



# **Natural Bioactive Molecules: An Alternative Approach to the Treatment and Control of COVID-19**

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**Abstract:** Several coronaviruses (CoVs) have been associated with serious health hazards in recent decades, resulting in the deaths of thousands around the globe. The recent coronavirus pandemic has emphasized the importance of discovering novel and effective antiviral medicines as quickly as possible to prevent more loss of human lives. Positive-sense RNA viruses with group spikes protruding from their surfaces and an abnormally large RNA genome enclose CoVs. CoVs have already been related to a range of respiratory infectious diseases possibly fatal to humans, such as MERS, SARS, and the current COVID-19 outbreak. As a result, effective prevention, treatment, and medications against human coronavirus (HCoV) is urgently needed. In recent years, many natural substances have been discovered with a variety of biological significance, including antiviral properties. Throughout this work, we reviewed a wide range of natural substances that interrupt the life cycles for MERS and SARS, as well as their potential application in the treatment of COVID-19.

Keywords: bioactive molecules; alkaloids; flavonoids; terpenoids; coronavirus; COVID-19; SARS-CoV-2

## 1. Introduction

Viral infections are currently considered as a public health concern. Viruses are nonliving submicroscopic agents that cause a variety of human ailments. They are often composed of RNA or DNA strains. Measles, human immunodeficiency virus (HIV), influenza, herpes simplex virus, and dengue virus are the most well-known viral diseases [1]. Coronaviruses (CoVs) are members of the *Coronaviridae* family and look like spiky rings under an electron microscope. Their surface is covered in spikes that aid in the attack and binding of living cells [2]. Among the many coronaviruses, notably, coronavirus disease 2019 (COVID-19), first recorded in China in December 2019, is caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). It has already spread across



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**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). the world, infecting millions of people [3,4]. COVID-19 has prevailed almost across the whole world and many countries' healthcare services have now reached their breaking point. The disease spreads through respiratory droplets and direct contact, and city and community disinfection have not proven to be successful in controlling this disease [5]. On 11 March 2020, the World Health Organization (WHO) classified the COVID-19 breakout a pandemic [6]. COVID-19 has affected over 219 countries and territories throughout the world, resulting in around 190 million reported cases and over four million fatalities [7]. Despite the findings of this viral entity and a greater understanding of its mechanism of transmission, SARS-CoV-2 continues to infect hundreds of thousands of individuals every day throughout the world. The new coronavirus, SARS-CoV-2, is closely linked to bat coronavirus, with over 88% nucleotide sequence identity, according to phylogenic research. As a result, it has been categorized as a severe acute respiratory syndrome coronavirus (SARS-CoV) (80% nucleotide similarity), such as the virus that produced the severe acute respiratory syndrome (SARS) epidemic in Guangdong Province, China, in 2002 and 2003, and similar to the Middle East respiratory syndrome coronavirus (MERS-CoV) (% nu-

in the Arabian Peninsula in 2012 [8]. In folk medicine, natural products and their derivatives are utilized to cure a variety of diseases, such as viral infections. Throughout the field of nutraceuticals, herbal drugs have a variety of applications [9–11]. By limiting virus replication, certain natural compounds have been shown to have antiviral characteristics. Aside from plant-derived chemicals, some marine natural products and biotechnologically generated compounds have also been demonstrated to have antiviral properties against various viruses [12–17]. Nature has a wide reservoir of compounds that may be used to research and produce medications for a variety of ailments, including viral infections. A large variety of herbal medications and their ingredients have proven antiviral efficacy in the past [18–20]. There is a scarcity of studies into the creation of anti-coronavirus therapies based on natural compounds. These medicines are vital not only for combating coronavirus, but also for preventing viral infection. Based on previous debates, the goal of this review is to assess the current situation of natural compounds and/or their compounds working against different species of coronavirus.

cleotide similarity) that produced the Middle East respiratory syndrome (MERS) epidemic

## 2. Pathogenesis of COVID-19

COVID-19, an extremely lethal disease produced by the SARS-CoV-2 virus, is a major public health concern around the world. In humans, the SARS-CoV-2 virus enters the lower respiratory tract and causes pneumonia [21]. In the earlier stages, it shows only a little respiratory dysfunction, but later move to a fatal respiratory dysfunction syndrome and hyper inflammation. The SARS-CoV and MERS-CoV diseases appear to have similar immunopathogenic features, where acute respiratory distress syndrome (ARDS) is the major cause of death in most of the infectious diseases, including COVID-19 disease. One of the most noticeable aspects is the cytokine storm, which is an unregulated systemic inflammatory process induced by immune cells producing pro-inflammatory cytokines and chemokines [22,23]. Increased blood stages of chemokines and cytokines, such as basic FGF2, GCSF, MIP1, PDGFB, TNF, GMCSF, IL1, IL9, IL10, IL1RA, IL7, IFN, IP10, IL8, FN, IP10, MCP1, MIP1, and VEGFA, are found in COVID-19 patients [24]. Identical to SARS and MERS-CoV infections, a robust inflammatory immune reaction is induced in serious forms of SARS-CoV-2 disease, which leads to organ failure, ARDS, and ultimately death [22]. The cytokine storm harms the lungs and many other internal organs, which is often the leading cause of death in acute COVID-19, including the heart, kidneys, and liver, which results in multiple organ failure [17,25–28].

## 3. Epidemiology

Transmission by people is the major pathway for getting infected with the SARS-CoV-2 virus, with droplets through sneezing and coughing inhaled via close contact. COVID-19

virus transmission can also occur through contaminated sites or fomites after interaction with the mouth, eye, or nose. Symptomatic patients have a high risk of spreading the virus to others [29]. Asymptomatic patients have less evidence of viral shedding, whereas critically ill patients seem to have higher viral shedding levels [30]. The virus is extremely virulent and was shown to have a lengthy spreading period in China, according to current trends in coronavirus illness epidemiology [31–37]. In the first coronavirus pandemic in Wuhan, an epidemiological analysis of 425 coronavirus patients was conducted, where 56% of the infected individuals were male, with a median age of 59 years. In total, 86.6% of the affected people were between the ages of 30 and 70 on 11 February 2020. Patients had a total mortality rate of 2.3%, with 80.9% of the recorded cases being moderate. Healthcare professionals accounted for about 3.8% of reported cases in hospitals, with 14.6% of these cases being chronic or serious. Infants infected with the virus were observed in only a few cases (2.1%). The coronavirus was found to have the greatest impact on people above the age of 80, representing 14.8% of all cases. For COVID-19 patients to be classed as chronic, the main elements must be present: a  $PaO_2/FiO_2$  higher than 300, oxygen saturated higher than or equal to 93%, respiration larger than or equal to 30 breath/min, and dyspnea, defined as a 50% involvement of the lungs in less than 24 to 48 h. The virus is infectious and at the critical phase in patients with multiple organ dysfunction, leads to septic shock and/or restricted breathing [29,38]. The suggested 14-day initial infection for coronavirus illness is based on the relevant incubation duration for coronaviruses with similar incubation periods after the initial exposure. Incubation takes an average of 5.2 days, with 95% Cl of 4.1 to 7.0, but it can take from 2 to 14 days. Gao et al. also discovered a 9-day average incubation period [39]. Another study revealed that the delta variant's incubation takes an average of 4 days [40]. Associated infections can be found in roughly 22–33% of infected people, and they may be greater in people with severe illnesses [29].

## 4. Etiology

The virus has an 88% sequencing resemblance to two bat-derived severe acute respiratory syndrome (SARS) coronaviruses, but is more distanced from SARS-CoV [41,42]. Here, as consequence, the virus was dubbed a novel coronavirus in 2019. A coronavirus seems to be a single-stranded, encapsulated ribonucleic acid with surface spikes that are 9 to 12 nm long and mimics the solar corona [43-45]. The spike (S) protein interacts with the angiotensin-converting enzyme 2 (ACE2) receptor and facilitates viral entrance into the host cell by mediating fusion of the enveloped and host cell membranes, and the coronavirus genome encodes four key structural protein molecules on the surface [46,47]. Glycosaminoglycans (GAGs) and sialic acid-containing oligosaccharides are examples of such molecules. GAGs are typically found on the cell's outer surface. They are particularly well suited to function as attachment factors to recruit viruses to the cell surfaces because of their position [48–50]. In mammals, heparan sulfate (HS) is one of the most common forms of GAGs. It is a sulfated linear polysaccharide found on the surface of practically all cell types as well as in the extracellular matrix. To produce HS proteoglycans (HSPGs), the HS chains are typically covalently attached as side chains to the core proteins (Figure 1) [51,52].

Many distinct enzymes produce HS in the Golgi apparatus. HS undergoes a number of changes during and after arrangement, including sulfation, acetylation, and epimerization, resulting in glycan structures with a high degree of variation in length, sulfation, and glucuronate/iduronate ratio. In distinct species, organs, tissues, and even at different ages and illness stages, there was significant diversity in the sulfation pattern and degree of HS [54,55]. Many viruses' attachment to host cells during transmission has been demonstrated to be regulated by the sequence and sulfation pattern of HS. Sialylation patterns of cell surface oligosaccharides showed similar results. The MERS-CoV S protein, for instance, selectively attaches  $\alpha 2$ ,3-linked sialic acids on the cell surface over  $\alpha 2$ ,6-linked sialic acids, and 5-*N*-glycolylation and 7,9-*O*-acetylation of sialic acids impairs their interaction. These data suggest that the distribution of distinct forms of HS/sialylated glycans and viral

tropism may have a linkage [56–58]. A deeper knowledge of their interaction could involve the development of novel antiviral drugs. Although there is presently inadequate evidence on the viral tropism of SARS-CoV-2, new research suggests that its tropism may not be totally connected with ACE2 expression. Other variables, such as proteases and glycans, may have a role in determining cellular sensitivity to this viral infection [59–61]. A recent study revealed that HS could bind to the SARS-CoV-2 spike protein's receptor-binding domain (RBD, the C-terminal region of the S1 subunit) and modify its shape. The fascinating potential that differences in HS and sialic acid properties could influence virus tropism prompted us to look into SARS-binding of CoV-2 to a variety of HS and sialic acid-containing oligosaccharides [62,63].



**Figure 1.** SARS-CoV-2 infection and entrance via a plausible pathway. Using the S protein projecting from the viral particle, SARS-CoV-2 may first engage with the HSPGs on the surface of susceptible cells early in the infection cycle. The virus's subsequent interaction with the high-affinity entry receptor ACE2 may be facilitated by this initial attachment. By cleaving the S protein at the S1/S2 and/or S2' sites, the transmembrane protease serine 2 (TMPRSS2) on the host cell surface and other host cell proteases may facilitate viral entry [53]. Adapted with permission from ref. [53]. Copyright 2021 Science China Press.

## 5. Structural Composition of SARS-CoV-2

With a genomic diameter varying from 27 to 32 kilobases in size (~125 nm or 0.125  $\mu$ m), SARS-CoV-2 belongs to the biggest RNA virus family. It is an enveloping RNA virus with a single-stranded (+ssRNA) positive-sense RNA genome, with a 5' cap shape and also a 3' poly-A tail [64]. SARS-CoV-2 shares some characteristics with other virus infections in this family. The envelope (E), membrane (M), spike (S), and nucleocapsid protein (N) are four critical structural proteins that influence the virus's function and shape [65]. The most important of these four proteins are N and S, the former of which aids in the optimal utilization of the capsid and the entire viral structure, and the latter of which aids in the future growth of the capsid, with the full viral framework facilitating virus attachment to host cells [66,67]. The three primary elements of the S protein are a huge ectodomain, a short intracellular tail, and also a single-pass transmembrane anchor. Those are all necessary for the host cells to be anchored. The S1 receptor attaching subunits and the S2

membrane fusion subunit are the two parts of the ectodomain (Figure 2). The clove-trimeric or crown shape in which such subunits are located gives the virus its name [68]. According to studies, SARS and SARS-CoV-2 have identical receptors within viral genomes, especially in the receptor-binding domain (RBD) and receptor-binding motif (RBM). Throughout SARS infection, the RBM of the S protein is intimately related to ACE2 in mammalian or host cells. The coronavirus's principal targets are indeed the kidneys, lungs, and gut, while the ACE2 protein is detected in a range of human organs [69–73]. Although no research has established that the virus may impair men's fertility or sexual potency because of the virus's new nature, doctors in Wuhan have speculated that the condition may influence sperm production, a low sperm rate, and the production of male sex hormones, resulting in low libido. SARS-CoV-2 also attacks host cells through ACE2 receptors, leading to COVID-19-related pneumonia, acute myocardial damage, and long-term cardiovascular injury [74].



**Figure 2.** The structure of the SARS-CoV-2 virus. Clathrin is a protein that plays a vital role in the formation of coated vesicles. The S protein is an important element of the viral shape that helps the virus to connect with the host receptor cells. The S protein comprises two parts: the receptor-binding component S1 and the membrane fusion component S2; the former attaches to the human host cell's ACE2 receptor, while the latter internalizes and forms a membrane fusion between the viral component and the ACE2 receptors. This causes the viral RNA to be released into the host cell, resulting in respiratory infection [75]. Adapted with permission from ref. [75]. Copyright 2020 Elsevier B.V.

#### 6. SARS-CoV-2: Suggested Mode of Action

SARS-CoV-2 shares similarities with SARS-CoV, but it does have a much higher risk of transmission and virulence; this higher rate of spread could be because of a gain of point mutations, which distinguishes this distinct virus from SARS-CoV. The 8a segment of SARS-CoV-2 is absent, the 8b and 3b segments are longer and also shorter, and the Nsp 2 and 3 proteins are changed (Figure 3) [75].

SARS-CoV-2 Nsp 2 is a mutation that is thought to be linked to the virus's capacity to be more infectious [76]. The SARS-CoV-2 orf8 and orf10 proteins are distinct. It can be helpful to learn more about the physiological functions of such proteins. Understanding the function of these proteins might be valuable. Moreover, a furin-like cleave site throughout the S protein has been revealed in novel pathogenic viruses, which is lacking in SARS-CoV but active in SARS-CoV-2. This might be the cause of SARS-CoV-2's increased pathogenicity. SARS-CoV-2 also interacts with almost the same ACE2 receptor as SARS-CoV, but with a significantly stronger affinity; this might explain the enhanced transmission rate and

ease with which it may infect other species. The S1 portion of the S protein, which has S1 just on the N terminal and S2 on the C terminal, contains RBD. The S2 domain of an S protein contains the fusion protein, an S2, an internal fusion peptide (FP), and two heptad-repeat domains surrounding the transmembrane domain (TM). SARS-CoV-2 and SARS-CoV has similar internal FPs [77]. SARS-CoV recruits proteases once it binds to the receptor and cleaves the S protein into S1 and S2 domains. S2 undergoes a structural shift as a result of the fragmentation, which is accompanied by the incorporation of the FP into the membrane and membrane fusion, allowing the virus to more readily enter the cell. ACE2 is broken and released into the additional membrane region by ADAM17 as soon as the virus penetrates the cell. Alveolar injury and higher pulmonary vascular permeability have both been associated with decreased ACE2 [78]. Once the virus's proteins are imported into the cell, the ORF3a protein is generated, which encodes for a  $Ca^{2+}$  ion pathway identical to SARS and SARS-CoV-2. ORF3a links with TRAF3 to stimulate the nuclear factor kappa B (NF-kB) pathway, which causes the pro-IL-1B gene e to be transcribed [79]. ORF3a also recruits an inflammasome complex with TRAF3. NLRP3, ASC, and caspase 1 make up this complex. A secondary signal, such as Ca<sup>2+</sup> influx, activation caspase, reactive oxygen species (ROS) generation, or mitochondrial injury convert pro-IL-1B to IL-B, culminating in cell proliferation. The extended ORF8b protein in SARS-CoV-2 also activates the inflammasome system via NLRP3 [80]. A cytokine storm happens because all of those channels are active, leading to the respiratory problem, which is a common sign of COVID-19. The JNK cascade, which is triggered by ORF3a, ORF3b, and ORF7a, may result in higher generation of pro-inflammatory mediators, resulting in greater lung injury (Figure 4) [81].



