REVIEW



Open Access

Extracellular matrix macromolecules: potential tools and targets in cancer gene therapy

Annele Sainio¹ and Hannu Järveläinen^{1,2*}

Abstract

Tumour cells create their own microenvironment where they closely interact with a variety of soluble and non-soluble molecules, different cells and numerous other components within the extracellular matrix (ECM). Interaction between tumour cells and the ECM is bidirectional leading to either progression or inhibition of tumourigenesis. Therefore, development of novel therapies targeted primarily to tumour microenvironment (TME) is highly rational. Here, we give a short overview of different macromolecules of the ECM and introduce mechanisms whereby they contribute to tumourigenesis within the TME. Furthermore, we present examples of individual ECM macromolecules as regulators of cell behaviour during tumourigenesis. Finally, we focus on novel strategies of using ECM macromolecules as tools or targets in cancer gene therapy in the future.

Keywords: Extracellular matrix, Macromolecules, Tumour microenvironment, Cancer, Gene therapy

Introduction

Normally, the extracellular matrix (ECM) is composed of a dynamic 3D network of macromolecules, particularly collagens, elastin, proteoglycans (PGs) and hyaluronan (HA), and other non-collagenous matrix glycoproteins [1,2]. In the ECM there are also ECM degrading enzymes as well as a variety of soluble factors such as growth factors, chemokines and cytokines [1]. Furthermore, within the ECM there are several cell types including fibroblasts, adipocytes, epithelial and endothelial cells as well as different immune cells [3]. The principal function of the ECM is to maintain normal architecture and homeostasis of a particular tissue. The composition of the ECM is unique to each tissue and it undergoes constant enzymatic and non-enzymatic modifications and remodeling processes through a biophysical dialogue between its components [4]. These modifications and remodeling processes result in versatile microenvironments, "niches", which in turn vitally regulate the behaviour of the cells within the ECM [5].

In cancer, the malignant cells are known to create their own tumour microenvironment (TME) which crucially affects both the malignant cells themselves and all other

²Division of Endocrinology, Department of Medicine, Turku University Hospital, Kiinamyllynkatu 4-8, Fl-20520 Turku, Finland





© 2014 Sainio and Järveläinen; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.

^{*} Correspondence: hannu.jarvelainen@utu.fi

¹Department of Medical Biochemistry and Genetics, University of Turku, Turku, Finland

Apart from the ECM macromolecules, growth factors and cytokines mentioned above, there are other essential groups of molecules regulating tumour initiation and progression. For example, overexpression of ECM degrading enzymes such as matrix metalloproteinases (MMPs) can promote tumourigenesis. Indeed, in ovarian cancer the expression of MMP-2 and MMP-9 has been shown to correlate with poor survival indicating increased disseminating capability of cancer cells [18,19]. In addition to MMPs, other members of the metzincin superfamily such as a disintegrin and metalloproteinases (ADAM) and ADAM with thrombospondin motifs (ADAMTS) are known to be critically involved in ECM turnover and remodeling during tumourigenesis [20-22]. The same applies to the family of lysyl oxidase (LOX) enzymes and transglutaminases that also represent central molecules in regulating ECM organization and tumour progression [23,24]. Furthermore, cell membrane adhesion molecules called integrins are importantly involved in the development of tumours. For example, in prostate cancer metastasis integrin $\alpha v\beta 6$ expression has been shown to induce the expression of MMP-2 which in turn mediates osteolysis via its matrix degrading activity [25]. Certainly, other molecules and mechanisms whereby TME is involved in tumourigenesis could be presented [5,26,27].

Tumour progression also requires angiogenesis. Normally angiogenesis is strictly regulated. However, in the TME various cells can overexpress angiogenesis stimulating growth factors like VEGF ensuring oxygen and nutrient delivery to the growing tumour cell mass [28,29]. In addition to certain growth factors, specific ECM macromolecules by themselves are known to be able to regulate angiogenesis [30]. As the central role of the ECM and its components in tumourigenesis has become a recognized fact, to study the regulatory functions of the ECM macromolecules in the TME is of vital importance [31]. In this review we will focus on the ECM macromolecules in cancer and on the possibility to exploit them as novel therapeutic tools or targets in cancer gene therapy.

Review

ECM macromolecules and cancer

As mentioned in the beginning, ECM macromolecules can be categorized into four main groups: collagens, elastin and microfibrillar proteins, PGs and HA, and other non-collagenous matrix glycoproteins [2]. From these main groups, a subclass of secreted proteins, socalled "matricellular proteins" can be segregated [32-34]. They comprise a group of ECM macromolecules including thrombospondin- 1 and –2, SPARC (secreted protein, acidic and rich in cysteine), tenascin C (TN-C) and osteopontin [34]. The matricellular proteins do not directly participate in the formation of structural elements but are rather involved in the modulation of cell-matrix interactions and cell function. For example, TN-C is often associated with increased invasiveness of tumour cells [35]. On the basis of the ability of individual ECM macromolecules to form fibers, classification into fiberforming and interfibrillary matrix molecules can also be made. Most of these ECM macromolecules have been shown to be variously associated with cancer as described below.

Collagen is the most prominent structural protein of the ECM and 28 collagen types have been identified [36]. Different collagens, particularly types I and III are often associated with cancers such as breast and pancreatic cancers resulting in increased stromal collagen accumulation and promotion of cancer progression [37-40]. In breast cancer, collagen accumulation together with increased expression of small leucine-rich proteoglycans (SLRPs) decorin and lumican has been shown to correlate with increased mammographic density [41]. Also degradation of specific collagens has been observed during metastasis and the resulting collagen fragments can recruit e.g. TAMs whose abundance in tumours predicts poor prognosis [39,42]. On the other hand, degradation of collagen type XVIII resulting in the formation of endostatin causes inhibition of angiogenesis and thereby retards tumour growth [43]. Furthermore, some collagens such as type XXII and XXIV have been referred to have prognostic value in certain cancers [44].

