



Draft Genome Sequences of *Bacillus* and *Paenibacillus* Species Isolated from Seeds of *Citrullus Ianata* (Watermelon), *Cucurbita moschata* (Butternut Squash), and *Cucurbita pepo* L. var. *pepo* L. (Pumpkin)

Eman M. Khalaf,^{a,b} Manish N. Raizada^a

^aDepartment of Plant Agriculture, University of Guelph, Guelph, Ontario, Canada ^bDepartment of Microbiology and Immunology, Faculty of Pharmacy, Damanhour University, Damanhour, Egypt

ABSTRACT Here, we announce the draft genome sequences of four endophytic bacilli isolated from surface-sterilized seeds of three cucurbit species, *Bacillus* sp. strains EKM417B and EKM420B (from *Citrullus lanata* [watermelon]) and EKM501B (from *Cucurbita moschata* [butternut squash]) and *Paenibacillus* sp. strain EKM301P (from *Cucurbita pepo* L. var. *pepo* L. [pumpkin]). These strains previously demonstrated biostimulant and biocontrol activities.

Dlant microbiomes have evolved to perform defensive/growth-promoting functions (1-3). 16S Illumina sequencing of cucurbit seeds revealed the dominance of spore-forming, Gram-positive bacteria (e.g., Bacillus/Paenibacillus genera) (4), consistent with aerobically cultivated microbiota (5). We isolated unique colonies from cultivated surface-sterilized cucurbit seeds. They were identified as Bacillus sp. strains EKM417B and EKM420B (from Citrullus lanata [watermelon]), Bacillus sp. strain EKM501B (from Cucurbita moschata [butternut squash]), and Paenibacillus sp. strain EKM301P (Cucurbita pepo L. var. pepo L. [pumpkin]) using 16S rRNA gene primer pair 799F and 1492R and then submitted to GenBank (accession numbers KT281355, KT281357, KT281359, and KT281432, respectively) (5). Since many commercial microbial fertilizers/biocontrol agents are Bacillus/Paenibacillus based (1, 2), these candidate endophytes were tested for beneficial traits in vitro/in planta (5, 6). All four strains showed in vitro protease activity (5) and acetoin/diacetyl production (volatiles) and suppressed Phytophthora capsici (6). Other in vitro traits were scored, albeit inconsistently, as follows: EKM417B/ EKM420B displayed pectinase and RNase activities; EKM417B secreted cellulase and reduced the disease severity of Podosphaera fuliginea (foliar fungal pathogen) in planta; EKM501B grew on N₂-free medium, produced indole-3-acetic acid (IAA/auxin) and ribonucleases, and suppressed Rhizoctonia solani; and EKM301P secreted cellulase and antagonized Fusarium graminearum and Rhizoctonia solani (5, 6).

The strains were cultured overnight on LB agar from original -80° C glycerol stocks. Single colonies were inoculated into lysogeny broth (overnight, 37°C, 250 rpm). Genomic DNA was extracted using DNeasy UltraClean microbial kits (Qiagen, catalog number 12224-50) and adjusted to 50 ng/µl. Libraries were constructed using TruSeq DNA Nano library prep kits (KAPA HyperPrep kit, catalog number KK8504) and then sequenced using the Illumina NovaSeq 6000 system to produce 1,461,384 (EKM417B), 1,659,509 (EKM420B), 1,823,824 (EKM501B), and 2,124,086 (EKM301P) raw reads in 150-bp paired-end format. Quality-trimmed reads (with a quality score of 30) were *de novo* assembled using the EvoCAT pipeline (Evogene Clustering and Assembly Toolbox) and identified using KmerFinder 3.2 (7) by conducting a BLAST search against *Bacillus velezensis* strain KD1 (GenBank

genome sequences of *Bacillus* and *Paenibacillus* species isolated from seeds of *Citrullus* lanata (watermelon), *Cucurbita moschata* (butternut squash), and *Cucurbita pepo* L. var. *pepo* L. (pumpkin). Microbiol Resour Announc 9:e00727-20. https://doi.org/10.1128/ MRA.00727-20.

Citation Khalaf EM, Raizada MN. 2020. Draft

Editor David A. Baltrus, University of Arizona Copyright © 2020 Khalaf and Raizada. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Eman M. Khalaf, emankhalaf@pharm.dmu.edu.eg.

