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REVIEW ARTICLE

Expanding the Arsenal Against Huntington's Disease-Herbal Drugs and Their Nanoformulations

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DOI: 10.2174/1570159X18666201109090824 Abstract: Huntington's disease (HD) is an autosomal fatal genetic disease in which degeneration of neuronal cells occurs in the central nervous system (CNS). Commonly used therapeutics are clude-monoamine depletors, antipsychotics, antidepressants, and tranquilizers. However, these drugs cannot prevent the psychotic, cognitive, and behavioral dysfunctions associated with HD. In addition to this, their chronic use is limited by their long-term side effects. Herbal drugs offer a plausible alternative to this and have shown substantial therapeutic effects against HD. Moreover, their safety profile is better in terms of side effects. However, due to limited drug solubility and permeability to reach the target site, herbal drugs have not been able to reach the stage of clinical exploration. In recent years, the paradigm of research has been shifted towards the development of herbal drugs based nanoformulations that can enhance their bioavailability and blood-brain barrier permeability. The present review covers the pathophysiology of HD, available biomarkers, phytomedicines explored against HD, ongoing clinical trials on herbal drugs exclusively for treating HD and their nanocarriers, along with their potential neuroprotective effects.

Keywords: Huntington's disorder, oxidative stress, herbal medicine, neuroprotective effects, blood-brain barrier, nanocarriers.

1. INTRODUCTION

The term HD was coined by Ohio based physician George Huntington in 1872, who described this disease for the first time [1]. HD is an autosomal fatal genetic disorder which is a progressive, genetically programmed ND that leads to depletion of psychological, cognitive, and motor functions. As per Huntington's disease Society of America (HDSA), HD patients show symptoms similar to those of Alzheimer's disease (AD), Parkinson's disease (PD), and amyotrophic lateral sclerosis (ALS) [2-7]. The symptoms associated with HD are "chorea" (abnormal autonomic movements), loss of rational abilities, and psychological disturbances. This abnormality occurs due to the mutation of the Huntingtin genes. Healthy neurons contain 6-35 repeats of units of cytosine-adenine-guanine (CAG) trinucleotide, while accumulation of mutant Huntingtin (mHTT) genes changes the translation process (more than 36 CAG repeats) [8, 9]. This process may lead to neuronal cell death and

cause degeneration of neurotransmitters within the central nervous system (CNS) [10]. After the first appearance of symptoms in an affected person, death usually occurs within 15 to 20 years [11]. Various biochemical alterations such as downregulation of γ -aminobutyric acid (GABA) and acetyl-choline (ACh), along with a decrease in their production enzymes, glutamate decarboxylase (GAD) and choline-acetyl transferase (CAT), respectively are seen in patients with HD [11-13].

Globally, 5 to 8 people in a population of 0.1 million are diagnosed with HD [14, 15]. The disease is reported to be more prevalent in Europe as compared to that in the USA, China, and India. A number of patients diagnosed with HD are extrapolated to increase from 58,176 in 2019 to 60,743 in 2024 [14].

2. ETIOPATHOGENESIS

2.1. Neuropsychiatric Disturbance

There is a broad range of HD neuropsychiatric symptoms, involving irritation, obsessive- compulsive behavior, depression, psychosis, and apathy. Prior to the knowledge of HD, this disease was categorized under psychiatric disorder because its symptoms were similar to psychiatric diseases. Later on, based on mechanistic studies, it was understood

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that the pathology of HD is associated with neurodegeneration in the brain [16, 17]. Irritation, depression, and apathy are neuropsychiatric symptoms that continuously manifest and get advanced with the progress of the disease [18].

2.2. Neurodegeneration

NDs can be classified by extrapyramidal and pyramidal motor disturbances that can lead to cognitive or behavioral changes in the body [19]. Neurodegeneration is a process that involves the degeneration of neurons due to aging of the brain or the influence of pathological factors that can damage the neurons. It has been seen that the loss of neurons in the brain is one of the significant health hazards. Cerebral malfunctioning occurs due to various NDs like AD, PD, HD, ALS, and multiple sclerosis [20-22]. Moreover, activation of excitatory neurotransmitters receptors such as N-methyl-Daspartate (NMDA) and α-amino-3-hydroxy-5-methyl-4isoxazolepropionic acid (AMPA) is also a leading cause of excitotoxicity and apoptosis of the neurons. These excitatory receptors can lead to excitotoxicity as observed in chronic NDs such as PD and HD. AMPA and NMDA receptors limit the neuronal entry of calcium ions by regulating calcium ions-permeability in the brain and CNS [23]. The factors that can cause degeneration of neurons are shown in Fig. (1).

2.3. Genetic Factors

mHTT genes work at the molecular level of cells. They are located in chromosome 4p16.3, 67 exons, and 3144 amino acids. Healthy human genes contain 5 to 35 CAG triplet

genes in rRNA exons. mHTTs protein causes a genetic mutation in cells and changes the translation process. Hence, CAG repeat increases from 36 to 121. A number of repeats of CAG depend on the age of onset of the disease [24-26].

2.4. Mitochondrial Dysfunction

Mitochondria play a crucial role in storing maximal bioenergy, adenosine triphosphate (ATP) in the body (eukaryotic cells). They regulate intracellular calcium homeostasis, which can lead to diminishment in the production of free radicals in the endoplasmic reticulum and reduces the apoptosis process. Indeed, mitochondrial dysfunction has been affected by an earlier pathological manifestation of HD. In HD, an increase in the level of polyglutamate occurs in the striatum and cerebral cortex parts of the brain. The mHTT protein is known to cause mitochondrial dysfunction in Huntington's patients. This mHTT protein binds with mitochondrial transporter II receptors and causes oxidative damage as well as mitochondrial dysfunction. The dysfunction of mitochondria results in lower intake of glucose metabolism and mitochondrial oxidation in cerebrospinal fluid (CSF), which has been clearly seen in post-mortem reports of brains of HD's patients [27-30]. It also increases lactate levels in both the CSF and cerebral cortical tissue [31, 32]. Deregulation of mitochondrial function by 3-nitropropionic acid (3-NP) mitochondrial toxin has been observed in various studies in which metabolic impairment occurred due to deficiency of energy, excitotoxicity, and oxidative stress (OS) [33-36].



Fig. (1). Factors that cause degeneration of neurons in HD. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

2.5. Oxidative Stress (OS)

The mechanism of OS in HD is still not clear. Some molecular hypotheses state that increase in the level of ROS. lipid peroxidation, and chromosomal mutation can be major factors for disease manifestation [37, 38]. Impairment in the electron transport chain, oxidative damage, and mitochondrial dysfunction can increase OS. 8-Hydroxy-2-deoxygonosine $(OH^{8}dG)$ is a biomarker of oxidative damage in DNA. As reported by Bogdanovet et al. (2001), the enhanced concentration of OH⁸dG may increase oxidative damage [39]. Accumulation of mHTT protein has been observed in HD's patients; hence, it may be implicated in the increase of OS [40, 41]. Increased concentrations of free radicals can predispose to excitotoxicity that can cause impairment of the mitochondrial functions, energy production, and metabolic inhibition [42, 43]. These mechanisms of OS are presented in Fig. (2).

3. BIOMARKERS INVOLVED IN THE PATHO-GENESIS

Biomarkers play an important role in evaluating and measuring pathogenic as well as biological processes and pharmacological responses against HD. The ideal biomarker must be reliable, accurate, and specific. For understanding new clinical strategies, understanding the biomarkers of a disease is very important. In order to assess the treatment response and monitor the progression of the disease, the Unified Huntington's Disease Rating Scale is currently in use [11].

Based on the method of identification, biomarkers of HD are divided into three categories. These include clinical, biofluid, and imaging biomarkers. Clinical biomarkers are used to measure the motor, cognitive and psychotic abnormalities related to HD. Neurotransmitters, microglial toxins, and mHTT protein are listed under the category of bio-fluid biomarkers. Various techniques that are used to quantify their levels in blood and CSF include high-performance liquid chromatography (HPLC), mass spectrometry(MS), timeresolved fluorescence energy transfer (TR-FRET), homogeneous time-resolved fluorescence (HTRF), and enzymelinked immune sorbent assay (ELISA) [45]. Imaging biomarkers are used for the detection of structural changes in brain with the help of imaging techniques such as MRI and [18F] MNI-659 PET [46, 47]. Various applications of these biomarkers are listed in Table 1.



Fig. (2). Pathophysiological mediators that are responsible for OS and HD. Mitochondrial dysfunction: Mitochondria are widely known as the powerhouse of cells as they generate energy in the form of adenosine triphosphate (ATP). mHTT genes bind with transporter II in mitochondria and cause mDNA damage and bioenergy failure. mHTT proteins increase the influx of Ca^{2+} in cytoplasm in mitochondria which leads to excitotoxicity and bioenergy failure, and ATP formation reduces. As a result of this, mitochondrial dysfunction and the generation of ROS take place. Neuro-inflammation: Microglia and astrocytes in the presence of ATP chemokines activate Toll-like receptor (TLR) and m1 receptor protein inflammation cytokines, which, in turn, increase the intracellular Ca^{2+} entry and ROS levels. M1 receptor protein inflammation cytokines also increase inflammatory mediators (IL6, TNF α) and OS, which give rise to neuroinflammation and degeneration of the neuronal cell. Accumulation of mHTT genes: The normal base DNA pair contains 5-35 repeated units of CAG chain in exon 1 cytoplasm. When alteration in base DNA pair occurs, mHTT genes bind with exon 1 and increase the CAG units from 36 to 121, which is responsible for OS. This leads to the misfolding of mHtt and the formation of their aggregates in neuronal nuclei and neuropils in the brains of HD patients. This misfolded mHtt exerts its neurotoxicity by disturbing a wide range of cellular functions due to its interaction with a variety of proteins, thus interrupting their function [44]. Increase ROS: Due to mHTT gene, the intracellular influx of Ca2+ increases. This process can enhance excitotoxicity and cause oxidative damage and OS. **OS:** The factors like accumulation of mHTT genes, neuroinflammation, high lipid concentration, and mitochondrial dysfunction can increase OS, and that is responsible for the progression of the disease. (*A higher resolution / colour version of this figure is available in the electronic copy of the article*).

Table 1. Biomarker of HD.

S. No.	Biomarkers	Mediator	Molecule	Sample	Methods	Comments	Refs.
1.	Clinical	Motor			Anti-saccade error rate	Understanding genetic and environmental factor for disease	[48]
	-	-			Digitomotography	Assessment of quantitative motor by finger tapping.	[46, 49]
		Cognitive	-	-	SDMT	-	[46]
2.	Bio-fluid	Immune system	IL-6,IL-8, IL-1β	CSF, Blood	MSD immunoassay, ELISA	Leukotriene inflammatory mediator activate NFkB and cause neuroinflammation	[50]
	-	-	TNF-α	CSF	MSD antibody- based tetraplex array	Tumour necrosis factor α inflammatory mediator activate NF κ B and cause neuroin- flammation	[51]
	-	Genetic HTT mutation	HTT Protein	Blood	TR-FRET, HTRF, ELISA	-	[45]
	-	-	mHTT protein	CSF, Blood	IP-FCM, ELISA, HTRF	mHTT protein can increase OS.	[52]
	-	Microglial markers	YKL-40, MCP1, Chi- totriosidase	CSF	ELISA	-	[53]
	-	Microglial toxins	3-HK, QUIN, ROS	-	-	-	-
	-	Neurodegeneration	neurofilament light (NfL)	CSF, Blood	ELISA	Analyse premanifest and manifest Huntington's pateints	[54]
	-	-	GABA	CSF	Radioreceptor assay, Ion-exchange fluorometry	Diminution of inhibitory neurotransmitter GABA	
	-	-	-		Ion-exchange chromatography, High resolution proton NMR spectroscopy and HPLC	-	[55]
	-	-	Choline	CSF	Radiochemical micro-method	-	[56]
	-	-	Dopamine	CSF	-	-	-
	-	Transglutaminase	Nε-(γ-l-glutamyl)-l-lysine (GGEL)	CSF	MS	-	[57]
	-	-	γ-glutamylspermidine, γ-glutamylputrescine, bis- γ glutamylputrescine	γ-glutamylspermidine, - γ-glutamylputrescine, bis- CSF γ glutamylputrescine -		-	[58]
3.	Imaging	Structural loss			MRI	Neurodegeneration seen in the brain	[46]
	-	PDE10 uptake			[18F]MNI-659 PET	-	[47]

4. TREATMENT STRATEGIES

Till now, none of the available drugs has been able to show complete relief in symptoms of the disease. Tetrabenazine, however, has been reported to show the most significant response in terms of reducing symptoms of motor abnormality (chorea) [59]. A combination of antipsychotic, anti-depressant, and anti-AD medicines is reported to reduce cognitive, psychotic, and motor abnormalities [60-62]. Tetrabenazine is a monoamine enzyme inhibitor that prevents the loss of adrenergic neurotransmitters in the synapse. It has been found to be useful in the treatment of hyperkinetic movement disorder. The major side effects of this drug are depression, exacerbation of depression, akathisia, restlessness, and psychotic problems [59]. Haloperidol is an an-

S. No.	Drugs	Animal	Dose (mg/kg)	Duration of Study	Side effect	Mechanism of Action	Results	Refs.
1.	Tetramethylpyrazine	Male Wistar rats	40 and 80	21 days	-	Reduce 3-NP neurotoxin	Effective against 3-NP induce HD model	[67]
2.	Rivastigmine	Male Wistar rats	0.5, 1,2	15 days	Drowsiness, loss of appe- tite/weight loss, diarrhea, weakness, dizziness	AChE inhibitor	Improved cognitive function	[61]
3.	Galantamine	Female Wistar rats	3.75, 7.5	21 days	Drowsiness, dizziness, loss of appetite, and weight loss	AChE inhibitor	Reduction in oxidative stress, Neuroprotective effect against 3-NP induced neuro- toxicity.	[62]
4.	Amantadine	Male Wistar rats	10, 40		Blurred vision, nausea, and loss of appetite, dry mouth, constipation, or trouble sleep- ing, leg swelling and skin discoloration	NMDA gluta- mate antagonist	Amantadine binds with NMDA receptor and in- creases dopamine in postsyn- aptic receptors and helps to improve neurological and psychological conditions associated in NDs.	[68]
5.	Haloperidol	Male Lister Hooded rats	1.5	112 day	dry mouth, constipation, sedation, tardive dyskinesia, parkinsonism, depression, extrapyramidal symptoms, neuroleptic malignant syn- drome	decrease dopa- mine	Used in the treatment of chronic neuroleptics and reduce locomotors activity in the brain	[63]
6.	Leveteracetam	Human	3,000 mg/day		Infection, asthenia, neurosis, drowsiness, headache, naso- pharyngitis, nervousness, abnormal behavior, agitation, anxiety, apathy.	Neuroprotective effect	Dose of 3,000 mg/day for 48 hour reduced the symptoms of chorea in HD	[69]
7.	Terabenzine	Human	50 mg/day		Drowsiness, sedated state, muscle rigidity, depersonal- ization depression, exacerba- tion of depression, akathisia, and restlessness.	Inhibition of MAO enzyme	Dose of 50 mg/day useful in treatment of hyperkinetic movement disorder	[59]

Table 2. Preclinical studies of synthetic drugs are reported for the treatment of HD.

tipsychotic drug that causes inhibition of dopamine in the limbic system of the brain and reduces psychotic symptoms of Huntington patients [63]. The side effects of this drug are suppression of movement, mood changes, breast enlargement, irregular menstrual periods, and loss of interest in sex. Acetylcholinesterase (AChE) enzyme inhibitors such as rivastigmine and galantamine can enhance the ACh levels in the brain and improve cognitive function [62]. These drugs are reported to show side effects such as dizziness, drowsiness, loss of appetite, and weight loss. The drugs that have been explored to treat HD in animal studies are listed in Table 2. Apart from the pharmacological approach, some nonpharmacological approaches such as psychotherapy, speech therapy, physical therapy and occupational therapy have also shown beneficial effects for the treatment of the disease. A combination of pharmacological and non-pharmacological therapy has been reported to work in a better way as compared to the use of a single modality [64]. Some in vitro cell line studies showed that the use of stem cells is able to reduce the degeneration of neurons by reducing CAG sequencing in genetic bases. The list of such studies is given in Table **3**. To date, a number of clinical and preclinical studies have been conducted on different drugs, but none of them has shown complete treatment of HD. Herbal drugs offer the third treatment strategy for HD. These have been reported to possess a better safety profile and are easily available as compared to synthetic drugs [65]. The use of herbal drugs in traditional medicines has shown neuroprotective effects in NDs [66]. Their antioxidant, anti-inflammatory, anti-apoptosis, and AChE enzyme inhibition have been reported to be responsible forthe treatment of cognitive, psychotic, and motor dysfunctions associated with HD [11].

5. NEUROPROTECTIVE HERBS

Herbal medicines contain complex mixtures of phytoconstituents and organic chemicals, including alkaloids, fatty acids, sterols, flavonoids, glycosides, saponins, terpenes, *etc*. Phytoconstituents present in some of the herbal drugs have shown pharmacological efficacy in reducing the symptoms

Table 3. Cell line studies of HD.

Disease	Cell line	Test	Results	Refs.
HD	LUMCi007-A, LUMCi007-B, LUMCi008-A, LUMCi008-B, LUMCi008C		Reducing CAG repeats	[70]
	ICGi018-A (iHD38Q-3) DNA fragment anal PCR-product		<i>In vitro</i> cell line studies reduce CAG18 and 38 repeats by PBMCs and iPSC line	[71]
	CSSi006-A (3681)	Sequencing	Reducing CAG repeats in fibroblasts (17 \pm 2 and 46 \pm 3 CAG repeats)	[72]
	CSSi004-A (2962)	Sequencing	Reducing CAG repeats in fibroblasts (17 \pm 1 and 43 \pm 2 CAG repeats)	[73]
	Genea090 human embryonic stem cell line	Sterility	The cell line is tested and found negative for Mycoplasma and any visible contamination	[74]
	Genea017 human embryonic stem cell line	Sterility	The cell line is tested and found negative for Mycoplasma and any visible contamination. <i>Mycoplasma</i> and any visible contamination	
	Herbal formula B401		Neuroprotective and angiogenesis effects in R6/2 mouse model of HD	[75]
	CurcuminSolid lipid nanoparticles (SLNs) (C-SLNs)	SDH Staining, Mitochondrial Oxidative Stress Parameters	Reduce ROS, mitochondrial dysfunction and lipid preroxidation.	[76]

of HD. In India, traditional herbal plants have been used for a number of diseases afflicting the nervous system. VataVyadh is a Sanskrit word that means disease related to the nervous system. Vata represents energy around the body, and disturbance of this process is called as VataVvadh. It ultimately leads to weakness, hypersensitivity, dementia, and chorea [77]. Certain herbal drugs possess phytoconstituents that enhance ACh levels in post synaptic neurons by inhibiting AChE in synapse and enhancing cognitive function [78, 79]. The *in vivo* studies have shown that certain herbal drugs and their phytochemicals exhibit a significant response against 3-NP neurotoxin. Several other pathways are also crucial in HD. Based on the concept of multi targets, network pharmacology-based analysis is employed to find out related proteins in disease networks. The network targeting method aims to find out the related mechanism of efficacious substances in a rational design way. Traditional Chinese medicine (TCM) prescriptions would be used for research and development against HD [80]. Virtual screening is performed to obtain drug molecules with high binding capacity from TCM. Mechanism of action and beneficial effects of herbal drugs are shown in Fig. (3).

5.1. Acorus Calamus (AC)

AC, also known as the Sweet flag, belongs to *Araceae* family. It acts as a brain and nervous system rejuvenator with beneficial memory-enhancing properties. It also improves learning efficiency, and reduces behavioral alteration. The major constituents of the plant are α -and β -asarone. β -Asarone has the ability to suppress beta-amyloid-induced neuronal apoptosis in the hippocampus through reversal down-regulation of Bcl-2, Bcl-w, caspase-3 activation, and phosphorylation of c-Jun terminal kinase (JNK) [81]. It has the potential to enhance dopaminergic nerve function. Therefore, it can play a key role in PD by increasing the amount of striatal extracellular dopamine and the expression of tyrosine hydroxylase in substantia nigra. It also improves the expression of DJ-1 genes in the striatum and thus acts as PD neuro-

protective [82]. The treatment of PD using AC indicates its neuroprotective action; hence, it could be used for the treatment of HD.

5.2. Allium Sativum (AS)

AS is one of the most widely researched herbs found in the ancient medical literature [83, 84]. AS belongs to the family Amaryllidaceae. The main bioactive compounds of AS are allicin (allyl2-propene thiosulfinate or diallylthiosulfinate) and alliin. S-allyl cysteine (SAC) is the major component of the extensively studied aged garlic extract (AGE) [85, 86]. SAC exerts antioxidant activity, both directly and indirectly. It also decreases protein oxidation and nitration. In addition to this, it is reported to reduce lipid peroxidation and DNA fragmentation. Dopamine levels, oxidative damage, and lipid peroxidation in 1-methyl-4-phenyl pyridinium and 6-hydroxydopamine (6-OHDA) models of PD were found to be downregulated by SAC. It decreased lipid peroxidation and mitochondrial dysfunction in 3-nitro propionic acid and quinolinic acid animal models of HD. It also increased the dismutase activity of manganese and superoxide copper/zinc and prevented changes in behavior. AGE activates the expression of significant genes required for neuronal survival, both directly and indirectly [87, 88].

5.3. Bacopa Monnieri (BM)

BM or *Herpestis monniera*, commonly referred to as Brahmi, belongs to the family *scrophulariaceae*. It is found throughout the Indian subcontinent and is categorized in Ayurveda as *Medhya Rasayana* [89-92]. It is used to treat epilepsy, insomnia, anxiety and is a memory enhancer [93, 94]. The significant chemical components present in the plant are tri-terpenoid saponins like dammarane, bacosides A and B [90, 95]. In addition to these significant components, it also contains other saponins, including bacopa saponin A-G [96-98], along with pseudo-jujubogenin, jujubogenin [99], bacopaside I-V, X, and N1 and N2 [100-102]. Brahmine, herpestine, and monnierin are also present in the plant



Fig. (3). Phytoconstituents and their target of HD. Note: Green boxes indicate the herbal drugs used to inhibit various molecular pathways and blue indicate the molecular targets (biomarkers). (A higher resolution / colour version of this figure is available in the electronic copy of the article).

[103, 104]. The potential of this plant in improving memory has been well reported [94, 105-107]. Bacoside A has been shown to be the main constituent to enhance memory [90, 108]. In clinical trials, BM showed significant dose- dependent memory enhancement activity [109]. BM also acts as a metal ion chelator [110], scavenges free radicals, and shows the antioxidant property in the body [111, 112]. The neuroprotective and memory improving potential of BM's extracts have also been reported. It exhibits antioxidant [112], antistress [113], antidepressant [114], anxiolytic [115], free radical scavenging [111], hepatoprotective [116] and antiulcerogenic activity [117]. 3-NP inactivates the succinate dehydrogenase cell enzyme (SDH) and the electron transport chain complex II-III [118, 119]. It also reduces ROS, malondialdehyde (MDA), and free fatty acid levels [120]. The oral intake of BM's leaf powder is reported to reduce basal concentrations of several oxidative markers and improve thiol-related antioxidant molecules, and antioxidant enzyme activity suggesting its significant antioxidant potential. Dietary BM's supplements were reported to lead to substantial protection against oxidative damage caused by neurotoxins in the brain [93]. BM has been found to be helpful in HD's therapy owing to its protective impact against stressmediated neuronal dysfunctions also [112].

