MITOGENOME ANNOUNCEMENT

Taylor & Francis Group

Taylor & Francis

OPEN ACCESS

The complete chloroplast genome sequence of *Spyridium parvifolium* var. *parvifolium* (family Rhamnaceae; tribe Pomaderreae)

Catherine Clowes^a (D), Rachael M. Fowler^a, Gillian K. Brown^b and Michael J. Bayly^a (D)

^aSchool of Biosciences, The University of Melbourne, Parkville, Australia; ^bDepartment of Environment and Science, Queensland Herbarium, Toowong, Australia

ABSTRACT

We assembled the complete chloroplast genome of the Australian shrub *Spyridium parvifolium* var. *parvifolium*. The genome was 161,012 bp in length, with a pair of inverted repeats (IRs) of 26,515 bp, separated by a large single copy (LSC) region of 88,814 bp and a small single copy region (SCC) of 19,168 bp. The GC content was 36.9%. In total, 130 genes were annotated, including 86 protein coding genes, 36 tRNA genes and 8 rRNA genes. Phylogenetic analysis of 56 chloroplast genes placed this genome of *S. parvifolium* var. *parvifolium* within the family Rhamnaceae.

ARTICLE HISTORY

Received 24 May 2018 Accepted 29 May 2018

KEYWORDS

Spyridium parvifolium; Rosales; Rhamnaceae; Ziziphus jujuba; chloroplast genome

Spyridium Fenzl is a member of the cosmopolitan Rhamnaceae family (VicFlora 2016), which includes the economically important Chinese Date, Ziziphus jujuba Mill. (Liu et al. 2014). Spyridium parvifolium (tribe Pomaderreae) is a shrubby, widespread, and morphologically variable species from south-eastern Australia (Jessop et al. 1986; Curtis and Morris 1993; VicFlora 2016; PlantNET n.d.). Several varieties of the species are sometimes recognized, and additional morphological variants have been identified (VicFlora 2016). Conflicting infraspecific taxonomies in different parts of Australia have implications for conservation management. In particular, two varieties (var. parvifolium and var. mole) are recognized in Tasmania, where both are listed as 'Threatened' under state legislation (Threatened Species Section 2016a, 2016b); in contrast, none of the varieties is currently recognized as distinct by the Australian Plant Census (CHAH 2016).

In this study, we report the complete chloroplast genome sequence of *S. parvifolium* var. *parvifolium* (GenBank accession MH234313). We generated this sequence to use as a reference in further chloroplast genome studies aimed at assessing phylogeography, genetic diversity, introgression, and infraspecific taxonomy of *S. parvifolium*.

Plant material was sampled from a population of var. *par-vifolium* at Sisters Beach, Tasmania, Australia (40°54′15.0′S 145°32′47.5′E; Permit Number: TFL 15171; Voucher Specimen: MELUD155066a). Total DNA was extracted from leaves dried *in silica* gel using a modified CTAB protocol (Shepherd and McLay 2011), prepared for sequencing using the protocol of Schuster et al. (2018), and sequenced on an Illumina NextSeq

550 (mid-output, 2×150 Paired End kit) at The Walter and Eliza Hall Institute of Medical Research (WEHI). The genome was assembled by mapping paired reads to the reference genome of *Ziziphus jujuba* (accession number KU351660). Contigs built in Spades 3.10.0 (Bankevich et al. 2012), CLC Genomics Workbench 10.0.1 and Geneious 10.2 (Kearse et al. 2012) were mapped to the consensus sequence for quality control. Annotations were transferred from the reference sequence, with reading frames reviewed and manually adjusted.

The complete chloroplast of *S. parvifolium* var. *parvifolium* was 161,012 bp in length. A pair of inverted repeats (IRs) of 26,515 bp were separated by a large single copy (LSC) region of 88,814 bp and a small single copy region (SCC) of 19,168 bp. The GC content of the chloroplast genome was 36.9%. In total 130 genes were annotated, including 86 protein coding genes, 36 tRNA genes, and 8 rRNA genes. One pseudogene was predicted (*InfA*) and two truncated repeats were recorded at IR boundaries (*rps19* and *ycf1*). Annotations were identical between the reported genome (*S. parvifolium* var. *parvifolium*) and the reference (*Ziziphus jujuba*) except for one copy of the *ycf1* gene which was annotated on the reported genome as protein coding while neither copy of the gene was annotated as protein coding on the reference genome.

The phylogenetic tree presented in this study (Figure 1) builds from the results of Hauenschild et al. (2016) and Cheon et al. (2018). This tree shows *S. parvifolium* var. *parvifolium* within the Rhamnaceae clade and most closely related to *Ziziphus jujuba* (tribe Paliureae).

CONTACT Catherine Clowes Clowes@student.unimelb.edu.au School of Biosciences, The University of Melbourne, Grattan Street, Parkville, 3010, Australia © 2018 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

