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# *Drosophila* wing imaginal discs respond to mechanical injury via slow $\text{InsP}_3\text{R}$ -mediated intercellular calcium waves

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Calcium signalling is a highly versatile cellular communication system that modulates basic functions such as cell contractility, essential steps of animal development such as fertilization and higher-order processes such as memory. We probed the function of calcium signalling in *Drosophila* wing imaginal discs through a combination of *ex vivo* and *in vivo* imaging and genetic analysis. Here we discover that wing discs display slow, long-range intercellular calcium waves (ICWs) when mechanically stressed *in vivo* or cultured *ex vivo*. These slow imaginal disc intercellular calcium waves (SIDICs) are mediated by the inositol-3-phosphate receptor, the endoplasmic reticulum (ER) calcium pump SERCA and the key gap junction component *Inx2*. The knockdown of genes required for SIDIC formation and propagation negatively affects wing disc recovery after mechanical injury. Our results reveal a role for ICWs in wing disc homeostasis and highlight the utility of the wing disc as a model for calcium signalling studies.

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Calcium ( $\text{Ca}^{2+}$ ) is a ubiquitous and highly versatile signal that modulates basic cellular functions, such as contractility and secretion, essential steps of animal development such as fertilization and proliferation, and higher-order processes such as learning and memory<sup>1–3</sup>. Intercellular calcium waves (ICWs) constitute an interesting aspect of  $\text{Ca}^{2+}$  signalling currently been untangled<sup>4,5</sup>.

ICWs have been observed in a variety of systems: *in vivo* and *ex vivo*<sup>4–7</sup>. In zebrafish<sup>8,9</sup> and *Xenopus*<sup>10–13</sup>, ICWs occur during important developmental milestones such as gastrulation<sup>2</sup>. However, the functional relevance of ICWs often remains open<sup>1,2</sup>.

ICWs have been reported to occur during specific stages of *Drosophila* embryonic development<sup>14</sup>. Recently, calcium flashes—a fast type of ICWs—have been shown to regulate inflammation and the response to injury in the fly embryo<sup>15</sup>. Further, it was also recently found that calcium signalling might regulate coordinated waves of actomyosin flow and cell constriction on wounding of the *Drosophila* pupal notum<sup>16</sup>.

Yet, calcium signalling has seldom been explored in wing imaginal discs, arguably one of the better-studied model organs. To redress this we decided to use our recent development of a system for *ex vivo* culturing of imaginal discs<sup>17</sup> and improvements in ultrasensitive genetically encoded calcium indicators, the GCamp6 reporters<sup>18</sup>.

Through *ex vivo* live imaging of wing discs expressing GCamp6s<sup>18</sup>, we uncover a slow type of ICW that we named slow imaginal disc ICW (SIDIC). Next, we characterize the molecular mechanism underlying the SIDICs through a combination of quantitative live imaging of ICWs and gene knockdown analysis. We find that the SIDICs are a cell-to-cell chain of intracellular  $\text{Ca}^{2+}$  release from the endoplasmic reticulum (ER)  $\text{Ca}^{2+}$  stores downstream of the inositol-3-phosphate receptor ( $\text{InsP}_3\text{R}$ ) that requires gap junctions for intercellular propagation. Further, we find that, *in vivo*, the SIDICs constitute a response to mechanical stress. Finally, we explore whether the SIDICs might play a role as an injury response. We find that the knockdown of the  $\text{InsP}_3\text{R}$  and the key gap junction component innexin2 (*Inx2*) affect the wound healing of wing discs negatively. Our results highlight a role for calcium signalling in the form of slow, long-range  $\text{InsP}_3\text{R}$ -mediated  $\text{Ca}^{2+}$  waves in the homeostasis of wing imaginal discs.

## Results

***Ex vivo*-cultured imaginal wing discs display ICWs.** To study calcium signalling activity in imaginal wing discs, we employed a previously developed *ex vivo* culturing setup that is amenable to high-resolution live imaging and supports normal levels of proliferation for up to 12 h (ref. 17). We employed a genetically encoded calcium indicator, UAS-GCamp6s, under the control of a wing pouch driver (*nubbin-Gal4*). We refer to this fly strain as *nub>GCamp6s*. Strikingly, we observed large SIDICs propagating across the wing imaginal disc explants (Fig. 1a,b and Supplementary Movie 1).

To better understand this phenomenon, we characterized the properties of the SIDICs (Supplementary Fig. 1A–J). The SIDIC wavefront is slow at  $\sim 0.4 \mu\text{m s}^{-1}$  (Supplementary Fig. 1A,B:  $0.41 \pm 0.17$ ,  $n = 10$  discs). During each wave, the recruited imaginal disc cells mobilize intracellular calcium for nearly 6 min (Supplementary Fig. 1C–E:  $6.18 \pm 1.97$ ,  $n = 10$  discs). *Ex vivo*, 75% ( $n = 41/55$  discs) of SIDICs propagate throughout the whole pouch with a magnitude (area covered by wave/pouch area) close to 1 (Supplementary Fig. 1F–G:  $0.99 \pm 0.01$ ,  $n = 10$  discs). In 13% of the discs the SIDICs were of smaller magnitude with only a fraction of the cells of the wing pouch being recruited ( $0.43 \pm 0.30$ ,  $n = 9$ ). Finally, an additional 12% did not display

