(6):1474–82.
- Young LJ. Regulating the social brain: a new role for CD38. Neuron 2007; 54(3):353–6. PMID: 17481389
- 77. Button KS, Ioannidis John P A, Mokrysz C, Nosek BA, Flint J, Robinson Emma S J et al. Power failure: why small sample size undermines the reliability of neuroscience. Nature Reviews. Neuroscience 2013; 14(5):365–76. doi: 10.1038/nrn3475 PMID: 23571845
- 78. Maxwell SE, Lau MY, Howard GS. Is psychology suffering from a replication crisis? What does \"failure to replicate\" really mean? The American psychologist 2015; 70(6):487–98. doi: 10.1037/a0039400 PMID: 26348332
- Open Science Collaboration. Estimating the reproducibility of psychological science. Science (New York, N.Y.) 2015; 349(6251):aac4716.
- Lundqvist D, Flykt A, Öhman A. The Karolinska directed emotional faces (KDEF). CD ROM from Department of Clinical Neuroscience, Psychology section, Karolinska Institutet 1998:91–630.
- O'Connor K, Hamm JP, Kirk IJ. Neurophysiological responses to face, facial regions and objects in adults with Asperger's syndrome: An ERP investigation. International Journal of Psychophysiology 2007; 63(3):283–93. PMID: <u>17267061</u>
- Langdell T. Recognition of faces: An approach to the study of autism. Journal of Child Psychology and Psychiatry 1978; 19(3):255–68. PMID: 681468
- 83. Hobson RP, Lee A. Hello and goodbye: A study of social engagement in autism. Journal of Autism and Developmental Disorders 1998; 28(2):117–27. PMID: 9586774
- 84. Neumann ID, Slattery DA. Oxytocin in General Anxiety and Social Fear: A Translational Approach. Biological Psychiatry 2015.
- 85. Liu J, Harris A, Kanwisher N. Stages of processing in face perception: an MEG study. Nature Neuroscience 2002; 5(9):910–6. PMID: 12195430
- 86. Baron-Cohen S, Hoekstra RA, Knickmeyer R, Wheelwright S. The autism-spectrum quotient (AQ)— Adolescent version. Journal of Autism and Developmental Disorders 2006; 36(3):343–50. PMID: 16552625
- 87. Yuan K, Maxwell S. On the Post Hoc Power in Testing Mean Differences. Journal of Educational and Behavioral Statistics 2005; 30(2):141–67.