

Clomiphene Citrate and Enclomiphene Hydrochloride Exposure Is Associated With Interfrontal Suture Fusion in Zebrafish

L'exposition au citrate de clomifène ou au chlorhydrate d'enclomiphène, des agents ovulatoires, est liée à la fusion de la suture métopique chez le poisson-zèbre

Plastic Surgery
2023, Vol. 31(4) 383-389
© 2021 The Author(s)



Article reuse guidelines:
sagepub.com/journals-permissions
DOI: 10.1177/22925503211057526
journals.sagepub.com/home/psg



Tamara A. Franz-Odendaal, PhD¹ and Michael Bezuhly, FRCSC²

Abstract

Background: The last several decades have witnessed an increase in metopic craniosynostosis incidence. Population-based studies suggest that pharmacological exposure in utero may be responsible. This study examined effects of the fertility drug clomiphene citrate (CC) on calvarial development in an established model for craniofacial development, the zebrafish *Danio rerio*. **Results:** Zebrafish larvae were exposed to clomiphene citrate or its isomer enclomiphene for five days at key points during calvarial development. Larvae were then raised to adulthood in normal rearing water. Zebrafish were analyzed using whole-mount skeletal staining. We observed differential effects on survivability, growth and suture formation depending on the treatment. Treatments with CC or enclomiphene at 5.5 mm SL led to increased fusion of the interfrontal suture ($p < .01$) compared to controls. **Conclusions:** Exposure to fertility drugs appears to affect development of the cranial vault, specifically the interfrontal suture, in zebrafish. Further research is required to identify the signaling mechanisms at play. This work suggests that fertility drug treatment may contribute to the increased incidence of metopic craniosynostosis observed globally.

Résumé

Historique : L'incidence de craniosynostoses métopiques a augmenté ces dernières décennies. D'après les études en population, l'exposition aux médicaments *in utero* pourrait en être responsable. La présente étude traite des effets d'un agent ovulatoire, le citrate de clomifène (CC), sur le développement de la voûte crânienne d'un modèle établi de développement crâniofacial, le poisson-zèbre *Danio rerio*. **Résultats :** Les chercheurs ont exposé des larves de poisson-zèbre au citrate de clomifène ou à son isomère, l'enclomiphène, sur une période de cinq jours à des moments clés du développement de la voûte crânienne. Ils les ont ensuite élevés jusqu'à l'âge adulte dans de l'eau normale. Ils ont analysé les poissons-zèbres par coloration du squelette entier et observé des effets différenciels sur la capacité de survie, la croissance et la formation des sutures en fonction du traitement. Les traitements au CC ou à l'enclomiphène d'une longueur standard de 5,5 mm ont entraîné une fusion accrue de la suture métopique ($p < 0,01$) par rapport aux sujets témoins. **Conclusions :** L'exposition aux agents ovulatoires semble influer sur le développement de la voûte crânienne, notamment la suture métopique, chez le poisson-zèbre. D'autres recherches devront être réalisées pour déterminer les mécanismes de signalisation en jeu. Selon la présente étude, les agents ovulatoires peuvent contribuer à l'incidence accrue de craniosynostose métopique sur la scène mondiale.

¹ Department of Biology, Mount Saint Vincent University, Mount Saint Vincent University, Halifax, Nova Scotia, Canada

² Division of Plastic Surgery, Dalhousie University, Halifax, Nova Scotia, Canada

Received July 12, 2021. Revised September 24, 2021. Accepted September 30, 2021.

Corresponding Author:

Michael Bezuhly, Division of Plastic Surgery, Dalhousie University, IWK Health Centre, 5850/5980 University Avenue, PO Box 9700, Halifax, Nova Scotia, Canada, B3 K 6R8.
Email: mbezuhly@dal.ca

Keywords

craniosynostoses, trigonocephaly, nonsyndromic, fertility agents, clomiphene, enclomiphene, zebrafish

Mots-clés

agents ovulatoires, clomifène, craniosynostoses, enclomiphène, poisson-zèbre, trigonocéphalie

Introduction

Over the last several decades, paediatric craniofacial centers around the world have reported a dramatic increase in the incidence of children born with metopic craniosynostosis, making it the second most common type of craniosynostosis.^{1–7}

