

RESEARCH ARTICLE

Environmental assessment of cytotoxic drugs in the Oncology Center of Cyprus

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Abstract

Background

Cytotoxic drugs constitute an important workplace hazard in the hospital environment. Our aim was to conduct an environmental assessment of hazardous drugs in the Oncology Center of Cyprus.

Methods

Wipe samples were obtained from 42 workplace areas of the Oncology Center including two pairs of gloves in an initial assessment, while 10 samples were obtained at follow-up 3 years later. Potential contamination with cyclophosphamide (CP), ifosphamide (IF) and 5-fluorouracil (5-FU) and other cytotoxic medications was examined using the GC-MSMS system (CP, IF) and the HPLC system with UV detection (5-FU) method, respectively.

Results

Wipe sample contamination was detected at 11.9% and 15% in the initial and follow-up assessment, respectively. Both pairs of gloves assessed were free from contamination. The results showed contamination with cyclophosphamide on the work space inside the isolator, on a day-care office phone and on the central pharmacy bench. Ifosphamide was only detected on the floor of a patient's room. Contamination with 5-fluorouracil was found only on the surface of a prepared IV infusion bag. The levels of contamination in the positive samples ranged from 0.05 to 10.12 ng/cm².

Conclusions

The overall percentage of sample contamination at the Oncology Center was very low compared to other centers around the world. In addition, the detected levels of contamination with cytotoxic drugs were relatively low with the exception of the workspace inside the

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biological safety cabinet. These results in both assessments may reflect the implementation of comprehensive control measures including employee training, technological equipment and effective cleaning procedures.

Introduction

Anti-neoplastic agents constitute a significant workplace hazard for health professionals in the hospital environment [1–3]. Such hazardous drugs, used in the treatment of cancer, have been associated with many adverse health effects following employee acute and/or chronic cumulative exposure. Cytotoxic drugs have been particularly associated with reproductive toxicity as documented by several scientific publications in the international literature [4–6]. However, such reported reproductive toxicity was linked to higher levels of workers' exposure to such drugs, usually observed in past decades [usually at the level of milligrams per milliliter (mg/mL) or (mg/cm²)] compared to current levels of potential exposure observed nowadays [nanograms per milliliter (ng/mL) or (ng/cm²)] [7–9].

Environmental assessment and biological monitoring has been extensively used in order to estimate and quantify the potential workplace exposure of health professionals to anti-neoplastic agents and evaluate associated health risks [10, 11]. Several methods have been employed to measure the level of environmental contamination with hazardous drugs in relevant occupational settings. Collecting surface wipe samples from different sites of the hospital environment constitutes one of the most widespread method used for environmental assessment studies around the world [12–14]. Sugiura et al reported multi-center environmental monitoring studies evaluating cyclophosphamide exposure in Japan with levels of contamination ranging from 50% to 80% among all samples collected [15, 16]. In a study conducted in 6 British Columbian hospital pharmacies, 61% of the samples tested positive for contamination with Cyclophosphamide (CP) or Methotrexate (MTX). It is worth mentioning that contamination was detected sometimes even after cleaning, implying that the cleaning protocols in British Columbia hospitals required further improvement [17]. In Sweden, a similar study in a hospital pharmacy showed that CP or Ifosfamide (IF) were detected and quantified in 96–100% of wipe samples obtained from several areas in the preparation unit, with the highest values observed in the dressing room. However, after the cleaning procedures were reviewed and a second measurement was conducted several months later with samples taken from the same sites, results showed significantly lower levels of contamination [18]. Another study performed in two hospitals in France, where positive air pressure isolators were used, demonstrated much lower levels of contamination especially in areas outside the isolators [19].

In the US, between 2000–2005, the closed-system drug transfer device (CSTD) was introduced, and a study of environmental assessment was conducted in 22 hospitals. When standard drug delivery devices were used, the levels of contamination with positive wipe samples were 78%, 54% and 33% for Cyclophosphamide, Ifosfamide and 5-fluorouracil, respectively. With the introduction of the closed-system drug transfer device, there was a significant decrease in the levels of contamination to 68%, 45% and 20%, respectively [20]. Several other studies have shown significant reduction in the levels of occupational and environmental contamination as well as exposure to hazardous drugs following the implementation of a CSTD device [21–24].

