

Non-invasive monitoring of hepatic steatosis via acoustic structure quantification of ultrasonography with MR spectroscopy as the reference standard

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Purpose: The purpose of this study was to prospectively evaluate whether monitoring hepatic steatosis by ultrasonography with acoustic structure quantification (ASQ) is feasible, using magnetic resonance spectroscopy (MRS) as the reference standard.

Methods: Thirty-six patients with suspected fatty liver disease underwent both ultrasonography with ASQ and MRS on the same day. After a mean follow-up period of 11.4 ± 2.5 months, follow-up ultrasonography with ASQ and MRS were performed on 27 patients to evaluate whether hepatic steatosis had improved. The focal disturbance (FD) ratio, as calculated using ASQ, and the hepatic fat fraction (HFF), estimated by MRS, were obtained at both initial and follow-up examinations. Pearson correlation coefficients were calculated to assess the correlations between ordinal values.

Results: The FD ratio showed a strong, negative linear correlation with the HFF after logarithmic transformation of both variables from the initial examinations of 36 patients ($\rho = -0.888$, $P < 0.001$) and the follow-up examinations of 27 patients ($\rho = -0.920$, $P < 0.001$). There was also a significant, negative linear correlation between the change in the logarithm of the FD ratio and the change in the logarithm of the HFF by MRS over the follow-up period ($\rho = -0.645$, $P < 0.001$). In 16 patients with an increased FD ratio on follow-up, the HFF on follow-up MRS significantly decreased, and high-density lipoprotein levels significantly increased, whereas low-density lipoprotein levels tended to decrease.

Conclusion: The FD ratio was significantly correlated with the HFF at both the initial and follow-up examinations, and there was also a significant correlation between changes in the FD ratio and changes in the HFF over the follow-up period.

Keywords: Hepatic steatosis; Acoustic structure quantification; Magnetic resonance spectroscopy; Monitoring hepatic steatosis

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Introduction

Hepatic steatosis is defined as the excessive and abnormal accumulation of intracellular lipids, primarily in the form of triglycerides. Clinically, the most common type of hepatic steatosis is nonalcoholic fatty liver disease (NAFLD), which includes a broad spectrum of diseases, ranging from simple steatosis to nonalcoholic steatohepatitis (NASH). In addition, NASH can progress to fibrosis/cirrhosis, and end-stage liver disease requiring liver transplantation can eventually develop when it is not appropriately managed [1–4]. Currently, NAFLD is the most common cause of chronic liver disease in Western countries, with a prevalence of approximately 30% in the general population, and the incidence of NAFLD is known to be increasing [5,6]. Therefore, NAFLD is a major emerging health problem, and the early detection and quantitative assessment of NAFLD are of crucial importance for preventing progression to advanced stages, such as cirrhosis.

Traditionally, liver biopsy has been considered the gold standard for the diagnosis and quantification of hepatic steatosis [7]. However, due to its invasive nature and procedure-related complications and mortality, repeated biopsies for monitoring NAFLD should rarely be performed. Therefore, hydrogen-1 (^1H) magnetic resonance spectroscopy (MRS), which is an accurate and reproducible method for assessing hepatic steatosis, has been used as an alternative, non-invasive standard of reference [8,9]. In addition, ^1H -MRS can be used to monitor hepatic steatosis [10]. However, its high cost and the limited availability of MRS compared to other non-invasive imaging modalities, such as ultrasonography (US), are disadvantages.

Recently, acoustic structure quantification (ASQ), which is a newly developed quantitative analysis method for US reported by Toshiba Medical Systems (Otawara, Japan), has been introduced to clinical practice. ASQ is based on comparisons between theoretical and real echo amplitude distributions [11–13], and the theoretical echo amplitude distribution of the liver region is considered using the Rayleigh distribution function, based on the assumption that the speckle pattern is generated by only ultrasound beam interference from very small scattering objects that are located closer than the ultrasound beam wavelength [11–13]. However, the real echo amplitude distribution of the liver parenchyma does not actually fit a Rayleigh distribution because of the presence of small structures, such as the hepatic vessel walls, which scatter the ultrasound beam and cause heterogeneity in the echo amplitude [11–13]. By comparing the theoretical echo amplitude distribution to a real distribution, ASQ can provide quantitative information regarding changes of parenchymal echotexture in diffuse liver diseases, such as steatosis. According to the results of recent studies, the focal