5.4. Centella Asiatica (CA)

CA, also known as *Hydrocotyle asiatica*, *Gotu kola*, Indian Pennywort, and *Jal brahmi* belongs to the family *Umbelliferae*. CA has exhibited many neuropharmacological effects that include memory enhancement [121, 122], increased neurite elongation, and nerve regeneration acceleration [123]. It has also been reported for its anti-oxidant properties [124, 125]. Triterpenoid saponins, including asiaticoside, Asian acid, madecassoside, and madecassic acid, are the most significant chemical constituents of CA [126, 127]. Other minor saponins present in CA are brahmoside and brahminoside [126, 128]. Various acids that are present in the plant are triterpene acids, betullic acid, brahmic acid, and isobrahmic acid [126, 128]. The essential oils that are present in plant leaves include monoterpenes such as bornyl acetate, α -pinene, β -pinene, and π -pinene [129]. In addition to these constituents, CA is also reported to contain flavones, sterols, and lipids. Attenuation of 3-NP-induced depletion of GSH, total thiols, and endogenous antioxidants level by CA has been reported in the striatum and other brain regions [130]. It also displayed protection against 3-NP-induced mitochondrial dysfunctions, viz., reduced SDH activity, enzymes in the electron transport chain, and reduced mitochondrial viability [130].

5.5. Coriandrum Sativum (CS)

CS, commonly known as coriander, belongs to Apiaceae family. It contains a number of flavonoids. The major phytoconstituents include glucoronides such as quercetin and polyphenols such as caffeic acid, protocatechinic acid, and glycitin. The flavonoid content of the plants is reported to be equivalent to 12.6 quercetin equivalents per gram, while polyphenolic content is equivalent to 12.2 gallic acid equivalents per gram [131, 132]. A study showed that the CS's extract enhanced concentrations of superoxide dismutase (SOD), glutathione, CAT, and total protein in the animal model. It also reduced the levels of cerebral infarction, lipid peroxidation (LPO), and calcium in the rats [133]. Scopolamine and diazepam-induced memory deficits were found to be reversed by leaf extracts of CS. It reduced reactive modifications in brain histology such as gliosis, lymphocytic infiltration, and cellular edema. It showed protective function in the states of cerebrovascular insufficiency. The leaves also demonstrated antioxidant properties in terms of free radical scavenging activity by 2, 2-diphenyl-1-picrylhydrazyl and lipoxygenase inhibition [134-136].

5.6. Curcuma Longa (CL)

The common name used for the CL is turmeric. It is a perennial herb and belongs to the family Zingiberaceae. It is used throughout the world, mainly in China, Japan, and India, as a pharmacotherapeutic [137]. It has a long history of use as a spice and household remedy to treat inflammation, skin diseases, wounds, as well as antibacterial and antiseptic agent [138]. CL contains different curcuminoids, sesquiterpenes, essential oil, and starch. Most of the curcuminoids are diarylheptanoid, curcumin being the most prevalent. Desmethoxycurcumin and bis-desmethoxycurcumin are the other two curcuminoids [138, 139]. CL shows a number of pharmacological actions such as antioxidant [140], antiinflammatory [141], choleretic, hepatoprotective, analgesic, antifungal, free radical scavenging, antiparasitic, antiviral, antibacterial [138, 142], and anti-mutagenic [143]. The antioxidant properties of turmeric are attributed to its direct scavenging of superoxide radicals, chelating action [140, 144, 145], and by induction of antioxidant enzymes such as glutathione-S-transferase, glutathione peroxidase, catalase, superoxide dismutase, and hemeoxygenase [145]. It shows anti-inflammatory action by restricting cyclooxygenase-2 pathway (COX-2).In various neurological disorders, it is reported to show neuroprotective action [146]. Curcumin alone or along with manganese complex provides protective action against vascular dementia due to its antioxidant activity [147-149], and it is also helpful in treating aging and memory dysfunction [150]. In one of the studies, it has been reported that chronic administration of curcumin enhanced body weight continuously and increased SDH activity in rats treated with 3-NP [150]. The reversed 3-NP-induced motor and cognitive impairment, along with a powerful antioxidant property, indicate that curcumin may be helpful in treating HD [150].

5.7. Galanthus Nivalis (GN)

GN, commonly known as snowdrop, belongs to the family *Amaryllidacea*. Galantamine, a tertiary isoquinoline alkaloid, is the main ingredient found in bulbs and flowers of GN. The neuroprotective activity of galantamine is due to this alkaloid. It is a reversible carbamates AChE inhibitor. Galantamine is an FDA approved drug that is used to treat AD. It can stimulate nicotinic receptors that further improve memory and cognition [151]. The drug allosterically modulates nicotinic receptors of ACh, particularly subtypes α 7 and α 3 β 4, to increase the release of ACh on cholinergic cells [152].

5.8. Ginkgo Biloba (GB)

GB is an ancient Chinese herbal plant having neuroprotective properties [153]. The active phytoconstituents are mainly obtained from the leaves and flowers of the plant. These include flavonoids (quercetin, isorhamnetins and kaempferol), bioflavonoids (bilobetin, sciadopitysin, 5-methoxybilobetol, isoginkgetin, ginkgetin and aimoflavone), proanthocyanidins, Trilactonic diterpenes (A-C ginkgolide and J-M ginkgolide), and sesquiterpenes (bilobalide) [154-156].

This plant's leaf extract has been reported to be effective against dementia, cardiovascular diseases, stress, tumor and led to increased peripheral and central blood flow [157]. It also showed numerous pharmacotherapeutic activities due to its antioxidant effect [158], anti-platelet activating factor activity, and inhibition of amyloid-beta (AB) peptide aggregation [159, 160]. In one of the studies, the extract of GB (100 mg / kg, i.p. for 15 days) reversed neurobehavioral deficits induced by 3-NP and also reduced striatal MDA [161]. The standardized extract of *Ginkgo Biloba* (EGb 761) also caused up and downregulation of the expression of Bclxl and striatal glyceraldehyde-3-phosphate dehydrogenase concentrations, respectively. These biochemical findings suggested the neuroprotective function of EGb 761 in HD [161]. The extracts of GB have been reported to commonly induce biphasic dose responses in a range of cell types and endpoints (e.g., cochlea neural stem cells, cell viability, cell proliferation) [162]. The magnitude and width of the low dose stimulation of these biphasic dose responses are similar to those reported for hormetic dose responses. These hormetic dose responses occur within direct stimulatory responses as well as in preconditioning experimental protocols, displaying acquired resistance within an adaptive homeodynamic and temporal framework and repeated measurement protocols. The demonstrated GB's dose responses further reflect the general occurrence of hormetic dose responses that consistently appear to be independent of the biological model, endpoint, inducing agent, and/or mechanism. These findings have important implications for consideration(s) of study designs involving dose selection, dose spacing, sample size, and statistical power [163].

5.9. Glycyrrhiza Glabra (G. glabra)

G. glabra, commonly referred to as Yashti-madhuh, belongs to the family *Leguminosae*. G. galabra contains an isoflavane glabridin. It has been reported to exert various pharmacological activities such as antiviral, anticancer, antiulcer, anti-diabetic, antioxidant, immunomodulatory, antiinflammatory, and anticonvulsant effects. Glabridin reduces the amount of MDA and glutathione, and increases the amount of SOD in the brain [164, 165]. G. glabra lowers the brain concentrations of neurotransmitters such as glutamate and dopamine and reduces the activity of AChE [166].

5.10. Lycopodium Serratum (LS)

LS belongs to the family *lycopodiaceae*. Alkaloid huperzine is obtained from LS extract [167]. As per the literature, this alkaloid shows AChE inhibition activity. Therefore, it increases the level of ACh in post synaptic receptors in the brain. LS elicits the same kind of effects as AChE inhibitor drugs [168]. The component huperzine is also used for the treatment of AD because it inhibits the ACh enzyme, acts as an antioxidant, and possesses anti-inflammatory properties [169]. Huperzine has been reported to have several neuroprotective effects such as apoptosis, the rectification of mitochondrial dysfunction, and anti-inflammatory effects [170].

5.11. Olea Europaea (OE)

It is commonly known as olive oil and belongs to the family Oleaceae [171]. Fruits' oils contain many nutritious chemical constituents such as triacylglycerols, glycerol, free fatty acid, pigments, phosphatides, and flavor compounds [172]. Olive oil is very nutritious to the health and is also used as cooking oil. Some of the pharmacological studies have reported the potential effects of olive oil and extravagant olive oil against cardiovascular diseases [173], AD [174, 175], PD [176, 177], MS, and cancer [178, 179]. In studies conducted by Visioli et al. (1998) and Tasset et al. (2011), potential effects of extravagant olive oil have been reported against HD due to its antioxidant property [180] and neuroprotective effects [181]. In pharmaceutical formulations such as emulsions, olive oil acts as a solubilizer. In one of the studies. Guo *el al.*, have used olive oil as a solubilizer for lycopene. The lycopene loaded microemulsions (LME) were prepared in which lycopene has been dissolved in olive oil. The potentials of microemulsion in improving bioavailability and brain-targeting efficiency following oral administration were investigated [182]. The pharmacokinetics and tissue distributions of optimized LME were evaluated in rats and mice, respectively. The pharmacokinetic study revealed a dramatic 2.10-folds enhancement of relative bioavailability with LME against the control lycopene dissolved in olive oil (LOO) dosage form in rats. Moreover, LME showed a preferential targeting distribution of lycopene toward the brain in mice, with the value of drug targeting index (DTI) up to 3.45 [182].

5.12. Plants Containing Trehalose

Trehalose was subsequently found in mosses, ferns, green algae, and liverworts [183]. It is found in many plants that grow in low and high altitudes, as well as in many organisms like bacteria, yeast, fungi, insects, invertebrates [184, 185]. It has been found in the literature that trehalose inhibits the formation of amyloid [186, 187]. Besides these, it also helps in inhibiting polyglutamine (polyQ) 3-mediated protein aggregation and reduced toxicity caused by Huntington's aggregates. Tanaka et al. (2004) and Sarkar et al. (2007) conducted a study on HD using HD R6/2 mouse model. It was found that trehalose helped in the inhibition of polyQ-induced pathology by stabilizing the partly unfolded mutant proteins [188, 189]. It has also been reported that, by offering neuroprotective activity against HD, trehalose increases autophagic activity against multiple aggregations of proteins such as mHTT [188].

5.13. Panax Ginseng (PG)

PG root is a well-known herb used in China, Japan, and Korea as a tonic to revitalize and restore adequate body metabolism for over more than 2,000 years [190]. The most prevalent species of PG are Asian ginseng and American ginseng (Panaxl quinquefolium L.) from the Araliaceae family. PG is a neuroprotective herb, and its neuroprotective potential can be used to prevent and treat neurodegenerative diseases such as AD, PD, HD, depression symptoms, and strokes [163, 191]. The major constituent that is responsible for the neuroprotective action of PG is ginsenoside. In recent years a number of studies have been reported on the role of ginsenosides in the prevention of NDs [192]. Moreover, the results of some of the clinical trials conducted on PG and its constituents, ginsenosides, and gintonin, revealed that they are safe [192]. PG contains tetracyclic dammarane, triterpenoids, saponin glycosides, and ginsenosides as their active constituents [193, 194]. Different studies (in vitro and in vivo) have shown positive results of ginseng in various pathological conditions such as cardiovascular diseases, CNS disorders, cancer, immune deficiency, and hepatotoxicity [194, 195]. It also has antioxidant [196], anti-apoptotic [196], anti-inflammatory [197], and immune-stimulating functions [195]. It helps in decreasing lipid peroxidation by inhibiting excitotoxicity and over-influx of Ca²⁺ into neurons. It retains concentrations of cellular ATP, preserves neuronal structural integrity, which helps in increasing cognitive performance [195]. Ginsenoside Rb1 and Rg3 are reported to exhibit protective effects by preventing Ca^{2+} influx through glutamate receptors on cortical neurons against glutamate-induced cell death [198]. Ginseng contains saponins that are NMDA glutamate antagonists. They reduce intracellular Ca^{2+} influx in the hippocampus; hence glutamate type NMDA receptors get inhibited, and this results in reduction

of the symptoms of HD [199]. Ginsenosides Rb1, Rb3, and Rd showed a neuroprotective impact on striatal neuronal harm caused by 3-NP [200-202].

5.14. Sesamum Indicum (SI)

Sesamol is obtained from the plant SI, frequently referred to as sesame, belonging to family Pedaliaceae. It is used in India and other East Asian nations as a healthy food [203]. The oil obtained from sesame is responsible for its pharmacological activities. Its active component, sesamol, is accountable for its antioxidant activity [204]. It helps in reducing hyperlipidemia, blood pressure, and lipid peroxidation by diminishing enzymatic and non-enzymatic oxidants stress. It also has tumour suppressant action [205]. Sesamol has been reported to have its protective effect against HD through suppression of the expression of nitric oxide (NOS) [206]. It is also reported to attenuate behavioral, biochemical, and cellular changes in 3-NP-induced animals [207]. It has been reported to protect the brain against memory impairment caused by 3-NP, OS, neuroinflammation in the neurons of the hippocampus, and thus increases synaptic plasticity and neurotransmission [208].

5.15. Solanum Lycopersicum (SL)

SL is commonly referred to as tomato and belongs to the family *Solanaceae*. Lycopene is a well-known carotenoid found in tomatoes and tomato-based goods in considerable quantities [209]. It has been reported to possess powerful neuroprotective [210], antioxidant [211, 212], antiproliferative, anticancer [213], anti-inflammatory [214], memory enhancing [215], and hypocholesterolemic properties [216]. It is a stronger singlet oxygen carotenoid quencher for vitamin E and glutathione [216]. Treatment with lycopene considerably helps in the reduction of multiple behavioral and biochemical changes induced by 3-NP, indicating its therapeutic potential against HD's symptoms [217].

5.16. Tinospora Cordifolia (TC)

TC belongs to the family Menispermaceae and is frequently known as Giloy. Phytochemical constituents such as alkaloids, steroids, diterpenoid lactones, aliphatics, and glycosides are present in giloy extract [218]. TC has been reported for memory enhancing property, immunostimulation, and enhancement of ACh synthesis [219]. It has powerful free radical scavenging characteristics and also reduces ROS and reactive nitrogen species as studied by paramagnetic resonance electron spectroscopy [220]. It also reduces the level of glutathione, gamma-glutamyl-cysteine ligase expression, copper-zinc superoxide dismutase genes, owing to which it can be used for the treatment of hypoxia, ischemia, and neuronal injury [220]. Additionally, TC is helpful in enhancing dopamine levels in the brain and improving cognitive and psychotic function [219].

5.17. Tripterygium Wilfordii (TW)

The root extract of TW has been extensively used as traditional Chinese medicine for the treatment of inflammation and autoimmune diseases such as rheumatic arthritis [221]. TW's root extract also showed neurotropic and neuroprotective effects [222]. Celastrol and Triptolide are the two major neuroprotective phytoconstituents that are isolated from the root extract of TW. It has many therapeutic potentials such as antioxidant [223], anti-inflammatory [221], anticancer [224], and insecticidal activity [225]. A pro-inflammatory study conducted on animals using 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine (MTPT) indicated that celastrol helped in improving the functions of dopaminergic cells, increasing the dopaminergic level [222]. By controlling the expression of the thermal shock protein gene in dopaminergic cells, it has also provided protection against 3-NPinduced striatal damage [222, 226].

5.18. Withania Somnifera (WS)

WS is also known by the common name Ashwagandha. It belongs to the family Solanaceae. For centuries, it has been used in Ayurvedic medicine [227]. Ashwagandha's root extract has been reported to possess antioxidant [228, 229], memory enhancing [230], anti-inflammatory [231], immunomodulatory [232], anti-stress [233], and anti-convulsant characteristics [234]. As an antioxidant, WS and its active ingredients (sitoindosides VII-X and withaferin A) increase catalase, ascorbic acid, endogenous superoxide dismutase, and reduce lipid peroxidation [235-237]. It functions as an anti-inflammatory agent through complement inhibition, the proliferation of lymphocytes, and delayed hypersensitivity. Different trials have shown that WS increases cortisol circulation, decreases tiredness, increases physical performance, and decreases refractory stress depression [238]. It also modulates different receptor systems for neurotransmitters in the CNS. Major active constituents of WS include steroidal lactones and alkaloids (collectively referred to as withanolides). Withaferin A, withanolide A, withanolide D-P, withanone, sitoindoside VII-X are the major isolated withanolides from WS. WS inhibits AChE and increases the level of ACh in the brain. The beneficial effects of herbal drugs against HD are listed in Table 4.

6. CURRENT ONGOING CLINICAL TRIALS

A limited number of clinical trials have been reported for the treatment of HD. This could be attributed to a lack of complete understanding of the underlying mechanism of the disease as the drugs used so far have been unable to provide complete relief to patients. Some of the trials that have been completed or are ongoing, are listed in Table **5**.

7. HORMESIS AND REDOX ASPECTS OF HERBAL DRUGS AND THEIR POTENTIAL CHEMICAL CONSTITUENTS

Hormesis is a biological process that has found its application in drug development, drug designing, and toxicological studies. It helps to rationalize the dose-response relationships [80, 256]. Hormetins are the chemical inducers of hormesis and possess a range of therapeutic applications, including protection against stress, toxin, and aging-related diseases [257, 258]. They have a protective effect at a low level and show deleterious effects at higher levels due to the narrow therapeutic window [259, 260]. Hormesis describes the phenomenon of pharmacological conditioning of the heart and brain where a low dose of pharmacological agents

Name of Herbal Medicine	Synonyms	Source	Bioactive Component	Structure	Animal Models	Effects	Refs.
AC	Sweet flag		α-and β-Asarone			Antioxidant, anti- inflammatory	[239]
AG	Ginseng	Whose root	Ginsenosides		-	Antioxidant, anti- apoptotic, anti- inflammatory, and immune-stimulating functions	[195- 197]
WS	Withania root, asgandh, winter cherry.	Dried Roots	Withaferin A	CH_3 O CH_3 O CH_3 O H_3C O	3-nitropropionic acid model	Reduce oxida- tive/nitrosative stress, inhibits complex II of the mitochondrial electron transport chain	[240]
ВМ	Kapotvadka, somvalli and saraswati	Aerial parts	Bacoside A,	H_{0} H_{0	3-nitropropionic acid induce model	Memory enhancer	[241]
			Bacoside B,	HO HO HO HO HO HO HO HO	3-nitropropionic acid induce model	Facilitates an- terograde memory	
CR	Celastrol (tripterine)		Celastrol		_	Anti-inflammatory, anti-oxidant, and inhibition of Pro- inflammatory cytokines.	[221]
CL	Indian saffron, curcuma, Turmeric, Haldi	Fresh rhizomes	Curcumin		3-nitropropionic acid-induced HD rat model and inhibitory response against AMPA receptor	Anti-oxidant, Anti- inflammatory and reduce excitotoxic- ity	[242]

Table 4.	Effect of herbal	drugs and	phytoconstituents.
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(Table 4) contd....

Name of Herbal Medicine	Synonyms	Source	Bioactive Component	Structure	Animal Models	Effects	Refs.
			Demethoxy curcumin	-	Rotenone-induced PD in rats	Anti-oxidant and Anti-inflammatory	
CS	Coriander	Leaves	Coriandrum sativum extract		Ischemic reperfusion insult in brain	Neuroprotective effects	[133]
GN	Snowdrop	Bulbs and flowers	Galantamine 0.1 mg/kg, s.c.		Scopolamine- induced amnesia model in mice	Reduce AChE enzyme and increase ACh in postsynaptic receptor	[243]
Ginkgo (GB)	Ginnan, maidenhair tree.	Leaf	Gingkolides A,B,C,J and M	-	3-nitropropionic acid model	Memory enhancer property and Anti- Platelet Activating Factor (Anti-PAF)	[161]
GG	Yashti- madhuh or liquorice	Stems	Glabridin	-	-	Antioxidant	[244]
CA	Spade leaf, Indian Pen- nywort, Mandukaparni		Asiatic Acid	HO, CH ₃ HO, CH ₃ HO, CH ₃ HO, CH ₃ CH ₃ C	_	Neuroprotective effect against harm caused by OS and Mitochondrial dysfunction	[245]
LS	Ground pines or creeping cedar, Qian Ceng Ta.	Leaves	Huperzine A	HN H ₂ N	_	Antioxidant and anti-inflammatory and reduce mito- chondrial dysfunc- tion	[170]
Persea Americana	Avocado	Peel, seed coat and seeds	Persea major methanolic extract (0.5 mg/ml)		Cellular viability assay, Glutamate uptake assay	Antioxidant capac- ity, increased glutamate uptake	[246]
OE	Olive- growing	Oil	Olive oil, Extravir- gin olive oil (20 mg/kg ip)	R_1 O O R_2 R_2 O O R_3	3-nitropropionic acid-induced HD- like rat model	Reduces oxidative damage	[181]
Sesamum indicum	Sesame, benne	Oil	Sesamol	O OH		Neuroprotective effect	[247]
TC	Giloe	Stem	<i>Tinospora cordifo- lia-</i> stem methano- lic extract		6-hydroxy dopamine (6-OHDA) lesion rat model, Cadmium- induced OS in Wistar rats	Anti-OS, Memory enhance and Increase dopamine level in to brain.	[248]

(Table 4) contd....

Name of Herbal Medicine	Synonyms	Source	Bioactive Component	Structure	Animal Models	Effects	Refs.
TW	Thunder god vine	Root extracts	Celastrol, Triptolide	HO O HO HO HO HO HO HO HO HO HO HO HO HO		Antioxidant effects	[249]
Fruits, vegetables, tea, cocoa and wine			Flavonoids			Effects against OS and Inflammation	[250]
Mosses ferns, green algae, and liverworts			Trehalose		induced damage in bovine spermatozoa	Antioxidant effects	[183, 251]
SL	Tomato	Hole fruits	Lycopene	CH_3	3-nitropropionic acid- induced HD rat model	Inhibition of cogni- tive dysfunction and motor abnormality and antioxidant effects	[252, 253]
Saccharomyces cerevisiae and Corynebacterium glutamicum	Rasberry	Fruit	Salidroside		Inhibit the SOD1 and HTT genes and also show anti- inflammatory effects.	Reduce the symp- toms of HD by acting oxidative stress and inflammation, and HTT genes.	[254]

Table 5.Ongoing clinical trial of HD.

Disease	Drug	Sample Size	Purpose	Phase	Status	Design	Study State	Study end
HD	THC, CBD	21	Treatment	Phase 2	Completed	DB, R, CO	December 30, 2011	February 1, 2013
	EGCG	54	Treatment	Phase 1	Completed	R	May 23, 2011	June 16, 2015
	PBT2	109	Treatment	Phase 2	Completed	DB, R	May 3, 2012	July 18, 2016
	DM/Q	22	Treatment	Phase 3	Recruiting	R	February 26, 2019	April 19, 2019
	Triheptanoin	10	Treatment	Phase 2	Completed		June 20, 2013	March 24, 2016
	SD-809	90	Treatment	Phase 3	Completed	R, DB	January 2, 2006	September 20, 2017
	Digoxin, Dimebon	12	Treatment	Phase 1	Completed	R	January 29, 2009	June 12, 2009
Chorea	SD-809	90	Treatment	Phase 3	Completed	R, DB	February 21, 2013	August 11, 2017
	Amantadine sulphate	30	Treatment	Phase 4	Completed	NR	July 31, 2009	June 28, 2011
HMD	Tetrabenazine				Available		March 24, 2008	February 26, 2020

Abbreviations: CO; Cross Over, CBC, Cannabidiol, DB; Double Blind, DM/Q; Dextromethorphan/quinidine, EGCG; (2)-epigallocatechin-3-gallate, NR, Non-Randomized, R; Randomized, THC; Delta-9-tetrahydrocannabinol, HMD; Hyperkinetic Movement Disorders Based on search of clinicaltrial.gov (https://clinicaltrials.gov/ct2/ results?cond=huntington+disease&term=&cntry=&state=&city=&dist=) [255] [Accessed May 26, 2020]. activate various downstream cascades that have cardio and neuroprotective potential, but at high doses, their protective effect gets attenuated [261-263]. The spectrum of hormetic results, such as increased development, reproduction, survival, and a decreased disease occurrence, indicates the presence of thousands of genes, thereby influencing basic biological processes through hormetic mechanisms. In one of the recently published studies, Moghaddam et al (2019) [264] mentioned hormesis effects of curcumin. It is one of the types of hormetic agents because, at a low dose, curcumin shows stimulatory effects, while at high doses, it shows inhibitory effects. For example, at a low dose, curcumin shows antioxidant and anti-inflammatory effects but in high dose curcumin is reported to cause autophagy and apoptosis or cell death [264, 265]. In another study, it is reported that quercetin showed antioxidant, anti-inflammatory, and neuroprotective effects at its low dose whereas, at higher doses, it caused toxicity into the body such as mitochondrial oxidative stress [261, 266]. Another herbal drug, celastrol that is extracted from TW, has proven its neuroprotective effects in preclinical studies. It is also called "Thunder of God Vine". This phytomedicine decreases the striatal lesion volume, which is induced by 3-NP at its low dose. At low doses, it showed antioxidant effects and reduced neuroinflammation induced by NF κ B and TNF α signaling pathways. But at a high dose, celastrol increased the blood pressure and caused hypertension [261]. BM's extract has many potential effects against neuronal diseases such as anxiety, depression, and various NDs. Whereas, its overdose causes dry mouth, stomach cramps, fatigue and bowel movement, etc. [94, 267]. Hence, hormetic processes should be considered because plant derivatives at low dose may provide pro-oxidants that are able to upregulate the expression of enzymes of innate detox pathways or, alternatively regulate the expression of vitagenes [259]. Various drugs shown in Fig. (3) have neuroprotective effects however, they also show hormesis effects.

