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Brown adipose tissue and regulation of human body weight

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Abstract

Background: Approximately 30% of the global population is affected by obesity. Traditional non-surgical measures for weight loss have limited efficacy and tolerability. Therefore, there is a need for novel, effective therapies. Brown adipose tissue (BAT) has been implicated in physiological energy expenditure, indicating that it could be targeted to achieve weight loss in humans. The use of ¹⁸F-fluorodeoxyglucose (¹⁸F-FDG) positron emission tomography—computed tomography—(PET-CT) imaging has enabled the discovery of functionally active BAT in the supraclavicular, subclavian, and thoracic spine regions of human adults. This review aims to discuss the reasons behind the renewed interest in BAT, assess whether it is metabolically important in humans, and evaluate its feasibility as a therapeutic target for treating obesity.

Sources of material: PubMed Central, Europe PMC, Medline.

Findings: In vivo studies have shown that BAT activity is regulated by thyroid hormones and the sympathetic nervous system. Furthermore, BAT uniquely contains uncoupling protein 1 (UCP1) that is largely responsible for non-shivering thermogenesis. Cold exposure can increase BAT recruitment through the browning of white adipose tissue (WAT); however, this technique has practical limitations that may preclude its use. Currently available medicines for humans, such as the β 3-adrenergic receptor agonist mirabegron or the farnesoid X receptor agonist obeticholic acid, have generated excitement, although adverse effects are a concern. Capsinoids represent a tolerable alternative, which require further investigation.

Conclusions: The use of currently available BAT-activating agents alone is unlikely to achieve significant weight loss in humans. A combination of BAT activation with physical exercise and modern, successful dietary strategies represents a more realistic option.

KEYWORDS

brown adipose tissue (BAT), capsaicin, capsinoids, cold activation, energy expenditure, farnesoid X receptor, mirabegron, obesity, thermogenesis

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1 | BACKGROUND

The global prevalence of obesity has been increasing exponentially over the last few decades. In 2014, over 2.1 billion individuals were estimated to be overweight or obese, accounting for approximately 30% of the world's population. Due to the numerous diseases associated with increased adiposity including type 2 diabetes and cancer, the economic burden of obesity is substantial and thus there is an urgent need for novel effective treatments.

Bariatric surgery is currently the most effective intervention for achieving weight loss, but it is accompanied by the costs of specialist equipment, inpatient stay, and the requirement for appropriately trained and specialised healthcare providers.² Moreover, not every obese individual is eligible for and/or chooses surgery due to perioperative and longer-term complications, and there is limited availability of bariatric surgeons.³ Therefore, there is a requirement for effective and accessible medical therapies.

Obesity is the consequence of a long-term imbalance between energy intake and energy expenditure; therefore, drugs have been developed to address this imbalance. For example, liraglutide is a glucagon-like peptide-1 (GLP1) agonist, which enhances satiety and delays gastric emptying, thus reducing calorie intake.⁴ This therapeutic agent is effective but nausea, constipation, and pancreatitis are potential side-effects.⁵ Conversely, monoamine reuptake inhibitors, such as sibutramine, cause weight loss by increasing energy expenditure, but the substantial risk of adverse cardiovascular outcomes resulted in its withdrawal from the market.⁶ Hence, there is still a need for additional safe and tolerable pharmacological agents to treat obesity.

Researchers have recently shifted their focus towards brown adipose tissue (BAT), which represents an alternative, potentially safer therapeutic target for increasing energy expenditure. Whilst white adipose tissue (WAT) is responsible for storing energy in the form of triglycerides, BAT has been implicated in physiological energy expenditure through a process called non-shivering thermogenesis. It has therefore been hypothesised that increasing BAT recruitment and activation may result in upregulated energy expenditure and consequent weight loss. Furthermore, studies suggest that WAT can be converted into thermogenic beige adipocytes in a process known as browning, which may represent an alternative therapeutic approach. This review will discuss the reasons behind the renewed interest in BAT, assess whether it is metabolically important in humans, and evaluate the feasibility of BAT activation and WAT browning in the treatment of obesity.

2 | BROWN ADIPOSE TISSUE (BAT)

BAT is distinct from WAT in both structure and function^{7,9,10}; its constituent adipocytes are characterised by numerous mitochondria that give BAT its characteristic brown colour.¹¹ These mitochondria contain uncoupling protein 1 (UCP1), which is responsible for the role of BAT in thermogenesis and increasing energy expenditure.¹¹ When

BAT is activated, fatty acids are utilised in the electron transport chain and UCP1 acts as an alternative channel to adenosine triphosphate (ATP) synthase for the passage of protons, thereby releasing the energy as heat instead of producing ATP (Figure 1).¹²

Consequently, the concept of therapeutically stimulating BAT, in order to increase energy expenditure by utilising fat, is of great interest to the scientific community. Glucose is also taken up by BAT, and therefore, enhanced BAT activation is likely to improve glucose metabolism and benefit patients with diabetes, which is a common comorbidity in obesity.¹³

Alongside white and brown adipocytes, scientists have discovered other adipocytes known as beige adipocytes (Figure 2). These cells typically exist within WAT depots, although data indicates that they may also be present alongside brown cells in BAT tissue. 14 Beige adipocytes express UCP1 in their mitochondria and thus can undertake thermogenesis. 15 Despite the similarities in the structures of brown and beige adipocytes, there are notable differences in their function. Brown adipocytes express high levels of UCP1 under normal conditions, whereas beige cells only express UCP1 in response to external stimuli, such as cold exposure or β3-adrenoceptor agonism.¹² Beige cells may also undergo thermogenesis via UCP1-independent mechanisms, such as ATP-dependent calcium cycling¹⁶ or creatine cycling.¹⁷ Additionally, the two types of cell have contrasting developmental lineages: brown adipocytes are derived from a specific subset of myogenic factor 5 positive dermatomyotome cells, whereas beige adipocytes classically arise in WAT depots and are derived from platelet-derived growth factor α positive precursors. 18,19 The key similarities and differences between brown and beige adipocytes are summarised in Table 1. It has been reported that WAT can be converted into beige adipose tissue, 8,20 which could represent an additional approach for treating obesity.

3 | BROWN FAT IN HUMAN ADULTS

Despite the implication of BAT in energy expenditure, the quantity of this tissue decreases from infancy to adulthood, and it was believed that adult humans only possessed insignificant quantities of BAT.²¹ Therefore, there was limited enthusiasm about the therapeutic activation of BAT for many years. However, recent innovations in imaging and laboratory techniques have led to fascinating discoveries about the presence and functionality of BAT in adulthood.