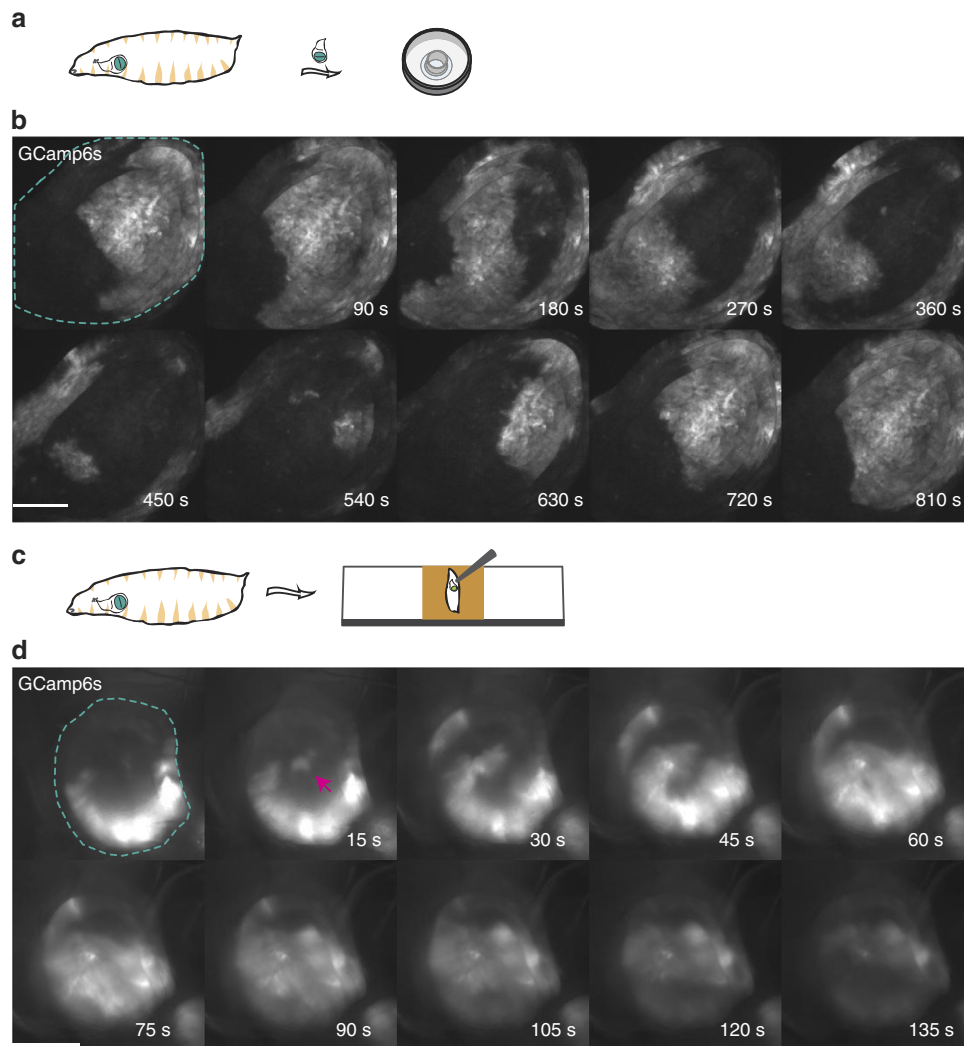
SIDIC activity. *Ex vivo*, the SIDICs recur with a relatively constant period (Supplementary Fig. 1H–J:  $13.9 \text{ min} \pm 3.4$ ,  $n = 22$  discs). In addition, per disc the spatio-temporal pattern of propagation is generally repeated (Fig. 1b and Supplementary Movie 1). Both the duration of the recurrence period and the spatio-temporal pattern of wave propagation vary more among discs than per disc.

**The SIDICs are induced by mechanical stress.** ICWs can be induced by mechanical stimulation and mechanical wounding *in vitro* in airway epithelial cells<sup>19</sup> and human urothelial cell monolayers<sup>20</sup>. Further, during our first trials at *in vivo* imaging of SIDICs we had found that SIDICs could only be observed if the larvae were squeezed in between a coverslip and a glass slide in a setup that resulted in mechanical stress for the larva (Supplementary Fig. 2). We also noted that sometimes the SIDICs seemed to occur after strong larval movements, again pointing towards mechanical stress as a trigger.

We therefore tested whether mechanical stress could trigger a SIDIC wave *in vivo*. We developed a setup in which we glued larvae, imaginal discs facing down, on a microscope slide with double sided adhesive tape (Fig. 1c). Next, we used blunt forceps to punch the imaginal discs through the larval cuticle, but crucially, without piercing the cuticle. We found that this manipulation induced two different calcium responses. First, there is a direct strong and localized increase in intracellular calcium levels (dashed lines in Fig. 3). Further, in 63% of perturbed discs ( $n = 10/16$ ) we observed a SIDIC wave appearing during the next 30 min (Fig. 1d and Supplementary Movie 2). In contrast, we only observed SIDICs in 9.8% ( $n = 4/41$ ) of unperturbed discs. The presence of SIDICs in unperturbed discs could indicate a different potential source of stimulation for the SIDICs (for example, mechanical stimulation induced by larval movements during foraging, before imaging) or that the discs were sometimes involuntarily mechanically stressed, while the larvae were glued onto the adhesive tape. The delay between the mechanical stimulation and the SIDICs suggests that these waves constitute a later response to injury and are in this respect different from the fast calcium flashes described in the embryo<sup>15</sup> or notum<sup>16</sup>.

The SIDICs induced by mechanical stress happen on a similar timeframe as those observed *ex vivo* and require several minutes to spread through the wing disc (Fig. 1d). The SIDICs are difficult to characterize *in vivo* due to larval movements. However, we found the following approximations: the wavefront speed is of  $0.76 \pm 0.42 \mu\text{m s}^{-1}$  ( $n = 8$ ); the duration of intracellular calcium mobilization seems shorter at  $3.7 \pm 1.2 \text{ min}$  ( $n = 8$ ); the magnitude is generally smaller and more variable ( $0.36 \pm 0.28$ ,  $n = 8$ ); and there seems to be no constant recurring period, although we did observe recurring waves ( $n = 5/16$ ) (Supplementary Movie 3). The disparities in wave magnitude could reflect differences in the type and magnitude of stimuli inducing the waves. *In vivo* the trigger is a transient mechanical stress, whereas *ex vivo* the stimuli are linked to the dissection and culturing conditions, and are likely to be stronger and constantly present.

**The SIDICs mobilize intracellular  $\text{Ca}^{2+}$  via  $\text{InsP}_3\text{R}$  and SERCA.** The magnitude of the SIDICs and the constant intensity of GCamp6s fluorescence during propagation hint to a calcium-induced calcium release mechanism of propagation. Calcium-induced calcium release can be induced by the  $\text{InsP}_3\text{R}$  and the ryanodine receptor (R<sub>YR</sub>), sometimes exclusively and sometimes in combination<sup>1–3,5</sup>. Previous analyses of R<sub>YR</sub> expression and function concluded that R<sub>YR</sub> was only important for muscle function in *Drosophila*<sup>21–25</sup>. We therefore focused on  $\text{InsP}_3\text{R}$ .