disturbance (FD) ratio calculated using the ASQ technique was significantly associated with the degree of hepatic steatosis assessed by histopathology, and there was a strong, negative correlation between the FD ratio and the estimated fat fraction assessed by MRS [12,13]. Given these findings, the FD ratio calculated using the ASQ technique could be used to monitor hepatic steatosis. However, the utility of the ASQ technique as a tool for monitoring hepatic steatosis has not been evaluated. Therefore, the purpose of our study was to prospectively evaluate whether monitoring hepatic steatosis using US with ASQ was feasible in a fatty liver disease patient cohort, using ^1H -MRS as the reference standard.

Materials and Methods

Study Population

The institutional review board of our hospital approved this prospective study, and written informed consent was obtained from all enrolled patients. From November 2014 to April 2015, we enrolled 36 patients who visited our institution for suspected or verified hepatic steatosis and received routine US examinations. The electronic medical records and past medical histories of all patients were reviewed to confirm that they did not have any past medical histories or clinical symptoms or signs of liver disease or systemic or malignant diseases. Laboratory findings, including liver function tests obtained within 3 months before the imaging examinations, were also checked in all of the patients. All patients underwent both US examinations that used the ASQ technique and ^1H -MRS examinations on the same day. After the initial imaging examination, lifestyle interventions were conducted in all of the enrolled patients, with dietary counseling aimed to reduce body weight. The patients were also instructed to perform at least 3 hours of moderate physical activity per week, and aerobic endurance exercise, including walking and swimming, was encouraged [14–17]. All of the enrolled patients were seen by a physician (M.S.P.) on a regular basis. After a mean and median follow-up of 11.4 ± 2.5 (standard deviation) months and 12.0 months (range, 6 to 15 months), respectively, follow-up US examinations using ASQ, ^1H -MRS examinations and laboratory tests, including liver function tests, were performed in 27 of the 36 patients to evaluate whether hepatic steatosis had improved; follow-up US and MRS examinations were done on the same day. Nine patients (9 of 36, 25.0%) refused to undergo follow-up imaging examinations. The characteristics of the patient cohort are summarized in Table 1.

US Examinations

All of the US examinations were performed by one of the two authors (J.Y.L. and D.H.L. with 20 and 9 years of experience,

respectively, in liver US imaging) using a US scanner (Aplio XG, Toshiba Medical Systems) with a 5-MHz convex transducer. All patients fasted for at least 6 hours before the imaging examinations. The US examinations were done with patients in the supine position with their right arm extended above their head to stretch the intercostal muscle for a better sonic window; the same patient position was used in both the initial and follow-up examinations. First, a B-mode US examination of the liver was performed, including transverse and sagittal images, to evaluate the presence of any possible focal liver lesions. Then, US images in ASQ mode were acquired 5 times each from the right intercostal view and the right subcostal view. All five regions of interest (ROIs) were placed in the same intercostal and right subcostal views. The display depth and transmit focus were set to 10 cm and 6 cm, respectively. The ROIs were as large as possible and were placed on the liver parenchyma; care was taken to avoid large hepatic vessels or artifacts. The value of the FD ratio was calculated by the system and was displayed

on the monitor (Fig. 1). The mean FD ratio of five measurements was calculated for each patient and recorded for further analysis. Detailed methods and a description of the ASQ technique and the calculation of the FD ratio are provided in the appendix.

