


REVIEW

Tumor heterogeneity and the potential role of liquid biopsy in bladder cancer

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Abstract

Bladder cancer (BC) is a heterogeneous disease that characterized by genomic instability and a high mutation rate. Heterogeneity in tumor may partially explain the diversity of responses to targeted therapies and the various clinical outcomes. A combination of cytology and cystoscopy is the standard methodology for BC diagnosis, prognosis, and disease surveillance. However, genomics analyses of single tumor-biopsy specimens may underestimate the mutational burden of heterogeneous tumors. Liquid biopsy, as a promising technology, enables analysis of tumor components in the bodily fluids, such as blood and urine, at multiple time points and provides a minimally invasive approach that can track the evolutionary dynamics and monitor tumor heterogeneity. In this review, we describe the multiple faces of BC heterogeneity at the genomic and transcriptional levels and how they affect clinical care and outcomes. We also summarize the outcomes of liquid biopsy in BC, which plays a potential role in revealing tumor heterogeneity. Finally, we discuss the challenges that must be addressed before liquid biopsy can be widely used in clinical treatment.

KEYWORDS

bladder cancer, genomic heterogeneity, transcriptome heterogeneity, liquid biopsy, circulating tumor DNA

Abbreviations: APC, APC regulator of WNT signaling pathway; BC, bladder cancer; BRCA1, BRCA1 DNA repair associated; cfDNA, cell-free DNA; CT, computed tomography; CTCs, circulating tumor cells; ctDNA, circulating tumor DNA; EBCCs, exudative bladder cancer cells; EGFR, epidermal growth factor receptor; ERBB2, receptor tyrosine-protein kinase erbB-2; FGFR3, fibroblast growth factor receptor 3; FRa, folate receptor α ; GLOBOCAN, Global Cancer Observatory: CANCER TODAY; GSTP1, glutathione s-transferase pi 1; ITH, Intra-tumoral heterogeneity; lncRNA, long non-coding RNA; MIBC, muscle-invasive bladder cancer; miRNA, microRNA; NAC, neoadjuvant chemotherapy; NGS, next-generation sequencing; NMIBC, non-muscle-invasive bladder cancer; PD-L1, programmed death-ligand 1; PIK3CA, phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha; RAF1, Raf-1 proto-oncogene; TCGA, The Cancer Genome Atlas; TGF β , transforming growth factor β ; TIG1, tazarotene-induced gene 1; TP53, tumor protein p53; UC, urothelial cancer; UcfDNA, urinary cell-free DNA

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1 | INTRODUCTION

Bladder cancer (BC) is the 10th most common cancer worldwide [1]. According to the Global Cancer Observatory: CANCER TODAY (GLOBOCAN), three quarters of all BC cases are predominantly male and more than 90% of patients are over 50 years old [2]. BC is a highly heterogeneous malignancy, especially in advanced stage [3]. Approximately 75% of BC patients are diagnosed with non-muscle-invasive bladder cancer (NMIBC), whereas, 10%–25% eventually develop muscle-invasive bladder cancer (MIBC) [4]. Patients with MIBC are offered cisplatin-based neoadjuvant chemotherapy (NAC), which may prolong median overall survival by 13–14 months with a response rate of 50% [5–7]. Non-response patients may lose the opportunity for additional therapy with their disease progression. The use of immune checkpoint blockade continues to break new ground in the management of MIBC, however, there are still many patients cannot benefit from these therapies [7, 8]. With the advent of precision oncology, more and more molecular subtyping is increasingly recognized in BC [9–13].

Several lines of evidence suggest that tumor heterogeneity occurs on multiple levels can lead to the distinct clinical outcomes of NMIBC and MIBC. For example, gain-of-function mutations of fibroblast growth factor receptor 3 (*FGFR3*) are more prevalent in low-grade NMIBC [14], whereas mutations in DNA damage repair genes [12, 15] and somatic mutations in receptor tyrosine-protein kinase *erbB-2* (*ERBB2*) [16] were associated with an excellent response to NAC in patients with MIBC. Previous studies have proven that pancreatic ductal adenocarcinoma is the most common type of pancreatic cancer featured with high intra-tumoral heterogeneity (ITH) and poor prognosis [17], whereas other studies have shown a more modest level of ITH in lung cancer [18], or varying degree of ITH between patients with high-grade ovarian cancer [19]. However, studies in BC have shown a low level of ITH within individual biopsies, but a large difference between primary tumors and metastatic regions [20, 21]. The heterogeneity of BC results in great variation between different patients or regions of the same tumor tissue can lead to great differences in treatment efficacy and drug resistance [20, 21].

Since obtaining tumor tissue for biopsies is invasive and technically difficult, biopsies are limited to very few sampling points in time and accessible sites or metastatic sites, such as the lung, bone, and brain [22]. These limitations may fail to detect clinically relevant resistance mutations and pose a significant challenge to the development of BC treatment strategy [23]. The focus of precision oncology

is increasingly turning to liquid biopsy. Liquid biopsy is used to analyze biomarkers in various body fluids, including blood, urine, and saliva [24–26]. It is noninvasive and can be repeated at multiple points in time. Recent studies have shown that attempts are now being made to use liquid biopsy as an alternative strategy to understand heterogeneity in different kinds of cancer, such as lung [27], breast [28], gastrointestinal [29, 30], and colorectal cancers [31]. With the application of liquid biopsy in BC gradually maturing in recent years, numerous studies have shown that liquid biopsy may play an important role in the management of patients with BC at different stages [32–35].

Here, we provide a comprehensive overview of BC heterogeneity at the genomic and transcriptional levels, which may predict disease progression and therapeutic response and could eventually affect clinical decision. We then discuss the current applications of liquid biopsy in BC research and analyze its potential for identifying tumor heterogeneity in clinical routine. The many challenges that need to be overcome in the future are also described.

2 | HETEROGENEITY IN BC

Improved technology has allowed scientists and clinicians to characterize the heterogeneity of tumor at multiple levels. Moreover, multi-region sequencing was performed in many cancer types, including lung [36], breast [37], kidney [38–40], rectal [41], colorectal [42], prostate cancers [43], and BC [44], which provides the opportunity to expose the etiologies of treatment failure and drug resistance. At the molecular level, MIBC is a heterogeneous disease that is characterized by genomic instability and a high somatic mutation rate (median, 5.5 per megabase), similar to non-small cell lung cancer and melanoma [45]. This high mutation rate provides the fuel for tumor evolution and tumor heterogeneity, and eventually poses fundamental challenges for treatment remission. Therefore, it is essential to develop a comprehensive understanding of BC heterogeneity between tumors over time (Figure 1).

2.1 | Genomic heterogeneity

Insights into the genomic landscape of MIBC, the consequences of heterogeneity at the individual level have now been well recognized by the different molecular subtypes. An analysis of 412 tumors and matched normal samples revealed that patients with high APOBEC-signature mutagenesis, which was strongly correlated with high mutation burden, showed a better overall survival

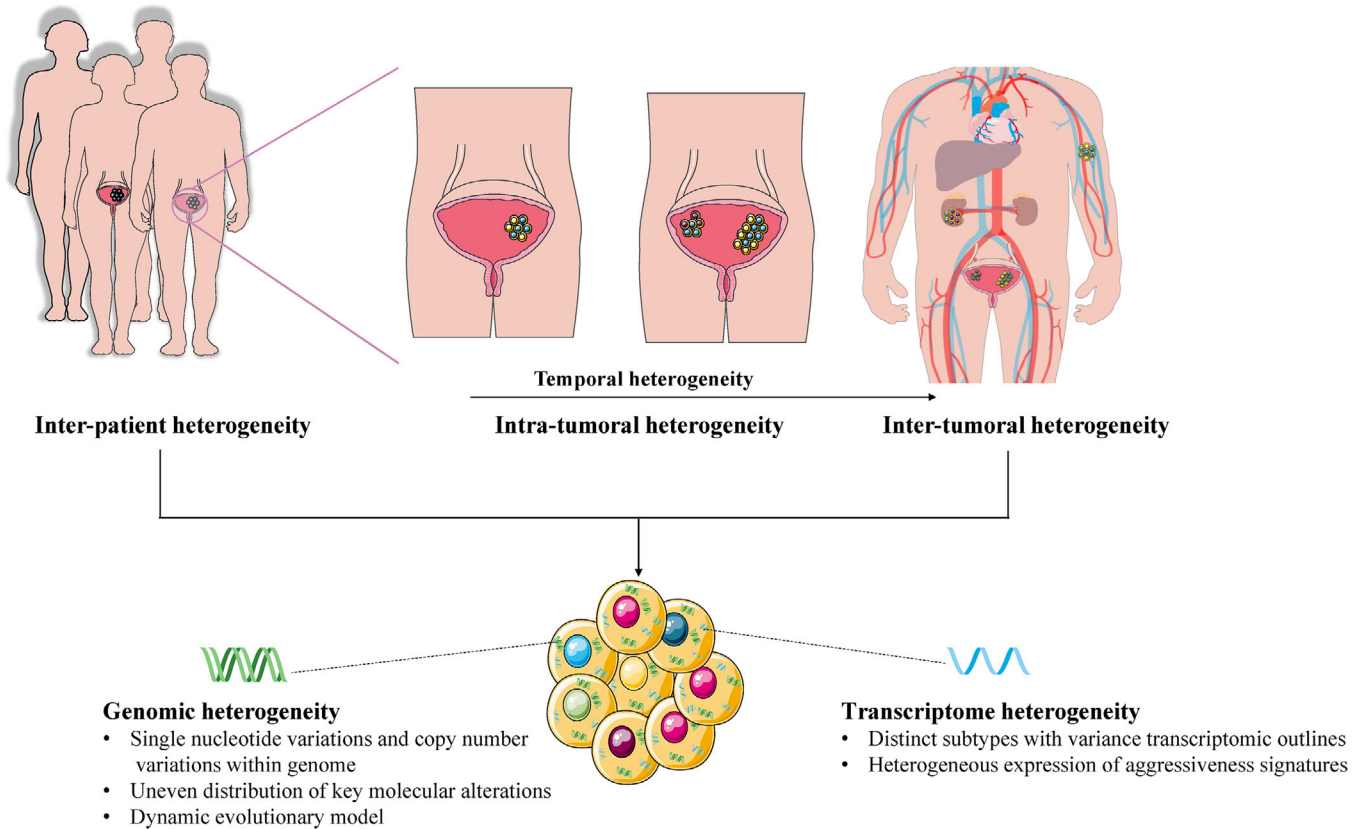


FIGURE 1 The multiple faces of bladder cancer heterogeneity. Bladder cancer is a heterogeneous disease that is characterized by genomic instability and a high mutation rate. In the disease course, bladder cancer generally becomes more heterogeneous. Inter-patient heterogeneity refers to heterogeneity between patients harboring tumors of the same histological type. In the same patients, tumor heterogeneity can be broadly divided into intra-tumoral and inter-tumoral heterogeneity. Intra-tumoral heterogeneity refers to the differences between distinct regions of one tumor. The variations between multiple primary tumors and/or metastatic sites are termed inter-tumoral heterogeneity. Temporal heterogeneity is present in tumor over time in the same patients. Colors denote the presence of sub-clones with different genomic and/or transcriptome features. Heterogeneity at the genomic level and the transcriptional level are also listed.

than those with other mutational signatures [46]. Moreover, next-generation sequencing (NGS) of 178 cancer-associated genes in 110 MIBC patients has demonstrated that *ERBB2* missense mutations are exclusively present in patients responding to NAC [16].

