



# Complete Genome Sequences of the Methicillin-Resistant Strain *Staphylococcus aureus* 17Gst354 and Its Prophage *Staphylococcus* Phage vB\_StaphS-IVBph354

Sonja Kittl,<sup>a</sup> Isabelle Brodard,<sup>a</sup> Gudrun Overesch,<sup>a</sup>  Peter Kuhnert,<sup>a</sup>  Joerg Jores,<sup>a</sup>  Fabien Labroussaa<sup>a</sup>

<sup>a</sup>Institute of Veterinary Bacteriology, Department of Infectious Diseases and Pathobiology, University of Bern, Bern, Switzerland

**ABSTRACT** We report the complete 2,783,931-bp circular genome sequence of the human methicillin-resistant strain *Staphylococcus aureus* 17Gst354, isolated from a nasal swab. The strain possessed an additional 4,397-bp plasmid. Moreover, we induced and sequenced its temperate phage *Staphylococcus* phage vB\_StaphS-IVBph354, which has a circular genome of 41,970 bp.

The livestock-associated (LA) methicillin-resistant *Staphylococcus aureus* (MRSA) strain 17Gst354 was isolated from a nasal swab of a healthy Swiss farmer in 2017 (Switzerland) (1). The study was approved by the Ethics Committee for Research of the Canton of Bern (Req-2017-00793). Genomic DNA was extracted as previously reported (2), quantified using a Qubit 4.0 fluorometer (Thermo Fisher Scientific), and then qualified and sized (12- to 15-kb fragments; no shearing) using the Advanced Analytical FEMTO Pulse system (Agilent). Multiplexed SMRTbell libraries were prepared according to the manufacturer's instructions using the SMRTbell Express template prep kit v2.0. Single-molecule real-time (SMRT) sequencing was performed at the Next Generation Sequencing Platform (University of Bern) on the Sequel system using the Sequel sequencing kit v3.0.

*Staphylococcus* phage vB\_StaphS-IVB354 was purified from strain 17Gst354 through three successive single-plaque isolations after mitomycin C induction, as previously reported (3). Phage genomic DNA was isolated from high-titer lysates using a phenol-chloroform extraction protocol (4). Library preparation was performed using the NEBNext Ultra II directional DNA library prep kit, and Illumina sequencing was performed at Eurofins Genomics (Ebersberg, Germany) on the Illumina NovaSeq 6000 platform in 2 × 150-bp sequencing mode. The raw reads were quality controlled using FastQC v0.11.9 (5) and LongQC v1.2.0 (6) for the Illumina and PacBio reads, respectively. Default parameters were used for all software unless otherwise specified.

A total of 166,155 long reads (average length, 8,827 bp;  $N_{50}$ , 10,217 bp; coverage, 494×) and 3,044,780 paired-end short reads obtained previously (1) (coverage, 297×; SRA accession number [SRX6491214](https://www.ncbi.nlm.nih.gov/sra/SRX6491214)) were used to assemble the complete 2,783,931-bp chromosome (G+C content, 32.9%) of strain 17Gst354 using Unicycler v0.4.4 (7). The chromosome was rotated to the first nucleotide of the start codon of the *dnaA* gene using the fixstart command of Circlator v1.5.5 (8). The chromosome sequence was annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (9). A total of 2,754 open reading frames (ORFs) were detected and included the genes *mecA* [staphylococcal cassette chromosome *mec* (SCC*mec*) type IVa(2B), as determined using SCC*mec*Finder v1.2 (<https://cge.cbs.dtu.dk/services/SCCmecFinder/>)], and *blaZ* (beta-lactamase), as well as 59 tRNAs and a set of six 5S, five 16S, and five 23S rRNAs. A 4,397-bp plasmid was also present. A BLASTN analysis (10) showed 99% identity with the plasmid pRIVM4390 previously isolated from other MRSA strains (11).

The presence of two prophages was detected *in silico* using PHASTER (12) at positions 337407 to 383619 and 2001104 to 2046421. The latter sequence matched to the genome

**Citation** Kittl S, Brodard I, Overesch G, Kuhnert P, Jores J, Labroussaa F. 2021. Complete genome sequences of the methicillin-resistant strain *Staphylococcus aureus* 17Gst354 and its prophage *Staphylococcus* phage vB\_staphS-IVBph354. Microbiol Resour Anounc 10: e00586-21. <https://doi.org/10.1128/MRA.00586-21>.

**Editor** Steven R. Gill, University of Rochester School of Medicine and Dentistry

**Copyright** © 2021 Kittl et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Fabien Labroussaa, [fabien.labroussaa@vetsuisse.unibe.ch](mailto:fabien.labroussaa@vetsuisse.unibe.ch).

