Contents lists available at ScienceDirect

Saudi Journal of Biological Sciences

journal homepage: www.sciencedirect.com

Original article Seed viability of five wild Saudi Arabian species by germination and X-ray tests

B.A. Al-Hammad *, B.S. Al-Ammari

Biology Department, College of Science and Humanities Studies, Prince Sattam Bin Abdulaziz University, Al-Kharj, Box 292, Riyadh 11942, Saudi Arabia

ARTICLE INFO

Article history: Received 23 August 2016 Revised 8 January 2017 Accepted 10 April 2017 Available online 12 April 2017

Keywords: Seed viability Seed germination Germination test X-ray test Dormancy Wild Saudi Arabian plant species

ABSTRACT

Our objective was to evaluate the usefulness of the germination vs. the X-ray test in determining the initial viability of seeds of five wild species (Moringa peregrina, Abrus precatorius, Arthrocnemum macrostachyum, Acacia ehrenbergiana and Acacia tortilis) from Saudi Arabia. Usually several days were required to determine the viability of all five species via germination tests. However, X-ray test will give immediate results on filled/viable seeds. Seeds of all species, except Acacia ehrenbergiana and Acacia tortilis showed high viability in both germination (96-72% at 25/15 °C, 94-70% at 35/25 °C) and X-ray (100-80%) test. Furthermore, there was a general agreement between the germination (19%, 14% at 25/15 °C and 17% and 12% at 35/25 °C) and X-ray (8%, 4%) tests in which seed viability of Acacia ehrenbergiana and Acacia tortilis was very low due to insect damaged embryo as shown in X-ray analysis. Seeds of Abruspreca torius have physical dormancy, which was broken by scarification in concentrated sulfuric acid (10 min), and they exhibited high viability in both the germination (83% at 25/15 °C and 81% at 35/25 °C) and X-ray (96%) tests. Most of the nongerminated seeds of the five species except those of Acacia ehrenbergiana and Acacia tortilis, were alive as judged by the tetrazolium test (TZ). Thus, for the five species examined, the X-ray test was proved to be a good and rapid predictor of seed viability. © 2017 Production and hosting by Elsevier B.V. on behalf of King Saud University. This is an open access

article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

The information about seed viability is very significant for the farmer and for the conservation of the seeds in the gene bank (Dhatchanamoorthy et al., 2016; David et al., 2016). Thus, several tests for seed viability have been carried out for testing the seed viability, such as germination, cutting, embryo excision, hydrogen peroxide, indigo carmine staining, tetrazolium staining and X-raying (Karrfalt, 2004; Antonisamy et al., 2015). All these tests except the X-ray test takes several days or weeks to complete, i. e. before the viability of the seeds is known. The X-ray method has been used in the assessment of seed viability of several species such as in Pinus spp. (Sahlen et al., 1995; David et al., 2016), corn,

* Corresponding author.

ELSEVIER

E-mail addresses: b.alhammad@psau.edu.sa, Allhammadb@gmail.com (B.A. Al-Hammad), Alammari.rr@gmail.com (B.S. Al-Ammari). Peer review under responsibility of King Saud University.

http://dx.doi.org/10.1016/j.sjbs.2017.04.004

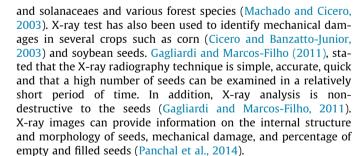
1319-562X/© 2017 Production and hosting by Elsevier B.V. on behalf of King Saud University.

This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).









Germination is a critical stage in the life cycle of the plants in general and particularly so in extreme environments such as desert and light and temperature are major factors that affect seed germination of desert and semi-desert plants. In contrast to X-ray analysis, seed germination tests are time consuming to determine seed viability, taking anywhere from a couple of weeks to a couple of years, depending on the seed dormancies involved. For example, seeds of Pinus species required at least three weeks to germinate (ISTA, 1985). Furthermore, a tetrazolium test (TZ) of seeds that fail to germinate takes at least another 24 h, e.g. Francoeuria crispa (Asteraceae), and Aeluropus massauensis (Poaceae), and it may be difficult if the TZ solution fails to penetrate some seeds or parts

Production and hosting by Elsevier

of seeds. Thus, although X-ray radiography provides a quick test for seed viability, it has also been used to determine maturity (viability) (Shen and Odén, 1999) and to predict early growth (germination) (Goodman et al., 2005) of tree seeds. However, no information is available on the relative benefits of using the germination vs. the X-ray test to evaluate seed viability of wild species from Saudi Arabia. Thus, the aim of this study was to compare the germination and X-ray test for determining seed viability of five species *Moringa peregrina*, *Abrus precatorius*, *Arthrocnemum macrostachyum*, *Acacia ehrenbergiana* and *Acacia tortilis* from Saudi Arabia.