Current strategies for the treatment of MIBC or other cancers are typically relied on the biopsy of a single primary or metastatic site. In some multifocal cancers, such as breast [37], kidney [38], rectal [41], prostate cancers [43], and BC [44], the majority of point mutations detected in different fragments are frequently unique to a single fragment. These findings indicate that gene mutations found in single biopsies will not necessarily be representative of mutations presented in the entire tumor and ITH has substantial obstacle to appropriate selection of precision therapies. For example, in a recent study, Heide *et al.* [44] used multi-regional whole-exome sequencing of 10 whole cystectomy specimens from BC to show an uneven distribution of key molecular alterations across distinct areas within a tumor. It intends to support an evolutionary model

whereby synchronous development and parallel evolution of important alterations are followed in a dynamic rather than static way. This ongoing evolution might ultimately lead to inter-tumoral heterogeneity, meaning the differences between the primary tumor and metastases.

The transfer of disease from local organ to another part of the body is termed “metastasis”. Although distant metastasis is the leading cause of cancer-related deaths, current risk allocation and treatment recommendations for BC metastasis are primarily based on the histological and molecular characteristics of the primary tumor. Genomic studies in several types of tumors have revealed regionally diverse mutational landscapes between primary tumors and metastatic sites [47, 38, 48], and genetic differences between metastatic and primary tumors may affect treatment efficacy [49]. The study on the genetic difference between primary BC and local metastasis also showed that there were a distinct differences between primary tumor and metastasis in the degree of genetic differentiation [50, 20].

As multiregional sampling and blood-based liquid biopsies become widely studied, a high degree of genomic diversity across different regions (spatial heterogeneity) and over time (temporal heterogeneity) will be explained. However, the difference between primary tumor, metastatic, and circulating tumor markers currently remain to be elucidated.

2.2 | Transcriptome heterogeneity

The classification of MIBC subtypes based on gene expression has become increasingly clear, and several reports have highlighted the clinical significance of molecular stratification of MIBC [51–55, 46, 56]. Seiler *et al.* [55] reported that patients with basal tumors should be prioritized for NAC, they found the first single-sample classifier to subtype MIBC, which may be suitable for integration into routine clinical practice. Moreover, patients with metastatic urothelial cancer (UC) and patients who responded to the anti-programmed death-ligand 1 (PD-L1) agent were associated with CD8⁺ T-effector cell phenotype and high neoantigen or tumor mutation burden. Lack of response was associated with a signature of transforming growth factor β (TGF β) signaling in fibroblasts [57]. In recent years, The Cancer Genome Atlas (TCGA) database and other groups have identified multiple intrinsic subtypes of BC based on gene expression profiling (Table 1). These studies divided BC into distinct subtypes with variant transcriptomic outlines and the response to specific treatment types.

The study by Thomsen *et al.* [58] revealed inter-tumoral heterogeneity in various subtypes and aggressiveness signatures, and they also suggested that multiple subclones can occur within a MIBC patient. However, these subtype sets are not uniform and confusing. Kamoun *et al.* [59] performed a network-based analysis of six independent MIBC classification systems to reach a consensus on MIBC molecular subtypes (luminal papillary, luminal non-specified, luminal unstable, stroma-rich, basal/squamous, and neuroendocrine-like). Their results showed that the overall survival was directly associated with the subtypes. For example, patients with luminal papillary subtype tumors had a better outcome, and the neuroendocrine-like subtype tumors were associated with the worst prognosis. For guided therapy, basal/squamous subtype tumors expressed high levels of immune checkpoint markers and epidermal growth factor receptor (*EGFR*), which may be associated with sensitivity to immunotherapies and *EGFR*-targeted therapies [59]. Therefore, these studies demonstrated that responses to chemotherapy or immunotherapy may be enriched in specific MIBC subtypes.

3 | NONINVASIVE MONITORING OF BC HETEROGENEITY

Practical molecular stratification of tumors will be critical to guide the use of emerging targeted therapies and immunotherapy in BC. However, in the setting of significant inter-tumoral heterogeneity or ITH, a single-site biopsy may not be representative for the entire tumor [41]. Moreover, longitudinal or simultaneous multi-site testing is not feasible because continuous tissue sampling is invasive and impinges on quality of life. Cystoscopy and urine cytology remain the gold standard for BC diagnosis, prognosis, and disease surveillance; unfortunately, cystoscopy is invasive and urine cytology is limited by its low sensitivity (20%–53%), especially in low-grade tumor [60]. Existing urinary biomarkers for BC, such as the nuclear matrix protein 22 [61, 62] and bladder cancer antigen, which are Food and Drug Administration-approved, are not widespread adopted because of the lack of sensitivity for low-grade tumor and may result in false positives because of inflammation and hematuria [62, 61].

As liquid biopsy is considered a non-invasive and repeatable test that allows dynamic assessment of specific molecular markers, many efforts have been made to identify new circulating/urinary biomarkers including cell-free DNA (cfDNA), circulating tumor cells (CTCs), circulating microRNAs (miRNAs), and exosomes, which are capable of diagnosing diseases, monitoring recurrence, and potentially predicting treatment response. The ongoing studies on the clinical significance of liquid biopsy are summarized in Table 2.

3.1 | cfDNA

Substantial evidence has demonstrated that cfDNA originated from tumor cells contains tumor-specific DNA alterations, generally referred as circulating tumor DNA (ctDNA), plasma tumor DNA, or urinary cell-free DNA (UcfDNA), which can be detected in the blood or urine of patients with BC [63–69].

Hypermethylation of the CpG island of promoter regions causes tumor suppressor gene silencing, and it has been reported for numerous gene sites in BC, such as hypermethylation at APC regulator of WNT signaling pathway (*APC*) [70], glutathione s-transferase pi 1 (*GSTP1*) [71], prostaglandin-endoperoxide synthase 2 (*PTGS2*) [72], and Reprimo (*RPRM*) [73]. However, the gene sites mentioned above were performed single gene analysis, and sensitivity was limited at 18% to 48% [66]. Using a methylation-specific PCR, Ellinger *et al.* [66] detected hypermethylation in at least one of the 3 genes (*GSTP1*, tazarotene-induced

TABLE 1 Identification of multiple molecular subtypes through transcriptome sequencing analysis

Reference	Samples	Molecular subtype	Main findings
Robertson <i>et al.</i> [46]	412 MIBC	Luminal-papillary, Luminal-infiltrated, Luminal, Basal-squamous, Neuronal	<ul style="list-style-type: none"> Five molecularly distinct consensus molecular subtypes were identified with potential clinical utility; The luminal-infiltrated subtype had increased expression of several immune markers, including PD-L1 and PD-1; Loss of TP53 and RB1 was a hallmark of small cell neuroendocrine cancer, which had the poorest survival.
Tan <i>et al.</i> [51]	2411 NMIBC and MIBC	Papillary-like, HER2-like, Luminal-like, Neural-like, Mesenchymal-like, Squamous-cell carcinoma-like	<ul style="list-style-type: none"> Six molecularly distinct consensus molecular subtypes were identified with potential clinical utility; NMIBC with high risk of progression, displayed the molecular features of MIBC.
Warrick <i>et al.</i> [52]	309 BC co-occurring with conventional urothelial carcinomas	Urothelia-like, Genomically unstable, Basal-squamous, Mesenchymal-like	<ul style="list-style-type: none"> Four molecularly distinct consensus molecular subtypes were identified with potential clinical utility; BC was often molecularly heterogeneous, particularly in the basal-squamous subtype; Among patients with more than one tumor histology, 39% demonstrated molecular heterogeneity among the different tumor histologists.
Sjödahl <i>et al.</i> [54]	307 MIBC	Urothelial-like, Genomically unstable, Epithelial-Infiltrated, SCCL/Mesenchymal Infiltrated, SCCL/UroB, Small-cell/Neuroendocrine-like	<ul style="list-style-type: none"> Six molecularly distinct consensus molecular subtypes were identified with potential clinical utility; There was a systematic disagreement in subtype classification determined by global mRNA profiling and by immunohistochemistry profiling at the tumor-cell level; A combination of tumor cell phenotype and global mRNA analysis was suggested as a method for adequate subtype classification of MIBC.
Seiler <i>et al.</i> [55]	305 MIBC	Claudin-low, Basal, Luminal-infiltrated, Luminal	<ul style="list-style-type: none"> Four molecularly distinct consensus molecular subtypes were identified with potential clinical utility; Higher RNA-based immune signatures were significantly associated with improved CR and PFS outcomes after pembrolizumab, but not after NAC.
Efstathiou <i>et al.</i> [56]	259 MIBC	Luminal, Luminal-infiltrated, Basal, Claudin-low	<ul style="list-style-type: none"> Four expression signatures of immune infiltration of MIBC were identified with potential clinical utility; Higher immune infiltration in MIBC was associated with improved disease-specific survival after trimodality therapy, whereas higher stromal infiltration was associated with shorter disease-specific survival after NAC and RC.

Abbreviations: NMIBC, non-muscle-invasive bladder cancer; MIBC, muscle-invasive bladder cancer; PD-L1, programmed death-ligand 1; PD-1, programmed death-1; TP53, tumor protein p53; RB1, RB transcriptional corepressor 1; HER2, human epidermal growth factor receptor 2; SCCL, squamous-cell carcinoma-like; UroB, urothelial-like B; CR, complete response; PFS, progression-free survival; NAC, neoadjuvant chemotherapy; RC, radical cystectomy.