Similarly to the richness of collagen types, over 30 mostly extracellular PG species have been identified [45,46]. With few exceptions PGs comprise a protein core to which one or more glycosaminoglycan (GAG) side chains are covalently linked. Many of the PGs, like certain family members of the SLRPs have been shown to be involved in the organization of the ECM [47]. For example, the SLRPs decorin, biglycan, fibromodulin and lumican are centrally involved in the regulation of collagen fibrillogenesis [48-51]. The above mentioned SLRPs are also variously involved in tumourigenesis. Especially decorin has a recognized regulatory role in tumour development, most notably via its ability to down regulate several members of the receptor tyrosine kinase (RTK) family members such as the epidermal growth factor receptor (EGFR) [52-54]. Furthermore, decorin can regulate the activity of the Met receptor, i.e., the receptor for hepatocyte growth factor, and the insulin-like growth factor receptor I (IGF-IR) [55,56]. Interestingly, decorin can also regulate tumour angiogenesis by reducing the production of e.g., VEGF [57,58]. Other mechanisms whereby decorin contributes to tumour growth will be discussed later on. Regarding lumican, it has been shown to variously modulate proliferation, migration and adhesion of cancer cells. For example, inhibition of tumour cell migration by lumican has been shown to be mediated via its interaction with $\alpha 2\beta 1$ integrin [59]. Similarly to decorin, lumican can also regulate cancer associated angiogenesis [60].

Hyaluronan is a versatile non-sulfated GAG that consists of repeating D-glucuronic acid and N-acetyl-D-glucosamine disaccharides. It has a widely recognized tumour promoting role in several cancer types such as prostate, ovarian and breast cancers via activating e.g., CD44 and RHAMM (receptor for hyaluronan-mediated motility) mediated signaling pathways including NFkB and mitogen-activated protein kinase (MAPK) pathways [61-63]. Indeed, its stromal accumulation is typical of progressed and undifferentiated tumours predicting poor prognosis [64-66].

Non-collagenous matrix glycoproteins like fibronectin have also been linked with cancers such as lung cancer, where its overexpression correlates with invasive and metastatic phenotype [67]. Fibronectin can bind to other ECM molecules and integrins including $\alpha_v\beta_3$ and thereby regulate among its other functions cell adhesion and Epithelia-Mesenchymal Transition (EMT) [68]. Different ECM molecules may also exist as tumour promoting splice variants as has been described e.g. for TN-C in breast cancer [69].

En bloc, ECM macromolecules are of central importance in cellular biology and they must act in concert in a finely regulated manner to maintain homeostasis and cellular functions within tissues and organs [70]. When the ECM is affected by inherited defects or its components are dysregulated, the proper 3D network of the matrix is lost and the cell-matrix interactions are hampered leading to or enabling different disease processes. Below, we will present some mechanisms whereby ECM macromolecules promote or alternatively inhibit tumourigenesis.

ECM macromolecules as regulators of tumourigenesis within the TME

Remodeling of the ECM is considered one of the earliest steps in the formation of TME which involves most ECM proteins [71]. Today we understand that there is an interaction between ECM macromolecules and cancer cells within each specific TME that can either facilitate or counteract the growth of solid tumours [9,72,73]. The TME can also restrict drug delivery to malignant cells [3,9]. As different ECM molecules possess central functional roles in various cellular events crucial in tumourigenesis like proliferation, adhesion, migration, survival and differentiation of the cells, they represent potential and novel targets and tools for pharmacotherapy [2].

With the help of the TME, cancer cells are able to exploit various mechanisms that can increase their malignancy and survival, e.g. they use autophagy to survive in nutrient-limited and hypoxic microenvironments [74]. On the other hand, autophagy has also been linked to suppression of tumourigenesis through elimination of p62 in tumour cells [75]. Interestingly, the SLRP decorin

has been found to act as a regulator of endothelial cell autophagy which results in the inhibition of angiogenesis and suppression of tumour growth [76]. More precisely, soluble decorin induces the expression of Peg3, a transcription factor usually silenced in cancer and causes inhibition of endothelial tubulogenesis and cell migration [77]. In addition to regulating autophagy, decorin is able to cause mitostatin-dependent mitophagy by inducing the expression of mitostatin via the Met receptor (the receptor for hepatocyte growth factor) resulting in concurrent decrease in VEGF-A transcription in breast cancer cells and subsequently inhibition of angiogenesis [78].

Other mechanisms that can be exploited by cancer cells via ECM macromolecules are Epithelia-Mesenchymal Transition (EMT) and its reverse process called Mesenchymal-Epithelial Transition (MET) [79]. The epithelial plasticity is a central part of normal development, but its regulators including myc, growth factors such as TGF- β , and β catenin can be misexpressed in cancers like breast [80], ovarian [81] and colon [82] cancers, respectively. This plasticity provides tumour cells with phenotypes which enable them to escape from the primary tumour. As breast cancer cells among other malignant cell types are known to secrete TN-C, an ECM macromolecule often associated with the invasive front of tumours [83,84], its expression together with the production of TGF-B1 can cause the malignant cells to undergo EMT [85]. Furthermore, integrins $\alpha v \beta 1$ and $\alpha v \beta 6$ are able to independently participate in this TN-C-induced EMT-like change [86]. The EMT also involves several other molecules including cadherins, focal adhesion molecules and proteolytic enzymes, e.g. MMPs [87]. All the above mechanistical possibilities have to be taken into account when novel therapies focusing on the ECM are developed.

ECM macromolecules as tools or targets in gene therapy

As discussed above, marked quantitative changes in the composition of the ECM macromolecules are a typical feature of different types of cancer. However, the distinction must be made whether these changes are the cause of the malignant disease or the result of it [7]. In spite of this, because individual ECM macromolecules can significantly influence cell behaviour and disease processes, modulation of the ECM composition provides a rational way to be used as an optional novel therapy in diseases like cancer. This modulation could be achieved with a number of approaches including gene therapy where specific ECM macromolecules are used as tools or targets.