¹H-MRS

All ¹H-MRS examinations were performed on a clinical 3.0-T MR scanner (Trio A Tim System, Siemens Healthcare, Erlangen, Germany) with a 32-channel phased-array surface coil on the same day of the US examinations. First, sagittal, coronal, and axial sections that covered the whole liver were obtained to position the spectroscopy acquisition voxel. Then, a signal voxel measuring 3×3×3 cm was placed in the right hepatic lobe dome area of Couinaud segment VII or VIII, with care taken to avoid large blood vessels or bile ducts and liver edges. For the fat fraction spectroscopy measurements, a modified stimulated-echo acquisition sequence (echo time [msec]/repetition time [msec]; 12, 24, 36, 48 and 72/3,000) was employed within the single voxel. Each acquisition was performed during a single breath hold with a scan time of 15 seconds. T2 correction for both the water and lipids was performed to calculate the hepatic fat fraction (HFF), and each T2 value of water and fat was calculated separately. By using an exponential fit of the points acquired at five different echoes by automatic fitting of water (4.7 ppm) and the major fat peak of methylene (1.3 ppm), extrapolation of fat and water integrals for an echo time of 0 msec was performed for fat quantification [8,18]. The calculated HFF was automatically displayed as a percentage in Distal Imaging and Communications in Medicine format and was recorded for further analysis.

Statistical Analysis

The estimated HFF by MRS and the FD ratio calculated using the ASQ technique are expressed as the mean±standard deviation throughout the results. We used Pearson correlation coefficients to assess the correlation between the FD ratio and the HFF. After the initial examination, 27 of the 36 patients underwent follow-up US examinations using the ASQ technique and ¹H-MRS examinations, and the changes in the FD ratio, as well as the estimated HFF by MRS, between the initial and follow-up examinations were calculated. Using the changes in FD ratio calculated by ASQ over the follow-up period, patients were classified into two groups depending on whether their FD ratio increased or decreased during follow-up. Differences in the estimated HFF on MRS, as well as in other characteristics of patients, between the initial and follow-up examinations were assessed by group using the Wilcoxon signed-rank test. All statistical analyses were performed using SPSS software version 21 (IBM Corp., Armonk, NY, USA), and MedCalc software version 12.1.0.0 (MedCalc Software, Mariakerke, Belgium).

Table 1. Baseline characteristics of the 36 patients with suspected fatty liver disease

Characteristic	Value
Age (yr)	50.4±13.3 (20–71)
Sex	
Male	21 (58.3)
Female	15 (41.7)
Diabetes	
Yes	10 (27.8)
No	26 (72.2)
Hypertension	
Yes	17 (47.2)
No	19 (52.8)
Hyperlipidemia	
Yes	21 (58.3)
No	15 (41.7)
Serum albumin (mg/dL)	4.5±0.2 (4.1–4.9)
Total bilirubin level (mg/dL)	0.90±0.31 (0.6–1.7)
Alanine aminotransferase (IU/dL)	49.6±43.2 (14–220)
Body weight (kg)	80.4±18.6 (55–124)
Body mass index (kg/m ²)	28.4±4.6 (22–39)
Total cholesterol (mg/dL)	194.2±38.5 (125–267)
Triglycerides (mg/dL)	164.8±92.3 (45–433)
Low-density lipoprotein (mg/dL)	124.3±44.3 (54–216)
High-density lipoprotein (mg/dL)	49.9±12.5 (32–90)
Estimated hepatic fat fraction by MRS (%)	16.7±9.7 (1.8–39.3)

Values are presented as mean±SD (range) or number (%).

MRS, magnetic resonance spectroscopy; SD, standard deviation.

Results

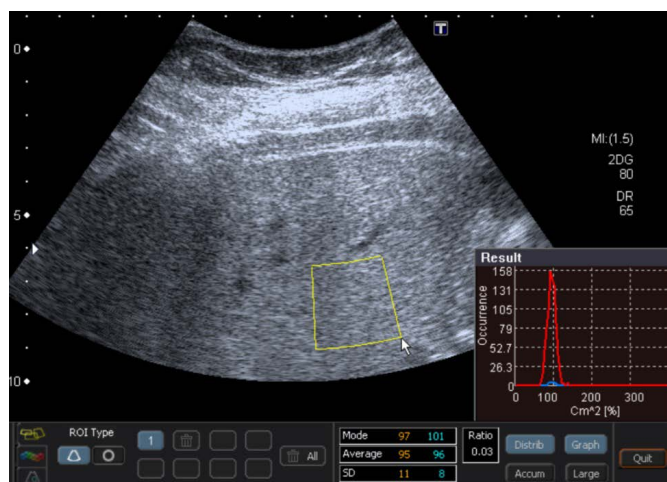
FD Ratio and Estimated HFF at the Initial Examinations