The gold standard imaging modality for assessing BAT activity is currently ¹⁸F-fluorodeoxyglucose (¹⁸F-FDG) positron emission tomography—computed tomography (PET-CT), which measures the uptake of ¹⁸F-FDG by metabolically active tissues.²² Through the use of this imaging technique, regions of functionally active BAT were discovered in the supraclavicular, subclavian, and thoracic spine regions of adult humans.^{23,24} Early imaging studies also observed greater levels of ¹⁸F-FDG uptake in female patients.²⁴ Although the use of ¹⁸F-FDG PET-CT imaging identified the presence of functional BAT in adults, the quantity of BAT present in children remains higher than in adults.²⁴ Subsequent studies have since corroborated these

FIGURE 1 Thermogenesis in the mitochondria of brown adipose tissue. Increased sympathetic activity and β 3-adrenoceptor stimulation drives a cAMP-dependent mechanism, resulting in glucose uptake and lipolysis of intracellular triglycerides. Thyroxine receptor stimulation enhances the effects of increased sympathetic activity. Fatty acids and glucose are utilised in the electron transport chain and activate UCP1, which uncouples the respiratory chain from the ATP synthesis by providing an alternative route for the passage of H⁺. The flow of H⁺ through UCP1 generates heat via non-shivering thermogenesis. Created with BioRender.com. ADP, adenosine diphosphate; ATP, adenosine triphosphate; cAMP, cyclic adenosine monophosphate; e⁻, electron; FADH₂, reduced flavin adenine dinucleotide; H⁺, proton; NADH, reduced nicotinamide adenine dinucleotide; UCP1, uncoupling protein 1

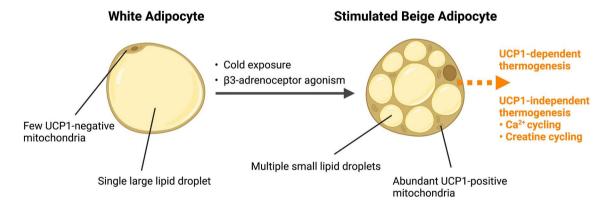


FIGURE 2 Beiging of white adipose tissue. Cold exposure and β 3-adrenoceptor agonism converts white adipocytes into stimulated beige adipocytes, which contain UCP1-positive mitochondria. Beige adipocytes can undergo thermogenesis via UCP1-dependent and UCP1-independent mechanisms. Created with BioRender.com. Ca²⁺, calcium ion; UCP1, uncoupling protein 1

findings regarding the effects of sex^{25} and age^{26} on the presence of BAT with the prevalence of detectable BAT reported to be over 2-fold greater in women than men^{25} and higher in younger age groups.²⁶

More recently, studies have discovered additional factors that appear to affect the quantity of BAT observed, including body mass index (BMI) and seasons.²⁷ ¹⁸F-FDG uptake following cold stimulation was higher in the winter compared to the summer, which may be due to the colder room temperature associated with winter.^{27,28}

There also appears to be an inverse correlation between BAT activity and BMI, as shown by van Marken Lichtenbelt $et~al.^{29}$ who found that mean BAT activity was significantly, 4-fold higher in lean participants (BMI <25) compared to overweight or obese participants (BMI \geq 25), despite non-significant differences in BAT volume. This is supported by the finding that basal and cold-induced fatty acid uptake by BAT is impaired in obesity. Leitner $et~al.^{31}$ also reported that lean men (mean BMI = 23.2) had over twice the amount of activated BAT compared to obese men (mean BMI = 34.8). Finally, Matsushita

TABLE 1 Summary of the key similarities and differences between brown and beige adipocytes

	Brown adipocytes	Beige adipocytes
Location	Present in supraclavicular, subclavian, and thoracic spine regions.	• Arise in white adipose tissue depots in response to external stimuli.
Cellular structure	Several small fat droplets.Abundant mitochondria and UCP1.	 Single fat droplet, with multiple smaller droplets after stimulation. Abundant mitochondria and UCP1 after stimulation.
Origin	Myogenic factor 5 positive precursors.	\bullet Platelet-derived growth factor α positive precursors.
Thermogenesis	UCP1-dependent.	 UCP1-dependent. UCP1-independent (Ca²⁺ cycling and creatine cycling).

Abbreviations: Ca²⁺, calcium ion; UCP1, uncoupling protein 1.

et al.²⁸ found that participants with detectable BAT had lower BMI, body fat mass, and abdominal fat area compared to those without detectable BAT. However, it should be noted that blood glucose, which may have been higher in the obese participants,²⁹ can compete with ¹⁸F-FDG and result in lower uptake measurements.³²

Although several ¹⁸F-FDG PET-CT imaging studies have been conducted to detect and quantify BAT in humans, it is difficult to perform objective comparisons. This is because these studies used different protocols, and experimental factors including the ambient temperature, the injected dose of ¹⁸F-FDG and the time from ¹⁸F-FDG injection to imaging can significantly influence the results.³³ The use of varying standard uptake value (SUV) thresholds also limits the comparability of these studies.^{23,24} In 2016, Chen *et al.*³³ proposed the standardised BARCIST 1.0 criteria, which should guide future imaging studies in order to improve comparability.

There are several limitations to the use of ¹⁸F-FDG PET-CT imaging. Firstly, this imaging technique does not conclusively detect the presence or absence of BAT; instead, it enables the active tissue to be visualised in a functional manner.³⁴ Therefore, new methods are required to identify inactive BAT in humans. Secondly, the high ionising radiation levels mean that there are ethical concerns with using this technique in children³⁴; and alternatives such as magnetic resonance imaging (MRI) could overcome this issue. Finally, the concept of ¹⁸F-FDG PET-CT imaging is based on the ability of BAT to metabolise glucose,²² but fatty acids are the main substrates for BAT.¹² Lipid tracers are also capable of detecting BAT³⁵ and this technology may help us to further understand the physiological roles of BAT.

Furthermore, the technique of ¹⁸F-FDG PET-CT imaging can yield false positives due to uptake of the tracer by muscles.²⁴ Therefore, studies that only used imaging could not conclusively prove the existence of BAT in adult humans. However, Virtanen *et al.*¹³ obtained biopsies from the tissues exhibiting high ¹⁸F-FDG uptake before quantifying UCP1 levels, and the results were highly suggestive of BAT. These transformational findings were supported by Zingaretti *et al.*,²⁶ who collected adipose tissue from the necks of surgical patients and discovered islands of UCP1-positive brown adipocytes. Taken together, ¹³, ²³, ²⁶ it can be concluded that substantial amounts of BAT are present in human adults, and that they occupy well-defined regions.

4 | BAT PHYSIOLOGY AND WEIGHT CONTROL

The discovery of active BAT in adults generated significant excitement in the scientific community. ¹³ Nevertheless, there was a need for greater understanding about BAT physiology in order to fully uncover its therapeutic potential. Firstly, assessing the contribution of BAT thermogenesis towards overall energy expenditure is required to ascertain whether BAT activation could actually achieve meaningful weight loss. Secondly, researchers have also investigated the precise mechanisms underlying BAT activation and induction to uncover specific targets for pharmaceutical interventions. These two topics are discussed below.

