



# **Pro-Inflammatory Profile of Adipokines in Obesity Contributes** to Pathogenesis, Nutritional Disorders, and Cardiovascular Risk in Chronic Kidney Disease

Sylwia Czaja-Stolc<sup>1,\*</sup>, Marta Potrykus<sup>2</sup>, Marta Stankiewicz<sup>1</sup>, Łukasz Kaska<sup>2</sup> and Sylwia Małgorzewicz<sup>1</sup>

- <sup>1</sup> Department of Clinical Nutrition, Medical University of Gdansk, 80-211 Gdańsk, Poland; marta.stankiewicz@gumed.edu.pl (M.S.); sylwia.malgorzewicz@gumed.edu.pl (S.M.)
- <sup>2</sup> Department of General, Endocrine and Transplant Surgery, Medical University of Gdansk, 80-211 Gdańsk, Poland; martapotrykus@gumed.edu.pl (M.P.); lukasz.kaska@gumed.edu.pl (Ł.K.)
- Correspondence: sylwia.czaja-stolc@gumed.edu.pl; Tel.: +48-(58)-349-27-24

Abstract: Obesity is a disease which leads to the development of many other disorders. Excessive accumulation of lipids in adipose tissue (AT) leads to metabolic changes, including hypertrophy of adipocytes, macrophage migration, changes in the composition of immune cells, and impaired secretion of adipokines. Adipokines are cytokines produced by AT and greatly influence human health. Obesity and the pro-inflammatory profile of adipokines lead to the development of chronic kidney disease (CKD) through different mechanisms. In obesity and adipokine profile, there are gender differences that characterize the male gender as more susceptible to metabolic disorders accompanying obesity, including impaired renal function. The relationship between impaired adipokines in the serum is additionally disturbed due to their insufficient excretion by the excretory system caused by renal pathology. Increased levels of adipokines affect the nutritional status and cardiovascular risk (CVR) of patients with CKD. This article aims to systematize the current knowledge on the influence of obesity, AT, and adipokine secretion disorders on the pathogenesis of CKD and their influence on nutritional status and CVR in patients with CKD.

Keywords: obesity; adipokines; chronic kidney disease; malnutrition; cardiovascular risk

# 1. Introduction

The global epidemic of overweight and obesity has been constantly increasing. According to the World Health Organization (WHO), the prevalence of obesity in the world has almost tripled in the last fifty years. In 2016, over 650 million adults were obese [1]. Newly emerging publications indicate that the COVID-19 pandemic, and the accompanying lockdown, contributed to the exacerbation of obesity [2,3]. Excessive body weight contributes to the development of many diseases, such as cardiovascular diseases (CVD), diabetes, and chronic kidney disease (CKD). Treatment of obesity and diseases caused by it is associated with a significant financial burden for the health care system [4,5].

The mean global incidence of CKD is 13.4%, but many patients go undiagnosed [6]. Patients with CKD are characterized by higher mortality compared to the general population. The main cause of death in patients treated conservatively and undergoing dialysis are cardiovascular diseases [7]. The risk of developing CVD is 5–10 times higher in the CKD population compared to people with normal kidney function. Traditional risk factors such as lipid disturbances, diabetes, and hypertension do not explain this phenomenon [8]. Furthermore, untraditional risk factors such as chronic inflammation, hypoalbuminemia, or oxidative stress, included in protein-energy wasting (PEW), have a negative impact on survival [9].



Citation: Czaja-Stolc, S.; Potrykus, M.; Stankiewicz, M.; Kaska, Ł.; Małgorzewicz, S. Pro-Inflammatory Profile of Adipokines in Obesity Contributes to Pathogenesis, Nutritional Disorders, and Cardiovascular Risk in Chronic Kidney Disease. *Nutrients* **2022**, *14*, 1457. https://doi.org/10.3390/ nu14071457

Academic Editors: Francesca Di Sole and Maria J. Barnes

Received: 18 February 2022 Accepted: 29 March 2022 Published: 31 March 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Obesity increases the risk of developing CKD in many ways. For instance, it contributes to chronic inflammation, oxidative stress, impaired secretion of adipokines and lipotoxicity, which results in nephron damage [10,11]. The aim of the study is to review and summarize the literature on obesity, adipose tissue (AT), and adipokines, and their impact on the risk of developing CKD as well as the occurrence of nutritional disorders and cardiovascular risk (CVR) in the above-mentioned group of patients. To find articles matching the topic of the manuscript, PubMed and WorldWideScience databases were searched for keywords "obesity", "adipose triglyceride lipase", "cardiovascular risk", and their various combinations. The bibliography of selected publications was also searched. It has been established that the articles considered in the search for information for the manuscript are to be published in the period from 2000 to 2022. All publications published earlier than 2000 and/or written in languages other than English and Polish were excluded.

#### 2. Heterogeneity of Adipose Tissue

AT, as an endocrine organ, is a source of various substances including adipokines [12]. AT is a miscellaneous tissue which consists of adipocytes, preadipocytes, fibroblasts, immune cells, epithelial cells of the blood and lymphatic systems, and stem cells [13,14].

Depending on the structure and function of adipocytes, AT is divided into white, brown, and beige tissues. White adipose tissue (WAT) stores fatty acids as an energy source and consists of one large, centrally placed droplet of triglycerides (TG) and other cell organelles pushed to the cell periphery [15]. In turn, adipocytes included in brown adipose tissue (BAT) contain numerous smaller droplets of fatty acids and a high number of mitochondria. BAT is rich in the mitochondrial protein—uncoupling protein 1 (UCP1), which dissipates energy in the form of heat and thus increases energy expenditure. Unlike WAT, BAT reduces hyperlipidemia and facilitates weight loss [16,17]. It is possible to induce UCP1 expression in white adipocytes, a process known as adipocyte browning. The cells created in this way are beige adipocytes which, despite their different origins, have a similar structure and function to brown adipocytes. Browning of adipocytes induced by various factors, including exposure to cold, insulin, specific nutrients, or the presence of an excess of nutrients, has a positive effect on health [17,18].

Additionally, WAT can be classified based on its distribution in the body. Although it is found almost throughout the body, there are several places where it is most abundant. These include subcutaneous, pericardial, gonadial, and visceral adipose tissue (VAT) divided into mesenteric, omental, and retroperitoneal AT [19,20]. As already mentioned, AT is scattered throughout the body. This type of AT is called discrete tissue-associated adipose depots. They are small clusters that are closely related to other anatomical structures and perform functions specific to these structures [19].

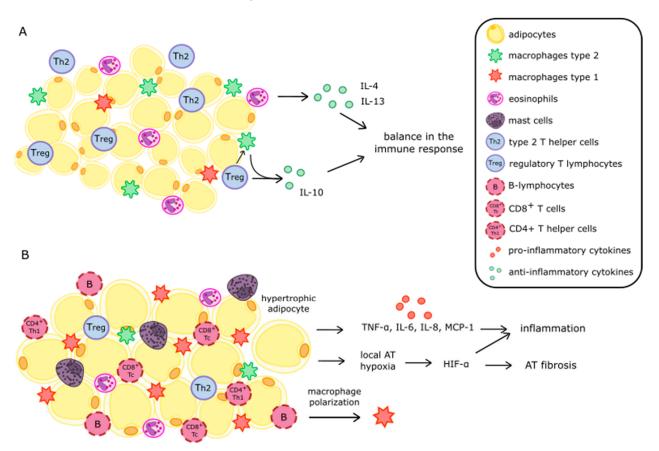
#### 3. Obesity-Induced Changes in Adipose Tissue

Obesity is characterized by an increase in and dysregulation of WAT, especially visceral fat, which is associated with altered adipokine secretion. When the threshold of adipocyte storage capacity for fatty acids is exceeded, the processes of hypertrophy and hyperplasia take place [17]. An increased size of fat cells (hypertrophy) and their increased number (hyperplasia) differ in the way they affect the organism. Hypertrophy is associated with metabolic disturbances, while hyperplasia shows protective activity [21]. Under conditions of positive energy balance, the size and number of adipocytes increases. Increased adipocytes secrete some hormones and cytokines that affect both pre-adipocyte recruitment and pre-adipocyte differentiation into adipocytes. In the case of a chronic positive energy balance, adipocytes become lipid-overloaded and reach a critical size, which leads to inhibition of cellular multiplication and an imbalance between the processes of hypertrophy and hyperplasia [22].

The hypertrophic adipocytes secrete pro-inflammatory cytokines such as tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin 6 (IL-6), interleukin 8 (IL-8), and monocyte chemoattractant

protein 1 (MCP-1) which, through the phosphorylation of insulin receptor substrate-1 (IRS-1), cause insulin resistance. The secretion of these cytokines also recruits macrophages and T-cells, which fuel inflammatory processes. Enlarged adipocytes also cause local tissue hypoxia, which results in the release of hypoxia-inducible factor (HIF- $\alpha$ ), leading to inflammation and fibrosis of the AT. If the capacity of the adipocytes is exceeded, lipids start to accumulate in the ectopic tissues, which deteriorates the functioning of these organs and/or tissues [22].

AT is characterized by the presence of various immune cells such as type 2 T-helper cells (Th2), regulatory T-lymphocytes (Treg), eosinophils and type 2 macrophages (M2 macrophages). Treg causes the secretion of anti-inflammatory interleukin 10 (IL-10) and promotes the formation of M2 macrophages which suppress inflammation through the secretion of IL-10. Eosinophils are also characterized by the secretion of anti-inflammatory cytokines interleukin 4 (IL-4) and interleukin 13 (IL-13). This composition of immune cells allows for homeostasis and anti-inflammatory effects [23]. Obesity causes disturbances in the composition of the immune system cells in AT. Hypertrophic adipocytes cause monocytes to migrate to AT and transform them into type 1 macrophages (M1 macrophages). The number of B lymphocytes also increases, which enhances the formation of M1 macrophages. A decrease in the number of eosinophils causes a fall in the secretion of anti-inflammatory cytokines—IL-4 and IL-13. In addition, obesity diminishes the amount of Treg in AT, which results in an increase in cytotoxic CD4<sup>+</sup>Th1 and CD8<sup>+</sup>Tc [23,24]. Figure 1 compares AT in people with obesity and normal body weight and the consequences of their metabolic activity.



