

# *Vibrio vulnificus* VvpE inhibits mucin 2 expression by hypermethylation via lipid raft-mediated ROS signaling in intestinal epithelial cells

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Mucin is an important physical barrier against enteric pathogens. VvpE is an elastase encoded by Gram-negative bacterium *Vibrio vulnificus*; however, the functional role of VvpE in intestinal mucin (Muc) production is yet to be elucidated. The recombinant protein (r) VvpE significantly reduced the level of Muc2 in human mucus-secreting HT29-MTX cells. The repression of Muc2 induced by rVvpE was highly susceptible to the knockdown of intelectin-1b (ITLN) and sequestration of cholesterol by methyl- $\beta$ -cyclodextrin. We found that rVvpE induces the recruitment of NADPH oxidase 2 and neutrophil cytosolic factor 1 into the membrane lipid rafts coupled with ITLN to facilitate the production of reactive oxygen species (ROS). The bacterial signaling of rVvpE through ROS production is uniquely mediated by the phosphorylation of ERK, which was downregulated by the silencing of the *PKC $\delta$* . Moreover, rVvpE induced region-specific methylation in the *Muc2* promoter to promote the transcriptional repression of *Muc2*. In two mouse models of *V. vulnificus* infection, the mutation of the *vvpE* gene from *V. vulnificus* exhibited an increased survival rate and maintained the level of Muc2 expression in intestine. These results demonstrate that VvpE inhibits Muc2 expression by hypermethylation via lipid raft-mediated ROS signaling in the intestinal epithelial cells.

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Gastrointestinal mucosal surfaces serve as the first line of defense against many bacterial and viral infections between the epithelium and the luminal content.<sup>1,2</sup> Damage and impairment of the mucus layer facilitate the invasion of pathogenic microorganisms and enhance epithelial cell–microbiota interactions to destabilize the homeostasis of the host immune responses.<sup>3</sup> Mucin (Muc) 2 is the major intestinal O-glycosylated protein produced by goblet cells, having an important role as a physiological barrier via mucus formation.<sup>3</sup> A lack of Muc2 in mice results in severe colitis propagated by an increased bacterial adhesion to the epithelium and intestinal permeability.<sup>4,5</sup> Several studies have been conducted to determine the factors that regulate *Muc2* gene expression, including growth factor,<sup>6</sup> transcription factors,<sup>7</sup> and the methylation status.<sup>8</sup> A recent report showed that bacterial pathogens affect a diverse set of epigenetic factors such as DNA methylation and histone modifications to regulate selective activation or silencing of specific host genes.<sup>9</sup> Specifically, it was proven that there is a close correlation between the silencing of *Muc2* gene expression and its promoter hypermethylation.<sup>10</sup> On the other hand, intelectin-1b (ITLN) is a lectin that recognizes sugar motifs specific to bacterial cells

walls.<sup>11</sup> ITLN is produced mainly by goblet cells in intestine.<sup>12</sup> ITLN is located in the brush border membrane, where it serves as a decoy pathogen receptor against microbial infection,<sup>13</sup> but the functional role of ITLN and its related signaling pathway on *Muc2* gene expression are yet to be elucidated.

*Vibrio (V.) vulnificus* is a Gram-negative food pathogen which causes septicemia, necrotizing wound infections, or gastroenteritis.<sup>14</sup> The majority of the virulence effect is known to derive from the secreted toxins encoded by cytolytic pore-forming hemolysin (*VvhA*)<sup>14</sup> and the multifunctional autoprocessing repeats-in-toxin (*MARTXVv*).<sup>14,15</sup> VvpE is a homolog of the hemagglutinin/protease, which is suspected to be a major elastase produced by *V. vulnificus*.<sup>16,17</sup> Previous work reported that VvpE, a 45-kDa protein, consists of a 35-kDa N-terminal domain for catalytic activity and a 10-kDa domain for its attachment to the substrate.<sup>18</sup> Although the functions of VvhA and MARTXVv and their roles in pathogenesis have been well studied<sup>14</sup> owing to their significant fulminating and destructive actions in human tissues, the VvpE appeared to be dispensable for the virulence effect of *V. vulnificus*.<sup>17</sup> However, it has been reported that the expression of *VvpE* is regulated by the quorum-sensing regulator in promoting the pathogenesis of

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**Abbreviations:** Cont, Control; EDTA, Ethylenediaminetetraacetic acid; EGTA, ethylene glycol tetraacetic acid; ERK, extracellular signal-regulated kinases; FBS, fetal bovine serum; HEPES, 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid; Ig, immunoglobulin; ITLN, intelectin-1b; JNK, c-Jun N-terminal kinases; MAPK, mitogen-activated protein kinases; M $\beta$ CD, methyl- $\beta$ -cyclodextrin; NAC, N-acetylcysteine; NCF1, neutrophil cytosolic factor 1; NOX2, NADPH oxidase 2; Pan-cad, pan-cadherin; PBS, phosphate-buffered saline; p38, p38 MAPK; PI, propidium iodide; PKC, protein kinase C; PVDF, polyvinylidene fluoride; RFI, relative fluorescence intensity; ROD, relative optical density; ROS, reactive oxygen species; rVvpE, recombinant protein VvpE; SDS-PAGE, sodium dodecyl sulfate-polyacrylamide gel electrophoresis; SE, standard errors; TBST, tris-buffer solution-tween 20; Veh, vehicle

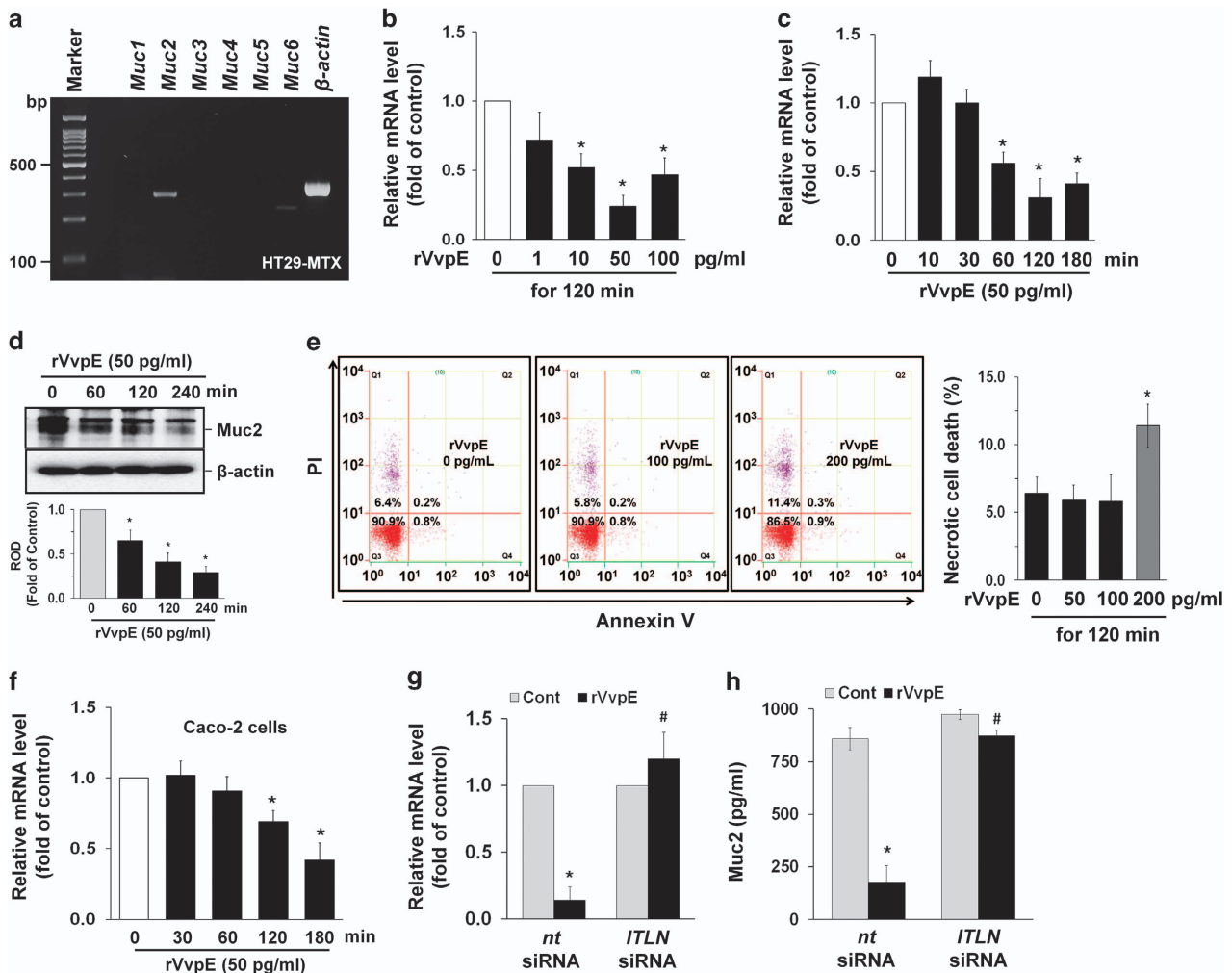
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*V. vulnificus*.<sup>19</sup> Thus, the functional role of VvpE remains a topic of much debate. In this study, therefore, we investigate the pathogenic mechanism of VvpE and its related signaling pathway in the regulation of the *Muc2* expression.