Challenges using gene therapy

Generally, the development of gene therapy sets several challenges [88]. These challenges include choice of the vector to deliver the gene, the delivered gene itself and the route of administration. Furthermore, viruses must target diseased cells with enough large numbers to achieve wanted effects and at the same time with minimal destruction of the normal cells [89,90]. Timing of the gene delivery is also a critical issue to be considered because several tumours undergo marked temporal changes, as can be seen e.g. during the initiation and progression of pancreatic cancer [91]. Some tumour types like pancreatic cancer can also contain vast amounts of HA resulting in exceptionally high interstitial fluid pressure in solid tumour tissue, which in turn can prevent perfusion and diffusion of small molecule therapeutics [92]. In addition to the above challenges, alterations in the chemomechanical environment of cancer cells has to be recognized [5,93-95].

When gene therapy for cancer using or targeting ECM macromolecules is developed, the challenges can become even more sophisticated. For example, regarding the timing of gene therapy it is noteworthy to take into account that collagen expression has been discovered to have a dual role in tumour progression; different collagen types can either restrict or promote tumour development depending on the stage of the tumour [6,96]. Furthermore, phenomenon known as desmoplastic reaction, i.e., the formation of a sclerotic stroma around the cancer cell population due to the overexpression of certain collagen types can make the gene targeting problematic [7]. This dense fibrotic deposition is typical of cancers like colorectal [97,98], pancreatic [99,100] and liver [101] cancers. In addition to the possible increased amount of ECM macromolecules around the cancer cells, most cancers are of epithelial origin and still maintain intercellular junctions despite their de-differentiated nature further restricting

gene targeting [102]. However, it is possible to overcome these challenges as is discussed in the following two sections. Thus, gene therapy may have a promising future.

Strategies to overcome ECM macromolecule formed barrier in cancer treatment using gene delivery

Excess accumulation of the ECM macromolecules in the TME plays a critical role in blocking the transport of therapeutics to the target cells. Specifically, fibrillar collagen and collagen-proteoglycan bonds can form a major limit to the delivery of gene vectors [103,104]. Even nanomedicine utilizing nanoparticles can confront this same problem [105]. Regarding drug penetration to the tumour, the use of ECM macromolecules in gene therapy could provide a solution. For example the use of decorin containing replicating oncolytic adenovirus can increase drug penetration to solid tumours leading to a dramatic anti-tumour effect [102]. This beneficial effect of decorin gene delivery has been suggested to be based on decreased expression of other ECM molecules within the tumour tissue [102]. The physical barrier formed by ECM macromolecules is also possible to overcome by the administration of hyaluronidase (HAase) or collagenase containing oncolytic adenoviruses, which have been found to lead to significant tumour regression [106-108]. Additionally, the use of HAase can enhance the action of immune effector cells via degradation of the HA formed halos around cancer cells like adherent fibrosarcoma cells [109]. Treatment with HAase has also been shown to stimulate hematopoiesis and to increase the number of neutrophils in the peripheral blood [110].



Applying the same ECM degrading principle with other enzymes such as MMPs can be valuable [111]. For example, adenoviral expression of MMP-8, which breaks down collagen type I, II, and III, is a potential strategy for improving ECM hindered drug penetration or viral spread [112]. Apart from degrading excess ECM in the tumour matrix, reducing ECM production or modifying ECM organization by means of pharmacological intervention offers potential strategies to improve drug penetration [113,114]. This kind of manipulation of the tumour matrix structure can be considered as "matrix normalization" [105,113,114].

Inhibition of tumour progression using selected ECM macromolecules via gene delivery

A myriad number of ECM macromolecules are known to be involved directly or indirectly in tumourigenesis [5]. This provides several possibilities for using ECM macromolecules as tools in gene therapy. For example, expression of human decorin cDNA via adenovirus mediated transfection has been demonstrated to induce specific and even distant apoptosis of cancer cells demonstrating a direct antitumourigenic effect of decorin [115,116]. In addition to decorin, there are other PGs such as lumican and its core protein called lumcorin that have been found to possess antitumourigenic functions and could therefore be used as a chosen molecule in gene therapy [117-119]. Furthermore, human fibronectin containing recombinant adenovirus has been shown to be a promising strategy as a novel gene therapy against metastatic breast cancer via its inhibitory effect on adhesion of cancer cells to ECM molecules [120]. Additional ECMbased gene therapy strategies also exist. Regarding HA, interactions between HA and its major cell surface receptor, CD44, have raised interests on the basis of influencing on CD44 variants (CD44v) of cancer cells [121]. For example, cell-specific delivery of shRNA (short hairpin RNA) targeting HA-CD44v6 has been shown to lead to marked inhibition of tumour growth in mice [122]. In the case of collagen, particularly the 20 kDa C-terminal cleavage product of collagen type XVIII (endostatin) provides a very potent candidate in gene therapy in the future [43]. Endostatin is able to markedly inhibit not only angiogenesis but also lymphangiogenesis [123]. Adenoviral transfection carrying endostatin to bladder cancer cells in mouse models has already been shown to significantly decrease tumour progression via angiogenesis inhibition [124]. In theory, gene delivery of specific ECM macromolecules such as collagen and decorin could also be applied to encapsulate the tumour mass, thus possibly restricting tumour growth and metastasis [7,58]. A summary of the desired effects of different ECM-based gene therapies on tumours is presented in Figure 1.

Conclusions

The ECM has a recognized role in the regulation of key cellular functions such as adhesion, proliferation, migration and apoptosis. Also the differentiation of normal and cancer associated cells is known to be highly dependent on the interaction of the cells with their microenvironment. Therefore, the ECM is critically involved in various disease processes including tumourigenesis where significant changes in the composition and structure of the ECM can be observed. Although it has not been clearly established whether the observed ECM changes are the cause or the result of various diseases including cancer, strategies to modulate the structure of the ECM offers therapeutical possibilities. These could be achieved e.g. via gene therapy utilizing specific ECM macromolecules as tools and/or targets. As discussed in this review, the current literature offers solid rationale to the development of gene therapies focusing primarily on the ECM. However, several challenges must still be resolved. These include choice of the vector and the delivered gene itself, timing of the delivery and route of administration. Nevertheless, ECM-based gene therapies form a very attractive and promising field of research that is likely to advance novel treatments of cancer in the future.

