

Amelioration of thioacetamide-induced liver toxicity in Wistar rats by rutin

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Abstract

This study was designed to evaluate the effect of rutin on hepatotoxicity induced by thioacetamide (TAA) in rats. Four groups of male Wistar rats consisting of six rats each were used: Group I: control group; Group II: rats receiving single injection of 300 mg kg⁻¹ body weight of TAA intraperitoneally; Group III: rats administered rutin (10 mg kg⁻¹ body weight) dissolved in saline orally for 2 weeks; and Group IV: rats administered rutin (10 mg kg⁻¹ body weight) dissolved in saline orally for 2 weeks followed by TAA injection last day of second week. All groups were sacrificed after 24 h of treatment and hepatic toxicity was analyzed with respect to liver toxicity markers, liver DNA fragmentation, and histology of liver tissue. Administration of TAA in Wistar rats resulted in significant increase of hepatic markers, DNA fragmentation in the hepatocytes, and changes in histology. Pretreatment of rats with rutin before 2 weeks of TAA assault resulted in the complete reversal of TAA-mediated hepatic toxicity ($P < 0.0001$ to $P < 0.01$) with concomitant restoration of DNA fragmentation. This study suggests rutin as a protective agent for restoration of toxicity caused by TAA.

Keywords

liver enzymes, oxidative stress, rutin, thioacetamide, toxicity

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Background

Thioacetamide (TAA) is an organosulfur, white crystalline compound having liver damaging and carcinogenic activity by causing cytomegaly.¹ It is used to induce an acute liver injury in rats. TAA gets metabolized to thioacetamide-S-oxide and acetamide immediately after administration to rats. Thioacetamide-S-oxide binds to macromolecules in a cell that are responsible for the change in cell permeability and Ca⁺⁺ uptake. This interruption of calcium stores increases nuclear volume, enlarges nucleoli, and inhibits mitochondrial activity eventually leading to hepatic necrosis.^{2,3} The toxicological studies have revealed the neurotoxicity of TAA in humans and animals.⁴ Generation of a large amount of reactive oxygen species (ROS) due to TAA can overwhelm the antioxidant defense mechanism and damage cellular ingredients such as lipids, proteins,

and DNA; this in turn can impair cellular structure and function.⁵ Regardless of the route of exposure, TAA rapidly distributes to nearby tissues and has also been reported to be a genotoxic, reproductive toxicant and rodent carcinogen.⁶ It is also reported that liver, kidney, spleen, and erythrocyte have significant binding capacity with TAA. Many studies

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were done to understand the changes elicited morphologically and biochemically that occur in the liver of TAA-treated rats.^{7–10}

Rutin has a strong antioxidant potential and is a derivative (glycoside) of quercetin. Based on the antioxidant potential of quercetin,^{11–14} it is hypothesized that rutin can have a significant therapeutic activity in diseases caused due to oxidative stress.¹⁵ Rutin is both water-soluble and alcohol-soluble antioxidant found in high amounts in plants maximally in citrus fruits. Rutin attaches to the iron ion Fe^{2+} , preventing its binding to hydrogen peroxide, and hence preventing the generation of a highly reactive free radical that can damage cells.¹⁶ Rutin has also been reported to inhibit the vascular endothelial growth factor *in vitro*.^{17,18} This in turn decreases the vascular permeability that is primary protection against certain injuries and inflammation. In this study, we, therefore, sought to evaluate the hepatoprotective role of rutin against TAA-induced oxidative stress.

Materials and methods

Chemicals

All the chemicals required in this study, including rutin and TAA, were purchased from Sigma Chemical Co., St Louis, USA. UV-kinetic diagnostic kits were procured from United Diagnostic Industry, Riyadh.

Animals

A total of 24 Wistar rats (male), weighing 100–120 g, were taken from the breeding laboratory of King Saud University and kept in departmental animal care facility. Rats were kept in polypropylene cages at room temperature with a 12 h light–dark cycle and relative humidity of $60\% \pm 15\%$. Animals were provided purified water *ad libitum* with standard laboratory rat chow. The study was approved by the animal ethics committee of Institutional Review Board of King Saud University, and the ethical guidelines for use and care of laboratory animals in experiments were strictly followed.