The mean estimated HFF at the initial MRS examinations of all 36 patients was $16.7\% \pm 9.7\%$ (range, 1.8% to 39.3%), and the mean FD ratio calculated using the ASQ technique at the initial US examinations was 0.07 ± 0.10 (range, 0.02 to 0.56). The correlation coefficient (ρ) between the FD ratio calculated using the ASQ technique and the estimated HFF by MRS was -0.526 (95%

confidence interval [CI], -0.239 to -0.729). However, a scatter plot of the estimated HFF and the FD ratio showed an exponential relationship. When these two variables were log-transformed, there was a strong, negative linear correlation between the logarithm of the FD ratio and the logarithm of the HFF (Fig. 2), with a correlation coefficient (ρ) of -0.888 (95% CI, -0.790 to -0.942 ; $P < 0.001$).

FD Ratio and Estimated HFF at the Follow-up Examinations

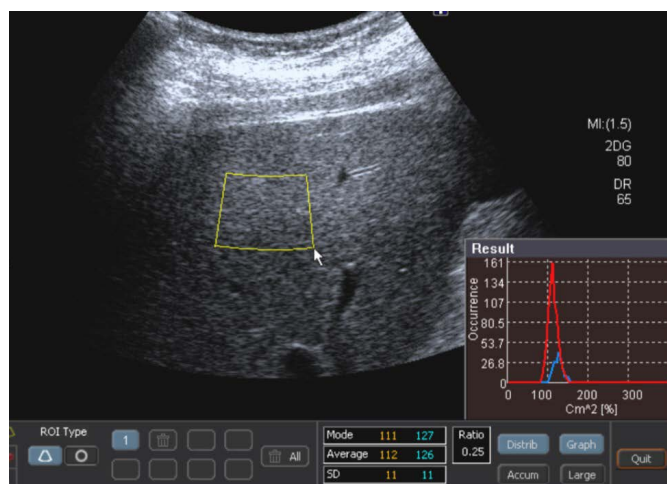
Follow-up US examinations using the ASQ technique and ^1H -MRS



A

Fat (%)	10.14% (95% CI, 9.95 to 10.34)		
R2 water	40.96/sec ($r^2=1.00$)		
	Water Sig.	Lipid Sig.	Lipid (%)
T2 corrected	$5.19e+07$	$5.86e+06$	10.14
TE=12	$3.25e+07$	$4.47e+06$	12.11
TE=24	$1.94e+07$	$3.98e+06$	17.01
TE=36	$1.16e+07$	$2.92e+06$	20.17
TE=48	$7.16e+06$	$2.63e+06$	26.86
TE=72	$2.77e+06$	$1.50e+06$	35.17
R2 (/sec)	40.96	18.30	
R ² fit	1.00	1.00	

B



C

Fat (%)	2.93% (95% CI, 2.75 to 3.11)		
R2 water	36.65/sec ($r^2=1.00$)		
	Water Sig.	Lipid Sig.	Lipid (%)
T2 corrected	$4.91e+07$	$1.48e+06$	2.93
TE=12	$3.15e+07$	$1.17e+06$	3.59
TE=24	$2.05e+07$	$7.35e+05$	3.47
TE=36	$1.34e+07$	$5.82e+05$	4.16
TE=48	$8.16e+06$	$4.74e+05$	5.49
TE=72	$3.55e+06$	$2.40e+05$	6.33
R2 (/sec)	36.65	25.16	
R ² fit	1.00	1.00	

D

Fig. 1. Initial and follow-up ultrasonography (US) examinations using the acoustic structure quantification (ASQ) technique and magnetic resonance spectroscopy (MRS) examinations in a 29-year-old man with fatty liver.