**Figure 3.** Graphical representation of the comparisons between SARS-CoV and SARS-CoV-2. N: nucleocapsid protein; E: envelope; M: membrane; S: spike.



**Figure 4.** Potential mode of action of SARS- CoV-2, illustrated as SARS-CoV-2 attached to its target ACE-2. After that, the S1 and S2 molecules are cleaved, and ADAM 17 sheds ACE-2. As a consequence, the body's level of Angiotensin II rises, resulting in respiratory discomfort. During contact, the virus fuses with the membrane and penetrates the cell, where protein synthesis and replication occur. ORF8b, ORF3a, and E proteins, and also the pathway of NF-κB, all stimulate the inflammasome path, resulting in cytokine stimulation in different ways. A cytokine storm ensues, leading in respiratory discomfort. Here, ACE2, angiotensin converting enzyme 2; NF-κB, nuclear factor- kappa B; ROS, reactive oxygen species; P65, P105, and P50 are the subunit of NF-κB transcription complex; TRAF3, TNF receptor associated factor 3; ATR maintain to genome integrity by stabilizing replication forks and by regulating cell cycle progression and DNA [75]. Adapted with permission from ref. [75]. Copyright 2020 Elsevier B.V.

#### 7. Therapeutic Approach against COVID-19

#### 7.1. Pharmacological Drugs

## 7.1.1. Remdesivir

Remdesivir (GS-5734) is an adenosine analog antiviral prodrug that was used to tackle the Ebola virus infection in Western Africa from December 2013 to January 2016. By attaching RNA-dependent RNA polymerase, remdesivir's active form (GS-441524) suppresses viral RNA multiplication [68,82-84]. Remdesivir is an antiviral drug that works against such a range of single-stranded viruses such as RNA and includes paramyxoviruses, filoviruses, pneumoviruses, and coronaviruses, including SARS- and MERS-CoV [84,85]. Remdesivir is a prodrug that is metabolized into GS-441524, an adenine nucleotide analogue that inhibits viral RNA production. Remdesivir functions initially in the target cells, decreasing viral RNA rates in a dose-dependent manner [82], which correlates to viral load dysfunction in vitro. For the Ebola virus, SARS-, and MERS-CoV, similar modes of action of remdesivir have been established in vitro [86,87]. In cultured cells experiments, remdesivir prevents SARS-CoV-2 illness in simian Vero E6 cells infected with SARS-CoV-2 at 1.76  $\mu$ M in an 90% effective concentration (EC<sub>90</sub>), a quantity achieved in wild monkey experiments. Remdesivir was also shown in virus-prone Huh-7 cells of human liver cancer to prevent SARS-CoV-2 infection [88]. Nowadays, remdesivir's efficacy in preventing and treating MERS-CoV infection in nonhuman primates has been shown [89]. Patient safety data for remdesivir was obtained from a randomized, clinical study of Ebola virus treatment that was done in August 2018 in response to the Ebola virus outbreak in the Democratic Republic of Congo. A sample of 175 individuals was given remdesivir at a

packed dose of 200 mg on Day 1 and then a maintenance dose of 100 mg for 13 days; nine of the people investigated had major adverse effects, demonstrating that remdesivir is a safe medicine [90].

#### 7.1.2. Chloroquine (CQ) and Hydroxychloroquine (HCQ)

CQ and HCQ are most commonly used as antimalarial drugs, having lysosomotropic effects. Particularly in comparison to CQ, HCQ is a less hazardous antimalarial drug made by combining chloroquine with an OH group. The effects of these antimalarial medications are widely found in the whole body, including in the lungs, according to pharmacokinetic studies [87]. They are now being researched as a potential therapy for SARS-CoV-2 because of their immunomodulating and anti-inflammatory properties [88]. Although preclinical research suggests that CQ and HCQ can reduce viral replication and may prevent COVID-19 infection, present evidence does not support their efficacy as a SARS-CoV-2 infection prophylaxis [91]. Experts recommended to utilize the CQ/HCQ regimen for SARS-CoV-2 infection prophylaxis, notably among healthcare workers who are at a higher risk of infection [92,93]. However, this opinion was refuted by data from a well-designed randomized controlled trial on 821 participants. Within four days of exposure, participants were randomly assigned to receive either HCQ or the placebo. The occurrence of novel COVID-19-related symptoms did not differ significantly between the two groups (11.8% versus 14.3%; P = 0.35) [94]. Another study also showed differences between CQ/HCQ and the placebo, but leading without showing any significant effect againt SARS-CoV-2 [95].

#### 7.1.3. Lopinavir/Ritonavir

Lopinavir is an extremely effective blocker of HIV protease, which is necessary for intracellular HIV replication. It was created in 1998 to combat HIV tolerance of the protease inhibitor ritonavir, which is produced by a variation of valine at position 82 (Val 82) in HIV protease's active site in reaction to ritonavir treatment. Because lopinavir decomposition is inhibited by ritonavir, concomitant oral treatment of ritonavir and lopinavir exceeded the in vitro antiviral half maximal effective concentration ( $EC_{50}$ ) of lopinavir in monkeys, dog, and rat plasma by over 50-fold after 8 h [96]. As a result, when taken in combination with other antiretroviral drugs, the combination of lopinavir and ritonavir has indeed been acknowledged as an excellent orally administered medicine in the treatment of HIV-infected persons [97,98].

#### 7.1.4. Tocilizumab

Tocilizumab is a humanized monoclonal antibody that targets the interleukin-6 receptor (IL-6R) and has been allowed by the FDA to treat rheumatoid arthritis. In critically sick COVID-19, the cytokine release storm (CRS) has been mediated by IL-6. As a result, it has been recommended as a treatment for such patients [73,99]. In China and Italy, tocilizumab has been used as an immuno-suppressive medication in COVID-19 patients with impressive results. In China, individuals with COVID-19 who were given tocilizumab, showed massive improvement, implying that tocilizumab could be very effective in curing people with acute disease [100–102].

## 7.1.5. Favipiravir

Toyama Chemical, Japan, developed favipiravir that inhibits the RNA-dependent RNA polymerase (RdRp) of RNA viruses and produces lethal RNA transversion mutations, resulting in a nonviable viral phenotype [103–105]. Favipiravir inhibits the replication of a wide range of RNA viruses, such as influenza A, Ebola, and Lassa viruses [106]. The favipiravir-treated individuals had a substantially superior therapeutic response, with quicker viral clearance and a higher incidence of improvement in chest imaging. Based on these promising results, China's National Medical Products Administration has authorized favipiravir as the country's first anti-COVID-19 drug [107].

# 7.1.6. Umifenovir

Umifenovir is a small indole-derivate molecule that protects against viral infection by inhibiting clathrin-mediated endocytosis, which hinders membrane fusion between the viral particle and the host cell's cytoplasmic membrane (Figure 5). In Russia and China, umifnovir is approved for the treatment and control of influenza A and B viruses, as well as for many other respiratory pathogens [106,108–110]. Around 36 COVID-19 patients had taken 400 mg umifenovir three times per day for nine days in a clinical pilot project in Wuhan, China, with 31 uncontrolled COVID-19 patients taking part as a control condition. When compared with the control group, umifenovir therapy was found to have a higher proclivity for reducing viral load as determined by RT-PCR, and also a reduced fatality rate of 0% vs. 16% [111]. Table 1 illustrates some combinational drugs that can act as potential targets for COVID-19.



Figure 5. The structures of some effective COVID-19 treatment pharmacological drugs.

Tabl	e 1.	Com	binational	drugs	as p	otential	l C	O	VII	D-19	ereatment.
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Drugs	<b>Targeted Virions Infection</b>	<b>Targeted Virions Modes</b>	References
HCQ and Itazoxanide	SARS-CoV-2	Used as adjuvant treatment in COVID-19	[111]
Darunavir and Umifenovir	SARS-CoV-2	Inhibition of viral replication reduces viral load	[112]
CQ and HCQ	SARS-CoV-2	Reduces viral loads in the lungs and increases pulmonary activity	[113]
Interferon beta, Lopinavir and ritonavir	MERS-CoV and SARS-CoV-2	The viral load was reduced little, and pulmonary function improved slightly	[114]
Ribavirin and Interferon-α	SARS-CoV-2	ARDS was reduced, as was mortality	[115]
Camostat mesilate HCQ	SARS-CoV-2	Blockers of the angiotensin receptor and inhibitors of the serine protease of the host cell	[116]

## 7.2. Natural Products for COVID-19 Treatment

Nonetheless, the expensive healthcare expenditure, lack of availability, unprecedented drug side effects, and ethical considerations regarding convalescent plasma therapy (CPT) make them difficult to execute globally. Furthermore, the problem can be made worse if the virus evolves into drug-resistant mutants, rendering antiviral medications worthless because most of them target specific viral proteins [117]. As a result, we can search for new therapies using natural products. Some of the potential natural products antiviral medications utilized for the management and prophylaxis of COVID-19 are reviewed in the following sections of this review (Table 2).

Sl. No.	Name of Natural Product	Class of the Compound	Source	<b>Biological Efficacy</b>	References
1	Daurisoline	Alkaloid	Rhizoma menispermi	Increased endolysosomal pH, lowered active cathepsin levels, and impaired V-type ATPase activity (EC of 10 μM)	[118,119]
2	Dauricine	Alkaloid	Rhizoma menispermi	Increased endolysosomal pH, lowered active cathepsin levels, and impaired V-type ATPase activity (EC of 10 μM)	[120]
3	Tetrandrine	Alkaloid	Stephania tetrandra	Increased endolysosomal pH in a concentrationdependent manner (EC of 1–10 μM)	[121,122]
4	Luteolin	Flavonoid	Rhodiola kirilowii	$IC_{50} = 4.5 \ \mu M$	[123,124]
5	Quercetin	Flavonoid	Allium cepa	$IC_{50} = 83.4 \ \mu M$	[125]
6	Kazinol A	Flavonoid	Broussonetia papyrifera	$IC_{50} = 84.8 \ \mu M$	[126]
7	Kazinol F	Biphenyl propanoids	Broussonetia papyrifera	$IC_{50} = 43.3 \ \mu M$	[127]
8	Kazinol J	Biphenyl propanoids	Broussonetia papyrifera	$IC_{50} = 64.2 \ \mu M$	[127]
9	Kaempferol	Flavonoid	Zingiber officinale	$IC_{50} = 16.3 \ \mu M$	[128]
10	Neobavaisoflavone	Flavonoid	Psoralea corylifolia	$IC_{50} = 18.3 \ \mu M$	[129]
11	Papyriflavonol A	Flavonoid	Broussonetia papyrifera	$IC_{50} = 3.7 \ \mu M$	[126]
12	Psoralidin	Flavonoid	Psoralea corylifolia	$IC_{50} = 4.2 \ \mu M$	[129]
13	Tomentin A	Flavonoid	Paulownia tomentosa	$IC_{50} = 6.2 \ \mu M$	[130]
14	Tomentin B	Flavonoid	Paulownia tomentosa	$IC_{50} = 6.1 \ \mu M$	[130]
15	Tomentin C	Flavonoid	Paulownia tomentosa	$IC_{50} = 11.6 \ \mu M$	[130]
16	Tomentin D	Flavonoid	Paulownia tomentosa	$IC_{50} = 12.5 \ \mu M$	[130]
17	Tomentin E	Flavonoid	Paulownia tomentosa	$IC_{50} = 5.0 \ \mu M$	[130]
18	Catechin	Flavonoid	Camellia sinensis	Elevated $Zn^{2+}$ level (2-fold increase at EC of 50 $\mu$ M)	[131]

Table 2. List of some natural products, their classification, and possible mechanism of action.

Sl. No.	Name of Natural Product	Class of the Compound	Source	<b>Biological Efficacy</b>	References
19	Epigallocatechin-3- gallate (EGCG)	Flavonoid	Camellia sinensis	Elevated intracellular Zn <sup>2+</sup> level (2-fold increase at EC of 50 μM)	[131]
20	Rutin	Flavonoid glycoside	Morus alba	Elevated intracellular Zn <sup>2+</sup> level (4-fold increase at EC of 50 μM)	[131]
21	Apigenin	Flavonoid	Adinandra nitida	30.3% suppression at EC of 500 μg/mL	[132,133]
22	Camellianin A	Flavonoid	Adinandra nitida	30.2% suppression at EC of 500 μg/mL	[132,133]
23	Camellianin B	Flavonoid	Adinandra nitida	40.7% suppression at EC of 500 μg/mL	[132,133]
24	Taxifolin	Flavonoid	Coreopsis tinctoria	$IC_{50} = 145.7 \ \mu M$	[134]
25	Myrtenal	Terpene	Elettaria cardamomum	Suppressed the action of V-type ATPase and reduced endolysosomal acidification (EC of 100 µM)	[135]
26	Pulsatilla saponin D	Triterpenoid saponin	Pulsatilla chinensis	Increased endolysosomal pH and downregulated cathepsins (EC of 1.25 µM)	[88]
27	Betulinic acid	Terpenoid	Breynia fruticose	$IC_{50} = 10.0 \ \mu M$	[136,137]
28	Leelamine	Terpene	Pinus sylvestris	Decreased endolysosomal acidity and suppressed cellular endocytosis (EC of 3 µM)	[138]
29	Curcumin	Polyphenol	Curcuma longa	$IC_{50} = 5.7 \ \mu M$	[137]
30	Caffeic acid	Phenolic acid	Ocimum basilicum	Elevated intracellular Zn <sup>2+</sup> level (3-fold increase at EC of 50 μM)	[131]
31	Catechol	Phenol	Allium cepa	Elevated intracellular Zn <sup>2+</sup> level (2-fold increase at EC of 50 μM)	[131]
32	Gallic acid	Phenolic acid	Syzygium aromaticum	Elevated intracellular Zn <sup>2+</sup> level (4-fold increase at EC of 50 μM)	[131]
33	Resveratrol	Polyphenol	Vitis vinifera	Elevated intracellular Zn <sup>2+</sup> level (7.5-fold increase at EC of 10 μM)	[139]
34	Methyl gallate	Phenolic acid	Tamarix hohenackeri	35.7% suppression at EC of 20 mg/mL	[140]
35	Tannic acid	Phenolic acid	Camellia sinensis	$IC_{50} = 5.7 \ \mu M$	[141]
36	Pd-C-I	Coumarin	Angelica decursiva	$IC_{50} = 6.8 \ \mu M$	[142]
37	Pd-C-II	Coumarin	Angelica decursiva	$IC_{50} = 12.4 \ \mu M$	[142]
38	Pd-C-III	Coumarin	Angelica decursiva	$IC_{50} = 15.3 \ \mu M$	[142]
39	Isorutarine	Coumarin	Angelica decursiva	$IC_{50} = 68.4 \ \mu M$	[142]
40	Ampleopsin C	Stilbenoid	Vitis thunbergiivar	$IC_{50} = 18.4 \ \mu M$	[143]

## Table 2. Cont.

## 7.2.1. Alkaloids Derivatives

Homoharringtonine is a cytotoxic alkaloid isolated in the *Cephalotaxus hainanensis* medicinal plant. It has been approved by the FDA as a treatment for resistant chronic myeloid leukemia. Homoharringtonine has the smallest half maximal inhibitory concentra-

tion (IC<sub>50</sub>) and has significant antiviral efficacy against a number of human and animal coronaviruses [144]. The antiallergic, antimalarial, antibacterial, and antiviral effects of isatin (1H-indole-2,3-dione), an oxidizing indole derivative, have been found in nature, including *Isatis tinctoria* and *Calanthe discolor*. SARS-CoV 3C-like protease (3CL<sup>pro</sup>) was suppressed in tiny doses by isatin derivatives [145–147]. Rhinovirus and SARS-CoV have identical protease architectures. In coronavirus-infected swine testicular cells, *Tylophora indica*'s tylophorine and 7-methoxycryptopleurine were found to limit viral prolification. In this investigation, 7-methoxycryptopleurine IC<sub>50</sub>:20 nM was found to be more effective than tylophorine (IC<sub>50</sub>:58 nM). In previous studies, tylophorine was demonstrated to affect viral RNA proliferation and cellular JAK2-mediated dominating nuclear factor kappa B (NF-kB) activation in CoV at doses of 0–1000 nM [148,149].