Products such as wine extract, green tea, grape seed, PA, CL, OE, and TC extracts are all known to contain a large variety of potent antioxidants in the form of polyphenols like phenolic acids, gallic acid, stilbenes, tannins, flavanols, resveratrol, and anthocyanins, etc. [179, 268] Polyphenolic compounds act as iron chelators, radical scavengers, and modulators of pro-survival genes. These polyphenols activate the endogenous enzymes like glutathione peroxidase, catalase, or superoxide dismutase that directly modulate the level of free radicals [269]. In NDs, neuronal stress response activates pro-survival pathways, which control the activation or modulation of protective genes called vitagenes. These vitagenes produce endogenous enzymes, heat shock proteins, heat shock protein 72 (Hsp72), heme oxygenase-1, sirtuins, and the thioredoxin/thioredoxin reductase system [260]. All these have potent anti-oxidant and anti-apoptotic activities against NDs. Polyphenols activate the vitagene system by upregulating the levels of antioxidant enzymes and sirtuin system, along with activation of heat shock transcription factors and Kelch-like erythroid cell-derived proteins with CNC homology [ECH]-associated protein 1)/antioxidant response element Keap1/Nrf2/ARE pathway that results in counteraction of pro-oxidant conditions in neuronal tissue [259]. In neuronal cells, mitochondria are the principal source of energy for their survival. In stressful conditions, neuronal cells compensate the energy demands of cells by changing the rate of mitochondrial fission and fusion. This process leads to excessive production of superoxide anions at the inner mitochondrial membrane that promotes the production of physiological or endogenous ROS. These mitochondria-derived ROS are involved in the aging process. The ROS directly modulates signal transduction pathways that enhance cellular proliferation [270]. These changes in mitochondrial activity interrupt the functionality of the mitochondrial network and promote the molecular abnormalities influencing mitochondrial dynamics. Since mitochondria play a critical role in neuronal physiology, impaired mitochondrial dynamics promote the NDs such as PD [176], AD [271, 272], and HD. Polyphenols like flavanols are known to have brain-permeability potential that directly benefits neuronal health. Several studies show that polyphenols have a neuroprotective role in NDs, for example, epigallocatechin gallate has neuroprotective potential in amyloid-betamediated neurotoxicity. Resveratrol acts by decreasing nuclear factor kappa-light-chain-enhancer of activated B cells (NFkB) level and microglia-induced neuroinflammation, thereby it protects the brain from the deleterious effect of ischemic injury. Polyphenols directly or indirectly modulate the levels of pro-and anti-inflammatory microRNAs in NDs [273-275]. Polyphenols have shown the potential to activate the mitochondrial biogenesis, in aged mice. They attenuated the deleterious effect of oxidative stress mediated damage and increased the physical endurance that resulted in prolonged survival of the animals [257, 276, 277].

8. NEED FOR NOVEL DRUG DELIVERY SYSTEMS

Herbal drugs have been reported to show very good neuroprotective effects; however, they have some limitations such as poor bioavailability, poor aqueous solubility, and lack of blood-brain barrier (BBB) permeability. Novel drug delivery systems have been reported to enhance the bioavailability, therapeutic efficacy [278], stability, and brain permeability [279] of the herbal drugs and reduce their side effects, which, on the other hand, is hard to be achieved through conventional drug delivery systems [280-283]. Herbal drugbased nanoparticles are reported to reduce first pass metabolism and improve their bioavailability because their small particle size (less than 200 nm) enables them to cross endothelial cells of BBB by transcytosis [284]. Glucose transporter 1 (GLUT1) or albumin transporters, lactoferrin receptors, transferrin receptors ligands can enhance receptormediated transcytosis [285]. The mechanisms of targeting of BBB of drug-loaded nanoparticles are shown in Fig. (4). There are a number of studies in which the plant extracts or their active constituents have been reported to enhance the pharmacokinetic properties such as Cmax and AUC, thereby increasing their oral bioavailability. Hence, they have been able to treat various types of NDs such as PD, AD. Some of the studies entailing about enhancement of oral bioavailability are listed in Table 6. It is important to note that there is very limited information available regarding the formulation of nanoparticles to treat HD, and they are limited to preclinical studies. However, based on the success rate of NDDS in treating other neurodegenerative diseases apart

S. No.	Plant Used	Phytoconstituent	NDDS	Outcomes	Refs.
01	Malus domestica, Allium cepa, etc	Fisetin (FS)	Self- nanoemulsifying drug delivery sys- tem (SNEDDS)	 Cmax of FS SNEDDS increased by 3.7 folds as compared to naïve FS AUC_{0-x} of FS SNEDDS increased by 1.5 folds as compared to naïve FS Increase in bioavailability (151.58%) 	[286]
02	Nigella sativa	Thymoquinone	SLNs	 Cmax of thymoquinone SLNs increased by 4.3 folds as compared to thymoquinone suspension AUC_{0-x} of thymoquinone SLNs increased by 6.19 folds as compared to thymoquinone suspension Increase in bioavailability (619.3%) 	[287]
03	Nigella sativa	Thymoquinone	SLNs	 Cmax of thymoquinone SLNs increased by 4.8 folds as compared to thymoquinone AUC_{0-∞} of thymoquinone SLNs increased by 5.53 folds as compared to thymoquinone Increase in bioavailability (553%) 	[288]
04	CL	Curcumin	SNEDDS	 Cmax of curcumin SNEDDS increased by 9.1 folds compared with naïve curcumin AUC_{0-x} of curcumin SNEDDS increased by 7.5 folds as compared to naïve curcumin Increase in bioavailability (754%) 	[289]
05	CL	Curcumin	Nanosuspensions	 Cmax of curcumin nanoparticles increased by 4.8 folds compared to naïve curcumin. AUC_{0-∞} of Curcumin nanoparticles increased by 5.5 folds compared with naïve curcumin. Increase in bioavailability (558%) 	[290]
06	CL	Curcumin	SLNs	 Cmax of curcumin nanoparticles increased by 49.27 folds compared to naïve curcumin. AUC_{0-x} of Curcumin SLNs increased by 39.06 folds as compared to naïve curcumin. Increase in bioavailability (3906%) 	[291]
07	CL	Curcumin	Phospholipid com- plex	 Cmax of curcumin phospholipid complex increased by 2.4 folds as compared to naïve curcumin AUC_{0-x} of Curcumin phospholipid complex increased by 5.19 folds as compared to naïve curcumin Increase in bioavailability (519%) 	[292]
08	GB, Allium cepa, Brassica oleracea var. italic etc	Quercetin	Zein nanoparticles	 Cmax of quercetin nanoparticles increased by 8.5 folds as compared to naïve quercetin AUC₀₋₁₂₀ of quercetin SLNs increased by 13.9 folds as compared to naïve quercetin Increase in bioavailability (1396%) 	[293]
09	GB, Allium cepa, Brassica oleracea var. italic etc	Quercetin	SLNs	 Cmax of quercetin SLNs increased by 2.07 folds as compared to naïve curcumin AUC_{0.48} of quercetin nanoparticles increased by 5.7 folds as compared with naïve curcumin Increase in bioavailability (571.4%) 	[294]
10	Camellia sinensis	Epigallocatechin- 3-Gallate (EGCG)	Nanolipidic parti- cles	 Cmax of EGCG nanoparticles increased by 6.04 folds as compared to EGCG 10% ethanolic extract AUC_{0-∞} of EGCG nanoparticles increased by 2.49 folds as compared to EGCG 10% ethanolic extract Increase in bioavailability (249%) 	[295]

Table 6. Pharmacokinetics parameters for the herbal drugs and their nanoparticles.

(Table 6) contd....

S. No.	Plant Used	Phytoconstituent	NDDS	Outcomes	Refs.
11	SL	Lycopene	Microemulsion	 Cmax of lycopene-loaded microemulsion increased by 1.8 folds as compared to lycopene devolved in olive oil AUC_{0-x} of lycopene-loaded microemulsion increased by 2.1 folds as compared to lycopene devolved in olive oil Increase in bioavailability (210%) 	[182]
12	TW	Celastrol	Silk Fibroin Nanoparticles (SFNPs)	 Initial concentration of celastrol SFNPs increased by 4.36 folds as compared to celastrol in PEG 300 AUC_{0-x} of celastrol SFNPs increased by 2.61 folds as compared to celastrol in PEG 300 Increase in bioavailability (261%) 	[296]
13	TW	Celastrol	Phytosomes	 Initial concentration of celastrol phytosomes increased by 5 folds as compared to celastrol AUC_{0-x} of celastrol phytosomes increased by 4.1 folds as compared to celastrol Increase in bioavailability (410%) 	[297]
14	Phaseolus vulgaris, Glycine max, etc	Genistein	Eudragit nanoparti- cles	 Cmax of genistein nanoparticles increased by 2.4 folds as compared to genistein suspension AUC_{0-x} of genistein nanoparticles increased by 2.4 folds compared to genistein suspension Increase in bioavailability (241%) 	[298]
15	GN	Galantamine	SLNs	 Volume of distribution of galantamine SLNs increased by 1.15 folds as compared to galantamine AUC_{0-x} of galantamine SLNs increased by 2.14 folds as compared to galantamine Increase in bioavailability (261%) 	[299]
16	Silybum marianum	Silymarin	Nanostructured Lipid Carrier (NLCs)	 Cmax of silymarin NLCs increased by 3.4 folds as compared to silymarin pellets AUC_{0-x} of silymarin NLCs increased by 2.2 folds as compared to silymarin pellets Increase in bioavailability (224%) 	[300]
17	Medagascar periwinkle	Vinpocetine	SLNs	 Cmax of vinpocetine SLNs increased by 3.2 folds as compared to vinpocetine AUC_{0-x} of vinpocetine SLNs increased by 4.16 folds as compared to vinpocetine Increase in bioavailability (416%) 	[301]
18	Trifolium pratense	Biochanin A	NLCs	 Cmax of biochanin PEG-NLCs increased by 15.74 folds as compared to biochanin AUC_{0-x} of biochanin PEG-NLCs increased by 2.89 folds as compared to biochanin Increase in bioavailability (289%) 	[302]
19	Glycine max	Genistein	Micellar emulsions (ME)	 AUC_{0-x} of genistein MEs increased by 2.36 folds compared to genistein Increase in bioavailability (236%) 	[303]

from HD, it is anticipated that they may provide success in treating HD also. Hence, there is a dire need to explore those delivery systems loaded with aforementioned phytoconstituents/extracts to treat HD. Nevertheless, in this review some studies that have been reported to treat HD are discussed in the subsequent sections.

8.1. Nanoliposomes

The herbal drugs loaded in nanoliposomes have the potential to cross the physiological membrane barriers of the body owing to the submicron size of vesicles. The drugs loaded in the vesicles can bypass first pass metabolism and enhance oral bioavailability. Ligand (such as GLUT1, lactoferrin, transferrin) based nanoliposomes prepared through surface modification methods have been able to deliver several proteins, antibodies, and peptides [304]. Ligands help the liposomes to permeate BBB by transcytosis. Nanoliposomes can also enter the brain by passive diffusion, where they release entrapped drugs by energy dependent mechanism or passive efflux [304]. The limitation of liposomes is their short half-life, due to which the drug gets easily metabolized by hydrolysis and oxidation [305]. Francesca *et al.* studied the effect of curcumin-loaded apoprotein E (Apo-E) derived peptide nanoliposomes on HD. The



Fig. (4). Mechanism of BBB permeability of the nanoparticles. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

liposomes were prepared by loading Apo-E in the dispersion of bovine brain sphingomyelin (Sm), cholesterol (Chol), and 1,2-stearoyl-sn-glycero-3-phosphoethanolamine-N-[maleimide (poly(ethylene glycol)-2000)] (mal-PEG-PhoEth) using thin film hydration method (Fig. 5A). The prepared liposomes showed particle size, PDI, and zeta potential of 132 ± 10 nm, 0.187, and -19.41 ± 0.09 mV, respectively. The ratio of Apo-E to Chol and Sm was kept constant. However, composition of Apo-E and mal-PEG-PhoEth were varied in the ratio of 1.2:1 and 1:5. During ApoE-liposome coupling, it was observed that when the ratio of peptide and mal-PEG-PhoEth was changed from 1:5 to 1.2:1, it resulted in increased density of ApoE on the surface of the liposome (Fig. **5B**). It was observed that around 70,000 molecules of lipids were on the surface of the liposome, having a particle size of 140 nm and consisted of reactive mol-PEG-PhoEth (2.5 mol). Coupling efficiency was found to be 70%. The molar ratio of peptide and mol-PEG- PhoEth (1:5 to 1:2:1) after the incubation period showed a high density of 1200 and low density of 400 peptide molecules per single nanoliposomes particle (Fig. 5C). The in vitro cell line study was done on rat's brain endothelial cells. The curcumin-nanoliposomes did not show any cytotoxicity as confirmed by 3-(4,5dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay. The liposomes were labeled by using a fluorescent dye and cellular uptake of the fluorescence labeled liposome was detected by using confocal laser scanning microscopy (CLSM). It was found that liposomes in the absence of surface functionalization did not show any membrane accretion and cellular uptake of fluorescence. The Rat Brain Endothelial cells (RBE₄) showed very less fluorescence at high and low density of peptides. It was observed that the green fluorescence of cells got increased with an increase in the high density of peptides. The green spots were near the nucleus below the plasma membrane. The liposomes coupled with peptide mApoE displayed effective uptake (Fig. 5D). The curcumin-nanoliposomes helped in treating HD by their interaction with low-density lipoprotein receptors via special Apo-E sequence amino acid and penetrated curcumin across the BBB through transcytosis without getting affected by lysosomal degradation. Hence, the obtained results revealed that ligand-based nanoliposomes successfully targeted BBB and protected the drug from degradation [306].

8.2. Solid Lipid Nanoparticles (SLNs)

SLNs contain a solid lipid matrix stabilized by lipid molecules and physiological emulsifiers. Homogenization is used for the preparation of SLNs, where high temperature and high pressure provided by thermodynamic and mechanical stress causes size reduction of drug particles [307, 308]. SLNs are excellent nanocarriers to enhance the bioavailability of drugs and are highly biocompatible. SLNs of size ranging from 0 to 1000nm can be prepare during high pressure



Fig. (5). (A) The Curcumin liposomes were prepared by thin film hydration method. (**B**) Fluorescence estimation of the amount of mApoE (dark bars) and dApoE peptide coupled to liposomes at different peptide-to-lipid molar ratios. (**C**) Cell-associated fluorescence was evaluated using FACS analysis. (**D**) The localization and distribution of ApoE-NLs within RBE4 cells [306] Copyright © 2011 Elsevier. (*A higher resolution / colour version of this figure is available in the electronic copy of the article*).



Fig. (6). (A) C-SLNs prepared by halogenation method (B) Neuroprotective effects of C-SLNs reported against 3-NP in rat brain. (C) Effect of C-SLN on mitochondrial cytochrome levels in the striatum of 3-NP-induced HD rats. (D) Effect of C-SLN on spontaneous locomotor activity in terms of total photo beam counts of 3-NP-induced HD rats [76] Copyright © 2013, Springer Nature. (A higher resolution / colour version of this figure is available in the electronic copy of the article).



Fig. (7). (A) Formulation of TQ-SLN prepared by homosimeation method. (**B**) TQ-SLN has been showed dose dependent significant effects against the inflammatory mediator such as TNF- α , IL-1 β , IL-6, iNOS, and COX2 (**C**) TQ-SLNs and TQ-S treatment attenuates the overexpression of GFAP in the striatal slices of 3-NP intoxicated animals. (**D**) TQ-SLNs and TQ-S treatment improve the expression of TH in the striatal slices upon 3-NP toxicity [312] Copyright © 2018, Springer Nature. (*A higher resolution / colour version of this figure is available in the electronic copy of the article*).

homogenization. Those in size range of 120-200 nm can easily cross endothelial cells of BBB by endocytosis [309]. Brain permeability of SLNs can be enhanced by their attachment with ligand (e.g., apolipoprotein E) [310]. Limitations of SLNs are their low drug loading capacity and poor entrapment efficiency. In one of the studies carried by Sandhir et al. (2014), C-SLNs reported positive results against 3-NP induced HD rats. In this study, 20 mg/kg and 40 mg/kg drug was administered for 7 days through oral route. C-SLNs were prepared by the homogenization method. In this formulation steric acid, lecithin taurocholate and curcumin were used. The formulation exhibited significantly dose dependent neuroprotective effects against neurotoxin (3-NP) (Fig. 6B). It also produced significant positive effect on mitochondrial cytochrome levels in striatum and spontaneous locomotor activity in total photobeam counts of 3-NP-induced HD rats [76] (Fig. 6C, D).

In another study, the authors have mentioned the pharmacological action of the thymoquinone (TQ). It is a strong antioxidant and also inhibits neuroinflammation. The drug still could not show the desired action in the in-vivo study [311] due to low solubility, leading to decrease drug absorption and bioavailability. Thereby, a drug cannot reach a desired concentration in the targeted side (the brain). Ramachandran et al. (2018) [312] prepared TQ-SLNs to enhance the bioavailability and brain permeability of the drug. TQ-SLNs were prepared by the homogenization method. (Fig. 7A) The inflammatory response was checked by polymerase chain reaction (PCR), and TQ-SLNs showed anti-inflammatory effects. TQ-SLNs and Thymoguinone suspension (TQ-S) were found to inhibit inflammatory mediators including TNF- α , IL-1 β , IL-6, iNOS, and COX2 (Fig. 7B). The anti-inflammatory effects of TQ-SLNs and TQ-S reported in the glial fibrillary acid protein (GFAP)



Fig. (8). (A) Zwitterionic poly(trehalose) can easily permeate BBB and inhibit polyglutamate aggregation in CNS. (**B**) **a**) Immunoblot analysis data of soluble huntingtin aggregates using (green fluorescent protein) GFP antibody. **b**) Quantification of band intensities of soluble huntingtin (tNhtt) shown in (**a**) using NIH image analysis software. Data are normalized against beta-actin. **c**) Dot blot analysis data of insoluble huntingtin aggregates using GFP antibody. **d**) Dot blot analysis using ubiquitin antibody [314] Copyright © 2017 American Chemical Society. (*A higher resolution / colour version of this figure is available in the electronic copy of the article*).

S. No.	Phytoconstituents	Formulation	Compassion	Animal Model	No. of Animal	Dose (mg/kg)	Duration of Study	Applications	Results	Biochemical Evaluation	Refs.
1.	Curcumin	SLNs	Steric acid, lecithin taurocho- late, Curcumin	Female wistar rats	20	40mg/kg p.o.	7 days	SLNs improved oral bioavailability of curcumin	Assessed its neuroprotective efficacy against 3- NP-induced HD	Reduced GSH levels and SOD activity, reduc- tion in mito- chondrial swelling, lipid peroxidation, protein carbonyls and ROS	[76]
2	poly(trehalose)	Polymeric nanoparticle	Sulfo-acrylate (to introduce SO3-), amino-acrylate (to introduce NH2), PEG-acrylate (to introduce PEG).	The transgenic mice for HD [strain B6CBA-Tg (HDexon1) 62Gpb/3J]		0.4 mg/mL corresponding to 50 μM Trehalose, i.p.	56 days and 84 days	Polymeric nanopar- ticles enhanced BBB permeability of the trehalose	Neuroprotective effects	Immuno- histochemical staining,	[314]
3.	Thymoquinone	SLNs	Steric acid, lecithin taurocho- late, Thymoquinone	Albino male rats	48	TQ-SLNs (10, 20mg/kg), TQ-SLNs (40,80mg/kg) p.o.	14 days	SLNs increased the solubility, bioavail- ability and absorp- tion of the thymo- quinone. It also enhanced drug payload and sustained drug release ability	Due to this SLNs thymoquinone acts as halting 3- NP induced inflammation and degeneration.		[312]

Table 7.	Herbal Nanoformulations reported for the treatment of HD in animal models.
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(Table 7) contd....

S. No.	Phytoconstituents	Formulation	Compassion	Animal Model	No. of Animal	Dose (mg/kg)	Duration of Study	Applications	Results	Biochemical Evaluation	Refs.
4.	Cholesterol	Nanaolipos omes		Mice	3	Chol-D6-loaded liposomes (200 µg/mouse)	2 days	Nanoliposomes enhanced brain delivery of cholesterol	Brain cholesterol (Chol) synthesis, which is essential for optimalsynap- tic transmission		[315]
5.	Selenium	Nanoliposomes		Worms		_		Selenium Nanopar- ticles enhanced bioavailability and therapeutic efficacy with low toxicity	-	20µM, Nano-Se played a dosage- dependent protective effect on the viability after the exposure to both stress stimuli	[316]
6.	Lithium	Microemulsion (NP03)		YAC128 mouse		20 μg Li/kg or 40 μg Li/kg body weigh <i>i.e.</i> 0.03 and 0.06 mEq/kg)	2 months	Microemulsion of lithium (NP03) reduced the toxicity of lithium and increased the absorptions at targeted site	NP03 improves motor function and rescues striatal pathology and testicular atrophy in YAC128 mice.		[317]
7.	Coenzyme Q10 (CoQ10)	Coenzyme Q10		R6/2 transgenic mouse	110	(CoQ10) 1000, 5000, 10000, or 20000 mg/kg/day and (HydroQ-sorb) 400, 1000, and 2000 mg/ kg/day	150 days		Showed neuropro- tective effects		[318]

and tyrosine hydroxylase (TH) against 3-NP induced animals model [312] are shown in Fig. 7C and D.

8.3. Polymeric Nanoparticles

The particle size of polymeric nanoparticles is approximately in the range of 10 nm to 1000 nm. These can be formulated in the form of nanospheres and nanocapsules. Nanospheres are made up of a matrix system. In nanocapsules, the drug is loaded into the cavity, which is made up of a polymeric membrane [313]. Debnath et al. (2017) reported the successful delivery of trehalose by enhancing its BBB permeability through poly(trehalose) nanoparticles. Poly(trehalose) nanoparticles were reported to be more potent as compared to trehalose molecules. They were found to inhibit polyglutamine aggregation in HD150Q cell in in vitro study. (Fig. 8A) Immunoblot analysis and Dot blot analysis of poly(trehalose) nanoparticles reduced polyglutamine levels and amyloid aggregation and also suppressed mHTT genes [314]. (Fig. 8B) Herbal drug nanoparticle formulation, animal models, and beneficial results are enlisted in Table 7.

CONCLUSION

The prevention of NDs is essential for the aged population globally. Phytoconstituents serve as novel medicinal therapies in the present scenario. Herbal drugs are reported to have multiple actions such as antioxidant, antiinflammatory, anti-proliferative and anti-apoptotic. Many of them are also reported to reduce AChE levels in synapses. Hence, they could offer a pragmatic alternative to the current synthetic drugs that are being used to treat HD. Various preclinical and clinical studies have been highlighted in the manuscript indicating a significant positive response against symptoms of HD. Despite having such therapeutic potential, the efficacy of herbal drugs has not been widely explored due to their poor solubility and pharmacokinetic properties. Herbal drugs incorporated in various nanocarriers such as nanoliposomes, microemulsions, SLNs, and polymeric nanoparticles have shown very good efficacy to treat HD due to the enhancement in their bioavailability ordirect targeting to specific cells. This has further helped in the reduction of their dose as well as toxicity. The major challenges associated with the formulation of herbal drug-loaded nanoparticles include the poor loading of drugs in the formulation, low stability of herbal drugs during their processing, difficulty during scale-up of the process, and low stability of nanoformulations. Hence, it is important to look at these issues prior to the start of pre-clinical studies. Upon getting successful pre-clinical reports, a thorough clinical study is required for their positioning into the market.