## Results

**Regulatory effect of VvpE on Muc2 expression.** To find the functional role of VvpE, we used mucus-secreting human HT29-MTX cells, which form a homogeneous population of polarized goblet cells. These types of cells have previously been widely used in the field of mucin-related research.<sup>20</sup> We first determined the existence of mucin isotypes in HT29-MTX cells. *Muc2* expression, followed by that of *Muc6*, is most abundant in these cells, whereas the expression levels

of *Muc1*, *Muc3*, *Muc4*, and *Muc5* were very low (Figure 1a). To evaluate the role of VvpE in the regulation of *Muc2* expression, HT29-MTX cells were exposed to various concentrations (0~100 pg/ml) of rVvpE for 120 min. rVvpE significantly reduced the expression of *Muc2* from 10 to 100 pg/ml compared with the cells with no treatment (Figure 1b). A decrease in *Muc2* mRNA expression and its protein level was observed after 60 min of incubation with 50 pg/ml of rVvpE (Figures 1c and d). Over 200 pg/ml of rVvpE induced a cytotoxic effect by promoting necrotic cell death; however, below 100 pg/ml of rVvpE had an inhibitory effect on *Muc2* expression without cytotoxicity (Figure 1e). In addition, rVvpE (50 pg/ml) also has the ability to inhibit *Muc2* expression in Caco-2 cells (Figure 1f). This result indicates that the functional role of rVvpE is reproducible in other type of human epithelial cells. Moreover, the level of *Muc2* mRNA



**Figure 1** The effect of rVvpE in Muc2 expression. (a) Expression of mucin (*Muc*) mRNAs in HT29-MTX cells is shown.  $n=4$ . Dose (b) and time (c) responses of rVvpE in *Muc2* expression are shown.  $n=5$ . \* $P<0.01$  versus the cells with no treatment. (d) The effect of rVvpE on protein expression of Muc2 was determined by western blot.  $n=4$ . \* $P<0.01$  versus the cells with no treatment. ROD, relative optical density. (e) Cells were incubated with 100 and 200 pg/ml of rVvpE for 120 min. Percentages of necrosis, survival, and apoptosis were measured by using PI/Annexin V staining and flow cytometry (left panel). Quantitative analysis of the percentage of necrotic cells (Q1) by FACS analysis is shown (right panel).  $n=5$ . \* $P<0.01$  versus the cells with no treatment. (f) The effect of rVvpE on expression of *Muc2* mRNA in Caco-2 cells was determined.  $n=3$ . \* $P<0.01$  versus the cells with no treatment. (g) HT29-MTX cells transfected with siRNAs for non-targeting (nt) control and *ITLN* were incubated with rVvpE for 120 min. The level of *Muc2* mRNA expression was measured by using real-time PCR.  $n=5$ . \* $P<0.01$  versus nt siRNA+Cont (boiled rVvpE, 50 pg/ml). # $P<0.05$  versus nt siRNA+rVvpE. (h) HT29-MTX cells transfected with *ITLN* siRNA were incubated with rVvpE for 4 h. The level of Muc2 protein was quantified by ELISA.  $n=5$ . \* $P<0.01$  versus nt siRNA+Cont (boiled rVvpE, 50 pg/ml). # $P<0.05$  versus nt siRNA+rVvpE

expression was not changed by treatment with trypsin at 5, 50, and 500 pg/ml for 180 min (Supplementary Figure S1), suggesting that the functional role of rVvpE in Muc2 repression is different from an outcome of general proteolysis of surface proteins. ITLN is thought to confer mucosal protection as a decoy pathogen receptor.<sup>11,13</sup> We further investigated the involvement of ITLN in the repression of Muc2 elicited by rVvpE (Figure 1g). Interestingly, the inhibitory effect of rVvpE on Muc2 expression was silenced by transfection with siRNA for ITLN. We also explored the ability of rVvpE to regulate Muc2 production in an enzyme-linked immunosorbent assay (ELISA) assay. In contrast to the control, 50 pg/ml of rVvpE evoked a substantial reduction of Muc2 secretion, whereas the repression activity of rVvpE on Muc2 was blocked by the silencing of ITLN (Figure 1h). These results suggest that rVvpE regulates the expression and secretion of Muc2 via ITLN.

**Involvement of a lipid raft and ROS production in Muc2 regulation.** It was previously demonstrated that ITLN is preferentially associated with microvillar lipid rafts.<sup>11,13</sup> We further determined the effect of rVvpE on the membrane location of ITLN by means of discontinuous sucrose density-gradient centrifugation. Although caveolin-1 was detected in fraction 5, flotillin-1 was observed in fractions 4–6 (Figure 2a, left panel), indicating that lipid rafts of HT29-MTX cells are presented in fractions 4–6. ITLN at basal condition was distributed in mainly fraction 4, which are correlated with membrane lipid rafts. Interestingly, 50 pg/ml of rVvpE altered the ITLN localization into the fraction 5, despite of the distribution of caveolin-1 and flotillin-1 was not changed (Figure 2a, right panel). Moreover, the subunits of NADPH oxidase (NOX) enzymes, NOX2 (gp91<sup>phox</sup>) and NCF1 (p47<sup>phox</sup>), were highly enriched in the fraction 6. However, rVvpE treatment resulted in translocations of NOX2 and NCF1 into fraction 5 enriched with lipid rafts proteins. Since this approach is qualitative at best, we further tried to quantify the results by using co-immunoprecipitation of ITLN with proteins related to the lipid rafts in the presence of rVvpE. It was noted that ITLN co-immunoprecipitated with caveolin-1, NOX2, and NCF1, and importantly, that these interactions were enhanced by the rVvpE treatment (Figure 2b). These results indicate that rVvpE has ability to cluster ITLN and NOX enzymes without altering their expressions in lipid raft. To confirm the structural importance of membrane rafts in the rVvpE-mediated signaling pathway, we employed the lipid raft sequester methyl- $\beta$ -cyclodextrin (M $\beta$ CD), which is known to deplete cholesterol from the cell membrane. Interestingly, the repression of Muc2 induced by rVvpE was significantly blocked by a pretreatment with M $\beta$ CD (Figure 2c). Since the tyrosine kinases are closely associated with the lipid raft, we further have confirmed whether rVvpE regulates activation of c-Src and FAK, as representative tyrosine kinases. rVvpE markedly increased the phosphorylation of c-Src and FAK from 10 min (Supplementary Figure S2a). Interestingly, pre-treatment with M $\beta$ CD blocked the c-Src and FAK activation induced by rVvpE (Supplementary Figure S2b). These mean that rVvpE regulates the lipid raft-mediated signaling pathway and thereby modifies the phosphoryrosination of c-Src and FAK and the localization of NOX. Clustering of NOX enzymes facilitates the

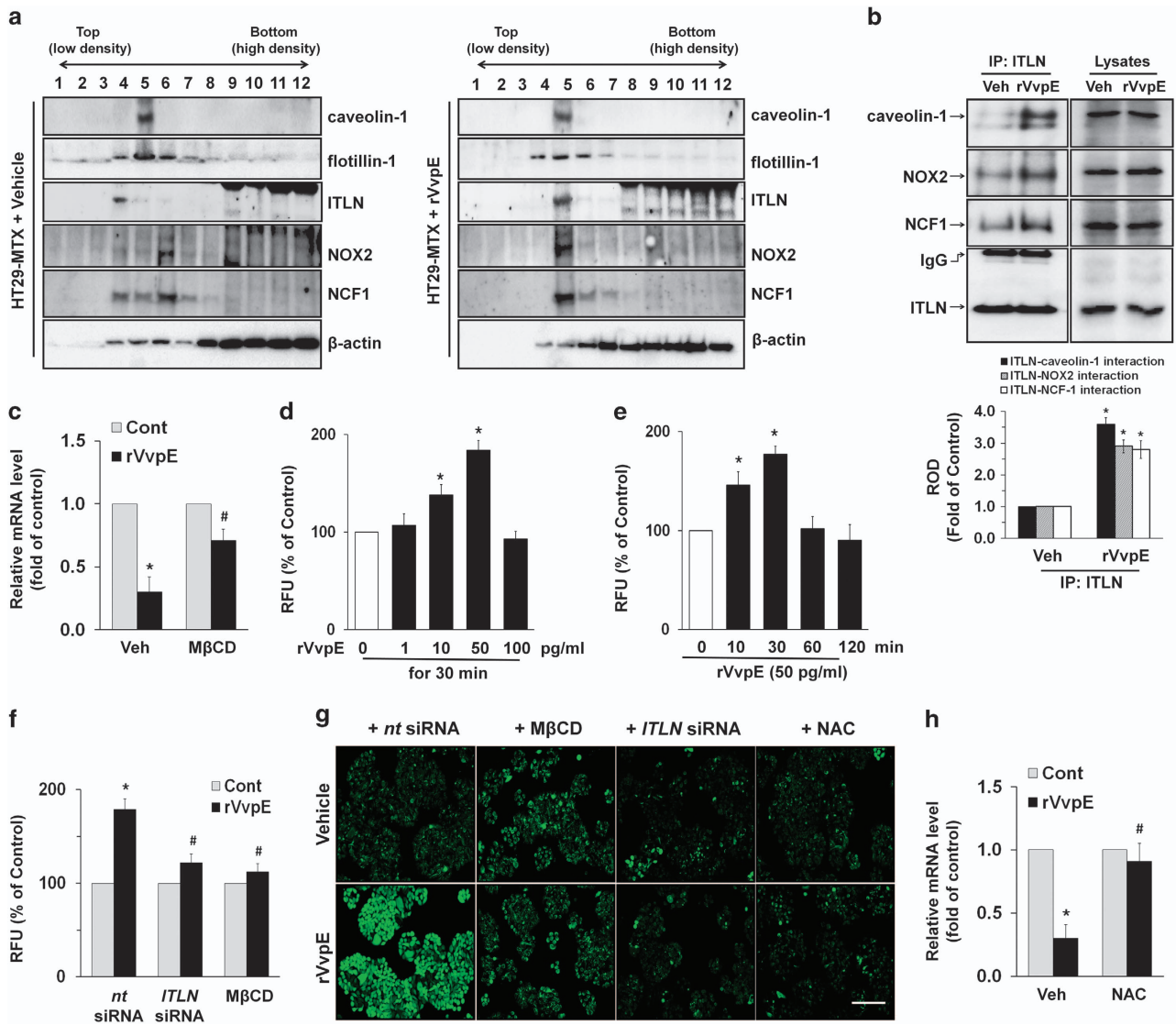
production of reactive oxygen species (ROS), which results in a prominent amplification of the transmembrane signal.<sup>21</sup> A significant increase in the ROS level appeared after incubation with 10–50 pg/ml for 30 min compared with the vehicle alone (Figure 2d). The increase in ROS production was transiently augmented between 10 and 30 min after incubation with 50 pg/ml of rVvpE (Figure 2e). In addition, a pretreatment with M $\beta$ CD as well as silencing of ITLN significantly blocked rVvpE-induced ROS production (Figure 2f). The regulatory effects of M $\beta$ CD, ITLN siRNA, and antioxidant, N-acetylcysteine (NAC) on ROS production were further visualized by staining HT29-MTX cells with a fluorescent dye, 2', 7'-dichlorofluorescein diacetate (CM-H<sub>2</sub>DCFDA) (Figure 2g). Consistently, the repression of Muc2 by rVvpE was significantly blocked by the treatment with NAC (Figure 2h). These data suggest an involvement of a lipid raft and ROS production in rVvpE-mediated Muc2 regulation.