**Figure 1.** Comparison of adipose tissue of humans with normal body weight (**A**) and with obesity (**B**). Abbreviations: AT—adipose tissue; IL-4,-6,-8,-10,-13—interleukin -4,-6,-8,-10,-13; TNF- $\alpha$ —tumor necrosis factor  $\alpha$ ; MCP-1—monocyte chemoattractant protein-1; HIF- $\alpha$ —hypoxia-inducible factor  $\alpha$ .

An increased amount of WAT, through increased release of pro-inflammatory cytokines, reduces BAT activity by inhibiting the induction of UCP1. It is associated with a decreased energy expenditure and might be another reason for difficulties in losing and maintaining weight, and obesity [17,25].

#### 4. Sexual Dimorphism in the Obesity-Induced Metabolic Profile

The central distribution of AT typical of men is correlated with a greater incidence of metabolic disorders than the gluteal–femoral distribution of AT. Additionally, a pear-type silhouette is associated with a reduced risk of metabolic disease and may have a protective effect against the consequences of obesity in both sexes [26].

Estrogens, which dominate in women, increase the level of leptin and other anorexic compounds such as cholecystokinin, brain-derived neurotrophic factor (BDNF), and apolipoprotein A-IV, and simultaneously reduce orexigenic substances, including ghrelin and melanin-concentrating hormone. The protective effect of estrogens against obesity is not only limited to reduced food consumption but also increased energy expenditure [27].

Gender not only affects the distribution and type of AT but also affects the type of metabolic activity of AT. Estrogens reduce inflammation in AT and make it more insulinsensitive. Estrogens reduce the activity of HIF- $\alpha$ , which is responsible for fibrosis and inflammation, by promoting the transcription of the enzyme prolyl hydroxylase domain enzyme 3 (PHD3) [27].

In turn, testosterone inhibits the activity of lipoprotein lipase, which reduces the uptake of fatty acids in the abdominal fat tissue. Chronic overexposure to free fatty acids (FFAs) in the circulation increases hepatic glucose production and elevates insulin production. It leads to a decrease in tissue sensitivity to insulin. This phenomenon is one of the reasons why men are more susceptible to insulin resistance and metabolic syndrome [26].

The greater deposition of VAT in men, and therefore a greater risk of high blood pressure and CVD, may be due to higher testosterone levels in men than in women. In addition, in postmenopausal women, decreased levels of estrogens also lead to increased VAT and higher blood pressure [28].AT concentrated around the kidneys is called perirenal adipose tissue (PRAT) and is part of visceral white adipose tissue. The morphology of PRAT is gender-dependent. In males, PRAT has a greater volume and thickness than in females with the same waist circumference. Additionally, in women, the expression of UCP1 in PRAT is higher than in men [29].

In the murine model, it was observed that the presence of the Y chromosome inhibits the expression of UCP1 in BAT. However, Chen et al. suggest that it is not the presence of the Y chromosome that leads to the gender differences in AT, but the increased number of X chromosomes. The authors showed that having a set of two X chromosomes led to a higher body weight to fat mass ratio compared to one X chromosome regardless of the number of Y chromosomes [30].

In people with obesity, there is sexual dimorphism in the levels of adipokines and their correlation with metabolic disorders. Walicka et al. observed that women had higher levels of adiponectin, leptin, and visfatin. In women, there was a negative correlation between the levels of adiponectin and insulin and the homeostasis model assessment of insulin resistance (HOMA-IR) index. In turn, in men, there was a positive correlation between the level of leptin and the HOMA-IR index [31]. It has been suggested that the leptin-to-adipokine (Lep/Adpn) ratio is an indicator of metabolic disorders. In the study by Selthofer-Relatić et al., in women, this ratio positively correlated only with anthropometric measurements. In men, however, the higher the Lep/Adpn ratio, the worse the results of total cholesterol, low-density lipoprotein (LDL) fraction, and TG are. This suggests that differences in the secretion of adipokines in different sexes affect the neurohormonal activity of AT. This leads to differences in the metabolic profile [32].

Females, at least before menopause, are more protected against obesity and inflammatory changes in AT. They are also more likely to maintain the proper functioning of kidneys. Across all classes of CKD, men show more kidney damage and a faster progression to end-stage renal disease (ESRD) compared to premenopausal women. After menopause, this effect wears off [33].

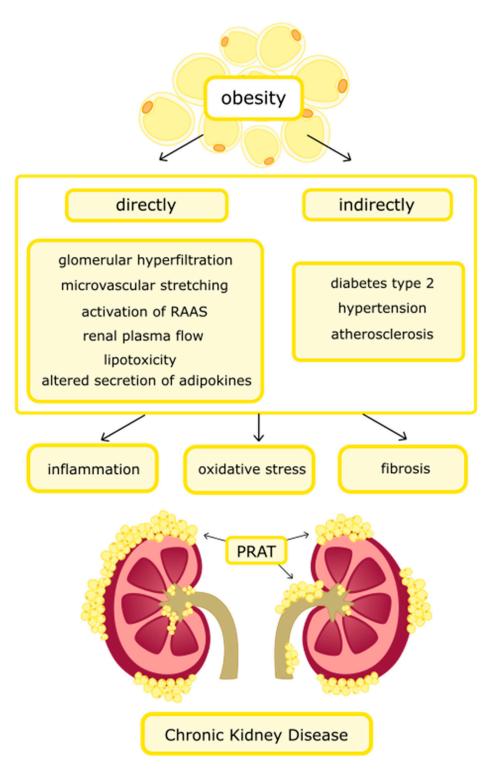
#### 5. Obesity and Chronic Kidney Disease

CKD is defined by the disturbance of the function or structure of the kidneys lasting more than three months. CKD is divided into five stages based on the value of the glomerular filtration rate (GFR) [34]. It is estimated that the mean global incidence of CKD is 13.4%, but many patients go undiagnosed and find out about the disease in the late stages of development [6].

Overweight and obesity contribute to the development of CKD among 15–30% of patients, but the mechanism of kidney damage is not fully understood [35]. Obesity increases the risk of developing CKD in a direct and indirect way, as is shown in Figure 2. The direct effect is associated with glomerular hyperfiltration (abnormally high GFR), increased renal plasma flow, microvascular stretching, renin-angiotensin-aldosterone system (RAAS) activation, altered secretion from AT, and lipotoxicity. All of these factors promote the formation of inflammation, oxidative stress, and fibrosis in the kidneys [11,36]. The pathophysiological mechanism described above leads to the development of obesity-related glomerulopathy (ORG) and is characterized by glomerulomegaly. If it is accompanied by glomerulosclerosis, it is called focal and segmental glomerulosclerosis (FSGS) [37]. The first symptom of ORG is usually microalbuminuria or clinically dominant proteinuria [38]. These disorders lead to progressive damage to nephrons and the development of CKD. The incidence of it is steadily increasing, however, due to relatively infrequent renal biopsy performed, exact statistics are not known [39,40]. Research is underway to discover new biomarkers of kidney damage. One of them is micro-RNA, which can be measured in the blood [41]. The indirect effect of CKD development is related to hypertension, atherosclerosis, and diabetes type 2 [35].

Hyperfiltration occurs as a result of dilatation of the afferent arteriole to the glomerulus, which is associated with an increase in reabsorption of glucose and sodium via sodium-glucose co-transporter–1 (SGLT1) and sodium–glucose co-transporter–2 (SGLT2), which leads to renal hypertrophy. The phenomenon manifested by an increase in the volume of glomerular tuft/capsule, tubular epithelium/lumen, and podocyte injury. These changes consequently lead to glomerulosclerosis [42,43]. A physiological glomerular hyperfiltration occurs after consuming a large amount of protein, sodium, and during pregnancy [44,45]. Glomerulomegaly is another renal pathology associated with obesity that occurs independently of microalbuminuria. These processes lead to impaired kidney function and complications associated with it [11,45].

Lipotoxicity is defined as the ectopic accumulation of lipids in organs other than AT. PRAT and accumulation of fatty acids in renal parenchyma cause the damage of tubulointerstitial tissue, proximal tubular epithelial cells, and endothelial cells [46]. Non-esterified fatty-acid (NEFA) accumulation leads to insulin resistance and apoptosis of podocytes which cannot regenerate [39,47,48]. Insulin resistance and hyperinsulinemia lead to activation of the sympathetic nervous system, which was confirmed by an increase in the concentration of catecholamines, and contributes to the occurrence of vasoconstriction [49]. Cumulation of NEFA in renal cells activates macrophages, which leads to transformation into foam cells [46]. Renal steatosis leads to the development of inflammation and fibrosis. Lipotoxicity contributes to decreased production of adenosine 5'-triphosphate (ATP) through inhibition of  $\beta$ -oxidation, thereby damaging the structure of the mitochondria [50]. PRAT compresses the kidneys, which causes an increase in hydrostatic pressure, reduces renal blood flow, and leads to CKD [51]. The influence of adipokines on the risk of developing CKD is described in the next section.



**Figure 2.** The direct and indirect impact of obesity on the development of chronic kidney disease. Abbreviations: RAAS—renin-angiotensin-aldosterone system; PRAT—perirenal adipose tissue.

# 6. Adipokines and Their Influence on Pathogenesis, Nutritional Disorders, and Cardiovascular Risk in Chronic Kidney Disease

# 6.1. Leptin

Leptin is a plasma protein encoded by the obesity gene (ob) that was first described in 1994. It is an anorexigenic hormone that increases energy expenditure. In obesity, increased concentration of leptin occurs, simultaneously with leptin resistance [52]. Reduced leptin activity on the hypothalamus impairs appetite suppression [53].

Enlarged adipocytes are characterized by increased secretion of leptin. Thus, adipokine, by binding to leptin receptors in the central nervous system, causes increased activity of the sympathetic nervous system, which, consequently, can lead to obesity-related hypertension [34]. Hypertension is inextricably linked with kidney failure. Although hypertension is one of the greatest risk factors for the development of CKD, it is also a result of impaired renal function. This is manifested by the fact that with the progression of CKD, the incidence of hypertension increases [54].

Leptin increases the expression of the transforming growth factor- $\beta$  1 (TGF- $\beta$ 1) gene and other fibrotic factors, such as collagen IV and fibronectin, which stimulate the proliferation of mesangial cells in the kidneys [55]. Excessive production of these cells can lead to glomerulosclerosis through mesangial hypertrophy in the glomeruli, thickening of the glomerular basement membrane, and increased extracellular matrix [11]. In turn, glomerulosclerosis, by increasing the permeability of the glomerular barrier, contributes to proteinuria and impaired renal function [56].