TABLE 2 Ongoing clinical trials on liquid biopsy in BC with/without therapeutic intervention

Reference	Trial no. [†]	Type of study/ starting date, study design	Status of the study	Solid tumors	Intervention/ treatment	Primary purpose	Estimated accrual/ country	Clinical findings
ctDNA								
NA	NCT04412070	Observational/2020, cohort, prospective	Not yet recruiting	BC	NA	Guide therapeutic decision	40, France	NA
NA	NCT04339933	Observational/2020, cohort, prospective	Not yet recruiting	BC	Diagnostic test: FGFR test	Predict treatment efficacy	92, Korea	NA
NA	NCT03837821	Interventional/2019, single group assignment	Recruiting	BC	Drug: abemaciclib	Predict treatment efficacy	20, US	NA
Park et al. [115]	NCT04197414	Observational/2019, cohort, prospective	Recruiting	BC, other solid cancers [‡]	NA	Differential diagnosis, monitor disease progression, predict treatment efficacy.	3000, Korean	SETD2 and DDXII mRNA can serve as non-invasive plasma biomarkers for predicting high-grade ccRCCs (AUC=0.971).
NA	NCT04138628	Interventional/2019, non-randomized, single group	Recruiting	BC, metastatic BC	Drug: atezolizumab	Monitor disease progression, predict treatment efficacy.	282, Denmark	NA
Rijnders et al. [116]	NCT03263039	Interventional/2017, single group assignment	Recruiting	TCC BC	Drug: pembrolizumab	Guide therapeutic decision	80, Nether- lands	The decrease of peripheral CD4 T cells expressing chemokine receptors is an early response marker during pembrolizumab treatment in mUC.
NA	NCT02546661	Interventional/2016, randomized, parallel assignment	Active, not recruiting	MIBC	<ul style="list-style-type: none"> • Drug: AZD4547 • Drug: MEDI4736 • Drug: olaparib • Drug: AZD1775 • Drug: vistusertib • Drug: AZD9150 • Drug: selumetinib 	Predict treatment efficacy	156, US	NA
NA	NCT03517332	Observational/2015, cohort, prospective	Unknow	BC, other solid cancers [§]	Diagnostic test: multiplexed primer and probe design developed	Assess process feasibility	10000, US	NA

(Continues)

TABLE 2 (Continued)

Reference	Trial no. ^a	Type of study/ starting date, study design	Status of the study	Solid tumors	Intervention/ treatment	Primary purpose	Estimated accrual/ country	Clinical findings
CTCs								
NA	NCT04280640	Observational/2020, cohort, prospective	Not yet recruiting	BC other solid cancers ^b	Other: blood draw	Predict treatment efficacy	40, US	NA
NA	NCT04358718	Interventional/2020, randomized, parallel assignment	Recruiting	BC	<ul style="list-style-type: none"> • Procedure: general anesthesia • Procedure: general anesthesia combined with epidural analgesia 	Monitor disease progression	58, China	NA
NA	NCT02716961	Interventional/2016, randomized, parallel assignment	Recruiting	Moderate-high risk NMIBC	Drug: gemcitabine, cisplatin	Predict treatment efficacy	208, China	NA
NA	NCT02080650	Interventional/2014, non-randomized, single group assignment	Completed	BC, other solid cancers ^c	<ul style="list-style-type: none"> • Device: mesenchymal-marker based ferrofluid • Device: EpCAM ferrofluid 	Assess device feasibility	62, US	NA
Choueiri et al. [117]	NCT01780545	Interventional/2013, randomized, parallel assignment	Completed	BC, UC	<ul style="list-style-type: none"> • Drug: OGX-427 • Drug: docetaxel 	Predict treatment efficacy	200, US	In the platinum-pretreated population of advanced UC, adding OGX-427 to Docetaxel provided a statistically significant improvement in OS.
Bellmunt et al. [118]	NCT01454089	Interventional/ October 2011, randomized, parallel assignment	Completed	Metastatic BC, urologic neoplasms, urinary tract neoplasms	<ul style="list-style-type: none"> • Drug: OGX-427 600 mg • Drug: OGX-427 1000 mg • Drug: placebo • Drug: gemcitabine • Drug: cisplatin • Drug: carboplatin 	Predict treatment efficacy	183, US	Advanced BC patients with poor prognosis benefited from apartorsen 600mg combined with first line GC. Apartorsen may be impacting the intrinsic biology of patients with poor risk factors.

(Continues)

TABLE 2 (Continued)

Reference	Trial no. [†]	Type of study/ starting date, study design	Status of the study	Solid tumors	Intervention/ treatment	Primary purpose	Estimated accrual/ country	Clinical findings
NA	NCT00829920	Observational/2008, cohort, prospective	Completed	BC	NA	Predict treatment efficacy	44, US	NA
NA	NCT02345473	Observational/2005, cohort, prospective	Completed	BC	Genetic: detection of circulating tumor cells in blood samples	Liquid biopsy characterization	59, Italy	NA
Cell-free RNA								
Abdelgawad et al. [119]	NCT03591367	Interventional/2018, single group assignment	Completed	NMIBC	<ul style="list-style-type: none"> • Diagnostic test: microRNAs-155 • Diagnostic test: hTERT 	Differential diagnosis	115, Egypt	As molecular urinary biomarkers: E2F3 (AUC=0.889) and hTERT (AUC=0.872) have the highest potential for prediction of the grade of NMIBC to either low or high grade.
Exosome								
NA	NCT04155359	Observational/2020, cohort, prospective	Recruiting	BC	NA	Differential diagnosis	3000, US	NA

Abbreviations: BC, bladder cancer; ctDNA, circulating tumor DNA; ccRCCs, clear cell renal cell carcinoma; FGFR, fibroblast-growth factor receptor; SETD2, SET domain-containing 2; DDX11, DEAD/H-box helicase II; TCC BC, transitional cell carcinoma of the bladder; mUC, metastatic urological cancer; EpCAM, epithelial cell adhesion molecule; UC, urological cancer; OS, overall survival; GC, gemcitabine and cisplatin; NMIBC, non-muscle-invasive bladder cancer; hTERT, human telomerase reverse transcriptase; E2F3, E2F transcription factor 3; AUC, area under curve; NA, not available.
[†]<http://clinicaltrials.gov/>.

[‡]Prostate cancer, renal cell cancer, ureter cancer.

[§]Colorectal cancer, pancreatic adenocarcinoma, gastric cancer, hepatocellular carcinoma, non-small cell lung cancer, melanoma, ovarian cancer, adrenocortical cancer, breast cancer.

[¶]Metastasis lung, metastasis to liver, gastrointestinal Cancer.

[‡]Prostate cancer, renal cell carcinoma, colorectal cancer, gastric cancer, pancreatic cancer, non-small cell lung cancer, advanced MET amplified solid tumor.

gene 1 (*TIG1*), and *APC*) in 80% of serum samples from BC patients undergoing cystectomy, with a specificity of 93% (Table 3). Furthermore, they investigated two small DNA fragments (124 bp *PTGS2* and 271 bp *RPRM* in serum DNA levels using the same cohorts, and an apoptosis index (ratio of 124 bp/271 bp fragments) has been identified to discriminate BC from benign prostate hyperplasia with high sensitivity (96%) and moderate specificity (62%) [68] (Table 3).

With the wide application of NGS technology, noninvasive testing of ctDNA in BC has been used to identify genomic aberrations and predict treatment response. For example, alterations in *BRCA1* DNA repair associated (*BRCA1*) and Raf-1 proto-oncogene (*RAF1*) genes appear to be negatively associated with clinical outcomes [63], and the sequencing results revealed high genomic concordance between the tissue DNA and ctDNA. Moreover, changes in ctDNA variant allele frequencies are closely correlated with duration of immunotherapy, antitumor activity, and clinical outcomes in BC [74]. A phase I trial conducted by Sundahl *et al.* [64] showed a rapid ctDNA fraction decline in patients who responded to treatment, whereas stable or increased fractions were detected in non-responders. This is the first trial to demonstrate that ctDNA fraction monitoring can be used to predict treatment response for metastatic urothelial carcinoma. Similarly, Birkenkamp-Demtroder *et al.* [65] observed that ctDNA levels of patients with metastatic relapse were significantly higher than those of patients without cancer. The median positive interval between ctDNA detection in plasma and diagnosis of relapse was 101 days after cystectomy, suggesting that ctDNA analysis may be more sensitive than computed tomography (CT) scanning in MIBC.

Besides ctDNA, urothelial tumors have close contact with urine, thus, urinary biomarkers are highly promising as noninvasive tools. Somatic mutations are reliably detected in urinary cell-pellet DNA and cfDNA [75–80], and the level of cfDNA in urine supernatants was associated with pathologic features and disease progression, which makes urine tumor DNA have great potential in clinical detection, especially in the early stages of BC [81, 82, 80, 79, 83, 84]. Current research on the clinical significance of UcfDNA detection for BC is increasing (Table 2).

Christensen *et al.* [83] found that NMIBC patients had high levels of cfDNA in serial urine supernatants, which was associated with later disease progression in NMIBC. Increased levels of *FGFR3* and Phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha (*PIK3CA*) mutated DNA in urine and plasma are indicative of later progression and metastasis in BC. Hirotsu *et al.* [81] compared detection rates of 71 UC commonly mutated genes in urine supernatants, urine precipitation, and plasma from 25 BC patients and 5 patients with cystitis and benign

tumor. The diagnostic sensitivity of the genetic analysis was higher in urine DNA (67%–78%) compared to cfDNA or conventional cytology (22%) in NMIBC. In addition, by using a high-throughput sequencing-based hybrid capture method, Dudley *et al.* [84] observed a high concordance for mutations between tumor and ucfDNA which enabled genotyping of multiple somatic aberration types across a broad genomic space in a single integrated assay, and they concluded that profiling of urine supernatants could have a significant value for BC early detection, sensitivity and specificity was achieved at 93% and 96% respectively (Table 3).

3.2 | CTCs

Although inter-tumoral heterogeneity, as reflected in different clinical phenotypes in different patients, has been partially explained with cfDNA, the cfDNA analysis cannot fully explain ITH because of its limited origin and accurate calling of copy-number aberrations [24].

Single-cell technologies now allow a comprehensive analysis of tumor cells in peripheral blood to unravel diverse aspects of metastasis, which can contribute to tumor heterogeneity. In some cases, it may be easier to distinguish tumor heterogeneity from CTCs than from the primary tumors. Recent molecular and clinical studies have shown that invasion may occur early in tumor development, and CTCs are released into circulation in the early stage of cancer [85]. Qi *et al.* [86] explored an effective strategy for enrichment, characterization, and quantification of CTCs based on the high expression of folate receptor α (*FR α*) in BC, which validated its diagnostic significance and demonstrated a modest sensitivity of 82.1% and specificity of 61.9%. This is so far the first study to confirm that *FR α* can be used as a tumor marker to detect CTCs in BC. New methods for isolating large numbers of CTCs are still being studied, which may improve the ability to perform a more comprehensive molecular analysis of most patients.