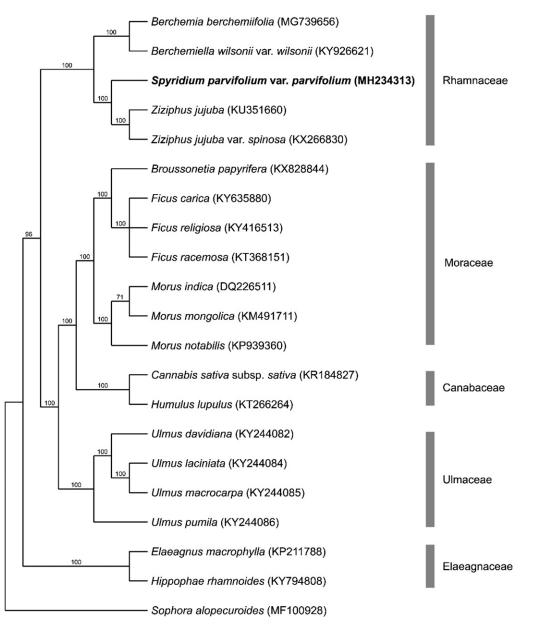


Figure 1. Bootstrap 50% majority rule consensus tree based on 56 protein coding chloroplast genes from 21 taxa including 20 species from the order Rosales and *Sophora alopecuroides* as the outgroup (CI = 0.8069 RI = 0.8744). Genes were aligned in MAFFT using default settings (Katoh et al. 2002). Sequences were analysed using maximum parsimony (MP) with PAUP 4.0a 161 using default settings (Swofford 2003). Bootstrap values are provided above branches. GenBank accessions are provided in brackets. *Spyridium parvifolium* var. *parvifolium* is highlighted in bold.

Acknowledgments

We acknowledge Mark Wapstra (ECOtas) for assisting with fieldwork. Jürgen Kellermann (State Herbarium of South Australia) for advice on *Spyridium*. Erin Batty and Todd McLay (The University of Melbourne) for support with DNA extractions. Stephen Wilcox (WEHI) for Illumina sequencing. Tanja Schuster (The University of Melbourne) for advice on preparation of this manuscript.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was supported by The University of Melbourne under an Australian Postgraduate Awards, The University of Melbourne Botany

Foundation under a Megan Klemm Postgraduate Research Award and a Sophie Ducker Postgraduate Scholarship, the Australasian Systematic Botany Society under a grant from the Hansjörg Eichler Scientific Research Fund, and the Ecological Society of Australia under a Holsworth Wildlife Research Endowment.

ORCID

Catherine Clowes D http://orcid.org/0000-0002-9466-753X Michael J. Bayly D http://orcid.org/0000-0001-6836-5493

References

Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, et al. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol. 19:455–477.

- CHAH. 2016. Autralian Plant Census (APC). [updated 2015 Jan 23; accessed 2016 Jul 25]. https://biodiversity.org.au/nsl/services/search?product=APC &tree.id=1133571&name=spyridium+parvifolium&inc._scientific=&inc. scientific=on&inc._cultivar=&max =100&display=apc&search=true.
- Cheon K-S, Kim K-A, Yoo K-O. 2018. The complete chloroplast genome sequence of Berchemia berchemiifolia (Rhamnaceae). Mitochondrial DNA Part B. 3:133–134.
- Curtis WM, Morris DI. 1993. The student's flora of Tasmania. Hobart: St. David's Park Publishing.
- Hauenschild F, Matuszak S, Muellner-Riehl AN, Favre A. 2016. Phylogenetic relationships within the cosmopolitan buckthorn family (Rhamnaceae) support the resurrection of Sarcomphalus and the description of Pseudoziziphus gen. nov. Taxon. 65:47–64.
- Katoh K, Misawa K, Kuma Ki, Miyata T. 2002. MAFFT: A novel method for rapid multiple sequence alignment based on fast Fourier transform. Nucleic Acids Res. 30:3059–3066.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, et al. 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics. 28:1647–1649.
- Jessop JP, Toelken HR, Black JM. 1986. Flora of South Australia. 4th ed. Adelaide: South Australian Government Printing Division.
- Liu M-J, Zhao J, Cai Q-L, Liu G-C, Wang J-R, Zhao Z-H, Liu P, Dai L, Yan G, Wang W-J, et al. 2014. The complex jujube genome provides insights into fruit tree biology. Nat Commun. 5:5315.

- PlantNET. n.d. The NSW Plant Information Network System. Royal Botanic Gardens and Domain Trust; [accessed 2018 Apr 30]. http://plantnet.rbgsyd.nsw.gov.au/cgi-bin/NSWfl.pl?page=nswfl&lvl=sp& name=Spyri-dium~parvifolium.
- Schuster TM, Setaro SD, Tibbits JFG, Batty EL, Fowler RM, McLay TGB, Wilcox S, Ades PK, Bayly MJ. 2018. Chloroplast variation is incongruent with classification of the Australian bloodwood eucalypts (genus Corymbia, family Myrtaceae). PloS One. 13:e0195034.

Shepherd LD, McLay TG. 2011. Two micro-scale protocols for the isolation of DNA from polysaccharide-rich plant tissue. J Plant Res. 124:311–314.

- Swofford DL. 2003. PAUP*:Phylogenetic analysis using parsimony, version 4.0a 161.
- Threatened Species Section. 2016a. Spyridium parvifolium var. molle (soft dustymiller): Species Management Profile for Tasmania's Threatened Species Link. Department of Primary Industries, Parks, Water and Environment, Tasmania; [accessed 2018 Apr 24]. Sequences were analysed using maximum parsimony (MP) with PAUP 4.0a 161 using default settings (Swofford 2003). http://www.threatenedspecieslink.tas. gov.au/Pages/Spyridium-parvifolium-var-molle.aspx.
- Threatened Species Section. 2016b. *Spyridium parvifolium* var. *parvifolium* (coast dustymiller): Species Management Profile for Tasmania's Threatened Species Link. Department of Primary Industries, Parks, Water and Environment, Tasmania; [accessed 2018 Apr 24]. http://www.threatened specieslink.tas.gov.au/Pages/Spyridium-parvifolium-var-parvifolium.aspx.
- VicFlora. 2016. Flora of Victoria. Royal Botanic Gardens Victoria; [accessed 2018 Apr 23]. https://vicflora.rbg.vic.gov.au.