#### Essential role of protein kinase C (PKC) in Muc2 regulation.

Given that ROS has an important role as signal messengers in regulating cellular functions through activation of protein kinase C (PKC),<sup>22</sup> we examined whether rVvpE induces the phosphorylation and translocation of PKC as an important downstream intermediate of ROS in mucus-secreting HT29-MTX cells. rVvpE significantly induced PKC phosphorylation from 30 min (Figure 3a). The phosphorylation of PKC by rVvpE was blocked by NAC (Figure 3b), indicating that the functional importance of ROS in this regulation. In an experiment to identify the specific PKC isoforms, the translocation of PKC $\delta$ , but not PKC $\alpha$  or PKC $\zeta$ , from the cytosol to the membrane compartment was observed after cells were treated with rVvpE (Figure 3c). The membrane translocation of PKC $\delta$  was further confirmed by immunofluorescence staining in rVvpE-treated HT29-MTX cells (Figure 3d). We also assessed the involvement of calcium influx during the repression of Muc2 induced by rVvpE. As shown in Figure 3e, 50 pg/ml of rVvpE induced an increase in calcium influx. A Ca<sup>2+</sup> ionophore A23187, which increases [Ca<sup>2+</sup>]<sub>i</sub>, was used as a positive control to validate the results. Importantly, the rVvpE-induced Muc2 repression was inhibited by transfection with PKC $\delta$  siRNA (Figure 3f). These data suggest a functional role of PKC $\delta$  in regulating rVvpE-mediated Muc2 repression.

#### Regulatory effect of VvpE on MAPK activation and Muc2 promoter methylation.

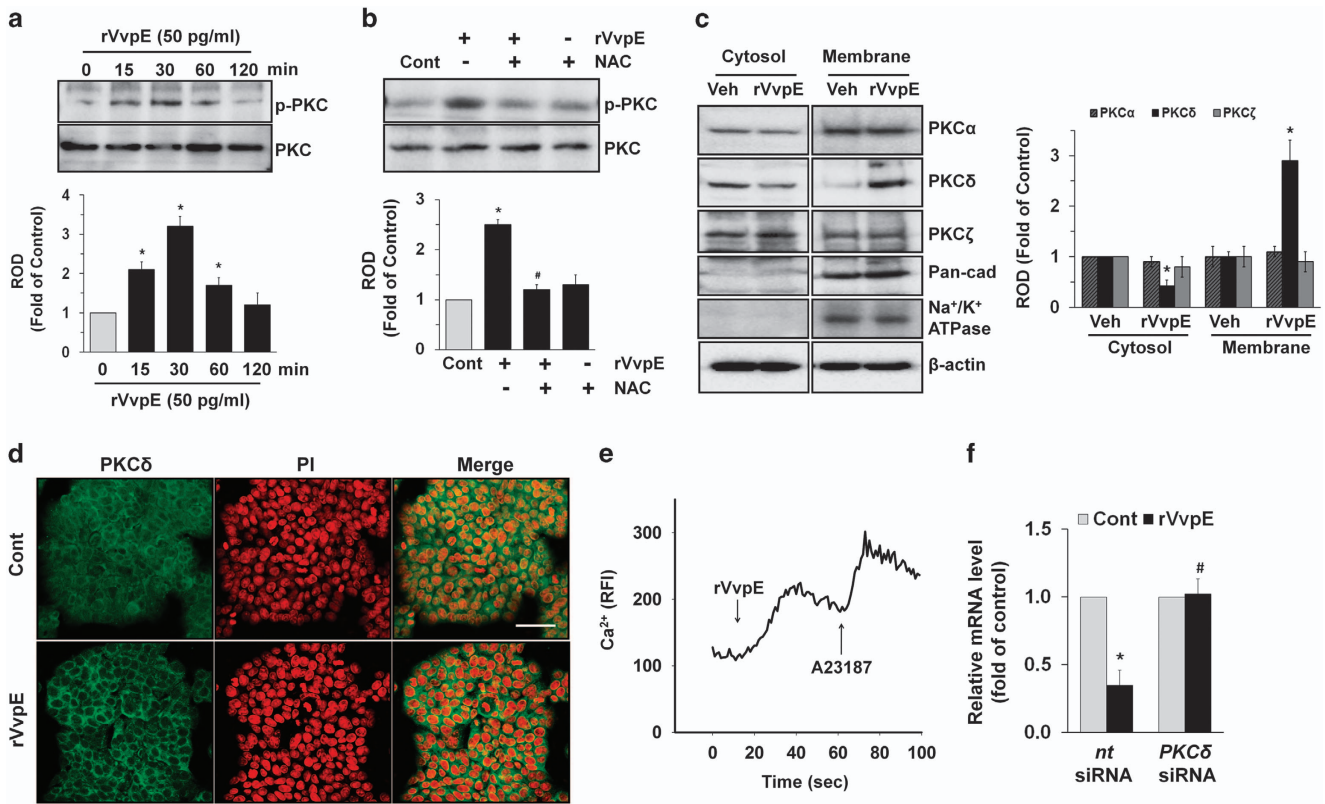
We then determined how rVvpE links to the activation of MAPKs, which are interesting candidates as downstream mediators of ROS and PKC in the regulation of mucin production.<sup>23</sup> rVvpE increased the phosphorylation of ERK between 60 and 120 min (Figure 4a) but did not affect the phosphorylation of JNK or p38 MAPK. Knockdown of ERK1/2 with specific ERK1/2 siRNA significantly blocked the Muc2 reduction induced by rVvpE (Figure 4b). In addition, the phosphorylation of ERK evoked by a treatment with rVvpE was markedly inhibited by transfection with PKC $\delta$  siRNA (Figure 4c). These data represent important evidence that ERK phosphorylation is regulated by the activation of PKC, as required for Muc2 reduction. Because ERK is known to affect gene methylation directly for epigenetic alteration,<sup>24</sup> the methylation status following the treatment with rVvpE was determined by means of methyl-specific PCR. A bisulfite treatment converted



**Figure 2** Involvement of a lipid raft and ROS production in *Muc2* regulation. HT29-MTX cells were incubated in the presence of rVvpE (50 pg/ml) for 30 min and then harvested. (a) Caveolin-enriched membrane fractions were prepared by discontinuous sucrose density-gradient fractionation, and the location of caveolin-1, flotillin-1, ITLN, and NOX2, and NCF1 was determined by western blot. (b) ITLN co-immunoprecipitated with caveolin-1, NOX2, and NCF1 (left side). Expression of caveolin-1, NOX2, NCF1, and ITLN in total cell lysates is shown in the right side.  $n=3$ .  $^*P<0.05$  versus Veh (boiled rVvpE, 50 pg/ml). ROD, relative optical density. (c) Cells were pretreated with M $\beta$ CD (0.1 mM) for 60 min prior to rVvpE exposure for 120 min. The expression level of *Muc2* mRNA was determined by real-time PCR.  $n=5$ .  $^*P<0.01$  versus Cont (boiled rVvpE, 50 pg/ml).  $^{\#}P<0.05$  versus rVvpE alone. Dose (d) and time (e) responses of rVvpE in ROS production are shown.  $n=5$ .  $^*P<0.05$  versus the cells with no treatment. RFU, Relative fluorescence units. (f) HT29-MTX cells were pretreated with M $\beta$ CD for 60 min or transfected with *ITLN* siRNA for 24 h prior to rVvpE (50 pg/ml) exposure for 30 min. The level of ROS production is shown. Error bars represent the means  $\pm$  S.E. ( $n=5$ ).  $^*P<0.05$  versus *nt* siRNA+Cont (boiled rVvpE, 50 pg/ml).  $^{\#}P<0.05$  versus *nt* siRNA+rVvpE. (g) ROS production (green) was visualized by confocal microscopy. Scale bars, 100  $\mu$ m.  $n=3$ . (h) Cells were pretreated with NAC (1 mM) for 30 min prior to rVvpE exposure for 120 min. The level of *Muc2* expression is shown.  $n=5$ .  $^*P<0.01$  versus Cont (boiled rVvpE, 50 pg/ml).  $^{\#}P<0.05$  versus rVvpE alone

cytosine residues in the genomic DNA to uracil, which were amplified as thymine during the subsequent PCR. We attempted to analyze the methylation status of the *Muc2* gene promoter containing three CpG sites (-193, -274, and -289), which are critical CpG sites responsible for the repression of *Muc2* expression.<sup>25</sup> As shown in Figure 4d, primers for -193 and -274 CpG sites that specifically amplified either the methylated or unmethylated form of the *Muc2* promoter produced 137 bp and 105 bp methylated bands from vehicle-treated HT29-MTX cells, respectively, indicating that the *Muc2* promoter was methylated in these alleles. However, the

methylated bands of the *Muc2* gene did not appear at the -289 CpG site. Interestingly, 50 pg/ml of rVvpE markedly increased the level of *Muc2* promoter methylation at the -274 CpG site, but not at -193 for 60 min. We further explored the ability of 50 pg/ml of rVvpE to induce methylation for 60 min and quantified the relative value of CpG methylation compared with an unmethylated form in the *Muc2* promoter using a real-time PCR analysis (Figure 4e). The relative level of *Muc2* promoter methylation at the -274 CpG site was significantly augmented by a treatment with rVvpE between 30 and 60 min compared with a control. As



