

Review

# Mineralocorticoid Receptor and Aldosterone-Related Biomarkers of End-Organ Damage in Cardiometabolic Disease

Stefania Gorini<sup>1</sup>, Vincenzo Marzolla<sup>1</sup>, Caterina Mammi<sup>1</sup>, Andrea Armani<sup>1</sup> and Massimiliano Caprio<sup>1,2,\*</sup>

- <sup>1</sup> Laboratory of Cardiovascular Endocrinology, IRCCS San Raffaele Pisana, Via di Val Cannuta 247, 00166 Rome, Italy; stefania.gorini@sanraffaele.it (S.G.); vincenzo.marzolla@sanraffaele.it (V.M.); caterina.mammi@sanraffaele.it (C.M.); andrea.armani@sanraffaele.it (A.A.)
- <sup>2</sup> Department of Human Sciences and Promotion of the Quality of Life, San Raffaele Roma Open University, 00166 Rome, Italy
- \* Correspondence: massimiliano.caprio@sanraffaele.it; Tel.: +39-06-5225-3419; Fax: +39-06-5225-5668

Received: 3 August 2018; Accepted: 12 September 2018; Published: 18 September 2018



Abstract: The mineralocorticoid receptor (MR) was first identified as a blood pressure regulator, modulating renal sodium handling in response to its principal ligand aldosterone. The mineralocorticoid receptor is also expressed in many tissues other than the kidney, such as adipose tissue, heart and vasculature. Recent studies have shown that MR plays a relevant role in the control of cardiovascular and metabolic function, as well as in adipogenesis. Dysregulation of aldosterone/MR signaling represents an important cause of disease as high plasma levels of aldosterone are associated with hypertension, obesity and increased cardiovascular risk. Aldosterone displays powerful vascular effects and acts as a potent pro-fibrotic agent in cardiovascular remodeling. Mineralocorticoid receptor activation regulates genes involved in vascular and cardiac fibrosis, calcification and inflammation. This review focuses on the role of novel potential biomarkers related to aldosterone/MR system that could help identify cardiovascular and metabolic detrimental conditions, as a result of altered MR activation. Specifically, we discuss: (1) how MR signaling regulates the number and function of different subpopulations of circulating and intra-tissue immune cells; (2) the role of aldosterone/MR system in mediating cardiometabolic diseases induced by obesity; and (3) the role of several MR downstream molecules as novel potential biomarkers of cardiometabolic diseases, end-organ damage and rehabilitation outcome.

Keywords: mineralocorticoid receptor; aldosterone; PBMC; NGAL; Gal-3; PTGDS; adipose tissue

# 1. Introduction

The mineralocorticoid receptor (MR) is a member of the nuclear receptor family and acts as a ligand-dependent transcription factor. It was initially identified to regulate blood pressure through its ability to modulate renal sodium handling in response to aldosterone [1–3]. Importantly, aldosterone is not the exclusive ligand of MR. Cortisol and aldosterone display similar affinity and specificity for the MR [4]. In tissues with low 11 $\beta$ -hydroxysteroid dehydrogenase type 2 (11 $\beta$ -HSD2) activity, which converts cortisol to inactive cortisone, MR activation is mainly regulated by circulating glucocorticoids [5].

It is now evident that the MR is expressed in many tissues other than the kidney. Importantly, MR is expressed in the heart [6,7], in all cell types of the vasculature, including smooth muscle cells (SMCs), endothelial cells (ECs) and fibroblasts, and has also been found in adipose tissue [8]. In this context, the MR has a relevant role in the control of cardiovascular and metabolic function [9–11].



Dysregulation of the aldosterone/MR signaling has been identified as an important cause of several diseases. Indeed, high plasma levels of aldosterone are strictly associated with hypertension, obesity and increased cardiovascular risk [12]. Several studies demonstrated that obese and hypertensive patients display increased plasma and urinary levels of aldosterone [13–15]. Molecular mechanisms underlying vascular changes in hypertension are not completely understood, but a role for aldosterone has been suggested. Accumulating evidence has demonstrated that aldosterone displays powerful vascular effects and acts as a potent pro-fibrotic agent in cardiovascular remodeling [16,17]. Indeed, MR activation in human coronary artery SMCs regulates several genes involved in vascular fibrosis, calcification and inflammation, such as collagen types I and III, the parathyroid hormone receptors and interleukin (IL)-16 [16]. The MR is known to regulate genes involved in inflammation and oxidative stress in human coronary ECs [10]. Reactive oxygen species (ROS) have also been suggested to mediate the detrimental effects of aldosterone in the vasculature through MR activation [18,19].

Ligand-independent transcriptional activation of the MR has also been described, since MR can be activated under conditions of high oxidative stress, even without any increase in circulating agonists [20]. It is now clear that several molecules, other than aldosterone, can activate MR. For instance, Rac1 represents an important activator of MR. It is a small GTPase belonging to the Rho family and it is involved in the activation of MR in the kidney and in the heart [21–23]. Rac1 overexpression in cardiomyocytes of rats upregulates MR transcription [24]. Its overexpression in a mouse model of pressure overload-induced heart failure (HF) can increase MR protein and MR target genes expression in the heart [23]. Clinical evidence suggests that the interaction between Rac1 and MR plays a major role in cardiovascular damage induced by high sodium intake in humans as Rac1 expression positively correlates with MR expression under high sodium intake dietary regimens [25].

In addition to its classical genomic effects, aldosterone elicits rapid actions that do not require transcription or translation. These effects can be mediated by crosstalk of the MR with several membrane-associated signaling pathways, including transactivation of tyrosine kinases (i.e., epidermal growth factor receptor (EGFR), platelet-derived growth factor receptor (PDGFR) and insulin-like-growth factor 1 receptor (IGF1R) [26], or G protein coupled receptors. Among these, G protein-coupled estrogen receptor 1 (GPER1) has been proposed as a novel aldosterone receptor, even if a direct binding or interaction between GPER1 and aldosterone still awaits demonstration [27]. Such immediate effects are mostly involved in ion transport, but play also a relevant role in extrarenal tissues, contributing to the pathophysiological effects of MR and leading to inflammation, fibrosis and organ damage. Notably, genomic and nongenomic MR signaling interact closely, and their combined effect determines the long-term impact of altered MR activation at the level of vessels, heart and kidney [28–30]. This aspect was extensively reviewed by Ruhs et al. [26].

This review discusses the global pathophysiological relevance of aldosterone and MR-related pathways in cardiometabolic disease and obesity. In this context, we discuss the role of potential novel biomarkers related to the aldosterone/MR system, that could help identify early stages of end-organ damage (heart, vessels, kidney, and adipose tissue) in cardiometabolic diseases, as well as the outcome of therapeutic intervention and rehabilitation.

#### 2. Cardio-Metabolic Effects of Altered Mineralocorticoid Receptor Activation

There is a large body of evidence that identifies the MR pathway as a valuable check-point for healthy or pathological cardiometabolic states. Indeed, an important role of MR activation in the pathogenesis of cardiometabolic diseases has been clarified [31]. Aldosterone, classically considered only as regulator of Na<sup>+</sup> reabsorption, is known to trigger cardiovascular and renal tissue damage through different pathways, which are, at least in part, independent of its renal-mediated effects on blood pressure [10]. Indeed, extrarenal effects of aldosterone are relevant for production of extracellular matrix (ECM) components, and elicit several specific tissue responses, such as hypertrophy, remodeling and fibrosis, which are pathogenic and contribute to end-organ damage [32].

Notably, preclinical studies have shown that aldosterone causes end-organ tissue damage only in the context of an inappropriate salt status [33]. Indeed, pioneering studies clarified that aldosterone causes myocardial fibrosis only in rats maintained on a high-salt diet [34].

These pieces of evidence have also been observed in humans. In environmental conditions where the average daily intake of sodium is very low, physiologically circulating aldosterone levels can be very high, in response to sodium deficiency, but do not determine any deleterious cardiovascular or renal effects. It is therefore important to remark that, when aldosterone levels are out of the physiological feedback control loop, and become inappropriate for salt status, they can induce cardiovascular damage [12].

Primary aldosteronism (PA) is due to an autonomous overproduction of aldosterone by the adrenal gland, entirely unrelated to salt status [35]. This leads to a condition characterized by severe hypertension, low renin levels and severely increased cardiovascular risk, with a higher incidence of stroke, atrial fibrillation, and myocardial infarction [36]. Interestingly, chronic exposure to aldosterone in primary aldosteronism has been also associated to altered glucose homeostasis, and, in general, with a greater prevalence of the metabolic syndrome [37,38]. Fallo et al. described a higher prevalence of metabolic syndrome in patients with PA compared to those with essential hypertension [39]. In particular, altered glucose metabolism represented the best-established component of metabolic syndrome among PA patients. A possible reason for this observation relies on the inhibitory effects of aldosterone upon glucose-stimulated insulin secretion, as suggested by in vitro studies on isolated pancreatic islets [40,41]. In line with preclinical data, several clinical studies show that insulin sensitivity is reduced in PA patients compared to hypertensive controls [42–44].

Importantly, Catena et al. showed that PA is associated with insulin resistance and that pharmacological MR antagonism can reverse the insulin resistance status of primary aldosteronism patients [45].

Evaluation of longitudinal changes in metabolic risk factors in the Framingham offspring study showed that aldosterone is correlated to the development of metabolic syndrome. This correlation was also apparent in longitudinal changes in blood pressure and plasma levels of high-density lipoprotein (HDL) cholesterol, suggesting aldosterone and its associated pathways as potential biomarkers for metabolic risks [46].

Consequently, increased cardiovascular morbidity and mortality is related to several blood pressure-independent factors in patients affected by PA, such as cardiac myocardial fibrosis [47,48], cardiac remodeling [49], atherosclerosis with plaque rupture [50] and arrhythmias [51]. Moreover, several clinical studies reveal that circulating aldosterone levels are reliable predictors of cardiovascular ischemia [36,52]. Finally, strong evidence emerged from clinical trials in patients with heart failure and previous myocardial infarction that demonstrated MR pharmacological blockade protects from mortality and end-organ damage [53].

## 3. Contribution of Different Immune Cells Subsets to Aldosterone-Induced Inflammation

The state of hypertension induced by excessive secretion of aldosterone in PA patients is mostly due to the promotion of myocardial and vascular fibrosis [54,55]. However, a relevant role of oxidative stress [56], perivascular inflammation, and infiltration of T lymphocytes and antigen-presenting cells (APCs) in vessels has been also described [57–59]. Indeed, it is known that excessive production of aldosterone leads to hypertension by a pro-inflammatory state promoted by T cell immunity [57]. Pioneering studies by Selye et al. in 1949 showed that desoxycorticosterone (DOC), the first mineralocorticoid to be discovered, could induce a pro-inflammatory effect [60]. This finding was based on the observation that DOC was able to worsen clinical symptoms of rheumatoid arthritis, as well as induce a strong pro-inflammatory effect in animal models. In vitro and in vivo studies demonstrated that MR activation by aldosterone exerts its effects on vasculature and the heart, in part, by inducing an increase in oxidative stress [61].

In addition, aldosterone is able to induce vascular and cardiac inflammation through increased expression of inflammatory biomarkers, such as fibrinogen and plasminogen activator inhibitor-1 (PAI-1) [62,63]. Mineralocorticoid receptor activation in ECs is known to contribute to the induction of cardiac inflammation and remodeling by promoting the expression of vascular cell adhesion molecule 1 (VCAM1), as shown in animal models of hypertension [64]. Moreover, endothelial MR activation by aldosterone leads to the overexpression of the intracellular adhesion molecule-1 (ICAM-1) via an MR-responsive element in ICAM-1 promoter region [10,65,66], thereby enabling leukocyte adhesion to coronary artery ECs.