Experimental protocol

Total 24 rats were divided into four different groups consisting of six rats each. The groups were categorized as: Group I: control group; Group II: rats

receiving TAA; Group III: rats receiving rutin; and Group IV: rats receiving rutin before TAA treatment.

Group I: control group received vehicle alone (saline; pH 7.8); Group II: TAA single intraperitoneally injection of 300 mg kg^{-1} of body weight in week 2; Group III: rutin 10 mg kg^{-1} of body weight daily for 2 weeks; Group IV: rutin 10 mg kg^{-1} of body weight daily for 2 weeks followed by TAA injection on last day of rutin treatment. TAA and rutin both were dissolved in 1 mL of saline; TAA was administered intraperitoneally, whereas rutin was administered orally. TAA dosage was selected on the basis of previous literature.¹⁹ The rats were sacrificed 24 h after the last treatment by asphyxiation with carbon dioxide.

Sample preparation

After 24 h of treatment, blood from retro orbital plexus under anesthesia was collected in clean tubes, centrifuged at $3000g$ for 20 min at 4°C and serum was stored at -20°C until further analysis. All the groups of rats were sacrificed and the livers dissected, weighed, and placed in the Petri dishes. Right lobes of the excised livers from all groups were processed for histology; the rest of the liver samples were homogenized and stored in -80°C until further analysis.

Serum enzyme analysis

Measurement of transaminases. Aspartate aminotransferase (AST) and alanine aminotransferase (ALT) were measured by method of McPherson and Pincus²⁰ using UV-kinetic diagnostic kits. AST and ALT are both tissue enzymes that catalyze the exchange of amino and keto groups between alpha amino and keto acids. Tissue toxicity releases enzymes to general circulation, hence increasing their levels. AST and ALT activities were determined by the decrease in extinction of NADH followed at 340 nm in a coupling reaction. Each unit of enzyme activity was defined as micromoles of NADH decomposed per minute using molar absorbance of $6.22 \times 10^3 \times \text{M}^{-1} \text{Cm}^{-1}$.

Measurement of alkaline phosphatase. Alkaline phosphatase (ALP) was calculated by the method of Bowlers and McComb²¹ by a colorimetric kinetic diagnostic kit. Hydrolysis of *p*-nitrophenyl phosphate by ALP

produces inorganic phosphate and *p*-nitrophenol which at alkaline pH yields yellow *p*-nitrophenoxide ions measured by following the absorbance increase at 405 nm per unit time. One unit of ALP activity was defined as micromoles of *p*-nitrophenol produced per minute using molar absorbance of $18.75 \times 10^3 \times \text{M}^{-1} \text{Cm}^{-1}$.

Measurement of lactate dehydrogenase. The quantitative determination of lactate dehydrogenase (LDH) is achieved using lactate to pyruvate kinetic method using UV-kinetic diagnostic kit. Activity was evaluated by measuring the extinction decrease in NADH at 340 nm.²⁰ One unit of LDH activity was defined as micromoles of NADH decomposed per minute using molar absorbance of $6.22 \times 10^3 \times \text{M}^{-1} \text{Cm}^{-1}$.

Measurement of total bilirubin. The endpoint determination of total bilirubin was done by the method of Malloy and Evelyn²² using a colorimetric endpoint detection diagnostic. The intensity of purple color was directly proportional to bilirubin concentration in the serum recorded within 1 min for unconjugated bilirubin and after 5 min for conjugated bilirubin after adding methanol at 540 nm. The total bilirubin value was represented as the sum of bilirubin glucuronide (conjugated) and azobilirubin (unconjugated). Values were expressed as milligrams per deciliter.

Protein estimation

The protein content in the samples (both serum and tissue homogenate) was measured by the modified method of Markwell et al.²³ Bovine serum albumin (BSA) was used to make the standard curve. The amount of protein was calculated later from the obtained standard curve. Protein values were expressed as milligrams per deciliter.