Leptin-induced vascular endothelial dysfunction is another mechanism that links altered adipokine secretion in obesity with impaired renal function. Leptin stimulates the formation of reactive oxygen species (ROS) that impair the vascular response to acetyl-choline, thereby initiating reduced bioavailability of nitric oxide and oxidative stress [57]. In addition, leptin modulates the immune system response. The hormone increases the number of T-helper cells and reduces the number of Treg. It also increases the phagocytic activity of macrophages and increases the TNF- $\alpha$ , IL-6, and interleukin 12 (IL-12). In the case of hyperleptinemia, the balance of pro- and anti-inflammatory processes is disturbed and inflammation occurs [58].

In CKD, metabolic degradation in the renal tubules and glomerular filtration is disturbed, which leads to an increase in the concentration of leptin in the blood. The concentration of leptin increases with the progression of the disease [11,59,60]. Korczyńska et al. observed increased expression of leptin in subcutaneous adipose tissue (SAT) among 5th stage CKD pre-dialysis and dialyzed patients [61]. In stages 3–5 of CKD, without dialysis, altered adipokines profile and insulin resistance were associated with VAT, SAT, and intrahepatic fat [62]. Leptin levels are higher in hemodialysis (HD) women than HD men [63].

Leptin influences nutritional status and CVR among the mentioned group of patients. Patients with CKD are at risk of developing PEW, which is associated with a high risk of mortality. The prevalence of PEW among dialyzed patients is up to 80% [64]. One of the mechanisms influencing their occurrence is leptin concentration disturbances. Leptin has catabolic properties such as increasing the metabolic rate and inducing anorexia [65,66]. The secretion of neuropeptides and neurotransmitters involved in the regulation of appetite occurs as a result of Janus Kinase-2 (JAK2) activation caused by the association of leptin with its receptors (ObRb). Leptin increases the secretion of  $\alpha$ -melanocyte-stimulating hormone ( $\alpha$ -MSH) and cocaine- and amphetamine-regulated transcript (CART), stimulating feeling of satiety, inhibits the synthesis of secretion of one of the strongest appetite stimulants, i.e., neuropeptide Y (NPY) [67–69]. The concentration of Serum NPY increases, but cerebrospinal fluid (CSF) NPY decreases with the progression of CKD. A low concentration of CSF NPY was associated with cachexia, reduced energy intake, and muscle-wasting among CKD patients [70]. It has been observed that higher serum NPY levels prognosticate cardiovascular mortality among dialysis patients [71,72].

Research has also shown that high levels of leptin among CKD patients are associated with inadequate energy and protein intake, and also with reduced levels of muscle mass [9]. Markaki et al. observed that higher leptin levels in HD and peritoneal dialysis patients were associated with greater fat mass index (FMI) and female gender [73]. On the other hand, other research suggests that patients with PEW have lower leptin levels, which could be related to decreased fat mass. Leptin levels are positively correlated with inflammatory markers, which also affects the risk of malnutrition development [74,75]. More research is needed to understand the mechanism by which leptin affects the occurrence of PEW.

It has been observed that higher serum NPY levels prognosticate cardiovascular mortality among dialysis patients. Leptin contributes to CVD by causing oxidative stress, inflammation, and endothelial cell proliferation [76]. This adipokine increases platelet aggregation and affects the concentration of vascular endothelial growth factor (VEGF), which leads to angiogenesis. Vascular endothelial dysfunction contributes to the development of atherosclerosis, which is the basis of many cardiovascular diseases [60,77]. Furthermore, leptin promotes cardiac hypertrophy by mitogen-activated protein kinase (MAPK) signaling [78].

Lu et al. observed that higher serum leptin levels among patients with CKD were a risk factor of aortic stiffness, measured as the carotid–femoral pulse wave velocity (cfPWV). The study was conducted among 205 patients with CKD stage 3–5 without dialysis and kidney transplantation (KT). Other risk factors of aortic stiffness were higher systolic blood pressure (SBP) and older age [79]. Similar research results were obtained in HD patients and those after KT. In addition, in Kuo et al.'s study, elevated serum leptin level among HD patients was correlated with body mass index (BMI) and fat mass [80,81]. One study found that leptin levels were associated with CVR among HD patients only with a larger waist circumference (>102 cm) [82].

However, low leptin concentration is an independent predictor of mortality among CKD patients. This was confirmed in a study carried out on HD patients and kidney transplant recipients (KTRs). The cohort study involving 1214 KTRs showed that, despite the adverse impact of high leptin level on graft function and severity of inflammation, the risk of mortality was 10% lower for every 10 ng/mL higher leptin concentration [83,84].

## 6.2. Adiponectin

Adiponectin, a 244 amino acid protein, plays an important role in insulin sensitivity by increasing fatty-acid oxidation and reducing gluconeogenesis. Adiponectin has two types of receptors—adipoR1 and adipoR2 [85,86]. The former is found in all tissues and the latter mainly in the liver. The adipoR1 receptors in the excretory system are present in the proximal tubule cells and the glomerulus cells, i.e., in the cells of the endothelium, podocytes, mesangium, and the epithelium of Bowman's capsule. Adiponectin crosses the glomerular filtration barrier, binds to the adipoR1 receptor on the above-mentioned structures and acts on them by activating the adenosine monophosphate-activated protein kinase (AMPK) pathways [86]. AMPK stimulation induces ATP formation processes, including fatty acid oxidation, which protects against obesity and metabolic disorders, and thus indirectly protects the proper functioning of the kidneys. However, in obesity, the concentration of adiponectin is reduced. Kidneys are exposed to the harmful effects of increased levels of pro-inflammatory adipokines and are devoid of the protective effects of adiponectin. Adiponectin may also play a role in the development of obesity-related albuminuria [86–88].

The reduced concentration of adiponectin causes translocation of the zonula occludens (ZO-1) proteins from the podocyte epithelial layer into the cytosol. The ZO-1 proteins are structural tight-junction proteins that are responsible for the tightness of the epithelium and the proper function of slit diaphragms. The slit diaphragm is the structure found between the foot processes of the podocytes and acts as a sieve to prevent high-molecular-weight proteins from entering the urine. The structural proteins of slit diaphragms are nephrin and podocin, which determine the proper functioning of this structure. Additionally, low levels of adiponectin have been associated with low levels of nephrin in the renal cortex, which increases the permeability of the filtration barrier [89]. Adiponectin has opposite actions to leptin. Contrary to the action of leptin, decreased levels of adiponectin result in increased synthesis of TGF- $\beta$ 1. This results in cell hypertrophy and increased collagen synthesis, which causes renal fibrosis [90].

Adiponectin has a protective effect on the vascular endothelium. It inhibits the action of the endothelial transcription factor—nuclear factor-kappa  $\beta$  (NF- $\kappa\beta$ ), through the activation of AMPK. The reduction in NF- $\kappa\beta$  activity in the endothelium inhibits the expression of

pro-inflammatory adhesion proteins such as vascular cell adhesion molecule-1 (VCAM-1), E-selectin, and intercellular adhesion molecule-1 (ICAM-1). It causes the adhesion of monocytes to the endothelium of blood vessels, which is a key point in the development of atherosclerosis. On the one hand, this directly affects the renal structures. If the damage to the blood vessels occurs within the kidneys, it leads to a reduction in blood flow and local ischemia of the cells. It causes cell death or damage to their structure and kidney damage. On the other hand, the ongoing inflammatory process further increases renal dysfunction [91].

In addition, the low concentration of adiponectin leads to increased activation of NADPH oxidase 4 (Nox4) and the formation of oxidative stress [92]. Excessive amounts of ROS produced by mitochondria can lead to cellular damage and the progression of renal dysfunction [93].

Adiponectin levels are reduced in obesity, atherosclerosis, and metabolic syndrome. Despite the frequent occurrence of metabolic disorders, patients with CKD treated conservatively have from two to three times higher concentrations of serum adiponectin compared to healthy people. In patients requiring dialysis, the concentrations are even higher [94,95]. After successful KT, adiponectin levels decrease [96].

Adiponectin affects the nutritional status and CVR of patients with CKD. Increased serum adiponectin level is associated with lower BMI, waist circumference, albumin level, female sex, and older age [97–99]. Inverse associations were also observed among adiponectin levels and SAT, VAT, total body fat, and lean body mass [100]. Moreover, studies suggest a negative correlation between adiponectin levels and hand-grip [101]. Adiponectin may also affect bone turnover. It has been observed in HD patients that higher levels of adiponectin correlate with a decline in bone mineral density [102]. A high concentration of adiponectin may reflect malnutrition, which indicates a poor prognosis. Hyun et al. assessed the nutritional status and adiponectin levels were associated with PEW independent of many other factors [103]. In another study conducted among dialysis patients, a positive correlation between adiponectin and Malnutrition Inflammation Score (MIS) was observed, which indicates worsened nutritional status [104,105].

Research results regarding the influence of adiponectin on CVR is inconclusive. It was considered that high levels of adiponectin should benefit the health of people with CKD, which has been confirmed in many studies where lower adiponectin levels were associated with a higher incidence of cardiovascular events [106–108]. On the other hand, different studies have observed that high levels of this adipokine are associated with a higher risk of death, including CVR. Hence, it is sometimes called the "adiponectin paradox" [109]. Menon et al. showed that an increase in serum adiponectin among stages 3–4 of CKD by  $1 \,\mu g/mL$  was associated with a 6% higher CVR [110]. In a MADRAD study of 501 HD patients, higher adiponectin levels were also associated with a higher risk of mortality. Moreover, the concentration of adiponectin was positively correlated with high-density lipoprotein (HDL) and negatively correlated with total cholesterol and LDL [100]. High levels of adiponectin are also connected with macroalbuminuria, which is an adverse cardiovascular risk factor [111]. A synergistic effect of high adiponectin concentration and low BMI on CVR was observed [112]. The mechanism of the impact of adiponectin on CVR is unknown. Adipokine may increase the occurrence of inflammation, interfere with hematopoiesis, and increase the concentration of atrial natriuretic peptide (ANP) and brain natriuretic peptide (BNP), which are indicators of heart failure [113,114].

It has been established that the Lep/Adpn ratio may be a better indicator of the risk of developing CVD and mortality in CKD patients than the concentration of adiponectin or leptin alone. Lep/Adpn ratio reflects a disturbed function of adipose tissue [115]. Frühbeck et al. proposed the opposite ratio: adiponectin-to-leptin (Adpn/Lep). A low Adpn/Lep ratio was associated with increased inflammation and the occurrence of oxidative stress, which increases the risk of CVD [116].