As per the MTS testing for virus-induced cytopathic impact, *Lycoris radiata* extract has high antiviral efficacy against SARS-CoV. The active ingredient in this extraction is lycorine, an alkaloid with an EC<sub>50</sub> of 15.7 1.2 nM, showing antiviral action. These results indicate that lycorine could be a promising candidate for developing novel antiviral medicines. In vitro, lycorine also inhibited the reproduction of coronaviruses such as HCoV-OC43 (EC<sub>50</sub>:0.15  $\mu$ M), MERS-CoV (EC<sub>50</sub>:1.63  $\mu$ M), and HCoV-NL63 (EC<sub>50</sub>:0.47  $\mu$ M), according to another study [150,151].

Anticancer, anti-inflammatory, and antioxidant effects are reported for bisbenzylisoquinoline alkaloids found in the roots of *Stephania tetrandra*. The main active *S. tetrandra* alkaloids that have also showed antiviral efficacy towards human coronavirus-OC43 (HCoV-OC43) (betacoronavirus 1) disease, are tetrandrine (IC<sub>50</sub>: 14.51  $\mu$ M), fangchinoline (IC<sub>50</sub>: 12.40  $\mu$ M), and cepharanthine (IC<sub>50</sub>: 10.54  $\mu$ M). Emetine, also an alkaloid, is the active ingredient of *Carapichea ipecacuanha* roots that contains anti-protozoal and vomiting medicines properties. Many coronaviruses, including HCoV-OC43 (EC<sub>50</sub>: 0.30  $\mu$ M), MERS-CoV (EC<sub>50</sub>: 0.34  $\mu$ M), and human coronavirus-NL63 (HCoV-NL63) (alphacoronavirus), were suppressed in vitro by emetine (EC<sub>50</sub>: 1.43  $\mu$ M). Additionally, emetine can protect host cells against MERS-CoV infection [152]. Many coronaviruses, such as HCoV-OC43 (EC<sub>50</sub>: 0.30  $\mu$ M), MERS-CoV (EC<sub>50</sub>: 0.34  $\mu$ M), and HCoV-NL63, were suppressed in vitro by emetine (EC<sub>50</sub>: 1.43  $\mu$ M). And HCoV-NL63, were suppressed in vitro by emetine (EC<sub>50</sub>: 0.34  $\mu$ M), and HCoV-NL63, were suppressed in vitro by emetine (EC<sub>50</sub>: 1.43  $\mu$ M). Moreover, emetine can protect host cells from MERS-CoV infection [152].

#### 7.2.2. Polyphenols and Flavonoids Derivatives

Twelve geranylated flavonoids, including five new compounds 8 (tomentin A–E) (2.39–2.43) identified from the traditional Chinese medicinal (TCM) plant *Paulownia tomentosa* (Thunb.) Steud., inhibited SARS Papain-Like Protease (PL<sup>pro</sup>) in a mixed-type manner, with IC<sub>50</sub> values ranging from 5.0 to 14.4  $\mu$ M. Tomentin A, B, and E were revealed to be the most effective PL<sup>pro</sup> inhibitors in this group, with IC<sub>50</sub> values of 6.2, 6.1, and 5.0  $\mu$ M, respectively. Each of these novel compounds with the dihydro-2H-pyran moiety inhibited more effectively than their parent compounds [130].

Similarly, six flavonoids isolated from *Cullen corylifolium* (L.) Medik. seeds (bavachinin, neobavaisoflavone, isobavachalcone, 4'-O-methylbavachalcone, psoralidin, and corylifol A) showed mixed-type inhibition against SARS-CoV PL<sup>pro</sup>, with IC<sub>50</sub> values ranging from 4.2 to 38.4  $\mu$ M [153].

Bioflavonoids, such as amentoflavone isolated from *Torreya nucifera*, have been shown to have noncompetitive  $3\text{CL}^{\text{pro}}$  inhibitory action with low micromolar IC<sub>50</sub> values. The most powerful inhibitor (IC<sub>50</sub> = 8.3 µM) was found to be amentoflavone (2.6), which was much more potent than the parent chemical apigenin (IC<sub>50</sub> = 280.8 µM). Other apigenin-containing flavones, such as luteolin (2.23) (IC<sub>50</sub> = 20.2 µM) and quercetin (2.29) (IC<sub>50</sub> = 23.8 µM), similarly inhibited  $3\text{CL}^{\text{pro}}$  more than the parent, showing that the apigenin moiety at position C-3' of flavones is crucial for effectiveness. Honeysuckle's primary flavonoid, luteolin, is a component of *Lianhua qingwen*, a TCM for COVID-19 [154].

Baicalin is a glycosylated flavonoid derived from *S. baicalensis* that displays antiviral activity against by the fRhK-4 cell line's prototype virus (EC<sub>50</sub> 12.5  $\mu$ g/mL). At concentra-

tions of 0.1  $\mu$ M and higher, baicalein reduced the cell damage caused by SARS-CoV-2 and enhanced the morphology of Vero E6 cells. Oral administration of 200 mg/kg crystal form  $\beta$  of baicalein to rats resulted in an effective concentration. Baicalein also reduced body weight loss, virus multiplication, and lung tissue lesions in hACE2 transgenic mice treated with SARS-CoV-2 [130,155,156].

Polyphenols obtained from *Angelica keiskei* have chalcones with a C-5 prenyl that exhibit strong inhibitory action against  $3CL^{pro}$  and  $PL^{pro}$  in vitro. Noncompetitive inhibition of  $PL^{pro}$  was shown by alkylated chalcones, with the most effective compounds being xanthoangelol E (IC<sub>50</sub>: 1.2  $\mu$ M) and xanthoangelol F (IC<sub>50</sub>: 5.6  $\mu$ M). According to the analysis of SAR, the perhydroxyl member of a chalcone is an alkylated chalcone with a stronger inhibitory effect [157].

Resveratrol is a stilbenoid found in *Vitis vinifera, Vaccinium macrocarpon,* and *Polygonum cuspidatum,* among other plants. Hepatoprotective, cardioprotective, neuroprotective, antiinflammatory, and antibacterial properties are only some of the pharmacological and therapeutic benefits of resveratrol. In vitro, resveratrol significantly inhibit MERS-CoV proliferation and reduced MERS-CoV infection. As a consequence, resveratrol is an important anti-MERS medication and could be a viable SARS-CoV2 antiviral [158,159].

The structure–activity relationship (SAR analysis) of quercetin-3-galactoside and its replaced analogues exposes (1) that the 4 OH groups upon a quercetin moiety are important for biological action; (2) that trying to remove the 7-OH decreases the 3CL<sup>pro</sup> inhibitory effect; (3) that the sugar moiety is important for action; and (4) that sugar alterations have no influence on inhibitor efficacy [160].

Myricetin and scutellarein are obtained from *Nigella sativa*, which have been reported in many studies. At concentrations of 0.01–10  $\mu$ M, myricetin and scutellarein inhibit SARS-CoV 3CL<sup>pro</sup>. Broussochalcone B, broussochalcone A, 4-hydroxyisolonchocarpin, papyriflavonol A,4,7-trihydroxyflavane, kazinol A, kazinol B, broussoflavan A, kazinol F, and kazinol J are all obtained from *Broussonetia papyrifera*, which are also responsible for inhibiting SARS-CoV [126,161,162].

## 7.2.3. Terpenoid Derivatives

Quinone-methide triterpenes are a kind of terpene found solely in the Celastraceae family of plants, such as *Tripterygium regelii*. With an IC<sub>50</sub> of 2.6–10.3  $\mu$ M, these compounds demonstrated modest inhibitory action towards 3CL<sup>pro</sup>. The presence of a quinone-methide molecule, according to SAR analysis, plays a substantial role in 3CL<sup>pro</sup> inhibition [163].

Saikosaponins are the main pharmacological active triterpenoids, usually as glucosides, found from TCM such as *Bupleurum* spp., *Heteromorpha* spp., and *Scrophularia scorodonia*, with antiviral and immunomodulatory potential [164]. Four saikosaponins, namely, saikosaponin A, B2, C, and D (5–25 M/L), demonstrate action towards human coronavirus-229E (CoV-229E) (alphacoronavirus) with EC<sub>50</sub> values of 8.6, 1.7, 19.9, and 13.2  $\mu$ M, respectfully; saikosaponin B2 suppressed viral adherence and penetration stages [165].

Triterpenoids and 3-friedelanol obtained from *Euphorbia neriifolia* leaves were tested in vitro for anti-HCoV efficacy in 2012. 3-Friedelanol with a triterpenoid showed through screening a more potential antimicrobial action and increased cellular viability after incubation with HCoV. Furthermore,  $3\beta$ -fridelanol showed strong inhibitory action towards  $3CL^{pro}$  [166,167].

*Glycyrrhiza glabra* and glycyrrhizin, its active ingredient, have antiviral action against a variety of viruses, including hepatitis A, B, C, varicella-zoster, HIV, and herpes simplex type-1 [168].

Salvia miltiorrhiza produces tanshinones with an abietane diterpene structure. Tanshinones have several biological actions, including anti-inflammatory, cardiovascular, and anti-tumor properties. These compounds preferentially block the SARS-CoV 3CL<sup>pro</sup> and PL<sup>pro</sup> enzymes, and their effectiveness varies depending mostly on enzyme subtype. Some tanshinones inhibit PL<sup>pro</sup> more potently (IC<sub>50</sub> varying from 0.8 to 30.0  $\mu$ M) [169].

Ferruginol, a natural phenol with a terpenoid substructure derived from *Sequoia sempervirens*, has been shown to have anticancer effects in humans with colon, breast, and lung malignancies. Furthermore, at 0–80  $\mu$ M, ferruginol, betulonic acid, betulinic acid, hinokinin, savinin, and curcumin are some of the compounds found in turmeric, which reduced SARS-CoV replication substantially [136,170]. The structures of some effective COVID-19 treatment natural products are shown in Figure 6.



Figure 6. Cont.





**Figure 6.** The structures of some effective COVID-19 treatment natural products (alkaloids, polyphenols, flavonoids, and terpenoids derivatives).

#### 7.2.4. Miscellaneous Compounds

Sivestrol, a natural substance derived from the fruit of *Aglaia foveolata*, has been demonstrated to exhibit highly potent in vitro cytotoxic activity against a number of human cancer cell lines. Furthermore, at doses of 0.6–2  $\mu$ M, this drug inhibited nondependent viral mRNA synthesis of HCoV-229E with an IC<sub>50</sub> of 40 nM [171,172].

One of the promising antibiotic treatments against SARS-CoV is valinomycin with a cyclododecadepsipeptide architecture, which was discovered in *Streptomyces tsusimaensis* and has minimum cytotoxicity and great efficiency against CoV [173].

Phycocyanin, lutein, polysaccharides, vitamins, and other phenolics were found to have antibacterial, anticancer, anti-inammatory, and other important pharmacological effects in marine microalgae belonging to the phyla Rhodophyta and Phaeophyta [120]. Hirata et al. [174] investigated the antiviral and antioxidative properties of phycocyanobilins, a type of tetrapyrrole chromophores found in some marine cyanobacteria. In silico molecular docking tests conducted by Pendyala and Patras in 2020 revealed that phycocyanobilins had a significant binding affinity for the SARS-CoV-2 main protease (M<sup>pro</sup>) and RdRp. Lectins are a type of molecule with a strong affinity for carbs. Griffithsin, a lectin produced from red algae, was investigated for its possible application, and tests have revealed that it has antiviral action against human immunodeficiency virus-1 (HIV-1) and hepatitis C [175–177]. A recent in vitro study by Millet et al. [178] revealed that griffithsin had inhibitory action against MERS-CoV. Esculetin ethyl ester from Axinella cf. corrugate, a marine sponge, demonstrated a high interaction with SARS-CoV-2 protease and could be employed as an anti-COVID-19 drug [179]. Carrageenans, a type of sulphated polysaccharide found in the sea, are considered to be virus inhibitors. They work by preventing the virus from attaching and then being internalized. As a result, Nagle et al. [180] hypothesized that these compounds could be used as coating materials on hygienic products to inhibit COVID-19 infection. In silico analyses have recently contributed in the identification of potential lead compounds for therapeutic development against the COVID-19 pandemic. In a molecular dynamic research, Khan et al. [181] reported four effective SARS-CoV-2 Mpro inhibitors from marine sources (fostularin 3,1-hexadecoxypropane-1, 2-diol, palmitoleic acid, 15 alphamethoxypuupehenol, and puupehedione) that can be used to disrupt the viral life cycle in the host.

Lactoferrin (LF), a transferrin-family glycoprotein found in many different of human secretions, is known to bind and transport iron and to play a key role in iron homeostasis regulation. In vitro studies on human intestinal, liver, and T cell lines revealed that LF possesses promising antiviral and antibacterial properties, as well as anti-inflammatory and immune-modulating properties [182–185]. Many investigations have shown that LF has potent antiviral activity against viruses such as hepatitis C virus, herpes simplex virus, human immunodeficiency virus, poliovirus, and rotavirus, with  $EC_{50}$  values in the micromolar range in vitro [186,187]. The LF antiviral action is most evident in the early stages of infection, when it prevents viral particles from entering host cells by binding directly to them or inhibiting the virus receptor or co-receptor on the host cell. Moreover, LF can inhibit some viruses from internalizing, including the SARS pseudovirus, by binding to cell-surface HSPGs, which have been demonstrated to be required co-factors for SARS-CoV-2 infection [188–190]. Furthermore, LF has been found to prevent the entry of murine coronavirus and human coronaviruses such as hCoV-NL63, which are closely related to SARS-CoV-2 [191,192]. SARS-CoV-2 uses endocytosis as a cell-entrance mechanism, and LF has been shown to preferentially suppress cathepsin L, a lysosomal peptidase important for endocytosis. The immunomodulatory and anti-inflammatory properties of LF are another essential component of its bioactivity [192–195]. In experimental settings simulating sepsis, LF was shown to lower IL6 and tumor necrosis factor-alpha (TNF- $\alpha$ ). Therefore, it is feasible that LF can control the hyperactive immunological and inflammatory response to SARS-CoV-2, which can lead to acute respiratory distress and death in some individuals. Overall, LF has the potential to be a non-toxic health supplement that can be used to prevent infection as well as a supplementary treatment for those who have COVID-19 [26,196,197].

Vitamin C is a water-soluble vitamin with antioxidant effects that supports the epithelial barrier against pathogen entrance and the cellular functioning of the innate and adaptive immune systems in the immune system. Environmental factors, such as air pollution, and the presence of diseases, such as type 2 diabetes, can affect vitamin C levels in the body [198]. Vitamin C deficiency affects the older population in particular because chronic or acute disorders are frequent in this group, and aging is linked to lower vitamin C levels [199–201]. Furthermore, vitamin C may control the cytokine storm, which is characterized by elevated levels of the pro-inflammatory cytokine IL-6, increasing the risk of respiratory failure necessitating mechanical ventilation in COVID-19 patients. Pretreatment with vitamin C, according to an in vivo study involving 12 healthy males, can lower the amounts of IL-6 generated by the vasoconstrictor endothelin-1 (ET-1), lowering vascular dysfunction. Furthermore, elevated ET-1 expression has been linked to pneumonia, pulmonary hypertension, interstitial lung fibrosis, and acute respiratory distress syndrome (ARDS) [89,202–204].

In pulmonary epithelial cells, the vitamin D receptor (VDR) is present. When VDR is activated, it produces defensins and catelicidins, peptides that have antiviral activity either directly or through immunological regulation. The decreased antiviral immune response in COVID-19 patients during vitamin D shortage may be attributed to a reduction in LL37 levels, an antimicrobial peptide generated from catelicidin [205,206]. Vitamin D may also help to reduce the severity of inflammatory reactions by inhibiting pro-inflammatory cytokines such TNF- $\alpha$  and IL-6, which are involved in the development of cytokine storm in COVID-19-related ARDS [207,208].