LIST OF ABBREVIATIONS

3-NP	=	Nitropropionic acid
6-OHDA	=	6-hydroxydopamine
AC	=	Acorus calamus
ACh	=	Acetylcholine

AChE	=	Acetylcholinesterase			=	Nanoemulsions			
AD	=	Alzheimer Disease	NFκB	NFκB		Nuclear Factor kappa-light-chain-			
ALS	=	Amyotrophic Lateral Sclerosis				enhancer of activated B cells			
Anti-PAF	=	Anti-Platelet Activating Factor	NLCs	5	=	Nanostructured Lipid Carrier			
AS	=	Allium sativum	OH8d	lG	=	8-hydroxy-2-deoxygonosine			
ATP	=	Adenosine Triphosphate	PD		=	Parkinson Disease			
Αβ	=	Amyloid beta	PG		=	Panax Ginseng			
BBB	=	Blood Brain Barrier	RNS		=	Reactive Nitrogen Species			
BM	=	Bacopa monnieri	ROS		=	Reactive Oxygen Species			
CA	=	Centella Asiatica	SDH		=	Succinate dehydrogenase cell enzyme			
CAG	=	Cytisine-Adenine-Guanine	SI		=	Sedamim Indicum			
CAT	=	Choline-Acetyl Transferase	SLNs		=	Solid Lipid Nanoparticles			
CL	=	Curcuma Longa	TC		=	Tinospora cordifolia			
CNS	=	Central Nerves System	TH		=	Tyrosine Hydroxylase			
COX-2	=	Cyclooxygenase-2	TLR		=	Toll-Like Receptor			
CS	=	Coriandrum sativum	TQ		=	Thymoquinone			
CSF	=	Cerebrospinal Fluid	TQM		=	Triterpenoid Quinone Methide			
DTI	=	Drug Targeting Index	TQ-S		=	Thymoquinone Suspension			
EGCG	=	Epigallocatechin gallate	TR-FI	RET	=	Time-resolved Fluorescence Energy Transfer			
ELISA	=	Enzyme-Linked Immune Sorbent Assay	WS		=	Withania Somnifera			
G. glabra	=	Glycyrrhiza glabra	CONSENT FOI		FOD				
GABA	=	γ-aminobutyric acid	CONSENT FOR PUBLICATION						
GAD	=	Glutamate Decarboxylase	INC	Not applicable.					
GB	=	Ginkgo biloba	FUNDING						
GFAP	=	Glial Fibrillary Acid Protein	None.						
GLUT1	=	Glucose Transporter1	CONFLICT OF INTEREST						
GN	=	Galanthus nivalis	Th	The authors declare no conflict of interest, financia					
HD	=	Huntington's Disease	otherv	omerwise.					
HDSA	=	Huntington's Disease Society of America	ACKNOWLEDGEMENTS						
HPLC	=	High-Performance Liquid Chromatography	Declared none.						
HTRF	=	Homogeneous Time Resolved Fluorescenc	REFERENCES						
JNK	=	c-Jun terminal kinase	[1]	Walker	r, F.O.	Huntington's disease. Lancet, 2007, 369(9557), 218-			
LDL	=	Low Density Lipoprotein		228. http://d	lx.doi.c	org/10.1016/S0140-6736(07)60111-1 PMID:			
LME	=	Lycopene loaded microemulsions	[2] Medicalnewstoday, what you need to know about Huntington's						
LOO	=	Lycopene dissolved in Olive Oil	 disease. https://www.medicalnewstoday.com/articles/159552 [3] Dey, A.; Bhattacharya, R.; Mukherjee, A.; Pandey, D.K. Natural products against Alzheimer's disease: Pharmaco-therapeutics and biotechnological interventions. <i>Biotechnol. Adv.</i>, 2017, 35(2), 178- 						
LS	=	Lycopodium Serratum							
MDA	=	Malondialdehyde		216. http://dx.doi.org/10.1016/i.biotechadv.2016.12 005 PM					
ME	= Microemulsion		[4]	280438	043897 Idin M.S.: Tewari, D.: Mamun, A.A.: Kabir, M.T.: Niaz, K.:				
mHTT	= Mutant Huntingtin			Wahed	d, M.I.I.; Barreto, G.E.; Ashraf, G.M. Circadian and sleep nction in Alzheimer's disease <i>Agoing Res Population 2020</i>				
MS	=	Mass Spectrometry		101046	dystunction in Alzheimer's disease. Ageing Res. Rev., 2020 , 60, 101046.				

http://dx.doi.org/10.1016/j.arr.2020.101046 PMID: 32171783

- [5] Uddin, M.S.; Kabir, M.T.; Tewari, D.; Mamun, A.A.; Mathew, B.; Aleya, L.; Barreto, G.E.; Bin-Jumah, M.N.; Abdel-Daim, M.M.; Ashraf, G.M. Revisiting the role of brain and peripheral Aβ in the pathogenesis of Alzheimer's disease. *J. Neurol. Sci.*, **2020**, *416*, 116974.
 - http://dx.doi.org/10.1016/j.jns.2020.116974 PMID: 32559516
- [6] Uddin, M.S.; Kabir, M.T.; Tewari, D.; Mathew, B.; Aleya, L. Emerging signal regulating potential of small molecule biflavonoids to combat neuropathological insults of Alzheimer's disease. *Sci. Total Environ.*, **2020**, 700, 134836. http://dx.doi.org/10.1016/j.scitotenv.2019.134836 PMID: 31704512
- [7] Anand, A.; Patience, A.A.; Sharma, N.; Khurana, N. The present and future of pharmacotherapy of Alzheimer's disease: A comprehensive review. *Eur. J. Pharmacol.*, **2017**, *815*, 364-375. http://dx.doi.org/10.1016/j.ejphar.2017.09.043 PMID: 28978455
- [8] Szlachcic, W.J.; Switonski, P.M.; Krzyzosiak, W.J.; Figlerowicz, M.; Figiel, M. Huntington disease iPSCs show early molecular changes in intracellular signaling, the expression of oxidative stress proteins and the p53 pathway. *Dis. Model. Mech.*, **2015**, *8*(9), 1047-1057.
 - http://dx.doi.org/10.1242/dmm.019406 PMID: 26092128 Warby, S.C.; Montpetit, A.; Hayden, A.R.; Carroll, J.B.; Butland,

[9]

- S.L.; Visscher, H.; Collins, J.A.; Semaka, A.; Hudson, T.J.; Hayden, M.R. CAG expansion in the Huntington disease gene is associated with a specific and targetable predisposing haplogroup. *Am. J. Hum. Genet.*, **2009**, *84*(3), 351-366. http://dx.doi.org/10.1016/j.ajhg.2009.02.003 PMID: 19249009
- [10] Li, X.; Valencia, A.; Sapp, E.; Masso, N.; Alexander, J.; Reeves, P.; Kegel, K.B.; Aronin, N.; Difiglia, M. Aberrant Rab11dependent trafficking of the neuronal glutamate transporter EAAC1 causes oxidative stress and cell death in Huntington's disease. J. Neurosci., 2010, 30(13), 4552-4561. http://dx.doi.org/10.1523/JNEUROSCI.5865-09.2010 PMID: 20357106
- [11] Choudhary, S.; Kumar, P.; Malik, J. Plants and phytochemicals for Huntington's disease. *Pharmacogn. Rev.*, 2013, 7(14), 81-91. http://dx.doi.org/10.4103/0973-7847.120505 PMID: 24347915
- [12] Dey, A.; Nandy, S.; Mukherjee, A.; Pandey, D.K. Plant Natural Products as Neuroprotective Nutraceuticals: Preclinical and Clinical Studies and Future Implications. *Proc. Natl. Acad. Sci. India Sect B: Biol. Sci.* 2020, pp. 1-15.
- [13] Pareek, H.; Thakur, P.; Ray, D. Modeling and docking studies of 4aminobutyrate aminotransferase for huntington's disease. *Int. J. Pharma Bio Sci.*, 2011, 2, 539-549.
- [14] Kay, C.; Hayden, M.R.; Leavitt, B.R. Epidemiology of Huntington disease. *Handbook of Clinical Neurology*; Elsevier, 2017, Vol. 144, pp. 31-46.
- [15] Uddin, M.S.; Tewari, D.; Sharma, G.; Kabir, M.T.; Barreto, G.E.; Bin-Jumah, M.N.; Perveen, A.; Abdel-Daim, M.M.; Ashraf, G.M. Molecular mechanisms of ER stress and UPR in the pathogenesis of Alzheimer's disease. *Mol. Neurobiol.*, **2020**, *57*(7), 2902-2919. http://dx.doi.org/10.1007/s12035-020-01929-y PMID: 32430843
- [16] Craufurd, D.; Thompson, J.C.; Snowden, J.S. Behavioral changes in Huntington disease. *Neuropsychiatry Neuropsychol. Behav. Neurol.*, 2001, 14(4), 219-226. PMID: 11725215
- [17] Tabrizi, S.J.; Langbehn, D.R.; Leavitt, B.R.; Roos, R.A.; Durr, A.; Craufurd, D.; Kennard, C.; Hicks, S.L.; Fox, N.C.; Scahill, R.I.; Borowsky, B.; Tobin, A.J.; Rosas, H.D.; Johnson, H.; Reilmann, R.; Landwehrmeyer, B.; Stout, J.C. TRACK-HD investigators. Biological and clinical manifestations of Huntington's disease in the longitudinal TRACK-HD study: cross-sectional analysis of baseline data. *Lancet Neurol.*, **2009**, 8(9), 791-801. http://dx.doi.org/10.1016/S1474-4422(09)70170-X PMID: 19646924
- [18] Tabrizi, S.J.; Scahill, R.I.; Owen, G.; Durr, A.; Leavitt, B.R.; Roos, R.A.; Borowsky, B.; Landwehrmeyer, B.; Frost, C.; Johnson, H.; Craufurd, D.; Reilmann, R.; Stout, J.C.; Langbehn, D.R. TRACK-HD Investigators. Predictors of phenotypic progression and disease onset in premanifest and early-stage Huntington's disease in the TRACK-HD study: analysis of 36-month observational data. *Lancet Neurol.*, 2013, 12(7), 637-649.

http://dx.doi.org/10.1016/S1474-4422(13)70088-7 PMID: 23664844

- [19] Dugger, B.N.; Dickson, D.W. Pathology of neurodegenerative diseases. *Cold Spring Harb. Perspect. Biol.*, 2017, 9(7), a028035. http://dx.doi.org/10.1101/cshperspect.a028035 PMID: 28062563
- [20] Clinart, M.E.N.A. Rare Disease Clinical Research Untapped potential in MENA. https://longtaal.com/wpcontent/uploads/2019/05/Rare-Disease-Clinical-Research-Untapped-potential-in-MENA.pdf2020.
- [21] Sheikh, S.; Haque, E.; Mir, S.S. Neurodegenerative diseases: multifactorial conformational diseases and their therapeutic interventions. J. Neurodegen. Dis., 2013, 2013,563481. http://dx.doi.org/10.1155/2013/563481
- [22] Bawari, S.; Tewari, D.; Argüelles, S.; Sah, A.N.; Nabavi, S.F.; Xu, S.; Vacca, R.A.; Nabavi, S.M.; Shirooie, S. Targeting BDNF signaling by natural products: Novel synaptic repair therapeutics for neurodegeneration and behavior disorders. *Pharmacol. Res.*, 2019, *148*, 104458.

http://dx.doi.org/10.1016/j.phrs.2019.104458 PMID: 31546015

- [23] Jayakar, S.S.; Dikshit, M. AMPA receptor regulation mechanisms: future target for safer neuroprotective drugs. *Int. J. Neurosci.*, 2004, 114(6), 695-734.
 - http://dx.doi.org/10.1080/00207450490430453 PMID: 15204061
- [24] Rubinsztein, D.C.; Leggo, J.; Coles, R.; Almqvist, E.; Biancalana, V.; Cassiman, J-J.; Chotai, K.; Connarty, M.; Crauford, D.; Curtis, A.; Curtis, D.; Davidson, M.J.; Differ, A.M.; Dode, C.; Dodge, A.; Frontali, M.; Ranen, N.G.; Stine, O.C.; Sherr, M.; Abbott, M.H.; Franz, M.L.; Graham, C.A.; Harper, P.S.; Hedreen, J.C.; Hayden, M.R. Phenotypic characterization of individuals with 30-40 CAG repeats in the Huntington disease (HD) gene reveals HD cases with 36 repeats and apparently normal elderly individuals with 36-39 repeats. Am. J. Hum. Genet., 1996, 59(1), 16-22. PMID: 8659522
- [25] Gusella, J.F.; MacDonald, M.E.; Lee, J.M. Genetic modifiers of Huntington's disease. *Mov. Disord.*, **2014**, *29*(11), 1359-1365. http://dx.doi.org/10.1002/mds.26001 PMID: 25154728
- [26] Rosenblatt, A.; Kumar, B.V.; Mo, A.; Welsh, C.S.; Margolis, R.L.; Ross, C.A. Age, CAG repeat length, and clinical progression in Huntington's disease. *Mov. Disord.*, **2012**, *27*(2), 272-276. http://dx.doi.org/10.1002/mds.24024 PMID: 22173986
- [27] Leenders, K.L.; Frackowiak, R.S.; Quinn, N.; Marsden, C.D. Brain energy metabolism and dopaminergic function in Huntington's disease measured *in vivo* using positron emission tomography. *Mov. Disord.*, **1986**, *1*(1), 69-77.
 - http://dx.doi.org/10.1002/mds.870010110 PMID: 2973559
- [28] Stahl, W.L.; Swanson, P.D. Biochemical abnormalities in Huntington's chorea brains. *Neurology*, **1974**, *24*(9), 813-819. http://dx.doi.org/10.1212/WNL.24.9.813 PMID: 4277376
- [29] Cramer, H.; Warter, J.M.; Renaud, B. Analysis of neurotransmitter metabolites and adenosine 3',5'-monophosphate in the CSF of patients with extrapyramidal motor disorders. *Adv. Neurol.*, 1984, 40, 431-435.
 PMID: 6198888
- [30] Gines, S.; Seong, I.S.; Fossale, E.; Ivanova, E.; Trettel, F.; Gusella, J.F.; Wheeler, V.C.; Persichetti, F.; MacDonald, M.E. Specific progressive cAMP reduction implicates energy deficit in presymptomatic Huntington's disease knock-in mice. *Hum. Mol. Genet.*, 2003, 12(5), 497-508.

http://dx.doi.org/10.1093/hmg/ddg046 PMID: 12588797

- [31] Koroshetz, W.J.; Jenkins, B.G.; Rosen, B.R.; Beal, M.F. Energy metabolism defects in Huntington's disease and effects of coenzyme Q10. Ann. Neurol., 1997, 41(2), 160-165. http://dx.doi.org/10.1002/ana.410410206 PMID: 9029064
- [32] Jenkins, B.G.; Koroshetz, W.J.; Beal, M.F.; Rosen, B.R. Evidence for impairment of energy metabolism *in vivo* in Huntington's disease using localized 1H NMR spectroscopy. *Neurology*, **1993**, 43(12), 2689-2695.

http://dx.doi.org/10.1212/WNL.43.12.2689 PMID: 8255479

- [33] Fatokun, A.A.; Smith, R.A.; Stone, T.W. Resistance to kynurenic acid of the NMDA receptor-dependent toxicity of 3-nitropropionic acid and cyanide in cerebellar granule neurons. *Brain Res.*, 2008, 1215, 200-207.
- http://dx.doi.org/10.1016/j.brainres.2008.04.013 PMID: 18486115 [34] Huang, Q-Y.; Yu, L.; Ferrante, R.J.; Chen, J-F. Mutant
- SOD1G93A in bone marrow-derived cells exacerbates 3-

nitropropionic acid induced striatal damage in mice. *Neurosci. Lett.*, **2007**, *418*(2), 175-180. http://dx.doi.org/10.1016/j.neulet.2007.03.038 PMID: 17418947

- [35] Alexi, T.; Borlongan, C.V.; Faull, R.L.; Williams, C.E.; Clark, R.G.; Gluckman, P.D.; Hughes, P.E. Neuroprotective strategies for basal ganglia degeneration: Parkinson's and Huntington's diseases. *Prog. Neurobiol.*, 2000, 60(5), 409-470. http://dx.doi.org/10.1016/S0301-0082(99)00032-5 PMID: 10697073
- [36] Alexi, T.; Hughes, P.E.; Faull, R.L.; Williams, C.E. 3-Nitropropionic acid's lethal triplet: cooperative pathways of neurodegeneration. *Neuroreport*, **1998**, *9*(11), R57-R64. http://dx.doi.org/10.1097/00001756-199808030-00001 PMID: 9721909
- [37] Farooqui, T.; Farooqui, A.A. Aging: an important factor for the pathogenesis of neurodegenerative diseases. *Mech. Ageing Dev.*, 2009, 130(4), 203-215.
- http://dx.doi.org/10.1016/j.mad.2008.11.006 PMID: 19071157
 [38] Dong, X.X.; Wang, Y.; Qin, Z.H. Molecular mechanisms of excitotoxicity and their relevance to pathogenesis of neurodegenerative diseases. *Acta Pharmacol. Sin.*, 2009, 30(4), 379-387. http://dx.doi.org/10.1038/aps.2009.24 PMID: 19343058
- [39] Bogdanov, M.B.; Andreassen, O.A.; Dedeoglu, A.; Ferrante, R.J.;
 Beal, M.F. Increased oxidative damage to DNA in a transgenic mouse model of Huntington's disease. *J. Neurochem.*, 2001, 79(6), 1246-1249. http://dx.doi.org/10.1046/j.1471-4159.2001.00689.x PMID:
 - 11752065] Chen, C-M. Mitochondrial dysfunction, metabolic deficits, and
- [40] Chen, C-M. Mitochondrial dysfunction, metabolic deficits, and increased oxidative stress in Huntington's disease. *Chang Gung Med. J.*, 2011, 34(2), 135-152. PMID: 21539755
- [41] Browne, S.E.; Ferrante, R.J.; Beal, M.F. Oxidative stress in Huntington's disease. *Brain Pathol.*, **1999**, *9*(1), 147-163. http://dx.doi.org/10.1111/j.1750-3639.1999.tb00216.x PMID: 9989457
- Sawa, A.; Tomoda, T.; Bae, B-I. Mechanisms of neuronal cell death in Huntington's disease. *Cytogenet. Genome Res.*, 2003, 100(1-4), 287-295. http://dx.doi.org/10.1159/000072864 PMID: 14526190
- [43] Túnez, I.; Tasset, I.; Pérez-De La Cruz, V.; Santamaría, A. 3-Nitropropionic acid as a tool to study the mechanisms involved in Huntington's disease: past, present and future. *Molecules*, 2010, *15*(2), 878-916. http://dx.doi.org/10.3390/molecules15020878 PMID: 20335954
- [44] Shirasaki, D.I.; Greiner, E.R.; Al-Ramahi, I.; Gray, M.; Boontheung, P.; Geschwind, D.H.; Botas, J.; Coppola, G.; Horvath, S.; Loo, J.A.; Yang, X.W. Network organization of the huntingtin proteomic interactome in mammalian brain. *Neuron*, 2012, 75(1), 41-57. http://dx.doi.org/10.1016/j.neuron.2012.05.024 PMID: 22794259
- [45] Weiss, A.; Träger, U.; Wild, E.J.; Grueninger, S.; Farmer, R.; Landles, C.; Scahill, R.I.; Lahiri, N.; Haider, S.; Macdonald, D.; Frost, C.; Bates, G.P.; Bilbe, G.; Kuhn, R.; Andre, R.; Tabrizi, S.J. Mutant huntingtin fragmentation in immune cells tracks Huntington's disease progression. J. Clin. Invest., 2012, 122(10), 3731-3736. http://dx.doi.org/10.1172/JCI64565 PMID: 22996692
- [46] Weir, D.W.; Sturrock, A.; Leavitt, B.R. Development of biomarkers for Huntington's disease. *Lancet Neurol.*, 2011, 10(6), 573-590. http://dx.doi.org/10.1016/S1474-4422(11)70070-9 PMID:
- 21601164
 [47] Russell, D.S.; Jennings, D.L.; Barret, O.; Tamagnan, G.D.; Carroll, V.M.; Caillé, F.; Alagille, D.; Morley, T.J.; Papin, C.; Seibyl, J.P.; Marek, K.L. Change in PDE10 across early Huntington disease assessed by [18F]MNI-659 and PET imaging. *Neurology*, 2016, 86(8), 748-754. http://dx.doi.org/10.1212/WNL.00000000002391 PMID: 26802091
- [48] Tabrizi, S.J.; Reilmann, R.; Roos, R.A.; Durr, A.; Leavitt, B.; Owen, G.; Jones, R.; Johnson, H.; Craufurd, D.; Hicks, S.L.; Kennard, C.; Landwehrmeyer, B.; Stout, J.C.; Borowsky, B.; Scahill, R.I.; Frost, C.; Langbehn, D.R. TRACK-HD investigators. Potential endpoints for clinical trials in premanifest and early Huntington's disease in the TRACK-HD study: analysis of 24 month observational data. *Lancet Neurol.*, 2012, 11(1), 42-53.

http://dx.doi.org/10.1016/S1474-4422(11)70263-0 PMID: 22137354

[49] Braisch, U.; Muche, R.; Rothenbacher, D.; Landwehrmeyer, G.B.; Long, J.D.; Orth, M. REGISTRY Investigators of the European Huntington's Disease Network and COHORT Investigators of the Huntington Study Group. Identification of symbol digit modality test score extremes in Huntington's disease. *Am. J. Med. Genet. B. Neuropsychiatr. Genet.*, **2019**, *180*(3), 232-245.