A. Upon initial examination, the patient's body weight was 115 kg, and he had a body mass index (BMI) of 36 kg/m^2 . On an ASQ-mode US image, the region of interest was placed on the liver parenchyma, and care was taken to avoid large hepatic vessels or artifacts. The focal disturbance (FD) ratio calculated by ASQ in this patient was 0.03. **B.** The estimated hepatic fat fraction (HFF) according to the initial MRS examination of this patient was 10.14%. **C.** During the 10-month follow-up period, the patient lost 21 kg of body weight, and his BMI at the follow-up examination was 30 kg/m^2 . The FD ratio calculated by ASQ for the follow-up examination was 0.25. **D.** The estimated HFF for the follow-up MRS examination was 2.93%. CI, confidence interval; TE, echo time.

examinations were performed on 27 patients, and the mean estimated HFF calculated at the follow-up MRS examinations was $14.0\% \pm 9.3\%$ (range, 1.0% to 32.0%). The mean FD ratio calculated using the ASQ technique at the follow-up US examinations was 0.10 ± 0.11 (range, 0.02 to 0.53). The correlation coefficient (ρ) between the FD ratio calculated using the ASQ technique for the follow-up examinations and the estimated HFF according to the follow-up MRS examinations in these 27 patients was -0.637 (95% CI, -0.338 to -0.819). A scatter plot of the estimated HFF and the FD ratio obtained from the follow-up examinations also showed an exponential relationship. There was a strong, negative linear correlation between the logarithm of the FD ratio obtained at the follow-up US examinations and the ASQ technique and the logarithm of the HFF of the follow-up MRS examinations, and the correlation coefficient (ρ) was -0.920 (95% CI, -0.831 to -0.963 ; $P < 0.001$) (Fig. 3).

Changes in the FD Ratio over the Follow-up Period and Correlation with the HFF

Among the 27 patients who underwent follow-up examination, the FD ratio calculated by ASQ was higher at the follow-up US examination than at the initial US examination in 16 patients and lower in 11 patients. The correlation coefficient (ρ) between the

change in the FD ratio and the change in the estimated HFF was -0.553 (95% CI, -0.219 to -0.771). A scatter plot of the change in the estimated HFF and the change in the FD ratio over the follow-up period also showed an exponential relationship. There was also a significant, negative linear correlation between the change in the logarithm of the FD ratio over the follow-up period and the change in the logarithm of the HFF over the follow-up period by MRS, and the correlation coefficient (ρ) was -0.645 (95% CI, -0.351 to -0.823 ; $P < 0.001$) (Fig. 4).

Changes in Patient Characteristics According to Changes in the FD Ratio

Changes in patient characteristics over the follow-up period according to changes in the FD ratio calculated using ASQ are summarized in Table 2. Body weight decreased over the course of follow-up, but a statistically significant difference was not found. The estimated HFF on follow-up MRS significantly decreased in the 16 patients with an increased FD ratio at follow-up ($P = 0.003$). In addition, high-density lipoprotein (HDL) levels significantly increased over follow-up in the 16 patients with an increased FD ratio ($P = 0.035$). There was a tendency for low-density lipoprotein (LDL) levels to decrease during follow-up in the 16 patients with increased FD ratio ($P = 0.084$). In the 11 patients with a decreased FD ratio, the

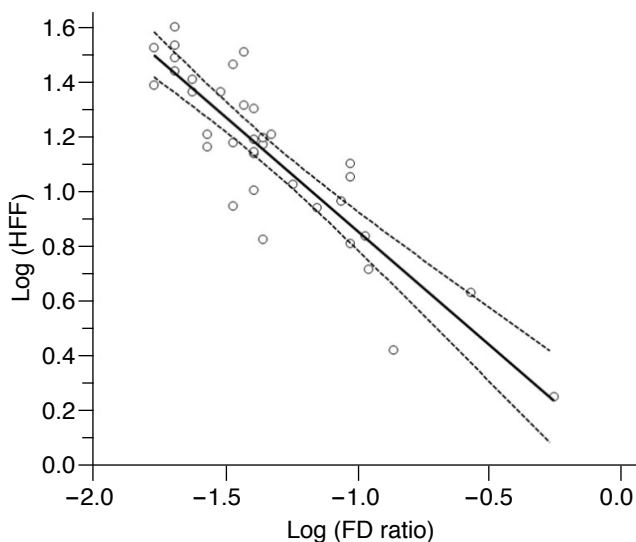


Fig. 2. Correlation between the focal disturbance (FD) ratio and the hepatic fat fraction (HFF) at the initial examinations of all 36 patients. Regression plot showing a strong negative linear correlation between the FD ratio and the HFF after the logarithmic transformation of both variables; the correlation coefficient was -0.888 ($P < 0.001$). The solid line indicates the regression line, and the lines with small dots are the 95% confidence intervals.