4.1 | Is BAT important for energy expenditure in humans?

Energy expenditure is predominantly driven by the basal metabolic rate, which is primarily dictated by body size and is responsible for up to 70% of total energy output. The remaining 30% is composed of the energy required for physical movement and thermogenesis.³⁶

To date, a variety of strategies have been used to elicit weight loss via increased energy expenditure with limited success. For example, physical exercise is a safe and natural way of increasing energy expenditure, but numerous studies have shown that the use of exercise alone results in minimal weight loss.³⁷ Thyroid hormone supplementation has also been trialled as a method of increasing the basal metabolic rate.³⁸ However, this approach is associated with the severe side effects of atrial fibrillation and osteoporosis, as well as hyperphagia that limits the weight loss.³⁸ Finally, recent studies have investigated an array of dietary strategies, including a meta-analysis by Ludwig *et al.*³⁹ showed that low carbohydrate diets initially reduce total energy expenditure before increasing it in the long term. Nevertheless, diets are notoriously difficult to sustain, and they represent an unrealistic solution for many obese individuals.⁴⁰

The contribution of BAT-specific thermogenesis to whole-body energy expenditure in humans has been postulated to be $5\%^{11}$ (70–90 kcal/day) or even less than 20 kcal/day in another study. This implies that merely activating the existing BAT may not increase energy expenditure sufficiently to achieve clinically significant weight

loss. However, converting WAT into BAT (or beige adipose tissue) to increase its contribution to whole-body energy expenditure could yield more impressive results.8 Interestingly, Chen et al.42 demonstrated that BAT volume in rodents was significantly increased following a Roux-en-Y gastric bypass, which might explain some of the weight loss benefits of bariatric surgery.

A combination of BAT activation with physical exercise and the modern, successful dietary strategies demonstrated in the DiRECT trial⁴³ may be far more successful than utilising each approach separately. Moreover, several studies have suggested that exercise itself may influence BAT activation and WAT beiging. Various exercise benefits are mediated by peroxisome proliferator-activated receptor γ co-activator 1α (PGC1-α), 44 and rodent studies have linked PGC1-α expression with the release of the hormone irisin8. In vitro and in vivo experiments showed that irisin induces UCP1 expression in white adipocytes, highlighting a link between exercise and WAT beiging.⁸ Notably, irisin was also present in human plasma at higher levels following exercise.8 In addition to WAT beiging, it has been postulated that exercise may stimulate BAT via the increased sympathetic nervous system activity that accompanies exercise, although further research is required.⁴⁵ Taken together, the use of physical exercise alongside other BAT activation approaches may have a synergistic effect on energy expenditure and weight loss.

4.2 The mechanism of BAT thermogenesis

As described above, BAT contains UCP1 that can be activated by fatty acids or glucose (Figure 1).13 UCP1 allows protons to bypass ATP synthase and uncouples oxidative phosphorylation from ATP synthesis, thus generating heat instead of ATP in a process called thermogenesis. 12 This process fluctuates in response to different stimuli: cold-induced thermogenesis maintains a normal body temperature.²⁹ whereas diet-induced thermogenesis compensates for overfeeding.11

Furthermore, rodent BAT expresses β3-adrenergic receptors (β3) and thyroid hormone receptors with studies demonstrating that the sympathetic nervous system acts synergistically with thyroid hormones to modulate the BAT activity. 46 β3-adrenoceptor stimulation by noradrenaline drives a cAMP-dependent mechanism that breaks down intracellular triglycerides to release free fatty acids, which act as a substrate for UCP1.⁴⁷ Another effect of sympathetic stimulation is the increased expression of the enzyme deiodinase type II, which upregulates the availability of local thyroid hormones.46 Conversely, the binding of thyroxine to thyroid hormone receptors enhances the effects of \(\beta \)-adrenoceptor stimulation on the BAT activity. 46 Notably, thyroid hormones have also been shown to induce WAT browning in mice, independently of the sympathetic nervous system, although the resultant beige adipose tissue did not demonstrate increased thermogenesis due to insufficient adrenergic stimulation.48

Alternative mechanisms for the regulation of BAT activation have been proposed. Studies by Schreiber et al.⁴⁹ and Shin et al.⁵⁰ unexpectedly showed that the intracellular production of free fatty acids by brown adipocytes driven by noradrenaline may not be essential for UCP1 activation. Therefore, one alternative model suggests that UCP1 is activated by external fatty acids derived from WAT instead; however, this model does not explain the functional significance of the sympathetic innervation of BAT.51 Another proposal is that there is an unknown, non-fatty acid, direct activator of UCP1 that is released following \(\beta \)-adrenoceptor stimulation.\(\beta \)1 Further research is required to evaluate the plausibility of these alternative mechanisms.

Research has provided further insight into the molecular biology of BAT. For example, Puigserver et al. 52 showed that nuclear receptors like peroxisome-proliferator-activated receptor y (PPARy) work with the PPARy coactivator 1a (PGC1a) to induce BAT thermogenesis. It may be possible to target these downstream targets in the future to enhance BAT activation. Meanwhile, Seale et al.53 identified PRDM16 as a key transcriptional regulator, which promotes the formation of brown adipocytes. Therefore, PRDM16 could serve as a promising therapeutic target for increasing the abundance of BAT.

5 | BAT AS A THERAPEUTIC TARGET FOR **WEIGHT LOSS**

Several studies have shown that cold exposure can increase BAT recruitment and energy expenditure in humans, 13,27,29,54 but it is unclear whether this would lead to significant weight loss. Three of these experiments were conducted by exposing participants to mildly cold temperatures of 16°C²⁹ and 19°C²⁷ for 2 h. Virtanen et al.¹³ exposed patients to a room temperature of 17-19°C for 2 h, and additionally, placed the subjects' feet in ice water during the scans. Similarly, participants in the study by Yoneshiro et al.⁵⁴ experienced 2-h cold exposure at 19°C whilst intermittently placing their legs on ice blocks. All 4 studies showed that cold exposure resulted in increased ¹⁸F-FDG uptake, ^{13,27,29,54} suggesting an increase in BAT recruitment, although this effect was diminished in older participants.²⁷ Moreover, a positive correlation was identified between BAT activity as per ¹⁸F-FDG uptake and the measured rise in energy expenditure following cold exposure, implying that BAT is a key contributor.54

To achieve clinically significant weight loss, cold-induced BAT activation would be required for longer than the 2-h periods described above. A 2016 study demonstrated that BAT can be reactivated and recruited in obese people by a 10-day period of cold acclimation, which involved exposure to 14-15°C temperatures for 6-h periods daily between days 3 and 10.55 Notably, there were consequent metabolic benefits observed including enhanced glucose uptake.⁵⁵ Longer studies have also been conducted: Lee et al.⁵⁶ demonstrated that a 1-month period of overnight temperature acclimation at 19°C significantly enhanced diet-induced thermogenesis and postprandial insulin sensitivity. 6 weeks of 2-h cold exposure at 17°C was trialled by Yoneshiro et al.,57 which was associated with

increased BAT activity, and notably, an accompanying 5.2% reduction in body fat mass.

