GENOME SEQUENCES





Complete Genome Sequence of *Escherichia coli* MT102, a Plasmid-Free Recipient Resistant to Rifampin, Azide, and Streptomycin, Used in Conjugation Experiments

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ABSTRACT We present here the complete genome sequence of *Escherichia coli* MT102, which is resistant to rifampin, azide, and streptomycin and is used as a recipient in plasmid transfer experiments. The sequence will be utilized for chromosomal read removal in plasmid sequence analyses obtained from transconjugants within this strain and in comprehensive genetic studies.

scherichia coli MT102 is an araD139 (ara-leu)7697 ∆lac thi hsdR derivate of E. coli K-12 substrain MC1000 (1, 2) and was constructed by Mogens Trier Hansen at Novo-Nordisk. E. coli MT102 is resistant to sodium azide, rifampin, and streptomycin due to missense mutations in the secA (3), rpoB (4), and rpsL (5) genes, respectively. This strain has been used in several studies characterizing mobile genetic elements carrying antimicrobial genes (6–8) and studies requiring a plasmid-free host (9, 10). E. coli MT102 was grown on MacConkey agar plates supplemented with rifampin (25 mg/liter) and sodium azide (25 mg/liter) overnight at 37°C. Bacterial genomic DNA for short-read Illumina sequencing was extracted using the DNeasy blood and tissue kit (catalog number 69506; Qiagen, Hilden, Germany), and a sequencing library was prepared using the Nextera XT kit (catalog number FC-131-1096; Illumina, San Diego, CA). Short reads were obtained by 2 \times 250-bp paired-end MiSeq sequencing (Illumina), yielding 3,312,280 reads. Reads were trimmed using Trimmomatic v0.36 (illuminaclip:TruSeq3-PE.fa:2:30:10 leading:3 trailing:3 slidingwindow:4:15 minlen:36) (11) in order to remove adaptor residues and low-quality ($Q \le 20$) ends. For the long reads, DNA was prepared using the Agencourt Genfind v2 kit (Beckman Coulter, Brea, CA) with a DynaMag-2 magnet (Thermo Fisher, Waltham, MA, USA). Libraries were prepared with the 1D ligation barcoding kit (catalog numbers SQK-LSK108 and EXP-NBD103; ONT, Oxford, United Kingdom) and sequenced in an R9.4 flow cell (catalog number FLO-MIN106) with a MinION Mk1B device (ONT), yielding 16,561 reads consisting of 138,366,031 bp in total, with an N_{so} read length of 14,053 bp. The fast5 reads were base called, demultiplexed, and converted to fastq format using Albacore v2.0.2 (ONT) with default settings. The adaptor sequences were removed using Porechop v0.2.2 (12) with default settings. Hybrid assembly of long and short reads was performed using Unicycler v0.4.0 (13), with default settings, and resulted in one circular contig of the bacterial chromosome. The length of the contig was 4,548,459 bp. The coverages of Illumina reads were $174 \times$ and $25 \times$ of the long reads from MinION. The short Illumina reads were mapped to the chromosomal contig using the "Map to reference" option in Geneious 9.0.5 (Biomatters Ltd., Auckland, New Zealand) and called variants using "Find variations/ SNPs..." based on the mapping for manual error correction, both with default settings. Moreover, MinION reads longer than 10 kb were mapped to the chromosomal contig to

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Received 2 April 2019 Accepted 28 April 2019 Published 16 May 2019 double-check the consensus sequence. Due to the manual error correction, the size of the final chromosomal sequence was increased when 489 bp was added.

The genome was annotated using NCBI Prokaryotic Genome Annotation Pipeline (14). The complete genome of *E. coli* MT102 consists of 4,548,948 bp, with a GC content of 50.7%, 4,288 coding sequences, 86 tRNAs, 22 rRNAs, and 15 noncoding RNAs (ncRNAs). The strain belongs to sequence type 10, which was identified using MLST-2.0 (15) by the Center for Genomic Epidemiology (CGE; DTU, Kongens Lyngby, Denmark).