Tissue analysis

DNA fragmentation assays for apoptosis. Apoptotic changes in the liver were evaluated colorimetrically by DNA fragmentation of known amount of DNA according to the procedure of Perandones et al.²⁴ Liver samples were homogenized in 700 μL hypotonic lysis buffer (0.2% Triton X-100, 10 mM Tris, 1 mM EDTA), incubated at 50°C for 2.4 h and centrifuged for 15 min at 11,000 r/min. The

supernatants contained small DNA fragments that were acid extracted and used for quantification by adding two volumes of diphenylamine (DPA) at 600 nm.

Histology

The right lobes of the excised livers from all groups were processed for histology. The processing was done as follows: fixation of the samples in a 10% neutral buffered formalin solution, block preparation in paraffin, section cutting (5–6 μm thick), and staining with hematoxylin–eosin stain. The processed sections were analyzed and photographed by a blinded expert pathologist without any prior information about the experimental groups.

Statistical analysis

Values were expressed as mean \pm standard deviation (SD). The data were generally statistically representative in terms of number, mean, and SD. Comparison between different groups was done using independent sample T-test for comparing two groups, and one-way analysis of variance (ANOVA) test was used for comparison between more than two groups with least significance difference as multiple comparison. Correlation between various variables was done by utilizing Pearson correlation coefficient (R) using linear regression with graphic representations. A probability value (*P* value) less than or equal to (0.05) was deliberated as significant. All statistical calculations were done using computer program SPSS (Statistical Package for Social Science) version (11.0).

Results

In the TAA-treated group (Group: II), the levels of all the liver marker enzymes were markedly elevated and there was an increase in total DNA fragmentation, indicating liver toxicity in this group of animals. In the other group (Group: IV) which received pretreatment of rutin, before TAA, the elevation of all the liver toxicity markers were reversed and DNA fragmentation was markedly reduced. Histopathological examination of liver sections showed the significant protection by rutin with no significant difference between control and rutin groups.