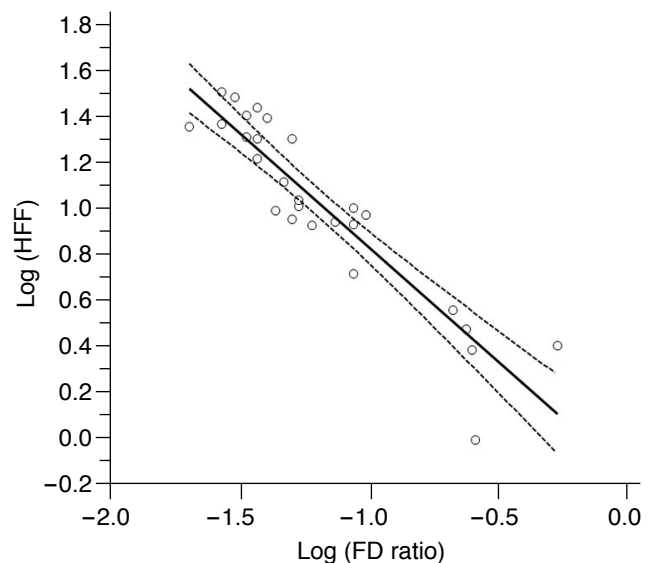


Fig. 3. Correlation between the focal disturbance (FD) ratio and the hepatic fat fraction (HFF) at the follow-up examinations of 27 patients. Regression plot showing a strong negative linear correlation between the FD ratio and the HFF after the logarithmic transformation of both variables; the correlation coefficient was -0.920 ($P < 0.001$). The solid line indicates the regression line, and the lines with small dots are the 95% confidence intervals.

Table 2. Changes in parameters in the 27 patients who underwent follow-up examinations according to the change in the FD ratio calculated by the ASQ technique at the follow-up examination

	Increased FD ratio on follow-up (n=16)			Decreased FD ratio on follow-up (n=11)		
	Initial	Follow-up	P-value ^{a)}	Initial	Follow-up	P-value ^{a)}
Body weight (kg)	82.8±16.9	79.8±13.5	0.158	76.1±16.8	74.2±16.7	0.260
Albumin (mg/dL)	4.6±0.2	4.5±0.2	0.287	4.5±0.2	4.5±0.3	0.887
Total bilirubin (mg/dL)	1.08±0.37	0.99±0.25	0.216	0.84±0.23	0.68±0.22	0.151
ALT (IU/dL)	59.8±56.5	47.8±32.9	0.780	34.6±16.8	30.0±12.6	0.406
Total cholesterol (mg/dL)	188.5±40.9	185.9±33.2	0.851	184.7±36.2	158.1±33.8	0.203
Triglycerides (mg/dL)	146.3±71.9	136.4±44.4	0.753	163.6±82.0	145.6±72.0	0.241
LDL (mg/dL)	128.6±44.4	109.8±35.7	0.084	108.3±38.4	100.9±37.3	0.241
HDL (mg/dL)	47.1±8.4	50.7±9.8	0.035	48.1±9.7	49.6±12.4	0.384
Fat fraction on MRS (%)	17.6±11.8	13.3±8.8	0.003	12.4±8.6	15.0±10.3	0.248

FD ratio, focal disturbance ratio; ASQ, acoustic structure quantification; ALT, alanine aminotransferase; LDL, low-density lipoprotein; HDL, high-density lipoprotein; MRS, magnetic resonance spectroscopy.

^{a)}The P-value was determined using the Wilcoxon signed-rank test.