Data availability. The complete genome sequence of *E. coli* MT102 and raw data have been deposited in GenBank under accession number CP034953 and SRA accession numbers SRX5367507 and SRX5367508.

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REFERENCES

- Cohen N, April R. 1980. Analysis of gene control signals by DNA fusion and cloning in *Escherichia coli*. J Mol Biol 138:179–207. https://doi.org/ 10.1016/0022-2836(80)90283-1.
- Steidle A, Allesen-Holm M, Riedel K, Berg G, Givskov M, Molin S, Eberl L. 2002. Identification and characterization of an *N*-acylhomoserine lactone-dependent quorum-sensing system in *Pseudomonas putida* strain IsoF. Appl Environ Microbiol 68:6371–6382. https://doi.org/10 .1128/AEM.68.12.6371-6382.2002.
- Oliver DB, Cabelli RJ, Dolan KM, Jarosik GP. 1990. Azide-resistant mutants of *Escherichia coli* alter the SecA protein, an azide-sensitive component of the protein export machinery. Proc Natl Acad Sci U S A 87:8227–8231. https://doi.org/10.1073/pnas.87.21.8227.
- Jin DJ, Gross CA. 1988. Mapping and sequencing of mutations in the Escherichia coli rpoB gene that lead to rifampicin resistance. J Mol Biol 202:45–58. https://doi.org/10.1016/0022-2836(88)90517-7.
- Timms AR, Steingrimsdottir H, Lehmann AR, Bridges BA. 1992. Mutant sequences in the *rpsL* gene of *Escherichia coli* B/r: mechanistic implications for spontaneous and ultraviolet light mutagenesis. Mol Gen Genet 232:89–96. https://doi.org/10.1007/BF00299141.
- Kutilova I, Janecko N, Cejkova D, Literak I, Papagiannitsis CC, Dolejska M. 2018. Characterization of *bla*_{KPC-3}-positive plasmids from an *Enterobacter aerogenes* isolated from a corvid in Canada. J Antimicrob Chemother 73:2573–2575. https://doi.org/10.1093/jac/dky199.
- Jamborova I, Dolejska M, Zurek L, Townsend AK, Clark AB, Ellis JC, Papousek I, Cizek A, Literak I. 2017. Plasmid-mediated resistance to cephalosporins and quinolones in *Escherichia coli* from American crows in the USA. Environ Microbiol 19:2025–2036. https://doi.org/10.1111/ 1462-2920.13722.
- 8. Papagiannitsis CC, Kutilova I, Medvecky M, Hrabak J, Dolejska M. 2017.

Characterization of the complete nucleotide sequences of IncA/ C2plasmids carrying In809-like integrons from Enterobacteriaceae isolates of wildlife origin. Antimicrob Agents Chemother 61:e01093-17. https://doi.org/10.1128/AAC.01093-17.

- Manefield M, de Nys R, Naresh K, Roger R, Givskov M, Peter S, Kjelleberg S. 1999. Evidence that halogenated furanones from Delisea pulchra inhibit acylated homoserine lactone (AHL)-mediated gene expression by displacing the AHL signal from its receptor protein. Microbiology 145: 283–291. https://doi.org/10.1099/13500872-145-2-283.
- Hansen LH, Sørensen SJ. 2000. Versatile biosensor vectors for detection and quantification of mercury. FEMS Microbiol Lett 193:123–127. https:// doi.org/10.1111/j.1574-6968.2000.tb09413.x.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. Microb Genom 3:e000132. https://doi.org/10.1099/mgen.0.000132.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Tatusova T, Dicuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614–6624. https://doi.org/10.1093/nar/gkw569.
- Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Ponten TS, Ussery DW, Aarestrup FM, Lund O. 2012. Multilocus sequence typing of total-genome-sequenced bacteria. J Clin Microbiol 50:1355–1361. https://doi.org/10.1128/JCM.06094-11.