Several epidemiologic studies have suggested that the increase in metopic craniosynostosis cannot be attributed solely to improved diagnosis and awareness of the condition.^{1,3,7} Some have speculated that the increased incidence and spectrum of severity observed in metopic craniosynostosis may be due to a dose-dependent *in utero* pharmacological exposure. Population-based studies have suggested that maternal use of thyroid replacement hormone, valproic acid, or folic acid may be associated with metopic craniosynostosis.^{6,8–10} Unfortunately, rates of use of these agents among the child-bearing age population have remained largely unchanged during the period over which the dramatic rise in metopic craniosynostosis has been observed.

We examined the association between an agent whose increased use has corresponded with the rise in metopic craniosynostosis cases, the fertility drug clomiphene citrate. According to data from the 2002 National Survey of Family Growth, 3.8% of women indicated they used the ovulation drug, a 27% increase from a similar survey conducted in 1995.^{11,12} The drug is comprised of a 60:40 mix of trans- (enclomiphene) and cis-isomers (zucloclomiphene), with the latter having a greater affinity for estrogen receptors. Previous population-based studies have identified an association between craniosynostosis and maternal clomiphene citrate use.^{13,14} Neither of these studies examined the impact of clomiphene citrate on specific type of craniosynostosis. The fact that clomiphene citrate is taken before conception has been cited as evidence against biologic plausibility; however, several investigators have found that zucloclomiphene, the more active isomer in clomiphene citrate preparations, accumulates over consecutive cycles and can be detected in plasma months after administration.^{15,16} Additionally, female mice injected with clomiphene citrate before ovulation in doses paralleling those in humans had an increased risk of exencephaly.¹⁷

In the current study, we investigated whether a causal link exists between clomiphene citrate or enclomiphene hydrochloride administration and metopic craniosynostosis using a zebrafish model. Zebrafish possess unique advantages over mammalian models, including a short reproductive cycle, large number of progeny, high degree of genetic and developmental conservation with humans, and transparent embryos which permits direct visualization of craniofacial morphogenesis.^{18,19} Interestingly,

however, the zebrafish differs from humans and murine models in that the interfrontal suture, the equivalent of the human metopic suture, remains patent throughout the lifespan of the animal.^{20,21} We hypothesized that exposure of fertilized zebrafish embryos and larvae to clomiphene citrate and/or enclomiphene hydrochloride will induce fusion of the interfrontal suture akin to metopic craniosynostosis in humans.

Materials and Methods

Animal Model and Maintenance

Wild-type progeny of AB zebrafish (*Danio rerio*) stocks originally from the Zebrafish International Research Center (ZIRC) were used for all experiments. Animals were raised according to national animal care guidelines and approval for these experiments was received from the Mount Saint Vincent University-Saint Mary's University Joint Animal Care Committee (protocol number 16-03). Larvae were raised to 5.5 mm or 6.5 mm standard length depending on the experimental group. Standard length (SL), defined as the distance from the rostral tip to the caudal peduncle (ie the base of the tail fin), was measured to the nearest decimal with the Nikon NIS Elements image analysis software. These specific SLs were selected as they are points critical to suture development in the zebrafish.²¹

Drug Treatment

Once the specified SL was reached, zebrafish larvae were then treated with either 100 nM clomiphene citrate (CC; Sigma-Aldrich, St. Louis, MO; n=20) or 60 nM enclomiphene hydrochloride (EH; Sigma-Aldrich; n=20) dissolved in 1% dimethyl sulfoxide (DMSO). The doses of each drug used correspond to human serum levels^{15,16} and have been shown to be tolerated by zebrafish.²² Maes et al. have previously shown that low concentrations of DMSO is well-tolerated by zebrafish at various ages (embryos and larvae).²³ Controls included a DMSO solvent control (n=20) at 5.5 mm SL and a no treatment group (n=20). The chemicals were added directly to rearing water for five days of continuous exposure and the fish were fed throughout the experimental period. Five days' duration was selected to ensure as high a dosage as possible that did not result in complete mortality. Water was changed daily and a fresh dose of drug added to the replacement water. After treatment, fish were transferred to new rearing containers containing regular water. Fish were raised for two

months to observe the long-term effects on the calvarial development. Two independent trials were conducted. The proportion of fish that survived to the endpoint in each treatment group was recorded. Fish surviving to two months were then euthanized with 0.1% MS222 Ethyl 3-aminobenzoate, methane sulfonic salt (Sigma-Aldrich). Samples were fixed overnight using 10% neutral buffered formalin.