**Figure 1 | Observation of intercellular calcium waves in imaginal wing discs *ex vivo* and *in vivo*.** (a) *Ex vivo* imaging setup. Dissected wing discs are cultured in WM1 inside a modified cell culture insert on an imaging disc and imaged with an inverted spinning-disc confocal microscope. Fly strains: *nub* > *GCamp6s* X *yw*. (b) Example of a SIDIC wave *ex vivo*. This wave propagated approximately along the antero-posterior axis of the wing disc and recruited all of the cells of the wing pouch. It is noteworthy how the wave starts to recur after 540 s. Scale bar, 100  $\mu\text{m}$ . Fly strains: *nub* > *GCamp6s* X *yw*. (c) *In vivo* imaging setup (inverted). Larvae were glued to a microscope slide, dorsally, with double-sided adhesive tape. We employed an inverted widefield fluorescence microscope. Fly strains: *nub* > *GCamp6s* X *yw*. (d) Observation of a SIDIC wave *in vivo*. The magenta arrowhead indicates the point where the SIDIC originates. Scale bar, 100  $\mu\text{m}$ . Fly strains: *nub* > *GCamp6s* X *yw*.

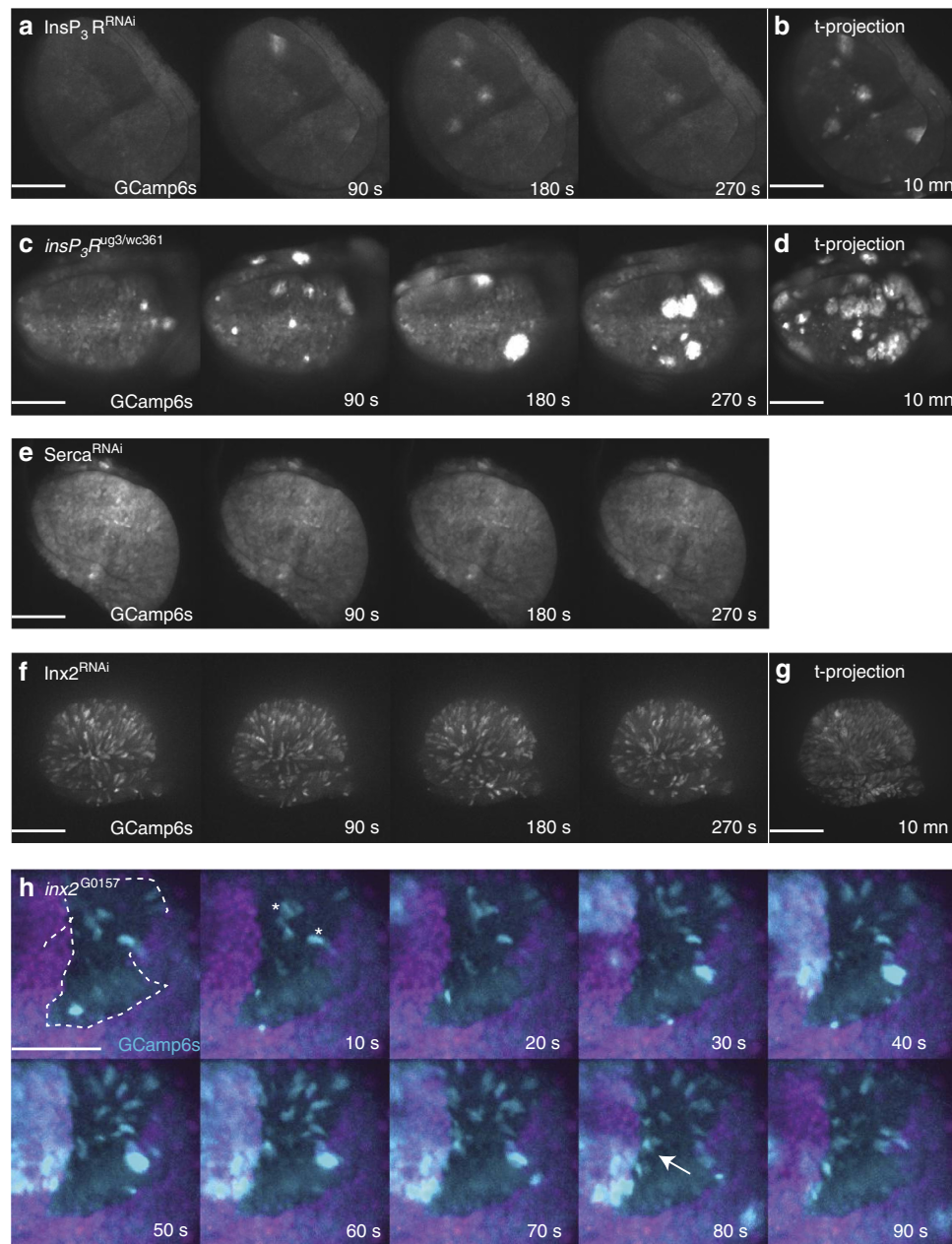
The knockdown of the  $\text{InsP}_3\text{R}$  abolished the SIDICs *ex vivo* (Fig. 2a,b and Supplementary Movie 4). During  $\text{InsP}_3\text{R}$  knockdown, in place of SIDICs we observed spontaneous occurrences of intracellular calcium release dispersed discretely throughout the wing pouch (Supplementary Fig. 3A and Supplementary Movie 4). These ‘bursts’ initiate in a small number of neighbouring cells and can propagate for one to two cell diameters (Supplementary Fig. 3B). The duration of calcium mobilization during a burst is shorter than during a SIDIC wave ( $44.3 \pm 18.5$  s,  $n = 23$  versus 6.18 min,  $1 \pm 1.97$ ). However, the speed at which these bursts propagate is comparable to that of the SIDICs wavefront ( $0.51 \pm 0.16 \mu\text{m s}^{-1}$ ,  $n = 18$ ). In some cases, the bursts can be quite large, involving up to  $\sim 50$  cells. However, these bursts never lead to the propagation of a SIDIC wave. We confirmed these RNA interference (RNAi) results by generating a *trans*-heterozygous hypomorph combinations of *InsP3R* alleles (*ug3/wc361*)<sup>26</sup>. The *trans*-heterozygous situation mimicked the RNAi experiments but the calcium activity seemed higher (Fig. 2c,d). This could indicate that the calcium bursts are a

result of residual  $\text{InsP}_3\text{R}$  activity present in both the knockdown and hypomorph situations.