http://dx.doi.org/10.1002/ajmg.b.32719 PMID: 30788902

- [50] Björkqvist, M.; Wild, E.J.; Thiele, J.; Silvestroni, A.; Andre, R.; Lahiri, N.; Raibon, E.; Lee, R.V.; Benn, C.L.; Soulet, D.; Magnusson, A.; Woodman, B.; Landles, C.; Pouladi, M.A.; Hayden, M.R.; Khalili-Shirazi, A.; Lowdell, M.W.; Brundin, P.; Bates, G.P.; Leavitt, B.R.; Möller, T.; Tabrizi, S.J. A novel pathogenic pathway of immune activation detectable before clinical onset in Huntington's disease. J. Exp. Med., 2008, 205(8), 1869-1877. http://dx.doi.org/10.1084/jem.20080178 PMID: 18625748
- [51] Rodrigues, F.B.; Byrne, L.M.; McColgan, P.; Robertson, N.; Tabrizi, S.J.; Zetterberg, H.; Wild, E.J. Cerebrospinal fluid inflammatory biomarkers reflect clinical severity in Huntington's disease. *PLoS One*, **2016**, *11*(9), e0163479.
 - http://dx.doi.org/10.1371/journal.pone.0163479 PMID: 27657730
- [52] Southwell, A.L.; Smith, S.E.; Davis, T.R.; Caron, N.S.; Villanueva, E.B.; Xie, Y.; Collins, J.A.; Ye, M.L.; Sturrock, A.; Leavitt, B.R.; Schrum, A.G.; Hayden, M.R. Ultrasensitive measurement of huntingtin protein in cerebrospinal fluid demonstrates increase with Huntington disease stage and decrease following brain huntingtin suppression. *Sci. Rep.*, **2015**, *5*(1), 12166. http://dx.doi.org/10.1038/srep12166 PMID: 26174131
- [53] Vinther-Jensen, T.; Budtz-Jørgensen, E.; Simonsen, A.H.; Nielsen, J.E.; Hjermind, L.E. YKL-40 in cerebrospinal fluid in Huntington's disease--a role in pathology or a nonspecific response to inflammation? *Parkinsonism Relat. Disord.*, 2014, 20(11), 1301-1303. http://dx.doi.org/10.1016/j.parkreldis.2014.08.011 PMID: 25219973
- [54] Constantinescu, R.; Romer, M.; Oakes, D.; Rosengren, L.; Kieburtz, K. Levels of the light subunit of neurofilament triplet protein in cerebrospinal fluid in Huntington's disease. *Parkinsonism Relat. Disord.*, 2009, 15(3), 245-248. http://dx.doi.org/10.1016/j.parkreldis.2008.05.012 PMID: 19056308
- [55] Nicoli, F.; Vion-Dury, J.; Maloteaux, J-M.; Delwaide, C.; Confort-Gouny, S.; Sciaky, M.; Cozzone, P.J. CSF and serum metabolic profile of patients with Huntington's chorea: a study by high resolution proton NMR spectroscopy and HPLC. *Neurosci. Lett.*, **1993**, *154*(1-2), 47-51.
- http://dx.doi.org/10.1016/0304-3940(93)90168-K PMID: 8361646
 [56] Saelens, J.K.; Allen, M.P.; Simke, J.P. Determination of acetylcholine and choline by an enzymatic assay. *Arch. Int. Pharmacodyn. Ther.*, **1970**, *186*(2), 279-286.
 PMID: 4331694
- [57] Jeitner, T.M.; Bogdanov, M.B.; Matson, W.R.; Daikhin, Y.; Yudkoff, M.; Folk, J.E.; Steinman, L.; Browne, S.E.; Beal, M.F.; Blass, J.P.; Cooper, A.J. N(ε)-(γ-L-glutamyl)-L-lysine (GGEL) is increased in cerebrospinal fluid of patients with Huntington's disease. J. Neurochem., 2001, 79(5), 1109-1112. http://dx.doi.org/10.1046/j.1471-4159.2001.00673.x PMID: 11739625
- [58] Jeitner, T.M.; Matson, W.R.; Folk, J.E.; Blass, J.P.; Cooper, A.J. Increased levels of γ-glutamylamines in Huntington disease CSF. J. Neurochem., 2008, 106(1), 37-44. http://dx.doi.org/10.1111/j.1471-4159.2008.05350.x PMID: 18422943
- [59] Guay, D.R. Tetrabenazine, a monoamine-depleting drug used in the treatment of hyperkinetic movement disorders. *Am. J. Geriatr. Pharmacother.*, **2010**, 8(4), 331-373. http://dx.doi.org/10.1016/j.amjopharm.2010.08.006 PMID: 20869622
- [60] Pidgeon, C.; Rickards, H. The pathophysiology and pharmacological treatment of Huntington disease. *Behav. Neurol.*, 2013, 26(4), 245-253.

http://dx.doi.org/10.1155/2013/705373 PMID: 22713409

[61] Kumar, P.; Kumar, A. Protective effect of rivastigmine against 3nitropropionic acid-induced Huntington's disease like symptoms: possible behavioural, biochemical and cellular alterations. *Eur. J. Pharmacol.*, **2009**, *615*(1-3), 91-101. http://dx.doi.org/10.1016/j.ejphar.2009.04.058 PMID: 19445928

- [62] Park, J-E.; Lee, S-T.; Im, W.S.; Chu, K.; Kim, M. Galantamine reduces striatal degeneration in 3-nitropropionic acid model of Huntington's disease. *Neurosci. Lett.*, **2008**, *448*(1), 143-147. http://dx.doi.org/10.1016/j.neulet.2008.10.020 PMID: 18938211
- [63] Besret, L.; Page, K.J.; Dunnett, S.B. The problem of antipsychotic treatment for functional imaging in Huntington's disease: receptor binding, gene expression and locomotor activity after sub-chronic administration and wash-out of haloperidol in the rat. *Brain Res.*, 2000, 853(1), 125-135. http://dx.doi.org/10.1016/S0006-8993(99)02311-2 PMID: 10627316
- [64] Wyant, K.J.; Ridder, A.J.; Dayalu, P. Huntington's disease update on treatments. *Curr. Neurol. Neurosci. Rep.*, 2017, 17(4), 33.
- http://dx.doi.org/10.1007/s11910-017-0739-9 PMID: 28324302
- [65] Iriti, M.; Vitalini, S.; Fico, G.; Faoro, F. Neuroprotective herbs and foods from different traditional medicines and diets. *Molecules*, 2010, 15(5), 3517-3555.
- http://dx.doi.org/10.3390/molecules15053517 PMID: 20657497
 [66] Ratheesh, G.; Tian, L.; Venugopal, J.R.; Ezhilarasu, H.; Sadiq, A.; Fan, T-P.; Ramakrishna, S. Role of medicinal plants in neurodegenerative diseases. *Biomanufacturing Rev.*, 2017, 2(1), 2. http://dx.doi.org/10.1007/s40898-017-0004-7
- [67] Danduga, R.C.S.R.; Dondapati, S.R.; Kola, P.K.; Grace, L.; Tadigiri, R.V.B.; Kanakaraju, V.K. Neuroprotective activity of tetramethylpyrazine against 3-nitropropionic acid induced Huntington's disease-like symptoms in rats. *Biomed. Pharmacother.*, **2018**, *105*, 1254-1268.
- http://dx.doi.org/10.1016/j.biopha.2018.06.079 PMID: 30021362
 [68] Nikolaus, S.; Wittsack, H-J.; Beu, M.; Antke, C.; Hautzel, H.; Wickrath, F.; Müller-Lutz, A.; De Souza Silva, M.A.; Huston, J.P.; Antoch, G.; Müller, H.W. Amantadine enhances nigrostriatal and mesolimbic dopamine function in the rat brain in relation to motor and exploratory activity. *Pharmacol. Biochem. Behav.*, 2019, 179, 156-170.

http://dx.doi.org/10.1016/j.pbb.2018.12.010 PMID: 30639878

[69] Zesiewicz, T.A.; Sullivan, K.L.; Hauser, R.A.; Sanchez-Ramos, J. Open-label pilot study of levetiracetam (Keppra) for the treatment of chorea in Huntington's disease. *Mov. Disord.*, 2006, 21(11), 1998-2001.

http://dx.doi.org/10.1002/mds.21061 PMID: 16941461

[70] Khursheed, R.; Singh, S.K.; Wadhwa, S.; Gulati, M.; Awasthi, A. Enhancing the potential preclinical and clinical benefits of quercetin through novel drug delivery systems. *Drug Discov. Today*, 2019.

PMID: 31707120

- [71] Malakhova, A.A.; Grigor'eva, E.V.; Malankhanova, T.B.; Pavlova, S.V.; Valetdinova, K.R.; Abramycheva, N.Y.; Vetchinova, A.S.; Illarioshkin, S.N.; Zakian, S.M. Generation of induced pluripotent stem cell line ICGi018-A from peripheral blood mononuclear cells of a patient with Huntington's disease. *Stem Cell Res. (Amst.)*, **2020**, *44*, 101743. http://dx.doi.org/10.1016/j.scr.2020.101743 PMID: 32179492
- [72] Rotundo, G.; Bidollari, E.; Ferrari, D.; Spasari, I.; Bernardini, L.; Consoli, F.; De Luca, A.; Santimone, I.; Lamorte, G.; Migliore, S.; Squitieri, F.; Vescovi, A.L.; Rosati, J. Generation of the induced pluripotent stem cell line CSSi006-A (3681) from a patient affected by advanced-stage Juvenile Onset Huntington's Disease. *Stem Cell Res. (Amst.)*, 2018, 29, 174-178.

http://dx.doi.org/10.1016/j.scr.2018.04.008 PMID: 29704769

- [73] Bidollari, E.; Rotundo, G.; Ferrari, D.; Candido, O.; Bernardini, L.; Consoli, F.; De Luca, A.; Santimone, I.; Lamorte, G.; Ilari, A.; Squitieri, F.; Vescovi, A.L.; Rosati, J. Generation of induced pluripotent stem cell line, CSSi004-A (2962), from a patient diagnosed with Huntington's disease at the presymptomatic stage. *Stem Cell Res. (Amst.)*, **2018**, *28*, 145-148. http://dx.doi.org/10.1016/j.scr.2018.02.014 PMID: 29486399
- [74] Dumevska, B.; Schaft, J.; McKernan, R.; Hu, J.; Schmidt, U. Derivation of Huntington Disease affected Genea090 human embryonic stem cell line. *Stem Cell Res. (Amst.)*, 2016, *16*(2), 519-521. http://dx.doi.org/10.1016/j.scr.2016.02.024 PMID: 27346026

- [75] Wang, S-E.; Lin, C-L.; Hsu, C-H.; Sheu, S-J.; Chien, C-T.; Wu, C-H. Treatment with a herbal formula B401 enhances neuroprotection and angiogenesis in the R6/2 mouse model of Huntington's disease. *Drug Des. Devel. Ther.*, **2015**, *9*, 887-900. PMID: 25733809
- [76] Sandhir, R.; Yadav, A.; Mehrotra, A.; Sunkaria, A.; Singh, A.; Sharma, S. Curcumin nanoparticles attenuate neurochemical and neurobehavioral deficits in experimental model of Huntington's disease. *Neuromol. Med.*, 2014, 16(1), 106-118. http://dx.doi.org/10.1007/s12017-013-8261-y PMID: 24008671
- [77] Rao, R.V.; Descamps, O.; John, V.; Bredesen, D.E. Ayurvedic medicinal plants for Alzheimer's disease: a review. *Alzheimers Res. Ther.*, 2012, 4(3), 22. http://dx.doi.org/10.1186/alzrt125 PMID: 22747839
- [78] Wu, P.F.; Zhang, Z.; Wang, F.; Chen, J.G. Natural compounds from traditional medicinal herbs in the treatment of cerebral ischemia/reperfusion injury. *Acta Pharmacol. Sin.*, **2010**, *31*(12), 1523-1531.

http://dx.doi.org/10.1038/aps.2010.186 PMID: 21127495

- [79] Sandhya, S.; Vinod, K.; Kumar, S. Herbs used for brain disorders. *Hygeia J Drugs Med*, 2010, 2, 38-45.
- [80] Wang, D.; Calabrese, E.J.; Lian, B.; Lin, Z.; Calabrese, V. Hormesis as a mechanistic approach to understanding herbal treatments in traditional Chinese medicine. *Pharmacol. Ther.*, **2018**, *184*, 42-50. http://dx.doi.org/10.1016/j.pharmthera.2017.10.013 PMID: 29080703
- [81] Geng, Y.; Li, C.; Liu, J.; Xing, G.; Zhou, L.; Dong, M.; Li, X.; Niu, Y. Beta-asarone improves cognitive function by suppressing neuronal apoptosis in the beta-amyloid hippocampus injection rats. *Biol. Pharm. Bull.*, 2010, 33(5), 836-843. http://dx.doi.org/10.1248/bpb.33.836 PMID: 20460763
- [82] Paterna, J-C.; Leng, A.; Weber, E.; Feldon, J.; Büeler, H. DJ-1 and Parkin modulate dopamine-dependent behavior and inhibit MPTPinduced nigral dopamine neuron loss in mice. *Mol. Ther.*, 2007, 15(4), 698-704.

http://dx.doi.org/10.1038/sj.mt.6300067 PMID: 17299411

- [83] Bayan, L.; Koulivand, P.H.; Gorji, A. Garlic: a review of potential therapeutic effects. Avicenna J. Phytomed., 2014, 4(1), 1-14. PMID: 25050296
- [84] Chauhan, N.B. Effect of aged garlic extract on APP processing and tau phosphorylation in Alzheimer's transgenic model Tg2576. J. Ethnopharmacol., 2006, 108(3), 385-394. http://dx.doi.org/10.1016/j.jep.2006.05.030 PMID: 16842945
- [85] Tatara, M.R.; Sliwa, E.; Dudek, K.; Mosiewicz, J.; Studzinski, T. Effect of aged garlic extract and allicin administration to sows during pregnancy and lactation on body weight gain and gastrointestinal tract development of piglets. PART I. Bull. Vet. Inst. Pulawy, 2005, 49(3), 349.
- [86] Pérez-Torres, I.; Torres-Narváez, J.C.; Pedraza-Chaverri, J.; Rubio-Ruiz, M.E.; Díaz-Diaz, E.; Del Valle-Mondragón, L.; Martínez-Memije, R.; Varela, L.E.; Guarner-Lans, V. Effect of the aged garlic extract on cardiovascular function in metabolic syndrome rats. *Molecules*, 2016, 21(11), 1425.

http://dx.doi.org/10.3390/molecules21111425 PMID: 27792195

- [87] Medina-Campos, O.N.; Barrera, D.; Segoviano-Murillo, S.; Rocha, D.; Maldonado, P.D.; Mendoza-Patiño, N.; Pedraza-Chaverri, J. Sallylcysteine scavenges singlet oxygen and hypochlorous acid and protects LLC-PK(1) cells of potassium dichromate-induced toxicity. *Food Chem. Toxicol.*, 2007, 45(10), 2030-2039.
- http://dx.doi.org/10.1016/j.fct.2007.05.002 PMID: 17576034
 [88] Roy, S.; Awasthi, H. Herbal medicines as neuroprotective agent: A mechanistic approach. *Int. J. Pharm. Pharm. Sci.*, 2017, 9, 1-7. http://dx.doi.org/10.22159/ijpps.2017v9i11.19444
- [89] Gohil, K.J.; Patel, J.A. A review on Bacopa monniera: current research and future prospects. *Intl. J. Green Pharmacy*, 2010, 4(1). http://dx.doi.org/10.4103/0973-8258.62156
- [90] Russo, A.; Borrelli, F. Bacopa monniera, a reputed nootropic plant: an overview. *Phytomedicine*, **2005**, *12*(4), 305-317. http://dx.doi.org/10.1016/j.phymed.2003.12.008 PMID: 15898709
- [91] Nandy, S.; Mukherjee, A.; Pandey, D.K.; Dey, A. Bacopa monnieri: The neuroprotective elixir from the East—Phytochemistry, pharmacology, and biotechnological improvement. Bioactive Natural products in Drug Discovery; Springer, 2020, pp. 97-126. http://dx.doi.org/10.1007/978-981-15-1394-7 2

- [92] Dey, A.; Hazra, A.; Nongdam, P.; Nandy, S.; Tikendra, L.; Mukherjee, A.; Banerjee, S.; Mukherjee, S.; Pandey, D. Enhanced bacoside content in polyamine treated *in-vitro* raised *Bacopa monnieri* (L.) Wettst. S. Afr. J. Bot., **2019**, *123*, 259-269. http://dx.doi.org/10.1016/j.sajb.2019.03.012
- [93] Shinomol, G.K.; Muralidhara, Bacopa monnieri modulates endogenous cytoplasmic and mitochondrial oxidative markers in prepubertal mice brain. *Phytomedicine*, **2011**, *18*(4), 317-326. http://dx.doi.org/10.1016/j.phymed.2010.08.005 PMID: 20850955
- [94] Calabrese, C.; Gregory, W.L.; Leo, M.; Kraemer, D.; Bone, K.; Oken, B. Effects of a standardized Bacopa monnieri extract on cognitive performance, anxiety, and depression in the elderly: a randomized, double-blind, placebo-controlled trial. J. Altern. Complement. Med., 2008, 14(6), 707-713. http://dx.doi.org/10.1089/acm.2008.0018 PMID: 18611150
- [95] Bammidi, S.R.; Volluri, S.S.; Chippada, S.C.; Avanigadda, S.; Vangalapati, M. A review on pharmacological studies of *Bacopa* monniera. J. Chem. Biol. Phys. Sci., 2011, 1(2), 250.
- [96] Hou, C-C.; Lin, S-J.; Cheng, J-T.; Hsu, F-L. Bacopaside III, bacopasaponin G, and bacopasides A, B, and C from *Bacopa monniera*. *J. Nat. Prod.*, **2002**, *65*(12), 1759-1763. http://dx.doi.org/10.1021/np020238w PMID: 12502309
- [97] Mahato, S.B.; Garai, S.; Chakravarty, A.K. Bacopasaponins E and F: two jujubogenin bisdesmosides from *Bacopa monniera*. *Phyto-chemistry*, 2000, 53(6), 711-714. http://dx.doi.org/10.1016/S0031-9422(99)00384-2 PMID: 10746885
- [98] Garai, S.; Mahato, S.B.; Ohtani, K.; Yamasaki, K. Dammaranetype triterpenoid saponins from *Bacopa monniera*. *Phytochemistry*, **1996**, 42(3), 815-820. http://dx.doi.org/10.1016/0031-9422(95)00936-1 PMID: 8768327
- [99] Garai, S.; Mahato, S.B.; Ohtani, K.; Yamasaki, K. Bacopasaponin D--a pseudojujubogenin glycoside from *Bacopa monniera*. *Phyto-chemistry*, **1996**, *43*(2), 447-449.
- http://dx.doi.org/10.1016/0031-9422(96)00250-6 PMID: 8862037
 [100] Kawai, K-I.; Shibata, S. Pseudojujubogenin, a new sapogenin from *Bacopa monniera*. *Phytochemistry*, **1978**, *17*(2), 287-289.
- http://dx.doi.org/10.1016/S0031-9422(00)94166-9
 [101] Murthy, P.B.S.; Raju, V.R.; Ramakrisana, T.; Chakravarthy, M.S.; Kumar, K.V.; Kannababu, S.; Subbaraju, G.V. Estimation of twelve bacopa saponins in Bacopa monnieri extracts and formulations by high-performance liquid chromatography. Chem. Pharm. Bull. (Tokyo), 2006, 54(6), 907-911.

http://dx.doi.org/10.1248/cpb.54.907 PMID: 16755069
[102] Chakravarty, A.K.; Garai, S.; Masuda, K.; Nakane, T.; Kawahara, N. Bacopasides III-V: three new triterpenoid glycosides from *Bacopa monniera*. *Chem. Pharm. Bull. (Tokyo)*, **2003**, *51*(2), 215-217. http://dx.doi.org/10.1248/cpb.51.215 PMID: 12576661

- [103] Chakravarty, A.K.; Sarkar, T.; Masuda, K.; Shiojima, K.; Nakane, T.; Kawahara, N. Bacopaside I and II: two pseudojujubogenin glycosides from *Bacopa monniera*. *Phytochemistry*, **2001**, *58*(4), 553-556.
 - http://dx.doi.org/10.1016/S0031-9422(01)00275-8 PMID: 11576596
- [104] Kapoor, R.; Srivastava, S.; Kakkar, P. Bacopa monnieri modulates antioxidant responses in brain and kidney of diabetic rats. Environ. Toxicol. Pharmacol., 2009, 27(1), 62-69.
- http://dx.doi.org/10.1016/j.etap.2008.08.007 PMID: 21783922
 [105] Vollala, V.R.; Upadhya, S.; Nayak, S. Learning and memoryenhancing effect of *Bacopa monniera* in neonatal rats. *Bratisl. Lek Listy*, 2011, *112*(12), 663-669.
 PMID: 22372329
- [106] Roodenrys, S.; Booth, D.; Bulzomi, S.; Phipps, A.; Micallef, C.; Smoker, J. Chronic effects of Brahmi (*Bacopa monnieri*) on human memory. *Neuropsychopharmacology*, 2002, 27(2), 279-281. http://dx.doi.org/10.1016/S0893-133X(01)00419-5 PMID: 12093601
- [107] Singh, H.; Dhawan, B. Neuropsychopharmacological effects of the Ayurvedic nootropic *Bacopa monniera* Linn. (Brahmi). *Indian J. Pharmacol.*, **1997**, *29*(5), 359. PMID: 31831931
- [108] Vohora, D.; Pal, S.N.; Pillai, K.K. Protection from phenytoininduced cognitive deficit by Bacopa monniera, a reputed Indian nootropic plant. J. Ethnopharmacol., 2000, 71(3), 383-390. http://dx.doi.org/10.1016/S0378-8741(99)00213-5 PMID: 10940574

- [109] Stough, C.; Lloyd, J.; Clarke, J.; Downey, L.A.; Hutchison, C.W.; Rodgers, T.; Nathan, P.J. The chronic effects of an extract of *Bacopa monniera* (Brahmi) on cognitive function in healthy human subjects. *Psychopharmacology (Berl.)*, **2001**, *156*(4), 481-484. http://dx.doi.org/10.1007/s002130100815 PMID: 11498727
- [110] Tripathi, Y.B.; Chaurasia, S.; Tripathi, E.; Upadhyay, A.; Dubey, G.P. Bacopa monniera Linn. as an antioxidant: mechanism of action. Indian J. Exp. Biol., 1996, 34(6), 523-526. PMID: 8792640
- [111] Russo, A.; Izzo, A.A.; Borrelli, F.; Renis, M.; Vanella, A. Free radical scavenging capacity and protective effect of *Bacopa monniera L*. on DNA damage. *Phytother. Res.*, **2003**, *17*(8), 870-875. http://dx.doi.org/10.1002/ptr.1061 PMID: 13680815
- [112] Bhattacharya, S.K.; Bhattacharya, A.; Kumar, A.; Ghosal, S. Antioxidant activity of *Bacopa monniera* in rat frontal cortex, striatum and hippocampus. *Phytother. Res.*, **2000**, *14*(3), 174-179. http://dx.doi.org/10.1002/(SICI)1099-1573(200005)14:3<174::AID-PTR624>3.0.CO;2-O PMID: 10815010
- [113] Chowdhuri, D.K.; Parmar, D.; Kakkar, P.; Shukla, R.; Seth, P.K.; Srimal, R.C. Antistress effects of bacosides of *Bacopa monnieri*: modulation of Hsp70 expression, superoxide dismutase and cytochrome P450 activity in rat brain. *Phytother. Res.*, **2002**, *16*(7), 639-645.

http://dx.doi.org/10.1002/ptr.1023 PMID: 12410544

- [114] Sairam, K.; Dorababu, M.; Goel, R.K.; Bhattacharya, S.K. RK. Antidepressant activity of standardized extract of *Bacopa monniera* in experimental models of depression in rats. *Phytomedicine*, 2002, 9(3), 207-211.
 - http://dx.doi.org/10.1078/0944-7113-00116 PMID: 12046860
- [115] Shanker, G.; Singh, H. Anxiolytic profile of standardized Brahmi extract. *Indian J. Pharmacol.*, 2000, 32(152), 5.
- [116] Sumathy, T.; Subramanian, S.; Govindasamy, S.; Balakrishna, K.; Veluchamy, G. Protective role of *Bacopa monniera* on morphine induced hepatotoxicity in rats. *Phytother. Res.*, **2001**, *15*(7), 643-645. http://dx.doi.org/10.1002/ptr.1007 PMID: 11746853
- Sairam, K.; Rao, C.V.; Babu, M.D.; Goel, R.K. Prophylactic and curative effects of *Bacopa monniera* in gastric ulcer models. *Phytomedicine*, 2001, 8(6), 423-430. http://dx.doi.org/10.1078/S0944-7113(04)70060-4 PMID: 11824516
- [118] Andreassen, O.A.; Ferrante, R.J.; Hughes, D.B.; Klivenyi, P.; Dedeoglu, A.; Ona, V.O.; Friedlander, R.M.; Beal, M.F. Malonate and 3-nitropropionic acid neurotoxicity are reduced in transgenic mice expressing a caspase-1 dominant-negative mutant. *J. Neurochem.*, **2000**, 75(2), 847-852. http://dx.doi.org/10.1046/j.1471-4159.2000.0750847.x PMID: 10899963
- [119] Coles, C.J.; Edmondson, D.E.; Singer, T.P. Inactivation of succinate dehydrogenase by 3-nitropropionate. J. Biol. Chem., 1979, 254(12), 5161-5167. PMID: 447637
- [120] Kim, G.W.; Copin, J-C.; Kawase, M.; Chen, S.F.; Sato, S.; Gobbel, G.T.; Chan, P.H. Excitotoxicity is required for induction of oxidative stress and apoptosis in mouse striatum by the mitochondrial toxin, 3-nitropropionic acid. J. Cereb. Blood Flow Metab., 2000, 20(1), 119-129.