Zinc's immunomodulatory and antiviral properties have made it and its ionophores potential COVID-19 targets [209,210]. Zinc is crucial for immune system integrity, and it plays a key role in cell maintenance, development, and activation throughout innate and adaptive immunological responses. It also helps to maintain the integrity of epithelial barriers, which are necessary for organism defense and pathogen prevention [211–213]. Zinc can regulate T cell growth and activity, thus decreasing the cytokine storm, which is accompanied by significant amounts of pro-inflammatory cytokines and chemokines that cause systemic immune response dysfunction, resulting in ARDS or multiple organ failure [214,215]. Natural killer (NK) cells and cytolytic T cells, both of which are important

in the elimination of viruses, bacteria, and tumor cells, are both affected by zinc deficiency [216,217]. Zinc's direct antiviral activity is another vital role, making it necessary for the immunological response to viral infection. Increased intracellular concentrations of this mineral can inhibit viral polyprotein processing and limit the replication of a number of RNA viruses [218–220]. Zinc can also improve interferon (IFN) cytokine signaling against RNA viruses and reduce ACE2 activity, which is required for SARS-CoV-2 entrance into host cells [221,222].

#### 8. Some Drawbacks of Antiviral Drugs on Human Body

## 8.1. Remdesivir

Researchers noted that remdesivir had certain negative impacts on the human body during a clinical study against SARS-CoV-2, such as improved productivity of liver enzymes, which might be responsible for liver dysfunction [223]. In recent times, experts in the United States documented many negative remdesivir impacts in individuals hospitalized with COVID-19. Moreover, remdesivir has been linked to an increased risk of allergic reactions, hypotension, breathing problems, as well as other human body anomalies in individuals with COVID-19, according to many study investigations. Researchers employed the first study to look at the efficacy of 5 or 10 days of remdesivir therapy for individuals with moderate bacterial meningitis. In the therapy group, vomiting, metabolic alkalosis, and migraine were more prevalent than customary [91,224].

## 8.2. Chloroquine (CQ) and Hydroxychloroquine (HCQ)

CQ can cause digestive side effects such as stomach pain, vomiting, sickness, and feces. CQ causes a variety of cardiovascular issues, including cardiac failure, lowers the blood pressure, a reduction in cardiac function, and dilatation. Finally, it has the potential to stop potassium from entering cells and has an influence on the chloride channels found in cardiac myocytes. CQ and HCQ can produce arrhythmias in COVID-19 patients, which can be decreased by mixing them with other medicinal medications, such as the antibiotic azithromycin, which has comparable cardiac actions. In other investigations, substantial rhythm problems have been linked to using CQ or HCQ, particularly in high dosages or in combination with the antibiotic azithromycin. It was also recognized that cardiac adverse effects were more common in women than in males. Heat transfer problems had the most common adverse effects, with ventricular hypertrophy, hypokinesia, and valve malfunction all being observed [225,226].

### 8.3. Lopinavir/Ritonavir

Patients who were given lopinavir/ritonavir experienced greater digestive problems as well as other comorbidities, which hampered their healing. In the lopinavir–ritonavir trial group, gastrointestinal problems were so much more prevalent in individuals with COVID-19 [227]. A clinical trial of 90 patients showed that about 14% receiving lopinavir–ritonavir were unable to conclude the 14-day regimen. Migraine, stomach discomfort, bloating, malnutrition, and anomalous stools were the most common gastrointestinal negative effects [228].

#### 8.4. Tocilizumab

In humans, tocilizumab has a number of negative health consequences, including upper respiratory infection, migraine, nasopharyngitis, site of injection response, and increased blood pressure [229].

#### 8.5. Favipiravir

Reduced locomotor activity, anemia, nausea, weight loss, enhanced demyelination in hepatocytes, and elevated blood concentrations of liver function enzymes are all side effects of favipiravir. It can potentially cause birth defects; thus, it should not be taken during pregnancy [230,231].

#### 8.6. Azithromycin

It has been confirmed that azithromycin is the cause of adverse effects, including nausea and dizziness. Nonetheless, treating COVID-19 patients with azithromycin and hydroxychloroquine has resulted in very serious complications, including the risk of mortality from abrupt heart failure [232–234].

## 9. Conclusions and Future Perspectives

COVID-19 is a life-threatening infection. It has sparked the interest of everyone on the planet, irrespective of age or educational level. The virus's effects have ceased all business, education, and travel operations, as well as disrupting people's daily lives across the world. Everyone who has been afflicted is hoping for a quick scientific reaction to put an end to the pandemic and lessen its devastating consequences. Unfortunately, developing a new drug is protracted, and the development of a new drug is improbable in a pandemic situation where an effective therapy is required right away. Natural products have been used for many years to treat viral infections and stimulate the immunological response of the host. Natural products have shown to be effective in past coronavirus illnesses, such as SARS and MERS; thus, natural products may be beneficial and provide hope during this new outbreak. In the combat against viruses, natural compounds can be used as both preventative and therapeutic agents. The following ways may be valuable in increasing and supporting research projects on COVID-19 treatment and prevention using natural products: more investigation on the use of natural chemicals as anti-coronavirus agents; quality-assurance studies for herbal extracts for use as immune-boosting medicines should be standardized; find novel targets for battling coronavirus; investigate the pharmacokinetics, pharmacodynamics, and toxicities of purified natural compounds; and expand new promiscuous drugs using SAR analysis and scientific in vivo and clinical studies.

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## References

- 1. Da Silva Antonio, A.; Wiedemann, L.S.M.; Veiga-Junior, V.F. Natural products' role against COVID-19. *RSC Adv.* 2020, 10, 23379–23393. [CrossRef]
- 2. Ali, I.; Alharbi, O.M. COVID-19: Disease, management, treatment, and social impact. *Sci. Total Environ.* **2020**, *728*, 138861. [CrossRef]
- 3. Florindo, H.F.; Kleiner, R.; Vaskovich-Koubi, D.; Acúrcio, R.C.; Carreira, B.; Yeini, E.; Tiram, G.; Liubomirski, Y.; Satchi-Fainaro, R. Immune-mediated approaches against COVID-19. *Nat. Nanotechnol.* **2020**, *15*, 630–645. [CrossRef] [PubMed]

- 4. Guessoum, S.B.; Lachal, J.; Radjack, R.; Carretier, E.; Minassian, S.; Benoit, L.; Moro, M.R. Adolescent psychiatric disorders during the COVID-19 pandemic and lockdown. *Psychiatry Res.* **2020**, *291*, 113264. [CrossRef]
- 5. Nandan, A.; Tiwari, S.; Sharma, V. Exploring alternative medicine options for the prevention or treatment of coronavirus disease 2019 (COVID-19)-A systematic scoping review. *medRxiv* 2020. [CrossRef]
- 6. World Health Organization. WHO COVID-19 Preparedness and Response Progress Report; World Health Organization: New York, NY, USA, 2020.
- Gawali Mangesh, B.; Sangle Rahul, R. Novel Corona Virus: Its Origin, Current Dignosis and Various Diseases Arises due to COVID-19. 2021. Available online: https://www.ajprd.com/index.php/journal/gateway/plugin/WebFeedGatewayPlugin/Rss (accessed on 24 September 2021).
- 8. Zhou, P.; Yang, X.-L.; Wang, X.-G.; Hu, B.; Zhang, L.; Zhang, W.; Si, H.-R.; Zhu, Y.; Li, B.; Huang, C.-L. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* **2020**, *579*, 270–273. [CrossRef] [PubMed]
- 9. Banerjee, N.; Mukhopadhyay, S. Viral glycoproteins: Biological role and application in diagnosis. *Virusdisease* **2016**, *27*, 1–11. [CrossRef]
- 10. Williamson, E.M.; Liu, X.; Izzo, A.A. Trends in use, pharmacology, and clinical applications of emerging herbal nutraceuticals. *Br. J. Pharmacol.* **2020**, 177, 1227–1240. [CrossRef] [PubMed]
- 11. Yimer, G.; Ekuadzi, E.; Fasinu, P.; de Melo, A.C.; Pillai, G. Traditional medicines for COVID-19: Perspectives from clinical pharmacologists. *Br. J. Clin. Pharmacol.* **2021**, *87*, 3455–3458. [CrossRef]
- 12. Al-Samydai, A.; Hajleh, M.; Akour, A.; Alabdallah, N.; Yousef, M.; Baqa'in, G.; Al-Saadi, A.; Al-Halaseh, L.; Aburjai, T. Phytotherapeutic approaches and ethnopharmacological responses against COVID-19. *Trop. J. Nat. Prod. Res.* **2021**, *5*, 1208–1214.
- 13. Jardim, A.C.G.; Shimizu, J.F.; Rahal, P.; Harris, M. Plant-derived antivirals against hepatitis c virus infection. *Virol. J.* **2018**, *15*, 34. [CrossRef]
- 14. Moghadamtousi, S.Z.; Nikzad, S.; Kadir, H.A.; Abubakar, S.; Zandi, K. Potential antiviral agents from marine fungi: An overview. *Mar. Drugs* 2015, *13*, 4520–4538. [CrossRef] [PubMed]
- 15. Neumann, H.; Neumann-Staubitz, P. Synthetic biology approaches in drug discovery and pharmaceutical biotechnology. *Appl. Microbiol. Biotechnol.* **2010**, *87*, 75–86. [CrossRef] [PubMed]
- Dhama, K.; Sharun, K.; Tiwari, R.; Dhawan, M.; Emran, T.B.; Rabaan, A.A.; Alhumaid, S. COVID-19 vaccine hesitancy Reasons and solutions to achieve a successful global vaccination campaign to tackle the ongoing pandemic. *Hum. Vaccin. Immunother.* 2021, 17, 3495–3499. [CrossRef]
- 17. Wang, S.-X.; Zhang, X.-S.; Guan, H.-S.; Wang, W. Potential anti-HPV and related cancer agents from marine resources: An overview. *Mar. Drugs* 2014, 12, 2019–2035. [CrossRef]
- 18. Boozari, M.; Hosseinzadeh, H. Natural products for COVID-19 prevention and treatment regarding to previous coronavirus infections and novel studies. *Phytother. Res.* **2021**, *35*, 864–876. [CrossRef]
- 19. Denaro, M.; Smeriglio, A.; Barreca, D.; De Francesco, C.; Occhiuto, C.; Milano, G.; Trombetta, D. Antiviral activity of plants and their isolated bioactive compounds: An update. *Phytother. Res.* **2020**, *34*, 742–768. [CrossRef]
- Lin, L.-T.; Hsu, W.-C.; Lin, C.-C. Antiviral natural products and herbal medicines. J. Tradit. Complement. Med. 2014, 4, 24–35. [CrossRef]
- Brufsky, A. Distinct viral clades of SARS-CoV-2: Implications for modeling of viral spread. J. Med. Virol. 2020, 92, 1386–1390. [CrossRef] [PubMed]
- 22. Wang, P.; Li, L.; Yang, H.; Cheng, S.; Zeng, Y.; Nie, L.; Zang, H. Chromatographic fingerprinting and quantitative analysis for the quality evaluation of Xinkeshu tablet. *J. Pharm. Anal.* **2012**, *2*, 422–430. [CrossRef] [PubMed]
- Yang, X.; Yu, Y.; Xu, J.; Shu, H.; Liu, H.; Wu, Y.; Zhang, L.; Yu, Z.; Fang, M.; Yu, T. Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: A single-centered, retrospective, observational study. *Lancet Respir. Med.* 2020, *8*, 475–481. [CrossRef]
- 24. Russell, C.D.; Millar, J.E.; Baillie, J.K. Clinical evidence does not support corticosteroid treatment for 2019-nCoV lung injury. *Lancet* 2020, 395, 473–475. [CrossRef]
- Tsang, H.F.; Chan, L.W.C.; Cho, W.C.S.; Yu, A.C.S.; Yim, A.K.Y.; Chan, A.K.C.; Ng, L.P.W.; Wong, Y.K.E.; Pei, X.M.; Li, M.J.W. An update on COVID-19 pandemic: The epidemiology, pathogenesis, prevention and treatment strategies. *Expert Rev. Anti-Infect. Ther.* 2021, 19, 877–888. [CrossRef]
- Mehta, P.; McAuley, D.F.; Brown, M.; Sanchez, E.; Tattersall, R.S.; Manson, J.J. COVID-19: Consider cytokine storm syndromes and immunosuppression. *Lancet* 2020, 395, 1033–1034. [CrossRef]
- 27. Mousavizadeh, L.; Ghasemi, S. Genotype and phenotype of COVID-19: Their roles in pathogenesis. *J. Microbiol. Immunol. Infect.* **2021**, *54*, 159–163. [CrossRef]
- 28. Nile, S.H.; Nile, A.; Qiu, J.; Li, L.; Jia, X.; Kai, G. COVID-19: Pathogenesis, cytokine storm and therapeutic potential of interferons. *Cytokine Growth Factor Rev.* **2020**, *53*, 66–70. [CrossRef] [PubMed]
- Rauf, A.; Abu-Izneid, T.; Olatunde, A.; Ahmed Khalil, A.; Alhumaydhi, F.A.; Tufail, T.; Shariati, M.A.; Rebezov, M.; Almarhoon, Z.M.; Mabkhot, Y.N. COVID-19 pandemic: Epidemiology, etiology, conventional and non-conventional therapies. *Int. J. Environ. Res. Public Health* 2020, *17*, 8155. [CrossRef] [PubMed]
- Bai, Y.; Yao, L.; Wei, T.; Tian, F.; Jin, D.-Y.; Chen, L.; Wang, M. Presumed asymptomatic carrier transmission of COVID-19. JAMA 2020, 323, 1406–1407. [CrossRef]