Taken together, these experiments suggest that the calcium bursts represent SIDICs that failed to propagate due to insufficient  $\text{InsP}_3\text{R}$  activity. This could indicate that intracellular  $\text{Ca}^{2+}$  has to be mobilized for a sufficiently long duration before the SIDICs can propagate from one cell to the next. However, these bursts could also point to other necessary components for SIDIC activity that remain to be uncovered.

On activation, the  $\text{InsP}_3\text{R}$  mobilizes  $\text{Ca}^{2+}$  from the intracellular  $\text{Ca}^{2+}$  stores of the ER. The ER calcium pump SERCA is required for the maintenance of the ER  $\text{Ca}^{2+}$  stores<sup>1,27</sup>. Consistent with the notion that the mobilization of intracellular  $\text{Ca}^{2+}$  from the ER is necessary for the propagation of the SIDICs, SERCA knockdown abolished the SIDICs (Fig. 2e and Supplementary Movie 5).

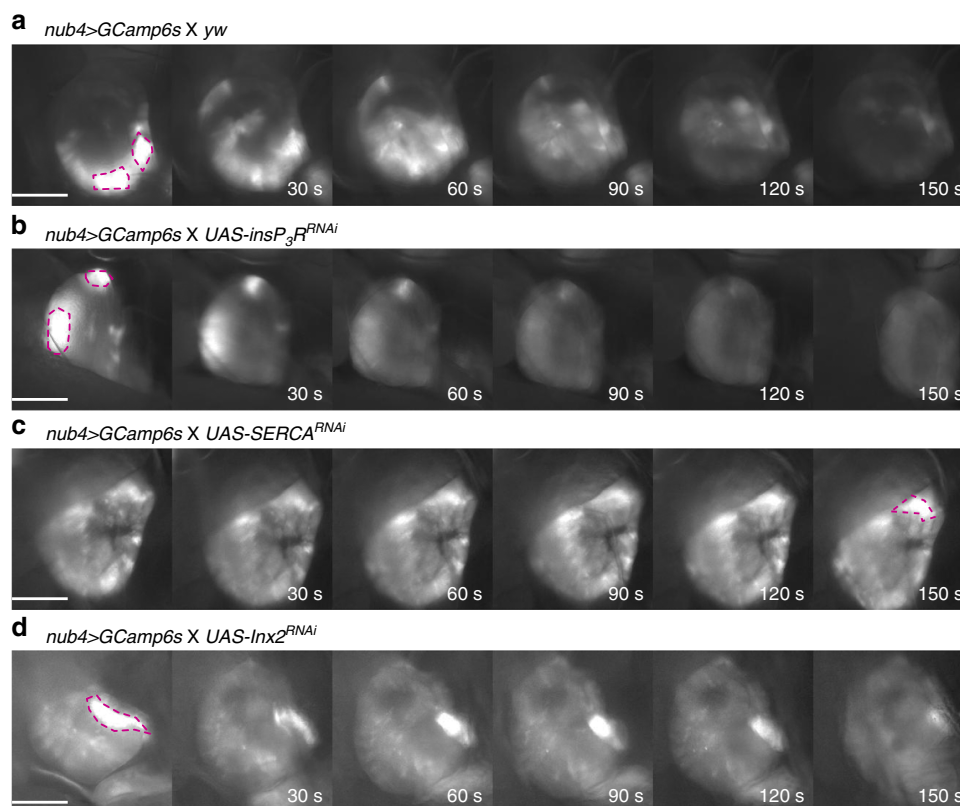
**The SIDICs require *Inx2* for propagation.** ICWs propagate via two major mechanisms: gap junctions and paracrine signalling



**Figure 2 | Characterization of key genes required for SIDIC wave propagation *ex vivo*.** (a) *InsP<sub>3</sub>R*<sup>RNAi</sup> prevents the formation of the SIDICs. The presence of groups of cells undergoing intracellular calcium transients, which we refer to as calcium bursts, are noteworthy. Scale bar, 100  $\mu$ m. Fly strains: *nub* > *GCaMP6s* X *UAS-InsP<sub>3</sub>R*<sup>RNAi</sup> (NIG 1063-R1). (b) Ten minutes time projection of *GCaMP6s* fluorescence for *InsP<sub>3</sub>R*<sup>RNAi</sup>. Scale bar, 100  $\mu$ m. Fly strains: *nub* > *GCaMP6s* X *UAS-InsP<sub>3</sub>R*<sup>RNAi</sup> (NIG 1063-R1). (c) *Trans*-heterozygous hypomorphs for *InsP<sub>3</sub>R* (*ug3/wc361*) *InsP<sub>3</sub>R*<sup>RNAi</sup>. The calcium transient activity is more intense, probably revealing higher residual *InsP<sub>3</sub>R* activity. Scale bar, 100  $\mu$ m. Fly strains: *InsP<sub>3</sub>R*<sup>ug3</sup> X *InsP<sub>3</sub>R*<sup>wc361</sup>. (d) Ten minutes time projection of *GCaMP6s* fluorescence for *InsP<sub>3</sub>R*<sup>ug3/wc361</sup>. Scale bar, 100  $\mu$ m. Fly strains: *InsP<sub>3</sub>R*<sup>ug3</sup> X *InsP<sub>3</sub>R*<sup>wc361</sup>. (e) RNAi-mediated knockdown of the ER calcium pump SERCA (*SERCA*<sup>RNAi</sup>) prevents the formation of the SIDICs consistent with the requirement of intracellular  $Ca^{2+}$  stores for the SIDICs. Scale bar, 100  $\mu$ m. Fly strains: *nub* > *GCaMP6s* X *UAS-SERCA*<sup>RNAi</sup> (BL 25928). Similar results were obtained with BL 44581. (f) An interfering RNA against the gap junction component *Inx2* (*Inx2*<sup>RNAi</sup>) prevents the formation of the SIDICs. Scale bar, 100  $\mu$ m. Fly strains: *nub* > *GCaMP6s* X *UAS-Inx2*<sup>RNAi</sup> (BL 29306). Similar results were obtained with *UAS-wizInx2*. (g) Ten minutes time projection of *GCaMP6s* fluorescence for *Inx2*<sup>RNAi</sup>. Scale bar, 100  $\mu$ m. Fly strains: *nub* > *GCaMP6s* X *UAS-Inx2*<sup>RNAi</sup> (BL 29306). (h) A SIDIC wave stopping at the boundary of an *Inx2* mutant clone generated by somatic recombination (*inx2*<sup>G0157</sup> -/-). The wave ends at the interior boundary of the clone (white arrow indicates increased *GCaMP6s* fluorescence at the inner boundary of the clone). The presence, inside the clone, of the same calcium 'sparkles' observed in *Inx2*<sup>RNAi</sup> discs are noteworthy. Scale bar, 50  $\mu$ m. Fly strains: *w*, *inx2*<sup>G0157</sup>, *FRT19A/FM7c* X *ubi* > *mRFPNLS,w,hsflp,FRT19*; *nubbin-Gal4,GCaMP6s*; *MKRS/Tm6b*.