http://dx.doi.org/10.1097/00004647-200001000-00016 PMID: 10616800

- [121] Veerendra Kumar, M.H.; Gupta, Y.K. Effect of different extracts of Centella asiatica on cognition and markers of oxidative stress in rats. *J. Ethnopharmacol.*, 2002, 79(2), 253-260. http://dx.doi.org/10.1016/S0378-8741(01)00394-4 PMID: 11801389
- [122] Wijeweera, P.; Arnason, J.T.; Koszycki, D.; Merali, Z. Evaluation of anxiolytic properties of Gotukola--(*Centella asiatica*) extracts and asiaticoside in rat behavioral models. *Phytomedicine*, **2006**, *13*(9-10), 668-676.

http://dx.doi.org/10.1016/j.phymed.2006.01.011 PMID: 16488124 [123] Soumyanath, A.; Zhong, Y.P.; Gold, S.A.; Yu, X.; Koop, D.R.;

Bourdette, D.; Gold, B.G. Centella asiatica accelerates nerve regeneration upon oral administration and contains multiple active fractions increasing neurite elongation *in-vitro*. J. Pharm. Pharmacol., **2005**, 57(9), 1221-1229. http://dx.doi.org/10.1211/jpp.57.9.0018 PMID: 16105244

- [124] Hussin, M.; Abdul-Hamid, A.; Mohamad, S.; Saari, N.; Ismail, M.; Bejo, M.H. Protective effect of *Centella asiatica* extract and powder on oxidative stress in rats. *Food Chem.*, 2007, 100(2), 535-541. http://dx.doi.org/10.1016/j.foodchem.2005.10.022
- [125] Kaur, P.; Pandey, D.K.; Dey, A.; Dwivedi, P.; Malik, T.; Gupta, R. Swertia spp.: A potential source of high-value bioactive components, pharmacology, and analytical techniques. *Bioactive Natural products in Drug Discovery*; Springer, **2020**, pp. 165-213. http://dx.doi.org/10.1007/978-981-15-1394-7 4
- [126] Singh, B.; Rastogi, R. A reinvestigation of the triterpenes of Centella asiatica. Phytochemistry, 1969, 8(5), 917-921. http://dx.doi.org/10.1016/S0031-9422(00)85884-7
- [127] Randriamampionona, D.; Diallo, B.; Rakotoniriana, F.; Rabemanantsoa, C.; Cheuk, K.; Corbisier, A-M.; Mahillon, J.; Ratsimamanga, S.; El Jaziri, M. Comparative analysis of active constituents in *Centella asiatica* samples from Madagascar: application for *ex situ* conservation and clonal propagation. *Fitoterapia*, **2007**, 78(7-8), 482-489.
 - http://dx.doi.org/10.1016/j.fitote.2007.03.016 PMID: 17560738
- [128] Singh, B.; Rastogi, R. Chemical examination of *Centella asiatica* linn—III: Constitution of brahmic acid. *Phytochemistry*, **1968**, 7(8), 1385-1393.
- http://dx.doi.org/10.1016/S0031-9422(00)85642-3
- [129] Asakawa, Y.; Matsuda, R.; Takemoto, T. Mono-and sesquiterpenoids from Hydrocotyle and Centella species. *Phytochemistry*, **1982**, 21(10), 2590-2592.
 - http://dx.doi.org/10.1016/0031-9422(82)85264-3
- [130] Shinomol, G.K.; Muralidhara, Prophylactic neuroprotective property of *Centella asiatica* against 3-nitropropionic acid induced oxidative stress and mitochondrial dysfunctions in brain regions of prepubertal mice. *Neurotoxicology*, **2008**, *29*(6), 948-957. http://dx.doi.org/10.1016/j.neuro.2008.09.009 PMID: 18930762
- [131] Bhat, S.; Kaushal, P.; Kaur, M.; Sharma, H. Coriander (*Corian-drum sativum L.*): Processing, nutritional and functional aspects. *Afr. J. Plant Sci.*, **2014**, 8(1), 25-33. http://dx.doi.org/10.5897/AJPS2013.1118
- [132] Deepa, B.; Anuradha, C.V. Antioxidant potential of *Coriandrum sativum L*. seed extract. *Indian J. Exp. Biol.*, 2011, 49(1), 30-38. PMID: 21365993
- [133] Vekaria, R.H.; Patel, M.N.; Bhalodiya, P.N.; Patel, V.; Desai, T.R.; Tirgar, P.R. Evaluation of neuroprotective effect of *Coriandrum Sativum linn*. against ischemicreperfusion insult in brain. *Intl. J. Phytopharmacol.*, **2012**, *2*, 186-193.
- [134] Liu, Q.F.; Jeong, H.; Lee, J.H.; Hong, Y.K.; Oh, Y.; Kim, Y-M.; Suh, Y.S.; Bang, S.; Yun, H.S.; Lee, K.; Cho, S.M.; Lee, S.B.; Jeon, S.; Chin, Y.W.; Koo, B.S.; Cho, K.S. *Coriandrum sativum* suppresses Aβ42-induced ROS increases, glial cell proliferation, and ERK activation. *Am. J. Chin. Med.*, **2016**, 44(7), 1325-1347. http://dx.doi.org/10.1142/S0192415X16500749 PMID: 27776428
- Singh, A.; Kaur, M.; Choudhary, A.; Kumar, B. Effect of *Butea monosperma* leaf extracts on cyclophosphamide induced clastogenicity and oxidative stress in mice. *Pharmacognosy Res.*, 2015, 7(1), 85-91. http://dx.doi.org/10.4103/0974-8490.147215 PMID: 25598640
- [136] Habibyar, A.F.; Sharma, N.; Khurana, N. PASS assisted prediction and pharmacological evaluation of hesperidin against scopolamine induced amnesia in mice. *Eur. J. Pharmacol.*, **2016**, 789, 385-394. http://dx.doi.org/10.1016/j.ejphar.2016.07.013 PMID: 27397428
- [137] Jain, S.; Shrivastava, S.; Nayak, S.; Sumbhate, S. Recent trends in *Curcuma longa Linn. Pharmacogn. Rev.*, 2007, 1(1).
- [138] Joe, B.; Vijaykumar, M.; Lokesh, B.R. Biological properties of curcumin-cellular and molecular mechanisms of action. *Crit. Rev. Food Sci. Nutr.*, **2004**, *44*(2), 97-111. http://dx.doi.org/10.1080/10408690490424702 PMID: 15116757
- [139] Akram, M.; Shahab-Uddin, A.A.; Usmanghani, K.; Hannan, A.; Mohiuddin, E.; Asif, M. *Curcuma longa* and curcumin: a review article. *Rom. J. Biol. Plant Biol.*, **2010**, *55*(2), 65-70.
- [140] Kunchandy, E.; Rao, M. Oxygen radical scavenging activity of curcumin. *Int. J. Pharm.*, **1990**, *58*(3), 237-240. http://dx.doi.org/10.1016/0378-5173(90)90201-E
- [141] Chainani-Wu, N. Safety and anti-inflammatory activity of curcumin: a component of tumeric (*Curcuma longa*). J. Altern. Complement. Med., 2003, 9(1), 161-168. http://dx.doi.org/10.1089/107555303321223035 PMID: 12676044

- [142] Chattopadhyay, I.; Biswas, K.; Bandyopadhyay, U.; Banerjee, R.K. Turmeric and curcumin: Biological actions and medicinal applications. *Curr. Sci. Bangalore*, 2004, 87, 44-53.
- [143] Simon, A.; Allais, D.P.; Duroux, J.L.; Basly, J.P.; Durand-Fontanier, S.; Delage, C. Inhibitory effect of curcuminoids on MCF-7 cell proliferation and structure-activity relationships. *Cancer Lett.*, **1998**, *129*(1), 111-116. http://dx.doi.org/10.1016/S0304-3835(98)00092-5 PMID: 9714342
- [144] Sreejayan, ; Rao, M.N. Nitric oxide scavenging by curcuminoids. J. Pharm. Pharmacol., 1997, 49(1), 105-107. http://dx.doi.org/10.1111/j.2042-7158.1997.tb06761.x PMID: 9120760
- [145] Daniel, S.; Limson, J.L.; Dairam, A.; Watkins, G.M.; Daya, S. Through metal binding, curcumin protects against lead- and cadmium-induced lipid peroxidation in rat brain homogenates and against lead-induced tissue damage in rat brain. *J. Inorg. Biochem.*, 2004, 98(2), 266-275.
- http://dx.doi.org/10.1016/j.jinorgbio.2003.10.014 PMID: 14729307 [146] Camacho-Barquero, L.; Villegas, I.; Sánchez-Calvo, J.M.; Talero,
- [146] Canacho-Barquero, E., Vinegas, I., Sanchez-Carvo, J.M., Farcho, E.; Sánchez-Fidalgo, S.; Motilva, V.; Alarcón de la Lastra, C. Curcumin, a *Curcuma longa* constituent, acts on MAPK p38 pathway modulating COX-2 and iNOS expression in chronic experimental colitis. *Int. Immunopharmacol.*, 2007, 7(3), 333-342. http://dx.doi.org/10.1016/j.intimp.2006.11.006 PMID: 17276891
- [147] Vajragupta, O.; Boonchoong, P.; Watanabe, H.; Tohda, M.; Kummasud, N.; Sumanont, Y. Manganese complexes of curcumin and its derivatives: evaluation for the radical scavenging ability and neuroprotective activity. *Free Radic. Biol. Med.*, **2003**, *35*(12), 1632-1644.

http://dx.doi.org/10.1016/j.freeradbiomed.2003.09.011 PMID: 14680686

- [148] Thiyagarajan, M.; Sharma, S.S. Neuroprotective effect of curcumin in middle cerebral artery occlusion induced focal cerebral ischemia in rats. *Life Sci.*, 2004, 74(8), 969-985. http://dx.doi.org/10.1016/j.lfs.2003.06.042 PMID: 14672754
- [149] Anand, A.; Khurana, P.; Chawla, J.; Sharma, N.; Khurana, N. Emerging treatments for the behavioral and psychological symptoms of dementia. *CNS Spectr.*, **2018**, *23*(6), 361-369. http://dx.doi.org/10.1017/S1092852917000530 PMID: 28911339
- [150] Frautschy, S.A.; Hu, W.; Kim, P.; Miller, S.A.; Chu, T.; Harris-White, M.E.; Cole, G.M. Phenolic anti-inflammatory antioxidant reversal of Abeta-induced cognitive deficits and neuropathology. *Neurobiol. Aging*, 2001, 22(6), 993-1005. http://dx.doi.org/10.1016/S0197-4580(01)00300-1 PMID: 11755008
- [151] Wattmo, C.; Jedenius, E.; Blennow, K.; Wallin, Å.K. Dose and plasma concentration of galantamine in Alzheimer's disease - clinical application. *Alzheimers Res. Ther.*, **2013**, *5*(1), 2. http://dx.doi.org/10.1186/alzrt156 PMID: 23286718
- [152] Prvulovic, D.; Hampel, H.; Pantel, J. Galantamine for Alzheimer's disease. Expert Opin. Drug Metab. Toxicol., 2010, 6(3), 345-354. http://dx.doi.org/10.1517/17425251003592137 PMID: 20113148
- [153] Nakanishi, K. Terpene trilactones from Gingko biloba: from ancient times to the 21st century. *Bioorg. Med. Chem.*, 2005, 13(17), 4987-5000.

http://dx.doi.org/10.1016/j.bmc.2005.06.014 PMID: 15990319

- [154] McKenna, D.J.; Jones, K.; Hughes, K. Efficacy, safety, and use of ginkgo biloba in clinical and preclinical applications. *Altern. Ther. Health Med.*, **2001**, 7(5), 70-86, 88-90. PMID: 11565403
- [155] Smith, J.V.; Luo, Y. Studies on molecular mechanisms of Ginkgo biloba extract. *Appl. Microbiol. Biotechnol.*, 2004, 64(4), 465-472. http://dx.doi.org/10.1007/s00253-003-1527-9 PMID: 14740187
- [156] DeFeudis, F.V.; Drieu, K. Ginkgo biloba extract (EGb 761) and CNS functions: basic studies and clinical applications. *Curr. Drug Targets*, 2000, 1(1), 25-58. http://dx.doi.org/10.2174/1389450003349380 PMID: 11475535
- [157] Ramassamy, C.; Longpré, F.; Christen, Y. Ginkgo biloba extract (EGb 761) in Alzheimer's disease: is there any evidence? *Curr. Alzheimer Res.*, **2007**, 4(3), 253-262. http://dx.doi.org/10.2174/156720507781077304 PMID: 17627482
- [158] Pietri, S.; Maurelli, E.; Drieu, K.; Culcasi, M. Cardioprotective and anti-oxidant effects of the terpenoid constituents of Ginkgo biloba extract (EGb 761). J. Mol. Cell. Cardiol., **1997**, 29(2), 733-742. http://dx.doi.org/10.1006/jmcc.1996.0316 PMID: 9140830

- [159] Mahadevan, S.; Park, Y. Multifaceted therapeutic benefits of Ginkgo biloba L.: chemistry, efficacy, safety, and uses. J. Food Sci., 2008, 73(1), R14-R19. http://dx.doi.org/10.1111/j.1750-3841.2007.00597.x PMID: 18211362
- [160] Amri, H.; Ogwuegbu, S.O.; Boujrad, N.; Drieu, K.; Papadopoulos, V. In vivo regulation of peripheral-type benzodiazepine receptor and glucocorticoid synthesis by Ginkgo biloba extract EGb 761 and isolated ginkgolides. *Endocrinology*, **1996**, 137(12), 5707-5718. http://dx.doi.org/10.1210/endo.137.12.8940403 PMID: 8940403
- [161] Mahdy, H.M.; Tadros, M.G.; Mohamed, M.R.; Karim, A.M.; Khalifa, A.E. The effect of Ginkgo biloba extract on 3nitropropionic acid-induced neurotoxicity in rats. *Neurochem. Int.*, 2011, 59(6), 770-778.
- http://dx.doi.org/10.1016/j.neuint.2011.07.012 PMID: 21827809
 [162] Zhao, B. Natural antioxidants for neurodegenerative diseases. *Mol. Neurobiol.*, 2005, 31(1-3), 283-293.
 http://dx.doi.org/10.1385/MN:31:1-3:283 PMID: 15953828
- [163] Calabrese, E.J.; Calabrese, V.; Tsatsakis, A.; Giordano, J.J. Hormesis and *Ginkgo biloba* (GB): Numerous biological effects of GB are mediated via hormesis. Ageing Res. Rev., 2020, 101019. http://dx.doi.org/10.1016/j.arr.2020.101019 PMID: 31931153
- [164] Teltumbde, A.; Wahurwagh, A.; Lonare, M.; Nesari, T. Effect of Yashtimadhu (*Glycyrrhiza glabra*) on intelligence and memory function in male adolescents. *Sch. J. App. Med. Sci*, **2013**, *I*(2), 90-95
- [165] Yu, X-Q.; Xue, C.C.; Zhou, Z-W.; Li, C-G.; Du, Y-M.; Liang, J.; Zhou, S-F. *In vitro* and *in vivo* neuroprotective effect and mechanisms of glabridin, a major active isoflavan from *Glycyrrhiza glabra* (licorice). *Life Sci.*, **2008**, *82*(1-2), 68-78. http://dx.doi.org/10.1016/j.lfs.2007.10.019 PMID: 18048062
- [166] Balamurugan, G.; Muralidharan, P.; Babu, V. Cerebroprotective effect of *Glycyrrhiza glabra* Linn. root extract on hypoxic rats. *Bangladesh J. Pharmacol.*, 2009, 4(1), 60-64.
- [167] Ding, R.; Fu, J-G.; Xu, G-Q.; Sun, B-F.; Lin, G-Q. Divergent total synthesis of the Lycopodium alkaloids huperzine A, huperzine B, and huperzine U. J. Org. Chem., 2014, 79(1), 240-250. http://dx.doi.org/10.1021/jo402419h PMID: 24299147
- [168] Chu, D.; Tian, J.; Liu, W.; Li, Z.; Li, Y. Poly(lactic-co-glycolic acid) microspheres for the controlled release of huperzine A: *in vitro* and *in vivo* studies and the application in the treatment of the impaired memory of mice. *Chem. Pharm. Bull. (Tokyo)*, 2007, 55(4), 625-628.
- http://dx.doi.org/10.1248/cpb.55.625 PMID: 17409558
 [169] Zangara, A. The psychopharmacology of huperzine A: an alkaloid with cognitive enhancing and neuroprotective properties of interest in the treatment of Alzheimer's disease. *Pharmacol. Biochem. Behav.*, 2003, 75(3), 675-686.
 http://dx.doi.org/10.1016/S0091-3057(03)00111-4 PMID: 12895686
- Zhang, H.Y.; Zheng, C.Y.; Yan, H.; Wang, Z.F.; Tang, L.L.; Gao, X.; Tang, X.C. Potential therapeutic targets of huperzine A for Alzheimer's disease and vascular dementia. *Chem. Biol. Interact.*, 2008, 175(1-3), 396-402. http://dx.doi.org/10.1016/j.cbi.2008.04.049 PMID: 18565502

[171] Boskou, D. *Olive oil: chemistry and technology*; Elsevier, **2015**.

- [172] Boskou, D.; Blekas, G.; Tsimidou, M. Olive oil composition. *Olive Oil*; Elsevier, **2006**, pp. 41-72.
- [173] Covas, M-I.; Konstantinidou, V.; Fitó, M. Olive oil and cardiovascular health. J. Cardiovasc. Pharmacol., 2009, 54(6), 477-482. http://dx.doi.org/10.1097/FJC.0b013e3181c5e7fd PMID: 19858733
- [174] Abuznait, A.H.; Qosa, H.; Busnena, B.A.; El Sayed, K.A.; Kaddoumi, A. Olive-oil-derived oleocanthal enhances β-amyloid clearance as a potential neuroprotective mechanism against Alzheimer's disease: *in vitro* and *in vivo* studies. ACS Chem. Neurosci., 2013, 4(6), 973-982. http://dx.doi.org/10.1021/cn400024q PMID: 23414128
- [175] Batarseh, Y.S.; Kaddoumi, A. Oleocanthal-rich extra-virgin olive oil enhances donepezil effect by reducing amyloid-β load and related toxicity in a mouse model of Alzheimer's disease. J. Nutr. Biochem., 2018, 55, 113-123.
- http://dx.doi.org/10.1016/j.jnutbio.2017.12.006 PMID: 29413486
 Brunetti, G.; Di Rosa, G.; Scuto, M.; Leri, M.; Stefani, M.; Schmitz-Linneweber, C.; Calabrese, V.; Saul, N. Healthspan maintenance and prevention of parkinson's-like phenotypes with hy-

droxytyrosol and oleuropein aglycone in C. elegans. Int. J. Mol. Sci., 2020, 21(7), 2588.

- http://dx.doi.org/10.3390/ijms21072588 PMID: 32276415
- [177] Di Rosa, G.; Brunetti, G.; Scuto, M.; Trovato Salinaro, A.; Calabrese, E.J.; Crea, R.; Schmitz-Linneweber, C.; Calabrese, V.; Saul, N. Healthspan enhancement by olive polyphenols in *C. elegans* wild type and Parkinson's models. *Int. J. Mol. Sci.*, **2020**, *21*(11), 3893.

http://dx.doi.org/10.3390/ijms21113893 PMID: 32486023

- [178] Materljan, E.; Materljan, M.; Materljan, B.; Vlacić, H.; Barićev-Novaković, Z.; Sepcić, J. Multiple sclerosis and cancers in Croatia-a possible protective role of the "Mediterranean diet". *Coll. Antropol.*, 2009, 33(2), 539-545.
 PMID: 19662776
- [179] Leri, M.; Scuto, M.; Ontario, M.L.; Calabrese, V.; Calabrese, E.J.; Bucciantini, M.; Stefani, M. Healthy effects of plant polyphenols: molecular mechanisms. *Int. J. Mol. Sci.*, **2020**, *21*(4), 1250. http://dx.doi.org/10.3390/ijms21041250 PMID: 32070025
- [180] Visioli, F.; Bellomo, G.; Galli, C. Free radical-scavenging properties of olive oil polyphenols. *Biochem. Biophys. Res. Commun.*, 1998, 247(1), 60-64.

http://dx.doi.org/10.1006/bbrc.1998.8735 PMID: 9636654

[181] Tasset, I.; Pontes, A.J.; Hinojosa, A.J.; de la Torre, R.; Túnez, I. Olive oil reduces oxidative damage in a 3-nitropropionic acidinduced Huntington's disease-like rat model. *Nutr. Neurosci.*, 2011, 14(3), 106-111. http://dx.doi.org/10.1179/1476830511Y.0000000005 PMID:

21756531

[182] Guo, Y.; Mao, X.; Zhang, J.; Sun, P.; Wang, H.; Zhang, Y.; Ma, Y.; Xu, S.; Lv, R.; Liu, X. Oral delivery of lycopene-loaded microemulsion for brain-targeting: preparation, characterization, pharmacokinetic evaluation and tissue distribution. *Drug Deliv.*, 2019, 26(1), 1191-1205. http://dx.doi.org/10.1080/10717544.2019.1689312 PMID:

http://dx.doi.org/10.1080/10/1/544.2019.168931 31738085

- [183] Lunn, J.E.; Delorge, I.; Figueroa, C.M.; Van Dijck, P.; Stitt, M. Trehalose metabolism in plants. *Plant J.*, **2014**, *79*(4), 544-567. http://dx.doi.org/10.1111/tpj.12509 PMID: 24645920
- [184] Chen, Q.; Haddad, G.G. Role of trehalose phosphate synthase and trehalose during hypoxia: from flies to mammals. J. Exp. Biol., 2004, 207(Pt 18), 3125-3129. http://dx.doi.org/10.1242/jeb.01133 PMID: 15299033
- [185] Kandror, O.; Bretschneider, N.; Kreydin, E.; Cavalieri, D.; Goldberg, A.L. Yeast adapt to near-freezing temperatures by STRE/Msn2,4-dependent induction of trehalose synthesis and certain molecular chaperones. *Mol. Cell*, 2004, *13*(6), 771-781. http://dx.doi.org/10.1016/S1097-2765(04)00148-0 PMID: 15053871
- [186] Arora, A.; Ha, C.; Park, C.B. Inhibition of insulin amyloid formation by small stress molecules. *FEBS Lett.*, 2004, 564(1-2), 121-125. http://dx.doi.org/10.1016/S0014-5793(04)00326-6 PMID: 15094052
- [187] Liu, R.; Barkhordarian, H.; Emadi, S.; Park, C.B.; Sierks, M.R. Trehalose differentially inhibits aggregation and neurotoxicity of beta-amyloid 40 and 42. *Neurobiol. Dis.*, 2005, 20(1), 74-81. http://dx.doi.org/10.1016/j.nbd.2005.02.003 PMID: 16137568
- [188] Sarkar, S.; Davies, J.E.; Huang, Z.; Tunnacliffe, A.; Rubinsztein, D.C. Trehalose, a novel mTOR-independent autophagy enhancer, accelerates the clearance of mutant huntingtin and α-synuclein. J. Biol. Chem., 2007, 282(8), 5641-5652. http://dx.doi.org/10.1074/jbc.M609532200 PMID: 17182613
- [189] Tanaka, M.; Machida, Y.; Niu, S.; Ikeda, T.; Jana, N.R.; Doi, H.; Kurosawa, M.; Nekooki, M.; Nukina, N. Trehalose alleviates polyglutamine-mediated pathology in a mouse model of Huntington disease. *Nat. Med.*, 2004, 10(2), 148-154. http://dx.doi.org/10.1038/nm985 PMID: 14730359
- [190] Nah, S.Y.; Kim, D.H.; Rhim, H. Ginsenosides: are any of them candidates for drugs acting on the central nervous system? CNS Drug Rev., 2007, 13(4), 381-404. http://dx.doi.org/10.1111/j.1527-3458.2007.00023.x PMID: 18078425
- [191] Flanagan, E.; Lamport, D.; Brennan, L.; Burnet, P.; Calabrese, V.; Cunnane, S.C.; de Wilde, M.C.; Dye, L.; Farrimond, J.A.; Emerson Lombardo, N.; Hartmann, T.; Hartung, T.; Kalliomäki, M.; Kuhnle,