#### 6.3. Zinc- $\alpha$ 2-Glycoprotein

Zinc- $\alpha$ 2-glycoprotein (ZAG) is a protein secreted by various organs, including AT and renal tubular cells, and is another adipokine that is reduced in obesity. Increased expression of the ZAG gene in AT is caused by glucocorticosteroids and androgens [117]. ZAG regulates the secretion of other adipokines in AT. It reduces the secretion of leptin and increases the secretion of adiponectin. Thus, its low concentration contributes to the persistence of obesity [118]. At physiological concentrations, ZAG increases the expression of peroxisome proliferator-activated receptor  $\gamma$  (PPAR $\gamma$ ) and early B-cell factor 2 (EBCF-2) genes by stimulating cyclic AMP (cAMP). This, in turn, initiates an increase in the synthesis of UCP1 and PR/SET domain 16 (Prdm16) proteins, which are involved in the browning process of WAT, which results in increased energy expenditure and leads to lipolysis [119].

ZAG is excreted in the urine due to damage to the basal glomerulus membrane, which may be caused by various glomerulopathies. In diabetic patients, urinary excretion of ZAG increases. It has been suggested that it may be a biomarker of diabetic nephropathy. There is a positive correlation between serum ZAG levels and the value of creatinine and eGFR. Urinary ZAG concentration correlates with the albumin/creatinine ratio [120,121].

Chronic inflammation and elevated leptin levels, which are common in obesity, reduce the secretion of ZAG in AT. Increasing the activity of ZAG in the event of an excessive energy balance may bring health benefits. A negative correlation was observed between ZAG and BMI and the body fat mass in obese subjects [122]. However, in states of catabolism such as cancer cachexia, this adipokine is associated with a deterioration in nutritional status [119].

Patients with CKD have significantly increased plasma levels of ZAG and increased synthesis in WAT compared to the healthy population, which may be associated with impaired renal excretion and the occurrence of uremia [123,124]. Moreover, inflammation and oxidative stress may increase ZAG secretion [125].

ZAG can be a new biomarker of malnutrition due to its lipolytic properties. It activates hormone-dependent lipase (HSL) via cAMP and stimulation of adenylate cyclase [126]. Overexpression of ZAG inhibits lipogenesis by reducing the expression of the genes of enzymes involved in this process, such as fatty-acid synthase (FAS) and acetyl-CoA carboxylase [127]. The serum concentration of ZAG is positively correlated with the serum levels of TG and negatively with HDL cholesterol and albumin levels [128,129].

Obese HD patients have a significantly lower concentration of this adipokine compared with patients with normal body weight [130]. Some studies revealed a negative correlation between the ZAG concentration and the percentage of body fat mass and the sum of skinfolds [131].

Bouchara et al. assessed the effect of ZAG on mortality in HD patients. High ZAG level is a predictor of all-cause mortality and CVR independent of age, or nutritional or metabolic status [126]. In another study, ZAG was negatively associated with TNF- $\alpha$  and VCAM-1, which is a marker of atherosclerosis [131].

#### 6.4. Adipose Triglyceride Lipase

Adipose triglyceride lipase (ATGL) is an enzyme encoded by the patatin-like phospholipase domain containing 2 (PNPLA2) gene. It catalyzes the first lipolysis reaction, i.e., the hydrolysis of triacylglyceride (TAG) to diacylglyceride (DAG). ATGL levels are lower in overweight and obese people than in healthy people with normal body weight [132]. Chronic ATGL adipokine deficiency increases the infiltration of immune cells into AT and enhances the inflammatory processes [133]. In terms of kidney function, ATGL deficiency leads to lipid accumulation, damage to the glomerular filtration barrier, and proteinuria. Decreased ATGL levels may increase intracellular ROS formation, which may result in F-actin fiber redistribution, foot-process fusion, and podocyte apoptosis [134]. ATGL gene expression is induced by ZAG in VAT and SAT [135].

As far as it is known, research on the comparison of ATGL concentration in people with normal kidney function and CKD has not been conducted.

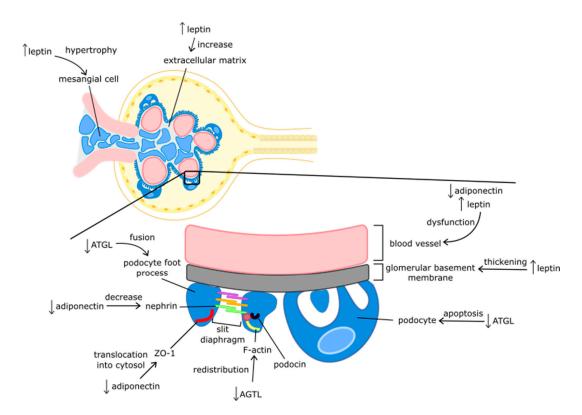
Alipoor et al. assessed the concentration of adipokines and nutritional status in HD patients and found that serum ATGL levels were significantly higher in moderate-wasting than in normal-to-mild wasting patients. The increase in ATGL levels was associated with a 21% increase in the severity of wasting. In addition, ATGL concentration was correlated with TG. ATGL levels are positively correlated with FFAs, which indicates that it affects lipolysis [136]. In another study, ATGL levels did not differ between people with normal and excessive body weight, although there was a direct relationship between ATGL levels and the percentage of body fat mass [130].

There are no studies on the effects of ATGL on CVR in patients with CKD. Salatzki et al. showed that in animal models with normal kidney function, the removal of ATGL in AT prevents pressure-induced left ventricular heart failure, which confirms the adverse effect of lipolysis on heart function [137]. However, other studies found an adverse effect of ATGL deficiency on the cardiovascular system. ATGL deficiency has been associated with increased wall thickness and cardiac fibrosis [138,139]. The impact of adipokines on nutritional status and CVR is summarized in Table 1 and on kidney structures in Figure 3.

**Table 1.** Profile of adipokines in chronic kidney disease and their influence on nutritional status and cardiovascular risk in CKD.

Activity	Leptin	Adiponectin	ZAG	ATGL
Plasma levels in obesity	$\uparrow$	$\downarrow$	$\downarrow$	$\downarrow$
Plasma levels in CKD	1	$\uparrow$	$\uparrow$	U
Malnutrition/PEW	†/↓	1	<b>†</b>	$\uparrow$
Oxidative stress	1	$\uparrow$	$\uparrow$	1
Inflammation	1	1	<u></u>	1
CVR	↑	$\uparrow/\downarrow$	1	Ŭ

Abbreviations: ZAG—zinc-alpha2-glycoprotein; ATGL—adipose triglyceride lipase; CKD—chronic kidney disease; PEW—protein energy-wasting; CVR—cardiovascular risk; U—unknown; ↑—increased levels; ↓—decreased levels; ↑/↓—conflicting results.



**Figure 3.** The influence of adipokines on the structures of the kidneys. Abbreviations: ATGL—adipose triglyceride lipase; ZO-1—zonula occludens; ↑—increased levels; ↓—decreased levels.

# 7. Conclusions

Obesity leads to the development of many other diseases through the occurrence of chronic inflammation, oxidative stress, and disorders of adipokine secretion. Obesity and the associated impaired secretion of adipokines contribute to the development of CKD and its progression. However, CKD also influences the concentration of adipokines, nutritional status, and CVR in patients with CKD. There is a mutual influence on the axis of the kidney and AT. Despite the broadening knowledge of obesity and its possible treatment methods, the number of obese people is still increasing. Correspondingly, although there is a lot of research on CKD, many mechanisms are not yet completely understood, for example, the mechanism of the development of nutritional disorders and CVR. In particular, ZAG and ATGL are adipokines that are not sufficiently studied. There is little literature on the concentration of these adipokines concerning the nutritional status and CVR in patients with CKD. Although the manuscript contains information summarizing many publications, not all existing articles on a given topic were considered. Additionally, some of the source texts are reviews and research publications. More research is needed on this topic.

**Author Contributions:** Writing—original draft preparation: S.C.-S. and M.P.; writing—review and editing: S.C.-S., M.P. and S.M.; supervision: M.S., Ł.K. and S.M. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

## References

- 1. World Health Statistics 2021: Monitoring Health for the SDGs, Sustainable Development Goals; World Health Organization: Geneva, Switzerland, 2021.
- Mattioli, A.V.; Sciomer, S.; Cocchi, C.; Maffei, S.; Gallina, S. Quarantine during COVID-19 outbreak: Changes in diet and physical activity increase the risk of cardiovascular disease. *Nutr. Metab. Cardiovasc. Dis.* 2020, 30, 1409–1417. [CrossRef]
- Pellegrini, M.; Ponzo, V.; Rosato, R.; Scumaci, E.; Goitre, I.; Benso, A.; Belcastro, S.; Crespi, C.; De Michieli, F.; Ghigo, E.; et al. Changes in Weight and Nutritional Habits in Adults with Obesity during the "Lockdown" Period Caused by the COVID-19 Virus Emergency. *Nutrients* 2020, *12*, 2016. [CrossRef]
- Divino, V.; Ramasamy, A.; Anupindi, V.R.; Eriksen, K.T.; Olsen, A.H.; DeKoven, M.; Meincke, H.H. Complication-specific direct medical costs by body mass index for 13 obesity-related complications: A retrospective database study. *J. Manag. Care Spec. Pharm.* 2021, 27, 210–222. [CrossRef] [PubMed]
- 5. Cawley, J.; Biener, A.; Meyerhoefer, C.; Ding, Y.; Zvenyach, T.; Smolarz, B.G.; Ramasamy, A. Direct medical costs of obesity in the United States and the most populous states. *J. Manag. Care Spec. Pharm.* **2021**, *27*, 354–366. [CrossRef]
- Hill, N.R.; Fatoba, S.T.; Oke, J.L.; Hirst, J.A.; O'Callaghan, C.A.; Lasserson, D.S.; Hobbs, F.D. Global Prevalence of Chronic Kidney Disease—A Systematic Review and Meta-Analysis. *PLoS ONE* 2016, *11*, e0158765. [CrossRef]
- 7. Lainščak, M. Cardiovascular Risk in Chronic Kidney Disease. EJIFCC 2009, 20, 73–76. [PubMed]
- 8. Srivastava, A.; Kaze, A.D.; McMullan, C.J.; Isakova, T.; Waikar, S.S. Uric Acid and the Risks of Kidney Failure and Death in Individuals with CKD. *Am. J. Kidney Dis.* **2018**, *71*, 362–370. [CrossRef] [PubMed]
- Castaneda-Sceppa, C.; Sarnak, M.J.; Wang, X.; Greene, T.; Madero, M.; Kusek, J.W.; Beck, G.; Kopple, J.D.; Levey, A.S.; Menon, V. Role of adipose tissue in determining muscle mass in patients with chronic kidney disease. *J. Ren. Nutr.* 2007, 17, 314–322. [CrossRef] [PubMed]
- Podkowińska, A.; Formanowicz, D. Chronic kidney disease as oxidative stress-and inflammatory-mediated cardiovascular disease. *Antioxidants* 2020, 9, 752. [CrossRef] [PubMed]
- 11. Martin-Taboada, M.; Vila-Bedmar, R.; Medina-Gómez, G. From Obesity to Chronic Kidney Disease: How Can Adipose Tissue Affect Renal Function? *Nephron* **2021**, *145*, 609–613. [CrossRef]
- Yang, Y.D.; Zheng, C.J.; Dong, Y.H.; Zou, Z.Y.; Lv, Y.; Wang, Z.H.; Yang, Z.G.; Wang, S.; Dong, B.; Ma, J. Sex difference in the mediation roles of an inflammatory factor (hsCRP) and adipokines on the relationship between adiposity and blood pressure. *Hypertens. Res.* 2019, 42, 903–911. [CrossRef] [PubMed]
- 13. Lenz, M.; Arts, I.C.W.; Peeters, R.L.M.; de Kok, T.M.; Ertaylan, G. Adipose tissue in health and disease through the lens of its building blocks. *Sci. Rep.* 2020, *10*, 10433. [CrossRef]