In Cyprus, there has been no previous study evaluating the potential contamination of hospital environment with hazardous drugs in the oncology units. The objective of our study was

to assess the potential workplace contamination of the main oncology center of Cyprus with three most frequently used cytotoxic drugs, namely Cyclophosphamide, Ifosfamide and 5-fluorouracil. In addition, we conducted a follow-up assessment study evaluating potential contamination with a number of other cytotoxic drugs three years later.

Methods

Hospital setting

An environmental contamination assessment was conducted at the Bank of Cyprus Oncology Center (BOCOC) in 2011 and a follow-up assessment was repeated in 2014 in the context of a European comparative study between different hospitals. BOCOC is the main oncology center of the island providing care to about 70% of the cancer patients in Cyprus. The Center has 32 beds, has a workforce of about 200 employees, and provides outpatient cancer treatment to about 60 patients on a daily basis. It has two inpatient wards, a central pharmacy and an outpatient day care facility. Preparation of cytotoxic drugs is performed by trained nurses in a specifically designed unit equipped with two biological safety cabinets that are externally vented. The cleaning/decontamination protocols of our center provides for daily cleaning of all sections including the biological safety cabinets. The center is required to prepare about 70 separate treatment protocols on a daily basis for in-patient treatments and for patients attending the day-care therapy unit. The units of Cytotoxic treatments are administered by nurses to patients in the wards and also in a daycare unit. Between the two environmental assessments we were able to introduce the use of a closed system drug transfer device (CSTD) in our Center.

Wipe sample collection

Wipe samples were taken from 42 workplace surfaces including two pairs of gloves for the initial assessment, while in the follow-up assessment, a total of 10 samples were obtained however they were tested for a total of 12 different drugs. The initial wipe samples were taken using the Cyto Wipe Kits from Exposure Control B.V. Monitoring and Consultancy (The Netherlands) [25]. Samples were obtained from all departments of the oncology center (central pharmacy, outpatient pharmacy, chemotherapy pharmacy, day care unit, patient wards, radiotherapy department and administration offices). In November 2011, wipe samples were taken and gloves were collected by a nurse under the supervision of the head of the pharmacy department. The detection limits for the analysis of cyclophosphamide, ifosfamide and 5-fluorouracil were 0.10, 0.10 and 5 ng/mL extract, respectively. The samples were sent to Exposure Control B.V. for analyses. The methodology used for the collection of the follow-up samples in 2014 was quite similar although the samples were sent to the Institute of Energy and Environmental Technology, IUTA (Germany). In addition, the same health professionals were involved in the sample collection as in the initial assessment. The second assessment was performed in the context of a European comparative study between different hospitals in Europe. A permission to use the results for our Center from the second study has been obtained. Although fewer samples were used in the second assessment, the samples were taken from the same departments of the Oncology Center. Nevertheless, the sites of samples collected were not identical to the first environmental assessment campaign. The sites from both wipe sample assessments are described in detail in Tables 1 and 2, respectively. Follow-up analyses were performed for the following hazardous drugs: 5-fluorouracil, Gemcitabine, Methotrexate, Topotecan, Irinotecan, Doxorubicin, Epirubicin, Ifosfamide, Cyclophosphamide, Etoposide, Docetaxel, and Paclitaxel with the corresponding detection limits: 0.008, 0.003, 0.003, 0.003, 0.003, 0.003, 0.003, 0.008, 0.008, 0.008, 0.02, 0.02 ng/cm², respectively. The following drugs

Table 1. Results of a wipe-sample environmental assessment conducted in 2011 evaluating potential contamination with cyclophosphamide (CP), ifosphamide (IF) and 5-fluorouracil (5FU) at the Bank of Cyprus Oncology Center—Initial assessment.