Despite the promising results to date, there are numerous limitations to these studies. 1-month⁵⁶ and 6-week⁵⁷ periods of cold exposure are still insufficient to achieve meaningful weight loss. Hence, future clinical trials should pilot the use of cold exposure for longer durations, or intense short bursts, and assess for a significant effect on weight loss. However, there may be issues with retaining participants in such trials; in the aforementioned studies of 1 month⁵⁶ and 6 weeks⁵⁷ duration, there was no dropout but only 5 and 12 subjects, respectively, underwent cold exposure, whereas participant withdrawal may become a greater issue in larger studies. Furthermore, cold exposure has been associated with an increase in free fatty acids,⁵⁸ which could be detrimental to the cardiovascular health of obese patients.⁵⁹

The use of cold exposure to recruit and activate BAT has provided valuable physiological insights but concerns about long-term tolerability and adverse effects may limit its use as a therapeutic approach for weight loss. Conversely, a variety of pharmacological approaches to target BAT have been trialled in rodents and humans, with varying levels of success, and these are discussed below.

5.1 | β3-adrenoceptor agonism

In an in vitro study, Cao *et al.* showed that β 3-agonism stimulates WAT browning by increasing UCP1 expression. The authors demonstrated that the effects of β 3 stimulation on WAT browning are mediated by the p38-MAPK signalling pathway, as these effects were not observed when a p38-MAPK inhibitor was administered. It is also important to note that β 3-agonism resulted in pleiotropic effects, including an increase in lipolysis and improved insulin sensitivity.

Mirabegron is a β3-agonist, which is currently licenced for treatment of bladder overactivity.⁶¹ Due to its presumed selectivity for β3-adrenoceptors, researchers have studied the use of mirabegron for BAT activation in rodents⁶² and humans.⁶¹ In obese mice, in vivo administration of mirabegron resulted in a 12% lower body weight, reduced adiposity and 14-fold increased gene expression of UCP1 compared to vehicle.⁶² In vitro experiments showed that mirabegron stimulates higher UCP1 expression in and browning of 3T3-L1 white preadipocytes; increased UCP1 expression was also observed in mouse brown preadipocytes.⁶² Conversely, a very modest upregulation in UCP1 was observed in BAT in vivo, 62 suggesting that UCP1 is almost maximally expressed in fully differentiated BAT and/or mirabegron does not increase BAT UCP1 expression. Calorimetry was not conducted in this study⁶² so it was not possible to quantify energy expenditure and assess its contribution to weight loss.

Clinical trials have provided conflicting results regarding the use of mirabegron. In 2015, Cypess *et al.*⁶³ found that administering mirabegron to healthy men for 12 weeks was associated with the higher BAT activity and the resting metabolic rate compared to participants receiving placebo. The authors calculated that the

observed energy expenditure could translate to the 5 kg/year weight loss required for FDA authorisation. However, the selected 200 mg/day dose is higher than the FDA-approved 50 mg/day dose (for bladder overactivity), and cardiovascular side-effects such as raised blood pressure and heart rate were observed due to off-target β 1- and/or β 2-agonism. δ 3

A more recent study corroborated these results. O'Mara et al. 64 also employed the 200 mg/day dose and found that 4-week therapy of mirabegron led to increased BAT activity and resting energy expenditure in healthy women. Again, mild cardiovascular symptoms were reported such as headaches and palpitations. 64 Taken together, 63.64 high-dose mirabegron is capable of stimulating BAT in men and women, but the side-effects indicate that an alternative with fewer off-target effects would be more suitable due to the potential for adverse cardiovascular events with longer periods of high-dose mirabegron administration. Neither study 63.64 demonstrated weight loss, but the promising effects on energy expenditure suggest the need for longer trials lasting at least 6 months, similar to other weight loss drug trials. 65

Whilst high doses of mirabegron can activate BAT in healthy humans, 64 other groups have demonstrated that the FDA-approved 50 mg/day dose did not result in a significant increase in resting energy expenditure or weight loss in obese participants after 12 weeks. 61 These results may be due to the inverse correlation between BAT and BMI. 29 However, mirabegron induced beiging of subcutaneous WAT and subsequently improved β -cell function in the insulin-resistant participants. 61 Therefore, this low dose of mirabegron should be administered for longer periods (e.g. 6 months) to assess whether adipose tissue beiging would eventually translate into meaningful weight loss.

An explanation for the modest performance of mirabegron in clinical trials to date was recently proposed by Blondin et al.⁶⁶ Here, they showed that the β2-adrenoceptor mediates BAT thermogenesis in humans.⁶⁶ unlike in rodents where β3 signalling is kev.⁶² These findings suggest that β3-agonism is unlikely to induce significant weight loss in humans, and that the high doses of mirabegron administered in the above studies^{63,64} increased BAT energy expenditure via non-selective β2-agonism. ⁶⁶ This explanation has since been challenged in a study by Cero et al.,67 where experiments involving primary cultures of human brown/beige adipocytes demonstrated that mirabegron activates BAT specifically via β3-agonism. Targeting β2-adrenoceptors could still be a more effective approach of activating BAT in humans, but pre-clinical testing would be difficult due to differential \(\beta 2\)-adrenoceptor distribution between rodents and humans. Furthermore, β2-adrenoceptors are present in multiple sites in the body, ⁶⁸ and hence, chronic β2-agonism in humans carries the risk of numerous side-effects, including tremors and tachycardia.

5.2 | Alternative approaches

Aside from $\beta 3$ -adrenoceptor agonism, other approaches have been trialled to therapeutically target BAT. Capsaicin is the active

component of chilli peppers, and it is believed to mediate the effects of chilli peppers on thermogenesis⁶⁹ by activating BAT and stimulating the browning of WAT. 70 However, capsaicin is also a chemical irritant, which is responsible for the pungency of chilli peppers, 69 and hence, it is unlikely to serve as a tolerable therapeutic. Instead, Yoneshiro et al.⁷¹ trialled the use of non-pungent capsaicin analogues called capsinoids in humans. Capsinoids lack the prominent taste of chilli peppers, so it was possible to perform a placebo-controlled trial. A small but significant elevation in energy expenditure was observed 1 h after the oral ingestion of capsinoids, but only in participants with detectable BAT.⁷¹ Therefore, capsinoids, which can be extracted from sweet chilli peppers, 72 may serve as a tolerable alternative to capsaicin for targeting BAT.