Abbreviations

ADAM: A disintegrin and metalloproteinases; ADAMTS: A disintegrin and metalloproteinases (ADAM) with thrombospondin motifs; CAF: Cancer-associated fibroblast: CD44v: CD44 variants: CSC: Cancer stem cells: ECM: Extracellular matrix; EGFR: Epidermal growth factor receptor; EMT: Epithelial-mesenchymal transition; GAG: Glycosaminoglycan; HA: Hyaluronan; HAase: Hyaluronidase; IGF-IR: Insulin-like growth factor receptor I; IL-6: Interleukin 6; MAPK: Mitogen-activated protein kinase; MET: Mesenchymal-epithelial transition: Met: Mesenchymal-epithelial transition [Met] receptor/receptor for hepatocyte growth factor; MMP: Matrix metalloproteinase; PG: Proteoglycan; RHAMM: Receptor for hyaluronan-mediated motility; RTK: Receptor tyrosine kinase; shRNA: short hairpin RNA; SLRP: Small leucine-rich proteoglycan; SPARC: Secreted protein, acidic and rich in cysteine; TAM: Tumour-associated macrophage; TGF-B: Transforming growth factor beta; TME: Tumour microenvironment; TN-C: Tenascin; TNF-α: Tumour necrosis factor α; VEGF: Vascular endothelial growth factor.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AS and HJ equally contributed to writing and editing the manuscript. Both writers have read and approved the final manuscript.

Acknowledgements

Financially this study was supported by Medical Research Fund (EVO) of Turku University Hospital and Cancer Foundation of South-Western Finland.

Received: 6 February 2014 Accepted: 23 April 2014 Published: 2 May 2014

References

 Hynes RO: The extracellular matrix: not just pretty fibrils. Science 2009, 326:1216–1219.

- Järveläinen H, Sainio A, Koulu M, Wight TN, Penttinen R: Extracellular matrix molecules: potential targets in pharmacotherapy. *Pharmacol Rev* 2009, 61:198–223.
- Hanahan D, Weinberg RA: Hallmarks of cancer: the next generation. Cell 2011, 144:646–674.
- Frantz C, Stewart KM, Weaver VM: The extracellular matrix at a glance. J Cell Sci 2010, 123:4195–4200.
- Lu P, Weaver VM, Werb Z: The extracellular matrix: a dynamic niche in cancer progression. J Cell Biol 2012, 196:395–406.
- Fang H, Declerck YA: Targeting the tumor microenvironment: from understanding pathways to effective clinical trials. *Cancer Res* 2013, 73:4965–4977.
- Sainio A, Järveläinen H: Extracellular matrix molecules in tumour microenvironment with special reference to desmoplastic reaction and the role of matrix proteoglycans and hyaluronan. J Carcinog Mutagen, in press.
- Balkwill FR, Capasso M, Hagemann T: The tumor microenvironment at a glance. J Cell Sci 2012, 125:5591–5596.
- Ye J, Wu D, Wu P, Chen Z, Huang J: The cancer stem cell niche: cross talk between cancer stem cells and their microenvironment. *Tumour Biol*, in press.
- Edin S, Wikberg ML, Dahlin AM, Rutegård J, Öberg Å, Oldenborg PA, Palmqvist R: The distribution of macrophages with a M1 or M2 phenotype in relation to prognosis and the molecular characteristics of colorectal cancer. *PLoS One* 2012, 7:e47045.
- Fridlender ZG, Sun J, Kim S, Kapoor V, Cheng G, Ling L, Worthen GS, Albelda SM: Polarization of tumor-associated neutrophil phenotype by TGF-beta: "N1" versus "N2" TAN. *Cancer Cell* 2009, 16:183–194.
- 12. Toomer KH, Chen Z: Autoimmunity as a double agent in tumor killing and cancer promotion. *Front Immunol* 2014, **5**:116.
- Wood SL, Pernemalm M, Crosbie PA, Whetton AD: The role of the tumormicroenvironment in lung cancer-metastasis and its relationship to potential therapeutic targets. *Cancer Treat Rev*, in press.
- Lee HO, Mullins SR, Franco-Barraza J, Valianou M, Cukierman E, Cheng JD: FAP-overexpressing fibroblasts produce an extracellular matrix that enhances invasive velocity and directionality of pancreatic cancer cells. BMC Cancer 2011, 11:245.
- Santoni M, Massari F, Amantini C, Nabissi M, Maines F, Burattini L, Berardi R, Santoni G, Montironi R, Tortora G, Cascinu S: Emerging role of tumorassociated macrophages as therapeutic targets in patients with metastatic renal cell carcinoma. *Cancer Immunol Immunother* 2013, 62:1757–1768.
- Zeng-Brouwers J, Beckmann J, Nastase MV, Iozzo RV, Schaefer L: De novo expression of circulating biglycan evokes an innate inflammatory tissue response via MyD88/TRIF pathways. *Matrix Biol*, in press.
- 17. Wight TN, Kang I, Merrilees MJ: Versican and the control of inflammation. *Matrix Biol*, in press.
- Davidson B, Goldberg I, Gotlieb WH, Kopolovic J, Ben-Baruch G, Nesland JM, Berner A, Bryne M, Reich R: High levels of MMP-2, MMP-9, MT1-MMP and TIMP-2 mRNA correlate with poor survival in ovarian carcinoma. *Clin Exp Metastasis* 1999, 17:799–808.
- Davidson B, Goldberg I, Gotlieb WH, Kopolovic J, Ben-Baruch G, Nesland JM, Reich R: The prognostic value of metalloproteinases and angiogenic factors in ovarian carcinoma. *Mol Cell Endocrinol* 2002, 187:39–45.
- Georgiadis D, Yiotakis A: Specific targeting of metzincin family members with small-molecule inhibitors: progress toward a multifarious challenge. *Bioorg Med Chem* 2008, 16:8781–8794.
- Lu X, Lu D, Scully M, Kakkar V: ADAM proteins therapeutic potential in cancer. Curr Cancer Drug Targets 2008, 8:720–732.
- Fontanil T, Rúa S, Llamazares M, Moncada-Pazos A, Quirós PM, García-Suárez O, Vega JA, Sasaki T, Mohamedi Y, Esteban MM, Obaya AJ, Cal S: Interaction between the ADAMTS-12 metalloprotease and fibulin-2 induces tumor-suppressive effects in breast cancer cells. Oncotarget, in press.
- Lentini A, Abbruzzese A, Provenzano B, Tabolacci C, Beninati S: Transglutaminases: key regulators of cancer metastasis. *Amino Acids* 2013, 44:25–32.
- Mayorca-Guiliani A, Erler JT: The potential for targeting extracellular LOX proteins in human malignancy. Oncol Targets Ther 2013, 6:1729–1735.
- Dutta A, Li J, Lu H, Akech J, Pratap J, Wang T, Zerlanko BJ, Fitzgerald TJ, Jiang Z, Birbe R, Wixted J, Violette SM, Stein JL, Stein GS, Lian JB, Languino

LR: Integrin avß6 promotes an osteolytic program in cancer cells by upregulating MMP2. *Cancer Res*, in press.