Sample Code	Department	Description Surface	Area Surface (cm ²)	Total Volume NaOH (mL)	[CP] (ng/mL NaOH)	CP (ng)	CP (ng/cm ²)	[IF] (ng/mL NaOH)	IF (ng)	IF (ng/cm ²)	[5FU] (ng/mL NaOH)	5FU (ng)
1	Central Pharmacy	Front bench	2500	157	0.84	132	0.05					
2	Central Pharmacy	Trolley	1936	157	ND							
3	Central Pharmacy	Floor	2500	157							ND	
4	Outpatient Pharmacy	Pharmacy elevator 1 st shelf	2500	157	ND							
5	Outpatient Pharmacy	Working bench	2500	157	ND							
6	Chemotherapy Pharmacy	Telephone	125	143	ND							
7	Chemotherapy Pharmacy	Outside vial		143				ND				
8	Chemotherapy Pharmacy	Shelve inside cabinet	1950	157	ND							
9	Chemotherapy Pharmacy	Floor	2500	157							ND	
10	Day Care	Staff kitchen—top of fridge next to microwave	1100	157	ND							
11	Day Care	Exit doors to waste bins	900	152	ND							
12	Day Care	Reception floor next to information booklets	2500	157	ND							
13	Day Care	Room A nursing desk	2500	157							ND	
14	Day Care	Chemo transfer box	945	157							ND	
15	Day Care	Infusion pump 012	900	155	ND							
16	Day Care	Room B, couch arm chair, right hand corner A	850	150	ND							
17	Day Care	Couch junior doctor's office	2500	157							ND	
18	Day Care	Pair disposable gloves from checker clean room		120							ND	
19	Day Care	Aseptic Unit—bench in preparation room	2500	157							ND	
20	Day Care	Prepared 50 ml syringe CP		143	ND							
21	Ward A	Prepared 1000 ml infusion bag 5FU		145							21.6	3132
22	Day Care	Inside transfer hatch from prep room to isolator	2500	157	ND							
23	Day Care	Floor under foot rest isolator	2500	157							ND	
24	Day Care	Work space inside isolator	2500	157	161.15	25301	10.12					
25	Day Care	Outpatient main reception desk end nearest corridor	2500	157							ND	

(Continued)

Table 1. (Continued)

Sample Code	Department	Description Surface	Area Surface (cm ²)	Total Volume NaOH (mL)	[CP] (ng/mL NaOH)	CP (ng)	CP (ng/cm ²)	[IF] (ng/mL NaOH)	IF (ng)	IF (ng/cm ²)	[5FU] (ng/mL NaOH)	5FU (ng)
26	Day Care	Outpatient consultation office phone room 324	125	143	0.41	59	0.47					
27	Day Care	Blue chemotherapy tray after washing	806	157	ND							
28	Ward A	Nursing station A	1500	157				ND			ND	
29	Ward A	Drug preparation area A	2500	157				ND				
30	Ward B	Drug preparation area B	2500	157							ND	
31	Ward A	Bed patient 's room (29)	120	150							ND	
32	Ward A	Floor patient's room (29)	2500	157				4.15	652	0.26		
33	Ward B	Infusion pump with stand 085	2500	157							ND	
34	Ward B	Floor patient's toilet (43)	2500	157							ND	
35	Ward B	Cap patient's toilet (43)	900	148							ND	
36	Ward	Desk junior doctor's office	2500	157							ND	
37	Ward	Waste bin room 44	1849	157							ND	
38	Ward A	Pair of gloves after administration 5FU		140							ND	
39	Ward	Table staff kitchen	2500	157							ND	
40	Administration	Front reception	2500	157	ND							
41	Administration	Fridge chemotherapy waste	1600	157	ND							
42	Radiotherapy	CT scan	2500	157	ND							

ND: Not Detected: Levels of Detection for CP and IF were < 0.10 ng/mL NaOH; and for 5FU were < 5.00 ng/mL NaOH.

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were selected in order for our study to be comparable to the European-wide environmental assessment study in which we participated at follow up. There was no change in environmental controls (hoods, rooms etc.) from the first wipe to the second wipe sample assessment.