- 31. Chavez, S.; Long, B.; Koyfman, A.; Liang, S.Y. Coronavirus Disease (COVID-19): A primer for emergency physicians. *Am. J. Emerg. Med.* **2021**, *44*, 220–229. [CrossRef]
- 32. Wax, R.S.; Christian, M.D. Practical recommendations for critical care and anesthesiology teams caring for novel coronavirus (2019-nCoV) patients. *Can. J. Anesth./J. Can. D'anesthésie* 2020, *67*, 568–576. [CrossRef] [PubMed]
- 33. Zou, L.; Ruan, F.; Huang, M.; Liang, L.; Huang, H.; Hong, Z.; Yu, J.; Kang, M.; Song, Y.; Xia, J. SARS-CoV-2 viral load in upper respiratory specimens of infected patients. *N. Engl. J. Med.* **2020**, *382*, 1177–1179. [CrossRef] [PubMed]
- 34. Al Battah, A.; Hammamy, R. Multiple sclerosis flare secondary to COVID-19 vaccine, a case report. *Authorea Prepr.* 2021. [CrossRef]
- 35. Blanchard-Rohner, G.; Didierlaurent, A.; Tilmanne, A.; Smeesters, P.; Marchant, A. Pediatric COVID-19: Immunopathogenesis, Transmission and Prevention. *Vaccines* **2021**, *9*, 1002. [CrossRef] [PubMed]
- 36. Jernigan, D.B.; CDC COVID; Response Team. Update: Public health response to the coronavirus disease 2019 outbreak—United States, 24 February 2020. *Morb. Mortal. Wkly. Rep.* **2020**, *69*, 216. [CrossRef]
- Stanzione, A.; Cuocolo, R.; Del Grosso, R.; Nardiello, A.; Romeo, V.; Travaglino, A.; Raffone, A.; Bifulco, G.; Zullo, F.; Insabato, L. Deep myometrial infiltration of endometrial cancer on MRI: A radiomics-powered machine learning pilot study. *Acad. Radiol.* 2021, 28, 737–744. [CrossRef]
- Li, Q.; Guan, X.; Wu, P.; Wang, X.; Zhou, L.; Tong, Y.; Ren, R.; Leung, K.S.; Lau, E.H.; Wong, J.Y. Early transmission dynamics in Wuhan, China, of novel coronavirus–infected pneumonia. N. Engl. J. Med. 2020, 382, 1199–1207. [CrossRef] [PubMed]
- Cheng, C.; Zhang, D.; Dang, D.; Geng, J.; Zhu, P.; Yuan, M.; Liang, R.; Yang, H.; Jin, Y.; Xie, J. The incubation period of COVID-19: A global meta-analysis of 53 studies and a Chinese observation study of 11 545 patients. *Infect. Dis. Poverty* 2021, 10, 119. [CrossRef]
- 40. Zhang, M.; Xiao, J.; Deng, A.; Zhang, Y.; Zhuang, Y.; Hu, T.; Li, J.; Tu, H.; Li, B.; Zhou, Y. Transmission dynamics of an outbreak of the COVID-19 delta variant B. 1.617. 2—Guangdong province, China, May–June 2021. *China CDC Wkly.* 2021, *3*, 584. [CrossRef]
- 41. Lai, C.-C.; Shih, T.-P.; Ko, W.-C.; Tang, H.-J.; Hsueh, P.-R. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and coronavirus disease-2019 (COVID-19): The epidemic and the challenges. *Int. J. Antimicrob. Agents* 2020, 55, 105924. [CrossRef]
- Leuzinger, K.; Osthoff, M.; Dräger, S.; Pargger, H.; Siegemund, M.; Bassetti, S.; Bingisser, R.; Nickel, C.H.; Tschudin-Sutter, S.; Khanna, N. Comparing immunoassays for SARS-Coronavirus-2 antibody detection in patients with and without laboratoryconfirmed SARS-Coronavirus-2 infection. *J. Clin. Microbiol.* 2021, 59, JCM-01381. [CrossRef]
- 43. Cheng, S.-C.; Chang, Y.-C.; Chiang, Y.-L.F.; Chien, Y.-C.; Cheng, M.; Yang, C.-H.; Huang, C.-H.; Hsu, Y.-N. First case of Coronavirus Disease 2019 (COVID-19) pneumonia in Taiwan. *J. Formos. Med. Assoc.* **2020**, *119*, 747–751. [CrossRef]
- 44. Cheng, Y.; Luo, R.; Wang, K.; Zhang, M.; Wang, Z.; Dong, L.; Li, J.; Yao, Y.; Ge, S.; Xu, G. Kidney disease is associated with in-hospital death of patients with COVID-19. *Kidney Int.* **2020**, *97*, 829–838. [CrossRef]
- 45. Erdmann, F.; Wellbrock, M.; Trübenbach, C.; Spix, C.; Schrappe, M.; Schüz, J.; Grabow, D.; Eichinger, M. Impact of the COVID-19 pandemic on incidence, time of diagnosis and delivery of healthcare among paediatric oncology patients in Germany in 2020: Evidence from the German Childhood Cancer Registry and a qualitative survey. *Lancet Reg. Health-Eur.* **2021**, *9*, 100188. [CrossRef]
- 46. Kanne, J.P. Chest CT Findings in 2019 Novel Coronavirus (2019-nCoV) Infections from Wuhan, China: Key Points for the Radiologist; Radiological Society of North America: Oak Brook, IL, USA, 2020.
- 47. Wu, J.; Pan, J.; Teng, D.; Xu, X.; Feng, J.; Chen, Y.-C. Interpretation of CT signs of 2019 novel coronavirus (COVID-19) pneumonia. *Eur. Radiol.* 2020, *30*, 5455–5462. [CrossRef]
- 48. Kalia, M.; Chandra, V.; Rahman, S.A.; Sehgal, D.; Jameel, S. Heparan sulfate proteoglycans are required for cellular binding of the hepatitis E virus ORF2 capsid protein and for viral infection. *J. Virol.* **2009**, *83*, 12714–12724. [CrossRef] [PubMed]
- Li, W.; Hulswit, R.J.; Widjaja, I.; Raj, V.S.; McBride, R.; Peng, W.; Widagdo, W.; Tortorici, M.A.; Van Dieren, B.; Lang, Y. Identification of sialic acid-binding function for the Middle East respiratory syndrome coronavirus spike glycoprotein. *Proc. Natl. Acad. Sci. USA* 2017, *114*, E8508–E8517. [CrossRef] [PubMed]
- 50. Sharun, K.; Tiwari, R.; Dhama, K.; Emran, T.B.; Rabaan, A.A.; Al Mutair, A. Emerging SARS-CoV-2 variants: Impact on vaccine efficacy and neutralizing antibodies. *Hum. Vaccin. Immunother.* **2021**, *17*, 3491–3494. [CrossRef] [PubMed]
- 51. Gomes, P.B.; Dietrich, C.P. Distribution of heparin and other sulfated glycosaminoglycans in vertebrates. *Comp. Biochem. Physiol. B Comp. Biochem.* **1982**, *73*, 857–863. [CrossRef]
- 52. Toledo, O.M.; Dietrich, C.P. Tissue specific distribution of sulfated mucopolysaccharides in mammals. *Biochim. Biophys. Acta Gen. Subj.* **1977**, *498*, 114–122. [CrossRef]
- 53. Hao, W.; Ma, B.; Li, Z.; Wang, X.; Gao, X.; Li, Y.; Qin, B.; Shang, S.; Cui, S.; Tan, Z. Binding of the SARS-CoV-2 spike protein to glycans. *Sci. Bull.* **2021**, *66*, 1205–1214. [CrossRef]
- Rabaan, A.A.; Al-Ahmed, S.H.; Muhammad, J.; Khan, A.; Sule, A.A.; Tirupathi, R.; Mutair, A.A.; Alhumaid, S.; Al-Omari, A.; Dhawan, M.; et al. Role of Inflammatory Cytokines in COVID-19 Patients: A Review on Molecular Mechanisms, Immune Functions, Immunopathology and Immunomodulatory Drugs to Counter Cytokine Storm. *Vaccine* 2021, 9, 436. [CrossRef]
- 55. Nader, H.B.; Chavante, S.F.; Dos-Santos, E.A.; Oliveira, F.W.; De-Paiva, J.F.; Jerônimo, S.M.B.; Medeiros, G.F.D.; De-Abreu, L.R.D.; Leite, E.L.; de-Sousa-Filho, J.F.; et al. Heparan sulfates and heparins: Similar compounds performing the same functions in vertebrates and invertebrates? *Braz. J. Med. Biol. Res.* 1999, 32, 529–538. [CrossRef] [PubMed]
- 56. Hilgard, P.; Stockert, R. Heparan sulfate proteoglycans initiate dengue virus infection of hepatocytes. *Hepatology* **2000**, *32*, 1069–1077. [CrossRef] [PubMed]

- 57. Rabaan, A.A.; Tirupathi, R.; Sule, A.A.; Aldali, J.; Al Mutair, A.; Alhumaid, S.; Muzaheed; Gupta, N.; Koritala, T.; Adhikari, R.; et al. Viral dynamics and real-time RT-PCR Ct values correlation with disease severity in COVID-19. *Diagnostics* **2021**, *11*, 1091. [CrossRef]
- 58. Tamhankar, M.; Gerhardt, D.M.; Bennett, R.S.; Murphy, N.; Jahrling, P.B.; Patterson, J.L. Heparan sulfate is an important mediator of Ebola virus infection in polarized epithelial cells. *Virol. J.* **2018**, *15*, 135. [CrossRef]
- 59. Bojkova, D.; McGreig, J.E.; McLaughlin, K.-M.; Masterson, S.G.; Widera, M.; Krähling, V.; Ciesek, S.; Wass, M.N.; Michaelis, M.; Cinatl, J. SARS-CoV-2 and SARS-CoV differ in their cell tropism and drug sensitivity profiles. *bioRxiv* 2020. [CrossRef]
- Guo, Y.-R.; Cao, Q.-D.; Hong, Z.-S.; Tan, Y.-Y.; Chen, S.-D.; Jin, H.-J.; Tan, K.-S.; Wang, D.-Y.; Yan, Y. The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak–an update on the status. *Mil. Med. Res.* 2020, 7, 11. [CrossRef] [PubMed]
- Hoffmann, M.; Kleine-Weber, H.; Schroeder, S.; Krüger, N.; Herrler, T.; Erichsen, S.; Schiergens, T.S.; Herrler, G.; Wu, N.-H.; Nitsche, A. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell* 2020, 181, 271–280.e8. [CrossRef]
- 62. Mycroft-West, C.; Su, D.; Elli, S.; Li, Y.; Guimond, S.; Miller, G.; Turnbull, J.; Yates, E.; Guerrini, M.; Fernig, D. The 2019 coronavirus (SARS-CoV-2) surface protein (Spike) S1 Receptor Binding Domain undergoes conformational change upon heparin binding. *bioRxiv* 2020. [CrossRef]
- 63. Wickramasinghe, I.A.; De Vries, R.; Gröne, A.; De Haan, C.; Verheije, M. Binding of avian coronavirus spike proteins to host factors reflects virus tropism and pathogenicity. *J. Virol.* **2011**, *85*, 8903–8912. [CrossRef] [PubMed]
- 64. Singh, B.; Datta, B.; Ashish, A.; Dutta, G. A comprehensive review on current COVID-19 detection methods: From Lab care to Point of care diagnosis. *Sens. Int.* **2021**, *2*, 100119. [CrossRef] [PubMed]
- 65. Schoeman, D.; Fielding, B.C. Coronavirus envelope protein: Current knowledge. Virol. J. 2019, 16, 69. [CrossRef]
- 66. Harapan, H.; Ryan, M.; Yohan, B.; Abidin, R.S.; Nainu, F.; Rakib, A.; Jahan, I.; Emran, T.B.; Ullah, I.; Panta, K.; et al. COVID-19 and dengue: Double punches for dengue-endemic countries in Asia. *Rev. Med. Virol.* **2021**, *31*, e2161. [CrossRef]
- 67. Walls, A.C.; Park, Y.-J.; Tortorici, M.A.; Wall, A.; McGuire, A.T.; Veesler, D. Structure, function, and antigenicity of the SARS-CoV-2 spike glycoprotein. *Cell* **2020**, *181*, 281–292.e6. [CrossRef] [PubMed]
- 68. Zumla, A.; Chan, J.F.; Azhar, E.I.; Hui, D.S.; Yuen, K.-Y. Coronaviruses—Drug discovery and therapeutic options. *Nat. Rev. Drug Discov.* **2016**, *15*, 327–347. [CrossRef]
- 69. Phan, T. Novel coronavirus: From discovery to clinical diagnostics. Infect. Genet. Evol. 2020, 79, 104211. [CrossRef] [PubMed]
- 70. Song, X.; Hu, W.; Yu, H.; Zhao, L.; Zhao, Y.; Zhao, Y. High expression of angiotensin-converting enzyme-2 (ACE2) on tissue macrophages that may be targeted by virus SARS-CoV-2 in COVID-19 patients. *bioRxiv* 2020. [CrossRef]
- Tai, W.; He, L.; Zhang, X.; Pu, J.; Voronin, D.; Jiang, S.; Zhou, Y.; Du, L. Characterization of the receptor-binding domain (RBD) of 2019 novel coronavirus: Implication for development of RBD protein as a viral attachment inhibitor and vaccine. *Cell. Mol. Immunol.* 2020, *17*, 613–620. [CrossRef]
- Yin, Y.; Wunderink, R.G. MERS, SARS and other coronaviruses as causes of pneumonia. *Respirology* 2018, 23, 130–137. [CrossRef]
  [PubMed]
- Zhang, L.; Lin, D.; Sun, X.; Curth, U.; Drosten, C.; Sauerhering, L.; Becker, S.; Rox, K.; Hilgenfeld, R. Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved α-ketoamide inhibitors. *Science* 2020, 368, 409–412. [CrossRef]
- 74. Huang, C.; Wang, Y.; Li, X.; Ren, L.; Zhao, J.; Hu, Y.; Zhang, L.; Fan, G.; Xu, J.; Gu, X. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020, *395*, 497–506. [CrossRef]
- 75. Vellingiri, B.; Jayaramayya, K.; Iyer, M.; Narayanasamy, A.; Govindasamy, V.; Giridharan, B.; Ganesan, S.; Venugopal, A.; Venkatesan, D.; Ganesan, H. COVID-19: A promising cure for the global panic. *Sci. Total Environ.* **2020**, *725*, 138277. [CrossRef]
- Lei, J.; Kusov, Y.; Hilgenfeld, R. Nsp3 of coronaviruses: Structures and functions of a large multi-domain protein. *Antivir. Res.* 2018, 149, 58–74. [CrossRef] [PubMed]
- 77. Coutard, B.; Valle, C.; de Lamballerie, X.; Canard, B.; Seidah, N.; Decroly, E. The spike glycoprotein of the new coronavirus 2019-nCoV contains a furin-like cleavage site absent in CoV of the same clade. *Antivir. Res.* 2020, 176, 104742. [CrossRef] [PubMed]
- 78. Li, Y.C.; Bai, W.Z.; Hashikawa, T. The neuroinvasive potential of SARS-CoV2 may play a role in the respiratory failure of COVID-19 patients. *J. Med. Virol.* **2020**, *92*, 552–555. [CrossRef] [PubMed]
- Siu, K.L.; Yuen, K.S.; Castano-Rodriguez, C.; Ye, Z.W.; Yeung, M.L.; Fung, S.Y.; Yuan, S.; Chan, C.P.; Yuen, K.Y.; Enjuanes, L. Severe acute respiratory syndrome Coronavirus ORF3a protein activates the NLRP3 inflammasome by promoting TRAF3-dependent ubiquitination of ASC. FASEB J. 2019, 33, 8865–8877. [CrossRef] [PubMed]
- 80. Shi, C.-S.; Nabar, N.R.; Huang, N.-N.; Kehrl, J.H. SARS-Coronavirus Open Reading Frame-8b triggers intracellular stress pathways and activates NLRP3 inflammasomes. *Cell Death Discov.* **2019**, *5*, 101. [CrossRef]
- Rabaan, A.A.; Al-Ahmed, S.H.; Garout, M.A.; Al-Qaaneh, A.M.; Sule, A.A.; Tirupathi, R.; Mutair, A.A.; Alhumaid, S.; Al-Omari, A.; Hasan, A.; et al. Diverse Immunological Factors Influencing Pathogenesis in Patients with COVID-19: A Review on Viral Dissemination, Immunotherapeutic Options to Counter Cytokine Storm and Inflammatory Responses. *Pathogens* 2021, 10, 565. [CrossRef]