Whole-Mount Alizarin Red Staining

A standard whole mount acid-free stain²⁴ was used to visualize the craniofacial skeleton in all samples. Bone was stained using Alizarin red S (Sigma-Aldrich). Briefly, the fish were stained overnight in Alizarin red staining solution. Fish were then rinsed in water and bleached in 3% hydrogen peroxide and 1% potassium hydroxide solution for 20 min. Soft tissues were digested with 1% trypsin (Thermo Fisher Scientific, Inc., Waltham, MA) and 2% sodium tetraborate (Sigma-Aldrich) in distilled water for three nights. All specimens were processed through an ascending series of glycerol in 1% potassium hydroxide solution then transferred to a storage solution of 100% glycerol.

Imaging

All samples were examined either using a Nikon SMZ1000 stereomicroscope or a Nikon Eclipse 50i compound microscope. Samples were photographed using a Nikon DXM120° C digital camera in all cases. Fish with insufficient staining were omitted from subsequent skull suture morphology assessment performed by an observer blinded to experimental group.

Statistical Analysis

Survival and body length, expressed as mean \pm standard error of mean, was compared between groups using a one-way ANOVA with Tukey's multiple comparison test. Proportion of fish surviving or exhibiting cranial abnormalities in each group were compared using a Fisher's exact test and Bonferroni correction for pairwise comparisons. Significance was set at $p < .05$. Statistical analyses were performed using Prism version 9.2.0 (GraphPad Software, San Diego, CA).

Results

Survival

While fish treated at 6.5 mm SL tended to have poorer survivability (25-30%) compared to fish treated at 5.5 mm SL (>50%) (Figure 1A), only EH treatment at 6.5 mm SL demonstrated a statistically significant reduction in survivability relative to untreated and DMSO controls ($p < .02$).

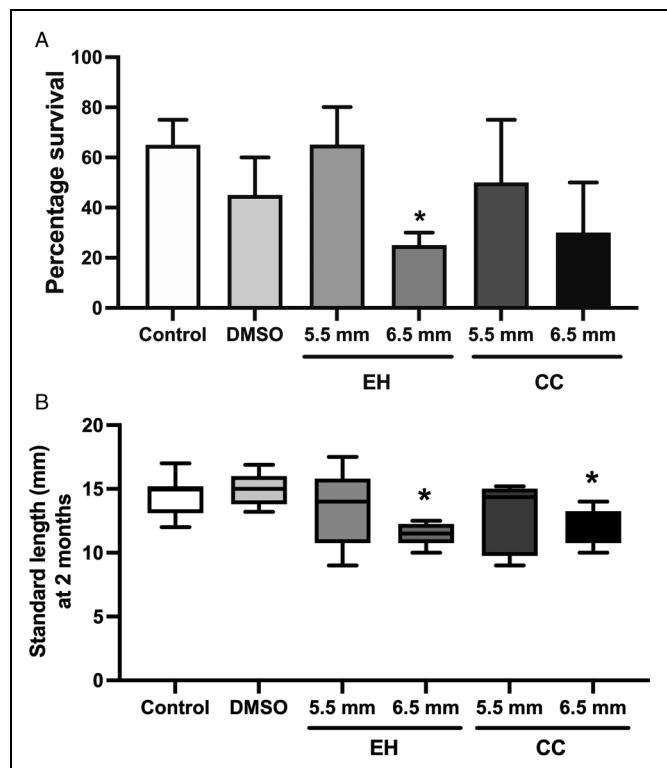


Figure 1. (A) Survivability of zebrafish at two months. Once animals had reached the defined standard length, treatments were initiated and continued until animals were sacrificed two months later. For the DMSO control group, animals were treated at 5.5 mm standard length. DMSO, dimethyl sulfoxide. EH, enclomiphene hydrochloride. CC, clomiphene citrate. * $p = .02$ compared to DMSO control, $n = 20$, experiment performed in duplicate (B) Body length at two months on pooled surviving fish, expressed as Tukey box plot. * $p = .04$ compared to DMSO control, $n \geq 5$.