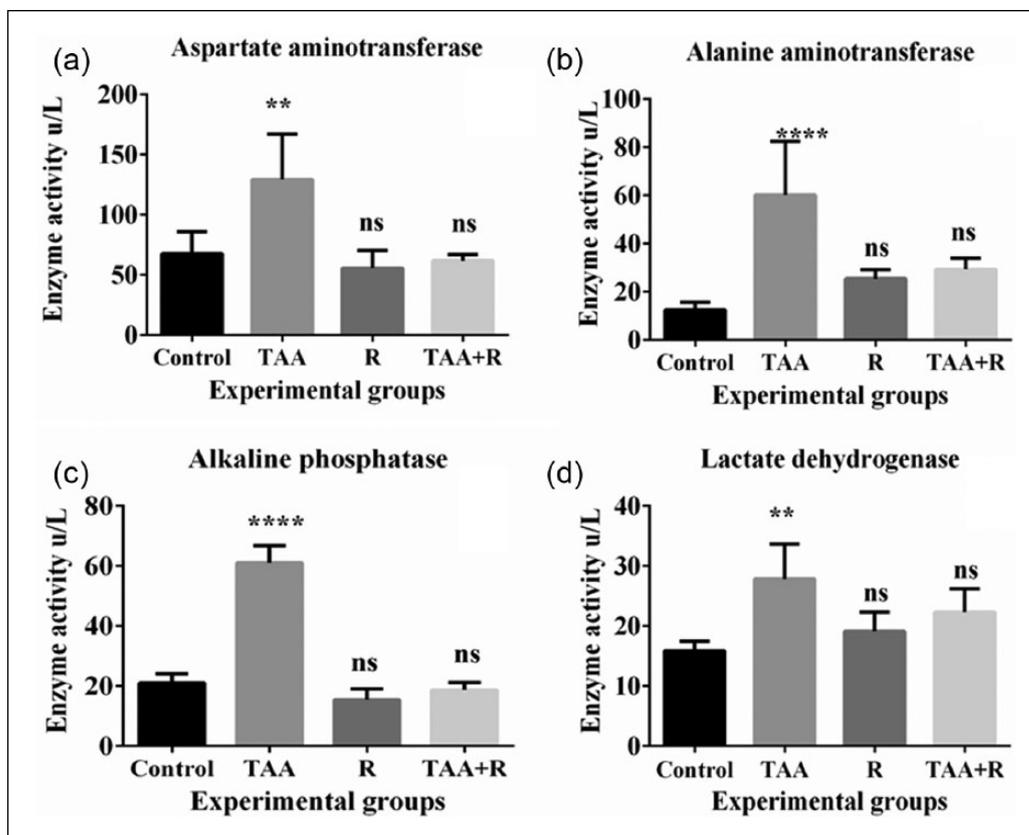


Figure 1. (a) ALT, (b) AST, (c) ALP, and (d) LDH enzyme activities in the control, thioacetamide (TAA), rutin (R), and TAA+rutin (TAA+R) groups in rat. Data between the groups were equated with an analysis of variance (ANOVA) and Tukey's multiple comparison tests.

**** $P < 0.0001$ when equated to control and ** $P < 0.01$ when equated to control ($n = 6$).

Figure 1(a) shows significant increased levels of AST in TAA-treated rats ($P < 0.0001$) when compared to control indicating the induction of liver damage. Rutin treatment alone caused non-significant decrease in the AST levels in rutin only treated rat group compared to control. Pretreatment of rutin in TAA group reversed the increased levels of AST and showed significant protection when compared with TAA-treated group. The decreased AST levels in group IV were insignificant compared to control group. Figure 1(b) shows increase in ALT levels by TAA significantly ($P < 0.0001$) and rutin non-significantly when compared to control. Rutin pretreatment followed by TAA assault reduced the levels when compared with TAA group and significant difference was observed when compared with this group and control group. It was depicted that rutin was completely protective with respect to ALT enzyme.

Figure 1(c) shows the significant increased levels of ALP in TAA group when compared to control. Pretreatment of rutin followed by TAA assault

showed significant protection as compared to TAA alone group ($P < 0.0001$). Figure 1(d) shows significant increase in levels of LDH in TAA alone treated groups ($P < 0.001$) and complete protection of LDH levels was observed in groups pretreated with rutin followed by TAA assault.

Figure 2(a) shows the concentration of bilirubin in all treated groups. There was significant effect observed on bilirubin content in TAA alone group ($P < 0.0001$); however, rutin alone treated group also showed significantly increased levels of bilirubin ($P < 0.001$). The increased levels reached to baseline when TAA was treated in combination with rutin. This further describes rutin as a protective flavonoid. Figure 2(b) shows the effect of TAA and rutin alone as well as together on hepatic protein content in rats of all groups. With the exposure to 300 mg TAA, protein levels were reduced significantly ($P < 0.01$) when compared to control. Rutin alone as well as along with TAA groups did not show any significant difference in hepatic protein content when compared to control.

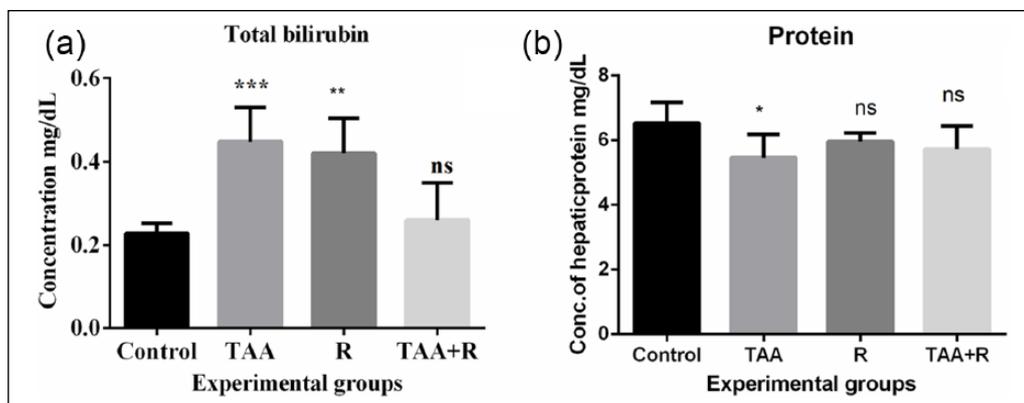


Figure 2. (a) Total bilirubin and (b) protein concentrations in the control, thioacetamide (TAA), rutin (R), and TAA + rutin (TAA + R) groups in rat. Data between the groups were equated with an analysis of variance (ANOVA) and Tukey's multiple comparison tests.