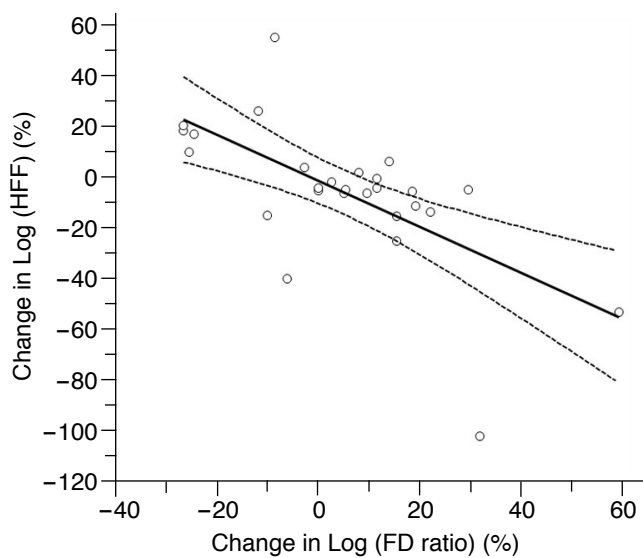


Fig. 4. Correlation between changes in the focal disturbance (FD) ratio and the hepatic fat fraction (HFF) over follow-up in the 27 patients who underwent follow-up examinations. Regression plot showing a significant negative linear correlation between changes in the logarithm of the FD ratio and the logarithm of the HFF; the correlation coefficient was -0.645 ($P < 0.001$). The solid line indicates the regression line, and the lines with small dots are the 95% confidence intervals.

estimated HFF increased on follow-up MRS, without a statistically significant difference ($P = 0.248$). Other patient characteristics were not significantly different at the follow-up examinations in the 11 patients with a decreased FD ratio.

Discussion

In our study, the FD ratio calculated using the ASQ technique showed a significant, negative correlation with the estimated HFF by MRS at both the initial and follow-up examinations, corresponding closely to the findings of a previous study by Son et al. [13]. In addition, there was a significant correlation between the change in the FD ratio over the follow-up period and the change in the HFF over the follow-up period by MRS. An increased FD ratio on follow-up US with the ASQ technique compared to the initial examination was significantly associated with a decreased HFF on follow-up MRS. Moreover, HDL levels significantly increased and LDL levels tended to decrease in the 16 patients with an increased FD ratio on the follow-up examination, which could indicate improvement of their lipid panels.

Because NAFLD encompasses a broad spectrum, ranging from simple steatosis to cirrhosis, and can progress into more advanced stages during the disease course, the quantitative assessment and monitoring of this disease are important for patient management. US is a non-invasive imaging modality for assessing liver disease with a relatively lower cost compared to magnetic resonance imaging, and it can be performed repeatedly during the disease course with no risk to the patient. Given this finding, US could potentially be a good imaging modality for monitoring fatty liver disease. When the echogenicity of the liver parenchyma increases compared to that of the renal cortical parenchyma on B-mode US, it is usually considered to indicate the presence of hepatic steatosis. However, the assessment of hepatic parenchymal echogenicity by B-mode US is subjective in nature. In addition, quantifying the

degree of hepatic steatosis is only barely possible by using B-mode US images alone. To overcome these drawbacks of B-mode US, several amendments, including the hepatorenal ratio, have been suggested for the quantitative assessment of hepatic steatosis, with relatively good performance [19–21]. However, although Mancini et al. [22] reported that the hepatorenal ratio showed an excellent correlation with the degree of steatosis evaluated by MRS, whether this ratio has sufficient diagnostic performance for assessing hepatic steatosis or could be used as an alternative tool to MRS remains questionable [13]. In addition, other studies have reported that additional processes, such as standardization or use of artificial neural networks, would be needed for the hepatorenal ratio to attain a sufficiently high diagnostic performance for the quantitative assessment of hepatic steatosis [21,23].