Activation of the MR by deoxycorticosterone acetate in the presence of high salt intake in mice represents a powerful model of hypertension and inflammation (DOCA-Salt model), which in turn leads to cardiovascular and renal fibrosis and cardiac remodeling [62,67]. Rickard et al. investigated the specific role of MR activation in ECs and studied vascular responses to aldosterone in EC-null MR (EC-MRKO) mice treated with DOCA-salt [68]. In the early stages (after eight days of treatment), macrophage infiltration and expression of myocardial proinflammatory genes (i.e., C-C chemokine receptor type 5 (CCR5) and inducible nitric oxide synthase (iNOS)) in EC-MRKO was prevented; mRNA levels of profibrotic genes in EC-MRKO mice (i.e., connective tissue growth factor (CTGF) and PAI-1) were significantly lower compared to wild type (WT) mice. Finally, CTGF expression and collagen deposition were significantly reduced in EC-MRKO mice. Reduced cardiac tissue macrophage infiltration determined the down-regulation of proinflammatory and profibrotic markers in the heart, along with a lower vascular expression of ICAM-1 and CTGF [68].

Interestingly, aldosterone amplifies its pro-inflammatory effects through the induction of osteopontin release in activated tissue macrophages and T-cells [69]. Recent studies also reveal that aldosterone induces renal tubulointerstitial inflammation/fibrosis and podocytes injury through the activation of the nucleotide-binding oligomerization domain (NOD)-like receptor family pyrin domain containing 3 (NLRP3) inflammasome, determining the expression of important inflammasome components, such as caspase 1, IL-1 $\beta$  and IL-18 [70,71]. Moreover eplerenone (a selective MR antagonist) is able to suppress the expression of critical inflammasome components, such as *Nlrp3* and *Caspase1*, in epididymal white adipose tissue (eWAT) and liver of obese mice. These data clearly show that MR represents a crucial player in the induction of inflammasome-mediated chronic inflammation in metabolic disorders [72].

There is a large body of evidence showing that lymphocytes are important players in the development of chronic hypertension, perivascular mononuclear cell infiltration, and renal injury. In 1976, Svendsen observed that, upon DOCA-salt treatment, mice with normal thymus function and nude mice with genetical aplasia of the thymus, both displayed a significant increase in blood pressure after three weeks [73]. After 2–3 months, however, blood pressure increased and cell infiltration around intrarenal vessels was significantly more pronounced in WT than in nude mice, together with degenerative changes in the kidney, such as wedge-shaped infarcts. Thymus grafting in nude mice before DOCA-salt treatment recovered the ability of DOCA-salt treatment to induce chronic hypertension and intrarenal vascular disease, as previously seen in mice with normal thymus function [73]. More recent studies clarified how MR signaling is able to regulate the number of circulating T cells in human subjects and their homing to lymph nodes [74]. Under physiological conditions, T lymphocytes do not seem to contribute to systemic blood pressure, but DOCA-salt treatment, as well as angiotensin II (AngII) infusion, in hypertensive animal models, display an increase in intravascular and circulating T cells [57,58]. DOCA-salt or AngII infusion is also able to increase IL-17 secretion by T lymphocytes, as well as IL-17 protein in the heart and vessel wall [57,75]. Interestingly, recent studies on peripheral blood mononuclear cells (PBMCs) from hypertensive patients showed an increased prevalence of cytotoxic CD8+ T cells compared to normal subjects [76]. Accordingly, Amador et al. demonstrated the presence of CD8+ and IL-17+ T cells in PBMCs and splenocytes of hypertensive DOCA-salt-treated mice. Such effects were prevented by spironolactone, suggesting a role for the mineralocorticoid receptor. Moreover, spironolactone was able to decrease

IL-17 expression and increase the synthesis of typical regulatory T cells (Treg) marker forkhead box P3 (FoxP3), indicating that MR blockade downregulates T helper 17 (Th17) and upregulates Treg cell polarization [77]. Li et al. recently demonstrated that pharmacological MR antagonism protects against cardiac dysfunction and hypertrophy induced by abdominal aortic constriction. Mineralocorticoid receptor antagonism decreased the accumulation and activation of CD4+ and CD8+ T cells in the murine heart. Moreover, T cell specific MR-knockout mice displayed reduced cardiac hypertrophy, fibrosis, and dysfunction after abdominal aortic constriction [78]. Interestingly, dendritic cells (DCs) express MR mRNA and protein, therefore they are able to respond to aldosterone [79]. Dendritic cells have the peculiar capacity to prime naive T cells (CD4+ and CD8+) modulating an adaptive immune response [80]. Herrada et al. demonstrated that aldosterone enhances CD8+ T cytotoxic cells activation in a DCs-dependent fashion [79]. Indeed, direct in vitro activation of T lymphocytes by aldosterone was not able to induce the overexpression of typical activation markers, such as IL-2 and CD69. On the other hand, pretreatment of DCs with aldosterone, followed by co-culture with purified T cells, determined the activation of CD8+ T cells, as shown by IL-2 and interferon-gamma (IFN- $\gamma$ ) secretion and CD69 upregulation, and CD4+ T lymphocytes polarization toward Th17 [79]. More specifically, aldosterone induced DCs to secrete IL-6 and transforming growth factor-beta (TGF- $\beta$ ), which in turn activate CD8+T cells and promote CD4+T cells towards a Th17 phenotype [81]. Moreover, aldosterone downregulates the programmed death-ligand 1 (PD-L1) in DCs. The programmed death-ligand 1 is one of the ligands that suppress CD8+T cell activation, and this mechanism further amplifies CD8+T cell activation [82,83] (Figure 1).



**Figure 1.** Effects of excess mineralocorticoid receptor (MR) activation on circulating and intra-tissue immune cells. Overactivation of MR upregulates CD8+ T cells and T helper 17 (Th17) cells in circulating peripheral blood mononuclear cells (PBMCs) and in immune cells infiltrating in the heart. On the other hand, MR antagonism is able to decrease Th17 polarization and to induce the T regulatory cell (Treg) phenotype. These cells subsets are primed by dendritic cells (DCs). Dendritic cells express MR and are induced by aldosterone to produce polarizing cytokines that are able to activate CD8+ T cells and to prime CD4+ T cells towards the Th17 phenotype. IL: interleukin; TGF- $\beta$ : transforming growth factor-beta; PD-L1: programmed death-ligand 1; IFN- $\gamma$ : interferon-gamma.

A precise characterization of cardiovascular inflammation is extremely important to gain more insight into the pathophysiology of aldosterone-related end-organ damage. Indeed, plasma cytokine

levels may represent a less-sensitive index of the underlying disease when compared to detailed immunophenotyping in the context of heart and vascular tissue. Therefore, a thorough description of immune cell populations in plasma and tissues might represent a more valuable approach to characterize chronic inflammation states that are dependent on the alteration of the MR/aldosterone pathway.

#### 4. Aldosterone as a Novel Marker of Obesity

Excessive activation of MR in adipose tissue contributes to several metabolic alterations often observed in obesity and metabolic syndrome. Obesity is determined by an excess of adipose tissue, in order to store excess lipids and calories, which results in white adipose tissue (WAT) expansion through two possible mechanisms: increase in cell number (hyperplasia) and/or cell size (hypertrophy) [84]. In turn, dysfunctional adipocytes promote macrophage recruitment within WAT through the production of several chemokines (e.g., monocyte chemoattractant protein-1 (MCP-1) and IL-8) [85,86]. This then contributes to several obesity-related complications, in particular low-grade chronic inflammation, fat mass expansion and insulin resistance [87–89]. In obesity states, infiltrating macrophages undergo a polarization shift from an anti-inflammatory phenotype (M2) to a proinflammatory one (M1) [90]. Several studies have shown that MR activation triggers adipose tissue inflammation [91]. In particular, aldosterone determines an up-regulation of several proinflammatory adipokines (e.g., tumor necrosis factor alpha (TNFα), MCP-1, IL-6, and leptin) and reduces adiponectin expression. The mineralocorticoid receptor pharmacological blockade is able to reduce the total number of hypertrophic adipocytes in murine models of obesity, with a subsequent modification of adipocyte secretory capacity [92]. The mineralocorticoid receptor is also able to affect macrophages polarization. Indeed, aldosterone promotes a classic proinflammatory profile in human monocyte-derived macrophages (M1), whereas the MR antagonist eplerenone elicits a switch to the anti-inflammatory profile (M2) [93]. Moreover, aldosterone favors an increase in intracellular ROS levels in murine preadipocytes, whereas MR blockade reverses such increase and reduces ROS production in the adipose tissue of obese mice [92]. Finally, MR antagonism in mice fed a high-fat diet has been shown to improve glucose tolerance and to prevent white fat expansion and body weight gain [72,94]. Altogether, these data demonstrate that MR activity plays a relevant role in the pathogenesis of the chronic low-grade inflammatory state and adipocyte dysfunction observed in obesity [95].

Our research group has characterized the effects of aldosterone and MR on adipocytes. We first demonstrated that MR expression in murine preadipocytes gradually increases along differentiation, driving the acquisition of the mature adipocyte phenotype via increased expression of peroxisome proliferator-activated receptor-gamma (PPAR $\gamma$ ) [96], the "master gene" of adipogenesis in mammals [97]. Pharmacological MR antagonism determines a marked antiadipogenic effect in both murine and human preadipocytes by decreasing the expression of PPAR $\gamma$  [98]. Accordingly, MR mRNA and protein have been both detected in human visceral adipose tissue (VAT) [99] and specific MR knockdown in primary human visceral preadipocytes significantly reduced PPAR $\gamma$  expression and disrupted adipose differentiation process [100]. Importantly, adipocyte MR expression is higher in obese subjects, as well as in VAT when compared with subcutaneous adipose tissue [101]. These findings strongly indicate that adipocyte MR is more abundant in obese subjects, and its excessive activation contributes to adipocyte hypertrophy and dysfunction, which are frequently observed in obesity states [11].

Obesity and metabolic syndrome are strictly associated with an increased risk of cardiovascular disease, including left ventricular hypertrophy, coronary artery disease, hypertension, congestive heart failure, and vascular stiffness [102]. A large body of evidence indicates an important contribution of aldosterone/MR system into development of metabolic syndrome [103,104]. A higher prevalence of metabolic syndrome and increased cardiovascular events have been observed in patients affected by primary aldosteronism, when compared to essential hypertension [39,105].

metabolic syndrome, and directly correlated to waist circumference, blood pressure and plasma triglycerides [106]. Obese subjects are characterized by high aldosterone and normal or low cortisol plasma levels [107,108]. This evidence supports the hypothesis that hyperaldosteronism and obesity could be linked by a mechanistic relationship, and the aldosterone/MR system may represent a mediator for cardiometabolic disease induced by obesity [108–110]. It is well established that unknown molecules secreted by adipose tissue are able to directly stimulate aldosterone production by the adrenal glands, and this effect is independent of renin-angiotensin-aldosterone system (RAAS) activation [111,112]. Recently, leptin has been proposed as one of the adipose tissue-derived products able to induce aldosterone synthesis. Huby et al. showed that leptin increases aldosterone synthase expression and function [113]. Leptin overexpression in obesity is able to directly stimulate the adrenals, leading to an increased production of aldosterone, which in turn binds and activates MR at the adipocyte level. Such a vicious cycle leads to adipose expansion, chronic inflammation, oxidative stress and subsequent increase of aldosterone-releasing factors production by adipocytes [110] (Figure 2). In consideration of these findings and in line with elevated aldosterone plasma levels observed in obese subjects [106], aldosterone plasma levels can be considered as a novel biomarker of obesity, as its secretion is correlated with WAT expansion and inflammatory state.