** $P < 0.001$ when compared to control ($n = 6$); *** $P < 0.0001$ when compared to control.

Figure 3 shows the extent of DNA fragmentation in all treated groups. DNA fragmentation was significantly increased in TAA-treated group, but complete protection of DNA strands was observed in groups pretreated with rutin pretreated group followed by TAA toxicity.

Figure 4 shows the changes produced by treatments with TAA and rutin alone and in combination. Normal rat liver hepatocytes have well-defined cytoplasm and nuclei. Treatment with TAA caused widespread intracellular vacuolization and aster (infiltration of inflammatory cells). Mild congestion was observed with congested portal area. Rutin-treated liver had nodular widened portal area with less inflammatory cell infiltration. Liver section from rutin pretreated group followed by TAA assault exhibited less changes in morphology as compared with TAA alone treated group. This group exhibited mild vacuolization and infiltration of inflammatory cells in portal area. Hepatocytes have dark eosinophilic cytoplasm with cells having heterochromatic nuclei.

Discussion

Liver, the largest glandular vital organ, plays various roles in regulating physiological processes. On the other hand, liver diseases have become the major causes of deaths all over world. Among them, drug-induced liver injury is one of the common causative factors that poses a major clinical and regulatory challenge.^{10,25} TAA at 300 mg kg^{-1} body weight as a single dose has been reported to cause oxidative stress.¹⁹ The aim of this study was

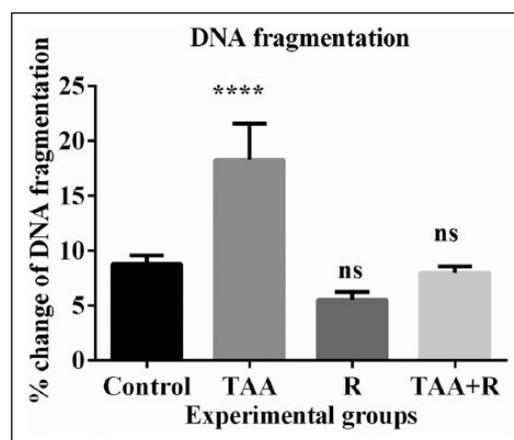


Figure 3. Percentage change of DNA fragmentation in the control, thioacetamide (TAA), rutin (R), and TAA + rutin (TAA + R) groups in rat. Data between the groups were compared with an analysis of variance (ANOVA) and Tukey's multiple comparison tests.

**** $P < 0.0001$ when compared to control ($n = 6$).

to examine the effect of flavonoid rutin against TAA-induced oxidative stress in rats. In this study, rutin caused significant reversal of increased liver enzyme activities, DNA fragmentation, and histopathological changes induced by TAA. Protein levels were reduced significantly in TAA-treated groups possibly due to the formation of protein adducts with TAA. TAA acts as an electrophilic agent and leads to formation of *s*-oxide that can covalently bind to the lysine residues forming adducts with sulfhydryl groups hence lowering the protein levels and causing significant damage. Protein adducts with TAA were also reported in previous studies.²⁶ This study reported the increase