- 14. Chait, A.; den Hartigh, L.J. Adipose Tissue Distribution, Inflammation and Its Metabolic Consequences, Including Diabetes and Cardiovascular Disease. *Front. Cardiovasc. Med.* **2020**, *7*, 22. [CrossRef] [PubMed]
- 15. El Hadi, H.; Di Vincenzo, A.; Vettor, R.; Rossato, M. Food Ingredients Involved in White-to-Brown Adipose Tissue Conversion and in Calorie Burning. *Front. Physiol.* **2019**, *9*, 1954. [CrossRef]
- 16. Bartelt, A.; Bruns, O.T.; Reimer, R.; Hohenberg, H.; Ittrich, H.; Peldschus, K.; Kaul, M.G.; Tromsdorf, U.I.; Weller, H.; Waurisch, C.; et al. Brown adipose tissue activity controls triglyceride clearance. *Nat. Med.* **2011**, *17*, 200–205. [CrossRef]
- 17. Roth, C.L.; Molica, F.; Kwak, B.R. Browning of White Adipose Tissue as a Therapeutic Tool in the Fight against Atherosclerosis. *Metabolites* **2021**, *11*, 319. [CrossRef]
- Orava, J.; Nuutila, P.; Lidell, M.E.; Oikonen, V.; Noponen, T.; Viljanen, T.; Scheinin, M.; Taittonen, M.; Niemi, T.; Enerbäck, S.; et al. Different metabolic responses of human brown adipose tissue to activation by cold and insulin. *Cell Metab.* 2011, 14, 272–279. [CrossRef] [PubMed]
- 19. Zwick, R.K.; Guerrero-Juarez, C.F.; Horsley, V.; Plikus, M.V. Anatomical, Physiological, and Functional Diversity of Adipose Tissue. *Cell Metab.* 2018, 27, 68–83. [CrossRef] [PubMed]
- 20. Wajchenberg, B.L. Subcutaneous and visceral adipose tissue: Their relation to the metabolic syndrome. *Endocr. Rev.* 2000, 21, 697–738. [CrossRef] [PubMed]
- 21. Zhang, Y.; Zitsman, J.L.; Hou, J.; Fennoy, I.; Guo, K.; Feinberg, J.; Leibel, R.L. Fat cell size and adipokine expression in relation to gender, depot, and metabolic risk factors in morbidly obese adolescents. *Obesity* **2014**, 22, 691–697. [CrossRef]
- 22. Longo, M.; Zatterale, F.; Naderi, J.; Parrillo, L.; Formisano, P.; Raciti, G.A.; Beguinot, F.; Miele, C. Adipose Tissue Dysfunction as Determinant of Obesity-Associated Metabolic Complications. *Int. J. Mol. Sci.* **2019**, *20*, 2358. [CrossRef]
- Góralska, M.; Majewska-Szczepanik, M.; Szczepanik, M. Immunological mechanisms involved in obesity and their role in metabolic syndrome. *Postepy Hig. Med. Dosw.* 2015, 69, 1384–1404.
- Thomas, D.; Apovian, C. Macrophage functions in lean and obese adipose tissue. *Metabolism* 2017, 72, 120–143. [CrossRef] [PubMed]
- 25. Kuryłowicz, A.; Puzianowska-Kuźnicka, M. Induction of Adipose Tissue Browning as a Strategy to Combat Obesity. *Int. J. Mol. Sci.* 2020, *21*, 6241. [CrossRef] [PubMed]
- Chang, E.; Varghese, M.; Singer, K. Gender and Sex Differences in Adipose Tissue. *Curr. Diabetes Rep.* 2018, 18, 69. [CrossRef] [PubMed]
- 27. Palmer, B.F.; Clegg, D.J. The sexual dimorphism of obesity. Mol. Cell. Endocrinol. 2015, 402, 113–119. [CrossRef]
- Hall, J.E.; do Carmo, J.M.; da Silva, A.A.; Wang, Z.; Hall, M.E. Obesity, kidney dysfunction and hypertension: Mechanistic links. *Nat. Rev. Nephrol.* 2019, 15, 367–385. [CrossRef]
- 29. Suzuki, M.; Murakami, M.; Shirai, M.; Hashimoto, O.; Kawada, T.; Matsui, T.; Funaba, M. Role of estradiol and testosterone in Ucp1 expression in brown/beige adipocytes. *Cell Biochem. Funct.* **2018**, *36*, 450–456. [CrossRef]
- Chen, X.; McClusky, R.; Chen, J.; Beaven, S.W.; Tontonoz, P.; Arnold, A.P.; Reue, K. The number of x chromosomes causes sex differences in adiposity in mice. *PLoS Genet.* 2012, *8*, e1002709. [CrossRef] [PubMed]
- 31. Walicka, M.; Bik, W.; Wolińska-Witort, E.; Marcinowska-Suchowierska, E. Gender dependent dimorphism in adipokines levels and its correlations with insulin resistance in extremely obese patients. *Postępy Nauk. Med.* **2013**, *24*, 262–266.
- 32. Selthofer-Relatić, K.; Radić, R.; Stupin, A.; Šišljagić, V.; Bošnjak, I.; Bulj, N.; Selthofer, R.; Delić Brkljačić, D. Leptin/adiponectin ratio in overweight patients—gender differences. *Diabetes Vasc. Dis. Res.* 2018, *15*, 260–262. [CrossRef]
- 33. Ecelbarger, C.M. Sex Differences in Renal Physiology and Pathophysiology. In *Sex Differences in Physiology*; Academic Press: Amsterdam, The Netherlands, 2016; pp. 105–124. [CrossRef]
- 34. National Kidney Foundation. KDOQI Clinical Practice Guidelines for Chronic Kidney Disease: Evaluation, Classification and Stratification. *Am. J. Kidney Dis.* **2002**, *39*, 1–266.
- 35. Rhee, C.M.; Ahmadi, S.F.; Kalantar-Zadeh, K. The dual roles of obesity in chronic kidney disease: A review of the current literature. *Curr. Opin. Nephrol. Hypertens.* 2016, 25, 208–216. [CrossRef]
- Wang, M.; Wang, Z.; Chen, Y.; Dong, Y. Kidney Damage Caused by Obesity and Its Feasible Treatment Drugs. *Int. J. Mol. Sci.* 2022, 23, 747. [CrossRef]
- 37. Praga, M.; Morales, E. The Fatty Kidney: Obesity and Renal Disease. Nephron 2017, 136, 273–276. [CrossRef]
- Denic, A.; Glassock, R.J. Obesity-Related Glomerulopathy and Single-Nephron GFR. *Kidney Int. Rep.* 2020, 5, 1126–1128. [CrossRef] [PubMed]
- 39. Yang, S.; Cao, C.; Deng, T.; Zhou, Z. Obesity-Related Glomerulopathy: A Latent Change in Obesity Requiring More Attention. *Kidney Blood Press. Res.* 2020, 45, 510–522. [CrossRef]
- 40. Herman-Edelstein, M.; Weinstein, T.; Chagnac, A. Obesity-Related Glomerulopathy: Clinical Management. *Semin. Nephrol.* 2021, 41, 358–370. [CrossRef]
- 41. Caus, M.; Eritja, A.; Bozic, M. Role of microRNAs in Obesity-Related Kidney Disease. Int. J. Mol. Sci. 2021, 22, 11416. [CrossRef]
- Chagnac, A.; Zingerman, B.; Rozen-Zvi, B.; Herman-Edelstein, M. Consequences of Glomerular Hyperfiltration: The Role of Physical Forces in the Pathogenesis of Chronic Kidney Disease in Diabetes and Obesity. *Nephron* 2019, 143, 38–42. [CrossRef] [PubMed]