**Figure 2.** Cross-talk between adipose tissue and adrenocortical cell. Leptin secreted by adipose tissue stimulates aldosterone secretion from adrenal cortex increasing aldosterone synthase expression and aldosterone production in adrenal cells. Aldosterone in turn binds and activates MR at adipocyte level, favoring adipocyte differentiation, hypertrophy and inflammation. This vicious cycle leads to adipose tissue expansion and inflammation, reactive oxygen species (ROS) production and increase in MR expression.

# 5. Mineralocorticoid Receptor Downstream Molecules: Novel Biomarkers of Cardiometabolic Diseases?

Cardiac remodeling secondary to hypertension is characterized by inflammation and fibrosis. It is considered as a major risk factor for cardiovascular morbidity and mortality, and represents a leading cause of chronic heart failure [114]. Cardiac remodeling begins with an inflammatory state, which promotes changes in ECM, resulting in myocardial fibrosis [115,116]. A dysregulated expression of metalloproteases (MMPs) determines an altered ECM remodeling during fibrosis. Activation of MMPs induces both the degradation of ECM structural components and the activation of growth factors able to promote inflammation [117]. Aldosterone has long been considered an important trigger for organ damage in hypertension [118]; in fact, its levels are increased in hypertensive patients and in

spontaneously hypertensive rats (SHR) [119,120]. Mineralocorticoid receptor activation leads to cardiac inflammation and fibrosis [121] and podocyte injury: it is now clear that the proinflammatory effects of aldosterone are mediated by NLPR3 inflammasome at the level of podocytes [71]. Most importantly, MR antagonism reduces mortality and morbidity in clinical trials [53,122,123].

Given the central role of MR in the development of cardiometabolic disease, we focused on three MR downstream molecules that recently emerged as specific mediators of MR activation.

#### 5.1. Neutrophil Gelatinase-Associated Lipocalin Protein

The neutrophil gelatinase-associated lipocalin protein (NGAL) has been identified as a novel MR target in the cardiovascular system [124]. The neutrophil gelatinase-associated lipocalin protein is a 25-kDa glycoprotein of the lipocalin superfamily [125] expressed by several cell types, including renal cells [126], ECs and SMCs [127,128], cardiomyocytes [124] and some immune cells subpopulations, such as neutrophils, macrophages, and DCs [128–131]. This protein is a marker of renal injury [132]. Elevated NGAL plasma levels have also been associated to increased mortality in patients with heart failure [133] independently from kidney dysfunction [134]. Accordingly, a recent study demonstrated that NGAL plays an important role in cardiovascular injury induced by aldosterone [135], and represents a mediator of cardiac inflammation and fibrosis in post myocardial infarction [136]. However, cell types involved in NGAL production in mineralocorticoid-induced organ damage have not yet been clearly determined. We previously discussed the role of immune cells MR expression in the progression of cardiometabolic diseases. Interestingly, elevated NGAL plasma levels have been detected in animal models in response to proinflammatory stimuli, as well as in patients affected by acute/chronic inflammatory states [137,138]. The secretion of NGAL by immune cells may play an important role in mediating mineralocorticoid-induced hypertension and cardiac injuries, since NGAL is a direct MR target [137,138]. In accordance, Buonafine et al. recently demonstrated that NGAL secretion by immune cells plays a pivotal role in mediating mineralocorticoid-induced cardiac injuries [139]. Mice lacking NGAL in their immune cells were protected against cardiac inflammation and fibrosis induced by nephrectomy-aldosterone (NAS) 200 µg/kg/day-salt 1% challenge [139]. In consideration of these data, NGAL could be an eligible biomarker in cardiovascular diseases due to altered mineralocorticoid activation, besides its well-known relevance as a biomarker of renal injury.

#### 5.2. Galectin-3

Hyperaldosteronism worsens fibrosis through the increase in the production of several proinflammatory molecules [140]. Galectin-3 (Gal-3) is a 29–35-kDa protein, member of the  $\beta$ -galactoside-binding lectin family, and it is expressed in several cell types such as fibroblasts [141], ECs [142], and inflammatory cells [143]. Recent evidence shows that Gal-3 mediates aldosterone-induced vascular remodeling and cardiac fibrosis [144,145]. Hypertensive aldosterone salt-treated rats showed increased Gal-3 expression at both mRNA and protein levels in the heart. Cotreatment with spironolactone or modified citrus pectin (MCP), a Gal-3 inhibitor [146], abolished cardiac Gal-3 mRNA and protein up-regulation. Interestingly, cardiac hypertrophy and dysfunction were prevented by spironolactone or MCP co-treatment [145].

In addition, pharmacological blockade of Gal-3 prevents the aldosterone-induced increase in inflammatory markers and in MMP activities, indicating Gal-3 as a possible novel mediator in cardiac inflammation. In human cardiac fibroblasts, Gal-3 inhibition was able to prevent the increase in inflammatory and fibrotic markers (MMP activities, and ECM components) induced by aldosterone [147]. These observations suggest a major role of Gal-3 in mediating aldosterone-induced cardiac remodeling due to myocardial inflammation and fibrosis, which in turn determines the development of HF. Accordingly, clinical studies show increased levels of Gal-3 in patients with HF [148]. Moreover, plasma levels of Gal-3 are correlated with serum ECM markers, and Gal-3 represents a prognostic factor in patients affected by coronary artery disease, given its role in plaque destabilization [149].

Cardiac fibrosis is also associated to obesity. High fat diet-fed animals show cardiac hypertrophy, fibrosis and an increase in superoxide anion and proinflammatory molecules production [150]. Ex vivo studies showed that aldosterone-activated MR promotes adipocyte differentiation and secretion of proinflammatory adipokines and leptin [96,151]. In obese subjects, MR expression is increased when compared with lean individuals, which has been shown in several preclinical models of obesity and metabolic syndrome [92,95,152]. In line with this, Gal-3 inhibition was recently found to prevent adipose tissue remodeling in obesity [153]. Ectopic fat in obese individuals shares some functional features with visceral adipose tissue, including leptin secretion. Interestingly, leptin secreted by epicardial fat can exert its action directly on the heart since epicardial fat leans closely against the myocardium [154]. Leptin is directly involved in cardiac fibrosis, exerting prooxidant and profibrotic effects, inducing cardiomyocytes hypertrophy [155–157], and affecting collagen turnover, as observed in high fat diet-fed mice. Galectin-3 is expressed in many tissues, including the heart, and its circulating levels significantly increase in obesity [158–160]. Given its ability to stimulate collagen deposition and exacerbate proinflammatory states, Gal-3 could be involved in leptin-induced cardiac collagen derangement [143,159]. To address this hypothesis, Martinez-Martinez et al. evaluated fibrosis and oxidative stress in cardiomyocytes from high fat diet-fed rats, as well as in vitro proliferation of cardiac fibromyoblasts extracted from rat heart exposed to elevated leptin levels. They showed that collagen synthesis induced by leptin is partly mediated by the production of Gal-3 [141]. Therefore, also taking into consideration that its plasma levels are increased in primary aldosteronism and obesity [141,159–161], Gal-3 emerges as a novel circulating biomarker of cardiac damage and cardiometabolic disfunction due to MR activation.

## 5.3. Lipocalin-Like Prostaglandin D2 Synthase

Experimental and clinical studies have clearly demonstrated that excess aldosterone is a risk factor for type-2 diabetes mellitus and metabolic syndrome [39]. Interestingly, pharmacological MR antagonism improves glucose tolerance and reduces insulin resistance in murine models [92, 94,95,152]. A mouse model selectively overexpressing MR in adipocytes (adipo-MR) displayed all the characteristics of metabolic syndrome [101]. Importantly, these mice showed an increase in lipocalin-like prostaglandin D2 synthase (PTGDS) mRNA expression in VAT.

The PTGDS is an enzyme involved in adipose tissue pathophysiology [162–166]. The increase in its expression is abolished in the presence of the MR antagonist spironolactone. In addition, the increase in PTGDS mRNA levels in VAT and subcutaneous adipose tissue (SAT) from genetically obese db/db mice are significantly correlated to increased MR mRNA levels from the same adipose depots. Moreover, upon aldosterone treatment, differentiated SW872 human adipocytes show increased expression of PTGDS mRNA levels, which is prevented by coincubation with spironolactone. Finally, in obese patients, VAT shows higher expression of PTGDS mRNA levels when compared to SAT, and again a positive correlation between PTGDS and MR mRNA levels is observed [101]. Altogether, these data suggest a direct control of MR in PTGDS transcription in adipocytes. Lipocalin-like prostaglandin D2 synthase emerges as a novel MR target in both mice and human adipocytes, and it might represent a novel tissutal marker of MR activation in adipocytes.

#### 6. Conclusions

Recent evidence shows that dysregulation of the aldosterone/MR system is strictly associated with several pathological states that are characterized by high cardiometabolic risk and end-organ damage, particularly at the level of the heart, vessels, kidney and adipose tissue. Diseases associated with an altered function of the aldosterone/MR system, such as hypertension, diabetes, chronic kidney diseases, obesity, heart failure, are distinguished by elevated mortality and costs.

To date, there are no validated clinical biomarkers of the aldosterone/MR system other than plasma circulating levels of aldosterone itself, plasma renin activity and electrolytes, in particular potassium [167]. However, plasma electrolytes only represent an indirect marker of the RAAS

status, and can be affected by several factors other than MR activation, such as plasma volume, salt intake, adrenergic tone, etc. Moreover, these readouts are not necessarily associated with organ damage, therefore they cannot be considered as a veritable signature of cardiometabolic diseases, fully able to identify high risk patients, eligible to intensive lifestyle or pharmacological intervention for cardiovascular protection.

Therefore, there is an unmet need for novel biomarkers that are able to detect the early stages of selective organ damage, mediated by the aldosterone-MR pathway.

An ideal diagnostic biomarker has to respect several criteria, such as a reasonable balance between cost and benefit, which favors a rapid and correct diagnosis, and should provide information on the patient health status [168].

Recently, the field of biomarkers has shifted from purely diagnostic aspects to risk stratification, therapeutic indications and prognosis. Novel biomarkers should preferably be involved in specific pathophysiological pathways, leading to the initiation or exacerbation of the disease. In this context, aldosterone-MR pathway has been carefully explored in the last years, due to its intimate connections with several comorbidities, and recent studies yielded potentially interesting novel biomarkers. Of course, more studies are deemed necessary to confirm their actual prognostic value, their ability to provide useful information on the patient health status beyond signs and symptoms or other already available techniques.

Here, we have discussed the impact of potential novel biomarkers related to the aldosterone/ MR system, which could help identify cardiovascular and metabolic detrimental conditions. Specifically, we focused on the effects of altered MR activation on distinct subpopulations of circulating and intra-tissue immune cells (Figure 1), and on MR downstream molecules (NGAL, Gal-3 and PTGDS), whose expression could represent a reliable biomarker of end-organ damage (Figure 3).