with an extracellular messenger<sup>5</sup>. The ligand of the *InsP<sub>3</sub>R*, IP3 diffuses through gap junctions<sup>5</sup>. Hence, we tested whether gap junctions were required for SIDIC propagation by knocking down the key gap junction component *Inx2* (ref. 28). Discs with impaired gap junction communication did not display SIDICs

(Fig. 2f and Supplementary Movie 6). Time-lapse analysis of *Inx2* knockdown experiments revealed a striking, sparkle-like pattern of intracellular  $Ca^{2+}$  release comprising either a single cell or small groups of adjacent cells (Supplementary Fig. 4A). These calcium pulses occur seemingly randomly throughout the wing



**Figure 3 | The SIDICs components identified *ex vivo* are required *in vivo*.** (a) Control wing discs of the *nub4>GCamp6s* genotype displaying a SIDIC after mechanical induction (same time lapse as Fig. 1d). SIDICs were observed in 63% of mechanical stimulation experiments during a 30 min time window (10/16). Scale bar, 100  $\mu$ m. Fly strains: *nub4>GCamp6s X yw*. (b) *InsP3R* knockdown prevents the formation of SIDICs, *in vivo* (0/14). Scale bar, 100  $\mu$ m. Fly strains: *nub4>GCamp6s X UAS-InsP3R<sup>RNAi</sup>* (NIG 1063-R1). (c) *SERCA<sup>RNAi</sup>* prevents the formation of SIDICs *in vivo* (0/7). Scale bar, 100  $\mu$ m. Fly strains: *nub4>GCamp6s X UAS-SERCA<sup>RNAi</sup>* (BL 25928). (d) *Inx2<sup>RNAi</sup>* prevents the formation of SIDICs, *in vivo* (0/9). Scale bar, 100  $\mu$ m. Fly strains: *nub4>GCamp6s X UAS-Inx2<sup>RNAi</sup>* (BL 29306). (a,b,c,d) Dashed lines highlight areas of immediate calcium release. These areas do not expand as a SIDIC wave.

disc epithelium and could sometimes be seen to propagate across neighbouring cells for short distances (Supplementary Fig. 4A). These results could support a role for gap junctions in SIDIC wave propagation. However, to rule out that loss of optimal gap junction function could be leading to general effects, for example, on calcium homeostasis, we performed *ex vivo* time-lapse analysis of wing discs with *inx2* mutant clones (*inx2<sup>G0157-7-</sup>*). Time-lapse analysis of *inx2* mutant clones generated by flippase recognition target (FRT)-mediated somatic recombination revealed that the SIDICs could not fully propagate across the clones and either stop at the clone border or propagate for a few cell diameters (Fig. 2h and Supplementary Movie 7). In addition, the same sparkles that were observed in the knockdown experiments could be observed inside the mutant clones (Fig. 2h and Supplementary Movie 7). We conclude that the SIDICs cannot fully propagate through cells that have impaired gap junction function. Here we identified *InsP3R*, *SERCA* and the key gap junction component *Inx2* as SIDIC components. Future work will be necessary to uncover additional SIDIC modulators and fully reveal the propagation mechanism.

#### RNAi targeting key SIDIC prevents SIDIC formation *in vivo*.

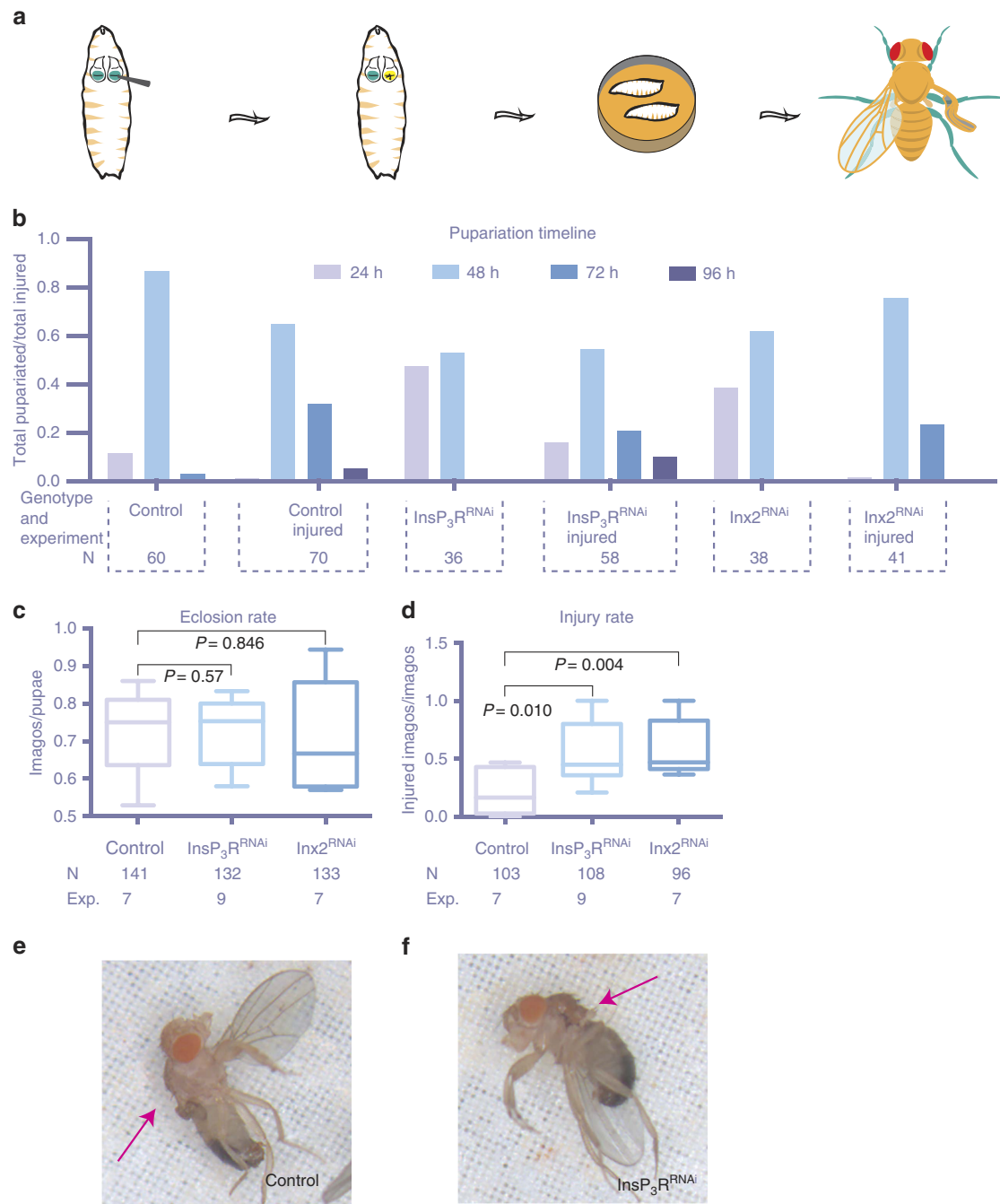
Having identified *InsP3R*, *SERCA* and *Inx2* as SIDIC components *ex vivo*, we asked whether the knockdown of these genes would prevent the formation of SIDICs *in vivo*. We stimulated SIDIC formation by striking the wing discs through the cuticle and monitoring them for a 30-min time window. As mentioned previously, striking wing discs *in vivo* triggers two