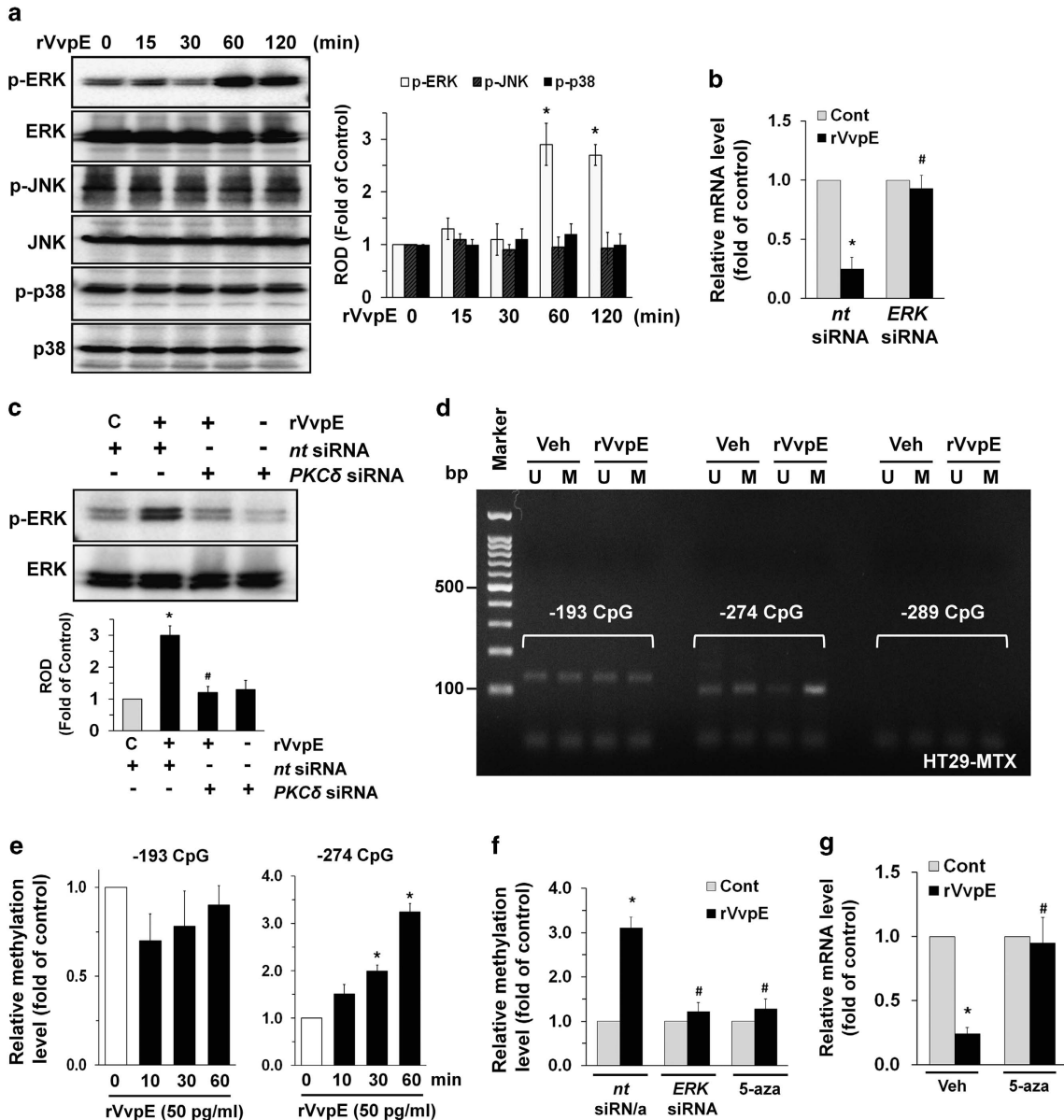
**Figure 3** Essential role of PKC in Muc2 regulation. (a) Phosphorylation of PKC in cells treated with rVvpE is shown.  $n = 3$ .  $*P < 0.01$  versus the cells with no treatment. ROD, Relative optical density. (b) Cells were pretreated with NAC (1 mM) prior to rVvpE exposure for 30 min. Phosphorylation of PKC is shown.  $n = 4$ .  $*P < 0.01$  versus Cont (boiled rVvpE, 50 pg/ml).  $^{\#}P < 0.05$  versus rVvpE alone. (c) Membrane translocation of PKC isoforms in cells treated with rVvpE for 30 min was determined by western blot. Pan-cadherin and Na<sup>+</sup>/K<sup>+</sup> ATPase were used as a plasma membrane control.  $n = 3$ .  $*P < 0.05$  versus Veh (boiled rVvpE, 50 pg/ml). (d) Membrane translocation of PKCδ (green) was determined by confocal microscopy. PI was used for nuclear counterstaining (red). Scale bars, 100 μm.  $n = 3$ . (e) Changes in [Ca<sup>2+</sup>]<sub>i</sub> were monitored by confocal microscopy, and data are expressed as relative fluorescence intensity (RFI, F/F0%, arbitrary unit). A23187 (10 μM) was used as a positive control.  $n = 4$ . (f) Cells were transfected with PKCδ siRNA for 24 h prior to rVvpE (50 pg/ml) exposure for 120 min. The level of Muc2 mRNA expression is shown.  $n = 4$ .  $*P < 0.01$  versus nt siRNA+Cont (boiled rVvpE, 50 pg/ml).  $^{\#}P < 0.05$  versus nt siRNA+rVvpE.

expected, the methylation status of the *Muc2* promoter containing the -193 CpG site was not changed by a treatment with rVvpE. Interestingly, the level of *Muc2* promoter methylation at the -274 CpG site was significantly inhibited by knockdown of *ERK1/2* and by treatment with DNA methylation inhibitor 5-azacytidine (5-aza) (Figure 4f). In addition, the pretreatment with 5-aza significantly blocked the repression of *Muc2* as induced by rVvpE (Figure 4g). These data indicate that the ERK-mediated hypermethylation of the *Muc2* promoter is responsible for the low level of *Muc2* expression in rVvpE-treated HT29-MTX cells.

To ensure the functional role of rVvpE and its related signaling molecules in regulation of transcriptional repression of *Muc2*, we further determined the effect of siRNAs and inhibitors on the activation of the signaling pathway induced by rVvpE using an ELISA assay (Figures 5a and b). We pretreated cells with MβCD and NAC to prove the involvement of lipid rafts-dependent ROS production, PKCδ and ERK1/2 siRNAs to confirm involvement PKCδ and ERK activation, and 5-aza to show involvement of *Muc2* promoter methylation. As expected, the repression of *Muc2* production induced by rVvpE was significantly abrogated by the aforementioned siRNAs and inhibitors for lipid rafts, ROS, PKC, ERK, and methylation, respectively. These results demonstrate the relevance of lipid

rafts-mediated ROS signaling pathway and hypermethylation in promoting rVvpE-mediated *Muc2* repression. The sequences of putative bacterial signaling pathways of VvpE are summarized in Figure 5c.

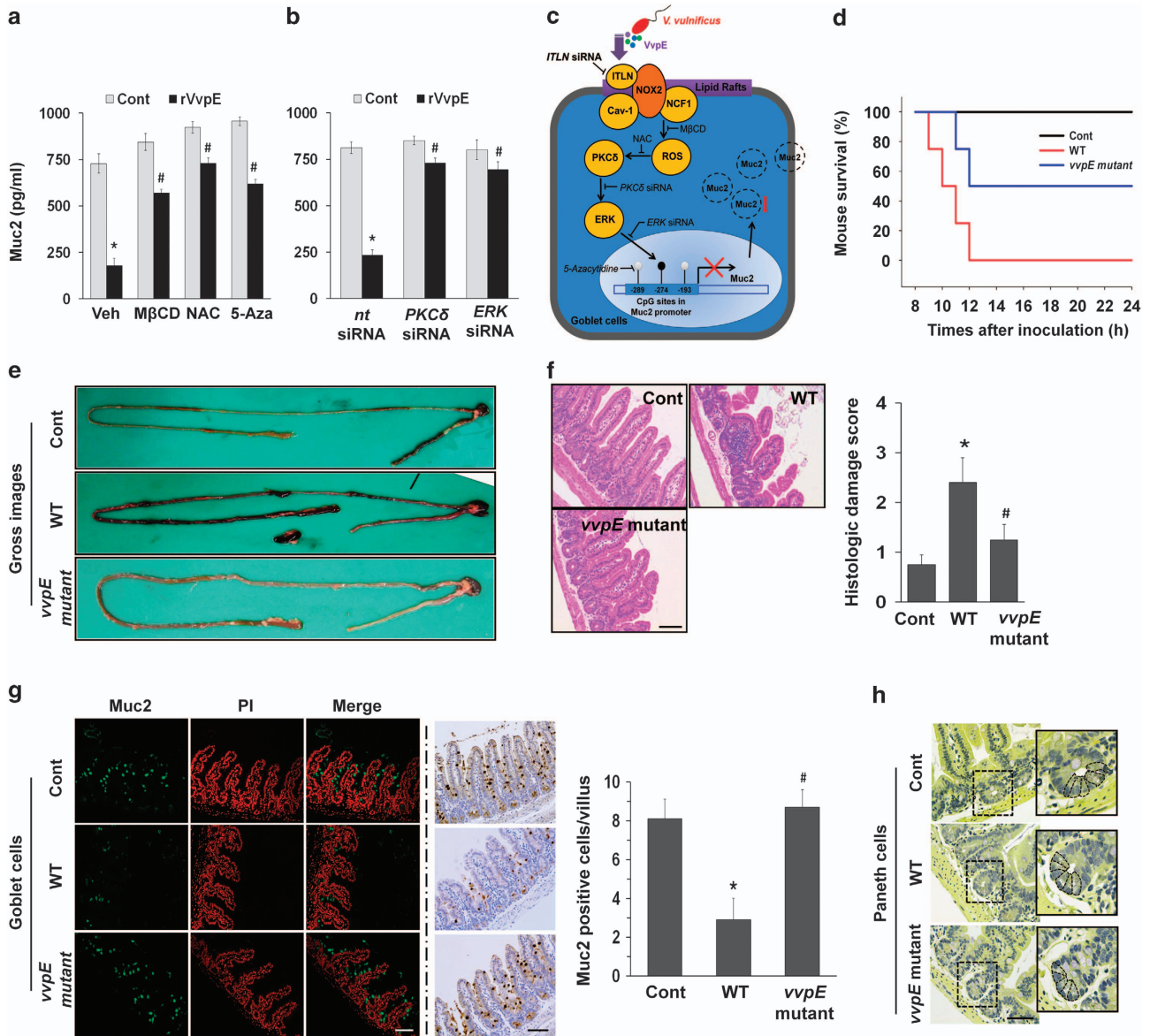
To evaluate the clinical relevance of *V. vulnificus* VvpE, we evaluated the effect of VvpE on mouse lethality. We used an iron-overloaded mouse model to bring the growth of *V. vulnificus* within the lethal level.<sup>26</sup> Seven-week-old iron-overloaded ICR mice inoculated intraperitoneally (i.p.) with boiled *V. vulnificus* (Cont), *V. vulnificus* (WT), and a mutant deficient in *vvpE* gene in *V. vulnificus* (*vvpE* mutant) at  $1.2 \times 10^2$  CFU/ml for 24 h, after which their survival rates were monitored (Figure 5d). All of the mice injected with WT were dead by 12 h post injection, while 5 out of the 10 mice injected with the *vvpE* mutant remained alive 24 h after i.p. inoculation, thus showing some degree of the attenuation of mouse lethality. These results indicate that VvpE makes an important contribution to the lethality of mice infected with WT. Finally, we further investigated the effect of VvpE on the expression of intestinal *Muc2* in mouse. Mice inoculated intragastrically (i.g.) with Cont, WT, and *vvpE* mutant at  $1.1 \times 10^9$  CFU/ml for 16 h. WT caused severe necrotizing enteritis of the intestine (Figure 5e), where it induced shortened villi heights accompanied by an expanded width and increased inflammation at