G.G.; La Fata, G.; Sala-Vila, A.; Samieri, C.; Smith, A.D.; Spencer, J.P.E.; Thuret, S.; Tuohy, K.; Turroni, S.; Vanden Berghe, W.; Verkuijl, M.; Verzijden, K.; Yannakoulia, M.; Geurts, L.; Vauzour, D. Nutrition and the ageing brain: Moving towards clinical applications. *Ageing Res. Rev.*, **2020**, *62*, 101079. http://dx.doi.org/10.1016/j.arr.2020.101079 PMID: 32461136

- [192] Rajabian, A.; Rameshrad, M.; Hosseinzadeh, H. Therapeutic potential of Panax ginseng and its constituents, ginsenosides and gintonin, in neurological and neurodegenerative disorders: a patent review. *Expert Opin. Ther. Pat.*, **2019**, *29*(1), 55-72. http://dx.doi.org/10.1080/13543776.2019.1556258 PMID: 30513224
- [193] Liu, C.X.; Xiao, P.G. Recent advances on ginseng research in China. J. Ethnopharmacol., 1992, 36(1), 27-38. http://dx.doi.org/10.1016/0378-8741(92)90057-X PMID: 1501490
- [194] Rausch, W-D.; Liu, S.; Gille, G.; Radad, K. Neuroprotective effects of ginsenosides. *Acta Neurobiol. Exp. (Warsz.)*, 2006, 66(4), 369-375.
 PMID: 17265697
- [195] Radad, K.; Gille, G.; Liu, L.; Rausch, W-D. Use of ginseng in medicine with emphasis on neurodegenerative disorders. J. Pharmacol. Sci., 2006, 100(3), 175-186. http://dx.doi.org/10.1254/jphs.CRJ05010X PMID: 16518078
- [196] Keum, Y-S.; Park, K-K.; Lee, J-M.; Chun, K-S.; Park, J.H.; Lee, S.K.; Kwon, H.; Surh, Y-J. Antioxidant and anti-tumor promoting activities of the methanol extract of heat-processed ginseng. *Cancer Lett.*, **2000**, *150*(1), 41-48. http://dx.doi.org/10.1016/S0304-3835(99)00369-9 PMID: 10755385
- [197] Li, X.; Li, S. Effects of total saponins of Sanchi (Panax pseudoginseng var. notoginseng) on TNF, NO and its mechanisms. *Chin. Tradit. Herbal Drugs*, **1999**, *30*(7), 514-517.
- Kim, Y.; Kim, S.; Markelonis, G.; Oh, T. Ginsenosides Rb1 and Rg3 protect cultured rat cortical cells from glutamate-induced neurodegeneration. J. Neurosci. Res., 53: 426-432. J. Neurosci. Res., 1998, 54(1), 123-123. http://dx.doi.org/10.1002/(SICI)1097-4547(19981001)54:1<123::AID-JNR13>3.0.CO;2-8 PMID: 9778146
- Kim, S.; Ahn, K.; Oh, T.H.; Nah, S-Y.; Rhim, H. Inhibitory effect of ginsenosides on NMDA receptor-mediated signals in rat hippocampal neurons. *Biochem. Biophys. Res. Commun.*, 2002, 296(2), 247-254. http://dx.doi.org/10.1016/S0006-291X(02)00870-7 PMID: 12163009
- [200] Lian, X.Y.; Zhang, Z.; Stringer, J.L. Protective effects of ginseng components in a rodent model of neurodegeneration. *Ann. Neurol.*, 2005, 57(5), 642-648. http://dx.doi.org/10.1002/ana.20450 PMID: 15852378
- [201] Kim, J-H.; Kim, S.; Yoon, I-S.; Lee, J-H.; Jang, B-J.; Jeong, S.M.; Lee, J-H.; Lee, B-H.; Han, J-S.; Oh, S.; Kim, H.C.; Park, T.K.; Rhim, H.; Nah, S.Y. Protective effects of ginseng saponins on 3nitropropionic acid-induced striatal degeneration in rats. *Neuropharmacology*, 2005, 48(5), 743-756. http://dx.doi.org/10.1016/j.neuropharm.2004.12.013 PMID: 15814108
- [202] Wu, J.; Jeong, H.K.; Bulin, S.E.; Kwon, S.W.; Park, J.H.; Bezprozvanny, I. Ginsenosides protect striatal neurons in a cellular model of Huntington's disease. *J. Neurosci. Res.*, **2009**, *87*(8), 1904-1912. http://dx.doi.org/10.1002/jnr.22017 PMID: 19185022
- [203] Sankar, D.; Sambandam, G.; Ramakrishna Rao, M.; Pugalendi, K.V. Modulation of blood pressure, lipid profiles and redox status in hypertensive patients taking different edible oils. *Clin. Chim. Acta*, 2005, 355(1-2), 97-104. http://dx.doi.org/10.1016/j.cccn.2004.12.009 PMID: 15820483
- [204] Baba, N.H.; Antoniades, K.; Habbal, Z. Effects of dietary canola, olive, and linolenic acid enriched olive oils on plasma lipids, lipid peroxidation and lipoprotein lipase activity in rats. *Nutr. Res.*, 1999, 19(4), 601-612.
 - http://dx.doi.org/10.1016/S0271-5317(99)00025-1
- [205] Kapadia, G.J.; Azuine, M.A.; Tokuda, H.; Takasaki, M.; Mukainaka, T.; Konoshima, T.; Nishino, H. Chemopreventive effect of resveratrol, sesamol, sesame oil and sunflower oil in the Epstein-Barr virus early antigen activation assay and the mouse skin twostage carcinogenesis. *Pharmacol. Res.*, **2002**, *45*(6), 499-505.

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http://dx.doi.org/10.1006/phrs.2002.0992 PMID: 12162952

[206] Hsu, D-Z.; Chen, K-T.; Li, Y-H.; Chuang, Y-C.; Liu, M-Y. Sesamol delays mortality and attenuates hepatic injury after cecal ligation and puncture in rats: role of oxidative stress. *Shock*, 2006, 25(5), 528-532. http://dx.doi.org/10.1097/01.shk.0000209552.95839.43 PMID: 16680019

[207] Kumar, P.; Kalonia, H.; Kumar, A. Sesamol attenuate 3nitropropionic acid-induced Huntington-like behavioral, biochemical, and cellular alterations in rats. J. Asian Nat. Prod. Res., 2009, 11(5), 439-450.

- http://dx.doi.org/10.1080/10286020902862194 PMID: 19504387
 [208] Hsu, D-Z.; Wan, C-H.; Hsu, H-F.; Lin, Y-M.; Liu, M-Y. The prophylactic protective effect of sesamol against ferric-nitrilotriacetate-induced acute renal injury in mice. *Food Chem. Toxicol.*, 2008, 46(8), 2736-2741.
 - http://dx.doi.org/10.1016/j.fct.2008.04.029 PMID: 18539378
- [209] Visioli, F.; Riso, P.; Grande, S.; Galli, C.; Porrini, M. Protective activity of tomato products on *in vivo* markers of lipid oxidation. *Eur. J. Nutr.*, 2003, 42(4), 201-206. http://dx.doi.org/10.1007/s00394-003-0415-5 PMID: 12923651
- [210] Hsiao, G.; Fong, T.H.; Tzu, N.H.; Lin, K.H.; Chou, D.S.; Sheu, J.R. A potent antioxidant, lycopene, affords neuroprotection against microglia activation and focal cerebral ischemia in rats. *in vivo*, 2004, 18(3), 351-356.
- [211] Rafi, M.M.; Yadav, P.N.; Reyes, M. Lycopene inhibits LPSinduced proinflammatory mediator inducible nitric oxide synthase in mouse macrophage cells. J. Food Sci., 2007, 72(1), S069-S074. http://dx.doi.org/10.1111/j.1750-3841.2006.00219.x PMID: 17995901
- [212] Vishwas, S.; Awasthi, A.; Corrie, L.; Kumar Singh, S.; Gulati, M. Multiple target-based combination therapy of galantamine, memantine and lycopene for the possible treatment of Alzheimer's disease. *Med. Hypotheses*, **2020**, *143*, 109879. http://dx.doi.org/10.1016/j.mehy.2020.109879 PMID: 32474382
- [213] Gunasekera, R.S.; Sewgobind, K.; Desai, S.; Dunn, L.; Black, H.S.; McKeehan, W.L.; Patil, B. Lycopene and lutein inhibit proliferation in rat prostate carcinoma cells. *Nutr. Cancer*, 2007, 58(2), 171-177.
- http://dx.doi.org/10.1080/01635580701328339 PMID: 17640163
- [214] Kuhad, A.; Sethi, R.; Chopra, K. Lycopene attenuates diabetesassociated cognitive decline in rats. *Life Sci.*, 2008, 83(3-4), 128-134.
 - http://dx.doi.org/10.1016/j.lfs.2008.05.013 PMID: 18585396
- [215] Akbaraly, N.T.; Faure, H.; Gourlet, V.; Favier, A.; Berr, C. Plasma carotenoid levels and cognitive performance in an elderly population: results of the EVA Study. J. Gerontol. A Biol. Sci. Med. Sci., 2007, 62(3), 308-316.

http://dx.doi.org/10.1093/gerona/62.3.308 PMID: 17389729

- [216] Atessahin, A.; Yilmaz, S.; Karahan, I.; Ceribasi, A.O.; Karaoglu, A. Effects of lycopene against cisplatin-induced nephrotoxicity and oxidative stress in rats. *Toxicology*, **2005**, *212*(2-3), 116-123. http://dx.doi.org/10.1016/j.tox.2005.04.016 PMID: 15946783
- [217] Kumar, P.; Kalonia, H.; Kumar, A. Lycopene modulates nitric oxide pathways against 3-nitropropionic acid-induced neurotoxicity. *Life Sci.*, **2009**, *85*(19-20), 711-718. http://dx.doi.org/10.1016/j.lfs.2009.10.001 PMID: 19822156
- [218] Rawal, A.K.; Muddeshwar, M.G.; Biswas, S.K. Rubia cordifolia, Fagonia cretica linn and Tinospora cordifolia exert neuroprotection by modulating the antioxidant system in rat hippocampal slices subjected to oxygen glucose deprivation. BMC Complement. Altern. Med., 2004, 4(1), 11. http://dx.doi.org/10.1186/1472-6882-4-11 PMID: 15310392
- [219] Kosaraju, J.; Chinni, S.; Roy, P.D.; Kannan, E.; Antony, A.S.; Kumar, M.N. Neuroprotective effect of Tinospora cordifolia ethanol extract on 6-hydroxy dopamine induced Parkinsonism. *Indian J. Pharmacol.*, 2014, 46(2), 176-180.

http://dx.doi.org/10.4103/0253-7613.129312 PMID: 24741189

- [220] Bairy, K.L.; Rao, Y.; Kumar Das, S.; Kumar, K.B. Efficacy of *Tinospora cordifolia* on learning and memory in healthy volunteers: A double-blind, randomized, placebo controlled study. *Iranian J. Pharmacol. Therap.*,2004, 3(2), 57-50.
- [221] Kim, D.H.; Shin, E.K.; Kim, Y.H.; Lee, B.W.; Jun, J.G.; Park, J.H.; Kim, J.K. Suppression of inflammatory responses by celastrol, a

quinone methide triterpenoid isolated from *Celastrus regelii. Eur. J. Clin. Invest.*, **2009**, *39*(9), 819-827. http://dx.doi.org/10.1111/j.1365-2362.2009.02186.x PMID: 19549173

- [222] Cleren, C.; Calingasan, N.Y.; Chen, J.; Beal, M.F. Celastrol protects against MPTP- and 3-nitropropionic acid-induced neurotoxicity. J. Neurochem., 2005, 94(4), 995-1004. http://dx.doi.org/10.1111/j.1471-4159.2005.03253.x PMID: 16092942
- [223] Allison, A.C.; Cacabelos, R.; Lombardi, V.R.; Álvarez, X.A.; Vigo, C. Celastrol, a potent antioxidant and anti-inflammatory drug, as a possible treatment for Alzheimer's disease. *Prog. Neuropsychopharmacol. Biol. Psychiatry*, **2001**, *25*(7), 1341-1357. http://dx.doi.org/10.1016/S0278-5846(01)00192-0 PMID: 11513350
- [224] Lee, J-H.; Choi, K.J.; Seo, W.D.; Jang, S.Y.; Kim, M.; Lee, B.W.; Kim, J.Y.; Kang, S.; Park, K.H.; Lee, Y-S.; Bae, S. Enhancement of radiation sensitivity in lung cancer cells by celastrol is mediated by inhibition of Hsp90. *Int. J. Mol. Med.*, 2011, 27(3), 441-446. PMID: 21249311
- [225] Avilla, J.; Teixidò, A.; Velázquez, C.; Alvarenga, N.; Ferro, E.; Canela, R. Insecticidal activity of Maytenus species (Celastraceae) nortriterpene quinone methides against codling moth, *Cydia pomo-nella* (L.) (*Lepidoptera: tortricidae*). J. Agric. Food Chem., 2000, 48(1), 88-92.
- http://dx.doi.org/10.1021/jf990008w PMID: 10637057
 [226] Zhang, Y-Q.; Sarge, K.D. Celastrol inhibits polyglutamine aggregation and toxicity though induction of the heat shock response. J. Mol. Med. (Berl.), 2007, 85(12), 1421-1428.
- http://dx.doi.org/10.1007/s00109-007-0251-9 PMID: 17943263
 [227] Winters, M. Ancient medicine, modern use: Withania somnifera and its potential role in integrative oncology. Altern. Med. Rev., 2006, 11(4), 269-277. PMID: 17176166
- [228] Yadava, S.A.; Hakkim, L.; Sathishkumar, F.; Sathishkumar, R. Antioxidant activity of *Withania somnifera* (L.) Dunal by different solvent extraction methods. *J. Pharm. Res.*, 2011, 4(5), 1428-1430.
- Bhattacharya, S.K.; Satyan, K.S.; Ghosal, S. Antioxidant activity of glycowithanolides from *Withania somnifera*. *Indian J. Exp. Biol.*, 1997, 35(3), 236-239.
 PMID: 9332168
- [230] Kulkarni, S.; Verma, A. Aswagandha and Brahmi: Nootropic and de-addiction profile of psychotropic indigenous plants. *Drugs Today (Barc)*, **1993**, *29*, 257-257.
- [231] Davis, L.; Kuttan, G. Effect of Withania somnifera on cell mediated immune responses in mice. J. Exper. Clin. Cancer Res., (East Lansing Mich.), 2002, 21(4), 585-590.
- [232] Davis, L.; Kuttan, G. Immunomodulatory activity of Withania somnifera. J. Ethnopharmacol., 2000, 71(1-2), 193-200. http://dx.doi.org/10.1016/S0378-8741(99)00206-8 PMID: 10904163
- [233] Bhattacharya, S.K.; Goel, R.K.; Kaur, R.; Ghosal, S. Anti stress activity of sitoindosides VII and VIII, new acylsterylglucosides from *Withania somnifera*. *Phytother. Res.*, **1987**, *1*(1), 32-37. http://dx.doi.org/10.1002/ptr.2650010108
- [234] Kulkarni, S.; Akula, K.K.; Dhir, A. Effect of Withania somnifera Dunal root extract against pentylenetetrazol seizure threshold in mice: possible involvement of GABAergic system. *Indian J. Exp. Biol.*, 2008, 46(6), 465-9.
- [235] Bhatnagar, M.; Sisodia, S.S.; Bhatnagar, R. Antiulcer and antioxidant activity of Asparagus racemosus Willd and Withania somnifera Dunal in rats. *Ann. N. Y. Acad. Sci.*, 2005, 1056(1), 261-278. http://dx.doi.org/10.1196/annals.1352.027 PMID: 16387694
- [236] Gupta, S.K.; Dua, A.; Vohra, B.P. Withania somnifera (Ashwagandha) attenuates antioxidant defense in aged spinal cord and inhibits copper induced lipid peroxidation and protein oxidative modifications. *Drug Metabol. Drug Interact.*, **2003**, *19*(3), 211-222.

http://dx.doi.org/10.1515/DMDI.2003.19.3.211 PMID: 14682611

- [237] Bhattacharya, S.K.; Bhattacharya, D.; Sairam, K.; Ghosal, S. Effect of Withania somnifera glycowithanolides on a rat model of tardive dyskinesia. *Phytomedicine*, 2002, 9(2), 167-170. http://dx.doi.org/10.1078/0944-7113-00089 PMID: 11995951
- [238] Singh, B.; Saxena, A.K.; Chandan, B.K.; Gupta, D.K.; Bhutani, K.K.; Anand, K.K. Adaptogenic activity of a novel, withanolide-

free aqueous fraction from the roots of *Withania somnifera* Dun. *Phytother. Res.*, **2001**, *15*(4), 311-318. http://dx.doi.org/10.1002/ptr.858 PMID: 11406854

- [239] Patel, V.; Jivani, N.; Patel, S. Medicinal plants with potential nootropic activity: a review. *Res. J. Pharm. Biol. Chem. Sci.*, 2014, 5(1), 1-11.
- [240] Kumar, P.; Kumar, A. Possible neuroprotective effect of Withania somnifera root extract against 3-nitropropionic acid-induced behavioral, biochemical, and mitochondrial dysfunction in an animal model of Huntington's disease. J. Med. Food, 2009, 12(3), 591-600.

http://dx.doi.org/10.1089/jmf.2008.0028 PMID: 19627208

- [241] Kumar, N.; Abichandani, L.; Thawani, V.; Gharpure, K.; Naidu, M.; Venkatm R,G. Efficacy of standardized extract of Bacopa monnieri (Bacognize®) on cognitive functions of medical students: a six-week, randomized placebo-controlled trial. *Evid-Based Complement Alternat. Med.*, 2016, 2016, 4103423.
- [242] Singh, S.; Jamwal, S.; Kumar, P. Piperine enhances the protective effect of curcumin against 3-NP induced neurotoxicity: possible neurotransmitters modulation mechanism. *Neurochem. Res.*, 2015, 40(8), 1758-1766.

http://dx.doi.org/10.1007/s11064-015-1658-2 PMID: 26160706

[243] Busquet, P.; Capurro, V.; Cavalli, A.; Piomelli, D.; Reggiani, A.; Bertorelli, R. Synergistic effects of galantamine and memantine in attenuating scopolamine-induced amnesia in mice. *J. Pharmacol. Sci.*, 2012, 120(4), 305-9.

http://dx.doi.org/10.1254/jphs.12166SC

- [244] Damle, M. Glycyrrhiza glabra (Liquorice)-a potent medicinal herb. International Journal of Herbal Medicine, 2014, 2(2), 132-136.
- [245] Prakash, A.; Kumar, A. Mitoprotective effect of *Centella asiatica* against aluminum-induced neurotoxicity in rats: possible relevance to its anti-oxidant and anti-apoptosis mechanism. *Neurol. Sci.*, 2013, 34(8), 1403-1409.

http://dx.doi.org/10.1007/s10072-012-1252-1 PMID: 23224641

- [246] Fedalto, M.L.; Ludka, F.K.; Tasca, C.I.; Molz, S. Neuroprotection of Persea major extract against oxygen and glucose deprivation in hippocampal slices involves increased glutamate uptake and modulation of A1 and A2A adenosine receptors. *Rev. Bras. Farmacogn.*, 2013, 23(5), 789-795.
- http://dx.doi.org/10.1590/S0102-695X2013000500011
 [247] Ren, B.; Yuan, T.; Diao, Z.; Zhang, C.; Liu, Z.; Liu, X. Protective effects of sesamol on systemic oxidative stress-induced cognitive impairments *via* regulation of Nrf2/Keap1 pathway. *Food Funct.*, 2018, 9(11), 5912-5924.

http://dx.doi.org/10.1039/C8FO01436A PMID: 30375618

- [248] Padma, V.V.; Baskaran, R.; Divya, S.; Priya, L.B.; Saranya, S. Modulatory effect of *Tinospora cordifolia* extract on Cd-induced oxidative stress in Wistar rats. *Integr. Med. Res.*, 2016, 5(1), 48-55. http://dx.doi.org/10.1016/j.imr.2015.12.005 PMID: 28462097
- [249] Zhou, J.; Xi, C.; Wang, W.; Fu, X.; Jinqiang, L.; Qiu, Y.; Jin, J.; Xu, J.; Huang, Z. Triptolide-induced oxidative stress involved with Nrf2 contribute to cardiomyocyte apoptosis through mitochondrial dependent pathways. *Toxicol. Lett.*, **2014**, *230*(3), 454-466. http://dx.doi.org/10.1016/j.toxlet.2014.08.017 PMID: 25169008
- [250] Suen, J.; Thomas, J.; Kranz, A.; Vun, S.; Miller, M. Effect of flavonoids on oxidative stress and inflammation in adults at risk of cardiovascular disease: a systematic review: *Healthcare*, 2016, 4(3),69.
- [251] Mizunoe, Y.; Kobayashi, M.; Sudo, Y.; Watanabe, S.; Yasukawa, H.; Natori, D.; Hoshino, A.; Negishi, A.; Okita, N.; Komatsu, M.; Higami, Y. Trehalose protects against oxidative stress by regulating the Keap1-Nrf2 and autophagy pathways. *Redox Biol.*, 2018, 15, 115-124.

http://dx.doi.org/10.1016/j.redox.2017.09.007 PMID: 29241092

[252] Tvrdá, E.; Kováčik, A.; Tušimová, E.; Paál, D.; Mackovich, A.; Alimov, J.; Lukáč, N. Antioxidant efficiency of lycopene on oxidative stress - induced damage in bovine spermatozoa. J. Anim. Sci. Biotechnol., 2016, 7(1), 50.

http://dx.doi.org/10.1186/s40104-016-0113-9 PMID: 27602206

- [253] Chen, D.; Huang, C.; Chen, Z. A review for the pharmacological effect of lycopene in central nervous system disorders. *Biomed. Pharmacother.*, **2019**, *111*, 791-801. http://dx.doi.org/10.1016/j.biopha.2018.12.151 PMID: 30616078
- [254] Kallscheuer, N.; Menezes, R.; Foito, A.; da Silva, M.H.; Braga, A.; Dekker, W.; Sevillano, D.M.; Rosado-Ramos, R.; Jardim, C.;

Oliveira, J.; Ferreira, P.; Rocha, I.; Silva, A.R.; Sousa, M.; Allwood, J.W.; Bott, M.; Faria, N.; Stewart, D.; Ottens, M.; Naesby, M.; Nunes Dos Santos, C.; Marienhagen, J. Identification and microbial production of the raspberry phenol salidroside that is active against Huntington's disease. *Plant Physiol.*, **2019**, *179*(3), 969-985. http://dx.doi.org/10.1104/pp.18.01074 PMID: 30397021