calcium-mediated reactions: a localized initial increase in intracellular calcium that does not propagate as a slow wave (dashed lines; Fig. 3) and later a SIDIC wave (Fig. 1d). All genotypes displayed the first type of reaction (in *SERCA<sup>RNAi</sup>* discs it seems delayed and of lower amplitude; Fig. 3c). However, although in control larvae 63% of stroked discs displayed a SIDIC wave during the time window of observation, we did not observe any SIDICs for *InsP3R<sup>RNAi</sup>* ( $n = 0/14$ ), *SERCA<sup>RNAi</sup>* ( $n = 0/7$ ) or *Inx2<sup>RNAi</sup>* ( $n = 0/9$ ) (Fig. 3a–d). Interestingly, the  $Ca^{2+}$  ‘sparkles’ that we observed *ex vivo* in *Inx2<sup>RNAi</sup>* wing discs could only be observed *in vivo* after mechanical stress (Supplementary Fig. 4B), revealing that these constitute a reaction to injury and culture, and not a basal change in gap junction function. The *in vivo* results confirm that the SIDICs observed *ex vivo* are a close proxy and adequate model of the *in vivo* phenomenon. Given the difficulty of accurately characterizing the SIDICs *in vivo*, an *ex vivo* model will be a useful tool to further identify required components and to continue dissecting the propagation mechanism of the SIDICs.

#### Knockdown of *InsP3R* and *Inx2* impairs recovery after injury.

We next asked what might be the function of the SIDICs during wing imaginal disc development. Previous studies have spotlighted the role of calcium signalling in wound healing and regeneration<sup>15,16,29–32</sup>. Further, our *in vivo* experiments had revealed that the SIDICs seemed to be a response to mechanical stress or injury.

Therefore, we wondered whether the ability of wing discs to recover after a mechanically induced injury was impaired in



**Figure 4 | The SIDICs components support wound healing of *Drosophila* wing imaginal discs. (a)** *In vivo* assay for recovery after mechanical injury. A blunt forceps tip is used to injure one disc per larva. The larvae that failed to heal eclose with heavily deformed or absent wings. **(b)** Pupariation timeline after mechanical injury. Control animals of all genotypes pupariate mainly during the first 48 h after a mock manipulation (no injury). Mechanical injury induced a delay in pupariation for all genotypes. Fly strains: *nub* > *GCamp6s* X *yw*, *nub* > *GCamp6s* X *UAS-InsP<sub>3</sub><sup>RNAi</sup>* (NIG 1063-R1), *nub* > *GCamp6s* X *UAS-Inx2<sup>RNAi</sup>* (BL 29306). **(c)** The eclosion rate after mechanical injury is not affected by RNAi lines against the key SIDICs components *InsP<sub>3</sub>* and *Inx2*: *yw* ( $0.72 \pm 0.11$ , 7 experiments,  $n = 103/141$  larvae); *InsP<sub>3</sub><sup>RNAi</sup>* ( $0.73 \pm 0.09$ , 8 experiments,  $n = 108/132$  larvae); *Inx2<sup>RNAi</sup>* ( $0.71 \pm 0.15$ , 7 experiments,  $n = 94/133$  larvae). ‘Whiskers’: min-max. *P*-values: Student’s *t*-test. Fly strains: *nub* > *GCamp6s* X *yw*, *nub* > *GCamp6s* X *UAS-InsP<sub>3</sub><sup>RNAi</sup>* (NIG 1063-R1 (6 exp.) or BL25937 (2 exp.)), *nub* > *GCamp6s* X *UAS-Inx2<sup>RNAi</sup>* (BL 29306). **(d)** Average injury rate after mechanical injury, per round of experiment. *InsP<sub>3</sub><sup>RNAi</sup>* and *Inx2<sup>RNAi</sup>* lead to a clear increase in the amount of injured animals: *yw* ( $0.22 \pm 0.19$ , 7 experiments, 103 animals); *InsP<sub>3</sub><sup>RNAi</sup>* ( $0.56 \pm 0.26$ , 9 experiments, 108 animals,  $P = 0.010$ ); *Inx2<sup>RNAi</sup>* ( $0.57 \pm 0.2$ , 7 experiments, 96 animals  $P = 0.004$ ). ‘Whiskers’: min-max. *P*-values: Student’s *t*-test. Fly strains: *nub* > *GCamp6s* X *yw*, *nub* > *GCamp6s* X *UAS-InsP<sub>3</sub><sup>RNAi</sup>* (NIG 1063-R1 (6 exp.)–BL25937 (2 exp.)), *nub* > *GCamp6s* X *UAS-Inx2<sup>RNAi</sup>* (BL 29306). **(e)** Control animal that failed to heal after mechanical wounding. It is noteworthy that the internal control wing is normal and can be easily differentiated from the injured wing. This picture is a close up view from Supplementary Fig. 6. Fly strains: *nub* > *GCamp6s* X *yw*. **(f)** *InsP<sub>3</sub><sup>RNAi</sup>* animal that failed to heal after mechanical wounding. It is noteworthy that the internal control wing is warped but can be easily differentiated from the injured wing. This picture is a close up view from Supplementary Fig. 6. Fly strains: *nub* > *GCamp6s* X *yw*, *nub* > *GCamp6s* X *UAS-InsP<sub>3</sub><sup>RNAi</sup>* (BL 25937).