2. Materials and methods

2.1. Study species

Five species Moringa peregrina, Abruspreca torius, Arthrocnemum macrostachyum, Acacia ehrenbergiana and Acacia tortilis with different uses were selected for study (Table 1).

2.2. Seed collection

Mature seeds Moringa peregrina, Abruspreca torius, Arthrocnemum macrostachyum, Acacia ehrenbergiana and Acacia tortilis were collected in Saudi Arabia at the location and on the date given in Table 2. The seeds were taken directly from at least 50 healthy plant individuals per species. Seeds were air dried, cleaned and stored in brown paper bags at room temperature (22 °C) for three weeks and then examined immediately.

2.3. Seed viability testing

Seed viability of the five species was determined using the germination test and the X-ray test.

2.4. Germination test

Seed germination tests were conducted using 9-cm Petri dishes containing two layers of filter paper (Whatman no. 1) moistened with 10 ml of distilled water, and five replicates of 20 seeds each for each species were used. Prior to the germination test, the water-impermeable seeds of *Abruspreca torius* were soaked in concentrated sulfuric acid (H₂SO₄) for 10 min to break dormancy. Petri dishes were randomly distributed in temperature-controlled incubators and their position was changed daily. Germination was

Table 1

The study species and their uses.

defined as the first emergence of the radicle (Sanjeewani et al., 2014). Newly-germinated seeds were counted each day for 30 d and subsequently removed from the Petri dishes. Seeds were incubated in a daily photoperiod (12 h light: 12 h dark) at alternating temperature regimes of 25/15 °C and 35/25 °C that simulate possible diurnal temperature fluctuations in the habitats of the eight species. At the end of the germination tests, nongerminated seeds were tested for viability using 2,3,5-triphenyl tetrazolium chloride (TTC) solution, as described by the International Seed Testing Association (1999). The seeds were soaked in 1% TTC solution for 4 days in a glass vial in the dark at 25 °C, and a red stained embryo was used as an indication of seed viability. The final germination percentage (%) was expressed as G (%) = $(A/B) \times 100$ (Panchal et al., 2014), where A is the total number of seeds germinated at the end of experiment (30 d) and B is the total number of seeds tested (100 seeds). Germination speed ($50\% = t_{50}$) was calculated according to Maguire (1962) as $GSI = G_1/N_1 + G_2/N_2 + \cdots + G_n/N_n$, where G_1 , G_2 , G_n are the number of germinated seeds and N_1 , N_2 , N_n the number of days.

2.5. X- ray radiography test

Two samples of 2 replications of 50 seeds of each species were radiographed with the aid of digital equipment (Faxitron X-ray brand, model MX-20 DC12) connected to a computer. The seeds were exposed to 18 KV/10 s. The X-ray plates were evaluated based on the presence and morphology of the embryo and endosperm. The percentage of seeds with a whole embryo, damaged embryo or no embryo was determined.

2.6. Statistical analysis

For each germination test, the results were expressed as the mean percentage \pm standard error, which were subjected to the *t*-test. *T*-statistics and probabilities indicate significance differences between treatments. Data of X-ray analysis were not statistically analyzed.

3. Results

3.1. Seed viability

3.1.1. Germination test

The germination percentage of all five species was higher at 25/15 °C than at 35/25 °C (Fig. 1). Regardless of the test tempera-

Species	Family	Uses	Reference(s)
Moringa peregrina	Moringaceae	Nutritional and medicinal properties	Elsayed et al. (2016)
Abrus precatorius	Fabaceae	Toxic seeds, the leaves and used to treat fevers coughs and colds	Martinez et al. (2012)
Arthrocnemum macrostachyum	Amaranthaceae	Possesses high quality of edible oil having unsaturation ranging from 70–80%	Weber et al. (2007)
Acacia ehrenbergiana	Fabaceae	Medicinal and economic and Grazing plant	Javed et al. (2013)
Acacia tortilis	Fabaceae	The seeds are ingested by birds	Jaouadi et al. (2015)

Table 2

Seed collection location and date of the five study species.