- [255] ClinicalTrials.gov. Huntington's Disease. https://clinicaltrials.gov/ct2/results?cond=huntington+disease&ter m=&cntry=&state=&city=&dist=
- [256] Calabrese, E.J.; Baldwin, L.A.; Holland, C.D. Hormesis: a highly generalizable and reproducible phenomenon with important implications for risk assessment. *Risk Anal.*, **1999**, *19*(2), 261-281. http://dx.doi.org/10.1111/j.1539-6924.1999.tb00404.x PMID: 10765404
- [257] Valerio, A.; D'Antona, G.; Nisoli, E. Branched-chain amino acids, mitochondrial biogenesis, and healthspan: an evolutionary perspective. Aging (Albany NY), 2011, 3(5), 464-478. http://dx.doi.org/10.18632/aging.100322 PMID: 21566257
- [258] Calabrese, V.; Santoro, A.; Trovato Salinaro, A.; Modafferi, S.; Scuto, M.; Albouchi, F.; Monti, D.; Giordano, J.; Zappia, M.; Franceschi, C.; Calabrese, E.J. Hormetic approaches to the treatment of Parkinson's disease: Perspectives and possibilities. *J. Neurosci. Res.*, 2018, 96(10), 1641-1662. http://dx.doi.org/10.1002/jnr.24244 PMID: 30098077
- [259] Calabrese, V.; Cornelius, C.; Dinkova-Kostova, A.T.; Iavicoli, I.; Di Paola, R.; Koverech, A.; Cuzzocrea, S.; Rizzarelli, E.; Calabrese, E.J. Cellular stress responses, hormetic phytochemicals and vitagenes in aging and longevity. *Biochim. Biophys. Acta*, 2012, 1822(5), 753-783.
- http://dx.doi.org/10.1016/j.bbadis.2011.11.002 PMID: 22108204
 [260] Bellia, F.; Vecchio, G.; Cuzzocrea, S.; Calabrese, V.; Rizzarelli, E. Neuroprotective features of carnosine in oxidative driven diseases. *Mol. Aspects Med.*, 2011, 32(4-6), 258-266. http://dx.doi.org/10.1016/j.mam.2011.10.009 PMID: 22020110
- [261] Calabrese, V.; Cornelius, C.; Dinkova-Kostova, A.T.; Calabrese, E.J.; Mattson, M.P. Cellular stress responses, the hormesis paradigm, and vitagenes: novel targets for therapeutic intervention in neurodegenerative disorders. *Antioxid. Redox Signal.*, 2010, 13(11), 1763-1811.
- http://dx.doi.org/10.1089/ars.2009.3074 PMID: 20446769
 [262] Calabrese, V.; Santoro, A.; Monti, D.; Crupi, R.; Di Paola, R.; Latteri, S.; Cuzzocrea, S.; Zappia, M.; Giordano, J.; Calabrese, E.J.; Franceschi, C. Aging and Parkinson's Disease: Inflammaging, neuroinflammation and biological remodeling as key factors in pathogenesis. *Free Radic. Biol. Med.*, 2018, *115*, 80-91. http://dx.doi.org/10.1016/j.freeradbiomed.2017.10.379 PMID: 29080843
- [263] Peters, V.; Calabrese, V.; Forsberg, E.; Volk, N.; Fleming, T.; Baelde, H.; Weigand, T.; Thiel, C.; Trovato, A.; Scuto, M.; Modafferi, S.; Schmitt, C.P. Protective actions of anserine under diabetic conditions. *Int. J. Mol. Sci.*, **2018**, *19*(9), 2751. http://dx.doi.org/10.3390/ijms19092751 PMID: 30217069
- [264] Moghaddam, N.S.A.; Oskouie, M.N.; Butler, A.E.; Petit, P.X.; Barreto, G.E.; Sahebkar, A. Hormetic effects of curcumin: What is the evidence? *J. Cell. Physiol.*, 2019, 234(7), 10060-10071. http://dx.doi.org/10.1002/jcp.27880 PMID: 30515809
- [265] Siracusa, R.; Šcuto, M.; Fusco, R.; Trovato, A.; Ontario, M.L.; Crea, R.; Di Paola, R.; Cuzzocrea, S.; Calabrese, V. Antiinflammatory and Anti-oxidant Activity of Hidrox[®] in Rotenone-Induced Parkinson's Disease in Mice. *Antioxidants*, **2020**, *9*(9), 824.
- http://dx.doi.org/10.3390/antiox9090824 PMID: 32899274
 [266] Costa, L.G.; Garrick, J.M.; Roquè, P.J.; Pellacani, C. Mechanisms of neuroprotection by quercetin: counteracting oxidative stress and
- more. Oxid. Med. Cell. Longev., 2016. 2016, 2986796. http://dx.doi.org/10.1155/2016/2986796
- [267] Peth-Nui, T.; Wattanathorn, J.; Muchimapura, S.; Tong-Un, T.; Piyavhatkul, N.; Rangseekajee, P.; Ingkaninan, K.; Vittaya-areekul, S. Effects of 12-week *Bacopa monnieri* consumption on attention, cognitive processing, working memory, and functions of both cholinergic and monoaminergic systems in healthy elderly volunteers. *Evid. Based Complement. Alternat. Med.*, **2012**, 2012, 606424.
- [268] Ulusoy, H.G.; Sanlier, N. A minireview of quercetin: from its metabolism to possible mechanisms of its biological activities. *Crit. Rev. Food Sci. Nutr.*, 2019, 1-14.

http://dx.doi.org/10.1080/10408398.2019.1683810 PMID: 31680558

- [269] Lotito, S.B.; Frei, B. Relevance of apple polyphenols as antioxidants in human plasma: contrasting *in vitro* and *in vivo* effects. *Free Radic. Biol. Med.*, 2004, 36(2), 201-211. PMID: 14744632
- [270] Dutson, A.J.; Todd, R.H.; Magleby, S.P.; Sorensen, C.D. A review of literature on teaching engineering design through project oriented capstone courses. J. Eng. Educ., 1997, 86(1), 17-28. http://dx.doi.org/10.1002/j.2168-9830.1997.tb00260.x
- [271] Miquel, S.; Champ, C.; Day, J.; Aarts, E.; Bahr, B.A.; Bakker, M.; Bánáti, D.; Calabrese, V.; Cederholm, T.; Cryan, J.; Dye, L.; Farrimond, J.A.; Korosi, A.; Layé, S.; Maudsley, S.; Milenkovic, D.; Mohajeri, M.H.; Sijben, J.; Solomon, A.; Spencer, J.P.E.; Thuret, S.; Vanden Berghe, W.; Vauzour, D.; Vellas, B.; Wesnes, K.; Willatts, P.; Wittenberg, R.; Geurts, L. Poor cognitive ageing: Vulnerabilities, mechanisms and the impact of nutritional interventions. *Ageing Res. Rev.*, **2018**, *42*, 40-55.
- http://dx.doi.org/10.1016/j.arr.2017.12.004 PMID: 29248758
- [272] Pilipenko, V.; Narbute, K.; Amara, I.; Trovato, A.; Scuto, M.; Pupure, J.; Jansone, B.; Poikans, J.; Bisenieks, E.; Klusa, V.; Calabrese, V. GABA-containing compound gammapyrone protects against brain impairments in Alzheimer's disease model male rats and prevents mitochondrial dysfunction in cell culture. *J. Neurosci. Res.*, **2019**, *97*(6), 708-726.

http://dx.doi.org/10.1002/jnr.24396 PMID: 30742328

- [273] Tili, E.; Michaille, J-J. Promiscuous effects of some phenolic natural products on inflammation at least in part arise from their ability to modulate the expression of global regulators, namely microR-NAs. *Molecules*, 2016, 21(9), 1263.
- http://dx.doi.org/10.3390/molecules21091263 PMID: 27657035
 [274] Scuto, M.; Di Mauro, P.; Ontario, M.L.; Amato, C.; Modafferi, S.; Ciavardelli, D.; Trovato, S.; A.; Maiolino, L.; Calabrese, V. Nutritional mushroom treatment in meniere's disease with *Coriolus versicolor*: A Rationale for therapeutic intervention in neuroinflammation and antineurodegeneration. *Int. J. Mol. Sci.*, 2019, *21*(1), 284. http://dx.doi.org/10.3390/ijms21010284 PMID: 31906226
- [275] Amara, I.; Scuto, M.; Zappalà, A.; Ontario, M.L.; Petralia, A.; Abid-Essefi, S.; Maiolino, L.; Signorile, A.; Trovato Salinaro, A.; Calabrese, V. *Hericium Erinaceus* Prevents DEHP-induced mitochondrial dysfunction and apoptosis in PC12 Cells. *Int. J. Mol. Sci.*, **2020**, *21*(6), 2138.

http://dx.doi.org/10.3390/ijms21062138 PMID: 32244920

- [276] Mishra, A.; Patni, P.; Hegde, S.; Aleya, L.; Tewari, D. Neuroplasticity and environment: A pharmacotherapeutic approach towards preclinical and clinical understanding; Current Opinion in Environmental Science & Health, 2020.
- [277] Sirangelo, I.; Borriello, M.; Vilasi, S.; Iannuzzi, C. Hydroxytyrosol inhibits protein oligomerization and amyloid aggregation in human insulin. *Int. J. Mol. Sci.*, **2020**, *21*(13), 4636. http://dx.doi.org/10.3390/ijms21134636 PMID: 32629793
- [278] Kamble, B.; Talreja, S.; Gupta, A.; Patil, D.; Pathak, D.; Moothedath, I.; Duraiswamy, B. Development and biological evaluation of *Gymnema sylvestre* extract-loaded nonionic surfactant-based niosomes. *Nanomedicine (Lond.)*, **2013**, *8*(8), 1295-1305.

http://dx.doi.org/10.2217/nnm.12.162 PMID: 23259778

- [279] Lockman, P.R.; Mumper, R.J.; Khan, M.A.; Allen, D.D. Nanoparticle technology for drug delivery across the blood-brain barrier. *Drug Dev. Ind. Pharm.*, 2002, 28(1), 1-13. http://dx.doi.org/10.1081/DDC-120001481 PMID: 11858519
- [280] Garbayo, E.; Ansorena, E.; Blanco-Prieto, M.J. Brain drug delivery systems for neurodegenerative disorders. *Curr. Pharm. Biotechnol.*, 2012, 13(12), 2388-2402.

http://dx.doi.org/10.2174/138920112803341761 PMID: 23016644

[281] Ou, H.; Cheng, T.; Zhang, Y.; Liu, J.; Ding, Y.; Zhen, J.; Shen, W.; Xu, Y.; Yang, W.; Niu, P.; Liu, J.; An, Y.; Liu, Y.; Shi, L. Surfaceadaptive zwitterionic nanoparticles for prolonged blood circulation time and enhanced cellular uptake in tumor cells. *Acta Biomater.*, 2018, 65, 339-348.

http://dx.doi.org/10.1016/j.actbio.2017.10.034 PMID: 29079515

[282] Zhang, Y.; Walker, J.B.; Minic, Z.; Liu, F.; Goshgarian, H.; Mao, G. Transporter protein and drug-conjugated gold nanoparticles capable of bypassing the blood-brain barrier. *Sci. Rep.*, **2016**, *6*, 25794. http://dx.doi.org/10.1038/srep25794 PMID: 27180729

- [283] Wang, S.; Su, R.; Nie, S.; Sun, M.; Zhang, J.; Wu, D.; Moustaid-Moussa, N. Application of nanotechnology in improving bioavailability and bioactivity of diet-derived phytochemicals. *J. Nutr. Bio-chem.*, 2014, 25(4), 363-376.
 - http://dx.doi.org/10.1016/j.jnutbio.2013.10.002 PMID: 24406273
- [284] Betzer, O.; Shilo, M.; Opochinsky, R.; Barnoy, E.; Motiei, M.; Okun, E.; Yadid, G.; Popovtzer, R. The effect of nanoparticle size on the ability to cross the blood-brain barrier: an *in vivo* study. *Nanomedicine (Lond.)*, **2017**, *12*(13), 1533-1546. http://dx.doi.org/10.2217/nnm-2017-0022 PMID: 28621578
- [285] Wiley, D.T.; Webster, P.; Gale, A.; Davis, M.E. Transcytosis and brain uptake of transferrin-containing nanoparticles by tuning avidity to transferrin receptor. *Proc. Natl. Acad. Sci. USA*, 2013, *110*(21), 8662-8667.
- http://dx.doi.org/10.1073/pnas.1307152110 PMID: 23650374
 [286] Kumar, R.; Kumar, R.; Khurana, N.; Singh, S.K.; Khurana, S.; Verma, S.; Sharma, N.; Kapoor, B.; Vyas, M.; Khursheed, R.; Awasthi, A.; Kaur, J.; Corrie, L. Enhanced oral bioavailability and neuroprotective effect of fisetin through its SNEDDS against rote-none-induced Parkinson's disease rat model. *Food Chem. Toxicol.*, 2020, 144, 111590.

http://dx.doi.org/10.1016/j.fct.2020.111590 PMID: 32710995

- [287] Singh, A.; Ahmad, I.; Akhter, S.; Jain, G.K.; Iqbal, Z.; Talegaonkar, S.; Ahmad, F.J. Nanocarrier based formulation of Thymoquinone improves oral delivery: stability assessment, *in vitro* and *in vivo* studies. *Colloids Surf. B Biointerfaces*, 2013, 102, 822-832.
- http://dx.doi.org/10.1016/j.colsurfb.2012.08.038 PMID: 23104039
 [288] Surekha, R.; Sumathi, T. An efficient encapsulation of thymoquinone using solid lipid nanoparticle for brain targeted drug delivery: physicochemical characterization, pharmacokinetics and biodistribution studies. *Intl. J. Pharm. Clin. Res.*, **2016**, 8(12), 1616-1624.
- [289] Kumar, B.; Singh, S.K.; Prakash, T.; Bhatia, A.; Gulati, M.; Garg, V.; Pandey, N.K.; Singh, S.; Melkani, I. Pharmacokinetic and pharmacodynamic evaluation of Solid self-nanoemulsifying delivery system (SSNEDDS) loaded with curcumin and duloxetine in attenuation of neuropathic pain in rats. *Neurol. Sci.*, **2020**, 1-13. PMID: 32885394
- [290] Ravichandran, R. Pharmacokinetic study of nanoparticulate curcumin: oral formulation for enhanced bioavailability. 2013. http://dx.doi.org/10.4236/jbnb.2013.43037
- [291] Kakkar, V.; Singh, S.; Singla, D.; Kaur, I.P. Exploring solid lipid nanoparticles to enhance the oral bioavailability of curcumin. *Mol. Nutr. Food Res.*, 2011, 55(3), 495-503. http://dx.doi.org/10.1002/mnfr.201000310 PMID: 20938993
- [292] Maiti, K.; Mukherjee, K.; Gantait, A.; Saha, B.P.; Mukherjee, P.K. Curcumin-phospholipid complex: Preparation, therapeutic evaluation and pharmacokinetic study in rats. *Int. J. Pharm.*, 2007, 330(1-2), 155-163.
- http://dx.doi.org/10.1016/j.ijpharm.2006.09.025 PMID: 17112692
 [293] Penalva, R.; González-Navarro, C.J.; Gamazo, C.; Esparza, I.; Irache, J.M. Zein nanoparticles for oral delivery of quercetin: Pharmacokinetic studies and preventive anti-inflammatory effects in a mouse model of endotoxemia. *Nanomedicine (Lond.)*, 2017, *13*(1), 103-110. http://dx.doi.org/10.1016/j.nano.2016.08.033 PMID: 27615118
- [294] Li, H.; Zhao, X.; Ma, Y.; Zhai, G.; Li, L.; Lou, H. Enhancement of gastrointestinal absorption of quercetin by solid lipid nanoparticles.
- J. Control. Release, 2009, 133(3), 238-244. http://dx.doi.org/10.1016/j.jconrel.2008.10.002 PMID: 18951932
- [295] Smith, A.; Giunta, B.; Bickford, P.C.; Fountain, M.; Tan, J.; Shytle, R.D. Nanolipidic particles improve the bioavailability and αsecretase inducing ability of epigallocatechin-3-gallate (EGCG) for the treatment of Alzheimer's disease. *Int. J. Pharm.*, **2010**, *389*(1-2), 207-212.

http://dx.doi.org/10.1016/j.ijpharm.2010.01.012 PMID: 20083179

- [296] Onyeabor, F.; Paik, A.; Kovvasu, S.; Ding, B.; Lin, J.; Wahid, M.A.; Prabhu, S.; Betageri, G.; Wang, J. Optimization of preparation and preclinical pharmacokinetics of celastrol-encapsulated silk fibroin nanoparticles in the rat. *Molecules*, **2019**, *24*(18), 3271. http://dx.doi.org/10.3390/molecules24183271 PMID: 31500392
- [297] Freag, M.S.; Saleh, W.M.; Abdallah, O.Y. Self-assembled phospholipid-based phytosomal nanocarriers as promising platforms for

improving oral bioavailability of the anticancer celastrol. *Int. J. Pharm.*, **2018**, *535*(1-2), 18-26. http://dx.doi.org/10.1016/j.ijpharm.2017.10.053 PMID: 29102699

- [298] Tang, J.; Xu, N.; Ji, H.; Liu, H.; Wang, Z.; Wu, L. Eudragit nanoparticles containing genistein: formulation, development, and bioavailability assessment. *Int. J. Nanomed.*, **2011**, *6*, 2429-2435. PMID: 22072878
- Misra, S.; Chopra, K.; Sinha, V.R.; Medhi, B. Galantamine-loaded solid-lipid nanoparticles for enhanced brain delivery: preparation, characterization, *in vitro* and *in vivo* evaluations. *Drug Deliv.*, 2016, 23(4), 1434-1443. http://dx.doi.org/10.3109/10717544.2015.1089956 PMID: 26405825
- [300] Shangguan, M.; Lu, Y.; Qi, J.; Han, J.; Tian, Z.; Xie, Y.; Hu, F.; Yuan, H.; Wu, W. Binary lipids-based nanostructured lipid carriers for improved oral bioavailability of silymarin. *J. Biomater. Appl.*, 2014, 28(6), 887-896.
 - http://dx.doi.org/10.1177/0885328213485141 PMID: 24008629
- [301] Luo, Y.; Chen, D.; Ren, L.; Zhao, X.; Qin, J. Solid lipid nanoparticles for enhancing vinpocetine's oral bioavailability. *J. Control. Release*, 2006, 114(1), 53-59.
 - http://dx.doi.org/10.1016/j.jconrel.2006.05.010 PMID: 16828192
- [302] Wang, L.; Luo, Q.; Lin, T.; Li, R.; Zhu, T.; Zhou, K.; Ji, Z.; Song, J.; Jia, B.; Zhang, C.; Chen, W.; Zhu, G. PEGylated nanostructured lipid carriers (PEG-NLC) as a novel drug delivery system for biochanin A. *Drug Dev. Ind. Pharm.*, 2015, 41(7), 1204-1212. http://dx.doi.org/10.3109/03639045.2014.938082 PMID: 25010850
- [303] Zhang, T.; Wang, H.; Ye, Y.; Zhang, X.; Wu, B. Micellar emulsions composed of mPEG-PCL/MCT as novel nanocarriers for systemic delivery of genistein: a comparative study with micelles. *Int. J. Nanomedicine*, 2015, 10, 6175-6184. PMID: 26491290
- [304] Noble, G.T.; Stefanick, J.F.; Ashley, J.D.; Kiziltepe, T.; Bilgicer, B. Ligand-targeted liposome design: challenges and fundamental considerations. *Trends Biotechnol.*, **2014**, *32*(1), 32-45. http://dx.doi.org/10.1016/j.tibtech.2013.09.007 PMID: 24210498
- [305] Himanshu, A.; Sitasharan, P.; Singhai, A. Liposomes as drug carriers. *IJPLS*, 2011, 2(7), 945-951.
- [306] Re, F.; Cambianica, I.; Zona, C.; Sesana, S.; Gregori, M.; Rigolio, R.; La Ferla, B.; Nicotra, F.; Forloni, G.; Cagnotto, A.; Salmona, M.; Masserini, M.; Sancini, G. Functionalization of liposomes with ApoE-derived peptides at different density affects cellular uptake and drug transport across a blood-brain barrier model. *Nanomedicine (Lond.)*, 2011, 7(5), 551-559.

http://dx.doi.org/10.1016/j.nano.2011.05.004 PMID: 21658472

- [307] Battaglia, L.; Gallarate, M. Lipid nanoparticles: state of the art, new preparation methods and challenges in drug delivery. *Expert Opin. Drug Deliv.*, **2012**, 9(5), 497-508. http://dx.doi.org/10.1517/17425247.2012.673278 PMID: 22439808
- [308] Mishra, V.; Bansal, K.K.; Verma, A.; Yadav, N.; Thakur, S.; Sudhakar, K.; Rosenholm, J.M. Solid lipid nanoparticles: Emerging colloidal nano drug delivery systems. *Pharmaceutics*, 2018, 10(4), 191. http://dx.doi.org/10.3390/pharmaceutics10040191 PMID:

http://dx.doi.org/10.3390/pharmaceutics10040191 PMID: 30340327

- [309] Gastaldi, L.; Battaglia, L.; Peira, E.; Chirio, D.; Muntoni, E.; Solazzi, I.; Gallarate, M.; Dosio, F. Solid lipid nanoparticles as vehicles of drugs to the brain: current state of the art. *Eur. J. Pharm. Biopharm.*, 2014, 87(3), 433-444.
- http://dx.doi.org/10.1016/j.ejpb.2014.05.004 PMID: 24833004
 [310] Michaelis, K.; Hoffmann, M.M.; Dreis, S.; Herbert, E.; Alyautdin, R.N.; Michaelis, M.; Kreuter, J.; Langer, K. Covalent linkage of apolipoprotein e to albumin nanoparticles strongly enhances drug

transport into the brain. J. Pharmacol. Exp. Ther., 2006, 317(3), 1246-1253.

http://dx.doi.org/10.1124/jpet.105.097139 PMID: 16554356

[311] Odeh, F.; Ismail, S.I.; Abu-Dahab, R.; Mahmoud, I.S.; Al Bawab, A. Thymoquinone in liposomes: a study of loading efficiency and biological activity towards breast cancer. *Drug Deliv.*, 2012, 19(8), 371-377.

http://dx.doi.org/10.3109/10717544.2012.727500 PMID: 23043626

[312] Ramachandran, S.; Thangarajan, S. Thymoquinone loaded solid lipid nanoparticles counteracts 3-Nitropropionic acid induced motor impairments and neuroinflammation in rat model of Huntington's disease. *Metab. Brain Dis.*, 2018, 33(5), 1459-1470.

- [313] Jawahar, N.; Meyyanathan, S. Polymeric nanoparticles for drug delivery and targeting: A comprehensive review. *Int. J. Health Allied Sci.*, 2012, 1(4), 217. http://dx.ioux.org/10.0102/078-244X_107822
 - http://dx.doi.org/10.4103/2278-344X.107832
- [314] Debnath, K.; Pradhan, N.; Singh, B.K.; Jana, N.R.; Jana, N.R. Poly (trehalose) nanoparticles prevent amyloid aggregation and suppress polyglutamine aggregation in a Huntington's Disease model mouse. ACS Appl. Mater. Interfaces, 2017, 9(28), 24126-24139. http://dx.doi.org/10.1021/acsami.7b06510 PMID: 28632387
- [315] Passoni, A.; Favagrossa, M.; Colombo, L.; Bagnati, R.; Gobbi, M.; Diomede, L.; Birolini, G.; Di Paolo, E.; Valenza, M.; Cattaneo, E.; Salmona, M. Efficacy of Cholesterol Nose-to-Brain Delivery for Brain Targeting in Huntington's Disease. ACS Chem. Neurosci., 2020, 11(3), 367-372.
- http://dx.doi.org/10.1021/acschemneuro.9b00581 PMID: 31860272
 [316] Cong, W.; Bai, R.; Li, Y-F.; Wang, L.; Chen, C. Selenium Nanoparticles as an efficient nanomedicine for the therapy of

Huntington's Disease. ACS Appl. Mater. Interfaces, 2019, 11(38), 34725-34735.

http://dx.doi.org/10.1021/acsami.9b12319 PMID: 31479233

[317] Pouladi, M.A.; Brillaud, E.; Xie, Y.; Conforti, P.; Graham, R.K.; Ehrnhoefer, D.E.; Franciosi, S.; Zhang, W.; Poucheret, P.; Compte, E.; Maurel, J.C.; Zuccato, C.; Cattaneo, E.; Néri, C.; Hayden, M.R. NP03, a novel low-dose lithium formulation, is neuroprotective in the YAC128 mouse model of Huntington disease. *Neurobiol. Dis.*, 2012, 48(3), 282-289.

http://dx.doi.org/10.1016/j.nbd.2012.06.026 PMID: 22796360

[318] Smith, K.M.; Matson, S.; Matson, W.R.; Cormier, K.; Del Signore, S.J.; Hagerty, S.W.; Stack, E.C.; Ryu, H.; Ferrante, R.J. Dose ranging and efficacy study of high-dose coenzyme Q10 formulations in Huntington's disease mice. *Biochim. Biophys. Acta*, 2006, 1762(6), 616-626.

http://dx.doi.org/10.1016/j.bbadis.2006.03.004 PMID: 16647250