Received 20 June 2020 Accepted 29 July 2020 Published 20 August 2020

Metrics	Data for bacterial strains ^a			
	B. velezensis EKM417B	B. velezensis EKM420B	B. cereus EKM501B	P. polymyxa EKM301P
Genome size (bp)	3,972,727	3,908,635	5,734,326	5,661,237
No. of clean reads	1,228,648	1,426,781	1,613,183	1,755,075
No. of contigs	310	83	105	198
N ₅₀ (bp)	996,524	996,284	239,839	531,804
Maximum scaffold length (bp)	1,083,982	1,084,468	876,274	1,218,405
Minimum scaffold length (bp)	206	206	202	203
Avg genome coverage (\times)	93	109	91	90
No. of predicted genes	3,693	3,553	5,055	4,560
G+C content (%)	45	47	36	52
GenBank accession no.	JAAMNY000000000	JAAMNX000000000	JAALFZ000000000	JAALFM00000000
SRA accession no.	SRR11043897	SRR11051662	SRR11051671	SRR11048276

^a The taxonomy of bacterial species was obtained from the updated GenBank databases.

accession number NZ_CP014990.2) (EKM417B and EKM420B), *Bacillus cereus* strain FORC087 (NZ_CP029454.1) (EKM501B), and *Paenibacillus polymyxa* strain SQR-21 (NZ_CP006872.1) (8) (EKM301P) as the top genome matches (Table 1). The protein predictions were completed using Prodigal (9) and then matched against the NCBI nonredundant protein database using Blastp (10). Peptide domains were identified using InterProScan 5.32-71.0 (11). Default parameters were used for all software unless otherwise specified. The statistics of the genomes are provided in Table 1.

Genome mining identified candidate genes involved in biofertilizer/biocontrol metabolic pathways, including those discussed above. These genes encode proteins involved in nitrogen fixation, phytase, alkaline phosphatase, carbon-nitrogen hydrolase, trehalose-6-phophate hydrolase, and tryptophan synthase (IAA/auxin production) (12– 14). Biocontrol/systemic resistance elicitor candidate genes encode hydrolytic enzymes (β -glucanase, chitinase, cellulase, proteases, pectin/pectate lyases, lipases, ribonucleases) (14–16), exopolysaccharide synthesis protein (colonization ability) (17), butanedioldehydrogenase-like (acetoin production) (18), iron-siderophore-like, polyketide synthase, nonribosomal peptide synthase (NRPS) (19), phenazine biosynthesis PhzF protein (except EKM301P) (20), bacteriocins (class IId [EKM417B/EKM420B], class IIb [EKM501B], and thiopeptide-type [EKM301P]) (21), and phenylalanine/histidine ammonia-lyases (except EKM301P) (22). The exception is that EKM501B lacked β -glucanase, pectate lyase, and phytase but encoded aerobactin siderophores (23). In conclusion, cucurbit seeds host bacilli as vectors that encode candidate beneficial traits for plants.

Data availability. This whole-genome shotgun project and the Illumina raw reads have been deposited in DDBJ/EMBL/GenBank and the SRA, respectively, under the accession numbers provided in Table 1.

ACKNOWLEDGMENTS

We thank Ada Viterbo and collaborators at Lavie Bio Ltd. for genome sequencing and bioinformatic support.

This research was supported by grants from the Ontario Ministry of Agriculture, Food, and Rural Affairs (OMAFRA) and the Natural Sciences and Engineering Research Council of Canada (NSERC).

REFERENCES

- Rybakova D, Cernava T, Köberl M, Liebminger S, Etemadi M, Berg G. 2016. Endophytes-assisted biocontrol: novel insights in ecology and the mode of action of *Paenibacillus*. Plant Soil 405:125–140. https://doi.org/ 10.1007/s11104-015-2526-1.
- Olishevska S, Nickzad A, Déziel E. 2019. Bacillus and Paenibacillus secreted polyketides and peptides involved in controlling human and plant pathogens. Appl Microbiol Biotechnol 103:1189–1215. https://doi .org/10.1007/s00253-018-9541-0.
- 3. Compant S, Saikkonen K, Mitter B, Campisano A, Mercado-Blanco J. 2016.

Editorial special issue: soil, plants and endophytes. Plant Soil 405:1–11. https://doi.org/10.1007/s11104-016-2927-9.

- Khalaf EM, Raizada MN. 2019. Cucurbit seeds: reservoirs of functional and antagonistic microbiomes. Plant Canada 2019, Guelph, Ontario, Canada, 7–10 July 2019.
- Khalaf EM, Raizada MN. 2016. Taxonomic and functional diversity of cultured seed associated microbes of the cucurbit family. BMC Microbiol 16:131. https://doi.org/10.1186/s12866-016-0743-2.
- 6. Khalaf EM, Raizada MN. 2018. Bacterial seed endophytes of domesti-

cated cucurbits antagonize fungal and oomycete pathogens including powdery mildew. Front Microbiol 9:42. https://doi.org/10.3389/fmicb .2018.00042.