- Tobar, A.; Ori, Y.; Benchetrit, S.; Milo, G.; Herman-Edelstein, M.; Zingerman, B.; Lev, N.; Gafter, U.; Chagnac, A. Proximal tubular hypertrophy and enlarged glomerular and proximal tubular urinary space in obese subjects with proteinuria. *PLoS ONE* 2013, *8*, e75547. [CrossRef] [PubMed]
- Ogna, A.; Forni Ogna, V.; Bochud, M.; Guessous, I.; Paccaud, F.; Burnier, M.; Wuerzner, G. Association between obesity and glomerular hyperfiltration: The confounding effect of smoking and sodium and protein intakes. *Eur. J. Nutr.* 2016, 55, 1089–1097. [CrossRef] [PubMed]
- 45. Helal, I.; Fick-Brosnahan, G.; Reed-Gitomer, B.; Schrier, R.W. Glomerular hyperfiltration: Definitions, mechanisms and clinical implications. *Nat. Rev. Nephrol.* **2012**, *8*, 293–300. [CrossRef]
- Opazo-Ríos, L.; Mas, S.; Marín-Royo, G.; Mezzano, S.; Gómez-Guerrero, C.; Moreno, J.A.; Egido, J. Lipotoxicity and Diabetic Nephropathy: Novel Mechanistic Insights and Therapeutic Opportunities. *Int. J. Mol. Sci.* 2020, 21, 2632. [CrossRef]
- de Vries, A.P.; Ruggenenti, P.; Ruan, X.Z.; Praga, M.; Cruzado, J.M.; Bajema, I.M.; D'Agati, V.D.; Lamb, H.J.; Pongrac Barlovic, D.; Hojs, R.; et al. Fatty kidney: Emerging role of ectopic lipid in obesity-related renal disease. *Lancet Diabetes Endocrinol.* 2014, 2, 417–426. [CrossRef]
- 48. Grigoraș, A.; Balan, R.A.; Căruntu, I.D.; Giușcă, S.E.; Lozneanu, L.; Avadanei, R.E.; Rusu, A.; Riscanu, L.A.; Amalinei, C. Perirenal Adipose Tissue-Current Knowledge and Future Opportunities. *J. Clin. Med.* **2021**, *10*, 1291. [CrossRef]
- Limberg, J.K.; Soares, R.N.; Power, G.; Harper, J.L.; Smith, J.A.; Shariffi, B.; Jacob, D.W.; Manrique-Acevedo, C.; Padilla, J. Hyperinsulinemia blunts sympathetic vasoconstriction: A possible role of β-adrenergic activation. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 2021, 320, R771–R779. [CrossRef] [PubMed]
- Jang, H.S.; Noh, M.R.; Kim, J.; Padanilam, B.J. Defective Mitochondrial Fatty Acid Oxidation and Lipotoxicity in Kidney Diseases. Front. Med. 2020, 7, 65. [CrossRef] [PubMed]
- 51. Huang, N.; Mao, E.W.; Hou, N.N.; Liu, Y.P.; Han, F.; Sun, X.D. Novel insight into perirenal adipose tissue: A neglected adipose depot linking cardiovascular and chronic kidney disease. *World J. Diabetes* 2020, *11*, 115–125. [CrossRef]
- 52. Farkhondeh, T.; Llorens, S.; Pourbagher-Shahri, A.M.; Ashrafizadeh, M.; Talebi, M.; Shakibaei, M.; Samarghandian, S. An Overview of the Role of Adipokines in Cardiometabolic Diseases. *Molecules* **2020**, *25*, 5218. [CrossRef] [PubMed]
- 53. Nasrallah, M.P.; Ziyadeh, F.N. Overview of the physiology and pathophysiology of leptin with special emphasis on its role in the kidney. *Semin. Nephrol.* **2013**, *33*, 54–65. [CrossRef]
- 54. Hamrahian, S.M.; Falkner, B. Hypertension in Chronic Kidney Disease. *Adv. Exp. Med. Biol.* 2017, 956, 307–325. [CrossRef] [PubMed]
- Zhu, Q.; Scherer, P.E. Immunologic and endocrine functions of adipose tissue: Implications for kidney disease. *Nat. Rev. Nephrol.* 2018, 14, 105–120. [CrossRef]
- 56. Podestà, M.A.; Ponticelli, C. Autoimmunity in Focal Segmental Glomerulosclerosis: A Long-Standing Yet Elusive Association. *Front. Med.* **2020**, *7*, 604961. [CrossRef] [PubMed]
- 57. Rüster, C.; Wolf, G. Adipokines promote chronic kidney disease. Nephrol. Dial. Transplant. 2013, 28, 8–14. [CrossRef] [PubMed]
- 58. Procaccini, C.; Jirillo, E.; Matarese, G. Leptin as an immunomodulator. Mol. Asp. Med. 2012, 33, 35–45. [CrossRef]
- 59. Lim, C.C.; Teo, B.W.; Tai, E.S.; Lim, S.C.; Chan, C.M.; Sethi, S.; Wong, T.Y.; Sabanayagam, C. Elevated serum leptin, adiponectin and leptin to adiponectin ratio is associated with chronic kidney disease in Asian adults. *PLoS ONE* 2015, *10*, e0122009. [CrossRef]
- Ambarkar, M.; Pemmaraju, S.V.; Gouroju, S.; Manohar, S.M.; Bitla, A.R.; Yajamanam, N.; Vishnubhotla, S. Adipokines and their Relation to Endothelial Dysfunction in Patients with Chronic Kidney Disease. *J. Clin. Diagn. Res.* 2016, 10, BC04–BC08. [CrossRef] [PubMed]
- Korczyńska, J.; Czumaj, A.; Chmielewski, M.; Śledziński, M.; Mika, A.; Śledziński, T. Increased Expression of the Leptin Gene in Adipose Tissue of Patients with Chronic Kidney Disease-The Possible Role of an Abnormal Serum Fatty Acid Profile. *Metabolites* 2020, 10, 98. [CrossRef]
- Navaneethan, S.D.; Kirwan, J.P.; Remer, E.M.; Schneider, E.; Addeman, B.; Arrigain, S.; Horwitz, E.; Fink, J.C.; Lash, J.P.; McKenzie, C.A.; et al. Adiposity, Physical Function, and Their Associations with Insulin Resistance, Inflammation, and Adipokines in CKD. *Am. J. Kidney Dis.* 2021, 77, 44–55. [CrossRef]
- 63. Ahamadi, F.; Bosorgmehr, R.; Razeghi, E. Relationship between serum leptin level and laboratory and anthropometric indices of malnutrition in patients on hemodialysis. *Indian J. Nephrol.* **2008**, *18*, 105–111. [CrossRef]
- 64. Koppe, L.; Fouque, D.; Kalantar-Zadeh, K. Kidney cachexia or protein-energy wasting in chronic kidney disease: Facts and numbers. *J. Cachexia Sarcopenia Muscle* **2019**, *10*, 479–484. [CrossRef]
- 65. Ko, Y.T.; Lin, Y.L.; Kuo, C.H.; Lai, Y.H.; Wang, C.H.; Hsu, B.G. Low serum leptin levels are associated with malnutrition status according to malnutrition-inflammation score in patients undergoing chronic hemodialysis. *Hemodial. Int.* 2020, 24, 221–227. [CrossRef]
- Steiner, A.A.; Romanovsky, A.A. Leptin: At the crossroads of energy balance and systemic inflammation. *Prog. Lipid Res.* 2007, 46, 89–107. [CrossRef]
- 67. Guzmán, A.; Hernández-Coronado, C.G.; Rosales-Torres, A.M.; Hernández-Medrano, J.H. Leptin regulates neuropeptides associated with food intake and GnRH secretion. *Ann. Endocrinol.* **2019**, *80*, 38–46. [CrossRef] [PubMed]
- 68. Mak, R.H.; Cheung, W.; Cone, R.D.; Marks, D.L. Orexigenic and anorexigenic mechanisms in the control of nutrition in chronic kidney disease. *Pediatr. Nephrol.* 2005, 20, 427–431. [CrossRef]