**Figure 3.** PTGDS, NGAL and Gal-3 as novel biomarkers in cardiovascular diseases induced by altered mineralocorticoid activation. MR activation by aldosterone induces the expression of different downstream molecules. PTGDS is expressed in adipose tissue, whereas NGAL and Gal-3 are expressed in the heart and vasculature; NGAL and Gal-3 are also detectable in the plasma. In obesity states, elevated leptin levels secreted by adipose tissue (in particular epicardial fat), directly activate heart MR, which in turn further promotes Gal-3 synthesis. All these molecules contribute to induce end-organ damage, through disarrangement of ECM and collagen. PTGDS: lipocalin-like prostaglandin D2 synthase; NGAL: neutrophil gelatinase-associated lipocalin; Gal-3: galectin-3.

**Author Contributions:** S.G. and M.C. conceived and wrote the manuscript; C.M. prepared the figures and revised the manuscript; and V.M. and A.A. wrote, in part, and revised the manuscript.

**Funding:** This research was funded by the Italian Ministry of Health (Ricerca Corrente and Bando 2011–2012 Progetti Collaborazione Ricercatori Italiani all'Estero, to M.C.) and by MIUR (Progetti di Ricerca di interesse Nazionale 2015—project code 2015ZTT5KB, to M.C., work package leader).

Acknowledgments: The authors wish to thank Amy Taheri from University of Zurich (CH) for language editing and proofreading the manuscript.

Conflicts of Interest: The authors declare no conflicts of interest.

# References

- 1. Rossier, B.C. Hormonal regulation of the epithelial sodium channel ENaC: N or P<sub>o</sub>? *J. Gen. Physiol.* 2002, 120, 67–70. [CrossRef] [PubMed]
- Rossier, B.C. Epithelial sodium channel (ENaC) and the control of blood pressure. *Curr. Opin. Pharmacol.* 2014, 15, 33–46. [CrossRef] [PubMed]
- 3. Rossier, B.C.; Staub, O.; Hummler, E. Genetic dissection of sodium and potassium transport along the aldosterone-sensitive distal nephron: Importance in the control of blood pressure and hypertension. *FEBS Lett.* **2013**, *587*, 1929–1941. [CrossRef] [PubMed]
- 4. Arriza, J.L.; Weinberger, C.; Cerelli, G.; Glaser, T.M.; Handelin, B.L.; Housman, D.E.; Evans, R.M. Cloning of human mineralocorticoid receptor complementary DNA: Structural and functional kinship with the glucocorticoid receptor. *Science* **1987**, *237*, 268–275. [CrossRef] [PubMed]
- Marzolla, V.; Armani, A.; Feraco, A.; De Martino, M.; Fabbri, A.; Rosano, G.; Caprio, M. Mineralocorticoid receptor in adipocytes and macrophages: A promising target to fight metabolic syndrome. *Steroids* 2014, *91*, 46–53. [CrossRef] [PubMed]
- Lombes, M.; Oblin, M.E.; Gasc, J.M.; Baulieu, E.E.; Farman, N.; Bonvalet, J.P. Immunohistochemical and biochemical evidence for a cardiovascular mineralocorticoid receptor. *Circ. Res.* 1992, 71, 503–510. [CrossRef] [PubMed]
- Lombes, M.; Alfaidy, N.; Eugene, E.; Lessana, A.; Farman, N.; Bonvalet, J.P. Prerequisite for cardiac aldosterone action. Mineralocorticoid receptor and 11 beta-hydroxysteroid dehydrogenase in the human heart. *Circulation* 1995, 92, 175–182. [CrossRef] [PubMed]
- Rondinone, C.M.; Rodbard, D.; Baker, M.E. Aldosterone stimulated differentiation of mouse 3T3-L1 cells into adipocytes. *Endocrinology* 1993, 132, 2421–2426. [CrossRef] [PubMed]
- 9. DuPont, J.J.; Jaffe, I.Z. 30 years of the mineralocorticoid receptor: The role of the mineralocorticoid receptor in the vasculature. *J. Endocrinol.* **2017**, 234, T67–T82. [CrossRef] [PubMed]
- Caprio, M.; Newfell, B.G.; La Sala, A.; Baur, W.; Fabbri, A.; Rosano, G.; Mendelsohn, M.E.; Jaffe, I.Z. Functional mineralocorticoid receptors in human vascular endothelial cells regulate intercellular adhesion molecule-1 expression and promote leukocyte adhesion. *Circ. Res.* 2008, *102*, 1359–1367. [CrossRef] [PubMed]
- 11. Armani, A.; Marzolla, V.; Fabbri, A.; Caprio, M. Cellular mechanisms of MR regulation of adipose tissue physiology and pathophysiology. *J. Mol. Endocrinol.* **2015**, *55*, R1–R10. [CrossRef] [PubMed]
- Funder, J.W. Aldosterone and mineralocorticoid receptors-physiology and pathophysiology. *Int. J. Mol. Sci.* 2017, *18*, 1032. [CrossRef] [PubMed]
- Bentley-Lewis, R.; Adler, G.K.; Perlstein, T.; Seely, E.W.; Hopkins, P.N.; Williams, G.H.; Garg, R. Body mass index predicts aldosterone production in normotensive adults on a high-salt diet. *J. Clin. Endocrinol. Metab.* 2007, 92, 4472–4475. [CrossRef] [PubMed]
- 14. Goodfriend, T.L.; Kelley, D.E.; Goodpaster, B.H.; Winters, S.J. Visceral obesity and insulin resistance are associated with plasma aldosterone levels in women. *Obes. Res.* **1999**, *7*, 355–362. [CrossRef] [PubMed]
- 15. Williams, J.S.; Williams, G.H. 50th anniversary of aldosterone. *J. Clin. Endocrinol. Metab.* **2003**, *88*, 2364–2372. [CrossRef] [PubMed]
- Jaffe, I.Z.; Mendelsohn, M.E. Angiotensin II and aldosterone regulate gene transcription via functional mineralocortocoid receptors in human coronary artery smooth muscle cells. *Circ. Res.* 2005, *96*, 643–650. [CrossRef] [PubMed]

- Jaffe, I.Z.; Tintut, Y.; Newfell, B.G.; Demer, L.L.; Mendelsohn, M.E. Mineralocorticoid receptor activation promotes vascular cell calcification. *Arterioscler. Thromb. Vasc. Biol.* 2007, 27, 799–805. [CrossRef] [PubMed]
- 18. Park, J.B.; Schiffrin, E.L. Cardiac and vascular fibrosis and hypertrophy in aldosterone-infused rats: Role of endothelin-1. *Am. J. Hypertens.* **2002**, *15*, 164–169. [CrossRef]
- 19. Harvey, A.P.; Montezano, A.C.; Hood, K.Y.; Lopes, R.A.; Rios, F.; Ceravolo, G.; Graham, D.; Touyz, R.M. Vascular dysfunction and fibrosis in stroke-prone spontaneously hypertensive rats: The aldosterone-mineralocorticoid receptor-Nox1 axis. *Life Sci.* **2017**, *179*, 110–119. [CrossRef] [PubMed]
- 20. Gomez-Sanchez, E.; Gomez-Sanchez, C.E. The multifaceted mineralocorticoid receptor. *Compr. Physiol.* **2014**, *4*, 965–994. [PubMed]
- Shibata, S.; Nagase, M.; Yoshida, S.; Kawarazaki, W.; Kurihara, H.; Tanaka, H.; Miyoshi, J.; Takai, Y.; Fujita, T. Modification of mineralocorticoid receptor function by Rac1 GTPase: Implication in proteinuric kidney disease. *Nat. Med.* 2008, *14*, 1370–1376. [CrossRef] [PubMed]
- 22. Nagase, M.; Fujita, T. Role of Rac1-mineralocorticoid-receptor signalling in renal and cardiac disease. *Nat. Rev. Nephrol.* **2013**, *9*, 86–98. [CrossRef] [PubMed]
- 23. Ayuzawa, N.; Nagase, M.; Ueda, K.; Nishimoto, M.; Kawarazaki, W.; Marumo, T.; Aiba, A.; Sakurai, T.; Shindo, T.; Fujita, T. Rac1-mediated activation of mineralocorticoid receptor in pressure overload-induced cardiac injury. *Hypertension* **2016**, *67*, 99–106. [CrossRef] [PubMed]
- Nagase, M.; Ayuzawa, N.; Kawarazaki, W.; Ishizawa, K.; Ueda, K.; Yoshida, S.; Fujita, T. Oxidative stress causes mineralocorticoid receptor activation in rat cardiomyocytes: Role of small GTPase Rac1. *Hypertension* 2012, 59, 500–506. [CrossRef] [PubMed]
- 25. Tapia-Castillo, A.; Carvajal, C.A.; Campino, C.; Hill, C.; Allende, F.; Vecchiola, A.; Carrasco, C.; Bancalari, R.; Valdivia, C.; Lagos, C.; et al. The expression of Rac1 and mineralocorticoid pathway-dependent genes are associated with different responses to salt intake. *Am. J. Hypertens.* **2015**, *28*, 722–728. [CrossRef] [PubMed]
- 26. Ruhs, S.; Nolze, A.; Hubschmann, R.; Grossmann, C. 30 years of the mineralocorticoid receptor: Nongenomic effects via the mineralocorticoid receptor. *J. Endocrinol.* **2017**, 234, T107–T124. [CrossRef] [PubMed]
- Wehling, M. Rapid actions of aldosterone revisited: Receptors in the limelight. J. Steroid Biochem. Mol. Biol. 2018, 176, 94–98. [CrossRef] [PubMed]
- Gekle, M.; Mildenberger, S.; Freudinger, R.; Grossmann, C. Altered collagen homeostasis in human aortic smooth muscle cells (HAoSMCs) induced by aldosterone. *Pflug. Arch.* 2007, 454, 403–413. [CrossRef] [PubMed]
- Huang, L.L.; Nikolic-Paterson, D.J.; Ma, F.Y.; Tesch, G.H. Aldosterone induces kidney fibroblast proliferation via activation of growth factor receptors and PI3K/MAPK signalling. *Nephron Exp. Nephrol.* 2012, 120, e115–e122. [CrossRef] [PubMed]
- 30. Krug, A.W.; Allenhofer, L.; Monticone, R.; Spinetti, G.; Gekle, M.; Wang, M.; Lakatta, E.G. Elevated mineralocorticoid receptor activity in aged rat vascular smooth muscle cells promotes a proinflammatory phenotype via extracellular signal-regulated kinase 1/2 mitogen-activated protein kinase and epidermal growth factor receptor-dependent pathways. *Hypertension* **2010**, *55*, 1476–1483. [PubMed]
- Zennaro, M.C.; Caprio, M.; Feve, B. Mineralocorticoid receptors in the metabolic syndrome. *Trends Endocrinol. Metab.* 2009, 20, 444–451. [CrossRef] [PubMed]
- 32. Marney, A.M.; Brown, N.J. Aldosterone and end-organ damage. *Clin. Sci. (Lond.)* 2007, 113, 267–278. [CrossRef] [PubMed]
- Catena, C.; Colussi, G.; Sechi, L.A. Aldosterone, organ damage and dietary salt. *Clin. Exp. Pharmacol. Physiol.* 2013, 40, 922–928. [CrossRef] [PubMed]
- 34. Brilla, C.G.; Weber, K.T. Mineralocorticoid excess, dietary sodium, and myocardial fibrosis. *J. Lab. Clin. Med.* **1992**, *120*, 893–901. [PubMed]
- 35. Rossi, G.P. Primary aldosteronism: A needle in a haystack or a yellow cab on fifth avenue? *Curr. Hypertens. Rep.* **2004**, *6*, 1–4. [CrossRef] [PubMed]
- Milliez, P.; Girerd, X.; Plouin, P.F.; Blacher, J.; Safar, M.E.; Mourad, J.J. Evidence for an increased rate of cardiovascular events in patients with primary aldosteronism. *J. Am. Coll. Cardiol.* 2005, 45, 1243–1248. [CrossRef] [PubMed]
- 37. Conn, J.W. Hypertension, the potassium ion and impaired carbohydrate tolerance. *N. Engl. J. Med.* **1965**, 273, 1135–1143. [CrossRef] [PubMed]