Storage, transportation and analysis of samples

On the initial assessment, all samples were stored frozen after sampling and during transport until sample preparation and analysis. The wipe samples were prepared by adding 140 mL of a 0.03 M NaOH solution. For the gloves, 120 or 140 mL solution was used. All samples were frozen right after collection and were sent to Exposure Control B.V., The Netherlands, where analyses were performed. Cyclophosphamide and ifosfamide were analysed using a Gas Chromatography–Mass Spectrometry (GC-MS) method on a GC-MSMS system showing increased sensitivity and specificity [26]. The analysis of 5-fluorouracil was performed on a High Performance Liquid Chromatography (HPLC) system with UV detection. The second environmental assessment was also performed using LC-MS/MS. The results were reported in nanograms per centimeters squared (ng/cm²).

Results

The results of the initial environmental assessment analyses of the wipe samples are presented in Table 1. The study findings on the initial assessment show contamination with

Table 2. Results of a wipe-sample environmental assessment conducted in 2014 evaluating potential contamination with 10 cytotoxic drugs at the Bank of Cyprus Oncology Center—Follow up assessment.

Sample Code	Department	Description Surface	Area (cm ²)	5-FU* ng/cm ²	Gemcitabine ng/cm ²	Methotrexate ng/cm ²	CP* ng/cm ²	Irinotecan ng/cm ²	Topotecan ng/cm ²	IF* ng/cm ²	Doxorubicin ng/cm ²	Epirubicin ng/cm ²	Etoposide ng/cm ²	Docetaxel ng/cm ²	Paclitaxel ng/cm ²
1	Clean room	Isolator	900	4.8	11	0.01	6.3	0.58	ND	0.093	ND	ND	ND	0.051	0.92
2	Day Care	Trolley	900	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
3	Ward	Lid of cytotoxic container	900	0.014	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
4	Day Care	Arm chair (pillow)	900	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
5	Day Care	Floor area under infusion stand	900	ND	0.0054	ND	0.17	ND	ND	ND	ND	ND	ND	0.91	0.037
6	Ward B	Telephone	900	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
7	Clean Room	Floor under the Isolator	900	ND	0.011	ND	0.011	ND	ND	0.21	ND	ND	ND	0.084	0.062
8	Pharmacy	Top checking counter	900	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
9	Day Care-Clean Room	Refrigerator door	900	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
10	Ward B	Top of checking counter	900	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND

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cyclophosphamide on the work surface inside the isolator as expected (biological safety cabinet), on an office phone at the daycare unit, and on the front of the bench in the central pharmacy. Ifosphamide was only observed on the floor of a patient's room at Ward A. Contamination with 5-fluorouracil was not found in the environment. A diluted IV bag was contaminated with 5-fluorouracil on the outside surface of the bag. Except for the work space inside the isolator and the IV bag, the levels of contamination were very low. The two pair of gloves examined were not found to be contaminated with 5-fluorouracil. Overall, only 11.9% of the samples were tested positive for any of the three cytotoxic drugs evaluated.

The results of the follow-up assessment are presented in Table 2. The follow-up assessment included a total of 12 different medications on 10 environmental samples. Based on the total number of samples multiplied by the total number of drugs tested, we found 18 positive samples with a percentage of contamination at 15% of the total samples tested. The lid of cytotoxic waste container tested positive for contamination with 5-FU, while the floor area under the infusion stand at the daycare unit tested positive for Gemcitabine, Cyclophosphamide, Docetaxel and Paclitaxel.

Discussion

The results of both environmental assessments showed quite low and similar levels of contamination in the Oncology Center of Cyprus. In the initial assessment, the contamination with cyclophosphamide or ifosphamide was detected mainly at the daycare unit and to a lesser extent in the central pharmacy and patient wards. Spread of contamination was not observed. The contamination with cyclophosphamide in the space inside the isolator, was expected to be relatively high, however the levels of contamination on the other positive samples were very low. Contamination with 5-fluorouracil was not found in the environment. This may probably be attributed to a higher detection limit for the analysis of 5-fluorouracil compared to cyclophosphamide and ifosphamide. Overall the percentage of positive samples was much lower

compared to other international scientific reports [11, 13, 15, 17, 18, 27–29]. Low levels of contamination were also observed on the follow-up assessment conducted three years later. The difference between the two assessments is not significant (12% vs 15%) and could most likely be attributed to the small number of samples used in the second assessment and the much bigger number of drugs implicated in the testing. The introduction of environmental assessment for many more cytotoxic drugs in the follow-up assessment (12 drugs compared to 3 in the first assessment) probably had an influence on our results with respect to the percentage of positive samples. We believe that the two assessments in general provide a similar picture of the low-level contamination in our Oncology Center. Except from the contamination detected in the clean room isolator and the floor underneath it, which was expected, only two other samples tested positive (the lid of cytotoxic waste container and the floor area under the infusion stand at the daycare). A sample obtained from a telephone on Ward B was negative for contamination on the follow-up assessment, compared to the wipe sample from a telephone in an office room that tested positive for contamination on the first assessment. The initial result was most likely related to unsafe practices of health professionals.