Although capsinoids may be capable of activating BAT, the findings of Yoneshiro et al. 71 indicate that they may be ineffective as a therapeutic when BAT is reduced or absent. Consequently, they may be of limited use in obesity where the presence of BAT is thought to be diminished.²⁹ However, a randomised controlled trial reported that capsinoid ingestion by participants with a mean BMI of 30.4 for 12 weeks resulted in significant abdominal fat loss. 73 Gastrointestinal adverse events, including dyspepsia and bowel disturbances, were noted, but they were not considered serious and did not result in withdrawal.⁷³ A series of in vitro and in vivo experiments then showed that capsaicin activates the transient receptor potential vanilloid 1 (TRPV1) channel, which results in the browning of WAT. 70,74 Thus, the prolonged use of capsinoids in obesity may convert WAT into beige adipose tissue, promote beige activity and increase energy expenditure to reduce abdominal adiposity.

There are additional novel therapeutics that have shown potential in rodents. Fang et al. 75 notably demonstrated that fexaramine, an intestine-specific farnesoid X receptor (FXR) agonist, exerts numerous beneficial effects on metabolism including WAT browning. Furthermore, fexaramine administration prevented dietinduced weight gain in obese mice.⁷⁵ This FXR agonist was minimally absorbed from the gut⁷⁵ (and thus may result in a lower risk of systemic toxicity), although further animal and human studies are warranted.

The metabolic effects of FXR-agonism in mice appeared to be secondary to the induction of fibroblast growth factor 15 (Fgf15) production, which has a human orthologue called FGF19, and the consequent changes to the composition of circulating bile acids. 75 In particular, chenodeoxycholic acid derivatives were present in higher proportions⁷⁵ and chenodeoxycholic acid has since been shown to activate human BAT. 76 Therefore, the findings of Fang et al. 75 may be translatable to humans and intestine-specific FXR agonism may be an effective therapeutic approach to achieve adipose tissue browning and subsequent weight loss. Currently, obeticholic acid is an example of an FXR agonist that is currently used in humans to treat primary biliary cholangitis, 77 and this agent has also been shown to stimulate BAT.⁷⁸ However, its effect on bile acids can commonly result in pruritus, as demonstrated by the PBC OCA International Study of Efficacy where over 50% of participants experienced pruritus as a

side-effect.⁷⁹ This tolerability issue may limit the use of obeticholic acid as a BAT-stimulating agent.

Ultimately, a variety of therapeutic agents have been studied to date, but there is a lack of strong evidence to suggest that a single drug can achieve meaningful weight loss in humans via BAT activation. However, combining pharmacological agents with behavioural interventions such as exercise and diets may result in greater calorie deficits and achieve better outcomes. BAT activation may also help to address some of the limitations of behavioural interventions; for example, significant weight loss from dieting is often accompanied by a slower resting metabolic rate, thus limiting the long-term effectiveness of the diet.80 Furthermore, the constrained total energy expenditure model describes the adaptation of the human body to increased physical activity by limiting energy expenditure on other physiological processes.⁸¹ The use of pharmacological agents to activate BAT could counteract these metabolic adaptations to lifestyle modifications and help sustain the benefits of physical activity and diets, although long-term clinical trials are necessary. Adverse effects are another key issue for many therapeutic agents, but strategies combining the use of multiple drugs may enable lower dosing to reduce the likelihood of severe toxicities. Nevertheless, this approach carries the risk of drug interactions, and thus further research is required.

CONCLUSION

It is evident that adult humans possess significant quantities of BAT¹³; yet the feasibility of utilising activated BAT for effecting significant weight loss in humans remains unclear. The portion of total energy expenditure that can be attributed to BAT activity in humans appears to be low, 11,41 and hence, BAT activation alone is unlikely to be a successful weight loss strategy. Additionally, BAT has been found in lean, healthy individuals but it is less detectable in the obese individuals that are the target patient group.²⁹ Further investigation is warranted to elucidate whether a lack of BAT causes an increase in weight or vice versa, in order to gain valuable insight into the pathophysiology of obesity.

As we learn more about the physiology of human BAT, therapeutics will continue to be developed and trialled. The β3-agonism approach using a currently available medicine, mirabegron, has shown the greatest potential to date. 63 However, this was associated with adverse cardiovascular side-effects due to off-target effects.⁶⁴ Furthermore, a greater understanding about the differences between rodent and human BAT will be necessary to improve upon the modest clinical trial results that have been observed so far.⁶⁶ A promising alternative strategy involves FXR agonism using obeticholic acid, although this is also limited by side-effects.⁷⁹ Ingesting non-pungent capsinoids to activate BAT may be the most tolerable approach,⁷¹ but further studies are required to assess whether clinically significant weight loss can be achieved in this manner.

The abundance of BAT in humans is inversely correlated with age²⁶ and BMI,²⁹ hence merely activating the existing BAT is unlikely

to achieve significant weight loss in the patients who need it the most. Drugs that are capable of converting WAT into beige adipose tissue may prove to be more successful in achieving meaningful weight loss, although there is still a lack of conclusive evidence as to whether this can be done safely. There are suggestions that BAT volume may be increased after bariatric surgery,⁴² and thus, BAT activation could be used to potentiate and sustain the weight loss benefits of surgery. Nevertheless, this suggestion requires confirmation in human studies. Consequently, the most realistic option for the near future is a combination of tolerable BAT-activating agents with physical exercise to maximise total energy expenditure, along-side healthy hypocaloric diets to achieve weight loss.

AUTHOR CONTRIBUTIONS

Conceptualization: Akash Srinivasan, Elissa Harb, Omar Kheder, Gishani Poopalasingam, Razi Rashid, and Chioma Izzi-Engbeaya; Writing—original draft, reviewing and editing: Akash Srinivasan, Elissa Harb, Omar Kheder, Gishani Poopalasingam, Razi Rashid, and Chioma Izzi-Engbeaya; Final approval: Akash Srinivasan, Elissa Harb, Omar Kheder, Gishani Poopalasingam, Razi Rashid, and Chioma Izzi-Engbeaya.

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CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

DATA AVAILABILITY STATEMENT

Data sharing not applicable to this article as no datasets were generated or analysed during the current study.

ETHICS STATEMENT

Not applicable.