- Kidambi, S.; Kotchen, J.M.; Grim, C.E.; Raff, H.; Mao, J.; Singh, R.J.; Kotchen, T.A. Association of adrenal steroids with hypertension and the metabolic syndrome in blacks. *Hypertension* 2007, 49, 704–711. [CrossRef] [PubMed]
- Fallo, F.; Veglio, F.; Bertello, C.; Sonino, N.; Della Mea, P.; Ermani, M.; Rabbia, F.; Federspil, G.; Mulatero, P. Prevalence and characteristics of the metabolic syndrome in primary aldosteronism. *J. Clin. Endocrinol. Metab.* 2006, 91, 454–459. [CrossRef] [PubMed]
- 40. Jin, H.M.; Zhou, D.C.; Gu, H.F.; Qiao, Q.Y.; Fu, S.K.; Liu, X.L.; Pan, Y. Antioxidant N-acetylcysteine protects pancreatic β-cells against aldosterone-induced oxidative stress and apoptosis in female db/db mice and insulin-producing MIN6 cells. *Endocrinology* **2013**, *154*, 4068–4077. [CrossRef] [PubMed]
- 41. Luther, J.M. Effects of aldosterone on insulin sensitivity and secretion. *Steroids* **2014**, *91*, 54–60. [CrossRef] [PubMed]
- Giacchetti, G.; Ronconi, V.; Turchi, F.; Agostinelli, L.; Mantero, F.; Rilli, S.; Boscaro, M. Aldosterone as a key mediator of the cardiometabolic syndrome in primary aldosteronism: An observational study. *J. Hypertens.* 2007, 25, 177–186. [CrossRef] [PubMed]
- 43. Sindelka, G.; Widimsky, J.; Haas, T.; Prazny, M.; Hilgertova, J.; Skrha, J. Insulin action in primary hyperaldosteronism before and after surgical or pharmacological treatment. *Exp. Clin. Endocrinol. Diabetes* **2000**, *108*, 21–25. [CrossRef] [PubMed]
- 44. Widimsky, J., Jr.; Sindelka, G.; Haas, T.; Prazny, M.; Hilgertova, J.; Skrha, J. Impaired insulin action in primary hyperaldosteronism. *Physiol. Res.* **2000**, *49*, 241–244. [PubMed]
- Catena, C.; Lapenna, R.; Baroselli, S.; Nadalini, E.; Colussi, G.; Novello, M.; Favret, G.; Melis, A.; Cavarape, A.; Sechi, L.A. Insulin sensitivity in patients with primary aldosteronism: A follow-up study. *J. Clin. Endocrinol. Metab.* 2006, *91*, 3457–3463. [CrossRef] [PubMed]
- 46. Ingelsson, E.; Pencina, M.J.; Tofler, G.H.; Benjamin, E.J.; Lanier, K.J.; Jacques, P.F.; Fox, C.S.; Meigs, J.B.; Levy, D.; Larson, M.G.; et al. Multimarker approach to evaluate the incidence of the metabolic syndrome and longitudinal changes in metabolic risk factors: The framingham offspring study. *Circulation* 2007, 116, 984–992. [CrossRef] [PubMed]
- Freel, E.M.; Mark, P.B.; Weir, R.A.; McQuarrie, E.P.; Allan, K.; Dargie, H.J.; McClure, J.D.; Jardine, A.G.; Davies, E.; Connell, J.M. Demonstration of blood pressure-independent noninfarct myocardial fibrosis in primary aldosteronism: A cardiac magnetic resonance imaging study. *Circ. Cardiovasc. Imaging* 2012, 5,740–747. [CrossRef] [PubMed]
- 48. Zennaro, M.C.; Boulkroun, S.; Fernandes-Rosa, F. An update on novel mechanisms of primary aldosteronism. *J. Endocrinol.* **2015**, 224, R63–R77. [CrossRef] [PubMed]
- Rossi, G.P.; Sacchetto, A.; Pavan, E.; Palatini, P.; Graniero, G.R.; Canali, C.; Pessina, A.C. Remodeling of the left ventricle in primary aldosteronism due to conn's adenoma. *Circulation* 1997, 95, 1471–1478. [CrossRef] [PubMed]
- Hillaert, M.A.; Lentjes, E.G.; Beygui, F.; Kemperman, H.; Asselbergs, F.W.; Nathoe, H.M.; Agostoni, P.; Voskuil, M.; Ivanes, F.; Jude, B.; et al. Measuring and targeting aldosterone and renin in atherosclerosis—A review of clinical data. *Am. Heart J.* 2011, *162*, 585–596. [CrossRef] [PubMed]
- Reil, J.C.; Hohl, M.; Selejan, S.; Lipp, P.; Drautz, F.; Kazakow, A.; Munz, B.M.; Muller, P.; Steendijk, P.; Reil, G.H.; et al. Aldosterone promotes atrial fibrillation. *Eur. Heart J.* 2012, *33*, 2098–2108. [CrossRef] [PubMed]
- 52. Barter, P.J.; Caulfield, M.; Eriksson, M.; Grundy, S.M.; Kastelein, J.J.; Komajda, M.; Lopez-Sendon, J.; Mosca, L.; Tardif, J.C.; Waters, D.D.; et al. Effects of torcetrapib in patients at high risk for coronary events. *N. Engl. J. Med.* **2007**, 357, 2109–2122. [CrossRef] [PubMed]
- Zannad, F.; McMurray, J.J.; Krum, H.; van Veldhuisen, D.J.; Swedberg, K.; Shi, H.; Vincent, J.; Pocock, S.J.; Pitt, B.; Group, E.-H.S. Eplerenone in patients with systolic heart failure and mild symptoms. *N. Engl. J. Med.* 2011, 364, 11–21. [CrossRef] [PubMed]
- 54. MacFadyen, R.J.; Barr, C.S.; Struthers, A.D. Aldosterone blockade reduces vascular collagen turnover, improves heart rate variability and reduces early morning rise in heart rate in heart failure patients. *Cardiovasc. Res.* **1997**, *35*, 30–34. [CrossRef]
- 55. Gerling, I.C.; Sun, Y.; Ahokas, R.A.; Wodi, L.A.; Bhattacharya, S.K.; Warrington, K.J.; Postlethwaite, A.E.; Weber, K.T. Aldosteronism: An immunostimulatory state precedes proinflammatory/fibrogenic cardiac phenotype. *Am. J. Physiol. Heart Circ. Physiol.* **2003**, *285*, H813–H821. [CrossRef] [PubMed]

- 56. Sun, Y.; Zhang, J.; Lu, L.; Chen, S.S.; Quinn, M.T.; Weber, K.T. Aldosterone-induced inflammation in the rat heart: Role of oxidative stress. *Am. J. Pathol.* **2002**, *161*, 1773–1781. [CrossRef]
- 57. Guzik, T.J.; Hoch, N.E.; Brown, K.A.; McCann, L.A.; Rahman, A.; Dikalov, S.; Goronzy, J.; Weyand, C.; Harrison, D.G. Role of the T cell in the genesis of angiotensin II induced hypertension and vascular dysfunction. *J. Exp. Med.* **2007**, *204*, 2449–2460. [CrossRef] [PubMed]
- 58. Marvar, P.J.; Thabet, S.R.; Guzik, T.J.; Lob, H.E.; McCann, L.A.; Weyand, C.; Gordon, F.J.; Harrison, D.G. Central and peripheral mechanisms of t-lymphocyte activation and vascular inflammation produced by angiotensin II-induced hypertension. *Circ. Res.* **2010**, *107*, 263–270. [CrossRef] [PubMed]
- Usher, M.G.; Duan, S.Z.; Ivaschenko, C.Y.; Frieler, R.A.; Berger, S.; Schutz, G.; Lumeng, C.N.; Mortensen, R.M. Myeloid mineralocorticoid receptor controls macrophage polarization and cardiovascular hypertrophy and remodeling in mice. *J. Clin. Investig.* 2010, *120*, 3350–3364. [CrossRef] [PubMed]
- 60. Selye, H.; Stone, H.; Timiras, P.S.; Schaffenburg, C. Influence of sodium chloride upon the actions of desoxycorticosteron acetate. *Am. Heart J.* **1949**, *37*, 1009–1016. [CrossRef]
- 61. Funder, J.W. Mineralocorticoid receptor activation and oxidative stress. *Hypertension* **2007**, *50*, 840–841. [CrossRef] [PubMed]
- 62. Brown, N.J. Aldosterone and vascular inflammation. Hypertension 2008, 51, 161–167. [CrossRef] [PubMed]
- 63. Moraes, L.A.; Paul-Clark, M.J.; Rickman, A.; Flower, R.J.; Goulding, N.J.; Perretti, M. Ligand-specific glucocorticoid receptor activation in human platelets. *Blood* **2005**, *106*, 4167–4175. [CrossRef] [PubMed]
- 64. Lother, A.; Furst, D.; Bergemann, S.; Gilsbach, R.; Grahammer, F.; Huber, T.B.; Hilgendorf, I.; Bode, C.; Moser, M.; Hein, L. Deoxycorticosterone acetate/salt-induced cardiac but not renal injury is mediated by endothelial mineralocorticoid receptors independently from blood pressure. *Hypertension* **2016**, 67, 130–138. [CrossRef] [PubMed]
- Marzolla, V.; Armani, A.; Mammi, C.; Moss, M.E.; Pagliarini, V.; Pontecorvo, L.; Antelmi, A.; Fabbri, A.; Rosano, G.; Jaffe, I.Z.; et al. Essential role of ICAM-1 in aldosterone-induced atherosclerosis. *Int. J. Cardiol.* 2017, 232, 233–242. [CrossRef] [PubMed]
- 66. Caprio, M.; Mammi, C.; Jaffe, I.Z.; Zennaro, M.C.; Aversa, A.; Mendelsohn, M.E.; Fabbri, A.; Rosano, G.M. The mineralocorticoid receptor in endothelial physiology and disease: Novel concepts in the understanding of erectile dysfunction. *Curr. Pharm. Des.* 2008, 14, 3749–3757. [CrossRef] [PubMed]
- Young, M.J.; Rickard, A.J. Mechanisms of mineralocorticoid salt-induced hypertension and cardiac fibrosis. *Mol. Cell. Endocrinol.* 2012, 350, 248–255. [CrossRef] [PubMed]
- Rickard, A.J.; Morgan, J.; Chrissobolis, S.; Miller, A.A.; Sobey, C.G.; Young, M.J. Endothelial cell mineralocorticoid receptors regulate deoxycorticosterone/salt-mediated cardiac remodeling and vascular reactivity but not blood pressure. *Hypertension* 2014, 63, 1033–1040. [CrossRef] [PubMed]
- 69. Tesch, G.H.; Young, M.J. Mineralocorticoid receptor signaling as a therapeutic target for renal and cardiac fibrosis. *Front. Pharmacol.* **2017**, *8*, 313. [CrossRef] [PubMed]
- Kadoya, H.; Satoh, M.; Sasaki, T.; Taniguchi, S.; Takahashi, M.; Kashihara, N. Excess aldosterone is a critical danger signal for inflammasome activation in the development of renal fibrosis in mice. *FASEB J.* 2015, 29, 3899–3910. [CrossRef] [PubMed]
- Bai, M.; Chen, Y.; Zhao, M.; Zhang, Y.; He, J.C.; Huang, S.; Jia, Z.; Zhang, A. NLRP3 inflammasome activation contributes to aldosterone-induced podocyte injury. *Am. J. Physiol. Renal. Physiol.* 2017, 312, F556–F564. [CrossRef] [PubMed]
- 72. Wada, T.; Ishikawa, A.; Watanabe, E.; Nakamura, Y.; Aruga, Y.; Hasegawa, H.; Onogi, Y.; Honda, H.; Nagai, Y.; Takatsu, K.; et al. Eplerenone prevented obesity-induced inflammasome activation and glucose intolerance. *J. Endocrinol.* **2017**, *235*, 179–191. [CrossRef] [PubMed]
- 73. Svendsen, U.G. Evidence for an initial, thymus independent and a chronic, thymus dependent phase of doca and salt hypertension in mice. *Acta Pathol. Microbiol. Scand. A* **1976**, *84*, 523–528. [CrossRef] [PubMed]
- Besedovsky, L.; Linz, B.; Born, J.; Lange, T. Mineralocorticoid receptor signaling reduces numbers of circulating human naïve T cells and increases their CD62L, CCR7, and CXCR4 expression. *Eur. J. Immunol.* 2014, 44, 1759–1769. [CrossRef] [PubMed]
- Madhur, M.S.; Lob, H.E.; McCann, L.A.; Iwakura, Y.; Blinder, Y.; Guzik, T.J.; Harrison, D.G. Interleukin 17 Promotes Angiotensin II-Induced Hypertension and Vascular Dysfunction. *Hypertension* 2010, 55, 500–507. [CrossRef] [PubMed]