**Figure 4** Regulatory effect of VvpE on MAPK activation and *Muc2* promoter methylation. (a) The effect of rVvpE on the expression of MAPK was determined by western blot.  $n = 4$ .  $*P < 0.01$  versus the cells with no treatment. ROD, relative optical density. (b) Cells were transfected with *ERK* siRNA for 24 h prior to rVvpE exposure for 120 min. The level of *Muc2* mRNA expression was measured by using real-time PCR.  $n = 4$ .  $*P < 0.01$  versus Cont (boiled rVvpE, 50 pg/ml).  $^{\#}P < 0.05$  versus rVvpE alone. (c) Cells were transfected with *PKCδ* siRNA for 24 h prior to rVvpE exposure for 60 min.  $n = 3$ .  $*P < 0.01$  versus nt siRNA+C (boiled rVvpE, 50 pg/ml).  $^{\#}P < 0.05$  versus nt siRNA+rVvpE. C, Control. ROD, relative optical density. (d) Genomic DNA treated with rVvpE for 60 min was prepared. Changes in the promoter methylation status of *Muc2* gene was determined by methyl-specific PCR analysis.  $n = 4$ . U, unmethylated form; M, methylated form. (e) Time responses of rVvpE in *Muc2* methylation status at -274 and -193 CpG sites are shown. The relative level of *Muc2* methylation is shown, compared to the unmethylated form.  $n = 4$ .  $*P < 0.05$  versus the cells with no treatment. (f) Cells were transfected with *ERK* siRNA for 24 h or pretreated with 5-azacytidine (5-aza) (1  $\mu$ M) for 30 min prior to rVvpE exposure for 60 min. The relative level of *Muc2* methylation is shown, compared to the unmethylated form.  $n = 4$ .  $*P < 0.05$  versus nt siRNA+Cont (boiled rVvpE, 50 pg/ml).  $^{\#}P < 0.05$  versus nt siRNA+rVvpE. (g) Cells were pretreated with 5-aza for 30 min prior to rVvpE exposure for 120 min. The level of *Muc2* mRNA expression was measured by using real-time PCR.  $n = 4$ .  $*P < 0.01$  versus Cont (boiled rVvpE, 50 pg/ml).  $^{\#}P < 0.05$  versus rVvpE alone

16 h infection, resulting in increased level of histopathological damage score, compared with control mice (Figure 5f). However, unlike WT, the mice infected with *vvpE* mutant almost completely maintained their intestinal villi structures, and the inflammation was silenced. The result of the immunofluorescence staining of *Muc2* revealed that mice

infected with WT for 16 h have fewer goblet cells expressing *Muc2*, which was negated by infection with *vvpE* mutant (Figure 5g). However, WT did not alter the number of paneth cells in the small intestine for 16 h infection (Figure 5h). Thus, these results indicate that VvpE has an important role in repression of *Muc2* during *V. vulnificus* infection.



**Figure 5** Role of VvpE and its related signal molecules on Muc2 production. **(a)** HT29-MTX cells were pretreated with MβCD, NAC, and 5-azacytidine prior to rVvpE (50 pg/ml) exposure for 4 h. The level of Muc2 protein was quantified by ELISA. Error bars represent the means ± S.E. ( $n = 5$ ). \* $P < 0.01$  versus Cont (boiled rVvpE, 50 pg/ml). # $P < 0.05$  versus rVvpE alone. **(b)** Cells were transfected with PKCδ siRNA and ERK siRNA prior to rVvpE (50 pg/ml) exposure for 4 h. \* $P < 0.01$  versus nt siRNA+Cont (boiled rVvpE, 50 pg/ml). # $P < 0.05$  versus nt siRNA+rVvpE alone. **(c)** A proposed model for VvpE-evoked signaling pathway in intestinal epithelial cells. VvpE has an important role in the targeting of *V. vulnificus* for goblet cells, where VvpE represses Muc2 expression via the regulation of ITLN-mediated ROS production and distinctive hypermethylation of Muc2 promoter. **(d)** Iron-overloaded ICR mice were received i.p. inoculation of WT, boiled WT (Cont), and vvpE mutant at  $1.2 \times 10^2$  CFU/ml for 24 h. Survival rate of mice infected with vvpE mutant relative to WT is shown.  $n = 10$ .  $P < 0.05$ . Mice were given i.g. inoculation of WT and vvpE mutant at  $1.1 \times 10^9$  CFU/ml and killed at 16 h later. **(e)** Gross morphologies of intestines are shown. **(f)** Representative ileum tissues stained with hematoxylin and eosin are shown (left panel).  $n = 5$ . Scale bars represent 100 μm. Average scores of histopathologic damage index from mouse ileum is shown (right panel).  $n = 5$ . \* $P < 0.01$  versus Cont. # $P < 0.05$  versus WT. **(g)** Expression of Muc2 in mouse ileum was examined by immunofluorescence (left panel, green) and immunohistochemical analysis (right panel, brown). PI (red) was used for nuclear counterstaining for immunofluorescence analysis. Scale bars represent 100 μm. The mean numbers of Muc2-labeled cells per villi are shown in the bar graph. \* $P < 0.01$  versus Cont. # $P < 0.01$  versus WT. Error bars represent the means ± S.E. from five number of villi and crypts from five different mice. **(h)** Paneth cells were stained using the phloxine-tartrazine staining method. There were no changes in the mean number of paneth cells per crypt ( $n = 5$ ). Scale bars, 50 μm

## Discussion

Our data demonstrate that *V. vulnificus* VvpE is the functional elastase responsible for Muc2 repression in the mouse and that VvpE, in acting through lipid raft-associated ITLN, inhibits Muc2 expression by stimulating the methylation of the Muc2 gene promoter through the ROS-dependent activation of PKCδ/ERK pathway in mucus-secreting HT29-MTX cells.

Many enteric pathogens directly degrade intestinal mucin by producing Pic,<sup>27</sup> StcE,<sup>28</sup> or Hap.<sup>29</sup> However, our result in the present study suggests that *V. vulnificus* uniquely regulates intestinal Muc2 expression by producing VvpE with modes of action that differ from those of other enteric pathogens. It is not clear whether the functional role of VvpE in the repression of Muc2 *in vivo* is a sequential result of the activation of different

types of cells in mouse intestine or, alternatively, an independent process involving other cellular signaling events. In addition, it is also possible that the VvpE would impact on the delivery of major toxins (e.g., VvhA, MARTXVv) during intestinal infection of *V. vulnificus*. However, it is clearly reported that the impairment of the mucus layer enhances the colonization of pathogens with establishment of the appropriate portal of entry, where it gains access more easily to epithelial cells and thereby promotes pathogen–host adherence mechanisms. Thus, these results suggest VvpE has important role in the pathogenesis of *V. vulnificus* in intestine. Although previous research has raised some serious doubts about the critical role of VvpE in mouse lethality,<sup>17</sup> our *in vivo* data revealed that VvpE is a necessary virulence factor responsible the mouse lethality of *V. vulnificus*. This means that the discrepancy with regard to the functional role of VvpE could be because of differences in the pathogenic isolate of *V. vulnificus* (MO6-24/O), the anesthesia method, or the injection location used in this study.<sup>30,31</sup> In addition, many relevant reports have suggested that VvpE causes tissue necrosis and increased vascular permeability, which are necessary for the invasion of this bacterium.<sup>32,33</sup> Thus, we suggest that VvpE is another important virulence factor of *V. vulnificus* responsible for mouse lethality and Muc2 repression.

Concerning the pathogenic mechanism of VvpE, we showed that a unique relationship between lipid raft-dependent ITLN recruitment and ROS production in the regulation of Muc2 expression. Our finding that ITLN is localized on lipid raft areas exposed on the membrane surface of mucus-secreting HT29-MTX cells is consistent with a previous report that ITLN was found in intestinal microvillar rafts, enriched in glycoproteins and glycolipids, where it provides a number of carbohydrate-binding sites for microbial adhesion.<sup>13</sup> However, ITLNs are proposed to serve as a host defense lectin to assist with the phagocytic clearance of microorganisms.<sup>34</sup> These differ from our data that ITLN is a functional signaling molecule which mediates the virulence effect of VvpE to repress Muc2 expression. Moreover, previous work revealed that transgenic mice overexpressing ITLN did not show enhanced pathogen clearance,<sup>35</sup> suggesting that ITLN has a critical role as host mediator of rVvpE in lipid rafts. The important finding of this study is that rVvpE triggers clustering of ITLN and NOX in the lipid raft for the ROS production, which results in a repression of Muc2. Increasing evidence has suggested that lipid rafts are clustered to form a redox signaling platform through gp91<sup>phox</sup> (NOX2) coupling with cytosolic factors that include p47<sup>phox</sup> (NCF1), p67<sup>phox</sup> (NCF1), and small GTPase Rac1, and that these processes subsequently produce superoxides and other ROS.<sup>36,37</sup> These results are further supported by a previous study in which several enteric toxins that interact with lipid rafts, including *Helicobacter* (*H.*) *pylori* vacuolating toxin<sup>38,39</sup> and *Clostridium* (*C.*) *perfringens* enterotoxin,<sup>40</sup> have shown the ability to regulate NOX aggregation via lipid raft clustering as an initial attachment platform, thus having a virulence effect on intestinal pathogenesis.