RNAi backgrounds that abrogate SIDIC wave activity in the wing pouch. For this purpose, we developed an *in vivo* mechanical wounding assay for wing discs and scored for adults that eclosed with damaged wings. Here we damaged one imaginal wing disc—by punching it carefully through the cuticle under direct observation with a fluorescent compound microscope—and scored for imagos with deformed wings (Fig. 4a). It is noteworthy that both *InsP<sub>3</sub>R<sup>RNAi</sup>* and *Inx2<sup>RNAi</sup>* expression alone resulted in slightly deformed adult wings (Supplementary Fig. 5) (*SERCA<sup>RNAi</sup>* results in heavily deformed adult wings; hence, we excluded this genotype from the analysis; Supplementary Fig. 5). To compensate for this, we only injured one wing disc, whereas the other provided an internal control. Crucially, the initial increase in reporter activity after injury can be used to visually control that the disc has indeed been injured, and that only one disc has been struck. Injured wings that had not healed properly resulted in heavily deformed stumps and could be unambiguously identified (Fig. 4e,f and Supplementary Fig. 6). Importantly, both wounding and scoring were performed ‘blind’ such that the experimenter ignored the genotypes of the animals being wounded/analysed.

*Drosophila* larvae delay their development after an injury, to allow the wounded organ to repair itself before pupariating<sup>33,34</sup>. Consistent with this, an analysis of the pupariation timeline following mechanical wounding revealed the expected delay in development relative to non-injured animals (Fig. 4b). We did not record a clear difference in this delay between *InsP<sub>3</sub>R<sup>RNAi</sup>* or *Inx2<sup>RNAi</sup>* and control animals. Further, the eclosion rate (number of flies/number of pupae) of control and knockdown groups was very similar (Fig. 4c), indicating that the degree of injury inflicted on each group had indeed been approximately equal.

Finally, we monitored the ratio of animals with a damaged wing (Fig. 4e,f). Evaluated over multiple rounds of experiments (seven to nine), the average ratio of injured animals per experiment was 0.216 ( $\pm 0.185$ ,  $n = 103$ , 7 experiments) for control animals, 0.556 ( $\pm 0.263$ ,  $n = 108$ , 9 experiments,  $P = 0.010$ ) for *InsP<sub>3</sub>R<sup>RNAi</sup>* and 0.573 ( $\pm 0.196$ ,  $n = 96$ , 7 experiments,  $P = 0.004$ ) for *Inx2<sup>RNAi</sup>* (Fig. 4d). These results indicate that in the absence of SIDIC activity, wound healing and regeneration can proceed; however, the rate at which they fail increases greatly.

The SIDICs are induced by mechanical stress *in vivo* and the knockdown of two components required for SIDIC formation and propagation affects the ability of the wing disc to heal after an injury. Based on these results, we infer that the SIDICs probably constitute a response to mechanical stress that contributes to the recovery of the wing imaginal disc after injury. However, we cannot exclude that *InsP<sub>3</sub>R* and *Inx2* have cell-autonomous (non-SIDIC related) functions during wound healing, and that their knockdown could negatively synergize with mechanical injury to perturb wound healing.

## Discussion

In this study, we describe the occurrence of slow, long-range ICWs in imaginal discs (SIDICs), *in vivo* and *ex vivo*. We identify the *InsP<sub>3</sub>R*, *SERCA* and *Inx2* as necessary components for SIDICs generation and propagation. Finally, we found that the SIDICs constitute a response to mechanical stress, probably supporting the wound healing and regeneration.

In comparison with previously reported calcium signals, such as the calcium flashes observed in the embryo<sup>15</sup> or the calcium transients in the pupal notum<sup>16</sup>, the SIDICs are different in several aspects: the propagation speed, the duration of the phenomenon and the latency between the source of stress and the generation of the wave. The dissimilarities in the type of calcium signal observed in wing discs and the embryo and pupal notum

could reflect inherent differences in the composition of the tissues studied. Alternatively, the SIDICs and the calcium flashes may be encoding different types of information that fulfill different purposes during wound healing and regeneration. In the embryo, the calcium flashes recruit hemocytes<sup>15</sup>, whereas in the pupal notum they coordinate cell contraction<sup>16</sup>. To further study the function of the SIDICs *in vivo*, a more sophisticated *in vivo* imaging setup will need to be developed. The *ex vivo* setup, however, proves to be a valuable substitute.

The wing disc has been an exquisite model for developmental genetics. Our work expands the utility of this model by revealing that it can be employed for the study of calcium signalling and ICWs. Wound healing and regeneration probably require a complex interplay between developmental pathways and the ability to coordinate cells during morphogenetic movements. Interestingly, calcium signalling has been proposed to be at the nexus of many signaling pathways<sup>1–3</sup> and this nexus function could be essential for its role during regeneration. It will be interesting to see whether the SIDICs link, and perhaps help to orchestrate, different signalling pathways and morphogenetic movements during wound healing and regeneration of *Drosophila* imaginal wing discs.

## Methods

**Ex vivo imaging.** *Ex vivo* imaging chambers were assembled with a life cell imaging dish (Zell-Kontakt) and a Millicell standing insert (PICMO1250) that we modified by removing its feet with a scalpel. Briefly, the wing discs were placed at the centre of the imaging dish, apical side facing down, in 20  $\mu$ l of culture medium. Next, the modified insert was gently placed on top of the discs, thereby trapping the discs under the membrane. Finally, 200  $\mu$ l of culture medium were added inside the insert. It is noteworthy that we did not employ an alginate gel in this study as previously described<sup>17</sup>. We used WM1 as culture medium: Schneider’s medium (Sigma), 6.2  $\mu$ g ml<sup>-1</sup> bovine insulin (Sigma) and 5% Fly extract (home made)<sup>17</sup>. Time-lapse recording was performed on a Zeiss-Andor Spinning disc microscope equipped with an Ixon3 camera and a  $\times 25$  Zeiss Neofluor water immersion objective. GCamp6s was excited with a 488 laser. Imaging was performed in a dark room at 21 °C. We acquired three-dimensional time-lapses with the IQ2 software. Z-stacks were acquired every 10 s with 15% laser power, a gain of 50 and an exposure of 60 ms.