- 76. Youn, J.C.; Yu, H.T.; Lim, B.J.; Koh, M.J.; Lee, J.; Chang, D.Y.; Choi, Y.S.; Lee, S.H.; Kang, S.M.; Jang, Y.; et al. Immunosenescent CD8+ T Cells and C-X-C Chemokine Receptor Type 3 Chemokines are Increased in Human Hypertension. *Hypertension* 2013, 62, 126–133. [CrossRef] [PubMed]
- 77. Amador, C.A.; Barrientos, V.; Pena, J.; Herrada, A.A.; Gonzalez, M.; Valdes, S.; Carrasco, L.; Alzamora, R.; Figueroa, F.; Kalergis, A.M.; et al. Spironolactone decreases doca-salt-induced organ damage by blocking the activation of T helper 17 and the downregulation of regulatory T lymphocytes. *Hypertension* 2014, 63, 797–803. [CrossRef] [PubMed]
- 78. Li, C.; Sun, X.N.; Zeng, M.R.; Zheng, X.J.; Zhang, Y.Y.; Wan, Q.; Zhang, W.C.; Shi, C.; Du, L.J.; Ai, T.J.; et al. Mineralocorticoid receptor deficiency in T cells attenuates pressure overload-induced cardiac hypertrophy and dysfunction through modulating T-cell activation. *Hypertension* 2017, 70, 137–147. [CrossRef] [PubMed]
- 79. Herrada, A.A.; Contreras, F.J.; Marini, N.P.; Amador, C.A.; Gonzalez, P.A.; Cortes, C.M.; Riedel, C.A.; Carvajal, C.A.; Figueroa, F.; Michea, L.F.; et al. Aldosterone promotes autoimmune damage by enhancing Th17-mediated immunity. *J. Immunol.* **2010**, *184*, 191–202. [CrossRef] [PubMed]
- Mellman, I.; Steinman, R.M. Dendritic cells: Specialized and regulated antigen processing machines. *Cell* 2001, 106, 255–258. [CrossRef]
- 81. Bettelli, E.; Carrier, Y.; Gao, W.; Korn, T.; Strom, T.B.; Oukka, M.; Weiner, H.L.; Kuchroo, V.K. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory t cells. *Nature* **2006**, 441, 235–238. [CrossRef] [PubMed]
- Zhang, Y.; Chung, Y.; Bishop, C.; Daugherty, B.; Chute, H.; Holst, P.; Kurahara, C.; Lott, F.; Sun, N.; Welcher, A.A.; et al. Regulation of T cell activation and tolerance by PDL2. *Proc. Natl. Acad. Sci. USA* 2006, 103, 11695–11700. [CrossRef] [PubMed]
- Elrefaei, M.; Baker, C.A.; Jones, N.G.; Bangsberg, D.R.; Cao, H. Presence of suppressor HIV-specific CD8+ T cells is associated with increased PD-1 expression on effector CD8+ T cells. *J. Immunol.* 2008, 180, 7757–7763. [CrossRef] [PubMed]
- 84. Jo, J.; Gavrilova, O.; Pack, S.; Jou, W.; Mullen, S.; Sumner, A.E.; Cushman, S.W.; Periwal, V. Hypertrophy and/or hyperplasia: Dynamics of adipose tissue growth. *PLoS Comput. Biol.* **2009**, *5*, e1000324. [CrossRef] [PubMed]
- 85. Sartipy, P.; Loskutoff, D.J. Monocyte chemoattractant protein 1 in obesity and insulin resistance. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 7265–7270. [CrossRef] [PubMed]
- 86. Bruun, J.M.; Pedersen, S.B.; Richelsen, B. Regulation of interleukin 8 production and gene expression in human adipose tissue in vitro. *J. Clin. Endocrinol. Metab.* **2001**, *86*, 1267–1273. [CrossRef] [PubMed]
- 87. Greenberg, A.S.; Obin, M.S. Obesity and the role of adipose tissue in inflammation and metabolism. *Am. J. Clin. Nutr.* **2006**, *83*, 461S–465S. [CrossRef] [PubMed]
- 88. Esser, N.; Legrand-Poels, S.; Piette, J.; Scheen, A.J.; Paquot, N. Inflammation as a link between obesity, metabolic syndrome and type 2 diabetes. *Diabetes Res. Clin. Pract.* **2014**, *105*, 141–150. [CrossRef] [PubMed]
- Lee, J.H.; Gao, Z.; Ye, J. Regulation of 11β-HSD1 expression during adipose tissue expansion by hypoxia through different activities of NF-κB and HIF-1α. *Am. J. Physiol. Endocrinol. Metab.* 2013, 304, E1035–E1041. [CrossRef] [PubMed]
- 90. Lumeng, C.N.; Bodzin, J.L.; Saltiel, A.R. Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *J. Clin. Investig.* **2007**, *117*, 175–184. [CrossRef] [PubMed]
- Hoppmann, J.; Perwitz, N.; Meier, B.; Fasshauer, M.; Hadaschik, D.; Lehnert, H.; Klein, J. The balance between gluco- and mineralo-corticoid action critically determines inflammatory adipocyte responses. *J. Endocrinol.* 2010, 204, 153–164. [CrossRef] [PubMed]
- 92. Hirata, A.; Maeda, N.; Hiuge, A.; Hibuse, T.; Fujita, K.; Okada, T.; Kihara, S.; Funahashi, T.; Shimomura, I. Blockade of mineralocorticoid receptor reverses adipocyte dysfunction and insulin resistance in obese mice. *Cardiovasc. Res.* 2009, *84*, 164–172. [CrossRef] [PubMed]
- 93. Labuzek, K.; Liber, S.; Buldak, L.; Machnik, G.; Liber, J.; Okopien, B. Eplerenone promotes alternative activation in human monocyte-derived macrophages. *Pharmacol. Rep.* **2013**, *65*, 226–234. [CrossRef]
- 94. Armani, A.; Cinti, F.; Marzolla, V.; Morgan, J.; Cranston, G.A.; Antelmi, A.; Carpinelli, G.; Canese, R.; Pagotto, U.; Quarta, C.; et al. Mineralocorticoid receptor antagonism induces browning of white adipose tissue through impairment of autophagy and prevents adipocyte dysfunction in high-fat-diet-fed mice. *FASEB J.* **2014**, *28*, 3745–3757. [CrossRef] [PubMed]

- 95. Guo, C.; Ricchiuti, V.; Lian, B.Q.; Yao, T.M.; Coutinho, P.; Romero, J.R.; Li, J.; Williams, G.H.; Adler, G.K. Mineralocorticoid Receptor Blockade Reverses Obesity-Related Changes in Expression of Adiponectin, Peroxisome Proliferator-Activated Receptor-gamma, and Proinflammatory Adipokines. *Circulation* 2008, 117, 2253–2261. [CrossRef] [PubMed]
- Caprio, M.; Feve, B.; Claes, A.; Viengchareun, S.; Lombes, M.; Zennaro, M.C. Pivotal role of the mineralocorticoid receptor in corticosteroid-induced adipogenesis. *FASEB J.* 2007, 21, 2185–2194. [CrossRef] [PubMed]
- Rosen, E.D.; Hsu, C.H.; Wang, X.; Sakai, S.; Freeman, M.W.; Gonzalez, F.J.; Spiegelman, B.M. C/EBPalpha induces adipogenesis through PPARgamma: A unified pathway. *Genes Dev.* 2002, 16, 22–26. [CrossRef] [PubMed]
- Caprio, M.; Antelmi, A.; Chetrite, G.; Muscat, A.; Mammi, C.; Marzolla, V.; Fabbri, A.; Zennaro, M.C.; Feve, B. Antiadipogenic effects of the mineralocorticoid receptor antagonist drospirenone: Potential implications for the treatment of metabolic syndrome. *Endocrinology* 2011, 152, 113–125. [CrossRef] [PubMed]
- 99. Urbanet, R.; Pilon, C.; Calcagno, A.; Peschechera, A.; Hubert, E.L.; Giacchetti, G.; Gomez-Sanchez, C.; Mulatero, P.; Toffanin, M.; Sonino, N.; et al. Analysis of insulin sensitivity in adipose tissue of patients with primary aldosteronism. *J. Clin. Endocrinol. Metab.* **2010**, *95*, 4037–4042. [CrossRef] [PubMed]
- 100. Armani, A.; Marzolla, V.; Rosano, G.; Caprio, M. Mineralocorticoid vs glucocorticoid receptors: Solo players or team mates in the control of adipogenesis? *Int. J. Obes. (Lond.)* **2014**, *38*, 1580–1581. [CrossRef] [PubMed]
- 101. Urbanet, R.; Nguyen Dinh Cat, A.; Feraco, A.; Venteclef, N.; El Mogrhabi, S.; Sierra-Ramos, C.; Alvarez de la Rosa, D.; Adler, G.K.; Quilliot, D.; Rossignol, P.; et al. Adipocyte mineralocorticoid receptor activation leads to metabolic syndrome and induction of prostaglandin D2 synthase. *Hypertension* 2015, 66, 149–157. [CrossRef] [PubMed]
- 102. Lavie, C.J.; Milani, R.V.; Ventura, H.O. Obesity and cardiovascular disease: Risk factor, paradox, and impact of weight loss. J. Am. Coll. Cardiol. 2009, 53, 1925–1932. [CrossRef] [PubMed]
- 103. Bochud, M.; Nussberger, J.; Bovet, P.; Maillard, M.R.; Elston, R.C.; Paccaud, F.; Shamlaye, C.; Burnier, M. Plasma aldosterone is independently associated with the metabolic syndrome. *Hypertension* 2006, 48, 239–245. [CrossRef] [PubMed]
- Garg, R.; Adler, G.K. Role of mineralocorticoid receptor in insulin resistance. *Curr. Opin. Endocrinol. Diabetes Obes.* 2012, 19, 168–175. [CrossRef] [PubMed]
- 105. Tirosh, A.; Garg, R.; Adler, G.K. Mineralocorticoid receptor antagonists and the metabolic syndrome. *Curr. Hypertens. Rep.* **2010**, *12*, 252–257. [CrossRef] [PubMed]
- 106. Min, S.H.; Kim, S.H.; Jeong, I.K.; Cho, H.C.; Jeong, J.O.; Lee, J.H.; Kang, H.J.; Kim, H.S.; Park, K.S.; Lim, S. Independent association of serum aldosterone level with metabolic syndrome and insulin resistance in korean adults. *Korean Circ. J.* 2018, 48, 198–208. [CrossRef] [PubMed]
- 107. Bjorntorp, P.; Rosmond, R. Obesity and cortisol. Nutrition 2000, 16, 924–936. [CrossRef]
- Kawarazaki, W.; Fujita, T. The role of aldosterone in obesity-related hypertension. *Am. J. Hypertens.* 2016, 29, 415–423. [CrossRef] [PubMed]
- 109. Calhoun, D.A.; Sharma, K. The role of aldosteronism in causing obesity-related cardiovascular risk. *Cardiol. Clin.* **2010**, *28*, 517–527. [CrossRef] [PubMed]
- 110. Infante, M.; Armani, A.; Mammi, C.; Fabbri, A.; Caprio, M. Impact of adrenal steroids on regulation of adipose tissue. *Compr. Physiol.* **2017**, *7*, 1425–1447. [PubMed]
- 111. Ehrhart-Bornstein, M.; Lamounier-Zepter, V.; Schraven, A.; Langenbach, J.; Willenberg, H.S.; Barthel, A.; Hauner, H.; McCann, S.M.; Scherbaum, W.A.; Bornstein, S.R. Human adipocytes secrete mineralocorticoid-releasing factors. *Proc. Natl. Acad. Sci. USA* 2003, 100, 14211–14216. [CrossRef] [PubMed]
- Marzolla, V.; Armani, A.; Zennaro, M.C.; Cinti, F.; Mammi, C.; Fabbri, A.; Rosano, G.M.; Caprio, M. The role of the mineralocorticoid receptor in adipocyte biology and fat metabolism. *Mol. Cell. Endocrinol.* 2012, 350, 281–288. [CrossRef] [PubMed]
- 113. Huby, A.C.; Antonova, G.; Groenendyk, J.; Gomez-Sanchez, C.E.; Bollag, W.B.; Filosa, J.A.; Belin de Chantemele, E.J. Adipocyte-derived hormone leptin is a direct regulator of aldosterone secretion, which promotes endothelial dysfunction and cardiac fibrosis. *Circulation* 2015, 132, 2134–2145. [CrossRef] [PubMed]
- 114. Diez, J. Arterial hypertension in patients with heart failure. *Heart Fail. Clin.* **2014**, *10*, 233–242. [CrossRef] [PubMed]