No	Species	Family	Collection-date	Location
1	Moringa peregrina	Moringaceae	13 January 2014	Al-Ula (317 South of Tabuk Region)
2	Abrus precatorius	Fabaceae	17 January 2014	Jabal Shada (Al-Baha Region)
3	Arthrocnemum macrostachyum	Amaranthaceae	8 June 2015	Darin Island (Arabian Gulf Coast)
4	Acacia ehrenbergiana	Fabaceae	12 October 2013	Al-Thummamah (55 km North–East of Riyadh)
5	Acacia tortilis	Fabaceae	11 October 2013	Al-Thummamah (55 km North-East of Riyadh)

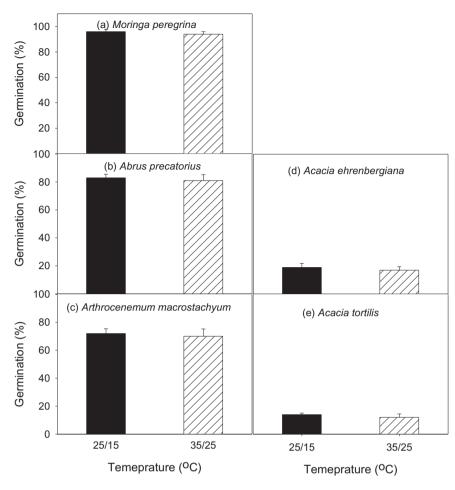


Fig. 1. The final germination percentages (mean ± Se) of (a) Moringa peregrina, (b) Abrus precatorius, (c) Arthrocnemum macrostachyum, (d) Acacia ehrenbergiana and (e) Acacia tortilis at two alternating temperatures.

Table 3

Time (days) taken to achieve 50% germination (t $_{50}$) of the five species at two alternating temperatures (12 h light/12 h dark).

Species	Alternating temperature (°C)		
	25/15	35/25	
Moringa peregrina	3.5	1	
Abrus precatorius	3.5	2	
Arthrocnemum macrostachyum	4	2	
Acacia ehrenbergiana	8	4	
Acacia tortilis	8	4	

Table 4

Fate of seeds that did not germinate at 25/15 and 35/25 °C (12 h light/12 h dark) Proportions (%) of original number germinating, remaining dormant and dead (as judged by the tetrazolium test) seeds of the in five species.

Species	Alternating temperature	Dormant	Dead
	(°C)	(%)	(%)
Moringa peregrina	25/15	4	0
	35/25	6	0
Abrus precatorius	25/15	10	7
	35/25	12	7
Arthrocnemum	25/15	20	8
macrostachyum	35/25	17	13
Acacia ehrenbergiana	25/15	1	80
	35/25	3	80
Acacia tortilis	25/15	0	86
	35/25	0	88

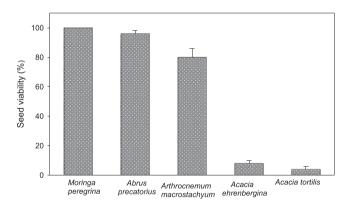


Fig. 2. Seed viability percentages (mean $\pm se$) of five wild species as judged by the X-ray test. ($\pm se = Standard error$).

ture, seeds of all species, except Acacia tortilis, Acacia ehrenbergiana germinated to \geq 70%, while those of Acacia ehrenbergiana, Acacia tortilis, germinated to only19%, 14% at 25/15 °C and 17% and 12% at 35/25 °C. Thus, temperature had no significant effect (P > 0.05) on germination of any of the species. Seeds of all species started to germinate after 3–4 days at both temperatures (25/15 °C and 35/25 °C). The **t**₅₀ for seeds of Acacia ehrenbergiana and Acacia tortilis was quite slow (8 days at 25/15 °C) (Table 3). In contrast, seeds of Moringa peregrina, Abruspreca torius and Arthrocnemum macrostachyum reached 50% within 3.5, 5 and 6 days, respectively, at 25/15 °C (Table 3). The **t**₅₀ of all five species decreased with

increasing temperature to 1–5 days at 35/25 °C (Table 3). The tetrazolium viability test (TZ) revealed that most non-germinated seeds of *Acacia ehrenbergiana*, *Acacia tortilis* (80%, 86% at 25/15 °C and 80% and 88% at 35/25 °C) were dead (Table 4). In contrast, most of the non-germinated seeds of the other species were alive (Table 4). The seeds of *Abruspreca torius* have physical dormancy (water-impermeable seed coat) that was broken by sulfuric acid, and the germination test showed very high viability of seeds (83% at 25/15 °C and 81% at 35/25 °C) (Fig. 1).