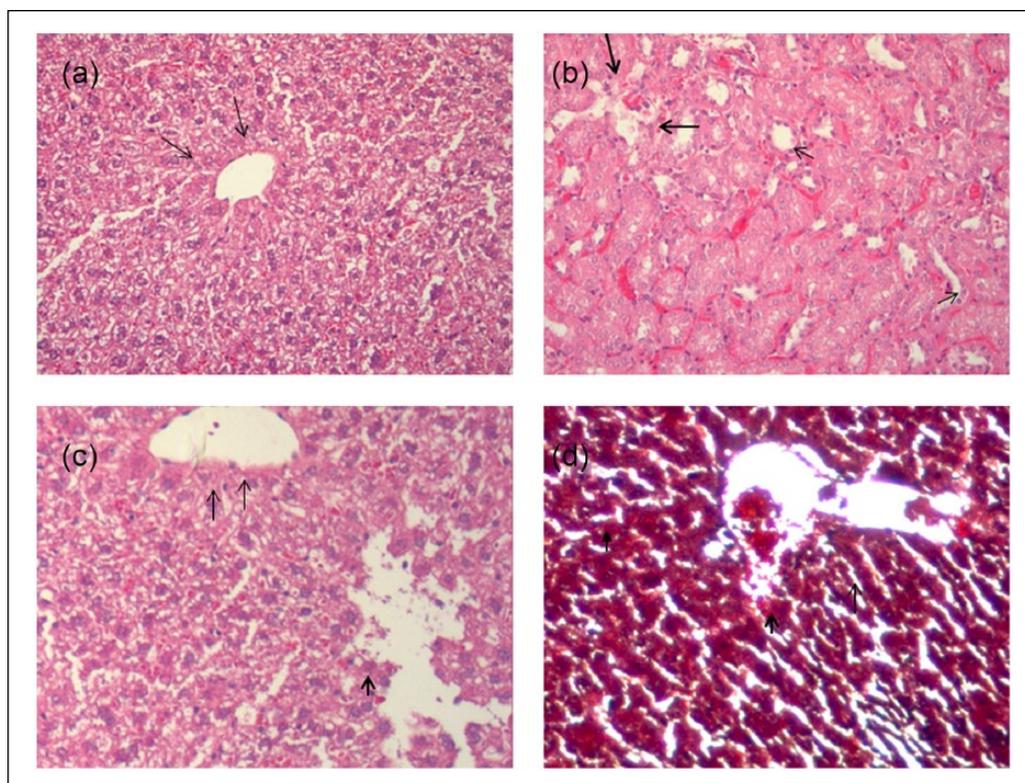


Figure 4. Liver sections of normal and treated groups at scale bar 100 μm . (a) Normal histological appearance of liver tissues with central vein and portal area shown by arrows. Hepatocytes have well-defined nuclei. (b) Thioacetamide-treated liver section with widespread intracellular vacuolization and infiltration of inflammatory cells (aster). Mild congestion is observed with congested portal area (thick arrows). (c) Rutin-treated liver section having nodular widened portal area with less inflammatory cell infiltration. (d) Rutin + TAA-treated liver section with mild vacuolization and mild infiltration of inflammatory cells in portal area (dark arrow). Hepatocytes have dark eosinophilic cytoplasm with cells having heterochromatic nuclei ($n=3$).

in liver transaminases (AST and ALT) and other enzymes LDH and ALP by TAA treatment which is concomitant to previous literature.⁵ Rutin exerted a powerful protective effect on the liver toxicity induced by different xenobiotics in other studies.²⁷ This study for the first time reports the significant protection by rutin pretreatment with TAA toxicity. Rutin at 10 mg kg^{-1} body weight for 2 weeks significantly restored the changes elicited by TAA assault on liver markers of injury and histology. When total bilirubin was evaluated in the control and treated groups, rutin alone group showed significant increase but when treated in combination the increased values reached to baseline showing the protective effect. These results were in accordance with previous study.²⁸ Bilirubin IX α has been recognized as a potent antioxidant. It is found to be toxic in infants but is considered to promote health in adults. Clinical literature from the last 10 years demonstrates that mildly increased serum bilirubin levels are significantly associated with decreased occurrence of chronic diseases,

including cardiovascular diseases (CVDs), and CVD-related mortality or risk factors.²⁹ In this study, rutin significantly reversed genotoxicity induced by TAA in liver tissue which was evaluated in terms of DNA fragmentation by DPA. The carcinogenicity of TAA has been demonstrated due to a disruption of function associated with spindle checkpoint immediately ahead of facilitation of cell proliferation.³⁰ However, rutin treatment alone had no effect on DNA fragmentation. Rutin probably caused remarkable changes in tissue morphology; however, it was at the same time protective too. Previously, we have shown quercetin as protective against acrylamide hepatotoxicity.³¹ In this study, we conclude that rutin which is a glycosidal derivative of quercetin has a hepatoprotective activity against TAA toxicity and prophylactic use of rutin against TAA poisoning is also suggested. These results contribute to the better understanding of rutin in the hepato-protection and reversal of genotoxicity, thus emphasizing its use in the human diet.

Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

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