- Bonnans, C.; Chou, J.; Werb, Z. Remodelling the extracellular matrix in development and disease. *Nat. Rev. Mol. Cell Biol.* 2014, 15, 786–801. [CrossRef] [PubMed]
- 116. Talman, V.; Ruskoaho, H. Cardiac fibrosis in myocardial infarction-from repair and remodeling to regeneration. *Cell Tissue Res.* **2016**, *365*, 563–581. [CrossRef] [PubMed]
- 117. Vanhoutte, D.; van Almen, G.C.; Van Aelst, L.N.; Van Cleemput, J.; Droogne, W.; Jin, Y.; Van de Werf, F.; Carmeliet, P.; Vanhaecke, J.; Papageorgiou, A.P.; et al. Matricellular proteins and matrix metalloproteinases mark the inflammatory and fibrotic response in human cardiac allograft rejection. *Eur. Heart J.* 2013, 34, 1930–1941. [CrossRef] [PubMed]
- 118. Zia, A.A.; Kamalov, G.; Newman, K.P.; McGee, J.E.; Bhattacharya, S.K.; Ahokas, R.A.; Sun, Y.; Gerling, I.C.; Weber, K.T. From aldosteronism to oxidative stress: The role of excessive intracellular calcium accumulation. *Hypertens. Res.* 2010, 33, 1091–1101. [CrossRef] [PubMed]
- 119. Tsybouleva, N.; Zhang, L.; Chen, S.; Patel, R.; Lutucuta, S.; Nemoto, S.; DeFreitas, G.; Entman, M.; Carabello, B.A.; Roberts, R.; et al. Aldosterone, through novel signaling proteins, is a fundamental molecular bridge between the genetic defect and the cardiac phenotype of hypertrophic cardiomyopathy. *Circulation* 2004, 109, 1284–1291. [CrossRef] [PubMed]
- Sowers, J.; Tuck, M.; Asp, N.D.; Sollars, E. Plasma aldosterone and corticosterone responses to adrenocorticotropin, angiotensin, potassium, and stress in spontaneously hypertensive rats. *Endocrinology* 1981, 108, 1216–1221. [CrossRef] [PubMed]
- 121. Iacobone, M.; Citton, M.; Viel, G.; Rossi, G.P.; Nitti, D. Approach to the surgical management of primary aldosteronism. *Gland Surg.* 2015, *4*, 69–81. [PubMed]
- 122. Pitt, B.; Zannad, F.; Remme, W.J.; Cody, R.; Castaigne, A.; Perez, A.; Palensky, J.; Wittes, J. The effect of spironolactone on morbidity and mortality in patients with severe heart failure. Randomized aldactone evaluation study investigators. *N. Engl. J. Med.* **1999**, *341*, 709–717. [CrossRef] [PubMed]
- 123. Pitt, B.; Remme, W.; Zannad, F.; Neaton, J.; Martinez, F.; Roniker, B.; Bittman, R.; Hurley, S.; Kleiman, J.; Gatlin, M.; et al. Eplerenone, a selective aldosterone blocker, in patients with left ventricular dysfunction after myocardial infarction. *N. Engl. J. Med.* 2003, *348*, 1309–1321. [CrossRef] [PubMed]
- 124. Latouche, C.; El Moghrabi, S.; Messaoudi, S.; Nguyen Dinh Cat, A.; Hernandez-Diaz, I.; Alvarez de la Rosa, D.; Perret, C.; Lopez Andres, N.; Rossignol, P.; Zannad, F.; et al. Neutrophil gelatinase-associated lipocalin is a novel mineralocorticoid target in the cardiovascular system. *Hypertension* 2012, *59*, 966–972. [CrossRef] [PubMed]
- Schmidt-Ott, K.M.; Mori, K.; Li, J.Y.; Kalandadze, A.; Cohen, D.J.; Devarajan, P.; Barasch, J. Dual action of neutrophil gelatinase-associated lipocalin. J. Am. Soc. Nephrol. 2007, 18, 407–413. [CrossRef] [PubMed]
- 126. Liu, F.; Yang, H.; Chen, H.; Zhang, M.; Ma, Q. High expression of neutrophil gelatinase-associated lipocalin (NGAL) in the kidney proximal tubules of diabetic rats. *Adv. Med. Sci.* 2015, 60, 133–138. [CrossRef] [PubMed]
- 127. Hamzic, N.; Blomqvist, A.; Nilsberth, C. Immune-induced expression of lipocalin-2 in brain endothelial cells: Relationship with interleukin-6, cyclooxygenase-2 and the febrile response. *J. Neuroendocrinol.* 2013, 25, 271–280. [CrossRef] [PubMed]
- 128. Eilenberg, W.; Stojkovic, S.; Piechota-Polanczyk, A.; Kaun, C.; Rauscher, S.; Groger, M.; Klinger, M.; Wojta, J.; Neumayer, C.; Huk, I.; et al. Neutrophil gelatinase-associated lipocalin (NGAL) is associated with symptomatic carotid atherosclerosis and drives pro-inflammatory state in vitro. *Eur. J. Vasc. Endovasc. Surg.* 2016, 51, 623–631. [CrossRef] [PubMed]
- 129. Kjeldsen, L.; Johnsen, A.H.; Sengelov, H.; Borregaard, N. Isolation and primary structure of ngal, a novel protein associated with human neutrophil gelatinase. *J. Biol. Chem.* **1993**, *268*, 10425–10432. [PubMed]
- Flo, T.H.; Smith, K.D.; Sato, S.; Rodriguez, D.J.; Holmes, M.A.; Strong, R.K.; Akira, S.; Aderem, A. Lipocalin 2 mediates an innate immune response to bacterial infection by sequestrating iron. *Nature* 2004, 432, 917–921. [CrossRef] [PubMed]
- Floderer, M.; Prchal-Murphy, M.; Vizzardelli, C. Dendritic cell-secreted lipocalin2 induces CD8+ T-cell apoptosis, contributes to t-cell priming and leads to a TH1 phenotype. *PLoS ONE* 2014, 9, e101881. [CrossRef] [PubMed]
- 132. Devarajan, P. Neutrophil gelatinase-associated lipocalin: A promising biomarker for human acute kidney injury. *Biomark. Med.* **2010**, *4*, 265–280. [CrossRef] [PubMed]