We believe that the low overall contamination levels with hazardous drugs seen in our study, may partly be attributed to the relatively small size of the Oncology Center where employees interact and cooperate within a somewhat family environment and are being concerned not only for their own health but also for their colleagues. It could also be attributed to the systematic and repeated training of nurses and pharmacists taking place in our Center regarding on cytotoxic drug management, which is being provided due to quality accreditation requirements. The training is provided on an annual basis and personnel are expected to attend all educational sessions. Furthermore, physicians are also expected to attend such training and all junior doctors do attend these training sessions. In addition, supervision of cleaning practices is done on a daily basis. The training consisted of two hour presentations from the head of pharmacy and the Occupational physician of the Center and was delivered to all nurses, pharmacists, and junior physicians of the Center. It concluded with a question/answer session.

The observed environmental contamination indicates several potential sources. A well-documented source of contamination is associated with spillage during preparation and administration of the cytotoxic drugs inside the isolator. This is also supported by the contamination found on the prepared IV infusion bag in the initial assessment and the floor area under the infusion stand in the follow-up study. In addition, another source of potential contamination includes the fact that the external vials may also be contaminated [30, 31]. Although somewhat expected, contamination inside the isolator and on the pharmacy bench can easily be transferred to prepared IV bags and further spread into the hospital environment. A similar limited spread was the observed contamination of the medical office phone found in the first assessment. Such findings support the need for health professionals to use gloves whenever they come into contact with prepared IV bags and other potential contaminated products, surfaces and/or vials associated with hazardous drugs. In addition, comprehensive cleaning protocols are essential [32, 33]. The rather high level of contamination on the workspace inside the isolator at the daycare unit and on the prepared IV infusion bag requires further investigation. Prevention and/or control of spillages in the isolator and the IV bags may be achieved through continuous employee training and comprehensive cleaning procedures, which would limit consequent spread of contamination with cytotoxic drugs in the hospital environment. Providing comprehensive cleaning of the isolator not only at the end of the day but also on an ad hoc basis related to spillages inside the isolator, is likely to minimize and/or eliminate spread of contamination and subsequent exposure of health professionals to such cytotoxic drugs.

Some limitations of our study need to be acknowledged. Although this is the first study conducted in an oncology hospital in Cyprus, we lack reference data from other local hospitals in

order to perform useful comparisons. In addition, due to cost limitations, we have used a relatively small number of environmental samples however we believe that the number is sufficient to obtain a comprehensive evaluation given the relatively small size of the Oncology Center. Our study was only focused on the environmental assessment for cytotoxic drugs. The samples from the two environmental assessments were analyzed in two different international laboratories using slightly different methodologies for detection. Therefore, direct comparison of the results of the two assessments should be viewed with caution. In addition, although the assessment samples were obtained from the same departments of the Oncology Center, they were not collected from identical sites used between the first and the second assessment campaign and therefore the comparison of the levels of contamination found should also be done with caution. The contamination was reported in nanograms per cm^2 and was calculated assuming 100% recovery of the sampling process and wipe efficiency. Based on the above, we may consider our results to represent potential underestimates of the true levels. The fact that the introduction of the closed system drug-transfer device did not have a substantial impact on decreasing the levels of positive samples in our Center requires further investigation. Perhaps the levels of contamination were quite low to begin with in the first assessment and there was little room for improvement. In addition, the contamination was not significantly associated with spillages in the patients wards, which are more likely to be related to the drug delivery devices. Finally, our study could have benefited from a parallel biological monitoring study to assess potential occupational exposure among health professionals.