- 82. Agostini, M.L.; Andres, E.L.; Sims, A.C.; Graham, R.L.; Sheahan, T.P.; Lu, X.; Smith, E.C.; Case, J.B.; Feng, J.Y.; Jordan, R. Coronavirus susceptibility to the antiviral remdesivir (GS-5734) is mediated by the viral polymerase and the proofreading exoribonuclease. *mBio* **2018**, *9*, e00221-18. [CrossRef]
- 83. De Wit, E.; Van Doremalen, N.; Falzarano, D.; Munster, V.J. SARS and MERS: Recent insights into emerging coronaviruses. *Nat. Rev. Microbiol.* **2016**, *14*, 523–534. [CrossRef] [PubMed]
- Lo, M.K.; Jordan, R.; Arvey, A.; Sudhamsu, J.; Shrivastava-Ranjan, P.; Hotard, A.L.; Flint, M.; McMullan, L.K.; Siegel, D.; Clarke, M.O. GS-5734 and its parent nucleoside analog inhibit Filo-, Pneumo-, and Paramyxoviruses. *Sci. Rep.* 2017, *7*, 43395. [CrossRef]
- 85. Sheahan, T.P.; Sims, A.C.; Graham, R.L.; Menachery, V.D.; Gralinski, L.E.; Case, J.B.; Leist, S.R.; Pyrc, K.; Feng, J.Y.; Trantcheva, I. Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses. *Sci. Transl. Med.* **2017**, *9*, 396. [CrossRef]
- Gordon, C.J.; Tchesnokov, E.P.; Feng, J.Y.; Porter, D.P.; Götte, M. The antiviral compound remdesivir potently inhibits RNAdependent RNA polymerase from Middle East respiratory syndrome coronavirus. J. Biol. Chem. 2020, 295, 4773–4779. [CrossRef]
- 87. Tchesnokov, E.P.; Feng, J.Y.; Porter, D.P.; Götte, M. Mechanism of inhibition of Ebola virus RNA-dependent RNA polymerase by remdesivir. *Viruses* **2019**, *11*, 326. [CrossRef]
- 88. Wang, M.; Cao, R.; Zhang, L.; Yang, X.; Liu, J.; Xu, M.; Shi, Z.; Hu, Z.; Zhong, W.; Xiao, G. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res.* **2020**, *30*, 269–271. [CrossRef] [PubMed]
- De Wit, E.; Feldmann, F.; Cronin, J.; Jordan, R.; Okumura, A.; Thomas, T.; Scott, D.; Cihlar, T.; Feldmann, H. Prophylactic and therapeutic remdesivir (GS-5734) treatment in the rhesus macaque model of MERS-CoV infection. *Proc. Natl. Acad. Sci. USA* 2020, 117, 6771–6776. [CrossRef] [PubMed]
- Mulangu, S.; Dodd, L.E.; Davey, R.T., Jr.; Tshiani Mbaya, O.; Proschan, M.; Mukadi, D.; Lusakibanza Manzo, M.; Nzolo, D.; Tshomba Oloma, A.; Ibanda, A. A randomized, controlled trial of Ebola virus disease therapeutics. *N. Engl. J. Med.* 2019, 381, 2293–2303. [CrossRef] [PubMed]
- Wang, W.; Zhang, D.; Du, R. Original: Remdesivir in adults with severe COVID-19: A randomised, double-blind, placebo. *Lancet* 2020, 395, 1569–1578. [CrossRef]
- Cohen, M.S. Hydroxychloroquine for the prevention of COVID-19—Searching for evidence. N. Engl. J. Med. 2020, 383, 585–586. [CrossRef] [PubMed]
- 93. Tilangi, P.; Desai, D.; Khan, A.; Soneja, M. Hydroxychloroquine prophylaxis for high-risk COVID-19 contacts in India: A prudent approach. *Lancet Infect. Dis.* 2020, 20, 1119–1120. [CrossRef]
- 94. Beigel, J.H.; Tomashek, K.M.; Dodd, L.E.; Mehta, A.K.; Zingman, B.S.; Kalil, A.C.; Hohmann, E.; Chu, H.Y.; Luetkemeyer, A.; Kline, S. Remdesivir for the treatment of COVID-19. *N. Engl. J. Med.* **2020**, *383*, 1813–1826. [CrossRef]
- 95. Nainu, F.; Abidin, R.S.; Bahar, M.A.; Frediansyah, A.; Emran, T.B.; Rabaan, A.A.; Dhama, K.; Harapnan, H. SARS-CoV-2 reinfection and implications for vaccine development. *Hum. Vaccin. Immunother.* **2020**, *16*, 3061–3073. [CrossRef] [PubMed]
- Sham, H.L.; Kempf, D.J.; Molla, A.; Marsh, K.C.; Kumar, G.N.; Chen, C.-M.; Kati, W.; Stewart, K.; Lal, R.; Hsu, A. ABT-378, a highly potent inhibitor of the human immunodeficiency virus protease. *Antimicrob. Agents Chemother.* 1998, 42, 3218–3224. [CrossRef] [PubMed]
- 97. Benson, C.A.; Deeks, S.G.; Brun, S.C.; Gulick, R.M.; Eron, J.J.; Kessler, H.A.; Murphy, R.L.; Hicks, C.; King, M.; Wheeler, D. Safety and antiviral activity at 48 weeks of lopinavir/ritonavir plus nevirapine and 2 nucleoside reverse-transcriptase inhibitors in human immunodeficiency virus type 1-infected protease inhibitor-experienced patients. *J. Infect. Dis.* 2002, 185, 599–607. [CrossRef]
- 98. Corbett, A.H.; Lim, M.L.; Kashuba, A.D. Kaletra (lopinavir/ritonavir). Ann. Pharmacother. 2002, 36, 1193–1203. [CrossRef]
- 99. Ortiz-Martínez, Y. Tocilizumab: A new opportunity in the possible therapeutic arsenal against COVID-19. *Travel Med. Infect. Dis.* **2020**, *37*, 101678. [CrossRef] [PubMed]
- 100. Cellina, M.; Orsi, M.; Bombaci, F.; Sala, M.; Marino, P.; Oliva, G. Favorable changes of CT findings in a patient with COVID-19 pneumonia after treatment with tocilizumab. *Diagn. Interv. Imaging* **2020**, *101*, 323. [CrossRef]
- 101. Chi, Z.; Zhao, W.; Jia-Wen, L.; Hong, Z.; Gui-Qiang, W. The Cytokine Release Syndrome (CRS) of Severe COVID-19 and Interleukin-6 Receptor (IL-6R) Antagonist Tocilizumab Man Be the Key to Reduce the Mortality. 2020. Available online: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7118634/pdf/main.pdf (accessed on 15 August 2021).
- Zhao, M. Cytokine storm and immunomodulatory therapy in COVID-19: Role of chloroquine and anti-IL-6 monoclonal antibodies. *Int. J. Antimicrob. Agents* 2020, 55, 105982. [CrossRef] [PubMed]
- 103. Furuta, Y.; Komeno, T.; Nakamura, T. Favipiravir (T-705), a broad spectrum inhibitor of viral RNA polymerase. *Proc. Jpn. Acad. Ser. B* 2017, 93, 449–463. [CrossRef] [PubMed]
- 104. Furuta, Y.; Takahashi, K.; Fukuda, Y.; Kuno, M.; Kamiyama, T.; Kozaki, K.; Nomura, N.; Egawa, H.; Minami, S.; Watanabe, Y. In vitro and in vivo activities of anti-influenza virus compound T-705. *Antimicrob. Agents Chemother.* 2002, 46, 977–981. [CrossRef] [PubMed]
- 105. Jin, Z.; Smith, L.K.; Rajwanshi, V.K.; Kim, B.; Deval, J. The ambiguous base-pairing and high substrate efficiency of T-705 (favipiravir) ribofuranosyl 5'-triphosphate towards influenza A virus polymerase. PLoS ONE 2013, 8, e68347. [CrossRef] [PubMed]
- 106. Blaising, J.; Polyak, S.J.; Pécheur, E.-I. Arbidol as a broad-spectrum antiviral: An update. Antivir. Res. 2014, 107, 84–94. [CrossRef] [PubMed]

- 107. Tu, Y.-F.; Chien, C.-S.; Yarmishyn, A.A.; Lin, Y.-Y.; Luo, Y.-H.; Lin, Y.-T.; Lai, W.-Y.; Yang, D.-M.; Chou, S.-J.; Yang, Y.-P. A review of SARS-CoV-2 and the ongoing clinical trials. *Int. J. Mol. Sci.* **2020**, *21*, 2657. [CrossRef] [PubMed]
- 108. Blaising, J.; Lévy, P.L.; Polyak, S.J.; Stanifer, M.; Boulant, S.; Pécheur, E.-I. Arbidol inhibits viral entry by interfering with clathrin-dependent trafficking. *Antivir. Res.* 2013, 100, 215–219. [CrossRef]
- 109. Rakib, A.; Nain, Z.; Islam, M.A.; Sami, S.A.; Mahmud, S.; Islam, A.; Ahmed, S.; Siddiqui, A.B.F.; Babu, S.M.O.F.; Hossain, P.; et al. A molecular modelling approach for identifying antiviral selenium-containing heterocyclic compounds that inhibit the main protease of SARS-CoV-2: An in silico investigation. *Brief. Bioinform.* 2021, 22, 1476–1498. [CrossRef]
- 110. Leneva, I.A.; Russell, R.J.; Boriskin, Y.S.; Hay, A.J. Characteristics of arbidol-resistant mutants of influenza virus: Implications for the mechanism of anti-influenza action of arbidol. *Antivir. Res.* 2009, *81*, 132–140. [CrossRef] [PubMed]
- Seo, J.-W.; Kim, D.; Yun, N.; Kim, D.-M. Clinical Update of Severe Fever with Thrombocytopenia Syndrome. *Viruses* 2021, 13, 1213. [CrossRef]
- 112. Cortegiani, A.; Ingoglia, G.; Ippolito, M.; Giarratano, A.; Einav, S. A systematic review on the efficacy and safety of chloroquine for the treatment of COVID-19. *J. Crit. Care* 2020, *57*, 279–283. [CrossRef]
- 113. Sanders, J.M.; Monogue, M.L.; Jodlowski, T.Z.; Cutrell, J.B. Pharmacologic treatments for coronavirus disease 2019 (COVID-19): A review. *JAMA* 2020, 323, 1824–1836. [CrossRef]
- 114. Sheahan, T.P.; Sims, A.C.; Leist, S.R.; Schäfer, A.; Won, J.; Brown, A.J.; Montgomery, S.A.; Hogg, A.; Babusis, D.; Clarke, M.O. Comparative therapeutic efficacy of remdesivir and combination lopinavir, ritonavir, and interferon beta against MERS-CoV. *Nat. Commun.* 2020, 11, 222. [CrossRef]
- 115. Dong, L.; Hu, S.; Gao, J. Discovering drugs to treat coronavirus disease 2019 (COVID-19). *Drug Discov. Ther.* 2020, 14, 58–60. [CrossRef]
- 116. Sanchis-Gomar, F.; Lavie, C.J.; Perez-Quilis, C.; Henry, B.M.; Lippi, G. Angiotensin-converting enzyme 2 and antihypertensives (angiotensin receptor blockers and angiotensin-converting enzyme inhibitors) in coronavirus disease 2019. *Mayo Clin. Proc.* 2020, 95, 1222–1230. [CrossRef] [PubMed]
- 117. Chakravarti, R.; Singh, R.; Ghosh, A.; Dey, D.; Sharma, P.; Velayutham, R.; Roy, S.; Ghosh, D. A review on potential of natural products in the management of COVID-19. *RSC Adv.* **2021**, *11*, 16711–16735. [CrossRef]
- 118. Wu, M.-Y.; Wang, S.-F.; Cai, C.-Z.; Tan, J.-Q.; Li, M.; Lu, J.-J.; Chen, X.-P.; Wang, Y.-T.; Zheng, W.; Lu, J.-H. Natural autophagy blockers, dauricine (DAC) and daurisoline (DAS), sensitize cancer cells to camptothecin-induced toxicity. *Oncotarget* 2017, *8*, 77673. [CrossRef] [PubMed]
- 119. Xue, L.; Liu, P. Daurisoline inhibits hepatocellular carcinoma progression by restraining autophagy and promoting cispaltininduced cell death. *Biochem. Biophys. Res. Commun.* 2021, 534, 1083–1090. [CrossRef] [PubMed]
- 120. Sathasivam, R.; Radhakrishnan, R.; Hashem, A.; Abd\_Allah, E.F. Microalgae metabolites: A rich source for food and medicine. *Saudi J. Biol. Sci.* **2019**, *26*, 709–722. [CrossRef]
- 121. Qiu, W.; Su, M.; Xie, F.; Ai, J.; Ren, Y.; Zhang, J.; Guan, R.; He, W.; Gong, Y.; Guo, Y. Tetrandrine blocks autophagic flux and induces apoptosis via energetic impairment in cancer cells. *Cell Death Dis.* **2014**, *5*, e1123. [CrossRef] [PubMed]
- 122. Wang, S.; Fu, J.-L.; Hao, H.-F.; Jiao, Y.-N.; Li, P.-P.; Han, S.-Y. Metabolic reprogramming by traditional Chinese medicine and its role in effective cancer therapy. *Pharmacol. Res.* **2021**, *170*, 105728. [CrossRef]
- 123. Nabavi, S.F.; Habtemariam, S.; Berindan-Neagoe, I.; Cismaru, C.A.; Schaafsma, D.; Ghavami, S.; Banach, M.; Aghaabdollahian, S.; Nabavi, S.M. Rationale for Effective Prophylaxis Against COVID-19 Through Simultaneous Blockade of Both Endosomal and Non-Endosomal SARS-CoV-2 Entry into Host Cell. *Clin. Transl. Sci.* **2021**, *14*, 431. [CrossRef]
- 124. Yi, L.; Li, Z.; Yuan, K.; Qu, X.; Chen, J.; Wang, G.; Zhang, H.; Luo, H.; Zhu, L.; Jiang, P. Small molecules blocking the entry of severe acute respiratory syndrome coronavirus into host cells. *J. Virol.* **2004**, *78*, 11334–11339. [CrossRef]
- 125. Kreutzberger, A.J.; Sanyal, A.; Ojha, R.; Pyle, J.D.; Vapalahti, O.; Balistreri, G.; Kirchhausen, T. Synergistic block of SARS-CoV-2 infection by combined drug inhibition of the host entry factors PIK fyve kinase and TMPRSS2 protease. J. Virol. 2021, 95, e00975-21. [CrossRef] [PubMed]
- 126. Park, J.-Y.; Yuk, H.J.; Ryu, H.W.; Lim, S.H.; Kim, K.S.; Park, K.H.; Ryu, Y.B.; Lee, W.S. Evaluation of polyphenols from Broussonetia papyrifera as coronavirus protease inhibitors. *J. Enzym. Inhib. Med. Chem.* **2017**, *32*, 504–512. [CrossRef] [PubMed]
- 127. Rabaan, A.A.; Al Mutair, A.; Alhumaid, S.; Al Alawi, Z.; Al Mohaini, M.; Alsalman, A.J.; Fawzy, M.; Al-Tawfiq, J.A.; Almahmoud, S.; Alfouzan, W.; et al. Modulation of host epigenome by coronavirus infections and developing treatment modalities for COVID-19 beyond genetics. *Eur. Rev. Med. Pharmacol. Sci.* 2021, 25, 5947–5964. [CrossRef]
- 128. Zhao, Y.; Niu, J.; Zhou, Q.; Chen, Y.; Gan, S.; Shen, X.; Zhang, N. Flavonoids Isolated from the genus Ficus and Their Biological Activities. *Med. Res.* 2021, *5*, 210004. [CrossRef]
- 129. Demeke, C.A.; Woldeyohanins, A.E.; Kifle, Z.D. Herbal medicine use for the management of COVID-19: A review article. *Metab. Open* **2021**, *12*, 100141. [CrossRef]
- Cho, J.K.; Curtis-Long, M.J.; Lee, K.H.; Kim, D.W.; Ryu, H.W.; Yuk, H.J.; Park, K.H. Geranylated flavonoids displaying SARS-CoV papain-like protease inhibition from the fruits of Paulownia tomentosa. *Bioorgan. Med. Chem.* 2013, 21, 3051–3057. [CrossRef]
- 131. Clergeaud, G.; Dabbagh-Bazarbachi, H.; Ortiz, M.; Fernández-Larrea, J.B.; O'Sullivan, C.K. A simple liposome assay for the screening of zinc ionophore activity of polyphenols. *Food Chem.* **2016**, *197*, 916–923. [CrossRef]
- 132. Liu, B.; Yang, J.; Ma, Y.; Yuan, E.; Chen, C. Antioxidant and angiotensin converting enzyme (ACE) inhibitory activities of ethanol extract and pure flavonoids from Adinandra nitida leaves. *Pharm. Biol.* **2010**, *48*, 1432–1438. [CrossRef]