Currently, a preliminary study comparing CTC and ctDNA sample collection in 16 metastatic UC patients has found a similar detection rate for CTCs and ctDNA. However, the CTCs count was not correlated with cell-free DNA or ctDNA fraction, and several clinically actionable mutations were detected in plasma that were not found in the matching tumor [87]. Therefore, the researchers suggested that CTCs and ctDNA can provide complementary information; ctDNA may be more effective for early detection, while CTCs may be more suitable for studying the biological features of circulating malignant cells as well as protein expression at the single cell level. The presence of CTCs has also been proposed to be associated with poor prognosis,

TABLE 3 The main studies that have investigated the role of liquid biopsy biomarkers in the diagnosis and/or prognosis of BC

Reference	Biomarkers	Patients	Controls	Molecular targets	Method	Sample type	Sensitivity (%)	Specificity (%)	Clinical application
Vandekerkhove <i>et al.</i> [3]	ctDNA	51 MIBC (37 with metastatic disease)	None	50 BC driver genes	Targeted sequencing	Plasma	NA	NA	Prognosis
Birkenkamp-Demtroder <i>et al.</i> [65]	ctDNA	26 MIBC	None	84 personalized assays targeting 61 genes	Tumor-specific ddPCR assays	Plasma	NA	NA	Predict recurrence
Ellinger <i>et al.</i> [66]	cfDNA	45 BC	45 BPH	GSTP1, TIG1, APC	Methylation-specific PCR	Serum	93.0	80.0	Diagnosis
Ellinger <i>et al.</i> [68]	cfDNA	45 BC	45 BPH	PTGS2/RPRM ratios	qRT-PCR	Serum	96.0	62.0	Diagnosis
Patel <i>et al.</i> [69]	cfDNA, UcfDNA	17 MIBC	None	8 BC common mutated genes	TAm-Seq, sWGS	Plasma	NA	NA	Predict recurrence, predict treatment response
						Urine super-natants	NA	NA	
						UCP	NA	NA	
Xu <i>et al.</i> [76]	UcfDNA	189 BC	166 hematuria	IQGAP3/BMP4 and IQGAP3/FAM107A ratios	RT-PCR	Urine super-natants	71.0	88.6	Diagnosis
Stasik <i>et al.</i> [78]	UcfDNA	53 BC	36 HCs	Two abundant point-mutations (C228T/C250T) in the TERT promoter	NGS	Urine super-natants	63.0	100.0	Diagnosis
						Urine sediment	77.0	97.0	
Springer <i>et al.</i> [79]	Urinary cell DNA	570 patients at risk for BC	188 HCs	11 UC common mutated genes	UroSEEK [†]	UCP	83.0	93.0	Diagnosis
Springer <i>et al.</i> [79]	Urinary cell DNA	322 BC after surgery	188 HCs	11 UC common mutated genes	UroSEEK [†]	UCP	68.0	80.0	Predict recurrence
Ward <i>et al.</i> [80]	Urinary cell DNA	120 early-stage BC	20 no-cancer controls, 89 cancer-free NMIBC	6 BC associated genes	Multiplex PCR, NGS	UCP	70.0	97.0	Diagnosis
Hirotsu <i>et al.</i> [81]	cfDNA, UcfDNA	25 NMIBC	5 cystitis and benign tumor	71 UC common mutated genes	Targeted sequencing	Plasma	NA	NA	Diagnosis
						Urine super-natants	67.0	NA	

(Continues)

TABLE 3 (Continued)

Reference	Biomarkers	Patients	Controls	Molecular targets	Method	Sample type	Sensitivity (%)	Specificity (%)	Clinical application
Dudley <i>et al.</i> [84]	UcfDNA	118 early-stage BC	67 HCs	460-gene sequencing panel	CAPP-Seq	Urine sediment	78.0	NA	Diagnosis
Qi <i>et al.</i> [86]	CTCs	57 BC	48 HCs, 15 benign urologic pathologies	Folate receptor α	Ligand-targeted PCR	Whole blood	82.1	61.9	Diagnosis
Zhang <i>et al.</i> [97]	Exosome	260 BC	260 HCs	lncRNA: PCAT-1, UBC1, SNHG16	qRT-PCR	Serum	80.0-85.0	75.0-78.0	Diagnosis
Zhan <i>et al.</i> [98]	Exosome	184 BC	184 HCs	lncRNA: MALATI, PCAT-1, SPRY4-IT1	qRT-PCR	Urine	62.5-70.2	85.0-85.6	Diagnosis
Armstrong <i>et al.</i> [100]	Exosome	85 BC	45 HCs	miR-21, miR-93, miR-200c, miR-940	qRT-PCR	Urine	88.0	78.0	Diagnosis
Long <i>et al.</i> [101]	Exosome	34 BC	9 HCs	miR-375, miR-146a	qRT-PCR	Urine	-	-	Diagnosis
Murakami <i>et al.</i> [102]	Exosome	173 BC	36 no-cancer controls	mRNA: SLC2A1, GPRC5A, α	qRT-PCR	Urine	SLC2A1: 64.0 GPRC5A: 54.0	SLC2A1: 75.0 GPRC5A: 72.0	Diagnosis
Lin <i>et al.</i> [103]	Exosome	129 BC	62 HCs	protein: α 1-antitrypsin, H2BIK	MALDI-TOF	Urine	KRT17: 68.0	KRT17: 58.0	Diagnosis
Chen <i>et al.</i> [104]	Exosome	28 BC	12 hematuria	protein: TACSTD2	LC-MRM/MS	Urine	65.0	75.6	Diagnosis
Du <i>et al.</i> [120]	Exosome	230 BC	230 HCs	lncRNA: uc004cox.4, GASS	qRT-PCR	Urine	80.0-84.5	78.2-85.0	Diagnosis

Abbreviations: BC, bladder cancer; cfDNA, circulating tumor DNA; cfDNA, cell-free DNA; UcfDNA, urine cell-free DNA; BPH, benign prostate hyperplasia; PTGS2, prostaglandin-endoperoxide synthase 2; RPRM, reprimo; qRT-PCR, quantitative real-time PCR; GSTP1, glutathione *S*-transferase p11; TIG1, lazaretone-induced gene 1; APC, APC regulator of WNT signaling pathway; ddPCR, droplet digital PCR assay; MIBC, muscle-invasive bladder cancer; TAM-Seq, tagged-amplicon sequencing; sWGS, shallow whole genome sequencing; UCP, urine cell pellet; NMIBC, non-muscle invasive bladder cancer; UC, urological cancer; HCs, healthy controls; IQGAP3, IQ motif containing GTPase activating protein 3; BMP4, bone morphogenetic protein 4; FAM107A, IQGAP3/family with sequence similarity 107A; CAPP-Seq, cancer personalized profiling by deep sequencing; NGS, next-generation sequencing technology; PCAT-1, prostate cancer associated transcript 1; UBC1, upregulated in bladder cancer 1; SNHG16, small nucleolar RNA host gene 16; MALATI, metastasis associated lung adenocarcinoma transcript 1; SPRY4-IT1, sprouty receptor tyrosine kinase signalling antagonist 4-intronic transcript 1; GASS, growth arrest specific 5; SLC2A1, solute carrier family 2 member 1; GPRC5A, G protein-coupled receptor class C group 5 member A; KRT17, keratin 17; H2BIK, histone H2B type 1-K; MALDI-TOF, matrix-assisted laser desorption ionization time-of-flight; TACSTD2, tumor-associated calcium signal transducer 2; LC-MS/MS, isotopic demethylation labeling coupled with liquid chromatography-tandem mass spectrometry; NA, not available.

[†]UroSEEK: a massively parallel sequencing-based assay.

and the amount of CTCs found in blood is often associated with short-term disease-free survival of metastatic BC [88].

3.3 | Circulating miRNAs and others

Other tumor-derived products also exist in bodily fluids, such as cell-free miRNAs [60–62], long non-coding RNAs (lncRNAs) [63], exosomes or tumor-educated platelets [61], which are of concern as liquid biopsy analyses with potential clinical significance.

miR-210 was upregulated in BC tissues, and the levels of miR-210 increased with advancing stage and grade [89]. Moreover, miRNA expression levels can also reflect tumor dynamics in MIBC. Yang *et al.* [90] compared miR-210 levels in matched serum samples obtained before and after surgery, and the results showed a significant reduction of miR-210 after surgery. The levels of serum miR-210 from patients with relapsed BC were upregulated and reached the levels of those in pre-operative patients. In summary, these studies suggest that serum miR-210 could be a potential noninvasive biomarker for screening, predicting, and monitoring MIBC.

Exosomes are vesicles of endocytic origin and continuously released from the bladder [91], which can mediate communication between cells through the transfer of nucleic acids [92–95] and proteins [96] in BC. Numerous studies have been conducted to investigate the role of exosomes in patients with BC. The results are very heterogeneous, but promising candidate nucleic acids biomarkers have been identified with higher sensitivities and specificities than plasma ctDNA. The major studies that have investigated the role of exosomes in the diagnosis of BC are compiled in Table 3 [86, 97–104]. Although the specific functional role of exosomes in BC heterogeneity remains unclear, exosomes have emerged as potential noninvasive disease biomarkers with potential clinical significance.

4 | CONCLUSIONS AND FUTURE PERSPECTIVES

BC is a heterogeneous disease with higher tumor burden among cancers. Unfortunately, there have been few advances in its clinical management due to a poor understanding of the correlations between its molecular and clinical features. This tumor heterogeneity might ultimately provide the fuel for the emergence of treatment resistance and, eventually, disease relapse.

An alternative strategy to understand tumor heterogeneity might be liquid biopsy, which can detect genetic alterations and tumor cells present in peripheral blood or urine of patients with BC [105, 99]. Peripheral blood- or urine-

based genomic analyses were flexible in processing, and there are no specific device required for isolation, which have the potential to diagnose and dynamically monitor BC recurrence [82, 78, 106, 69, 65, 79]. Therefore, cfDNA analysis has now become one of the major methods to evaluate BC heterogeneity without the sampling bias of tissue biopsy. However, there are still remaining challenges of widely used cfDNA. At present, majority of studies on liquid biopsy in BC are based on ctDNA analysis and focus on patients at advanced stages. Since the use of ctDNA for early detection is probably very limited by the low detection rates in early-stage cancer, highly sensitive methods and sufficient sample volumes are needed to detect trace amounts of ctDNA [107, 108].