3.2. X-ray radiography test

Exposure of seeds to 18 V radiation for 10 s enabled clear visualization of the embryo and endosperm. Based on embryo

morphology as seen by X-ray radiography, 100%, 96% and 80%, of the seeds of *Moringa peregrina*, *Abrus precatorius*, and *Arthrocnemum macrostachyum*, respectively were viable(Figs. 2 and 3a–c), while only 8% and 4% of *Acacia ehrenbergiana* and *Acacia tortilis* seeds were viable (Figs. 2 and 3d and e). Endosperm was not present in seeds of *Arthrocnemum macrostachyum* (Fig. 3c).

4. Discussion

The results show that seeds of *Moringa peregrina* and *Arthrocnemum macrostachyum* do not appear to have genetically fixed (innate) mechanism of dormancy since untreated seeds samples from several plants had 96%, 72% germination at 25/15 °C, and

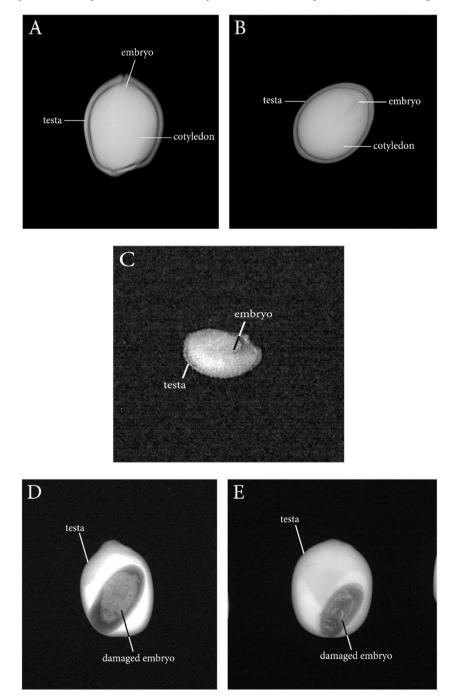


Fig. 3. X-ray photograph of Moringa peregrina seeds (A) and Abrus precatorius seeds (B) showing, testa, embryo and cotyledon (X5). X-ray photograph of Arthrocnemum macrostachyum seeds (C), Acacia ehrenbergiana seeds (D) and Acacia tortilis seeds (E) showing, testa, embryo, cotyledon and damaged embryo (X5).

94%, 70% germination at 35/25 °C respectively (Fig. 1). The seeds of Abruspreca torius have physical dormancy (water-impermeable seed coat) that was broken by sulfuric acid, and the final germination percentage (%) ranged between 83% at 25/15 °C and 81% at 35/25 °C (Fig. 1). Acacia ehrenbergiana and Acacia tortilis seeds showed very low germination (Fig. 1). Previous studies showed that seeds of different populations of Moringa peregrina collected from Egypt germinated easily (>94%) (Gomaa and Picó, 2011). Our study about seed viability of Acacia ehrenbergiana and Acacia tortilis confirms the earlier works of El Atta (1993). Who found that the embryo of Acacia nilotica from sudan was damaged by larvae of Caryedon serratus Olivier. Loth et al. (2005), showed that the optimum temperature of seed germination of Acacia tortilis from Tanzania ranged between 21 and 23 °C and the seeds of this species that had absorbed water lost their viability when kept above 35.5 °C.