**Received** 8 June 2021

**Accepted** 14 June 2021

**Published** 8 July 2021

sequence of *Staphylococcus* phage vB\_StaphS-IVBph354. This genome was assembled into one circular contig from a total of 4,789,426 paired-end short reads using Unicycler v0.4.4. The mean depth (coverage, 34,061×) was determined using the coverage command of SAMtools v1.11 (13) after mapping with the Burrows-Wheeler Alignment (BWA) tool v0.7.17 (14).

Phage vB\_StaphS-IVBph354 has a genome of 41,970 bp with a G+C content of 33% and includes 65 ORFs. Whole-genome alignment using EMBOSS Stretcher (15) showed that the vB\_StaphS-IVBph354 genome is identical to that of another  $\beta$ -hemolysin-converting integrase group 3 ( $\Phi$ Sa3) phage, *Staphylococcus* phage P282 (GenBank accession number [NC\\_048634.1](#)) (16), except that the vB\_StaphS-IVBph354 genome carries a classical *attB* site (5'-TGTATCCAAACTGG-3') and a frameshift insertion (5'-GAGCGAAAGA-3'), which extend the corresponding ORF (locus tag, HWA89\_gp54).

**Data availability.** The Illumina reads are available under the following SRA accession numbers: [SRX6491214](#) (strain 17Gst354) and [SRX10584153](#) (*Staphylococcus* phage vB\_StaphS-IVBph354). The PacBio reads are available under [SRX10576957](#). The assemblies can be accessed under the GenBank accession numbers [MW924889](#) (*Staphylococcus* phage vB\_StaphS-IVBph354), [CP073065](#) (strain 17Gst354), and [CP073064](#) (17Gst354 plasmid).

## ACKNOWLEDGMENTS

This work was supported by the University of Bern and the Swiss Federal Food Safety and Veterinary Office (Bern, Switzerland) (reference number 071-00000-75). The PacBio sequencing was performed at the Next Generation Sequencing Platform, University of Bern.

## REFERENCES

- Kittl S, Brodard I, Heim D, Andina-Pfister P, Overesch G. 2020. Methicillin-resistant *Staphylococcus aureus* strains in Swiss pigs and their relation to isolates from farmers and veterinarians. *Appl Environ Microbiol* 86: e01865-19. <https://doi.org/10.1128/AEM.01865-19>.
- Pitcher DG, Saunders NA, Owen RJ. 1989. Rapid extraction of bacterial genomic DNA with guanidium thiocyanate. *Lett Appl Microbiol* 8:151-156. <https://doi.org/10.1111/j.1472-765X.1989.tb00262.x>.
- Gutiérrez D, Martínez B, Rodríguez A, García P. 2010. Isolation and characterization of bacteriophages infecting *Staphylococcus epidermidis*. *Curr Microbiol* 61:601-608. <https://doi.org/10.1007/s00284-010-9659-5>.
- Center for Phage Technology. 2018. Protocol for phage DNA extraction with phenol:chloroform. <https://cpt.tamu.edu/wordpress/wp-content/uploads/2018/09/Phage-DNA-Extraction-by-PhenolChloroform-Protocol.pdf>.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data.
- Fukasawa Y, Ermini L, Wang H, Carty K, Cheung M-S. 2020. LongQC: a quality control tool for third generation sequencing long read data. *G3 (Bethesda)* 10:1193-1196. <https://doi.org/10.1534/g3.119.400864>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
- Hunt M, De Silva N, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. *Genome Biol* 16:294. <https://doi.org/10.1186/s13059-015-0849-0>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614-6624. <https://doi.org/10.1093/nar/gkw569>.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403-410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
- Bukowski M, Piwowarczyk R, Madry A, Zagorski-Przybylo R, Hydzik M, Wladyka B. 2019. Prevalence of antibiotic and heavy metal resistance determinants and virulence-related genetic elements in plasmids of *Staphylococcus aureus*. *Front Microbiol* 10:805. <https://doi.org/10.3389/fmicb.2019.00805>.
- Arndt D, Grant JR, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better, faster version of the PHAST phage search tool. *Nucleic Acids Res* 44:W16-W21. <https://doi.org/10.1093/nar/gkw387>.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25:2078-2079. <https://doi.org/10.1093/bioinformatics/btp352>.
- Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 25:1754-1760. <https://doi.org/10.1093/bioinformatics/btp324>.
- Myers EW, Miller W. 1988. Optimal alignments in linear space. *Comput Appl Biosci* 4:11-17. <https://doi.org/10.1093/bioinformatics/4.1.11>.
- Kraushaar B, Hammerl JA, Kienöl M, Heinig ML, Sperling N, Thanh MD, Reetz J, Jäckel C, Fetsch A, Hertwig S. 2017. Acquisition of virulence factors in livestock-associated MRSA: lysogenic conversion of CC398 strains by virulence gene-containing phages. *Sci Rep* 7:2004. <https://doi.org/10.1038/s41598-017-02175-4>.