- Hadler-Olsen E, Winberg JO, Uhlin-Hansen L: Matrix metalloproteinases in cancer: their value as diagnostic and prognostic markers and therapeutic targets. *Tumour Biol* 2013, 34:2041–2051.
- 27. Xiong J, Balcioglu HE, Danen EH: Integrin signaling in control of tumor growth and progression. Int J Biochem Cell Biol 2013, 45:1012–1015.
- Folkman J, Watson K, Ingber D, Hanahan D: Induction of angiogenesis during the transition from hyperplasia to neoplasia. *Nature* 1989, 339:58–61.
- Carmeliet P, Jain RK: Angiogenesis in cancer and other diseases. Nature 2000, 407:249–257.
- Ingber DE, Folkman J: How does extracellular matrix control capillary morphogenesis? Cell 1989, 58:803–805.
- 31. Quail DF, Joyce JA: Microenvironmental regulation of tumor progression and metastasis. *Nat Med* 2013, **19**:1423–1437.
- Sage EH, Bornstein P: Extracellular proteins that modulate cell-matrix interactions. SPARC, tenascin, and thrombospondin. J Biol Chem 1991, 266:14831–14834.
- 33. Bornstein P: Diversity of function is inherent in matricellular proteins: an appraisal of thrombospondin 1. *J Cell Biol* 1995, **130**:503–506.
- Bornstein P, Sage EH: Matricellular proteins: extracellular modulators cell function. Curr Opin Cell Biol 2002, 14:608–616.
- Varga I, Hutóczki G, Szemcsák CD, Zahuczky G, Tóth J, Adamecz Z, Kenyeres A, Bognár L, Hanzély Z, Klekner A: Brevican, neurocan, tenascin-C and versican are mainly responsible for the invasiveness of low-grade astrocytoma. Pathol Oncol Res 2012, 18:413–420.
- 36. Gordon MK, Hahn RA: Collagens. Cell Tissue Res 2010, 339:247-257.
- Kauppila S, Stenbäck F, Risteli J, Jukkola A, Risteli L: Aberrant type I and type III collagen gene expression in human breast cancer in vivo. *J Pathol* 1998, 186:262–268.
- Shields MA, Dangi-Garimella S, Redig AJ, Munshi HG: Biochemical role of the collagen-rich tumour microenvironment in pancreatic cancer progression. *Biochem J* 2012, 441:541–552.
- 39. Fang M, Yuan J, Peng C, Li Y: Collagen as a double-edged sword in tumor progression. *Tumour Biol* 2013: in press.
- Xiong G, Deng L, Zhu J, Rychahou PG, Xu R: Prolyl-4-hydroxylase α subunit 2 promotes breast cancer progression and metastasis by regulating collagen deposition. *BMC Cancer* 2014, 14:1.
- Alowami S, Troup S, Al-Haddad S, Kirkpatrick I, Watson PH: Mammographic density is related to stroma and stromal proteoglycan expression. *Breast Cancer Res* 2003, 5:R129–R135.
- 42. Bingle L, Brown NJ, Lewis CE: The role of tumour-associated macrophages in tumour progression: implications for new anticancer therapies. *J Pathol* 2002, **196:**254–265.
- O'Reilly MS, Boehm T, Shing Y, Fukai N, Vasios G, Lane WS, Flynn E, Birkhead JR, Olsen BR, Folkman J: Endostatin: an endogenous inhibitor of angiogenesis and tumor growth. *Cell* 1997, 88:277–285.
- Misawa K, Kanazawa T, Imai A, Endo S, Mochizuki D, Fukushima H, Misawa Y, Mineta H: Prognostic value of type XXII and XXIV collagen mRNA expression in head and neck cancer patients. *Mol Clin Oncol* 2014, 2:285–291.
- Järveläinen H, Wight TN: Vascular proteoglycans. In Proteoglycans in lung disease. Edited by Garg HG, Roughley PJ, Hales CA. New York: Marcel Dekker Inc; 2002:291–321.
- Schaefer L, Iozzo RV: Biological functions of the small leucine-rich proteoglycans: from genetics to signal transduction. J Biol Chem 2008, 283:21305–21309.
- lozzo RV: The family of the small leucine-rich proteoglycans: key regulators of matrix assembly and cellular growth. Crit Rev Biochem Mol Biol 1997, 32:141–174.
- Danielson KG, Baribault H, Holmes DF, Graham H, Kadler KE, Iozzo RV: Targeted disruption of decorin leads to abnormal collagen fibril morphology and skin fragility. J Cell Biol 1997, 136:729–743.
- Chakravarti S, Magnuson T, Lass JH, Jepsen KJ, LaMantia C, Carroll H: Lumican regulates collagen fibril assembly: skin fragility and corneal opacity in the absence of lumican. J Cell Biol 1998, 141:1277–1286.
- Svensson L, Aszódi A, Reinholt FP, Fässler R, Heinegård D, Oldberg A: Fibromodulin-null mice have abnormal collagen fibrils, tissue organization, and altered lumican deposition in tendon. *J Biol Chem* 1999, 274:9636–9647.