In conclusion, this is the first study conducted in Cyprus examining the potential environmental contamination with cytotoxic drugs in an oncology hospital setting. Our results show a relatively low level of contamination compared to a number of similar studies around the world. The outcome of our study in both assessments as well as the relatively low levels of environmental contamination with hazardous drugs could be attributed to an array of workplace practices including the use of a specifically designed isolator unit with biological safety cabinets externally vented through a separate ventilation system for the whole isolator unit. Furthermore, the involvement of highly trained employees in the drug preparation processes who are also receiving annual refresher courses. The maintenance of a continuous monitoring system for dilution procedures run by trained pharmacy personnel could also play a role along with the adoption of the European Oncology Pharmacy protocol for monitoring spillages in the workplace. In addition to the above measures, the introduction of the closed system drug transfer device (CSTD) in our Center between the two environmental assessments may have played a role. Finally, we believe that the cleaning protocols followed in our Center including a daily routine cleaning of the isolator system are likely to have an impact to the low contamination levels found. The slightly higher percentage of the positive samples in the second assessment may reflect the smaller number of samples obtained and the much larger number of medications tested. We believe that the above combination of workplace control measures may constitute a good practice that could be followed by other centers around the world in order to lower potential contamination levels and reduce associated exposure of health professionals to hazardous drugs in the workplace.

Supporting information

S1 Data.
(XLSX)

S2 Data.
(XLSX)

Author Contributions

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References

1. WHO-IARC Monographs on the evaluation of the carcinogenic risk of chemicals to humans. Vol. 24: Some Pharmaceutical Drugs. 337 Seiten. International Agency for Research on Cancer, Lyon, Food / Nahrung 1982; 26(2):206–207.
2. NIOSH Alert. Preventing Occupational Exposures to Antineoplastic and Other Hazardous Drugs in Health Care Settings. Available from: <https://www.cdc.gov/niosh/docs/2004-165/pdfs/2004-165.pdf> [Accessed November 10th 2019].
3. NIOSH List of Antineoplastic and Other Hazardous Drugs in Healthcare Settings. Available from: <https://www.cdc.gov/niosh/docs/2016-161/pdfs/2016-161.pdf> [Accessed November 10th 2019].
4. Ahlborg G, Hemminki K. Reproductive Effects of Chemical Exposures in Health Professions. *Journal of Occupational and Environmental Medicine*. 1995; 37(8): 957–961. <https://doi.org/10.1097/00043764-199508000-00012> PMID: 8520960
5. Department of Health and Human Services, Centers for Disease Control and Prevention, National Institute for Occupational Safety and Health. Reproductive Risks Associated with Hazardous Drug Exposures in Healthcare Workers and Recommendations for Reducing Exposures. Available from: <https://www.cdc.gov/niosh/docket/archive/pdfs/NIOSH-279/CIBXX-reporRisk-279.pdf> [Accessed November 10th 2019].
6. Connor T, Lawson C, Polovich M, McDiarmid M. Reproductive Health Risks Associated With Occupational Exposures to Antineoplastic Drugs in Health Care Settings. *Journal of Occupational and Environmental Medicine*. 2014; 56(9): 901–910. <https://doi.org/10.1097/JOM.0000000000000249> PMID: 25153300
7. Connor T, McDiarmid M. Preventing Occupational Exposures to Antineoplastic Drugs in Health Care Settings. *CA: A Cancer Journal for Clinicians*. 2006; 56(6): 354–365.
8. Villarini M, Gianfredi V, Levorato S, Vannini S, Salvatori T, Moretti M. Occupational exposure to cytostatic/antineoplastic drugs and cytogenetic damage measured using the lymphocyte cytokinesis-block micronucleus assay: A systematic review of the literature and meta-analysis. *Mutat Res*. 2016; 770: 35–45. <https://doi.org/10.1016/j.mrrev.2016.05.001> PMID: 27894689
9. Boiano J, Steege A, Sweeney M. Adherence to Safe Handling Guidelines by Health Care Workers Who Administer Antineoplastic Drugs. *Journal of Occupational and Environmental Hygiene*. 2014; 11(11): 728–740. <https://doi.org/10.1080/15459624.2014.916809> PMID: 24766408
10. Turci R, Sottani C, Spagnoli G, Minoia C. Biological and environmental monitoring of hospital personnel exposed to antineoplastic agents: a review of analytical methods. *Journal of Chromatography B*. 2003; 789(2): 169–209.
11. Kibby T. A review of surface wipe sampling compared to biologic monitoring for occupational exposure to antineoplastic drugs. *J Occup Environ Hyg*. 2017 Mar; 14(3):159–174. <https://doi.org/10.1080/15459624.2016.1237026> PMID: 27676216
12. Jeronimo M, Colombo M, Astrakianakis G, Hon CY. A surface wipe sampling and LC-MS/MS method for the simultaneous detection of six antineoplastic drugs commonly handled by healthcare workers.