In fact that both the germination and X-ray tests indicated that a high percentage of the seeds of Moringa peregrina, Abrus precatorius, and Arthrocnemum macrostachyum was viable (Figs. 1 and 2) while only a low percentage of the seeds of Acacia ehrenbergiana and Acacia tortilis was viable. As revealed by X-ray, the embryo in most seeds of Acacia ehrenbergiana and Acacia tortilis was damaged (Fig. 3d and e). Reliable results of both germination and X-ray test concerning seed viability for the five species agreed completely with results from previous studies. Further, using X-ray, Ferguson and Tuner (1971) found that the low germination of cotton seeds was due to damage that seeds sustained during harvest. Shen and Odén (1999) also found that X-raying seeds is an effective way to detect embryo damage and determine if seeds are filled. Although the X-ray analysis for seeds of the five species was very fast (few seconds), simple and accurate, the germination test was relatively slow but nonetheless accurate. The seeds required 2-4 days to start germination, and the speed of germination represented by t₅₀ varied with the species and test temperature (Table 3). The time required for reaching 50% germination of Moringa peregrina, Abrus precatorius, and Arthrocnemum macrostachyum was 3.5 days at 25/15 °C and 1-2 days at 35/25 °C. Seeds of Acacia ehrenbergiana and Acacia tortilis had a t₅₀ of 8 days at 25/15 °C, while the t₅₀ for these two species was decreased to 4 days at 35/25 °C A Similar result was reported by Gomaa and Picó (2011), who found that seeds from different populations of Moringa peregrina required 3 days to start germination. Also, Chanyenga et al. (2012) reported that the seeds of Widdringtonia whytei required about 16 days to start germination at 20 °C and 15/25 °C and 21 days to start germination at 15 °C and 10/20 °C. On the other hand, Vitis et al. (2014) reported that the t_{50} of Malcolmia littorea (Brassicaceae) seeds decreased with increasing temperatures (9 days at 5 °C and 2 days at 25 °C). The speed of germination (t50) of different populations of several species of Amaranthaceae from Saudi Arabia (e.g. Salicornia europaea agg., Suaeda aegyptiaca, Suaeda maritima, Suaeda vermiculata and Suaeda monoica) decreased with an increase in temperature (Al-Turki, 1992).

5. Conclusions

From the present investigation, we concluded that the X-ray test was easier and quicker than the germination test for the five wild species from Saudi Arabia. In germination tests for these five species, at least 2–4 days were required for seeds to start germination, and the speed of germination (t_{50}) in all the species decreased with an increase in temperature from at 25/15 °C to 35/25 °C. Seed viability as determined by germination and X-ray tests was high for three species (*Moringa peregrina, Abrus precatorius*, and

Arthrocnemum macrostachyum) and low for two species (Acacia ehrenbergiana and Acacia tortilis). X-ray analysis showed that the embryo in most of the Acacia ehrenbergiana, Acacia tortilis seeds was damaged. Thus, while both tests give an accurate assessment of seed viability, the X-ray test gave the fastest results as well as an explanation for low viability in the case of Acacia ehrenbergiana and Acacia tortilis.

Conflict of interest statement

None declared.