Compared to the peripheral blood collection, urine biopsy can be truly collected “non-invasively”, and sampling peripheral fluids in close proximity to diseased organs has been proved to improve the sensitivity of the detection of tumor mutated DNA [109, 69]. Therefore, urine-based genetic analysis is an ideal liquid biopsy for detecting tumor-derived DNA and may be more precise in reflecting tumor mutational profiles than plasma, especially in early-stage BCs [81, 69].

Attention should also be paid to the effects of confounders of apoptosis and aging on cfDNA analysis in non-tumor patients. For example, benign somatic heterogeneity may result from somatic mosaicism, which is the accumulation of mutations during development and aging, resulting in the production of cells with different genotypes in the same individual. Fernandez-Cuesta *et al.* [110] assessed the presence of tumor protein p53 (*TP53*) mutations detected at very low fractions in the cfDNA, and they detected *TP53* mutations in 49% small cell lung cancer patients and 11.4% of non-cancer controls. Therefore, detection of a mutation in a cancer driver gene in cfDNA cannot be equated with evidence of tumor.

In addition to genomic aberrations not covered by ctDNA analysis, CTC analysis provides unique insights into tumor heterogeneity, including transcriptional heterogeneity using single-cell RNA sequencing [111]. However, there is very low number of CTCs available in early stage disease, and different tumor regions within an individual patient might not present the same propensity of CTCs. Therefore, it is still unclear whether CTCs analysis could lead to a potential biological bias with the ITH. In the near future, besides the development of novel high-throughput technologies, the combined molecular characterization of ctDNA and CTCs or other liquid biopsy may provide complementary information, for example, combining ctDNA- and CTC-derived analyses can identify the T790M mutation in *EGFR*-mutant non-small cell lung cancer patients in whom the concurrent biopsy is negative or uncertain [112].

Other tumor products such as cell-free RNA or exosomes also have high sensitivity and specificity in the diagnosis of BC, whereas as exosomes can be released into the peripheral circulation by many types of cells, the key challenges is to discriminate exosomes derived from either tumor or normal cells.

Recent advances in detection and sequencing technology have contributed to clinical determination of genomic heterogeneity in subpopulations of BC patients. For example, the detection of invasive mesenchymal exudative bladder cancer cells (EBCCs) from urine using a microfluidic enrichment device has a high sensitivity (93.3%), therefore, the enumeration and cytological analysis of EBCCs may serve as a complementary tool for clinical real-time detection [113]. In nasopharyngeal cancer, a label-free and modification-free nanotechnology based on surface-enhanced Raman spectroscopy was employed for cfDNA analysis with an ideal diagnostic sensitivity of 83.3% and specificity of 82.5% [114]. These advancements have increased the sensitivity of non-invasive screening for early diagnosis. In addition, the extent of genomic and transcriptional heterogeneity at the individual cell level remains largely unknown within primary bladder tumors and metastases. Large-scale clinical trials involving various stages of BC will be needed to determine the clinical value of spatial and temporal variation in the genomic and transcriptional landscape of BC to guide clinical treatment.

DECLARATIONS

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

Not applicable.

CONSENT FOR PUBLICATION

Not applicable.

AVAILABILITY OF DATA AND MATERIALS

Not applicable.

COMPETING INTERESTS

The authors declare that they have no competing interests.

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AUTHORS' CONTRIBUTIONS

Conception: all authors. Manuscript writing: HM Huang. Manuscript revision: HX Li. All authors read and approved the final manuscript.

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REFERENCES

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 2018;68(6):394-424. <https://doi.org/10.3322/caac.21492>.
2. GLOBOCAN. <http://globocan.iarc.fr>.
3. Vandekerckhove G, Todenhofer T, Annala M, Struss WJ, Wong A, Beja K, et al. Circulating Tumor DNA Reveals Clinically Actionable Somatic Genome of Metastatic Bladder Cancer. *Clin Cancer Res.* 2017;23(21):6487-97. <https://doi.org/10.1158/1078-0432.CCR-17-1140>.
4. Sanli O, Dobruch J, Knowles MA, Burger M, Alemozaffar M, Nielsen ME, et al. Bladder cancer. *Nat Rev Dis Primers.* 2017;3:17022. <https://doi.org/10.1038/nrdp.2017.22>.
5. Pietzak EJ, Zabor EC, Bagrodia A, Armenia J, Hu W, Zehir A, et al. Genomic Differences Between "Primary" and "Secondary" Muscle-invasive Bladder Cancer as a Basis for Disparate Outcomes to Cisplatin-based Neoadjuvant Chemotherapy. *Eur Urol.* 2019;75(2):231-9. <https://doi.org/10.1016/j.eururo.2018.09.002>.
6. Grossman HB, Natale RB, Tangen CM, Speights VO, Vogelzang NJ, Trump DL, et al. Neoadjuvant chemotherapy plus cystectomy compared with cystectomy alone for locally advanced bladder cancer. *N Engl J Med.* 2003;349(9):859-66. <https://doi.org/10.1056/NEJMoa022148>.
7. Necchi A, Anichini A, Raggi D, Briganti A, Massa S, Lucianò R, et al. Pembrolizumab as Neoadjuvant Therapy Before Radical Cystectomy in Patients With Muscle-Invasive Urothelial Bladder Carcinoma (PURE-01): An Open-Label, Single-Arm, Phase II Study. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology.* 2018;Jco1801148. <https://doi.org/10.1200/jco.18.01148>.
8. Eckstein M, Sikic D, Strissel PL, Erlmeier F, Germany BC. Evolution of PD-1 and PD-L1 Gene and Protein Expression in Primary Tumors and Corresponding Liver Metastases of Metastatic Bladder Cancer. *Eur Urol.* 2018;74(4):527-9. <https://doi.org/10.1016/j.eururo.2018.06.028>.
9. Hong B, Yang Y, Guo S, Duoerkun S, Deng X, Chen D, et al. Intra-tumour molecular heterogeneity of clear cell renal cell carcinoma reveals the diversity of the response to targeted therapies using patient-derived xenograft models. *Oncotarget.* 2017;8(30):49839-50. <https://doi.org/10.18632/oncotarget.17765>.
10. Morrison CD, Liu P, Woloszynska-Read A, Zhang J, Luo W, Qin M, et al. Whole-genome sequencing identifies genomic heterogeneity at a nucleotide and chromosomal level in bladder cancer. *Proc Natl Acad Sci U S A.* 2014;111(6):E672-81. <https://doi.org/10.1073/pnas.1313580111>.
11. Ku SY, Gleave ME, Beltran H. Towards precision oncology in advanced prostate cancer. *Nature reviews Urology.* 2019. <https://doi.org/10.1038/s41585-019-0237-8>.

