



## Complete Genome Sequence of *Aggregatibacter actinomycetemcomitans* Strain IDH781

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We report here the complete genomic sequence and methylome of *Aggregatibacter actinomycetemcomitans* strain IDH781. This rough strain is used extensively as a model organism to characterize localized aggressive periodontitis pathogenesis, the basic biology and oral cavity colonization of *A. actinomycetemcomitans*, and its interactions with other members of the oral microbiome.

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A ggregatibacter actinomycetemcomitans is a Gram-negative nonmotile facultative anaerobe of the oral microbiota implicated in the development of localized aggressive periodontitis (LAP) (1, 2). Despite the lack of a complete genomic sequence, IDH781, a rough strain, is commonly used as a model for pathogenesis and basic bacteriology studies (3–11). Rough strains of *A. actinomycetemcomitans* display the classic star-shaped colony morphology observed in clinical isolates from LAP patients, so they are more appropriate for the study of *A. actinomycetemcomitans* biology than those displaying a smooth colony phenotype (12, 13).

IDH781 was grown under anaerobic conditions (10% hydrogen, 10% carbon dioxide, and 80% nitrogen) on brain heart infusion agar supplemented with glucose, sodium bicarbonate, and yeast extract. The rough phenotype was confirmed microscopically. Cells were scraped from agar plates and lysed in 625  $\mu$ g/ml proteinase K, 1.25 mg/ml lysozyme, and 2% sodium dodecyl sulfate. Genomic DNA was purified by phenol-chloroform/isoamyl extraction (14). DNA integrity was verified by agarose gel electrophoresis, purity was evaluated spectrophotometrically, and concentration was determined fluorometrically.

Sequencing libraries were constructed using the Pacific Biosciences 20-kb template preparation protocol and Sage Science's BluePippin size-selection system with a 5-kb fragment size cutoff. Pacific Biosciences single-molecule real-time (SMRT) sequencing was performed on an RSII instrument using a single SMRT cell with P6-C4 chemistry and a 3-h movie, producing 7,601 polymerase reads ( $N_{50}$ , 11,624 nucleotides [nt]) and 153,117 postfiltered subreads ( $N_{50}$ , 6,741 nt). *De novo* assembly with the HGAP assembler (version 2.3) yielded a single contig supported by a mean coverage of 283-fold (15).

The genome was circularized by permutation to start at the *dnaA* gene and remove terminal duplications using Circlator

(version 1.0.2), followed by resequencing using RS\_Modification\_and\_Motif\_Analysis (version 2.3) to correct errors at the original contig break and to detect DNA methylation motifs based on significant interpulse duration signals (15, 16). Two well-supported m6A motifs were detected, GATC and AGGAG (bold indicates the methylated residue), with >98% of motifs in the genome detected. Seven other unique nonpalindromic modifications were detected at low frequency; these consisted of 2 putative m6A motifs (34% and 30%), 1 putative m4C motif (11%), and 4 unknown base modifications (20%, 15%, 8%, and 4%). The significance and importance of these low-frequency interpulse duration signals remain unknown.

The final assembly was 2,291,252 bp, with a G+C content of 44.3%, consistent with other completed *A. actinomycetemcomitans* genomes. To verify strain identity and detect possible horizontal gene transfer events, genomic intervals were taxonomically assigned using Taxator-tk (version 1.3.1e) with the nonredundant-microbial\_20140513 database (refpack from http://research.bifo.helmholtz-hzi.de/software) (17). All classified regions (39.8% of the genome) were assigned to the species *A. actinomycetemcomitans*.

Annotation was performed by NCBI using the Prokaryotic Genome Automated Annotation Pipeline (PGAAP, bestplaced reference protein; GeneMarkS+; version 3.3) (18). The chromosome contains 2,206 genes, with 2,129 coding sequences, 19 rRNAs, and 54 tRNAs for all 20 amino acids plus selenocysteine, 4 noncoding RNAs (ncRNAs), and 3 predicted clustered regularly interspaced short palindromic repeats (CRISPRs).

Accession number(s). The complete genome sequence of *A. actinomycetemcomitans* strain IDH781 has been deposited in GenBank under the accession number CP016553. The version described here is the first version.