- Goldberg M, Rapoport O, Septier D, Palmier K, Hall R, Embery G, Young M, Ameye L: Proteoglycans in predentin: the last 15 micrometers before mineralization. *Connect Tissue Res* 2003, 44(Suppl 1):184–188.
- Reed CC, Waterhouse A, Kirby S, Kay P, Owens RT, McQuillan DJ, Iozzo RV: Decorin prevents metastatic spreading of breast cancer. Oncogene 2005, 24:1104–1110.
- Goldoni S, Seidler DG, Heath J, Fassan M, Baffa R, Thakur ML, Owens RT, McQuillan DJ, Iozzo RV: An antimetastatic role for decorin in breast cancer. Am J Pathol 2008, 173:844–855.
- Bi X, Pohl NM, Qian Z, Yang GR, Gou Y, Guzman G, Kajdacsy-Balla A, Iozzo RV, Yang W: Decorin-mediated inhibition of colorectal cancer growth and migration is associated with E-cadherin in vitro and in mice. *Carcinogenesis* 2012, 33:326–330.
- Goldoni S, Humphries A, Nyström A, Sattar S, Owens RT, McQuillan DJ, Ireton K, Iozzo RV: Decorin is a novel antagonistic ligand of the Met receptor. J Cell Biol 2009, 185:743–754.
- Iozzo RV, Buraschi S, Genua M, Xu SQ, Solomides CC, Peiper SC, Gomella LG, Owens RC, Morrione A: Decorin antagonizes IGF receptor I (IGF-IR) function by interfering with IGF-IR activity and attenuating downstream signaling. J Biol Chem 2011, 286:34712–34721.
- Grant DS, Yenisey C, Rose RW, Tootell M, Santra M, Iozzo RV: Decorin suppresses tumor cell-mediated angiogenesis. Oncogene 2002, 21:4765–4777.
- Salomäki HH, Sainio AO, Söderström M, Pakkanen S, Laine J, Järveläinen HT: Differential expression of decorin by human malignant and benign vascular tumors. J Histochem Cytochem 2008, 56:639–646.
- Brézillon S, Pietraszek K, Maquart FX, Wegrowski Y: Lumican effects in the control of tumour progression and their links with metalloproteinases and integrins. *FEBS J* 2013, 280:2369–2381.
- 60. Nikitovic D, Papoutsidakis A, Karamanos NK, Tzanakakis GN: Lumican affects tumor cell functions, tumor-ECM interactions, angiogenesis and inflammatory response. *Matrix Biol*, in press.
- Craig EA, Parker P, Camenisch TD: Size dependent regulation of Snail2 by hyaluronan: its role in cellular invasion. *Glycobiology* 2009, 19:890–898.
- 62. Telmer PG, Tolg C, McCarthy JB, Turley EA: How does a protein with dual mitotic spindle and extracellular matrix receptor functions affect tumor susceptibility and progression? *Commun Integr Biol* 2011, **4**:182–185.
- Dicker KT, Gurski LA, Pradhan-Bhatt S, Witt RL, Farach-Carson MC, Jia X: Hyaluronan: A simple polysaccharide with diverse biological functions. *Acta Biomater*, in press.
- Lipponen P, Aaltomaa S, Tammi R, Tammi M, Agren U, Kosma VM: High stromal hyaluronan level is associated with poor differentiation and metastasis in prostate cancer. *Eur J Cancer* 2001, 37:849–856.
- Anttila MA, Tammi RH, Tammi MI, Syrjänen KJ, Saarikoski SV, Kosma VM: High levels of stromal hyaluronan predict poor disease outcome in epithelial ovarian cancer. *Cancer Res* 2000, 60:150–155.
- Auvinen P, Tammi R, Parkkinen J, Tammi M, Agren U, Johansson R, Hirvikoski P, Eskelinen M, Kosma VM: Hyaluronan in peritumoral stroma and malignant cells associates with breast cancer spreading and predicts survival. Am J Pathol 2000, 156:529–536.
- Shuman Moss LA, Stetler-Stevenson WG: Influence of stromal components on lung cancer carcinogenesis. J Carcinog Mutagen 2013, 13(8). doi: 10.4172/2157-2518.513-008.
- Jia D, Yan M, Wang X, Hao X, Liang L, Liu L, Kong H, He X, Li J, Yao M: Development of a highly metastatic model that reveals a crucial role of fibronectin in lung cancer cell migration and invasion. *BMC Cancer* 2010, 10:364.
- Hancox RA, Allen MD, Holliday DL, Edwards DR, Pennington CJ, Guttery DS, Shaw JA, Walker RA, Pringle JH, Jones JL: Tumour-associated tenascin-C isoforms promote breast cancer cell invasion and growth by matrix metalloproteinase-dependent and independent mechanisms. *Breast Cancer Res* 2009, 11:R24.
- Lukashev ME, Werb Z: ECM signaling: orchestrating cell behaviour and misbehaviour. Trends Cell Biol 1998, 8:437–441.
- Botti G, Cerrone M, Scognamiglio G, Anniciello A, Ascierto PA, Cantile M: Microenvironment and tumor progression of melanoma: new therapeutic prospectives. J Immunotoxicol 2013, 10:235–252.
- Theocharis AD, Skandalis SS, Tzanakakis GN, Karamanos NK: Proteoglycans in health and disease: novel roles for proteoglycans in malignancy and their pharmacological targeting. *FEBS J* 2010, 277:3904–3923.
- 73. Skandalis SS, Aletras AJ, Gialeli C, Theocharis AD, Afratis N, Tzanakakis GN, Karamanos NK: **Targeting the tumor proteasome as a mechanism to**

control the synthesis and bioactivity of matrix macromolecules. *Curr Mol Med* 2012, **12**:1068–1082.