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PEER REVIEW

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REFERENCES

 Tremmel M, Gerdtham UG, Nilsson PM, Saha S. Economic burden of obesity: a systematic literature review. Int J Environ Res Publ Health. 2017;14(4):435. https://doi.org/10.3390/ijerph14040435

- Doble B, Wordsworth S, Rogers CA, et al. What are the real procedural costs of bariatric surgery? A systematic literature review of published cost analyses. Obes Surg. 2017;27(8):2179-2192. https://doi.org/10.1007/s11695-017-2749-8
- Ahmad A, Laverty AA, Aasheim E, Majeed A, Millett C, Saxena S. Eligibility for bariatric surgery among adults in England: analysis of a national cross-sectional survey. JRSM Open. 2014;5(1):204253 3313512479. https://doi.org/10.1177/2042533313512479
- Astrup A, Carraro R, Finer N, et al. Safety, tolerability and sustained weight loss over 2 years with the once-daily human GLP-1 analog, liraglutide. *Int J Obes.* 2012;36(6):843-854. https://doi.org/10.1038/ ijo.2011.158
- Pi-Sunyer X, Astrup A, Fujioka K, et al. A randomized, controlled trial of 3.0 mg of liraglutide in weight management. N Engl J Med. 2015;373(1):11-22. https://doi.org/10.1056/NEJMoa1411892
- James WPT, Caterson ID, Coutinho W, et al. Effect of sibutramine on cardiovascular outcomes in overweight and obese subjects. N. Engl J Med. 2010;363(10):905-917. https://doi.org/10.1056/NEJMoa10 03114
- Cannon B, Nedergaard J. Brown adipose tissue: function and physiological significance. *Physiol Rev.* 2004;84(1):277-359. https://doi.org/10.1152/physrev.00015.2003
- Boström P, Wu J, Jedrychowski MP, et al. A PGC1α-dependent myokine that drives browning of white fat and thermogenesis. Nature. 2012;481(7382):463-468. https://doi.org/10.1038/nature 10777
- Rosenwald M, Wolfrum C. The origin and definition of brite versus white and classical brown adipocytes. Adipocyte. 2014;3(1):4-9. https://doi.org/10.4161/adip.26232
- Giralt M, Villarroya F. White, brown, beige/brite: different adipose cells for different functions? *Endocrinology*. 2013;154(9):2992-3000. https://doi.org/10.1210/en.2013-1403
- van Marken Lichtenbelt WD, Schrauwen P. Implications of nonshivering thermogenesis for energy balance regulation in humans. Am J Physiol Regul Integr Comp Physiol. 2011;301(2):R285-R296. https://doi.org/10.1152/ajpregu.00652.2010
- Fedorenko A, Lishko PV, Kirichok Y. Mechanism of fatty-acid-dependent UCP1 uncoupling in Brown fat mitochondria. *Cell*. 2012;151(2):400-413. https://doi.org/10.1016/j.cell.2012.09.010
- Virtanen KA, Lidell ME, Orava J, et al. Functional brown adipose tissue in healthy adults. N. Engl J Med. 2009;360(15):1518-1525. https://doi.org/10.1056/NEJMoa0808949
- Jespersen NZ, Larsen TJ, Peijs L, et al. A classical brown adipose tissue mRNA signature partly overlaps with brite in the supraclavicular region of adult humans. *Cell Metab* 2013;17(5):798-805. https://doi.org/10.1016/j.cmet.2013.04.011
- Wu J, Boström P, Sparks LM, et al. Beige adipocytes are a distinct type of thermogenic fat cell in mouse and human. *Cell*. 2012;150(2): 366-376. https://doi.org/10.1016/j.cell.2012.05.016
- Ikeda K, Kang Q, Yoneshiro T, et al. UCP1-independent signaling involving SERCA2b-mediated calcium cycling regulates beige fat thermogenesis and systemic glucose homeostasis. *Nat Med.* 2017; 23(12):1454-1465. https://doi.org/10.1038/nm.4429
- Kazak L, Chouchani ET, Jedrychowski MP, et al. A creatine-driven substrate cycle enhances energy expenditure and thermogenesis in beige fat. Cell. 2015;163(3):643-655. https://doi.org/10.1016/j.cell. 2015.09.035
- Ikeda K, Maretich P, Kajimura S. The common and distinct features of Brown and beige adipocytes. *Trends Endocrinol Metabolism*. 2018; 29(3):191-200. https://doi.org/10.1016/j.tem.2018.01.001
- Ying T, Simmons RA. The role of adipocyte precursors in development and obesity. Front Endocrinol. 2021;11. https://doi.org/10.3389/fendo.2020.613606. https://www.frontiersin.org/articles/10.3389/fendo.2020.613606

- formulation. *Dis Models Mech.* 2012;5(5):608-613. https://doi.org/10.1242/dmm.009837
- Terada E, Ashida K, Ohe K, Sakamoto S, Hasuzawa N, Nomura M. Brown adipose activation and reversible beige coloration in adipose tissue with multiple accumulations of 18F-fluorodeoxyglucose in sporadic paraganglioma: a case report. Clin Case Rep. 2019;7(7): 1399-1403. https://doi.org/10.1002/ccr3.2259
- 21. Heaton JM. The distribution of brown adipose tissue in the human. *J Anat.* 1972;112(Pt 1):35-39.
- Fischer JGW, Maushart CI, Becker AS, et al. Comparison of [18F] FDG PET/CT with magnetic resonance imaging for the assessment of human brown adipose tissue activity. EJNMMI Res. 2020;10(1):85. https://doi.org/10.1186/s13550-020-00665-7
- Hany TF, Gharehpapagh E, Kamel EM, Buck A, Himms-Hagen J, von Schulthess GK. Brown adipose tissue: a factor to consider in symmetrical tracer uptake in the neck and upper chest region. Eur J Nucl Med Mol Imag. 2002;29(10):1393-1398. https://doi.org/10.1007/ s00259-002-0902-6
- Yeung HWD, Grewal RK, Gonen M, Schöder H, Larson SM. Patterns of (18)F-FDG uptake in adipose tissue and muscle: a potential source of false-positives for PET. J Nucl Med Official Publ Soc Nucl Med. 2003;44(11):1789-1796.
- Cypess AM, Lehman S, Williams G, et al. Identification and importance of Brown adipose tissue in adult humans. N Engl J Med. 2009; 360(15):1509-1517. https://doi.org/10.1056/NEJMoa0810780
- Zingaretti MC, Crosta F, Vitali A, et al. The presence of UCP1 demonstrates that metabolically active adipose tissue in the neck of adult humans truly represents brown adipose tissue. Faseb J official Publ Fed Am Soc Exp Biol. 2009;23(9):3113-3120. https://doi.org/10.1096/fj.09-133546
- Saito M, Okamatsu-Ogura Y, Matsushita M, et al. High incidence of metabolically active brown adipose tissue in healthy adult humans: effects of cold exposure and adiposity. *Diabetes*. 2009;58(7): 1526-1531. https://doi.org/10.2337/db09-0530
- Matsushita M, Yoneshiro T, Aita S, Kameya T, Sugie H, Saito M. Impact of brown adipose tissue on body fatness and glucose metabolism in healthy humans. *Int J Obes*. 2014;38(6):812-817. https:// doi.org/10.1038/ijo.2013.206
- van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, et al. Cold-activated brown adipose tissue in healthy men. N. Engl J Med. 2009;360(15):1500-1508. https://doi.org/10.1056/NEJMoa08 08718
- Saari TJ, Raiko J, U-Din M, et al. Basal and cold-induced fatty acid uptake of human brown adipose tissue is impaired in obesity. *Sci Rep.* 2020;10(1):14373. https://doi.org/10.1038/s41598-020-71197-2
- Leitner BP, Huang S, Brychta RJ, et al. Mapping of human brown adipose tissue in lean and obese young men. Proc Natl Acad Sci U. S. A. 2017;114(32):8649-8654. https://doi.org/10.1073/pnas.170 5287114
- Waqas K, van Haard PMM, Postema JWA, Schweitzer DH. Diabetes mellitus-related fractional glucose uptake in men and women imaged with 18F-FDG PET-CT. J Endocr Soc. 2019;3(4):773-783. https://doi.org/10.1210/js.2019-00001
- Chen KY, Cypess AM, Laughlin MR, et al. Brown adipose reporting criteria in imaging STudies (BARCIST 1.0): recommendations for standardized FDG-PET/CT experiments in humans. *Cell Metab.* 2016;24(2):210-222. https://doi.org/10.1016/j.cmet.2016. 07.014
- Alcalá M, Calderon-Dominguez M, Serra D, Herrero L, Viana M. Mechanisms of impaired Brown adipose tissue recruitment in obesity. Front Physiol. 2019;10. https://doi.org/10.3389/fphys.2019.00094. https://www.frontiersin.org/articles/10.3389/fphys.2019.00094
- Paulus A, van Marken Lichtenbelt W, Mottaghy FM, Bauwens M. Brown adipose tissue and lipid metabolism imaging. *Methods*. 2017; 130:105-113. https://doi.org/10.1016/j.ymeth.2017.05.001
- Blundell JE, Caudwell P, Gibbons C, et al. Role of resting metabolic rate and energy expenditure in hunger and appetite control: a new