An influx of Ca<sup>2+</sup> on PKC-mediated MAPK activation is linked to bacterial stratagems to modulate the host signaling pathway. *H. pylori* and *C. perfringens* have been shown to elicit

Ca<sup>2+</sup> response in the gut.<sup>41</sup> Moreover, multiple signaling processes, such as those acting through the Ca<sup>2+</sup> and PKC/MAPK pathways, were rapidly activated in target cells through ROS production.<sup>42,43</sup> These previous results are consistent with our current finding that rVvpE induces an influx of Ca<sup>2+</sup> on PKC/MAPK activation, in that PKC is required for ROS production to repress Muc2 expression. Indeed, ITLN was shown to evoke Ca<sup>2+</sup> signaling to recognize the pathogen infection.<sup>11</sup> Interestingly, rVvpE uniquely regulated Muc2 repression through two distinct proteins, PKC $\delta$  and ERK. A previous report showed *H. pylori* activates novel PKC $\delta$  to control MAPK pathways.<sup>44</sup> However, other authors showed that the *H. pylori* infection regulated p38 MAPK activation in promoting ROS signaling pathway.<sup>45</sup> These differ from our data that rVvpE increases ERK phosphorylation via the activation of PKC $\delta$  during ROS production. On the other hand, a previous report showed conventional PKC $\alpha$  activation was found to require for the impairing intestinal barrier function induced by enteropathogenic *Escherichia coli* infection,<sup>46</sup> suggesting that the cellular pathways differ in different bacterial pathogens type. These data indicate that rVvpE selectively regulates specific PKC isozymes and MAPK phosphorylation. In keeping with the unique signaling pathway of rVvpE repressing Muc2 expression, our study showed that rVvpE induced the region-specific methylation of the Muc2 promoter at the –274 site through ERK and that the inhibition of DNA methylation blocks rVvpE-induced Muc2 reduction. Promoter methylation of cytosine residues at CpG dinucleotides is an important epigenetic mechanism in promoting the transcriptional repression of Muc2.<sup>10</sup> Compelling evidence supports the role of PKC $\delta$  and ERK in the hypermethylation of tumor-suppressor genes and the pathogenesis of colon cancer.<sup>24</sup> These data provide the first evidence that bacterial pathogen promotes hypermethylation of the Muc2 promoter to repress Muc2 expression.

In conclusion, our results suggest that VvpE regulates DNA methylation via the activation of lipid rafts-mediated ROS signaling pathway to promote the transcriptional repression of Muc2. Thus, highlighting the new pathogenic signaling pathways involved in VvpE-induced Muc2 repression may provide potential therapeutic strategies for *V. vulnificus* infections in intestine.

## Materials and Methods

**Chemicals.** Fetal bovine serum (FBS) was purchased from BioWhittaker (Walkersville, MO, USA). The following antibodies were purchased: p-PKC and PKC antibodies (Cell Signaling Technology, Danvers, MA, USA); NOX2 antibody (BD Biosciences, Franklin Lakes, NJ, USA); NCF1 antibody (LifeSpan Biosciences, Seattle, WA, USA); ITLN and Na<sup>+</sup>/K<sup>+</sup> ATPase antibodies (abcam, Cambridge, MA, USA); p-ERK1/2, ERK, p-JNK, JNK, p-p38 MAPK, p38 MAPK, p-c-Src, c-Src, p-FAK, FAK, pan-cadherin, PKC $\alpha$ , PKC $\delta$ , and PKC $\zeta$  antibodies (Santa Cruz Biotechnology, Paso Robles, CA, USA); Horseradish peroxidase (HRP)-conjugated goat anti-rabbit and goat anti-mouse IgG antibodies (Jackson ImmunoResearch, West Grove, PA, USA); 2', 7'-dichlorofluorescein diacetate (CM-H<sub>2</sub>DCFDA) was obtained from Invitrogen (Carlsbad, CA, USA). A23187, methyl- $\beta$ -cyclodextrin (M $\beta$ CD), *N*-acetyl-L-cysteine (NAC) and 5-azacytidine were purchased from Sigma Chemical Company (St. Louis, MO, USA). The concentrations of all of pharmacological inhibitors listed did not show any significant cytotoxic effects by themselves as confirmed by FACS analysis in each experiment. All other reagents were of the highest purity commercially available and were used as received.



**Cells.** Mucus-secreting human intestinal epithelial (HT29-MTX) and human colorectal epithelial (Caco-2) cells were purchased from American Type Culture Collection (ATCC, Manassas, VA, USA). HT29-MTX and Caco-2 cells were grown at 37°C in 5% CO<sub>2</sub> in a McCoy's 5A Medium supplemented with 10% FBS and antibiotics (10 units/ml penicillin G and 10 µg/ml streptomycin). HT29-MTX cells have been used to study adhesion and invasion of pathogens because of its physiologically relevant characteristics that are responsible for the mucus layer formation.<sup>20,47</sup> Caco-2 cells were used as an alternative epithelial cell line to confirm the role of rVvpE in *Muc2* mRNA expression.

**Overexpression and purification of the recombinant elastolytic protease VvpE.** To find the functional role of VvpE in HT29-MTX cells, we have previously prepared a recombinant protein of VvpE (rVvpE). Briefly, the coding region of *vvpE* encoding an elastolytic protease was amplified and then subcloned into a His<sub>6</sub> tag expression vector, pET29a(+) (Novagen, Madison, WI, USA), resulting pKS1202.<sup>19</sup> The His-tagged VvpE protein was overexpressed in *E. coli* BL21 (DE3) and then purified by affinity chromatography according to the manufacturer's procedure (Qiagen, Valencia, CA, USA).

**Flow cytometry.** HT29-MTX cells were synchronized in the G<sub>0</sub>/G<sub>1</sub> phase by culture in serum-free media for 24 h before incubation of rVvpE. The necrotic cell death of HT29-MTX cells was detected with an Annexin V and PI staining kit (BD Biosciences) according to the manufacturer's instructions. Briefly, the cells were detached with 0.05% trypsin/EDTA and 1 × 10<sup>5</sup> cells were resuspended with Annexin V binding buffer (0.1 M HEPES/ NaOH (pH 7.4), 1.4 M NaCl, 25 mM CaCl<sub>2</sub>). And then the cells were stained with Annexin V (25 µg/ml) and PI (125 ng/ml), and incubated for 15 min at room temperature in the dark. The sample was read by flow cytometry and analyzed using CXP software (Beckman Coulter, Brea, CA, USA).

**Small interfering (si)RNA transfection.** Cells were grown until 75% of the surface of the plate and transfected for 24 h with either a siRNAs specific for *ITLN* and *PKCδ* (GE Dharmacon, Lafayette, CO, USA) or non-targeting (*nt*) siRNA as a negative control (GE Dharmacon) with HiPerFect Transfection Reagent (Qiagen) according to the manufacturer's instructions. The transient knockdown of *ERK1/2* was achieved by transfection with specific amounts of siRNA (100 nM) obtained from Cell Signaling. The siRNA efficacy for *ITLN*, *PKCδ*, and *ERK1/2* were determined by western blot (Supplementary Figure S3).

**RT-PCR.** Total RNA was extracted using the RNeasy Plus Mini Kit (Qiagen). Reverse transcription was carried out with 3 µg of RNA using a Maxime RT premix kit (iNTRON Biotechnology, Sungnam, Korea). The cDNAs (5 µl) for mucin isoforms were amplified using the primers described in Supplementary Table S1. The real-time quantifications of pro-inflammatory cytokines and *Muc2* were performed using a Rotor-Gene 6000 real-time thermal cycling system (Corbett Research, Mortlake, NSW, Australia) with a QuantiMix SYBR Kit (PhileKorea Technology, Daejeon, Korea) according to the manufacturer's instructions.  $\beta$ -actin was used as an endogenous control.

**Mucin (Muc) 2 ELISA.** HT29-MTX cells plated on 60-mm culture dishes were grown in FBS-free medium for 24 h and divided into groups according to the experimental protocol. The *Muc2* concentration in the culture medium was quantified by an ELISA (Elabscience Biotech, Wuhan, Hubei, China) according to the manufacturer's instructions.

**Western blot analysis and subcellular fractionation.** Western blotting was performed as previously described with minor modifications.<sup>48</sup> The subcellular fractionation method for the isolation of membrane and cytosolic proteins was previously reported.<sup>49</sup>

**Immunoprecipitation.** Interaction of *ITLN* with NOX, NCF1 or caveolin-1 was analyzed by immunoprecipitation and western blotting. Cells were lysed with lysis buffer (1% Triton X-100 in 50 mM Tris-HCl pH 7.4 containing 150 mM NaCl, 5 mM EDTA, 2 mM Na<sub>3</sub>VO<sub>4</sub>, 2.5 mM Na<sub>4</sub>PO<sub>7</sub>, 100 mM NaF, 200 nM microcystin lysine-arginine, and protease inhibitors). Cell lysates (400 µg) were mixed with 10 µg of each antibodies. The samples were incubated for 4 h, mixed with Protein A/G PLUS-agarose immunoprecipitation reagent (Pierce, Rockford, IL, USA) and then incubated for an additional 12 h. The beads were washed four times, and the bound proteins were released from the beads by boiling in SDS-PAGE sample buffer for 5 min. Samples were analyzed by western blotting.

### Detergent-free purification of caveolin-rich membrane fraction.

HT29-MTX cells were washed twice with ice-cold phosphate-buffered saline (PBS), scraped into 2 ml of 500 mM sodium carbonate (pH 11.0), transferred to a plastic tube, and homogenized with a Sonicator 250 apparatus (Branson Ultrasonic, Danbury, CT, USA) using three 20 s bursts. The homogenate was adjusted to 45% sucrose by the addition of 2 ml 90% sucrose prepared in 2 (*N*-morpholino) ethanesulfonic acid (MES)-buffered solution consisting of 25 mM MES-buffer solution (pH 6.5) and 0.15 M NaCl and placed at the bottom of an ultracentrifuge tube. A 5%–35% discontinuous sucrose gradient was formed (4 ml each of 5% and 35% sucrose, both in MES-buffer solution containing 250 mM sodium carbonate) and centrifuged at 40 000xg for 20 h in a Beckman SW41 Rotor (Beckman Coulter). Twelve fractions were collected and analyzed by 12% SDS-PAGE.

### Intracellular reactive oxygen species detection.

2', 7'-Dichlorofluorescein diacetate (CM-H<sub>2</sub>DCFDA) was used to detect the general ROS production. To quantify the intracellular ROS levels, the cells treated with 10 mM DCF-DA were rinsed twice with ice-cold PBS and then scraped. A 100 µl cell suspension was loaded into a 96-well plate and examined using a luminometer (Victor3; Perkin-Elmer, MA, USA) and a fluorescent plate reader at excitation and emission wavelengths of 485 and 535 nm, respectively.