- 74. Chen N, Karantza-Wadsworth V: Role and regulation of autophagy in cancer. *Biochim Biophys Acta* 2009, **1793:**1516–1523.
- Mathew R, Karp CM, Beaudoin B, Vuong N, Chen G, Chen HY, Bray K, Reddy A, Bhanot G, Gelinas C, Dipaola RS, Karantza-Wadsworth V, White E: Autophagy suppresses tumorigenesis through elimination of p62. *Cell* 2009, 137:1062–1075.
- 76. Neill T, Torres A, Buraschi S, Iozzo RV: Decorin has an appetite for endothelial cell autophagy. *Autophagy* 2013, **9**:1626–1628.
- Buraschi S, Neill T, Goyal A, Poluzzi C, Smythies J, Owens RT, Schaefer L, Torres A, Iozzo RV: Decorin causes autophagy in endothelial cells via Peg3. Proc Natl Acad Sci U S A 2013, 110:E2582–E2591.
- Neill T, Torres A, Buraschi S, Owens RT, Hoek JB, Baffa R, Iozzo RV: Decorin induces mitophagy in breast carcinoma cells via PGC-1α and mitostatin. *J Biol Chem*, in press.
- 79. Micalizzi DS, Farabaugh SM, Ford HL: Epithelial-mesenchymal transition in cancer: parallels between normal development and tumor progression. *J Mammary Gland Biol Neoplasia* 2010, **15**:117–134.
- Trimboli AJ, Fukino K, de Bruin A, Wei G, Shen L, Tanner SM, Creasap N, Rosol TJ, Robinson ML, Eng C, Ostrowski MC, Leone G: Direct evidence for epithelial-mesenchymal transitions in breast cancer. *Cancer Res* 2008, 68:937–945.
- Vergara D, Merlot B, Lucot JP, Collinet P, Vinatier D, Fournier I, Salzet M: Epithelial-mesenchymal transition in ovarian cancer. *Cancer Lett* 2010, 291:59–66.
- Brabletz T, Hlubek F, Spaderna S, Schmalhofer O, Hiendlmeyer E, Jung A, Kirchner T: Invasion and metastasis in colorectal cancer: epithelialmesenchymal transition, mesenchymal-epithelial transition, stem cells and beta-catenin. *Cells Tissues Organs* 2005, **179**:56–65.
- Tsunoda T, Inada H, Kalembeyi I, Imanaka-Yoshida K, Sakakibara M, Okada R, Katsuta K, Sakakura T, Majima Y, Yoshida T: Involvement of large tenascin-C splice variants in breast cancer progression. *Am J Pathol* 2003, 162:1857–1867.
- Oskarsson T, Acharyya S, Zhang XH, Vanharanta S, Tavazoie SF, Morris PG, Downey RJ, Manova-Todorova K, Brogi E, Massagué J: Breast cancer cells produce tenascin C as a metastatic niche component to colonize the lungs. Nat Med 2011, 17:867–874.
- Maschler S, Grunert S, Danielopol A, Beug H, Wirl G: Enhanced tenascin-C expression and matrix deposition during Ras/TGF-beta-induced progression of mammary tumor cells. *Oncogene* 2004, 23:3622–3633.
- Katoh D, Nagaharu K, Shimojo N, Hanamura N, Yamashita M, Kozuka Y, Imanaka-Yoshida K, Yoshida T: Binding of αvβ1 and αvβ6 integrins to tenascin-C induces epithelial-mesenchymal transition-like change of breast cancer cells. Oncogenesis 2013, 2:e65.
- 87. Ghersi G: Roles of molecules involved in epithelial/mesenchymal transition during angiogenesis. *Front Biosci* 2008, **13:**2335–2355.
- Wirth T, Parker N, Ylä-Herttuala S: History of gene therapy. Gene 2013, 525:162–169.
- Liu TC, Galanis E, Kirn D: Clinical trial results with oncolytic virotherapy: a century of promise, a decade of progress. *Nat Clin Pract Oncol* 2007, 4:101–117.
- Haseley A, Alvarez-Breckenridge C, Chaudhury AR, Kaur B: Advances in oncolytic virus therapy for glioma. *Recent Pat CNS Drug Discov* 2009, 4:1–13.
- Lunardi S, Muschel RJ, Brunner TB: The stromal compartments in pancreatic cancer: are there any therapeutic targets? *Cancer Lett* 2014, 343:147–155.
- Provenzano PP, Cuevas C, Chang AE, Goel VK, Von Hoff DD, Hingorani SR: Enzymatic targeting of the stroma ablates physical barriers to treatment of pancreatic ductal adenocarcinoma. *Cancer Cell* 2012, 21:418–429.
- 93. Mierke CT: Endothelial cell's biomechanical properties are regulated by invasive cancer cells. *Mol Biosyst* 2012, 8:1639–1649.
- 94. Mierke CT: Physical break-down of the classical view on cancer cell invasion and metastasis. *Eur J Cell Biol* 2013, **92**:89–104.
- Denais C, Lammerding J: Nuclear mechanics in cancer. Adv Exp Med Biol 2014, 773:435–470.
- Zou X, Feng B, Dong T, Yan G, Tan B, Shen H, Huang A, Zhang X, Zhang M, Yang P, Zheng M, Zhang Y: Up-regulation of type I collagen during tumorigenesis of colorectal cancer revealed by quantitative proteomic analysis. J Proteomics 2013, 94:473–485.