**In vivo imaging of squeezed larvae.** Larvae were glued dorsally unto double-sided adhesive tape on a microscope slide. Imaging was performed with a Zeiss Axiovert microscope using a  $\times 10$  air objective under the control of ZEN (Zeiss). We used a Zeiss MRm camera with a 200 ms exposure and acquired images every 330 ms.

**In vivo imaging of immobilized larvae.** Handmade imaging chambers were constructed as shown in Fig. 1. Larvae were glued ventrally on a microscope slide on double-sided adhesive tape. Two ‘chamber walls’ consisting of three layers of sticky tape were deposited adjacent to the larvae. Finally, a coverslip was added on top of the chamber. This setup enables one to gently flatten the larvae to better position the wing discs perpendicular to the objective and provide a source of mechanical stimulation. Time-lapse imaging was performed on an AxioPlan 2 microscope equipped with an Axiocam HRC under the control of Axiovision 4.7. Exposure was set to 100 ms and the time interval to 6 s.

**Staging.** We staged larvae by restricting egg deposition to ~8–12 h. Before conducting experiments, larvae were sorted visually under a binocular to refine the staging according to the following criteria: size, length, body colour and appearance of the spiracles.

**RNAi experiments.** Crosses were kept at 25 °C and shifted to 29 °C, 24 h after egg deposition, to increase Gal4 and RNAi activity.

**Inx2<sup>G0157</sup> clone induction.** *w, inx2<sup>G0157</sup>, FRT19A/FM7c*; were crossed to *ubi > mRFPNLS,w,hsflp,FRT19; nubbin-Gal4,GCamp6s; MKRS/Tm6b*, maintained at 25 °C and passed every 24 h. Clone induction was done 48 h after egg deposition at 37 °C for 30 min. *inx2<sup>G0157</sup>, FRT19A/ubi > mRFPNLS,w,hsFLP,FRT19; nubbin-Gal4,GCamp6s; +/Tm6b* roaming larvae were selected with a fluorescent binocular microscope.

All *Drosophila* strains used are listed in the Supplementary Methods.

**Statistics.** All results were tested for normality (D'agostino and Pearson Omnibus normality test), to define which statistical test to perform. However, as normality can be difficult to judge in small sample sizes, we performed Student's *t*-tests and Mann-Whitney *U*-tests for all comparisons. All the results presented in this study were significant at a 95% threshold for both tests. We show *t*-tests in this study, as all our samples passed the normality test. All tests were performed with the software package PRISM from Graph Pad and Excel from Microsoft. Sample sizes were not determined before performing the experiments. No sample exclusion criteria were employed. For mechanical wounding assay the following blinding strategy was followed: genotypes were replaced by a numeric code unknown to the experimenter and changed for each experiment.

**Mechanical wounding assay.** Four-day-old larvae of the correct genotype were collected with a sieve, rinsed with water and selected for GCaMP6s basal fluorescence with a binocular microscope. Next, the larvae were dried with a paper towel and glued, ventrally, to a microscope slide covered with double-sided adhesive tape. A blunt forceps was then used to tap the wing discs without piercing the cuticle, while monitoring GCaMP6s fluorescence to detect the injury. Afterwards, the larvae were detached from the adhesive tape with a drop of water and transferred to an apple agar petri dish. Recovering larvae were then placed in a fly incubator at 25 °C. At due time, freshly eclosed flies were anaesthetized with CO<sub>2</sub> on a fly pad and scored for injured wings.

**Time-lapse processing.** First, the original files were transformed into three-dimensional time lapses with the stack to hyperstack function in Fiji (<http://fiji.sc/Fiji>). Second, the hyperstacks were subject to a maximum intensity projection to generate two-dimensional time lapses. All calculations were performed on these two-dimensional time lapses.

**Cellular calcium pulse duration calculation.** For each sample, we selected a region of interest (ROI) of approximately the size of a cell, randomly, in the trajectory of a wave. We used the ROI Intensity Evolution plugin of ICY (<http://icy.bioimageanalysis.org/>)<sup>35</sup> to produce an excel file of the average fluorescence intensity per time frame of the ROI. Finally, we used Excel to calculate the duration of an average cell pulse.

**Wave magnitude calculation.** The area of the pouch was measured in ICY<sup>35</sup> by employing the baseline fluorescence of GCaMP6s and morphological landmarks. The magnitude of a wave was calculated in the following way. First, the time lapse was cropped in time to include only one wave or 10 min. The 10-min crops were required for genotypes in which wave activity was diminished. Finally, the area covered by the wave was calculated by performing a maximum intensity projection over time of the 10-min per one wave time lapses with the Intensity Projection plugin in ICY<sup>35</sup>. The wave magnitude was calculated as wave area per pouch area.

**Wavefront speed calculation.** We calculated the wavefront speed manually by defining two semi-parallel lines in (superimposed on the time lapses and perpendicular to the wavefront) in ICY<sup>35</sup> and measuring the time required by the wavefront to cross the distance between the two lines.

**Wave period calculation.** For each time lapse, we generated a ROI outlining the pouch by employing the baseline fluorescence of GCaMP6s and morphological landmarks. Next, we used the ROI Intensity Evolution plugin of ICY<sup>35</sup> to produce an Excel file of the average fluorescence intensity of the ROI per time frame. Finally, we used Excel to calculate the average interval in between peaks of fluorescence for the ROI.

**Data availability.** The authors declare that all data supporting the findings of this study are available within the article and its Supplementary Information files or from the corresponding author upon reasonable request.

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### Author contributions

S.R. designed and performed the experiments, and analysed the data. S.R. and K.B. wrote the manuscript. S.R. produced the figures and illustrations.

### Additional information

**Supplementary Information** accompanies this paper at <http://www.nature.com/naturecommunications>

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