- Anal Bioanal Chem. 2015 Sep; 407(23):7083–92. <https://doi.org/10.1007/s00216-015-8868-y> PMID: 26141324
13. Marie P, Christophe C, Manon R, Marc M, Charleric B, Patrice V. Environmental monitoring by surface sampling for cytotoxics: a review. *Environ Monit Assess*. 2017 Jan; 189(2):52. <https://doi.org/10.1007/s10661-016-5762-9> PMID: 28063118
 14. Pretty JR, Connor TH, Spasojevic I, Kurtz KS, McLaurin JL, B'Hymer C, et al. Sampling and mass spectrometric analytical methods for five antineoplastic drugs in the healthcare environment. *J Oncol Pharm Pract*. 2012 Mar; 18(1):23–36. <https://doi.org/10.1177/1078155210389215> PMID: 21183556
 15. Sugiura S, Asano M, Kinoshita K, Tanimura M, Nabeshima T. Risks to health professionals from hazardous drugs in Japan: a pilot study of environmental and biological monitoring of occupational exposure to cyclophosphamide. *J Oncol Pharm Pract*. 2011; 17(1): 14–9. <https://doi.org/10.1177/1078155209358632> PMID: 20179165
 16. Sugiura S, Nakanishi H, Asano M, Hashida T, Tanimura M, Hama T et al. Multicenter study for environmental and biological monitoring of occupational exposure to cyclophosphamide in Japan. *J Oncol Pharm Pract*. 2011; 17(1): 20–8. <https://doi.org/10.1177/1078155210369851> PMID: 20472603
 17. Chu WC, Hon CY, Danyluk Q, Chua PP, Astrakianakis G. Pilot assessment of the antineoplastic drug contamination levels in British Columbian hospitals pre- and post-cleaning. *J Oncol Pharm Pract*. 2012; 18(1): 46–51. <https://doi.org/10.1177/1078155211402106> PMID: 21737485
 18. Hedmer M, Georgiadi A, Bremberg ER, Jönsson BA, Eksborg S. Surface contamination of cyclophosphamide packaging and surface contamination with antineoplastic drugs in a hospital pharmacy in Sweden. *Ann Occup Hyg*. 2005; 49(7): 629–37. <https://doi.org/10.1093/annhyg/mei042> PMID: 16126760
 19. Crauste-Manciet S, Sessink PJ, Ferrari S, Jomier JY, Brossard D. Environmental contamination with cytotoxic drugs in healthcare using positive air pressure isolators. *Ann Occup Hyg*. 2005 Oct; 49(7): 619–28. <https://doi.org/10.1093/annhyg/mei045> PMID: 16126757
 20. Sessink P, Connor T, Jorgenson J, Tyler T. Reduction in surface contamination with antineoplastic drugs in 22 hospital pharmacies in the US following implementation of a closed-system drug transfer device. *Journal of Oncology Pharmacy Practice*. 2011; 17(1): 39–48. <https://doi.org/10.1177/1078155210361431> PMID: 20156932
 21. Wick C, Slawson MH, Jorgenson JA, Tyler LS. Using a closed-system protective device to reduce personnel exposure to antineoplastic agents. *Am J Health Syst Pharm*. 2003; 60(22): 2314–20. <https://doi.org/10.1093/ajhp/60.22.2314> PMID: 14652980
 22. Yoshida J, Tei G, Mochizuki C, Masu Y, Koda S, Kumagai S. Use of a closed system device to reduce occupational contamination and exposure to antineoplastic drugs in the hospital work environment. *Ann Occup Hyg*. 2009; 53(2): 153–60. <https://doi.org/10.1093/annhyg/men081> PMID: 19261696