- Deng X, den Bakker HC, Hendriksen RS. 2016. Genomic epidemiology: whole-genome-sequencing-powered surveillance and outbreak investigation of foodborne bacterial pathogens. Annu Rev Food Sci Technol 7:353–374. https://doi.org/10.1146/annurev-food-041715-033259.
- Li S, Yang D, Qiu M, Shao J, Guo R, Shen B, Yin X, Zhang R, Zhang N, Shen Q. 2014. Complete genome sequence of *Paenibacillus polymyxa* SQR-21, a plant growth-promoting rhizobacterium with antifungal activity and rhizosphere colonization ability. Genome Announc 2:e00281-14. https:// doi.org/10.1128/genomeA.00281-14.
- Hyatt D, Chen G-L, LoCascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. Nat Commun 11:119. https://doi.org/10.1186/1471-2105 -11-119.
- Pruitt KD, Tatusova T, Maglott DR. 2007. NCBI reference sequences (RefSeq): a curated non-redundant sequence database of genomes, transcripts and proteins. Nucleic Acids Res 35:D61–D65. https://doi.org/ 10.1093/nar/gkl842.
- Quevillon E, Silventoinen V, Pillai S, Harte N, Mulder N, Apweiler R, Lopez R. 2005. InterProScan: protein domains identifier. Nucleic Acids Res 33:W116–W120. https://doi.org/10.1093/nar/gki442.
- Xie J, Shi H, Du Z, Wang T, Liu X, Chen S. 2016. Comparative genomic and functional analysis reveal conservation of plant growth promoting traits in *Paenibacillus polymyxa* and its closely related species. Sci Rep 6:21329. https://doi.org/10.1038/srep21329.
- Wang J, Li R, Zhang H, Wei G, Li Z. 2020. Beneficial bacteria activate nutrients and promote wheat growth under conditions of reduced fertilizer application. BMC Microbiol 20:38. https://doi.org/10.1186/ s12866-020-1708-z.
- Elshaghabee FMF, Rokana N, Gulhane RD, Sharma C, Panwar H. 2017. Bacillus as potential probiotics: status, concerns, and future perspectives. Front Microbiol 8:1490. https://doi.org/10.3389/fmicb.2017.01490.
- 15. Grady EN, MacDonald J, Liu L, Richman A, Yuan ZC. 2016. Current

knowledge and perspectives of *Paenibacillus*: a review. Microb Cell Fact 15:203. https://doi.org/10.1186/s12934-016-0603-7.

- Soltani M, Ghosh K, Hoseinifar SH, Kumar V, Lymbery AJ, Roy S, Ringø E. 2019. Genus *Bacillus*, promising probiotics in aquaculture: aquatic animal origin, bio-active components, bioremediation and efficacy in fish and shellfish. Rev Fish Sci Aquac 27:331–379. https://doi.org/10.1080/ 23308249.2019.1597010.
- Chen L, Heng J, Qin S, Bian K. 2018. A comprehensive understanding of the biocontrol potential of *Bacillus velezensis* LM2303 against *Fusarium* head blight. PLoS One 13:e0198560. https://doi.org/10.1371/journal .pone.0198560.
- Peng G, Zhao X, Li Y, Wang R, Huang Y, Qi G. 2019. Engineering *Bacillus velezensis* with high production of acetoin primes strong induced systemic resistance in *Arabidopsis thaliana*. Microbiol Res 227:126297. https://doi.org/10.1016/j.micres.2019.126297.
- Fan B, Wang C, Song X, Ding X, Wu L, Wu H, Gao X, Borriss R. 2018. Bacillus velezensis FZB42 in 2018: the Gram-positive model strain for plant growth promotion and biocontrol. Front Microbiol 9:2491. https:// doi.org/10.3389/fmicb.2018.02491.
- Padaria JC, Tarafdar A, Raipuria R, Lone SA, Gahlot P, Shakil NA, Kumar J. 2016. Identification of phenazine-1-carboxylic acid gene (phc CD) from *Bacillus pumilus* MTCC7615 and its role in antagonism against *Rhizoctonia solani*. J Basic Microbiol 56:999–1008. https://doi.org/10.1002/jobm .201500574.
- Cotter PD, Ross RP, Hill C. 2013. Bacteriocins: a viable alternative to antibiotics? Nat Rev Microbiol 11:95–105. https://doi.org/10.1038/ nrmicro2937.
- Mandal SM, Chakraborty D, Dey S. 2010. Phenolic acids act as signaling molecules in plant-microbe symbioses. Plant Signal Behav 5:359–368. https://doi.org/10.4161/psb.5.4.10871.
- Zawadzka AM, Abergel RJ, Nichiporuk R, Andersen UN, Raymond KN. 2009. Siderophore-mediated iron acquisition systems in *Bacillus cereus*: identification of receptors for anthrax virulence- associated petrobactin. Biochemistry 48:3645–3657. https://doi.org/10.1021/bi8018674.