12. Vidotto T, Nersesian S, Graham C, Siemens DR, Koti M. DNA damage repair gene mutations and their association with tumor immune regulatory gene expression in muscle invasive bladder cancer subtypes. *Journal for immunotherapy of cancer*. 2019;7(1):148. <https://doi.org/10.1186/s40425-019-0619-8>.
13. Burrell RA, McGranahan N, Bartek J, Swanton C. The causes and consequences of genetic heterogeneity in cancer evolution. *Nature*. 2013;501(7467):338-45. <https://doi.org/10.1038/nature12625>.
14. Lee SH, Hu W, Matulay JT, Silva MV, Owczarek TB, Kim K, et al. Tumor Evolution and Drug Response in Patient-Derived Organoid Models of Bladder Cancer. *Cell*. 2018;173(2). <https://doi.org/10.1016/j.cell.2018.03.017>.
15. Yin M, Grivas P, Wang QE, Mortazavi A, Emamekhoo H, Holder SL, et al. Prognostic Value of DNA Damage Response Genomic Alterations in Relapsed/Advanced Urothelial Cancer. *Oncologist*. 2020. <https://doi.org/10.1634/theoncologist.2019-0851>.
16. Groenendijk FH, de Jong J, Fransen van de Putte EE, Michaut M, Schlicker A, Peters D, et al. ERBB2 Mutations Characterize a Subgroup of Muscle-invasive Bladder Cancers with Excellent Response to Neoadjuvant Chemotherapy. *Eur Urol*. 2016;69(3):384-8. <https://doi.org/10.1016/j.eururo.2015.01.014>.
17. Peng J, Sun B-F, Chen C-Y, Zhou J-Y, Chen Y-S, Chen H, et al. Single-cell RNA-seq highlights intra-tumoral heterogeneity and malignant progression in pancreatic ductal adenocarcinoma. *Cell Res*. 2019;29(9):725-38. <https://doi.org/10.1038/s41422-019-0195-y>.
18. Zhang J, Fujimoto J, Zhang J, Wedge DC, Song X, Zhang J, et al. Intratumor heterogeneity in localized lung adenocarcinomas delineated by multiregion sequencing. *Science*. 2014;346(6206):256-9. <https://doi.org/10.1126/science.1256930>.
19. Bashashati A, Ha G, Tone A, Ding J, Prentice LM, Roth A, et al. Distinct evolutionary trajectories of primary high-grade serous ovarian cancers revealed through spatial mutational profiling. *J Pathol*. 2013;231(1):21-34. <https://doi.org/10.1002/path.4230>.
20. Thomsen MB, Nordentoft I, Lamy P, Høyer S, Vang S, Hedegaard J, et al. Spatial and temporal clonal evolution during development of metastatic urothelial carcinoma. *Molecular oncology*. 2016;10(9):1450-60. <https://doi.org/10.1016/j.molonc.2016.08.003>.
21. Lamy P, Nordentoft I, Birkenkamp-Demtröder K, Thomsen MB, Villesen P, Vang S, et al. Paired Exome Analysis Reveals Clonal Evolution and Potential Therapeutic Targets in Urothelial Carcinoma. *Cancer Res*. 2016;76(19):5894-906. <https://doi.org/10.1158/0008-5472.Can-16-0436>.
22. Keller L, Pantel K. Unravelling tumour heterogeneity by single-cell profiling of circulating tumour cells. *Nat Rev Cancer*. 2019;19(10):553-67. <https://doi.org/10.1038/s41568-019-0180-2>.
23. Agarwal N, Pal SK, Hahn AW, Nussenzveig RH, Pond GR, Gupta SV, et al. Characterization of metastatic urothelial carcinoma via comprehensive genomic profiling of circulating tumor DNA. *Cancer*. 2018;124(10):2115-24. <https://doi.org/10.1002/cncr.31314>.
24. Wan JCM, Massie C, Garcia-Corbacho J, Mouliere F, Brenton JD, Caldas C, et al. Liquid biopsies come of age: towards implementation of circulating tumour DNA. *Nature reviews Cancer*. 2017;17(4):223-38. <https://doi.org/10.1038/nrc.2017.7>.
25. Alix-Panabières C, Pantel K. Clinical Applications of Circulating Tumor Cells and Circulating Tumor DNA as Liquid Biopsy. *Cancer Discov*. 2016;6(5):479-91. <https://doi.org/10.1158/2159-8290.Cd-15-1483>.
26. Zeng C, Stroup EK, Zhang Z, Chiu BCH, Zhang W. Towards precision medicine: advances in 5-hydroxymethylcytosine cancer biomarker discovery in liquid biopsy. *Cancer Commun (Lond)*. 2019;39(1):12. <https://doi.org/10.1186/s40880-019-0356-x>.
27. Abbosh C, Birkbak NJ, Wilson GA, Jamal-Hanjani M, Constantin T, Salari R, et al. Phylogenetic ctDNA analysis depicts early-stage lung cancer evolution. *Nature*. 2017;545(7655):446-51. <https://doi.org/10.1038/nature22364>.
28. Murtaza M, Dawson SJ, Pogrebniak K, Rueda OM, Provenzano E, Grant J, et al. Multifocal clonal evolution characterized using circulating tumour DNA in a case of metastatic breast cancer. *Nature communications*. 2015;6:8760. <https://doi.org/10.1038/ncomms9760>.
29. Parikh AR, Leshchiner I, Elagina L, Goyal L, Levovitz C, Siravegna G, et al. Liquid versus tissue biopsy for detecting acquired resistance and tumor heterogeneity in gastrointestinal cancers. *Nature medicine*. 2019;25(9):1415-21. <https://doi.org/10.1038/s41591-019-0561-9>.
30. Shen L. Liquid biopsy: a powerful tool to monitor trastuzumab resistance in HER2-positive metastatic gastric cancer. *Cancer Commun (Lond)*. 2018;38(1):72. <https://doi.org/10.1186/s40880-018-0344-6>.
31. Kyrochristos ID, Roukos DH. Comprehensive intra-individual genomic and transcriptional heterogeneity: Evidence-based Colorectal Cancer Precision Medicine. *Cancer treatment reviews*. 2019;80:101894. <https://doi.org/10.1016/j.ctrv.2019.101894>.
32. Cheng T, Roth B, Choi W, Black PC, Dinney C, McConkey DJ. Fibroblast growth factor receptors-1 and -3 play distinct roles in the regulation of bladder cancer growth and metastasis: implications for therapeutic targeting. *PLoS ONE*. 2013;8(2):e57284. <https://doi.org/10.1371/journal.pone.0057284>.
33. Huang Y, Gao S, Wu S, Song P, Sun X, Hu X, et al. Multilayered molecular profiling supported the monoclonal origin of metastatic renal cell carcinoma. *International journal of cancer*. 2014;135(1):78-87. <https://doi.org/10.1002/ijc.28654>.
34. Nel I, Gauler TC, Bublitz K, Lazaridis L, Goergens A, Giebel B, et al. Circulating Tumor Cell Composition in Renal Cell Carcinoma. *PLoS ONE*. 2016;11(4):e0153018. <https://doi.org/10.1371/journal.pone.0153018>.
35. Woo HK, Park J, Ku JY, Lee CH, Sunkara V, Ha HK, et al. Urine-based liquid biopsy: non-invasive and sensitive AR-V7 detection in urinary EVs from patients with prostate cancer. *Lab on a chip*. 2018;19(1):87-97. <https://doi.org/10.1039/c8lc01185k>.
36. de Bruin EC, McGranahan N, Mitter R, Salm M, Wedge DC, Yates L, et al. Spatial and temporal diversity in genomic instability processes defines lung cancer evolution. *Science*. 2014;346(6206):251-6. <https://doi.org/10.1126/science.1253462>.
37. Appierto V, Di Cosimo S, Reduzzi C, Pala V, Cappelletti V, Daidone MG. How to study and overcome tumor heterogeneity with circulating biomarkers: The breast cancer case. *Semin Cancer Biol*. 2017;44:106-16. <https://doi.org/10.1016/j.semcancer.2017.04.007>.
38. Gerlinger M, Rowan AJ, Horswell S, Math M, Larkin J, Endesfelder D, et al. Intratumor heterogeneity and branched

- evolution revealed by multiregion sequencing. *N Engl J Med*. 2012;366(10):883-92. <https://doi.org/10.1056/NEJMoa1113205>.
39. Sankin A, Hakimi AA, Mikkilineni N, Ostrovnaya I, Silk MT, Liang Y, et al. The impact of genetic heterogeneity on biomarker development in kidney cancer assessed by multi-regional sampling. *Cancer Med*. 2014;3(6):1485-92. <https://doi.org/10.1002/cam4.293>.
 40. Gerlinger M, Horswell S, Larkin J, Rowan AJ, Salm MP, Varela I, et al. Genomic architecture and evolution of clear cell renal cell carcinomas defined by multiregion sequencing. *Nat Genet*. 2014;46(3):225-33. <https://doi.org/10.1038/ng.2891>.
 41. Bettoni F, Masotti C, Habr-Gama A, Correa BR, Gama-Rodrigues J, Vianna MR, et al. Intratumoral Genetic Heterogeneity in Rectal Cancer: Are Single Biopsies representative of the entirety of the tumor? *Ann Surg*. 2017;265(1):e4-e6. <https://doi.org/10.1097/SLA.0000000000001937>.
 42. Lu Y-W, Zhang H-F, Liang R, Xie Z-R, Luo H-Y, Zeng Y-J, et al. Colorectal Cancer Genetic Heterogeneity Delineated by Multi-Region Sequencing. *PLoS ONE*. 2016;11(3):e0152673. <https://doi.org/10.1371/journal.pone.0152673>.
 43. Boutros PC, Fraser M, Harding NJ, de Borja R, Trudel D, Lalonde E, et al. Spatial genomic heterogeneity within localized, multifocal prostate cancer. *Nat Genet*. 2015;47(7):736-45. <https://doi.org/10.1038/ng.3315>.
 44. Heide T, Maurer A, Eipel M, Knoll K, Geelvink M, Veeck J, et al. Multiregion human bladder cancer sequencing reveals tumour evolution, bladder cancer phenotypes and implications for targeted therapy. *J Pathol*. 2019;248(2):230-42. <https://doi.org/10.1002/path.5250>.
 45. Lawrence MS, Stojanov P, Polak P, Kryukov GV, Cibulskis K, Sivachenko A, et al. Mutational heterogeneity in cancer and the search for new cancer-associated genes. *Nature*. 2013;499(7457):214-8. <https://doi.org/10.1038/nature12213>.
 46. Robertson AG, Kim J, Al-Ahmadie H, Bellmunt J, Guo G, Cherniack AD, et al. Comprehensive Molecular Characterization of Muscle-Invasive Bladder Cancer. *Cell*. 2018;174(4):1033. <https://doi.org/10.1016/j.cell.2018.07.036>.
 47. Yachida S, Jones S, Bozic I, Antal T, Leary R, Fu B, et al. Distant metastasis occurs late during the genetic evolution of pancreatic cancer. *Nature*. 2010;467(7319):1114-7. <https://doi.org/10.1038/nature09515>.
 48. Ding L, Ellis MJ, Li S, Larson DE, Chen K, Wallis JW, et al. Genome remodelling in a basal-like breast cancer metastasis and xenograft. *Nature*. 2010;464(7291). <https://doi.org/10.1038/nature08989>.
 49. Popławski AB, Jankowski M, Erickson SW, Díaz de Ståhl T, Partridge EC, Crasto C, et al. Frequent genetic differences between matched primary and metastatic breast cancer provide an approach to identification of biomarkers for disease progression. *Eur J Hum Genet*. 2010;18(5):560-8. <https://doi.org/10.1038/ejhg.2009.230>.
 50. Sauter G, Moch H, Gasser TC, Mihatsch MJ, Waldman FM. Heterogeneity of chromosome 17 and erbB-2 gene copy number in primary and metastatic bladder cancer. *Cytometry*. 1995;21(1):40-6. <https://doi.org/10.1002/cyto.990210109>.
 51. Tan TZ, Rouanne M, Tan KT, Huang RY, Thierry JP. Molecular Subtypes of Urothelial Bladder Cancer: Results from a Meta-cohort Analysis of 2411 Tumors. *Eur Urol*. 2019;75(3):423-32. <https://doi.org/10.1016/j.eururo.2018.08.027>.
 52. Warrick JI, Sjö Dahl G, Kaag M, Raman JD, Merrill S, Shuman L, et al. Intratumoral Heterogeneity of Bladder Cancer by Molecular Subtypes and Histologic Variants. *European urology*. 2019;75(1):18-22. <https://doi.org/10.1016/j.eururo.2018.09.003>.
 53. Hurst CD, Alder O, Platt FM, Droop A, Stead LF, Burns JE, et al. Genomic Subtypes of Non-invasive Bladder Cancer with Distinct Metabolic Profile and Female Gender Bias in KDM6A Mutation Frequency. *Cancer cell*. 2017;32(5):701-15.e7. <https://doi.org/10.1016/j.ccell.2017.08.005>.
 54. Sjö Dahl G, Eriksson P, Liedberg F, Höglund M. Molecular classification of urothelial carcinoma: global mRNA classification versus tumour-cell phenotype classification. *J Pathol*. 2017;242(1):113-25. <https://doi.org/10.1002/path.4886>.
 55. Seiler R, Ashab HAD, Erho N, van Rhijn BWG, Winters B, Douglas J, et al. Impact of Molecular Subtypes in Muscle-invasive Bladder Cancer on Predicting Response and Survival after Neoadjuvant Chemotherapy. *Eur Urol*. 2017;72(4):544-54. <https://doi.org/10.1016/j.eururo.2017.03.030>.
 56. Efsthathiou JA, Mouw KW, Gibb EA, Liu Y, Wu CL, Drumm MR, et al. Impact of Immune and Stromal Infiltration on Outcomes Following Bladder-Sparing Trimodality Therapy for Muscle-Invasive Bladder Cancer. *Eur Urol*. 2019;76(1):59-68. <https://doi.org/10.1016/j.eururo.2019.01.011>.
 57. Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SA, Behjati S, Biankin AV, et al. Signatures of mutational processes in human cancer. *Nature*. 2013;500(7463):415-21. <https://doi.org/10.1038/nature12477>.
 58. Thomsen MBH, Nordentoft I, Lamy P, Vang S, Reinert L, Mapendano CK, et al. Comprehensive multiregional analysis of molecular heterogeneity in bladder cancer. *Sci Rep*. 2017;7(1):11702. <https://doi.org/10.1038/s41598-017-11291-0>.
 59. Kamoun A, de Reyniès A, Allory Y, Sjö Dahl G, Robertson AG, Seiler R, et al. A Consensus Molecular Classification of Muscle-invasive Bladder Cancer. *Eur Urol*. 2019. <https://doi.org/10.1016/j.eururo.2019.09.006>.
 60. Lotan Y, Roehrborn CG. Sensitivity and specificity of commonly available bladder tumor markers versus cytology: results of a comprehensive literature review and meta-analyses. *Urology*. 2003;61(1).
 61. Konety BR. Molecular markers in bladder cancer: a critical appraisal. *Urol Oncol*. 2006;24(4):326-37.
 62. Chou R, Gore JL, Buckley D, Fu R, Gustafson K, Griffin JC, et al. Urinary Biomarkers for Diagnosis of Bladder Cancer: A Systematic Review and Meta-analysis. *Ann Intern Med*. 2015;163(12):922-31. <https://doi.org/10.7326/M15-0997>.
 63. Grivas P, Lalani AA, Pond GR, Nagy RJ, Faltas B, Agarwal N, et al. Circulating Tumor DNA Alterations in Advanced Urothelial Carcinoma and Association with Clinical Outcomes: A Pilot Study. *European urology oncology*. 2019. <https://doi.org/10.1016/j.euo.2019.02.004>.
 64. Sundahl N, Vandekerckhove G, Decaestecker K, Meireson A, De Visschere P, Fonteyne V, et al. Randomized Phase 1 Trial of Pembrolizumab with Sequential Versus Concomitant Stereotactic Body Radiotherapy in Metastatic Urothelial Carcinoma. *Eur Urol*. 2019;75(5):707-11. <https://doi.org/10.1016/j.eururo.2019.01.009>.
 65. Birkenkamp-Demtroder K, Christensen E, Nordentoft I, Knudsen M, Taber A, Hoyer S, et al. Monitoring Treatment Response