- Franz MJ, VanWormer JJ, Crain AL, et al. Weight-loss outcomes: a systematic review and meta-analysis of weight-loss clinical trials with a minimum 1-year follow-up. J Am Diet Assoc. 2007;107(10): 1755-1767. https://doi.org/10.1016/j.jada.2007.07.017
- Hollingsworth DR, Amatruda TT, Scheig R. Quantitative and qualitative effects of L-triiodothyronine in massive obesity. Metabolism Clin Exp. 1970;19(11):934-945. https://doi.org/10.1016/0026-0495 (70)90040-5
- Ludwig DS, Dickinson SL, Henschel B, Ebbeling CB, Allison DB. Do lower-carbohydrate diets increase total energy expenditure? An updated and reanalyzed meta-analysis of 29 controlled-feeding studies. J Nutr. 2021;151(3):482-490. https://doi.org/10.1093/jn/ nxaa350
- Buchanan K, Sheffield J. Why do diets fail? An exploration of dieters' experiences using thematic analysis. J Health Psychol. 2017;22(7): 906-915. https://doi.org/10.1177/1359105315618000
- Muzik O, Mangner T, Granneman J. Assessment of oxidative metabolism in Brown fat using PET imaging. Front Endocrinol. 2012;3. https://doi.org/10.3389/fendo.2012.00015. https://www.frontiersin. org/article/10.3389/fendo.2012.00015
- 42. Chen Y, Yang J, Nie X, Song Z, Gu Y. Effects of bariatric surgery on change of Brown adipocyte tissue and energy metabolism in obese mice. *Obes Surg.* 2018;28(3):820-830. https://doi.org/10.1007/s11 695-017-2899-8
- 43. Lean MEJ, Leslie WS, Barnes AC, et al. Durability of a primary careled weight-management intervention for remission of type 2 diabetes: 2-year results of the DiRECT open-label, cluster-randomised trial. *Lancet. Diabetes & Endocrinol.* 2019;7(5):344-355. https://doi.org/10.1016/S2213-8587(19)30068-3
- Handschin C, Spiegelman BM. The role of exercise and PGC1α in inflammation and chronic disease. *Nature*. 2008;454(7203):463-469. https://doi.org/10.1038/nature07206
- Sanchez-Delgado G, Martinez-Tellez B, Olza J, Aguilera CM, Gil Á, Ruiz JR. Role of exercise in the activation of Brown adipose tissue. Ann Nutr Metabol. 2015;67(1):21-32. https://doi.org/10.1159/00 0437173
- Sentis SC, Oelkrug R, Mittag J. Thyroid hormones in the regulation of brown adipose tissue thermogenesis. *Endocr Connect*. 2021;10(2): R106-R115. https://doi.org/10.1530/EC-20-0562
- Crichton PG, Lee Y, Kunji ERS. The molecular features of uncoupling protein 1 support a conventional mitochondrial carrier-like mechanism. *Biochimie*. 2017;134:35-50. https://doi.org/10.1016/j.biochi.20 16.12.016
- Johann K, Cremer AL, Fischer AW, et al. Thyroid-hormone-induced browning of white adipose tissue does not contribute to thermogenesis and glucose consumption. *Cell Rep* 2019;27(11):3385-3400. e3. https://doi.org/10.1016/j.celrep.2019.05.054
- Schreiber R, Diwoky C, Schoiswohl G, et al. Cold-induced thermogenesis depends on ATGL-mediated lipolysis in cardiac muscle, but not Brown adipose tissue. *Cell Metab.* 2017;26(5):753-763. e7. https://doi.org/10.1016/j.cmet.2017.09.004
- Shin H, Ma Y, Chanturiya T, et al. Lipolysis in Brown adipocytes is not essential for cold-induced thermogenesis in mice. *Cell Metab.* 2017; 26(5):764-777. e5. https://doi.org/10.1016/j.cmet.2017.09.002
- 51. Cannon B, Nedergaard J. What ignites UCP1? *Cell Metab.* 2017; 26(5):697-698. https://doi.org/10.1016/j.cmet.2017.10.012
- Puigserver P, Wu Z, Park CW, Graves R, Wright M, Spiegelman BM. A cold-inducible coactivator of nuclear receptors linked to adaptive thermogenesis. Cell. 1998;92(6):829-839. https://doi.org/10.1016/ s0092-8674(00)81410-5
- Seale P, Bjork B, Yang W, et al. PRDM16 controls a brown fat/ skeletal muscle switch. *Nature*. 2008;454(7207):961-967. https:// doi.org/10.1038/nature07182