- Zhang, L.; Reed, F.; Herzog, H. Leptin signalling on arcuate NPY neurones controls adiposity independent of energy balance or diet composition. J. Neuroendocrinol. 2020, 32, e12898. [CrossRef]
- 70. Lu, L.; Zou, Y.C.; Wang, M.; Huang, Y.F.; Chen, D.X.; Wei, L.B. Neuropeptide Y levels are associated with nutritional status and cardiovascular events in adults with chronic kidney disease. *Eur. J. Clin. Nutr.* **2015**, *69*, 717–721. [CrossRef]
- 71. Zoccali, C.; Ortiz, A.; Blumbyte, I.A.; Rudolf, S.; Beck-Sickinger, A.G.; Malyszko, J.; Spasovski, G.; Carriazo, S.; Viggiano, D.; Kurganaite, J.; et al. Neuropeptide Y as a risk factor for cardiorenal disease and cognitive dysfunction in chronic kidney disease: Translational opportunities and challenges. *Nephrol. Dial. Transplant.* 2021, *37*, ii14–ii23. [CrossRef]
- 72. Zoccali, C.; D'Arrigo, G.; Leonardis, D.; Pizzini, P.; Postorino, M.; Tripepi, G.; Mallamaci, F. Neuropeptide Y predicts cardiovascular events in chronic kidney disease patients: A cohort study. *J. Hypertens.* **2019**, *37*, 1359–1365. [CrossRef] [PubMed]
- 73. Markaki, A.; Grammatikopoulou, M.G.; Venihaki, M.; Kyriazis, J.; Perakis, K.; Stylianou, K. Associations of adiponectin and leptin levels with protein-energy wasting, in end stage renal disease patients. *Endocrinol. Nutr.* **2016**, *63*, 449–457. [CrossRef]
- 74. de Oliveira, R.B.; Liabeuf, S.; Okazaki, H.; Lenglet, A.; Desjardins, L.; Lemke, H.D.; Vanholder, R.; Choukroun, G.; Massy, Z.A. The clinical impact of plasma leptin levels in a cohort of chronic kidney disease patients. *Clin. Kidney J.* 2013, 6, 63–70. [CrossRef] [PubMed]
- Chiu, T.T.; Liao, S.C.; Lee, W.C.; Lee, P.S.; Ng, H.Y.; Chien, Y.S.; Lee, C.T. Gelsolin and adipokines are associated with protein-energy wasting in hemodialysis patients. *Artif. Organs* 2015, *39*, 150–155. [CrossRef] [PubMed]
- Katsiki, N.; Mikhailidis, D.P.; Banach, M. Leptin, cardiovascular diseases and type 2 diabetes mellitus. *Acta Pharmacol. Sin.* 2018, 39, 1176–1188. [CrossRef] [PubMed]
- Korczyńska, J.; Czumaj, A.; Chmielewski, M.; Świerczyński, J.; Śledziński, T. The Causes and Potential Injurious Effects of Elevated Serum Leptin Levels in Chronic Kidney Disease Patients. *Int. J. Mol. Sci.* 2021, 22, 4685. [CrossRef]
- 78. Sweeney, G. Cardiovascular effects of leptin. *Nat. Rev. Cardiol.* **2010**, *7*, 22–29. [CrossRef]
- 79. Lu, J.W.; Chi, P.J.; Lin, Y.L.; Wang, C.H.; Hsu, B.G. Serum leptin levels are positively associated with aortic stiffness in patients with chronic kidney disease stage 3–5. *Adipocyte* **2020**, *9*, 206–211. [CrossRef]
- 80. Tsai, J.P.; Lee, M.C.; Chen, Y.C.; Ho, G.J.; Shih, M.H.; Hsu, B.G. Hyperleptinemia is a risk factor for the development of central arterial stiffness in kidney transplant patients. *Transplant. Proc.* **2015**, *47*, 1825–1830. [CrossRef]
- 81. Kuo, C.H.; Lin, Y.L.; Lee, C.J.; Wang, C.H.; Lai, Y.H.; Liou, H.H.; Hsu, B.G. Hyperleptinemia positively associated with central arterial stiffness in hemodialysis patients. *PLoS ONE* **2018**, *13*, e0190694. [CrossRef]
- Zoccali, C.; Postorino, M.; Marino, C.; Pizzini, P.; Cutrupi, S.; Tripepi, G. Waist circumference modifies the relationship between the adipose tissue cytokines leptin and adiponectin and all-cause and cardiovascular mortality in haemodialysis patients. *J. Intern. Med.* 2011, 269, 172–181. [CrossRef]
- Scholze, A.; Rattensperger, D.; Zidek, W.; Tepel, M. Low serum leptin predicts mortality in patients with chronic kidney disease stage 5. *Obesity* 2007, 15, 1617–1622. [CrossRef] [PubMed]
- 84. Molnar, M.Z.; Nagy, K.; Remport, A.; Gaipov, A.; Fülöp, T.; Czira, M.E.; Kovesdy, C.P.; Mucsi, I.; Mathe, Z. Association Between Serum Leptin Level and Mortality in Kidney Transplant Recipients. *J. Ren. Nutr.* **2017**, *27*, 53–61. [CrossRef] [PubMed]
- Achari, A.E.; Jain, S.K. Adiponectin, a Therapeutic Target for Obesity, Diabetes, and Endothelial Dysfunction. *Int. J. Mol. Sci.* 2017, 18, 1321. [CrossRef]
- 86. Christou, G.A.; Kiortsis, D.N. The role of adiponectin in renal physiology and development of albuminuria. *J. Endocrinol.* **2014**, 221, R49–R61. [CrossRef]
- 87. Iwabu, M.; Okada-Iwabu, M.; Yamauchi, T.; Kadowaki, T. Adiponectin/AdipoR Research and Its Implications for Lifestyle-Related Diseases. *Front. Cardiovasc. Med.* 2019, *6*, 116. [CrossRef] [PubMed]
- 88. Cammisotto, P.G.; Bendayan, M. Adiponectin stimulates phosphorylation of AMP-activated protein kinase alpha in renal glomeruli. *J. Mol. Histol.* **2008**, *39*, 579–584. [CrossRef]
- 89. Ohmori, T.; De, S.; Tanigawa, S.; Miike, K.; Islam, M.; Soga, M.; Era, T.; Shiona, S.; Nakanishi, K.; Nakazato, H.; et al. Impaired NEPHRIN localization in kidney organoids derived from nephrotic patient iPS cells. *Sci. Rep.* **2021**, *11*, 3982. [CrossRef]
- 90. Zhao, D.; Zhu, X.; Jiang, L.; Huang, X.; Zhang, Y.; Wei, X.; Zhao, X.; Du, Y. Advances in understanding the role of adiponectin in renal fibrosis. *Nephrology* **2021**, *26*, 197–203. [CrossRef]
- Ouchi, N.; Kihara, S.; Arita, Y.; Okamoto, Y.; Maeda, K.; Kuriyama, H.; Hotta, K.; Nishida, M.; Takahashi, M.; Muraguchi, M.; et al. Adiponectin, an adipocyte-derived plasma protein, inhibits endothelial NF-kappaB signaling through a cAMP-dependent pathway. *Circulation* 2000, 102, 1296–1301. [CrossRef]
- 92. Sharma, K.; Ramachandrarao, S.; Qiu, G.; Usui, H.K.; Zhu, Y.; Dunn, S.R.; Ouedraogo, R.; Hough, K.; McCue, P.; Chan, L.; et al. Adiponectin regulates albuminuria and podocyte function in mice. *J. Clin. Investig.* **2008**, *118*, 1645–1656. [CrossRef]
- Galvan, D.L.; Green, N.H.; Danesh, F.R. The hallmarks of mitochondrial dysfunction in chronic kidney disease. *Kidney Int.* 2017, 92, 1051–1057. [CrossRef]
- 94. Miricescu, D.; Balan, D.G.; Tulin, A.; Stiru, O.; Vacaroiu, I.A.; Mihai, D.A.; Popa, C.C.; Enyedi, M.; Nedelea, A.S.; Nica, A.E.; et al. Impact of adipose tissue in chronic kidney disease development (Review). *Exp. Ther. Med.* **2021**, *21*, 539. [CrossRef] [PubMed]
- 95. Huang, J.W.; Yen, C.J.; Chiang, H.W.; Hung, K.Y.; Tsai, T.J.; Wu, K.D. Adiponectin in peritoneal dialysis patients: A comparison with hemodialysis patients and subjects with normal renal function. *Am. J. Kidney Dis.* **2004**, *43*, 1047–1055. [CrossRef]

- Chudek, J.; Adamczak, M.; Karkoszka, H.; Budziński, G.; Ignacy, W.; Funahashi, T.; Matsuzawa, Y.; Cierpka, L.; Kokot, F.; Wiecek, A. Plasma adiponectin concentration before and after successful kidney transplantation. *Transplant. Proc.* 2003, 35, 2186–2189. [CrossRef]
- Kamimura, M.A.; Canziani, M.E.; Sanches, F.R.; Velludo, C.M.; Carrero, J.J.; Bazanelli, A.P.; Draibe, S.A.; Cuppari, L. Variations in adiponectin levels in patients with chronic kidney disease: A prospective study of 12 months. *Braz. J. Nephrol.* 2012, 34, 259–265. [CrossRef]
- Kaynar, K.; Kural, B.V.; Ulusoy, S.; Cansiz, M.; Akcan, B.; Misir, N.; Yaman, S.; Kaya, N. Is there any interaction of resistin and adiponectin levels with protein-energy wasting among patients with chronic kidney disease. *Hemodial. Int.* 2014, 18, 153–162. [CrossRef] [PubMed]
- 99. Song, S.H.; Oh, T.R.; Choi, H.S.; Kim, C.S.; Ma, S.K.; Oh, K.H.; Ahn, C.; Kim, S.W.; Bae, E.H. High serum adiponectin as a biomarker of renal dysfunction: Results from the KNOW-CKD study. *Sci. Rep.* **2020**, *10*, 5598. [CrossRef]
- Rhee, C.M.; Nguyen, D.V.; Moradi, H.; Brunelli, S.M.; Dukkipati, R.; Jing, J.; Nakata, T.; Kovesdy, C.P.; Brent, G.A.; Kalantar-Zadeh, K. Association of Adiponectin with Body Composition and Mortality in Hemodialysis Patients. *Am. J. Kidney Dis.* 2015, 66, 313–321. [CrossRef]
- Kobayashi, H.; Otsuka, H.; Yanai, M.; Haketa, A.; Hara, M.; Hishiki, M.; Abe, M.; Soma, M. Adiponectin is not associated with renal function decline in community-dwelling elderly adults. *Medicine* 2018, 97, e10847. [CrossRef]
- 102. Okuno, S.; Ishimura, E.; Norimine, K.; Tsuboniwa, N.; Kagitani, S.; Yamakawa, K.; Yamakawa, T.; Sato, K.K.; Hayashi, T.; Shoji, S.; et al. Serum adiponectin and bone mineral density in male hemodialysis patients. *Osteoporos. Int.* 2012, 23, 2027–2035. [CrossRef] [PubMed]
- 103. Hyun, Y.Y.; Lee, K.B.; Oh, K.H.; Ahn, C.; Park, S.K.; Chae, D.W.; Yoo, T.H.; Cho, K.H.; Kim, Y.S.; Hwang, Y.H. Serum adiponectin and protein-energy wasting in pre-dialysis chronic kidney disease. *Nutrition* 2017, *33*, 254–260. [CrossRef] [PubMed]
- 104. Lee, Y.J.; Cho, S.; Kim, S.R. The association between serum adiponectin levels and nutritional status of hemodialysis patients. *Ren. Fail.* 2011, 33, 506–511. [CrossRef] [PubMed]
- 105. Dervisoglu, E.; Eraldemir, C.; Kalender, B.; Kir, H.M.; Caglayan, C. Adipocytokines leptin and adiponectin, and measures of malnutrition-inflammation in chronic renal failure: Is there a relationship? *J. Ren. Nutr.* **2008**, *18*, 332–337. [CrossRef]
- Rao, M.; Li, L.; Tighiouart, H.; Jaber, B.L.; Pereira, B.J.; Balakrishnan, V.S. Plasma adiponectin levels and clinical outcomes among haemodialysis patients. *Nephrol. Dial. Transplant.* 2008, 23, 2619–2628. [CrossRef] [PubMed]
- Becker, B.; Kronenberg, F.; Kielstein, J.T.; Haller, H.; Morath, C.; Ritz, E.; Fliser, D. Renal insulin resistance syndrome, adiponectin and cardiovascular events in patients with kidney disease: The mild and moderate kidney disease study. *J. Am. Soc. Nephrol.* 2005, 16, 1091–1098. [CrossRef] [PubMed]
- Abdallah, E.; Waked, E.; Nabil, M.; El-Bendary, O. Adiponectin and cardiovascular outcomes among hemodialysis patients. *Kidney Blood Press. Res.* 2012, 35, 247–253. [CrossRef]
- 109. Kalkman, H.O. An Explanation for the Adiponectin Paradox. Pharmaceuticals 2021, 14, 1266. [CrossRef] [PubMed]
- 110. Menon, V.; Li, L.; Wang, X.; Greene, T.; Balakrishnan, V.; Madero, M.; Pereira, A.A.; Beck, G.J.; Kusek, J.W.; Collins, A.J.; et al. Adiponectin and mortality in patients with chronic kidney disease. *J. Am. Soc. Nephrol.* **2006**, *17*, 2599–2606. [CrossRef]
- 111. Kim, H.Y.; Bae, E.H.; Ma, S.K.; Chae, D.W.; Choi, K.H.; Kim, Y.S.; Hwang, Y.H.; Ahn, C.; Kim, S.W. Association of serum adiponectin level with albuminuria in chronic kidney disease patients. *Clin. Exp. Nephrol.* **2016**, *20*, 443–449. [CrossRef] [PubMed]
- 112. Choi, S.H.; Ku, E.J.; Hong, E.S.; Lim, S.; Kim, K.W.; Moon, J.H.; Kim, K.M.; Park, Y.J.; Park, K.S.; Jang, H.C. High serum adiponectin concentration and low body mass index are significantly associated with increased all-cause and cardiovascular mortality in an elderly cohort, "adiponectin paradox": The Korean Longitudinal Study on Health and Aging (KLoSHA). *Int. J. Cardiol.* 2015, 183, 91–97. [CrossRef] [PubMed]
- 113. Kim, H.; Yun, H.R.; Park, S.; Jhee, J.H.; Park, J.T.; Yoo, T.H.; Lee, K.B.; Kim, Y.H.; Sung, S.A.; Lee, J.; et al. High serum adiponectin is associated with anemia development in chronic kidney disease: The results from the KNOW-CKD study. *Cytokine* 2018, 103, 1–9. [CrossRef] [PubMed]
- 114. Tsukamoto, O.; Fujita, M.; Kato, M.; Yamazaki, S.; Asano, Y.; Ogai, A.; Okazaki, H.; Asai, M.; Nagamachi, Y.; Maeda, N.; et al. Natriuretic peptides enhance the production of adiponectin in human adipocytes and in patients with chronic heart failure. *J. Am. Coll. Cardiol.* 2009, 53, 2070–2077. [CrossRef]
- 115. Park, J.T.; Yoo, T.H.; Kim, J.K.; Oh, H.J.; Kim, S.J.; Yoo, D.E.; Lee, M.J.; Shin, D.H.; Han, S.H.; Han, D.S.; et al. Leptin/adiponectin ratio is an independent predictor of mortality in nondiabetic peritoneal dialysis patients. *Perit. Dial. Int.* 2013, 33, 67–74. [CrossRef] [PubMed]
- 116. Frühbeck, G.; Catalán, V.; Rodríguez, A.; Ramírez, B.; Becerril, S.; Salvador, J.; Colina, I.; Gómez-Ambrosi, J. Adiponectin-leptin Ratio is a Functional Biomarker of Adipose Tissue Inflammation. *Nutrients* **2019**, *11*, 454. [CrossRef] [PubMed]
- 117. Russell, S.T.; Tisdale, M.J. The role of glucocorticoids in the induction of zinc-alpha2-glycoprotein expression in adipose tissue in cancer cachexia. *Br. J. Cancer* 2005, *92*, 876–881. [CrossRef] [PubMed]
- Banaszak, M.; Górna, I.; Przysławski, J. Zinc and the Innovative Zinc-α2-Glycoprotein Adipokine Play an Important Role in Lipid Metabolism: A Critical Review. *Nutrients* 2021, *13*, 2023. [CrossRef]
- Elattar, S.; Dimri, M.; Satyanarayana, A. The tumor secretory factor ZAG promotes white adipose tissue browning and energy wasting. *FASEB J.* 2018, 32, 4727–4743. [CrossRef] [PubMed]