- and Metastatic Relapse in Advanced Bladder Cancer by Liquid Biopsy Analysis. *Eur Urol.* 2018;73(4):535-40. <https://doi.org/10.1016/j.eururo.2017.09.011>.
66. Ellinger J, El Kassem N, Heukamp LC, Matthews S, Cubukluoz F, Kahl P, et al. Hypermethylation of cell-free serum DNA indicates worse outcome in patients with bladder cancer. *J Urol.* 2008;179(1):346-52.
 67. Birkenkamp-Demtroder K, Nordentoft I, Christensen E, Hoyer S, Reinert T, Vang S, et al. Genomic Alterations in Liquid Biopsies from Patients with Bladder Cancer. *Eur Urol.* 2016;70(1):75-82. <https://doi.org/10.1016/j.eururo.2016.01.007>.
 68. Ellinger J, Bastian PJ, Ellinger N, Kahl P, Perabo FG, Büttner R, et al. Apoptotic DNA fragments in serum of patients with muscle invasive bladder cancer: a prognostic entity. *Cancer Lett.* 2008;264(2):274-80. <https://doi.org/10.1016/j.canlet.2008.01.038>.
 69. Patel KM, van der Vos KE, Smith CG, Mouliere F, Tsui D, Morris J, et al. Association Of Plasma And Urinary Mutant DNA With Clinical Outcomes In Muscle Invasive Bladder Cancer. *Sci Rep.* 2017;7(1):5554. <https://doi.org/10.1038/s41598-017-05623-3>.
 70. Maruyama R, Toyooka S, Toyooka KO, Harada K, Virmani AK, Zöchbauer-Müller S, et al. Aberrant promoter methylation profile of bladder cancer and its relationship to clinicopathological features. *Cancer Res.* 2001;61(24):8659-63.
 71. Gutiérrez MI, Siraj AK, Khaled H, Koon N, El-Rifai We, Bhatia K. CpG island methylation in Schistosoma- and non-Schistosoma-associated bladder cancer. *Mod Pathol.* 2004;17(10):1268-74.
 72. Friedrich MG, Chandrasoma S, Siegmund KD, Weisenberger DJ, Cheng JC, Toma MI, et al. Prognostic relevance of methylation markers in patients with non-muscle invasive bladder carcinoma. *Eur J Cancer.* 2005;41(17):2769-78.
 73. Takahashi T, Suzuki M, Shigematsu H, Shivapurkar N, Echebiri C, Nomura M, et al. Aberrant methylation of Reprimo in human malignancies. *Int J Cancer.* 2005;115(4):503-10.
 74. Raja R, Kuziora M, Brohawn PZ, Higgs BW, Gupta A, Dennis PA, et al. Early Reduction in ctDNA Predicts Survival in Patients with Lung and Bladder Cancer Treated with Durvalumab. *Clin Cancer Res.* 2018;24(24):6212-22. <https://doi.org/10.1158/1078-0432.ccr-18-0386>.
 75. Ward DG, Gordon NS, Boucher RH, Pirrie SJ, Baxter L, Ott S, et al. Targeted deep sequencing of urothelial bladder cancers and associated urinary DNA: a 23-gene panel with utility for non-invasive diagnosis and risk stratification. *BJU international.* 2019;124(3):532-44. <https://doi.org/10.1111/bju.14808>.
 76. Xu Y, Kim YH, Jeong P, Piao XM, Byun YJ, Kang HW, et al. Diagnostic value of combined IQGAP3/BMP4 and IQGAP3/FAM107A expression ratios in urinary cell-free DNA for discriminating bladder cancer from hematuria. *Urol Oncol.* 2019;37(1):86-96. <https://doi.org/10.1016/j.urolonc.2018.10.023>.
 77. Togneri FS, Ward DG, Foster JM, Devall AJ, Wojtowicz P, Alyas S, et al. Genomic complexity of urothelial bladder cancer revealed in urinary cfDNA. *Eur J Hum Genet.* 2016;24(8):1167-74. <https://doi.org/10.1038/ejhg.2015.281>.
 78. Stasik S, Salomo K, Heberling U, Froehner M, Sommer U, Baretton GB, et al. Evaluation of TERT promoter mutations in urinary cell-free DNA and sediment DNA for detection of bladder cancer. *Clin Biochem.* 2019;64:60-3. <https://doi.org/10.1016/j.clinbiochem.2018.11.009>.
 79. Springer SU, Chen C-H, Rodriguez Pena MDC, Li L, Douville C, Wang Y, et al. Non-invasive detection of urothelial cancer through the analysis of driver gene mutations and aneuploidy. *Elife.* 2018;7. <https://doi.org/10.7554/eLife.32143>.
 80. Ward DG, Baxter L, Gordon NS, Ott S, Savage RS, Beggs AD, et al. Multiplex PCR and Next Generation Sequencing for the Non-Invasive Detection of Bladder Cancer. *PLoS ONE.* 2016;11(2):e0149756. <https://doi.org/10.1371/journal.pone.0149756>.
 81. Hirotsu Y, Yokoyama H, Amemiya K, Hagimoto T, Daimon H, Hosaka K, et al. Genomic profile of urine has high diagnostic sensitivity compared to cytology in non-invasive urothelial bladder cancer. *Cancer science.* 2019;110(10):3235-43. <https://doi.org/10.1111/cas.14155>.
 82. Hayashi Y, Fujita K, Matsuzaki K, Matsushita M, Kawamura N, Koh Y, et al. Diagnostic potential of TERT promoter and FGFR3 mutations in urinary cell-free DNA in upper tract urothelial carcinoma. *Cancer science.* 2019;110(5):1771-9. <https://doi.org/10.1111/cas.14000>.
 83. Christensen E, Birkenkamp-Demtroder K, Nordentoft I, Hoyer S, van der Keur K, van Kessel K, et al. Liquid Biopsy Analysis of FGFR3 and PIK3CA Hotspot Mutations for Disease Surveillance in Bladder Cancer. *Eur Urol.* 2017;71(6):961-9. <https://doi.org/10.1016/j.eururo.2016.12.016>.
 84. Dudley JC, Schroers-Martin J, Lazzareschi DV, Shi WY, Chen SB, Esfahani MS, et al. Detection and Surveillance of Bladder Cancer Using Urine Tumor DNA. *Cancer Discov.* 2019;9(4):500-9. <https://doi.org/10.1158/2159-8290.CD-18-0825>.
 85. Paterlini-Brechot P, Benali NL. Circulating tumor cells (CTC) detection: clinical impact and future directions. *Cancer Lett.* 2007;253(2):180-204. <https://doi.org/10.1016/j.canlet.2006.12.014>.
 86. Qi F, Liu Y, Zhao R, Zou X, Zhang L, Li J, et al. Quantitation of rare circulating tumor cells by folate receptor α ligand-targeted PCR in bladder transitional cell carcinoma and its potential diagnostic significance. *Tumour Biol.* 2014;35(7):7217-23. <https://doi.org/10.1007/s13277-014-1894-0>.
 87. Chalfin HJ, Glavaris SA, Gorin MA, Kates MR, Fong MH, Dong L, et al. Circulating Tumor Cell and Circulating Tumor DNA Assays Reveal Complementary Information for Patients with Metastatic Urothelial Cancer. *European urology oncology.* 2019. <https://doi.org/10.1016/j.euo.2019.08.004>.
 88. Naoe M, Ogawa Y, Morita J, Omori K, Takeshita K, Shichijyo T, et al. Detection of circulating urothelial cancer cells in the blood using the CellSearch System. *Cancer.* 2007;109(7):1439-45. <https://doi.org/10.1002/cncr.22543>.
 89. Zaravinos A, Radojicic J, Lambrou GI, Volanis D, Delakas D, Stathopoulos EN, et al. Expression of miRNAs involved in angiogenesis, tumor cell proliferation, tumor suppressor inhibition, epithelial-mesenchymal transition and activation of metastasis in bladder cancer. *J Urol.* 2012;188(2):615-23. <https://doi.org/10.1016/j.juro.2012.03.122>.
 90. Yang Y, Qu A, Liu J, Wang R, Liu Y, Li G, et al. Serum miR-210 Contributes to Tumor Detection, Stage Prediction and Dynamic Surveillance in Patients with Bladder Cancer. *PLoS ONE.* 2015;10(8):e0135168. <https://doi.org/10.1371/journal.pone.0135168>.
 91. Hiltbrunner S, Mints M, Eldh M, Rosenblatt R, Holmström B, Alamdari F, et al. Urinary Exosomes from Bladder