- Yoneshiro T, Aita S, Matsushita M, et al. Brown adipose tissue, whole-body energy expenditure, and thermogenesis in healthy adult men. *Obesity*. 2011;19(1):13-16. https://doi.org/10.1038/oby. 2010.105
- Hanssen MJW, van der Lans AAJJ, Brans B, et al. Short-term cold acclimation recruits Brown adipose tissue in obese humans. *Diabetes*. 2016;65(5):1179-1189. https://doi.org/10.2337/db15-1372
- Lee P, Smith S, Linderman J, et al. Temperature-acclimated brown adipose tissue modulates insulin sensitivity in humans. *Diabetes*. 2014;63(11):3686-3698. https://doi.org/10.2337/db14-0513
- 57. Yoneshiro T, Aita S, Matsushita M, et al. Recruited brown adipose tissue as an antiobesity agent in humans. *J Clin Investigation*. 2013; 123(8):3404-3408. https://doi.org/10.1172/JCI67803
- Chondronikola M, Volpi E, Børsheim E, et al. Brown adipose tissue activation is linked to distinct systemic effects on lipid metabolism in humans. *Cell Metab.* 2016;23(6):1200-1206. https://doi.org/10.10 16/j.cmet.2016.04.029
- Zhang HW, Zhao X, Guo YL, et al. Free fatty acids and cardiovascular outcome: a Chinese cohort study on stable coronary artery disease. *Nutr Metab*. 2017;14(1):41. https://doi.org/10.1186/s12986-017-0 195-1
- Cao W, Medvedev AV, Daniel KW, Collins S. beta-Adrenergic activation of p38 MAP kinase in adipocytes: cAMP induction of the uncoupling protein 1 (UCP1) gene requires p38 MAP kinase. J Biol Chem. 2001;276(29):27077-27082. https://doi.org/10.1074/jbc.M10 1049200
- Finlin BS, Memetimin H, Zhu B, et al. The β3-adrenergic receptor agonist mirabegron improves glucose homeostasis in obese humans. J Clin Investigation. 2020;130(5):2319-2331. https://doi.org/10.11 72/JCI134892
- Hao L, Scott S, Abbasi M, et al. Beneficial metabolic effects of mirabegron in vitro and in high-fat diet-induced obese mice. *J Pharmacol Exp Ther*. 2019;369(3):419-427. https://doi.org/10.1124/jpet.118. 255778
- 63. Cypess AM, Weiner LS, Roberts-Toler C, et al. Activation of human brown adipose tissue by a β3-adrenergic receptor agonist. *Cell Metab* 2015;21(1):33-38. https://doi.org/10.1016/j.cmet.2014.12.009
- 64. O'Mara AE, Johnson JW, Linderman JD, et al. Chronic mirabegron treatment increases human brown fat, HDL cholesterol, and insulin sensitivity. *J Clin Invest*. 2020;130(5):2209-2219. https://doi.org/10. 1172/JCI131126
- Fidler MC, Sanchez M, Raether B, et al. A one-year randomized trial of lorcaserin for weight loss in obese and overweight adults: the BLOSSOM trial. J Clin Endocrinol Metab. 2011;96(10):3067-3077. https://doi.org/10.1210/jc.2011-1256
- Blondin DP, Nielsen S, Kuipers EN, et al. Human Brown adipocyte thermogenesis is driven by β2-AR stimulation. *Cell Metab* 2020; 32(2):287-300. e7. https://doi.org/10.1016/j.cmet.2020.07.005
- Cero C, Lea HJ, Zhu KY, Shamsi F, Tseng YH, Cypess AM. β3-Adrenergic receptors regulate human brown/beige adipocyte lipolysis and thermogenesis. *JCI insight*. 2021;6(11):139160. https://doi. org/10.1172/jci.insight.139160
- Krief S, Lönnqvist F, Raimbault S, et al. Tissue distribution of beta 3adrenergic receptor mRNA in man. J Clin Invest. 1993;91(1):344-349. https://doi.org/10.1172/jci116191

- Ludy MJ, Mattes RD. The effects of hedonically acceptable red pepper doses on thermogenesis and appetite. *Physiology Behav.* 2011; 102(3-4):251-258. https://doi.org/10.1016/j.physbeh.2010.11.018
- Baskaran P, Krishnan V, Ren J, Thyagarajan B. Capsaicin induces browning of white adipose tissue and counters obesity by activating TRPV1 channel-dependent mechanisms. Br J Pharmacol. 2016; 173(15):2369-2389. https://doi.org/10.1111/bph.13514
- 71. Yoneshiro T, Aita S, Kawai Y, Iwanaga T, Saito M. Nonpungent capsaicin analogs (capsinoids) increase energy expenditure through the activation of brown adipose tissue in humans. *Am J Clin Nutr.* 2012;95(4):845-850. https://doi.org/10.3945/ajcn.111.018606
- Sasahara I, Furuhata Y, Iwasaki Y, et al. Assessment of the biological similarity of three capsaicin analogs (Capsinoids) found in nonpungent chili pepper (CH-19 Sweet) fruits. Biosci, Biotechnol, Biochem. 2010;74(2):274-278. https://doi.org/10.1271/bbb.90570
- Snitker S, Fujishima Y, Shen H, et al. Effects of novel capsinoid treatment on fatness and energy metabolism in humans: possible pharmacogenetic implications. Am J Clin Nutr. 2009;89(1):45-50. https://doi.org/10.3945/ajcn.2008.26561
- Montanari T, Boschi F, Colitti M. Comparison of the effects of browning-inducing capsaicin on two murine adipocyte models. Front Physiol. 2019;10. https://doi.org/10.3389/fphys.2019.01380. https:// www.frontiersin.org/article/10.3389/fphys.2019.01380
- Fang S, Suh JM, Reilly SM, et al. Intestinal FXR agonism promotes adipose tissue browning and reduces obesity and insulin resistance. Nat Med. 2015;21(2):159-165. https://doi.org/10.1038/nm.3760
- Broeders EPM, Nascimento EBM, Havekes B, et al. The bile acid chenodeoxycholic acid increases human Brown adipose tissue activity. *Cell Metab* 2015;22(3):418-426. https://doi.org/10.1016/j.cm et.2015.07.002
- Roberts SB, Ismail M, Kanagalingam G, et al. Real-world effectiveness of obeticholic acid in patients with primary biliary cholangitis. Hepatol Commun. 2020;4(9):1332-1345. https://doi.org/10.1002/ hep4.1518
- Zhang H, Dong M, Liu X. Obeticholic acid ameliorates obesity and hepatic steatosis by activating brown fat. Exp Ther Med. 2021; 22(3):991. https://doi.org/10.3892/etm.2021.10423
- Nevens F, Andreone P, Mazzella G, et al. A placebo-controlled trial of obeticholic acid in primary biliary cholangitis. N Engl J Med. 2016;375(7):631-643. https://doi.org/10.1056/NEJMoa1509840
- Fothergill E, Guo J, Howard L, et al. Persistent metabolic adaptation 6 years after the Biggest Loser competition. *Obesity*. 2016;24(8): 1612-1619. https://doi.org/10.1002/oby.21538
- Pontzer H, Durazo-Arvizu R, Dugas L, et al. Constrained total energy expenditure and metabolic adaptation to physical activity in adult humans. Curr Biol CB. 2016;26(3):410-417. https://doi.org/10.1016/j. cub.2015.12.046

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