- 133. Van Deursen, V.M.; Damman, K.; Voors, A.A.; van der Wal, M.H.; Jaarsma, T.; van Veldhuisen, D.J.; Hillege, H.L. Prognostic value of plasma neutrophil gelatinase-associated lipocalin for mortality in patients with heart failure. *Circ. Heart Fail.* **2014**, *7*, 35–42. [CrossRef] [PubMed]
- 134. Wu, G.; Li, H.; Fang, Q.; Jiang, S.; Zhang, L.; Zhang, J.; Hou, X.; Lu, J.; Bao, Y.; Xu, A.; et al. Elevated circulating lipocalin-2 levels independently predict incident cardiovascular events in men in a population-based cohort. *Arterioscler. Thromb. Vasc. Biol.* 2014, 34, 2457–2464. [CrossRef] [PubMed]
- 135. Tarjus, A.; Martinez-Martinez, E.; Amador, C.; Latouche, C.; El Moghrabi, S.; Berger, T.; Mak, T.W.; Fay, R.; Farman, N.; Rossignol, P.; et al. Neutrophil gelatinase-associated lipocalin, a novel mineralocorticoid biotarget, mediates vascular profibrotic effects of mineralocorticoids. *Hypertension* 2015, *66*, 158–166. [CrossRef] [PubMed]
- 136. Martinez-Martinez, E.; Buonafine, M.; Boukhalfa, I.; Ibarrola, J.; Fernandez-Celis, A.; Kolkhof, P.; Rossignol, P.; Girerd, N.; Mulder, P.; Lopez-Andres, N.; et al. Aldosterone target NGAL (neutrophil gelatinase-associated lipocalin) is involved in cardiac remodeling after myocardial infarction through NFκβ pathway. *Hypertension* 2017, 70, 1148–1156. [CrossRef] [PubMed]
- 137. Han, M.; Li, Y.; Liu, M.; Li, Y.; Cong, B. Renal neutrophil gelatinase associated lipocalin expression in lipopolysaccharide-induced acute kidney injury in the rat. *BMC Nephrol.* **2012**, *13*, 25. [CrossRef] [PubMed]
- 138. Shashidharamurthy, R.; Machiah, D.; Aitken, J.D.; Putty, K.; Srinivasan, G.; Chassaing, B.; Parkos, C.A.; Selvaraj, P.; Vijay-Kumar, M. Differential role of lipocalin 2 during immune complex-mediated acute and chronic inflammation in mice. *Arthritis Rheum.* 2013, 65, 1064–1073. [CrossRef] [PubMed]
- 139. Buonafine, M.; Martinez-Martinez, E.; Amador, C.; Gravez, B.; Ibarrola, J.; Fernandez-Celis, A.; El Moghrabi, S.; Rossignol, P.; Lopez-Andres, N.; Jaisser, F. Neutrophil gelatinase-associated lipocalin from immune cells is mandatory for aldosterone-induced cardiac remodeling and inflammation. *J. Mol. Cell. Cardiol.* 2018, 115, 32–38. [CrossRef] [PubMed]
- 140. Azibani, F.; Benard, L.; Schlossarek, S.; Merval, R.; Tournoux, F.; Fazal, L.; Polidano, E.; Launay, J.M.; Carrier, L.; Chatziantoniou, C.; et al. Aldosterone inhibits antifibrotic factors in mouse hypertensive heart. *Hypertension* **2012**, *59*, 1179–1187. [CrossRef] [PubMed]
- 141. Martinez-Martinez, E.; Jurado-Lopez, R.; Valero-Munoz, M.; Bartolome, M.V.; Ballesteros, S.; Luaces, M.; Briones, A.M.; Lopez-Andres, N.; Miana, M.; Cachofeiro, V. Leptin induces cardiac fibrosis through galectin-3, mtor and oxidative stress: Potential role in obesity. *J. Hypertens.* **2014**, *32*, 1104–1114. [CrossRef] [PubMed]
- 142. Thijssen, V.L.; Hulsmans, S.; Griffioen, A.W. The galectin profile of the endothelium: Altered expression and localization in activated and tumor endothelial cells. *Am. J. Pathol.* **2008**, *172*, 545–553. [CrossRef] [PubMed]
- 143. Papaspyridonos, M.; McNeill, E.; de Bono, J.P.; Smith, A.; Burnand, K.G.; Channon, K.M.; Greaves, D.R. Galectin-3 is an amplifier of inflammation in atherosclerotic plaque progression through macrophage activation and monocyte chemoattraction. *Arterioscler. Thromb. Vasc. Biol.* 2008, 28, 433–440. [CrossRef] [PubMed]
- 144. Calvier, L.; Miana, M.; Reboul, P.; Cachofeiro, V.; Martinez-Martinez, E.; de Boer, R.A.; Poirier, F.; Lacolley, P.; Zannad, F.; Rossignol, P.; et al. Galectin-3 mediates aldosterone-induced vascular fibrosis. *Arterioscler. Thromb. Vasc. Biol.* 2013, 33, 67–75. [CrossRef] [PubMed]
- 145. Calvier, L.; Martinez-Martinez, E.; Miana, M.; Cachofeiro, V.; Rousseau, E.; Sadaba, J.R.; Zannad, F.; Rossignol, P.; Lopez-Andres, N. The impact of galectin-3 inhibition on aldosterone-induced cardiac and renal injuries. *JACC Heart Fail.* **2015**, *3*, 59–67. [CrossRef] [PubMed]
- Glinsky, V.V.; Raz, A. Modified citrus pectin anti-metastatic properties: One bullet, multiple targets. *Carbohydr. Res.* 2009, 344, 1788–1791. [CrossRef] [PubMed]
- 147. Martinez-Martinez, E.; Calvier, L.; Fernandez-Celis, A.; Rousseau, E.; Jurado-Lopez, R.; Rossoni, L.V.; Jaisser, F.; Zannad, F.; Rossignol, P.; Cachofeiro, V.; et al. Galectin-3 blockade inhibits cardiac inflammation and fibrosis in experimental hyperaldosteronism and hypertension. *Hypertension* **2015**, *66*, 767–775. [CrossRef] [PubMed]
- 148. Lopez-Andres, N.; Rossignol, P.; Iraqi, W.; Fay, R.; Nuee, J.; Ghio, S.; Cleland, J.G.; Zannad, F.; Lacolley, P. Association of galectin-3 and fibrosis markers with long-term cardiovascular outcomes in patients with heart failure, left ventricular dysfunction, and dyssynchrony: Insights from the CARE-HF (cardiac resynchronization in heart failure) trial. *Eur. J. Heart Fail.* **2012**, *14*, 74–81. [CrossRef] [PubMed]

- Maiolino, G.; Rossitto, G.; Pedon, L.; Cesari, M.; Frigo, A.C.; Azzolini, M.; Plebani, M.; Rossi, G.P. Galectin-3 predicts long-term cardiovascular death in high-risk patients with coronary artery disease. *Arterioscler. Thromb. Vasc. Biol.* 2015, *35*, 725–732. [CrossRef] [PubMed]
- 150. Martinez-Martinez, E.; Lopez-Andres, N.; Jurado-Lopez, R.; Rousseau, E.; Bartolome, M.V.; Fernandez-Celis, A.; Rossignol, P.; Islas, F.; Antequera, A.; Prieto, S.; et al. Galectin-3 participates in cardiovascular remodeling associated with obesity. *Hypertension* **2015**, *66*, 961–969. [CrossRef] [PubMed]
- 151. Hirata, A.; Maeda, N.; Nakatsuji, H.; Hiuge-Shimizu, A.; Okada, T.; Funahashi, T.; Shimomura, I. Contribution of glucocorticoid-mineralocorticoid receptor pathway on the obesity-related adipocyte dysfunction. *Biochem. Biophys. Res. Commun.* **2012**, *419*, 182–187. [CrossRef] [PubMed]
- 152. Wada, T.; Kenmochi, H.; Miyashita, Y.; Sasaki, M.; Ojima, M.; Sasahara, M.; Koya, D.; Tsuneki, H.; Sasaoka, T. Spironolactone improves glucose and lipid metabolism by ameliorating hepatic steatosis and inflammation and suppressing enhanced gluconeogenesis induced by high-fat and high-fructose diet. *Endocrinology* 2010, 151, 2040–2049. [CrossRef] [PubMed]
- 153. Martinez-Martinez, E.; Calvier, L.; Rossignol, P.; Rousseau, E.; Fernandez-Celis, A.; Jurado-Lopez, R.; Laville, M.; Cachofeiro, V.; Lopez-Andres, N. Galectin-3 inhibition prevents adipose tissue remodelling in obesity. *Int. J. Obes. (Lond.)* **2016**, *40*, 1034–1038. [CrossRef] [PubMed]
- 154. Iacobellis, G.; Malavazos, A.E.; Corsi, M.M. Epicardial fat: From the biomolecular aspects to the clinical practice. *Int. J. Biochem. Cell Biol.* **2011**, *43*, 1651–1654. [CrossRef] [PubMed]
- 155. Rahmouni, K. Leptin-induced sympathetic nerve activation: Signaling mechanisms and cardiovascular consequences in obesity. *Curr. Hypertens. Rev.* **2010**, *6*, 104–209. [CrossRef] [PubMed]
- Rahmouni, K.; Morgan, D.A.; Morgan, G.M.; Mark, A.L.; Haynes, W.G. Role of selective leptin resistance in diet-induced obesity hypertension. *Diabetes* 2005, 54, 2012–2018. [CrossRef] [PubMed]
- 157. Schram, K.; Sweeney, G. Implications of myocardial matrix remodeling by adipokines in obesity-related heart failure. *Trends Cardiovasc. Med.* **2008**, *18*, 199–205. [CrossRef] [PubMed]
- 158. Kim, H.; Lee, J.; Hyun, J.W.; Park, J.W.; Joo, H.G.; Shin, T. Expression and immunohistochemical localization of galectin-3 in various mouse tissues. *Cell Biol. Int.* **2007**, *31*, 655–662. [CrossRef] [PubMed]
- 159. De Boer, R.A.; van Veldhuisen, D.J.; Gansevoort, R.T.; Muller Kobold, A.C.; van Gilst, W.H.; Hillege, H.L.; Bakker, S.J.; van der Harst, P. The fibrosis marker galectin-3 and outcome in the general population. *J. Intern. Med.* **2012**, 272, 55–64. [CrossRef] [PubMed]
- 160. Weigert, J.; Neumeier, M.; Wanninger, J.; Bauer, S.; Farkas, S.; Scherer, M.N.; Schnitzbauer, A.; Schaffler, A.; Aslanidis, C.; Scholmerich, J.; et al. Serum galectin-3 is elevated in obesity and negatively correlates with glycosylated hemoglobin in type 2 diabetes. *J. Clin. Endocrinol. Metab.* 2010, *95*, 1404–1411. [CrossRef] [PubMed]
- 161. Lin, Y.H.; Chou, C.H.; Wu, X.M.; Chang, Y.Y.; Hung, C.S.; Chen, Y.H.; Tzeng, Y.L.; Wu, V.C.; Ho, Y.L.; Hsieh, F.J.; et al. Aldosterone induced galectin-3 secretion in vitro and in vivo: From cells to humans. *PLoS ONE* **2014**, *9*, e95254. [CrossRef] [PubMed]
- 162. Jowsey, I.R.; Murdock, P.R.; Moore, G.B.; Murphy, G.J.; Smith, S.A.; Hayes, J.D. Prostaglandin D<sub>2</sub> synthase enzymes and PPARγ are co-expressed in mouse 3T3-L1 adipocytes and human tissues. *Prostaglandins Other Lipid Mediat.* 2003, 70, 267–284. [CrossRef]
- 163. Ragolia, L.; Palaia, T.; Hall, C.E.; Maesaka, J.K.; Eguchi, N.; Urade, Y. Accelerated Glucose Intolerance, Nephropathy, and Atherosclerosis in Prostaglandin D<sub>2</sub> Synthase Knock-out Mice. *J. Biol. Chem.* 2005, 280, 29946–29955. [CrossRef] [PubMed]
- 164. Ragolia, L.; Hall, C.E.; Palaia, T. Lipocalin-type prostaglandin D<sub>2</sub> synthase stimulates glucose transport via enhanced GLUT4 translocation. *Prostaglandins Other Lipid Mediat*. **2008**, *87*, 34–41. [CrossRef] [PubMed]
- 165. Chowdhury, A.A.; Hossain, M.S.; Rahman, M.S.; Nishimura, K.; Jisaka, M.; Nagaya, T.; Shono, F.; Yokota, K. Sustained expression of lipocalin-type prostaglandin D synthase in the antisense direction positively regulates adipogenesis in cloned cultured preadipocytes. *Biochem. Biophys. Res. Commun.* 2011, 411, 287–292. [CrossRef] [PubMed]
- 166. Virtue, S.; Masoodi, M.; Velagapudi, V.; Tan, C.Y.; Dale, M.; Suorti, T.; Slawik, M.; Blount, M.; Burling, K.; Campbell, M.; et al. Lipocalin prostaglandin D synthase and PPARγ2 coordinate to regulate carbohydrate and lipid metabolism in vivo. *PLoS ONE* **2012**, *7*, e39512. [CrossRef] [PubMed]

- 167. Emdin, M.; Fatini, C.; Mirizzi, G.; Poletti, R.; Borrelli, C.; Prontera, C.; Latini, R.; Passino, C.; Clerico, A.; Vergaro, G. Biomarkers of activation of renin-angiotensin-aldosterone system in heart failure: How useful, how feasible? *Clin. Chim. Acta* 2015, 443, 85–93. [CrossRef] [PubMed]
- 168. De Buyzere, M.; Gruson, D. Biomarkers in heart failure. Clin. Chim. Acta 2015, 443, 1–2. [CrossRef] [PubMed]



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).