- 133. Nguyen, H.Q.; Nguyen, T.N.L.; Doan, T.N.; Nguyen, T.T.N.; Phạm, M.H.; Le, T.L.; Sy, D.T.; Chu, H.H.; Chu, H.M. Complete chloroplast genome of novel Adrinandra megaphylla Hu species: Molecular structure, comparative and phylogenetic analysis. *Sci. Rep.* **2021**, *11*, 11731. [CrossRef]
- 134. Sun, S.; Huang, S.; Shi, Y.; Shao, Y.; Qiu, J.; Sedjoah, R.-C.A.-A.; Yan, Z.; Ding, L.; Zou, D.; Xin, Z. Extraction, isolation, characterization and antimicrobial activities of non-extractable polyphenols from pomegranate peel. *Food Chem.* 2021, 351, 129232. [CrossRef]
- 135. Martins, B.X.; Arruda, R.F.; Costa, G.A.; Jerdy, H.; de Souza, S.B.; Santos, J.M.; de Freitas, W.R.; Kanashiro, M.M.; de Carvalho, E.C.Q.; Sant'Anna, N.F. Myrtenal-induced V-ATPase inhibition-A toxicity mechanism behind tumor cell death and suppressed migration and invasion in melanoma. *Biochim. Biophys. Acta Gen. Subj.* 2019, 1863, 1–12. [CrossRef] [PubMed]
- Wen, C.-C.; Kuo, Y.-H.; Jan, J.-T.; Liang, P.-H.; Wang, S.-Y.; Liu, H.-G.; Lee, C.-K.; Chang, S.-T.; Kuo, C.-J.; Lee, S.-S. Specific plant terpenoids and lignoids possess potent antiviral activities against severe acute respiratory syndrome coronavirus. *J. Med. Chem.* 2007, 50, 4087–4095. [CrossRef]
- Malekmohammad, K.; Rafieian-Kopaei, M. Mechanistic Aspects of Medicinal Plants and Secondary Metabolites against Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). Curr. Pharm. Des. 2021, 27, 3996–4007. [CrossRef]
- 138. Panigrahi, G.K.; Sahoo, S.K.; Sahoo, A.; Behera, S.; Sahu, S.; Dash, A.; Satapathy, K.B. Bioactive molecules from plants: A prospective approach to combat SARS-CoV-2. *Adv. Tradit. Med.* **2021**, 1–14. [CrossRef]
- 139. Koh, Y.-C.; Ho, C.-T.; Pan, M.-H. Recent Advances in Health Benefits of Stilbenoids. J. Agric. Food Chem. 2021, 69, 10036–10057. [CrossRef]
- 140. Xing, Y.; Liao, J.; Tang, Y.; Zhang, P.; Tan, C.; Ni, H.; Wu, X.; Li, N.; Jia, X. ACE and platelet aggregation inhibitors from Tamarix hohenackeri Bunge (host plant of Herba Cistanches) growing in Xinjiang. *Pharmacogn. Mag.* **2014**, *10*, 111. [PubMed]
- Al Shukor, N.; Van Camp, J.; Gonzales, G.B.; Staljanssens, D.; Struijs, K.; Zotti, M.J.; Raes, K.; Smagghe, G. Angiotensin-converting enzyme inhibitory effects by plant phenolic compounds: A study of structure activity relationships. *J. Agric. Food Chem.* 2013, *61*, 11832–11839. [CrossRef]
- Ali, M.Y.; Seong, S.H.; Jung, H.A.; Choi, J.S. Angiotensin-I-converting enzyme inhibitory activity of coumarins from Angelica decursiva. *Molecules* 2019, 24, 3937. [CrossRef] [PubMed]
- 143. Lin, Y.-S.; Lu, Y.-L.; Wang, G.-J.; Chen, L.-G.; Wen, C.-L.; Hou, W.-C. Ethanolic extracts and isolated compounds from small-leaf grape (Vitis thunbergii var. taiwaniana) with antihypertensive activities. J. Agric. Food Chem. 2012, 60, 7435–7441. [CrossRef]
- 144. Cao, J.; Forrest, J.C.; Zhang, X. A screen of the NIH Clinical Collection small molecule library identifies potential anti-coronavirus drugs. *Antivir. Res.* 2015, 114, 1–10. [CrossRef] [PubMed]
- 145. Mahmud, S.; Uddin, M.A.R.; Paul, G.K.; Shimu, M.S.S.; Islam, S.; Rahman, E.; Islam, A.; Islam, M.S.; Promi, M.M.; Emran, T.B.; et al. Virtual screening and molecular dynamics simulation study of plant derived compounds to identify potential inhibitor of main protease from SARS-CoV-2. *Brief. Bioinform.* 2021, 22, 1402–1414. [CrossRef] [PubMed]
- 146. Khan, F.A.; Maalik, A. Advances in pharmacology of isatin and its derivatives: A review. *Trop. J. Pharm. Res.* **2015**, *14*, 1937–1942. [CrossRef]
- 147. Liu, W.; Zhu, H.-M.; Niu, G.-J.; Shi, E.-Z.; Chen, J.; Sun, B.; Chen, W.-Q.; Zhou, H.-G.; Yang, C. Synthesis, modification and docking studies of 5-sulfonyl isatin derivatives as SARS-CoV 3C-like protease inhibitors. *Bioorgan. Med. Chem.* 2014, 22, 292–302. [CrossRef]
- 148. Yang, C.-W.; Lee, Y.-Z.; Hsu, H.-Y.; Shih, C.; Chao, Y.-S.; Chang, H.-Y.; Lee, S.-J. Targeting coronaviral replication and cellular JAK2 mediated dominant NF-κB activation for comprehensive and ultimate inhibition of coronaviral activity. *Sci. Rep.* 2017, 7, 4105. [CrossRef] [PubMed]
- 149. Yang, C.-W.; Lee, Y.-Z.; Kang, I.-J.; Barnard, D.L.; Jan, J.-T.; Lin, D.; Huang, C.-W.; Yeh, T.-K.; Chao, Y.-S.; Lee, S.-J. Identification of phenanthroindolizines and phenanthroquinolizidines as novel potent anti-coronaviral agents for porcine enteropathogenic coronavirus transmissible gastroenteritis virus and human severe acute respiratory syndrome coronavirus. *Antivir. Res.* 2010, 88, 160–168. [CrossRef]
- 150. Li, S.-y.; Chen, C.; Zhang, H.-q.; Guo, H.-y.; Wang, H.; Wang, L.; Zhang, X.; Hua, S.-n.; Yu, J.; Xiao, P.-g. Identification of natural compounds with antiviral activities against SARS-associated coronavirus. *Antivir. Res.* 2005, *67*, 18–23. [CrossRef] [PubMed]
- 151. Shen, L.; Niu, J.; Wang, C.; Huang, B.; Wang, W.; Zhu, N.; Deng, Y.; Wang, H.; Ye, F.; Cen, S. High-throughput screening and identification of potent broad-spectrum inhibitors of coronaviruses. *J. Virol.* **2019**, *93*, e00023-19. [CrossRef]
- 152. Weber, C.; Opatz, T. Bisbenzylisoquinoline Alkaloids. Alkaloids Chem. Biol. 2019, 81, 1–114. [CrossRef]
- 153. Kim, D.W.; Seo, K.H.; Curtis-Long, M.J.; Oh, K.Y.; Oh, J.-W.; Cho, J.K.; Lee, K.H.; Park, K.H. Phenolic phytochemical displaying SARS-CoV papain-like protease inhibition from the seeds of Psoralea corylifolia. *J. Enzym. Inhib. Med. Chem.* **2014**, 29, 59–63. [CrossRef]
- Runfeng, L.; Yunlong, H.; Jicheng, H.; Weiqi, P.; Qinhai, M.; Yongxia, S.; Chufang, L.; Jin, Z.; Zhenhua, J.; Haiming, J. Lianhuaqingwen exerts anti-viral and anti-inflammatory activity against novel coronavirus (SARS-CoV-2). *Pharmacol. Res.* 2020, 156, 104761. [CrossRef]
- 155. Chen, F.; Chan, K.; Jiang, Y.; Kao, R.; Lu, H.; Fan, K.; Cheng, V.; Tsui, W.; Hung, I.; Lee, T. In vitro susceptibility of 10 clinical isolates of SARS coronavirus to selected antiviral compounds. *J. Clin. Virol.* **2004**, *31*, 69–75. [CrossRef] [PubMed]

- 156. Song, J.; Zhang, L.; Xu, Y.; Yang, D.; Yang, S.; Zhang, W.; Wang, J.; Tian, S.; Yang, S.; Yuan, T. The comprehensive study on the therapeutic effects of baicalein for the treatment of COVID-19 in vivo and in vitro. *Biochem. Pharmacol.* **2021**, *183*, 114302. [CrossRef]
- 157. Park, J.-Y.; Ko, J.-A.; Kim, D.W.; Kim, Y.M.; Kwon, H.-J.; Jeong, H.J.; Kim, C.Y.; Park, K.H.; Lee, W.S.; Ryu, Y.B. Chalcones isolated from Angelica keiskei inhibit cysteine proteases of SARS-CoV. J. Enzym. Inhib. Med. Chem. 2016, 31, 23–30. [CrossRef] [PubMed]
- 158. Lin, S.-C.; Ho, C.-T.; Chuo, W.-H.; Li, S.; Wang, T.T.; Lin, C.-C. Effective inhibition of MERS-CoV infection by resveratrol. *BMC Infect. Dis.* 2017, *17*, 144. [CrossRef]
- 159. Nassiri-Asl, M.; Hosseinzadeh, H. Review of the pharmacological effects of Vitis vinifera (Grape) and its bioactive compounds. *Phytother. Res. Int. J. Devoted Pharmacol. Toxicol. Eval. Nat. Prod. Deriv.* **2009**, 23, 1197–1204.
- 160. Chen, L.; Li, J.; Luo, C.; Liu, H.; Xu, W.; Chen, G.; Liew, O.W.; Zhu, W.; Puah, C.M.; Shen, X. Binding interaction of quercetin-3-β-galactoside and its synthetic derivatives with SARS-CoV 3CLpro: Structure–activity relationship studies reveal salient pharmacophore features. *Bioorgan. Med. Chem.* 2006, 14, 8295–8306. [CrossRef]
- 161. Pandey, P.; Khan, F.; Mazumder, A.; Rana, A.K.; Srivastava, Y. Inhibitory Potential of Dietary Phytocompounds of Nigella sativa against Key Targets of Novel Coronavirus (COVID-19). *Indian J. Pharm. Educ. Res.* **2021**, *55*, 190–197. [CrossRef]
- 162. Yu, M.-S.; Lee, J.; Lee, J.M.; Kim, Y.; Chin, Y.-W.; Jee, J.-G.; Keum, Y.-S.; Jeong, Y.-J. Identification of myricetin and scutellarein as novel chemical inhibitors of the SARS coronavirus helicase, nsP13. *Bioorgan. Med. Chem. Lett.* 2012, 22, 4049–4054. [CrossRef] [PubMed]
- Rakib, A.; Sami, S.A.; Islam, M.A.; Ahmed, S.; Faiz, F.B.; Khanam, B.H.; Marma, K.K.S.; Rahman, M.; Uddin, M.M.N.; Nainu, F.; et al. Epitope-Based Immunoinformatics Approach on Nucleocapsid Protein of Severe Acute Respiratory Syndrome-Coronavirus-2. *Molecules* 2020, 25, 5088. [CrossRef]
- 164. Chikhale, R.; Sinha, S.K.; Wanjari, M.; Gurav, N.S.; Ayyanar, M.; Prasad, S.; Khanal, P.; Dey, Y.N.; Patil, R.B.; Gurav, S.S. Computational assessment of Saikosaponins as adjuvant treatment for COVID-19: Molecular docking, dynamics, and network pharmacology analysis. *Mol. Divers.* 2021, 25, 1889–1904. [CrossRef]
- 165. Cheng, P.W.; Ng, L.T.; Chiang, L.C.; Lin, C.C. Antiviral effects of saikosaponins on human coronavirus 229E in vitro. *Clin. Exp. Pharmacol. Physiol.* **2006**, *33*, 612–616. [CrossRef] [PubMed]
- 166. Chang, F.-R.; Yen, C.-T.; Ei-Shazly, M.; Lin, W.-H.; Yen, M.-H.; Lin, K.-H.; Wu, Y.-C. Anti-human coronavirus (anti-HCoV) triterpenoids from the leaves of Euphorbia neriifolia. *Nat. Prod. Commun.* **2012**, *7*, 1934578X1200701103. [CrossRef]
- 167. Diniz, L.R.L.; Perez-Castillo, Y.; Elshabrawy, H.A.; de Sousa, D.P. Bioactive terpenes and their derivatives as potential SARS-CoV-2 proteases inhibitors from molecular modeling studies. *Biomolecules* **2021**, *11*, 74. [CrossRef] [PubMed]
- 168. Mahmud, S.; Paul, G.K.; Afroze, M.; Islam, S.; Gupt, S.B.R.; Razu, M.H.; Biswas, S.; Zaman, S.; Uddin, M.S.; Khan, M.; et al. Efficacy of Phytochemicals Derived from *Avicennia officinalis* for the Management of COVID-19: A Combined In Silico and Biochemical Study. *Molecules* 2021, 26, 2210. [CrossRef]
- Park, J.-Y.; Kim, J.H.; Kim, Y.M.; Jeong, H.J.; Kim, D.W.; Park, K.H.; Kwon, H.-J.; Park, S.-J.; Lee, W.S.; Ryu, Y.B. Tanshinones as selective and slow-binding inhibitors for SARS-CoV cysteine proteases. *Bioorgan. Med. Chem.* 2012, 20, 5928–5935. [CrossRef]
- Son, K.-H.; Oh, H.-M.; Choi, S.-K.; Han, D.C.; Kwon, B.-M. Anti-tumor abietane diterpenes from the cones of Sequoia sempervirens. *Bioorgan. Med. Chem. Lett.* 2005, 15, 2019–2021. [CrossRef]
- 171. Kim, S.; Hwang, B.Y.; Su, B.-N.; Chai, H.; Mi, Q.; Kinghorn, A.D.; Wild, R.; Swanson, S.M. Silvestrol, a potential anticancer rocaglate derivative from Aglaia foveolata, induces apoptosis in LNCaP cells through the mitochondrial/apoptosome pathway without activation of executioner caspase-3 or-7. *Anticancer Res.* 2007, 27, 2175–2183.
- 172. Müller, C.; Schulte, F.W.; Lange-Grünweller, K.; Obermann, W.; Madhugiri, R.; Pleschka, S.; Ziebuhr, J.; Hartmann, R.K.; Grünweller, A. Broad-spectrum antiviral activity of the eIF4A inhibitor silvestrol against corona-and picornaviruses. *Antivir. Res.* 2018, 150, 123–129. [CrossRef]
- 173. Wu, C.-Y.; Jan, J.-T.; Ma, S.-H.; Kuo, C.-J.; Juan, H.-F.; Cheng, Y.-S.E.; Hsu, H.-H.; Huang, H.-C.; Wu, D.; Brik, A. Small molecules targeting severe acute respiratory syndrome human coronavirus. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 10012–10017. [CrossRef]
- 174. Mutiawati, E.; Fahriani, M.; Mamada, S.S.; Fajar, J.K.; Frediansyah, A.; Maliga, H.A.; Ilmawan, M.; Emran, T.B.; Ophinni, Y.; Ichsan, I.; et al. Anosmia and dysgeusia in SARS-CoV-2 infection: Incidence and effects on COVID-19 severity and mortality, and the possible pathobiology mechanisms-a systematic review and meta-analysis. *F1000Research* 2021, 10, 40. [CrossRef]
- 175. Lusvarghi, S.; Bewley, C.A. Griffithsin: An antiviral lectin with outstanding therapeutic potential. *Viruses* **2016**, *8*, 296. [CrossRef] [PubMed]
- 176. Meuleman, P.; Albecka, A.; Belouzard, S.; Vercauteren, K.; Verhoye, L.; Wychowski, C.; Leroux-Roels, G.; Palmer, K.E.; Dubuisson, J. Griffithsin has antiviral activity against hepatitis C virus. *Antimicrob. Agents Chemother.* 2011, 55, 5159–5167. [CrossRef] [PubMed]
- 177. Sumon, T.A.; Hussain, M.; Hasan, M.; Hasan, M.; Jang, W.J.; Bhuiya, E.H.; Chowdhury, A.A.M.; Sharifuzzaman, S.; Brown, C.L.; Kwon, H.-J. A revisit to the research updates of drugs, vaccines, and bioinformatics approaches in combating COVID-19 pandemic. *Front. Mol. Biosci.* 2021, 7, 493. [CrossRef] [PubMed]
- 178. Millet, J.K.; Séron, K.; Labitt, R.N.; Danneels, A.; Palmer, K.E.; Whittaker, G.R.; Dubuisson, J.; Belouzard, S. Middle East respiratory syndrome coronavirus infection is inhibited by griffithsin. *Antivir. Res.* **2016**, *133*, 1–8. [CrossRef]
- 179. Vijayaraj, R.; Altaff, K.; Rosita, A.S.; Ramadevi, S.; Revathy, J. Bioactive compounds from marine resources against novel corona virus (2019-nCoV): In silico study for corona viral drug. *Nat. Prod. Res.* **2020**, 1–5. [CrossRef]