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## REFERENCES

- Slots J, Ting M. 1999. Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis in human periodontal disease: occurrence and treatment. Periodontol 2000 20:82–121. http://dx.doi.org/10.1111/j.1600 -0757.1999.tb00159.x.
- Slots J, Reynolds HS, Genco RJ. 1980. Actinobacillus actinomycetemcomitans in human periodontal disease: a cross-sectional microbiological investigation. Infect Immun 29:1013–1020.
- Shanmugam M, Gopal P, El Abbar F, Schreiner HC, Kaplan JB, Fine DH, Ramasubbu N. 2015. Role of exopolysaccharide in *Aggregatibacter actinomycetemcomitans*-induced Bone resorption in a Rat model for periodontal disease. PLoS One 10:e0117487. http://dx.doi.org/10.1371/ journal.pone.0117487.
- Haubek D, Poulsen K, Asikainen S, Kilian M. 1995. Evidence for absence in northern Europe of especially virulent clonal types of *Actinobacillus actinomycetemcomitans*. J Clin Microbiol 33:395–401.
- Fine DH, Furgang D, Schreiner HC, Goncharoff P, Charlesworth J, Ghazwan G, Fitzgerald-Bocarsly P, Figurski DH. 1999. Phenotypic variation in Actinobacillus actinomycetemcomitans during laboratory growth: implications for virulence. Microbiology 145:1335–1347. http:// dx.doi.org/10.1099/13500872-145-6-1335.
- Fine DH, Karched M, Furgang D, Sampathkumar V, Velusamy S, Godboley D. 2015. Colonization and persistence of labeled and "foreign" strains of Aggregatibacter actinomycetemcomitans inoculated into the mouths of rhesus monkeys. J Oral Biol (Northborough) 2:10. http:// dx.doi.org/10.13188/2377-987X.1000005.
- Takashima E, Konishi K. 2008. Characterization of a quinol peroxidase mutant in *Aggregatibacter actinomycetemcomitans*. FEMS Microbiol Lett 286:66–70. http://dx.doi.org/10.1111/j.1574-6968.2008.01253.x.

- Yue G, Kaplan JB, Furgang D, Mansfield KG, Fine DH. 2007. A second Aggregatibacter actinomycetemcomitans autotransporter adhesin exhibits specificity for buccal epithelial cells in humans and Old World primates. Infect Immun 75:4440–4448. http://dx.doi.org/10.1128/IAI.02020-06.
- Fine DH, Goncharoff P, Schreiner H, Chang KM, Furgang D, Figurski D. 2001. Colonization and persistence of rough and smooth colony variants of *Actinobacillus actinomycetemcomitans* in the mouths of rats. Arch Oral Biol 46:1065–1078. http://dx.doi.org/10.1016/S0003 -9969(01)00067-X.
- Fine DH, Kaplan JB, Furgang D, Karched M, Velliyagounder K, Yue G. 2010. Mapping the epithelial-cell-binding domain of the *Aggregatibacter actinomycetemcomitans* autotransporter adhesin Aae. Microbiology 156: 3412–3420. http://dx.doi.org/10.1099/mic.0.037606-0.
- Kaplan JB, Velliyagounder K, Ragunath C, Rohde H, Mack D, Knobloch JK, Ramasubbu N. 2004. Genes involved in the synthesis and degradation of matrix polysaccharide in *Actinobacillus actinomycetemcomitans* and *Actinobacillus pleuropneumoniae* biofilms. J Bacteriol 186: 8213–8220. http://dx.doi.org/10.1128/JB.186.24.8213-8220.2004.
- Preus HR, Sunday GJ, Haraszthy VI, Zambon JJ. 1992. Rapid identification of Actinobacillus actinomycetemcomitans based on analysis of 23S ribosomal RNA. Oral Microbiol Immunol 7:372–375. http://dx.doi.org/ 10.1111/j.1399-302X.1992.tb00639.x.
- Slots J. 1982. Selective medium for isolation of *Actinobacillus actinomyce-temcomitans*. J Clin Microbiol 15:606–609.
- 14. Sambrook J, Russell DW. 2006. Purification of nucleic acids by extraction with phenol:chloroform. Cold Spring Harb Protoc 2006:p=pdb.prot4455. http://dx.doi.org/10.1101/pdb.prot4455.
- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. http://dx.doi.org/ 10.1038/nmeth.2474.
- Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. Genome Biol 16:294. http://dx.doi.org/10.1186/s13059 -015-0849-0.
- Dröge J, Gregor I, McHardy AC. 2015. Taxator-tk: precise taxonomic assignment of metagenomes by fast approximation of evolutionary neighborhoods. Bioinformatics 31:817–824. http://dx.doi.org/10.1093/ bioinformatics/btu745.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Acids Res 44: 6614–6624. http://dx.doi.org/10.1093/nar/gkw569.