





Complete Genome Sequences of Three *Listeria monocytogenes* Strains from Microgreens Obtained with MinION and MiSeq Sequencing

Sohail Naushad, Damit Mathews, Marc-Olivier Duceppe, Mingsong Kang, Lin Ru Wang, Mongsheng Huang

^aOttawa Laboratory–Fallowfield, Canadian Food Inspection Agency, Ottawa, Ontario, Canada ^bGreater Toronto Area laboratory, Canadian Food Inspection Agency, Toronto, Ontario, Canada

ABSTRACT *Listeria monocytogenes* is a Gram-positive, rod-shaped, non-spore-forming bacterium which is an important foodborne bacterial pathogen for human worldwide with 20-30% mortality. Here, we report circular complete genome sequences of three *Listeria monocytogenes* strains isolated from the samples of microgreens in Canada.

isteria monocytogenes is a Gram-positive bacterium found in various environments, animals, and food and is an important foodborne pathogen for humans worldwide with 20 to 30% mortality (1). We report complete genome sequences of *L. monocytogenes* GTA-L407, GTA-L409, and GTA-L411 strains (Table 1), isolated from three different samples of microgreens taken in Canada in 2018. The strains were isolated by enrichment in UVM broth (30°C, 24 h), followed by isolation on Oxford agar plate (35°C, 48 h) and RAPID'*L.mono* agar plates (35°C for 24 h), followed by the confirmation with Haemolysis on Blood agar plates, motility testing on Trypticase soy agar with yeast extract (30°C, 24 h), and rapid identification with Vitek (bioMérieux, Canada) (2).

Genomic DNA (gDNA) was extracted from overnight culture (grown from a single colony) in Brain and Heart Infusion medium using Maxwell 16 Cell DNA purification kit (Promega, US) for Illumina sequencing or NanoBind CBB Big DNA kit (Circulomics, US) for Nanopore sequencing. The gDNA was quantified using Qubit (ThermoFisher Scientific, US). Paired-end Illumina library was prepared using Nextera XT Library Preparation kit (Illumina, US) and sequenced for $(2 \times 300 \text{ bp})$ cycles on Illumina MiSeq. Nanopore sequencing library was generated using the 1D Native barcoding gDNA protocol (EXP-NBD104 and SQK-LSK109) (Oxford Nanopore Technologies, UK), and sequenced using a FLO-MIN106 (R9.4.1) flow cell in MinION MK1C. Basecalling was performed using Super Accuracy mode in Guppy v5.0.11, trimming using Porechop v0.2.3 (3) and filtration using Filtlong v0.2.1 (4). Long reads assembly was performed using Flye v2.7 (5), corrected using Medaka v1.4.4 and polished with Illumina MiSeq reads using a combination of NextPolish v1.4.0, ntEdit v1.3.5 and Polypolish v0.5.0 after trimming/ filtering with fastp v0.23.2. Circularity and genome rotation using dnaA as the starting point was determined using fixstart plugin from Circlator v1.5.5 (6). Sequencing coverage depth was determined using Minimap2 v2.17 (7) and Samtools v1.13 (8) for long reads and BWA v0.7.17 and Samtools for short reads. Gene prediction and annotation were performed using NCBI Prokaryotic Genome Annotation Pipeline (PGAP)-v6.0 (9).

Antimicrobial resistance (AMR) genes were identified using ResFinder v4.1.5 (10) and RGI v5.2.0 (11). The plasmids were identified by mlplasmids v1.0.0 (12). Prophage sequences were analyzed using PHASTER (13). Default parameters of pipelines were used except otherwise noted (https://github.com/OLF-Bioinformatics/nanopore).

Each strain contains a single chromosome of an average 2,878,201 bp, had $299\times$ (nanopore) and $134\times$ (MiSeq) coverage on average (Table 1), 67 tRNAs and no plasmids and no intact prophage sequences (>90 PHASTER-Score). Each strain contains only *fosX* AMR gene in both ResFinder and RGI analyses. The annotated genome on average had 2,766 coding

Editor David A. Baltrus, University of Arizona
© Crown copyright 2022. This is an openaccess article distributed under the terms of
the Creative Commons Attribution 4.0
International license.

Address correspondence to Sohail Naushad, sohail.naushad@inspection.gc.ca, or Hongsheng Huang, hongsheng.huang@inspection.gc.ca.

hongsheng.huang@inspection.gc.ca.
The authors declare no conflict of interest.