 23. Siderov J, Kirsa S, McLaughlan R. Reducing workplace cytotoxic surface contamination using a closed-system drug transfer device. *J Oncol Pharm Pract*. 2010; 16(1): 19–25. <https://doi.org/10.1177/1078155209352543> PMID: 19965949
 24. Bartel SB, Tyler TG, Power LA. Multicenter evaluation of a new closed system drug-transfer device in reducing surface contamination by antineoplastic hazardous drugs. *Am J Health Syst Pharm*. 2018 Feb 15; 75(4): 199–211. <https://doi.org/10.2146/ajhp160948> PMID: 29339374
 25. Sessink PJ, Anzion RB, Van den Broek PH, Bos RP. Detection of contamination with antineoplastic agents in a hospital pharmacy department. *Pharm Weekbl Sci*. 1992 Feb 21; 14(1): 16–22. <https://doi.org/10.1007/bf01989220> PMID: 1553250
 26. Colombo M, Jeronimo M, Astrakianakis G, Apte C, Hon CY. Wipe Sampling Method and Evaluation of Environmental Variables for Assessing Surface Contamination of 10 Antineoplastic Drugs by Liquid Chromatography/Tandem Mass Spectrometry. *Ann Work Expo Health*. 2017 Oct 1; 61(8): 1003–1014. <https://doi.org/10.1093/annweh/wxx070> PMID: 29028255
 27. Connor TH, Anderson RW, Sessink PJ, Broadfield L, Power LA. Surface contamination with antineoplastic agents in six cancer treatment centers in Canada and the United States. *Am J Health Syst Pharm*. 1999; 56(14): 1427–32. <https://doi.org/10.1093/ajhp/56.14.1427> PMID: 10428450
 28. Bigelow S, Schulz H, Dobish R, Chambers CR. Antineoplastic agent workplace contamination study: the Alberta Cancer Board Pharmacy perspective Phase III. *J Oncol Pharm Pract*. 2009; 15(3): 157–60. <https://doi.org/10.1177/1078155208101097> PMID: 19171554
 29. Maeda S, Miyawaki K, Matsumoto S, Oishi M, Miwa Y, Kurokawa N. Evaluation of environmental contaminations and occupational exposures involved in preparation of chemotherapeutic drugs. *Yakugaku Zasshi*. 2010; 130(6): 903–10. <https://doi.org/10.1248/yakushi.130.903> PMID: 20519870
 30. Hama K, Fukushima K, Hirabatake M, Hashida T, Kataoka K. Verification of surface contamination of Japanese cyclophosphamide vials and an example of exposure by handling. *J Oncol Pharm Pract*. 2012; 18(2): 201–6. <https://doi.org/10.1177/1078155211419543> PMID: 21947739

31. Fleury-Souverain S, Nussbaumer S, Mattiuzzo M, Bonnabry P. Determination of the external contamination and cross-contamination by cytotoxic drugs on the surfaces of vials available on the Swiss market. *J Oncol Pharm Pract.* 2014; 20(2): 100–11. <https://doi.org/10.1177/1078155213482683> PMID: [23676511](https://pubmed.ncbi.nlm.nih.gov/23676511/)
32. Hon CY, Chua PP, Danyluk Q, Astrakianakis G. Examining factors that influence the effectiveness of cleaning antineoplastic drugs from drug preparation surfaces: a pilot study. *J Oncol Pharm Pract.* 2014; 20(3): 210–6. <https://doi.org/10.1177/1078155213497070> PMID: [23929731](https://pubmed.ncbi.nlm.nih.gov/23929731/)
33. Viegas S, Oliveira AC, Carolino E, Pádua M. Occupational exposure to cytotoxic drugs: the importance of surface cleaning to prevent or minimise exposure. *Arh Hig Rada Toksikol.* 2018; 69(3): 238–249. <https://doi.org/10.2478/aiht-2018-69-3137> PMID: [30285944](https://pubmed.ncbi.nlm.nih.gov/30285944/)