- Bhatt, A.; Arora, P.; Prajapati, S.K. Can Algal Derived Bioactive Metabolites Serve as Potential Therapeutics for the Treatment of SARS-CoV-2 Like Viral Infection? *Front. Microbiol.* 2020, 11, 596374. [CrossRef]
- Rampogu, S.; Gajula, R.G.; Lee, G.; Kim, M.O.; Lee, K.W. Unravelling the Therapeutic Potential of Marine Drugs as SARS-CoV-2 Inhibitors: An Insight from Essential Dynamics and Free Energy Landscape. *Comput. Biol. Med.* 2021, 135, 104525. [CrossRef] [PubMed]
- 182. Campione, E.; Cosio, T.; Rosa, L.; Lanna, C.; Di Girolamo, S.; Gaziano, R.; Valenti, P.; Bianchi, L. Lactoferrin as protective natural barrier of respiratory and intestinal mucosa against coronavirus infection and inflammation. *Int. J. Mol. Sci.* 2020, 21, 4903. [CrossRef]
- Ikeda, M.; Nozaki, A.; Sugiyama, K.; Tanaka, T.; Naganuma, A.; Tanaka, K.; Sekihara, H.; Shimotohno, K.; Saito, M.; Kato, N. Characterization of antiviral activity of lactoferrin against hepatitis C virus infection in human cultured cells. *Virus Res.* 2000, 66, 51–63. [CrossRef]
- Lang, J.; Yang, N.; Deng, J.; Liu, K.; Yang, P.; Zhang, G.; Jiang, C. Inhibition of SARS pseudovirus cell entry by lactoferrin binding to heparan sulfate proteoglycans. *PLoS ONE* 2011, 6, e23710. [CrossRef]
- Celik, I.; Yadav, R.; Duzgun, Z.; Albogami, S.; El-Shehawi, A.M.; Idroes, R.; Tallei, T.E.; Emran, T.B. Interactions of the receptor binding domain of SARS-CoV-2 variants with hACE2: Insights from molecular docking analysis and molecular dynamic simulation. *Biology* 2021, 10, 880. [CrossRef]
- 186. Van der Strate, B.; Beljaars, L.; Molema, G.; Harmsen, M.; Meijer, D. Antiviral activities of lactoferrin. *Antivir. Res.* 2001, 52, 225–239. [CrossRef]
- 187. Wakabayashi, H.; Oda, H.; Yamauchi, K.; Abe, F. Lactoferrin for prevention of common viral infections. *J. Infect. Chemother.* **2014**, 20, 666–671. [CrossRef] [PubMed]
- 188. Clausen, T.M.; Sandoval, D.R.; Spliid, C.B.; Pihl, J.; Perrett, H.R.; Painter, C.D.; Narayanan, A.; Majowicz, S.A.; Kwong, E.M.; McVicar, R.N. SARS-CoV-2 infection depends on cellular heparan sulfate and ACE2. *Cell* 2020, 183, 1043–1057.e15. [CrossRef] [PubMed]
- 189. Elnagdy, S.; AlKhazindar, M. The potential of antimicrobial peptides as an antiviral therapy against COVID-19. *ACS Pharmacol. Transl. Sci.* **2020**, *3*, 780–782. [CrossRef] [PubMed]
- 190. Jenssen, H.; Hancock, R.E. Antimicrobial properties of lactoferrin. Biochimie 2009, 91, 19–29. [CrossRef] [PubMed]
- 191. De Haan, C.A.; Li, Z.; Te Lintelo, E.; Bosch, B.J.; Haijema, B.J.; Rottier, P.J. Murine coronavirus with an extended host range uses heparan sulfate as an entry receptor. *J. Virol.* **2005**, *79*, 14451–14456. [CrossRef] [PubMed]
- 192. Milewska, A.; Zarebski, M.; Nowak, P.; Stozek, K.; Potempa, J.; Pyrc, K. Human coronavirus NL63 utilizes heparan sulfate proteoglycans for attachment to target cells. *J. Virol.* **2014**, *88*, 13221–13230. [CrossRef]
- 193. Actor, J.K.; Hwang, S.-A.; Kruzel, M.L. Lactoferrin as a natural immune modulator. *Curr. Pharm. Des.* **2009**, *15*, 1956–1973. [CrossRef] [PubMed]
- 194. Ou, X.; Liu, Y.; Lei, X.; Li, P.; Mi, D.; Ren, L.; Guo, L.; Guo, R.; Chen, T.; Hu, J. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. *Nat. Commun.* **2020**, *11*, 1620. [CrossRef]
- 195. Pišlar, A.; Mitrović, A.; Sabotič, J.; Pečar Fonović, U.; Perišić Nanut, M.; Jakoš, T.; Senjor, E.; Kos, J. The role of cysteine peptidases in coronavirus cell entry and replication: The therapeutic potential of cathepsin inhibitors. *PLoS Pathog.* 2020, 16, e1009013. [CrossRef]
- 196. Chang, R.; Ng, T.B.; Sun, W.-Z. Lactoferrin as potential preventative and adjunct treatment for COVID-19. *Int. J. Antimicrob. Agents* **2020**, *56*, 106118. [CrossRef] [PubMed]
- 197. Machnicki, M.; Zimecki, M.; Zagulski, T. Lactoferrin regulates the release of tumour necrosis factor alpha and interleukin 6 in vivo. *Int. J. Exp. Pathol.* **1993**, *74*, 433. [PubMed]
- 198. Jovic, T.H.; Ali, S.R.; Ibrahim, N.; Jessop, Z.M.; Tarassoli, S.P.; Dobbs, T.D.; Holford, P.; Thornton, C.A.; Whitaker, I.S. Could vitamins help in the fight against COVID-19? *Nutrients* **2020**, *12*, 2550. [CrossRef] [PubMed]
- 199. Abobaker, A.; Alzwi, A.; Alraied, A.H.A. Overview of the possible role of vitamin C in management of COVID-19. *Pharmacol. Rep.* **2020**, *72*, 1517–1528. [CrossRef]
- 200. Syahrul, S.; Maliga, H.A.; Ilmawan, M.; Fahriani, M.; Mamada, S.S.; Fajar, J.K.; Frediansyah, A.; Syahrul, F.N.; Imran, I.; Haris, S.; et al. Hemorrhagic and ischemic stroke in patients with coronavirus disease 2019: Incidence, risk factors, and pathogenesis-a systematic review and meta-analysis. *F1000Res.* **2021**, *10*, 34. [CrossRef]
- 201. Schorah, C. The level of vitamin C reserves required in man: Towards a solution to the controversy. *Proc. Nutr. Soc.* **1981**, *40*, 147–154. [CrossRef]
- 202. Feyaerts, A.F.; Luyten, W. Vitamin C as prophylaxis and adjunctive medical treatment for COVID-19? *Nutrition* **2020**, *79*, 110948. [CrossRef]
- 203. Hiedra, R.; Lo, K.B.; Elbashabsheh, M.; Gul, F.; Wright, R.M.; Albano, J.; Azmaiparashvili, Z.; Patarroyo Aponte, G. The use of IV vitamin C for patients with COVID-19: A case series. *Expert Rev. Anti-Infect. Ther.* 2020, 18, 1259–1261. [CrossRef]
- 204. Sözener, Z.C.; Cevhertas, L.; Nadeau, K.; Akdis, M.; Akdis, C.A. Environmental factors in epithelial barrier dysfunction. J. Allergy Clin. Immunol. 2020, 145, 1517–1528. [CrossRef]
- Crane-Godreau, M.A.; Clem, K.J.; Payne, P.; Fiering, S. Vitamin D deficiency and air pollution exacerbate COVID-19 through suppression of antiviral peptide LL37. Front. Public Health 2020, 8, 232. [CrossRef] [PubMed]
- 206. Klotman, M.E.; Chang, T.L. Defensins in innate antiviral immunity. Nat. Rev. Immunol. 2006, 6, 447–456. [CrossRef] [PubMed]

- 207. Munshi, R.; Hussein, M.H.; Toraih, E.A.; Elshazli, R.M.; Jardak, C.; Sultana, N.; Youssef, M.R.; Omar, M.; Attia, A.S.; Fawzy, M.S. Vitamin D insufficiency as a potential culprit in critical COVID-19 patients. *J. Med. Virol.* **2021**, *93*, 733–740. [CrossRef]
- Orru, B.; Szekeres-Bartho, J.; Bizzarri, M.; Spiga, A.; Unfer, V. Inhibitory effects of Vitamin D on inflammation and IL-6 release. A further support for COVID-19 management. *Eur. Rev. Med. Pharm. Sci* 2020, 24, 8187–8193.
- Brewer, J.; Marti, J.G.; Brufsky, A. Potential interventions for SARS-CoV-2 infections: Zinc showing promise. J. Med. Virol. 2020, 93, 1201–1203. [CrossRef] [PubMed]
- 210. Hoang, B.X.; Han, B. A possible application of hinokitiol as a natural zinc ionophore and anti-infective agent for the prevention and treatment of COVID-19 and viral infections. *Med. Hypotheses* **2020**, *145*, 110333. [CrossRef]
- 211. Maares, M.; Haase, H. Zinc and immunity: An essential interrelation. Arch. Biochem. Biophys. 2016, 611, 58-65. [CrossRef]
- 212. Shin, K.; Fogg, V.C.; Margolis, B. Tight junctions and cell polarity. Annu. Rev. Cell Dev. Biol. 2006, 22, 207–235. [CrossRef]
- Sturniolo, G.C.; Fries, W.; Mazzon, E.; Di Leo, V.; Barollo, M.; D'inca, R. Effect of zinc supplementation on intestinal permeability in experimental colitis. J. Lab. Clin. Med. 2002, 139, 311–315. [CrossRef]
- Coperchini, F.; Chiovato, L.; Croce, L.; Magri, F.; Rotondi, M. The cytokine storm in COVID-19: An overview of the involvement of the chemokine/chemokine-receptor system. *Cytokine Growth Factor Rev.* 2020, 53, 25–32. [CrossRef]
- 215. Rahman, M.T.; Idid, S.Z. Can Zn be a critical element in COVID-19 treatment? *Biol. Trace Elem. Res.* 2021, 199, 550–558. [CrossRef] [PubMed]
- 216. Prasad, A.S. Zinc: Mechanisms of host defense. J. Nutr. 2007, 137, 1345–1349. [CrossRef]
- 217. Truong-Tran, A.Q.; Carter, J.; Ruffin, R.; Zalewski, P.D. New insights into the role of zinc in the respiratory epithelium. *Immunol. Cell Biol.* **2001**, *79*, 170–177. [CrossRef]
- 218. Lanke, K.; Krenn, B.; Melchers, W.; Seipelt, J.; Van Kuppeveld, F. PDTC inhibits picornavirus polyprotein processing and RNA replication by transporting zinc ions into cells. *J. Gen. Virol.* 2007, *88*, 1206–1217. [CrossRef] [PubMed]
- Kumar, R.; Yeni, C.M.; Utami, N.A.; Masand, R.; Asrani, R.K.; Patel, S.K.; Kumar, A.; Yatoo, M.I.; Tiwari, R.; Natesan, S.; et al. SARS-CoV-2 infection during pregnancy and pregnancy-related conditions: Concerns, challenges, management and mitigation strategies–a narrative review. J. Infect. Public Health 2021, 14, 863–875. [CrossRef]
- 220. te Velthuis, A.; Van Den Worm, S.; Sims, A.; Baric, R.; Snijder, E. Zn<sup>2+</sup> inhibits coronavirus and arterivirus RNA polymerase activity in vitro. *PLoS Pathog.* **2010**, *6*, e1001176. [CrossRef]
- Berg, K.; Bolt, G.; Andersen, H.; Owen, T.C. Zinc potentiates the antiviral action of human IFN-α tenfold. J. Interferon Cytokine Res. 2001, 21, 471–474. [CrossRef] [PubMed]
- 222. McCarty, M.F.; DiNicolantonio, J.J. Nutraceuticals have potential for boosting the type 1 interferon response to RNA viruses including influenza and coronavirus. *Prog. Cardiovasc. Dis.* 2020, 63, 383. [CrossRef] [PubMed]
- Ison, M.G.; Wolfe, C.; Boucher, H.W. Emergency use authorization of remdesivir: The need for a transparent distribution process. JAMA 2020, 323, 2365–2366. [CrossRef]
- 224. Spinner, C.D.; Gottlieb, R.L.; Criner, G.J.; López, J.R.A.; Cattelan, A.M.; Viladomiu, A.S.; Ogbuagu, O.; Malhotra, P.; Mullane, K.M.; Castagna, A. Effect of remdesivir vs standard care on clinical status at 11 days in patients with moderate COVID-19: A randomized clinical trial. *JAMA* 2020, 324, 1048–1057. [CrossRef] [PubMed]
- 225. Leecharoen, S.; Wangkaew, S.; Louthrenoo, W. Ocular side effects of chloroquine in patients with rheumatoid arthritis, systemic lupus erythematosus and scleroderma. *J. Med. Assoc. Thail.* **2007**, *90*, *52*.
- 226. Mubagwa, K. Chloroquine cardiac effects and toxicity. A short update. Int J Antimicrob Agents 2020, 56, 106057. [CrossRef]
- 227. Tumilaar, S.G.; Fatimawali, F.; Niode, N.J.; Effendi, Y.; Idroes, R.; Adam, A.A.; Rakib, A.; Emran, T.B.; Tallei, T.E. The potential of leaf extract of Pangium edule Reinw as HIV-1 protease inhibitor: A computational biology approach. *J. Appl. Pharm. Sci.* 2021, 11, 101–110. [CrossRef]
- 228. Wijayasinghe, Y.S.; Bhansali, P.; Viola, R.E.; Kamal, M.A.; Poddar, N.K. Natural products: A rich source of antiviral drug lead candidates for the management of COVID-19. *Curr. Pharm. Des.* **2021**, *27*, 3526–3550. [CrossRef]
- 229. Lan, S.-H.; Lai, C.-C.; Huang, H.-T.; Chang, S.-P.; Lu, L.-C.; Hsueh, P.-R. Tocilizumab for severe COVID-19: A systematic review and meta-analysis. *Int. J. Antimicrob. Agents* 2020, *56*, 106103. [CrossRef] [PubMed]
- Hashemian, S.M.; Farhadi, T.; Velayati, A.A. A review on favipiravir: The properties, function, and usefulness to treat COVID-19. Expert Rev. Anti-Infect. Ther. 2021, 19, 1029–1037. [CrossRef]
- Rahman, A.; Niloofa, R.; Jayarajah, U.; De Mel, S.; Abeysuriya, V.; Seneviratne, S.L. Hematological Abnormalities in COVID-19: A Narrative Review. Am. J. Trop. Med. Hyg. 2021, 104, 1188. [CrossRef]
- 232. Bessière, F.; Roccia, H.; Delinière, A.; Charrière, R.; Chevalier, P.; Argaud, L.; Cour, M. Assessment of QT intervals in a case series of patients with coronavirus disease 2019 (COVID-19) infection treated with hydroxychloroquine alone or in combination with azithromycin in an intensive care unit. *JAMA Cardiol.* 2020, *5*, 1067–1069. [CrossRef]
- 233. Kremer, C. Azithromycin—A new macrolide. Prim. Care Update OB/GYNS 2002, 9, 174–175. [CrossRef]
- Mercuro, N.J.; Yen, C.F.; Shim, D.J.; Maher, T.R.; McCoy, C.M.; Zimetbaum, P.J.; Gold, H.S. Risk of QT interval prolongation associated with use of hydroxychloroquine with or without concomitant azithromycin among hospitalized patients testing positive for coronavirus disease 2019 (COVID-19). *JAMA Cardiol.* 2020, *5*, 1036–1041. [CrossRef] [PubMed]