In the ASQ technique, the theoretical echo amplitude distribution, which is believed to be a function of the Rayleigh distribution, is compared to a real echo amplitude distribution, which does not actually fit a Rayleigh distribution, mainly due to the presence of small structures, including hepatic vessels, that scatter the US waves [11–13]. As the degree of hepatic steatosis progresses, the small structures in the liver, including small vessels, are blurred with the increased hepatic parenchymal echogenicity; in other words, hepatic parenchymal echogenicity might become more homogeneous. In such cases, the real echo amplitude distribution of the liver parenchyma can become similar to the theoretical echo amplitude distribution (i.e., the Rayleigh distribution) [13]. Comparing the theoretical echo amplitude distribution to a real distribution, the FD ratio calculated using the ASQ technique could provide quantitative information regarding the hepatic parenchymal echo texture changes that occur during the progression of hepatic steatosis. Our study results clearly showed a significant correlation between the FD ratio calculated by the ASQ technique and the estimated HFF by MRS at both the initial and follow-up examinations. In addition, the change in the FD ratio over the follow-up period was significantly correlated with the change in the estimated HFF by MRS, and an increased FD ratio on follow-up examination was significantly associated with a decreased HFF on follow-up MRS. Considering this result, the FD ratio calculated using the ASQ technique could potentially be used to quantitatively monitor fatty liver disease.

Currently, NAFLD is considered to be a hepatic manifestation of metabolic syndrome, which is characterized by the concomitant existence of type 2 diabetes, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease [24]. In addition, Sinn et al. [25] recently reported that NAFLD was significantly associated with the development of coronary calcified plaque, independent of cardiovascular and metabolic risk factors. Therefore, accurate evaluation and monitoring of NAFLD could be quite important for

reducing the potential mortality and morbidity from cardiovascular disease. In this regard, quantitative assessment of hepatic steatosis using US with the ASQ technique would be helpful in the management of NAFLD patients, as our study results showed that the FD ratio calculated using the ASQ technique was significantly correlated with the estimated HFF on MRS, which is a reference standard for non-invasive methods of diagnosing NAFLD. In addition, an increased FD ratio at the follow-up examination was also associated with significantly increased HDL levels and a tendency for decreased LDL levels, potentially indicating improvements in the lipid panel. Considering our study results, US examinations using the ASQ technique could be a promising monitoring method not only for hepatic steatosis, but also for metabolic syndrome.

Our study had some limitations. First, in our study, ¹H-MRS examinations, instead of histologic examinations, were used as the reference standard to quantitatively assess hepatic steatosis. Therefore, the relationship between the FD ratio calculated using the ASQ technique and histologic changes could not be evaluated in our study. However, many clinical trials use ¹H-MRS as the reference standard to assess the degree of hepatic steatosis due to its accuracy and its quantitative and noninvasive nature. Second, the number of patients was relatively small, especially those undergoing follow-up examinations. Third, we did not perform liver biopsy or elastography. Therefore, we could not assess the effect of fibrosis or inflammation on the FD ratio. In addition, according to the results of the recent animal study conducted by Lee et al. [26], the degrees of both steatosis and fibrosis were significant factors affecting the FD ratio calculated by ASQ, although the correlation coefficient for steatosis was higher than that for fibrosis. Indeed, it is unclear whether the FD ratio calculated using the ASQ technique can provide good diagnostic performance for quantitatively assessing hepatic steatosis in patients with both steatosis and inflammation/fibrosis. Further studies with prospective designs and large numbers of patients with various stages of fatty liver disease, including steatosis and cirrhosis, are warranted to address this issue. Fourth, as the MRS signal voxel was placed on the right liver dome, where accurate US evaluation would be difficult, there could have been some differences between the area evaluated using the ASQ technique and the area assessed by MRS; therefore, there may be some limitations regarding the accuracy of the correlation between the FD ratio calculated by ASQ and the estimated HFF by MRS.

In conclusion, the FD ratio calculated using the ASQ technique was significantly correlated with the estimated HFF by MRS at both the initial and follow-up examinations, and there was a significant correlation between the changes in the FD ratio and the changes in the HFF over the follow-up period, suggesting that the FD ratio can be used to quantitatively monitor fatty liver disease.

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Conflict of Interest

No potential conflict of interest relevant to this article was reported.

Author Contributions

Conceptualization: Lee JY. Data acquisition: Lee DH, Lee JY, Park MS. Data analysis or interpretation: Lee DH. Drafting of the manuscript: Lee DH. Critical revision of the manuscript: Park MS, Han JK, Lee JY. Approval of the final version of the manuscript: all authors.

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