Received 17 March 2022 Accepted 16 May 2022 Published 6 June 2022

July 2022 Volume 11 Issue 7 10.1128/mra.00277-22

TABLE 1 Genomic characteristics of the complete genome sequences of 3 Listeria monocytogenes isolated from microgreens

		No. of Reads	spe	(hn)	Coverage (x)	(x)					
Microgreens				Nanopore			Chromosome	Predicted GC	gC	GenBank	SRA
type	Strain	MinION Miseq	Miseq	reads	MinION Miseq	Mised	size (bp)	CDSs	(%)	no.	nanopore/miseq
Microgreens-mix	Alicrogreens-mix GTA-L407 (also referred as CFIAFB20180112)	135,617	696'288	30,366	398	133	2,865,342	2765	38.06	CP092058	SRR17965221/
											SRR17965215
Sunflower	GTA-L409 (also referred as CFIAFB20180114)	59,643	770,315 37,519	37,519	291	121	2,879,234	2761	38.01	CP092057	SRR17965220/
											SRR17965224
Spring pea	GTA-L411 (also referred as CFIAFB20180116)	41,880	972,281 37,256	37,256	209	148	2,890,028	2774	38.01	CP092056	SRR17965219/
											SRR17965223

July 2022 Volume 11 Issue 7 10.1128/mra.00277-22 2

sequences (CDSs) and 38.08 GC%, similar to 2,889 CDS and 37.88 GC% for *L. monocytogenes* on average in NCBI (accessed on 2022-03-07).

Data availability. The assembled closed genome sequences of the three isolates and their Base-called MinION and MiSeq sequenced reads in fastq format have been deposited in GenBank under the BioProject number PRJNA803486. The accession numbers are provided in Table 1.

ACKNOWLEDGMENTS

This work was funded by the Canadian Food Inspection Agency. We thank the Microbiology Laboratory of CFIA-GTA Lab for technical support received for bacterial isolation.

REFERENCES

- Radoshevich L, Cossart P. 2018. Listeria monocytogenes: towards a complete picture of its physiology and pathogenesis. Nat Rev Microbiol 16: 32–46. https://doi.org/10.1038/nrmicro.2017.126.
- Pagotto F, Hebert K, Farber J. 2011. Isolation of Listeria monocytogenes and other Listeria spp. from foods and environmental sample

 — MFHPB-30: the compendium of analytical methods (Vol 2), HPB methods for the microbiological analysis of foods, Health Products and Food Branch, Health Canada.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. Microb Genom 3:e000132. https://doi.org/10.1099/mgen.0.000132.
- Kang M, Chmara J, Duceppe MO, Phipps-Todd B, Huang H. 2020. Complete genome sequence of a Canadian *Klebsiella michiganensis* strain, obtained using Oxford Nanopore Technologies sequencing. Microbiol Resour Announc 9. https://doi.org/10.1128/MRA.00960-20.
- Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, errorprone reads using repeat graphs. Nat Biotechnol 37:540–546. https://doi .org/10.1038/s41587-019-0072-8.
- Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularisation of genome assemblies using long sequencing reads. Genome Biol 16:294. https://doi.org/10.1186/s13059-015-0849-0.
- Li H. 2018. Minimap2: pairwise alignment for nucleotide sequences. Bioinformatics 34:3094–3100. https://doi.org/10.1093/bioinformatics/bty191.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis 0.011w?>G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The sequence alignment/map format and SAMtools. Bioinformatics 25: 2078–2079. https://doi.org/10.1093/bioinformatics/btp352.

- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/ nar/qkw569.
- Bortolaia V, Kaas RS, Ruppe E, Roberts MC, Schwarz S, Cattoir V, Philippon A, Allesoe RL, Rebelo AR, Florensa AF, Fagelhauer L, Chakraborty T, Neumann B, Werner G, Bender JK, Stingl K, Nguyen M, Coppens J, Xavier BB, Malhotra-Kumar S, Westh H, Pinholt M, Anjum MF, Duggett NA, Kempf I, Nykäsenoja S, Olkkola S, Wieczorek K, Amaro A, Clemente L, Mossong J, Losch S, Ragimbeau C, Lund O, Aarestrup FM. 2020. ResFinder 4.0 for predictions of phenotypes from genotypes. J Antimicrob Chemother 75:3491–3500. https://doi.org/10.1093/jac/dkaa345.
- 11. Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Bouchard M, Edalatmand A, Huynh W, Nguyen AV, Cheng AA, Liu S, Min SY, Miroshnichenko A, Tran HK, Werfalli RE, Nasir JA, Oloni M, Speicher DJ, Florescu A, Singh B, Faltyn M, Hernandez-Koutoucheva A, Sharma AN, Bordeleau E, Pawlowski AC, Zubyk HL, Dooley D, Griffiths E, Maguire F, Winsor GL, Beiko RG, Brinkman FSL, Hsiao WWL, Domselaar GV, McArthur AG. 2020. CARD 2020: antibiotic resistome surveillance with the comprehensive antibiotic resistance database. Nucleic Acids Res 48:D517–D525. https://doi.org/10.1093/nar/gkz935.
- Arredondo-Alonso S, Rogers MRC, Braat JC, Verschuuren TD, Top J, Corander J, Willems RJL, Schurch AC. 2018. mlplasmids: a user-friendly tool to predict plasmid- and chromosome-derived sequences for single species. Microb Genom 4.
- Arndt D, Grant JR, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better, faster version of the PHAST phage search tool. Nucleic Acids Res 44: W16–W21. https://doi.org/10.1093/nar/gkw387.

July 2022 Volume 11 Issue 7 10.1128/mra.00277-22