### Measurement of calcium influx.

Changes in intracellular calcium concentrations were monitored using Fluo-3-AM (Invitrogen, Carlsbad, CA, USA) as previously reported with minor modifications.<sup>48</sup> Fluorescence was excited at 488 nm and the emitted light was observed at 515 nm. All analyses of calcium influx were processed in a single cell, and the results are expressed as the fluorescent intensity.

### Immunofluorescence and immunohistochemical analysis.

Either HT29-MTX cells or ileum frozen sections were fixed in 4% paraformaldehyde in PBS for 10 min at room temperature, permeabilized in 0.1% Triton X-100 in PBS for 5 min, and blocked in PBS containing 5% (v/v) normal goat serum (NGS) for 30 min at room temperature. Samples were then stained with primary antibody for overnight at 4 °C. Following three washes with PBS, the samples were incubated with Alexa 488-conjugated goat anti-rabbit/mouse IgM (Invitrogen), and counterstained with PI in PBS containing 5% (v/v) NGS for 2 h. After washing with PBS, samples were mounted on slides and visualized with an Olympus FluoView 300 confocal microscope (Melville, NY, USA) with x400 objective. For immunohistochemical analysis, ileum frozen tissues incubated with primary antibody for overnight at 4 °C were treated with biotinylated secondary antibody solution (Vectastain Elite ABC kit, Vector Laboratories, CA, USA) for 1 h at room temperature. Sections were washed with PBS, incubated in the ABC reagent for 1 h at room temperature, washed again and incubated in a peroxidase solution. Sections were then counterstained with hematoxylin, dehydrated, and coverslipped. Images were acquired using an Axioskop 2 plus microscope equipped with an AxioCam MRc5 CCD camera (Zeiss, Thornwood, NY, USA). Other samples were subjected to hematoxylin and eosin staining for histological examinations.

### Methylation analysis.

Genomic DNA from HT29-MTX cells was prepared with the QIAamp DNA Mini Kit (Qiagen, Valencia, CA, USA). The extracted DNA was treated with sodium bisulfite using EzWay DNA Methylation Detection Kit according to the manufacturer's instructions (KOMABIOTECH, Seoul, Korea). The methylation status of *Muc2* gene in HT29-MTX cells was determined by methyl-specific PCR analysis. We conducted methyl-specific PCR of *Muc2* gene promoter at three CpG sites (–289, –274, –193) which have functional role in regulation of *Muc2* expression as described previously.<sup>25</sup> The primer sequences for each CpG sites of *Muc2* promoter and size of products are summarized in Supplementary Table S2.

### Bacterial strains, plasmids, and culture media.

The strains and plasmids used in this study are listed in Table 1. All *V. vulnificus* strains (M06-24/O and M06-24/O *vvpE*) are isogenic and naturally resistant to polymyxin B. Unless otherwise noted, *V. vulnificus* strains were grown in Luria Bertani medium supplemented with 2.0% (wt/vol) NaCl (LBS) at 30 °C. All media components were purchased from Difco (Difco Laboratories, Detroit, MI, USA). *V. vulnificus* were grown to mid-log phase ( $A_{600} = 0.500$ ) corresponding to 2 × 10<sup>8</sup> CFU/ml and centrifuged at 6000 × g for 5 min. The pellet was washed with PBS and adjusted to desired colony-forming unit (CFU)/ml based on the  $A_{600}$  determined using a UV-VIS spectrophotometer (UV-1800, Kyoto, Shimadzu, Japan) to estimate culture density.

**Table 1** Plasmids and bacterial strains used in this study

Strain or plasmid	Relevant characteristics	Reference or source
<i>Bacterial strains</i>		
<i>V. vulnificus</i>		
M06-24/O	Clinical isolate; virulent; WT	Laboratory collection
CMM111	M06-24/O <i>vvpE</i> ::pKC9844; elastase deficient; <i>vvpE</i> mutant	Jeong et al. <sup>17</sup>
<i>E. coli</i>		
BL21 (DE3)	F <sup>-</sup> <i>ompT hsdS<sub>B</sub> (r<sub>B</sub>m<sub>B</sub>) gal dcm</i> (DE3)	Laboratory collection
DH5α	λ φ80d <i>lacZ</i> ΔM15 Δ( <i>lacZYA-argF</i> )U169 <i>recA1 endA1 hsdR17 (r<sub>K</sub> m<sub>K</sub>) supE44 thi-1 gyrA relA1</i> ; plasmid replication	Laboratory collection
S17-λ <i>pir</i>	λ- <i>pir</i> lysogen; <i>thi pro hsdR hsdM<sup>*</sup> recA</i> RP4-2 Tc::Mu-Km::Tn7;Tp <sup>r</sup> Sm <sup>r</sup> ; host for π-requiring plasmids; conjugal donor	Laboratory collection
<i>Plasmids</i>		
pET29a(+)	His <sub>6</sub> tag fusion expression vector; Kn <sup>r</sup>	Novagen
pKS1202	pET29a(+) with 1,827-bp <i>vvpE</i> ; Kn <sup>r</sup>	Kim et al. <sup>19</sup>
pVSV102	<i>OriV<sub>R6Ky</sub> oriT<sub>RP4</sub> oriV<sub>pES213</sub> Kn<sup>r</sup> gfp</i>	Dunn et al. <sup>51</sup>

Abbreviations: Kn<sup>r</sup>, kanamycin resistant; Sm<sup>r</sup>, streptomycin resistant; Tp<sup>r</sup>, trimethoprim resistant

**Mouse models.** All animal procedures were performed following the National Institutes of Health Guidelines for the Humane Treatment of Animals, with approval from the Institutional Animal Care and Use Committee of Seoul National University (SNU-140108-4). To determine the mouse lethality, seven-week-old mice ( $n=10$ ) given 250 mg/kg iron dextran were received i.p. inoculation of boiled *V. vulnificus* (Cont), *V. vulnificus* (WT), and a mutant deficient in *vvpE* gene in *V. vulnificus* (M06-24/O *vvpE*, *vvpE* mutant) ( $1.2 \times 10^2$  CFU/ml) and the survival rate of the mice was recorded for 24 h. To confirm the expression of intestinal Muc2, mice ( $n=5$ ) were received i.g. inoculation of Cont, WT or *vvpE* mutant at  $1.1 \times 10^9$  CFU/ml for 16 h and sacrificed. We anticipated that 16 h was the minimum duration for colonization activities of the *V. vulnificus* colonization according to the previous our report.<sup>19</sup> The ileum tissues were embedded in OCT compound and stored at  $-70^\circ\text{C}$ . Samples were then cut into 6-μm-thick frozen sections.

**Histologic damage score.** Histological parameters were determined in a blinded fashion by two experienced gastrointestinal pathologists as previously described<sup>50</sup> with some modification. Briefly, scores were assigned as follows: 0 = no damage (normal); 1 = slight submucosal and/or lamina propria separation (mild); 2 = moderate separation of the submucosa and/or lamina propria and/or edema in the submucosa and muscular layers (moderate); 3 = severe separation of the submucosa and/or lamina propria and/or severe edema in the submucosa and muscular layers with regional villous sloughing (severe); or 4 = loss of villi and necrosis (necrosis).

**Paneth cells staining.** Frozen sections of ileal tissues were stained with phloxine-tartrazine to visualize paneth cells according to the Lendrum reaction. Briefly sections were treated with alum hematoxylin for 1 min, 0.5% phloxine in 0.5% aqueous calcium chloride for 20 min, and then finally differentiated with a saturated solution of tartrazine in 2-ethoxy ethanol (Sigma, St. Louis, MO, USA). With this technique, paneth granules are stained bright red.

**Statistical analysis.** Results are expressed as means ± standard errors (S.E.). All experiments were analyzed by ANOVA, followed in some cases by a comparison of treatment means with a control using the Bonferroni–Dunn test. Differences were considered statistically significant at  $P < 0.05$ .

### Conflict of Interest

The authors declare no conflict of interest.