- 120. Wang, Y.; Li, Y.M.; Zhang, S.; Zhao, J.Y.; Liu, C.Y. Adipokine zinc-alpha-2-glycoprotein as a novel urinary biomarker presents earlier than microalbuminuria in diabetic nephropathy. *J. Int. Med. Res.* **2016**, *44*, 278–286. [CrossRef] [PubMed]
- 121. Woźny, Ł.A.; Morawiecka-Pietrzak, M.; Jaszczura, M.; Ziora, K.; Grzeszczak, W. The new adipokine zinc-α2-glycoprotein (ZAG) as a link between adipose tissue and kidney? *Endokrynol. Pol.* **2019**, *70*, 171–189. [CrossRef] [PubMed]
- 122. Mracek, T.; Ding, Q.; Tzanavari, T.; Kos, K.; Pinkney, J.; Wilding, J.; Trayhurn, P.; Bing, C. The adipokine zinc-alpha2-glycoprotein (ZAG) is downregulated with fat mass expansion in obesity. *Clin. Endocrinol.* **2010**, *72*, 334–341. [CrossRef] [PubMed]
- 123. Sörensen-Zender, I.; Beneke, J.; Schmidt, B.M.; Menne, J.; Haller, H.; Schmitt, R. Zinc-alpha2-glycoprotein in patients with acute and chronic kidney disease. *BMC Nephrol.* **2013**, *14*, 145. [CrossRef]
- 124. Pelletier, C.C.; Koppe, L.; Croze, M.L.; Kalbacher, E.; Vella, R.E.; Guebre-Egziabher, F.; Géloën, A.; Badet, L.; Fouque, D.; Soulage, C.O. White adipose tissue overproduces the lipid-mobilizing factor zinc α2-glycoprotein in chronic kidney disease. *Kidney Int.* **2013**, *83*, 878–886. [CrossRef]
- 125. Gao, D.; Trayhurn, P.; Bing, C. Macrophage-secreted factors inhibit ZAG expression and secretion by human adipocytes. *Mol. Cell. Endocrinol.* **2010**, 325, 135–142. [CrossRef] [PubMed]
- 126. Bouchara, A.; Yi, D.; Pastural, M.; Granjon, S.; Selag, J.C.; Laville, M.; Arkouche, W.; Pelletier, S.; Fouque, D.; Soulage, C.O.; et al. Serum levels of the adipokine zinc-alpha2-glycoprotein (ZAG) predict mortality in hemodialysis patients. *Kidney Int.* 2018, 94, 983–992. [CrossRef]
- 127. Gong, F.Y.; Zhang, S.J.; Deng, J.Y.; Zhu, H.J.; Pan, H.; Li, N.S.; Shi, Y.F. Zinc-alpha2-glycoprotein is involved in regulation of body weight through inhibition of lipogenic enzymes in adipose tissue. *Int. J. Obes.* **2009**, *33*, 1023–1030. [CrossRef] [PubMed]
- 128. Wei, X.; Liu, X.; Tan, C.; Mo, L.; Wang, H.; Peng, X.; Deng, F.; Chen, L. Expression and Function of Zinc-α2-Glycoprotein. *Neurosci. Bull.* **2019**, *35*, 540–550. [CrossRef]
- 129. Yeung, D.C.; Lam, K.S.; Wang, Y.; Tso, A.W.; Xu, A. Serum zinc-alpha2-glycoprotein correlates with adiposity, triglycerides, and the key components of the metabolic syndrome in Chinese subjects. J. Clin. Endocrinol. Metab. 2009, 94, 2531–2536. [CrossRef]
- 130. Hosseinzadeh-Attar, M.J.; Mahdavi-Mazdeh, M.; Yaseri, M.; Zahed, N.S.; Alipoor, E. Comparative Assessment of Serum Adipokines Zinc-α2-glycoprotein and Adipose Triglyceride Lipase, and Cardiovascular Risk Factors Between Normal Weight and Obese Patients with Hemodialysis. *Arch. Med. Res.* 2017, *48*, 459–466. [CrossRef]
- Leal, V.O.; Lobo, J.C.; Stockler-Pinto, M.B.; Farage, N.E.; Abdalla, D.S.; Leite, M.J.; Mafra, D. Is zinc-α2-glycoprotein a cardiovascular protective factor for patients undergoing hemodialysis? *Clin. Chim. Acta* 2012, 413, 616–619. [CrossRef] [PubMed]
- 132. Yang, L.; Chen, S.J.; Yuan, G.Y.; Zhou, L.B.; Wang, D.; Wang, X.Z.; Chen, J.J. Association of serum adipose triglyceride lipase levels with obesity and diabetes. *Genet. Mol. Res.* **2014**, *13*, 6746–6751. [CrossRef] [PubMed]
- 133. Schoiswohl, G.; Stefanovic-Racic, M.; Menke, M.N.; Wills, R.C.; Surlow, B.A.; Basantani, M.K.; Sitnick, M.T.; Cai, L.; Yazbeck, C.F.; Stolz, D.B.; et al. Impact of Reduced ATGL-Mediated Adipocyte Lipolysis on Obesity-Associated Insulin Resistance and Inflammation in Male Mice. *Endocrinology* 2015, *156*, 3610–3624. [CrossRef] [PubMed]
- 134. Chen, W.; Jiang, Y.; Han, J.; Hu, J.; He, T.; Yan, T.; Huang, N.; Zhang, Q.; Mei, H.; Liao, Y.; et al. ATGL deficiency induces podocyte apoptosis and leads to glomerular filtration barrier damage. *FEBS J.* **2017**, *284*, 1070–1081. [CrossRef]
- 135. Ceperuelo-Mallafré, V.; Näf, S.; Escoté, X.; Caubet, E.; Gomez, J.M.; Miranda, M.; Chacon, M.R.; Gonzalez-Clemente, J.M.; Gallart, L.; Gutierrez, C.; et al. Circulating and adipose tissue gene expression of zinc-alpha2-glycoprotein in obesity: Its relationship with adipokine and lipolytic gene markers in subcutaneous and visceral fat. *J. Clin. Endocrinol. Metab.* 2009, 94, 5062–5069. [CrossRef] [PubMed]
- 136. Alipoor, E.; Esmaillzadeh, A.; Mahdavi-Mazdeh, M.; Yaseri, M.; Zahed, N.S.; Hosseinzadeh-Attar, M.J. The relationship of serum adipokines with malnutrition inflammation score in haemodialysis. *Eur. J. Clin. Investig.* **2017**, *47*, 545–554. [CrossRef] [PubMed]
- 137. Salatzki, J.; Foryst-Ludwig, A.; Bentele, K.; Blumrich, A.; Smeir, E.; Ban, Z.; Brix, S.; Grune, J.; Beyhoff, N.; Klopfleisch, R.; et al. Adipose tissue ATGL modifies the cardiac lipidome in pressure-overload-induced left ventricular failure. *PLoS Genet.* 2018, 14, e1007171. [CrossRef] [PubMed]
- 138. Haemmerle, G.; Lass, A.; Zimmermann, R.; Gorkiewicz, G.; Meyer, C.; Rozman, J.; Heldmaier, G.; Maier, R.; Theussl, C.; Eder, S.; et al. Defective lipolysis and altered energy metabolism in mice lacking adipose triglyceride lipase. *Science* 2006, 312, 734–737. [CrossRef] [PubMed]
- 139. Wölkart, G.; Schrammel, A.; Dörffel, K.; Haemmerle, G.; Zechner, R.; Mayer, B. Cardiac dysfunction in adipose triglyceride lipase deficiency: Treatment with a PPARα agonist. *Br. J. Pharmacol.* **2012**, *165*, 380–389. [CrossRef]