- Cancer Patients Show a Residual Cancer Phenotype despite Complete Pathological Downstaging. *Sci Rep.* 2020;10(1):5960. <https://doi.org/10.1038/s41598-020-62753-x>.
92. Yin X, Zheng X, Liu M, Wang D, Sun H, Qiu Y, et al. Exosomal miR-663b targets Ets2-repressor factor to promote proliferation and the epithelial-mesenchymal transition of bladder cancer cells. *Cell Biol Int.* 2020;44(4):958-65. <https://doi.org/10.1002/cbin.11292>.
 93. Huang C-S, Ho J-Y, Chiang J-H, Yu C-P, Yu D-S. Exosome-Derived LINC00960 and LINC02470 Promote the Epithelial-Mesenchymal Transition and Aggressiveness of Bladder Cancer Cells. *Cells.* 2020;9(6). <https://doi.org/10.3390/cells9061419>.
 94. Abbastabar M, Sarfi M, Golestani A, Karimi A, Pourmand G, Khalili E. Tumor-derived urinary exosomal long non-coding RNAs as diagnostic biomarkers for bladder cancer. *EXCLI J.* 2020;19:301-10. doi:10.17179/excli2019-1683.
 95. Chen C, Luo Y, He W, Zhao Y, Kong Y, Liu H, et al. Exosomal long noncoding RNA LNMAT2 promotes lymphatic metastasis in bladder cancer. *J Clin Invest.* 2020;130(1):404-21. <https://doi.org/10.1172/JCI130892>.
 96. Erozcenci LA, Böttger F, Bijnsdorp IV, Jimenez CR. Urinary exosomal proteins as (pan-)cancer biomarkers: insights from the proteome. *FEBS Lett.* 2019;593(13):1580-97. <https://doi.org/10.1002/1873-3468.13487>.
 97. Zhang S, Du L, Wang L, Jiang X, Zhan Y, Li J, et al. Evaluation of serum exosomal lncRNA-based biomarker panel for diagnosis and recurrence prediction of bladder cancer. *J Cell Mol Med.* 2019;23(2):1396-405. <https://doi.org/10.1111/jcmm.14042>.
 98. Zhan Y, Du L, Wang L, Jiang X, Zhang S, Li J, et al. Expression signatures of exosomal long non-coding RNAs in urine serve as novel non-invasive biomarkers for diagnosis and recurrence prediction of bladder cancer. *Mol Cancer.* 2018;17(1):142. <https://doi.org/10.1186/s12943-018-0893-y>.
 99. Yu X, Wang R, Han C, Wang Z, Jin X. A Panel of Urinary Long Non-coding RNAs Differentiate Bladder Cancer from Urocystitis. *J Cancer.* 2020;11(4):781-7. <https://doi.org/10.7150/jca.37006>.
 100. Armstrong DA, Green BB, Seigne JD, Schned AR, Marsit CJ. MicroRNA molecular profiling from matched tumor and biofluids in bladder cancer. *Mol Cancer.* 2015;14:194. <https://doi.org/10.1186/s12943-015-0466-2>.
 101. Long JD, Sullivan TB, Humphrey J, Logvinenko T, Summerhayes KA, Kozinn S, et al. A non-invasive miRNA based assay to detect bladder cancer in cell-free urine. *Am J Transl Res.* 2015;7(11):2500-9.
 102. Murakami T, Yamamoto CM, Akino T, Tanaka H, Fukuzawa N, Suzuki H, et al. Bladder cancer detection by urinary extracellular vesicle mRNA analysis. *Oncotarget.* 2018;9(67):32810-21. doi:10.18632/oncotarget.25998.
 103. Lin S-Y, Chang C-H, Wu H-C, Lin C-C, Chang K-P, Yang C-R, et al. Proteome Profiling of Urinary Exosomes Identifies Alpha 1-Antitrypsin and H2B1K as Diagnostic and Prognostic Biomarkers for Urothelial Carcinoma. *Sci Rep.* 2016;6:34446. <https://doi.org/10.1038/srep34446>.
 104. Chen C-L, Lai Y-F, Tang P, Chien K-Y, Yu J-S, Tsai C-H, et al. Comparative and targeted proteomic analyses of urinary microparticles from bladder cancer and hernia patients. *J Proteome Res.* 2012;11(12):5611-29. <https://doi.org/10.1021/pr3008732>.
 105. Metzenmacher M, Váraljai R, Hegedüs B, Cima I, Forster J, Schramm A, et al. Plasma Next Generation Sequencing and Droplet Digital-qPCR-Based Quantification of Circulating Cell-Free RNA for Noninvasive Early Detection of Cancer. *Cancers (Basel).* 2020;12(2). <https://doi.org/10.3390/cancers12020353>.
 106. Russo IJ, Ju Y, Gordon NS, Zeegers MP, Cheng KK, James ND, et al. Toward Personalised Liquid Biopsies for Urothelial Carcinoma: Characterisation of ddPCR and Urinary cfDNA for the Detection of the TERT 228 G>A/T Mutation. *Bladder cancer (Amsterdam, Netherlands).* 2018;4(1):41-8. <https://doi.org/10.3233/blc-170152>.
 107. Newman AM, Bratman SV, To J, Wynne JF, Eclov NC, Modlin LA, et al. An ultrasensitive method for quantitating circulating tumor DNA with broad patient coverage. *Nature medicine.* 2014;20(5):548-54. <https://doi.org/10.1038/nm.3519>.
 108. Newman AM, Lovejoy AF, Klass DM, Kurtz DM, Chabon JJ, Scherer F, et al. Integrated digital error suppression for improved detection of circulating tumor DNA. *Nature biotechnology.* 2016;34(5):547-55. <https://doi.org/10.1038/nbt.3520>.
 109. Wang Y, Springer S, Mulvey CL, Silliman N, Schaefer J, Sausen M, et al. Detection of somatic mutations and HPV in the saliva and plasma of patients with head and neck squamous cell carcinomas. *Sci Transl Med.* 2015;7(293):293ra104. <https://doi.org/10.1126/scitranslmed.aaa8507>.
 110. Fernandez-Cuesta L, Perdomo S, Avogbe PH, Leblay N, Delhomme TM, Gaborieau V, et al. Identification of Circulating Tumor DNA for the Early Detection of Small-cell Lung Cancer. *EBioMedicine.* 2016;10:117-23. <https://doi.org/10.1016/j.ebiom.2016.06.032>.
 111. Ledergor G, Weiner A, Zada M, Wang S-Y, Cohen YC, Gatt ME, et al. Single cell dissection of plasma cell heterogeneity in symptomatic and asymptomatic myeloma. *Nature medicine.* 2018;24(12):1867-76. <https://doi.org/10.1038/s41591-018-0269-2>.
 112. Sundaresan TK, Sequist LV, Heymach JV, Riely GJ, Jänne PA, Koch WH, et al. Detection of T790M, the Acquired Resistance EGFR Mutation, by Tumor Biopsy versus Noninvasive Blood-Based Analyses. *Clinical cancer research : an official journal of the American Association for Cancer Research.* 2016;22(5):1103-10. <https://doi.org/10.1158/1078-0432.ccr-15-1031>.
 113. Khoo BL, Bouquerel C, Durai P, Anil S, Goh B, Wu B, et al. Detection of Clinical Mesenchymal Cancer Cells from Bladder Wash Urine for Real-Time Detection and Prognosis. *Cancers (Basel).* 2019;11(9). <https://doi.org/10.3390/cancers11091274>.
 114. Lin D, Wu Q, Qiu S, Chen G, Feng S, Chen R, et al. Label-free liquid biopsy based on blood circulating DNA detection using SERS-based nanotechnology for nasopharyngeal cancer screening. *Nanomedicine.* 2019;22:102100. <https://doi.org/10.1016/j.nano.2019.102100>.
 115. Park JS, Lee HJ, Almujaalhem A, Althubiany HH, A AA, Jang WS, et al. Prediction of High-Grade Clear Cell Renal Cell Carcinoma Based on Plasma mRNA Profiles in Patients with Localized Pathologic T1N0M0 Stage Disease. *Cancers (Basel).* 2020;12(5). <https://doi.org/10.3390/cancers12051182>.
 116. Rijnders M, Balcioglu HE, Robbrecht D, Boormans JL, Aarts MJB, Hamberg P, et al. Early response marker during pembrolizumab treatment in metastatic urothelial cancer: Temporal shift in peripheral CD4 T cells expressing chemokine receptors. 2020;38(15_suppl):5033-. https://doi.org/10.1200/JCO.2020.38.15_suppl.5033.

117. Choueiri TK, Hahn NM, Werner L, Regan MM, Rosenberg JE, Pal S, et al. The Borealis-2 clinical trial: A randomized phase 2 study of OGX-427 (apatorsen) plus docetaxel versus docetaxel alone in relapsed/refractory metastatic urothelial cancer. *Journal of Clinical Oncology*. 2014; 33(15_suppl):TPS4577-TPS. https://doi.org/10.1200/JCO.2017.35.6_suppl.289.
118. Bellmunt J, Eigel BJ, Senkus-Konefka E, Loriot Y, Twardowski P, Castellano DE et al. First-line randomized phase II study of gemcitabine/cisplatin plus apatorsen or placebo in patients with advanced bladder cancer: The International Borealis-1 trial. 2015;33(15_suppl):4503-. https://doi.org/10.1200/jco.2015.33.15_suppl.4503.
119. Abdelgawad A, Laymon M, Hashem A, Abdellatif MM, Magdy D, Eissa LA, et al. MP61-13 DO URINARY-BASED MOLECULAR BIOMARKER HAVE THE ABILITY TO PREDICT THE GRADE OF NON-MUSCLE INVASIVE BLADDER CANCER GRADE? 2020;203(Supplement 4):e940-e. <https://doi.org/10.1097/JU.0000000000000936.013>.
120. Du L, Duan W, Jiang X, Zhao L, Li J, Wang R, et al. Cell-free lncRNA expression signatures in urine serve as novel non-invasive biomarkers for diagnosis and recurrence prediction of bladder cancer. *J Cell Mol Med*. 2018;22(5):2838-45. <https://doi.org/10.1111/jcmm.13578>.

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