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- Ivanov II, Honda K. Intestinal commensal microbes as immune modulators. *Cell Host Microbe* 2012; **12**: 496–508.
- Ivanov AI, Parkos CA, Nusrat A. Cytoskeletal regulation of epithelial barrier function during inflammation. *Am J Pathol* 2010; **177**: 512–524.
- Parlato M, Yeretssian G. NOD-like receptors in intestinal homeostasis and epithelial tissue repair. *Int J Mol Sci* 2014; **15**: 9594–9627.
- Shan M, Gentile M, Yeiser JR, Walland AC, Bornstein VU, Chen K et al. Mucus enhances gut homeostasis and oral tolerance by delivering immunoregulatory signals. *Science* 2013; **342**: 447–453.
- Heazlewood CK, Cook MC, Eri R, Price GR, Tauro SB, Taupin D et al. Aberrant mucin assembly in mice causes endoplasmic reticulum stress and spontaneous inflammation resembling ulcerative colitis. *PLoS Med* 2008; **5**: e54.
- Wang L, Cao H, Liu L, Wang B, Walker WA, Acra SA et al. Activation of epidermal growth factor receptor mediates mucin production stimulated by p40, a *Lactobacillus rhamnosus* GG-derived protein. *J Biol Chem* 2014; **289**: 20234–20244.
- Gopal A, Iyer SC, Gopal U, Devaraj N, Halagowder D. *Shigella dysenteriae* modulates BMP pathway to induce mucin gene expression *in vivo* and *in vitro*. *PLoS One* 2014; **9**: e111408.
- Hamada T, Goto M, Tsutsumida H, Nomoto M, Higashi M, Sugai T et al. Mapping of the methylation pattern of the *MUC2* promoter in pancreatic cancer cell lines, using bisulfite genomic sequencing. *Cancer Lett* 2005; **227**: 175–184.
- Takahashi K. Influence of bacteria on epigenetic gene control. *Cell Mol Life Sci* 2014; **71**: 1045–1054.
- Vincent A, Perrais M, Desseyn JL, Aubert JP, Pigny P, Van Seuningen I. Epigenetic regulation (DNA methylation, histone modifications) of the 11p15 mucin genes (*MUC2*, *MUC5AC*, *MUC5B*, *MUC6*) in epithelial cancer cells. *Oncogene* 2007; **26**: 6566–6576.
- Tsuji S, Uehori J, Matsumoto M, Suzuki Y, Matsuhiwa A, Toyoshima K et al. Human intelectin is a novel soluble lectin that recognizes galactofuranose in carbohydrate chains of bacterial cell wall. *J Biol Chem* 2001; **276**: 23456–23463.
- Pemberton AD, Knight PA, Gamble J, Colledge WH, Lee JK, Pierce M et al. Innate BALB/c enteric epithelial responses to *Trichinella spiralis*: inducible expression of a novel goblet cell lectin, intelectin-2, and its natural deletion in C57BL/10 mice. *J Immunol* 2004; **173**: 1894–1901.
- Wrackmeyer U, Hansen GH, Seya T, Danielsen EM. Intelectin: a novel lipid raft-associated protein in the enterocyte brush border. *Biochemistry* 2006; **45**: 9188–9197.
- Jeong HG, Satchell KJ. Additive function of *Vibrio vulnificus* MARTX(Vv) and VvhA cytolysins promotes rapid growth and epithelial tissue necrosis during intestinal infection. *PLoS Pathog* 2012; **8**: e1002581.
- Lee JH, Kim MW, Kim BS, Kim SM, Lee BC, Kim TS et al. Identification and characterization of the *Vibrio vulnificus* rtxA essential for cytotoxicity *in vitro* and virulence in mice. *J Microbiol* 2007; **45**: 146–152.
- Finkelstein RA, Boesman-Finkelstein M, Chang Y, Hase CC. *Vibrio cholerae* hemagglutinin/ protease, colonial variation, virulence, and detachment. *Infect Immun* 1992; **60**: 472–478.
- Jeong KC, Jeong HS, Rhee JH, Lee SE, Chung SS, Starks AM et al. Construction and phenotypic evaluation of a *Vibrio vulnificus* vvpE mutant for elastolytic protease. *Infect Immun* 2000; **68**: 5096–5106.
- Miyoshi S, Wakae H, Tomochika K, Shinoda S. Functional domains of a zinc metalloprotease from *Vibrio vulnificus*. *J Bacteriol* 1997; **179**: 7606–7609.
- Kim SM, Park JH, Lee HS, Kim WB, Ryu JM, Han HJ et al. LuxR homologue SmcR is essential for *Vibrio vulnificus* pathogenesis and biofilm detachment, and its expression is induced by host cells. *Infect Immun* 2013; **81**: 3721–3730.

20. Lesuffleur T, Barbat A, Dussaulx E, Zweibaum A. Growth adaptation to methotrexate of HT-29 human colon carcinoma cells is associated with their ability to differentiate into columnar absorptive and mucus-secreting cells. *Cancer Res* 1990; **50**: 6334–6343.
21. Zhang C, Hu JJ, Xia M, Boini KM, Brimson C, Li PL. Redox signaling via lipid raft clustering in homocysteine-induced injury of podocytes. *Biochim Biophys Acta* 2010; **1803**: 482–491.
22. Wu WS, Wu JR, Hu CT. Signal cross talks for sustained MAPK activation and cell migration: the potential role of reactive oxygen species. *Cancer Metastasis Rev* 2008; **27**: 303–314.
23. Lai Wing Sun K, Correia JP, Kennedy TE. Netrins: versatile extracellular cues with diverse functions. *Development* 2011; **138**: 2153–2169.
24. Lu R, Wang X, Chen ZF, Sun DF, Tian XQ, Fang JY. Inhibition of the extracellular signal-regulated kinase/mitogen-activated protein kinase pathway decreases DNA methylation in colon cancer cells. *J Biol Chem* 2007; **282**: 12249–12259.
25. Okudaira K, Kakar S, Cun L, Choi E, Wu Decamillis R, Miura S et al. MUC2 gene promoter methylation in mucinous and non-mucinous colorectal cancer tissues. *Int J Oncol* 2010; **36**: 765–775.
26. Hor LI, Chang YK, Chang CC, Lei HY, Ou JT. Mechanism of high susceptibility of iron-overloaded mouse to *Vibrio vulnificus* infection. *Microbiol Immunol* 2000; **44**: 871–878.
27. Gutierrez-Jimenez J, Arciniega I, Navarro-Garcia F. The serine protease motif of Pic mediates a dose-dependent mucolytic activity after binding to sugar constituents of the mucin substrate. *Microb Pathog* 2008; **45**: 115–123.
28. Pillai L, Sha J, Erova TE, Fadl AA, Khajanchi BK, Chopra AK. Molecular and functional characterization of a ToxR-regulated lipoprotein from a clinical isolate of *Aeromonas hydrophila*. *Infect Immun* 2006; **74**: 3742–3755.
29. Silva AJ, Pham K, Benitez JA. Haemagglutinin/protease expression and mucin gel penetration in El Tor biotype *Vibrio cholerae*. *Microbiology* 2003; **149**: 1883–1891.
30. Olivier V, Queen J, Satchell KJ. Successful small intestine colonization of adult mice by *Vibrio cholerae* requires ketamine anesthesia and accessory toxins. *PLoS One* 2009; **4**: e7352.
31. Lee JH, Rhee JE, Park U, Ju HM, Lee BC, Kim TS et al. Identification and functional analysis of *vibrio vulnificus* SmcR, a novel global regulator. *J Microbiol Biotechnol* 2007; **17**: 325–334.
32. Jones MK, Oliver JD. *Vibrio vulnificus*: disease and pathogenesis. *Infect Immun* 2009; **77**: 1723–1733.
33. Miyoshi S. *Vibrio vulnificus* infection and metalloprotease. *J Dermatol* 2006; **33**: 589–595.
34. Tsuji S, Yamashita M, Hoffman DR, Nishiyama A, Shinohara T, Ohtsu T et al. Capture of heat-killed *Mycobacterium bovis* bacillus Calmette-Guerin by intelectin-1 deposited on cell surfaces. *Glycobiology* 2009; **19**: 518–526.
35. Voehringer D, Stanley SA, Cox JS, Completo GC, Lowary TL, Locksley RM. *Nippostrongylus brasiliensis*: identification of intelectin-1 and -2 as Stat6-dependent genes expressed in lung and intestine during infection. *Exp Parasitol* 2007; **116**: 458–466.
36. Li PL, Zhang Y, Yi F. Lipid raft redox signaling platforms in endothelial dysfunction. *Antioxid Redox Signal* 2007; **9**: 1457–1470.
37. Yang CS, Lee JS, Rodgers M, Min CK, Lee JY, Kim HJ et al. Autophagy protein Rubicon mediates phagocytic NADPH oxidase activation in response to microbial infection or TLR stimulation. *Cell Host Microbe* 2012; **11**: 264–276.
38. Fassino S, Svrakic D, Abbate-Daga G, Leombruni P, Amianto F, Stanic S et al. Anorectic family dynamics: temperament and character data. *Compr Psychiatry* 2002; **43**: 114–120.
39. Kusumoto K, Kawahara T, Kuwano Y, Teshima-Kondo S, Morita K, Kishi K et al. Ecabet sodium inhibits *Helicobacter pylori* lipopolysaccharide-induced activation of NADPH oxidase 1 or apoptosis of guinea pig gastric mucosal cells. *Am J Physiol Gastrointest Liver Physiol* 2005; **288**: G300–G307.
40. van der Goot FG, Harder T. Raft membrane domains: from a liquid-ordered membrane phase to a site of pathogen attack. *Semin Immunol* 2001; **13**: 89–97.
41. Fasano A. Toxins and the gut: role in human disease. *Gut* 2002; **50**: III9–III14.
42. Lee SH, Lee YJ, Han HJ. Effect of arachidonic acid on hypoxia-induced IL-6 production in mouse ES cells: Involvement of MAPKs, NF- $\kappa$ B, and HIF-1 $\alpha$ . *J Cell Physiol* 2010; **222**: 574–585.
43. Lee SH, Na SI, Heo JS, Kim MH, Kim YH, Lee MY et al. Arachidonic acid release by H<sub>2</sub>O<sub>2</sub> mediated proliferation of mouse embryonic stem cells: involvement of Ca<sup>2+</sup>/PKC and MAPKs-induced EGFR transactivation. *J Cell Biochem* 2009; **106**: 787–797.
44. Brandt S, Wessler S, Hartig R, Backert S. *Helicobacter pylori* activates protein kinase C  $\delta$  to control Raf in MAP kinase signalling: role in AGS epithelial cell scattering and elongation. *Cell Motil Cytoskeleton* 2009; **66**: 874–892.
45. Ki MR, Lee HR, Goo MJ, Hong IH, Do SH, Jeong DH et al. Differential regulation of ERK1/2 and p38 MAP kinases in VacA-induced apoptosis of gastric epithelial cells. *Am J Physiol Gastrointest Liver Physiol* 2008; **294**: G635–G647.
46. Malladi V, Shankar B, Williams PH, Balakrishnan A. Enteropathogenic *Escherichia coli* outer membrane proteins induce changes in cadherin junctions of Caco-2 cells through activation of PKC $\alpha$ . *Microbes Infect* 2004; **6**: 38–50.
47. Gagnon M, Zihler Berner A, Chervet N, Chassard C, Lacroix C. Comparison of the Caco-2, HT-29 and the mucus-secreting HT29-MTX intestinal cell models to investigate *Salmonella* adhesion and invasion. *J Microbiol Methods* 2013; **94**: 274–279.
48. Lee SJ, Jung YH, Oh SY, Yun SP, Han HJ. Melatonin enhances the human mesenchymal stem cells motility via melatonin receptor 2 coupling with G $\alpha$ q in skin wound healing. *J Pineal Res* 2014; **57**: 393–407.
49. Cox B, Emili A. Tissue subcellular fractionation and protein extraction for use in mass-spectrometry-based proteomics. *Nat Protoc* 2006; **1**: 1872–1878.
50. Clark JA, Doelle SM, Halpern MD, Saunders TA, Holubec H, Dvorak K et al. Intestinal barrier failure during experimental necrotizing enterocolitis: protective effect of EGF treatment. *Am J Physiol Gastrointest Liver Physiol* 2006; **291**: G938–G949.
51. Dunn AK, Millikan DS, Adin DM, Bose JL, Stabb EV. New rfp- and pES213-derived tools for analyzing symbiotic *Vibrio fischeri* reveal patterns of infection and lux expression *in situ*. *Appl Environ Microbiol* 2006; **72**: 802–810.



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