- Coulson-Thomas VJ, Coulson-Thomas YM, Gesteira TF, de Paula CA, Mader AM, Waisberg J, Pinhal MA, Friedl A, Toma L, Nader HB: Colorectal cancer desmoplastic reaction up-regulates collagen synthesis and restricts cancer cell invasion. *Cell Tissue Res* 2011, 346:223–236.
- Karagiannis GS, Petraki C, Prassas I, Saraon P, Musrap N, Dimitromanolakis A, Diamandis EP: Proteomic signatures of the desmoplastic invasion front reveal collagen type XII as a marker of myofibroblastic differentiation during colorectal cancer metastasis. Oncotarget 2012, 3:267–285.
- Merika EE, Syrigos KN, Saif MW: Desmoplasia in pancreatic cancer. Can we fight it? Gastroenterol Res Pract 2012, 2012:781765.
- 100. Whatcott CJ, Posner RG, Von Hoff DD, Han H: Desmoplasia and chemoresistance in pancreatic cancer. In *Pancreatic Cancer and Tumor Microenvironment*. Edited by Grippo PJ, Munshi HG. Trivandrum: Transworld Research Network; 2012. Chapter 8.
- Kocabayoglu P, Friedman SL: Cellular basis of hepatic fibrosis and its role in inflammation and cancer. Front Biosci (Schol Ed) 2013, 5:217–230.
- Choi IK, Strauss R, Richter M, Yun CO, Lieber A: Strategies to increase drug penetration in solid tumors. Front Oncol 2013, 3:193.
- Netti PA, Berk DA, Swartz MA, Grodzinsky AJ, Jain RK: Role of extracellular matrix assembly in interstitial transport in solid tumors. *Cancer Res* 2000, 60:2497–2503.
- 104. McKee TD, Grandi P, Mok W, Alexandrakis G, Insin N, Zimmer JP, Bawendi MG, Boucher Y, Breakefield XO, Jain RK: Degradation of fibrillar collagen in a human melanoma xenograft improves the efficacy of an oncolytic herpes simplex virus vector. *Cancer Res* 2006, 66:2509–2513.
- 105. Chauhan VP, Jain RK: Strategies for advancing cancer nanomedicine. Nat Mater 2013, 12:958–962.
- Kuriyama N, Kuriyama H, Julin CM, Lamborn K, Israel MA: Pretreatment with protease is a useful experimental strategy for enhancing adenovirusmediated cancer gene therapy. *Hum Gene Ther* 2000, 11:2219–2230.
- 107. Kuriyama N, Kuriyama H, Julin CM, Lamborn KR, Israel MA: Protease pretreatment increases the efficacy of adenovirus-mediated gene therapy for the treatment of an experimental glioblastoma model. *Cancer Res* 2001, **61**:1805–1809.
- Guedan S, Rojas JJ, Gros A, Mercade E, Cascallo M, Alemany R: Hyaluronidase expression by an oncolytic adenovirus enhances its intratumoral spread and suppresses tumor growth. *Mol Ther* 2010, 18:1275–1283.
- McBride WH, Bard JB: Hyaluronidase-sensitive halos around adherent cells. Their role in blocking lymphocyte-mediated cytolysis. J Exp Med 1979, 149:507–515.
- Zyuz'kov GN, Zhdanov W, Dygai AM, Gol'dberg ED: Role of hyaluronidase in the regulation of hemopoiesis. Bull Exp Biol Med 2007, 144:840–845.
- Kessenbrock K, Plaks V, Werb Z: Matrix metalloproteinases: regulators of the tumor microenvironment. *Cell* 2010, 141:52–67.
- Cheng J, Sauthoff H, Huang Y, Kutler DI, Bajwa S, Rom WN, Hay JG: Human matrix metalloproteinase-8 gene delivery increases the oncolytic activity of a replicating adenovirus. *Mol Ther* 2007, 15:1982–1990.
- 113. Brown E, McKee T, di Tomaso E, Pluen A, Seed B, Boucher Y, Jain RK: Dynamic imaging of collagen and its modulation in tumors in vivo using second-harmonic generation. *Nat Med* 2003, 9:796–800.
- 114. Diop-Frimpong B, Chauhan VP, Krane S, Boucher Y, Jain RK: Losartan inhibits collagen I synthesis and improves the distribution and efficacy of nanotherapeutics in tumors. Proc Natl Acad Sci U S A 2011, 108:2909–2914.
- 115. Tralhão JG, Schaefer L, Micegova M, Evaristo C, Schönherr E, Kayal S, Veiga-Fernandes H, Danel C, Iozzo RV, Kresse H, Lemarchand P: In vivo selective and distant killing of cancer cells using adenovirus-mediated decorin gene transfer. FASEB J 2003, 17:464–466.
- 116. Boström P, Sainio A, Kakko T, Savontaus M, Söderström M, Järveläinen H: Localization of decorin gene expression in normal human breast tissue and in benign and malignant tumors of the human breast. *Histochem Cell Biol* 2013, **139**:161–171.
- 117. Coulson-Thomas VJ, Coulson-Thomas YM, Gesteira TF, Andrade de Paula CA, Carneiro CR, Ortiz V, Toma L, Kao WW, Nader HB: Lumican expression, localization and antitumor activity in prostate cancer. *Exp Cell Res* 2013, 319:967–981.
- 118. de Wit M, Belt EJ, Delis-van Diemen PM, Carvalho B, Coupé VM, Stockmann HB, Bril H, Beliën JA, Fijneman RJ, Meijer GA: Lumican and versican are associated with good outcome in stage II and III colon cancer. *Ann Surg Oncol* 2013, **20**(Suppl 3):S348–S359.

- Page 8 of 8
- Pietraszek K, Brézillon S, Perreau C, Malicka-Blaszkiewicz M, Maquart FX, Wegrowski Y: Lumican - derived peptides inhibit melanoma cell growth and migration. *PLoS One* 2013, 8:e76232.
- 120. He ZH, Lei Z, Zhen Y, Gong W, Huang B, Yuan Y, Zhang GM, Wang XJ, Feng ZH: Adeno-associated virus-mediated expression of recombinant CBD-Hepll polypeptide of human fibronectin inhibits metastasis of breast cancer. Breast Cancer Res Treat 2014, 143:33–45.
- Misra S, Heldin P, Hascall VC, Karamanos NK, Skandalis SS, Markwald RR, Ghatak S: Hyaluronan-CD44 interactions as potential targets for cancer therapy. *FEBS J* 2011, 278:1429–1443.
- 122. Misra S, Hascall VC, De Giovanni C, Markwald RR, Ghatak S: Delivery of CD44 shRNA/nanoparticles within cancer cells: perturbation of hyaluronan/CD44v6 interactions and reduction in adenoma growth in Apc Min/+MICE. J Biol Chem 2009, 284:12432–12446.
- 123. Seppinen L, Pihlajaniemi T: The multiple functions of collagen XVIII in development and disease. *Matrix Biol* 2011, **30**:83–92.
- 124. Pan JG, Luo RQ, Zhou X, Han RF, Zeng GW: Potent antitumor activity of the combination of HSV-TK and endostatin by adeno-associated virus vector for bladder cancer in vivo. *Clin Lab* 2013, **59**:1147–1158.

doi:10.1186/2052-8426-2-14

Cite this article as: Sainio and Järveläinen: Extracellular matrix macromolecules: potential tools and targets in cancer gene therapy. *Molecular and Cellular Therapies* 2014 2:14.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at www.biomedcentral.com/submit