References

- Antonisamy, P., Duraipandiyan, V., Ignacimuthu, S., Kim, J.-H., 2015. Anti-diarrhoeal activity of friedelin isolated from *Azima tetracantha* lam. in wistar rats. S. Ind. J. Biol. Sci. 1, 34–37.
- AL-Turki, T.A., 1992. Systematic and ecological studies of Suaeda and Salicornia from Saudi Arabia and Britain Ph.D. University of East Anglia, UK.
- Chanyenga, T.F., Geldenhuys, C.J., Sileshi, G.W., 2012. Germination response and viability of an endangered tropical conifer *Widdringtonia whytei* seeds to temperature and light. S. Afr. J. Bot. 81, 25–28.
- Cicero, S.M., Banzatto-Junior, H.L., 2003. Avaliacao do relacionamento entre danosmecanicos e vigor, emsementes de milho, pormeio da analise de imagens. Image analysis for evaluating the relationship between mechanical damage and seed vigour in maize. RevistaBrasileira de Sementes 25, 29–36.
- David, R.H.A., Ceasar, S.A., Thirugnanasambantham, K., Ignacimuthu, S., 2016. Genetic engineering of crop plants for drought tolerance: role of transcription factors. S. Ind. J. Biol. Sci. 2, 272–286.
- Dhatchanamoorthy, N., Balachandran, N., Ayyanar, M., 2016. Notes on some rare plant collections from the southern Coromandel Coast, India. S. Ind. J. Biol. Sci. 2, 256–263.
- El Atta, H.A., 1993. The effect of Caryedonserratus Olivier (Co; Bruchidea) on viability and germination of seeds of *Acacia nilotica* (L. Willd. Ex Del.) in the Sudan. For. Ecol. Manage. 57 (1–4), 169–177.
- Elsayed, E.A., Sharaf-Eldin, M.A., El-Enshasy, H.A., Wadaanl, M., 2016. In vitro assessment of anticancer properties of *Moringaperegrina* essential seed oil on different cell lines. Pakistan J. Zool. 48 (3), 853–859.
- Ferguson, D., Tuner, J.H., 1971. Influence of unfilled cotton seed upon emergence and vigor. Crop Sci. 11, 713–715.
- Gagliardi, B., Marcos-Filho, J., 2011. Relationship between germination and bell pepper seed structure assessed by the X-ray test. Sci. Agric. 86 (4), 411–416.
- Gomaa, N.H., Picó, F. Xavier, 2011. Seed germination, seedling traits, and seed bank of the tree *Moringa peregrine* (Moringaceae) in a hyper-arid environment. Am. J. Bot. 98 (6), 1024–1030.
- Goodman, R.C., Jacobs, D.F., Karrfalt, R.B., 2005. Evaluating desiccation sensitivity of Quercus rubra acorns using X-ray. Can. J. For. Res. 35, 2823–2831.
- International Seed Testing Association-ISTA, 1985. International rules for seed testing. Seed Sci. Technol. 13, 299–513.
- International Seed Testing Association, 1999. Biochemical test for viability. Seed Sci. Technol. 27 (Suppl.), 33–35.
- Jaouadi, W., Mechergui, K., Ammari, Y., Hamrouni, L., Hanana, M., Khouja, M.L., 2015. Ethnobotanical an ethnopharmacological study of Acacia tortilis (Forssk) Hayne subspraddiana (Savi) in a North African pseudo-Savanna. Phytothérapi. http://dx.doi.org/10.1007/s10298-015-0951-1.
- Javed, S.B., Anis, M., Khan, P.R., Aref, I.M., 2013. In vitro regeneration and multiplication for mass propagation of *Acacia ehrenbergiana* Hayne: a potential reclaiment of denude arid lands. Agroforest. Syst. 87, 621–629.
- Karrfalt, R.P., 2004. Seed testing. Chapter 5 In: Woody Plants Seed Manual URL. Dry Branch (GA): USDA Forest Service, National Seed Laboratory, 24p.
- Loth, P.E., de Boer, W.F., Heitkonig, M.A., Prins, H.H.T., 2005. Germination strategy of East African Savanna tree Acacia tortilis. J. Trop. Ecol. 21, 509–517.
- Machado, C.F., Cicero, S.M., 2003. AROEIRA-BRANCA. [Lithraea molleoides(Vell.) Engl. -ANACARDIACEAE] Seed quality evaluation by the X-ray test. Scientia Agricola 60, 393–397.
- Maguire, J.D., 1962. Speed of germination: aid in selection and evaluation for seedling emergence and vigour. Crop Sci. 2, 176–177.
- Martinez, N., Angélica, A., Mariana, A., Rosa, B., Edmundo, C., 2012. Toxic effects of *Abrusprecatorius* L. seeds on laboratory rats. Emir. J. Food Agric. 24 (2), 159–164.
- Panchal, K.P., Pandya, N.R., Albert, Susy., Gandhi, Dhara J., 2014. A X-ray image analysis for assessment of forage seed quality. Int. J. Plant Anim. Environ. Sci. 4, 103–109.
- Sahlen, K., Bergsten, U., Wiklund, K., 1995. Determination of viable and dead Scots pine seeds of different anatomical maturity after freezing using IDX method. Seed Sci. Technol. 23, 405–414.

1429

- Sanjeewani, B.L.G., Jayasuriya, K.M.G.G., Fernando, M.T.R., Damunupola, J.W., 2014. Storage and germination treatments for seeds of an ornamentally important Palm, *Livistona rotundifolia* (Lam.) Mart. J. Natl. Sci. Found. Sir Lanka 41 (4), 273– 277.
- Shen, T.Y., Odén, P.C., 1999. Activity of sucrose synthase, soluble acid invertase and fumarase in germinating seeds of Scots pine (*Pinus sylvestris* L.) of different quality. Seed Sci. Technol. 27, 825–838.
- Vitis, M.D., Seal, C.E., Ulian, T., Pritchard, H.W., Magrini, S., Fabrini, G., Mattana, E., 2014. Rapid adaptation of seed germination requirements of the threatened Mediterranean species *Malcolmialittorea* (Brassicaceae) and implications for its reintroduction. S. Afr. J. Bot. 94, 46–50. Weber, D.J., Ansari, R., Gul, B., Khan, M.A., 2007. Potential of halophytes as source of
- edible oil. J. Arid Environ. 68 (2), 315-321.