Growth

Differences in body length were observed after treatment. Based on normative zebrafish growth data, mean SL at 2 months is 14 mm. Among the untreated and DMSO controls, only 15% of fish were less than 14 mm SL at two months of age, whereas 60% of all EH and CC treated fish were less than 14 mm SL. Notably, significant differences in body lengths were observed for fish treated at 6.5 mm SL compared to solvent controls (Figure 1B) indicating an overall stunted growth after treatment with either agent.

Calvarial and Skeletal Development

Whole-mount bone staining the skulls revealed significant effects on calvarial and suture development. In general, the head shapes of treated fish were broader than those of controls, particularly at the level of the occipitoparietal skull. No gross ocular abnormalities were observed. All control fish demonstrated normal skull morphology (Figure 2A), with patent interfrontal sutures (Figure 2B). Of the few DMSO control fish that

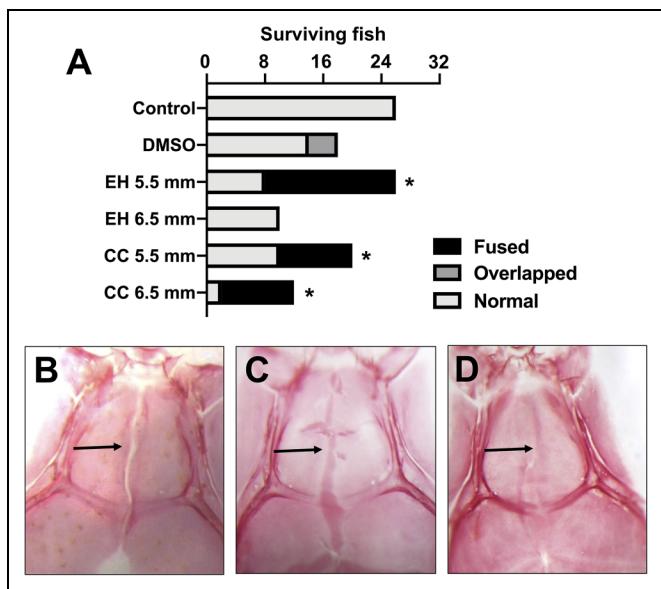


Figure 2. (A) Interfrontal suture morphology in pooled surviving fish. * $p < .01$, $n \geq 5$. (B) Representative whole mount Alizarin red stain of untreated control fish indicating patent interfrontal suture (arrow). (B) Representative stain of control fish showing patent interfrontal suture. (C) Representative stain of a fish showing overlapping interfrontal suture (arrow). (D) Representative stain of a fish showing fused/absent interfrontal suture (arrow).

exhibited cranial vault abnormalities, these were characterized by overlapping sutures (Figure 2C). In contrast, fish treated with EH and CC showed fusion of the interfrontal suture (Figure 2D). Fusion of the interfrontal suture was observed in fish treated with EH and CC at 5.5 mm SL (69% and 50%, respectively; $p < .01$ compared to controls). Furthermore, compared to controls treatment with CC at 6.5 mm SL caused fusion of the interfrontal suture in 83% of the fish ($p < .001$). Owing to poor survivability, a significant difference was not observed between fish treated with EH at 6.5 mm SL and controls.

As calvarial abnormalities have been shown to be associated with rib anomalies and vertebral fusion,²⁵ we also examined the remainder of the zebrafish skeleton to assess if these contributed to the shorter body length observed in treated animals. As demonstrated in Figure 3, no such anomalies were observed.

Discussion

Our results provide direct evidence, for the first time, that the active compounds in common fertility drugs, clomiphene citrate and enclomiphene hydrochloride, can affect calvarial development, specifically at the level of the interfrontal suture. Clomiphene citrate is a non-steroidal ovulation-inducing drug that selectively binds estrogen receptors. Although its fertility activity is due principally to its inhibition of negative feedback of gonadotropin release from the hypothalamic-pituitary-gonadal axis, it also has direct effects on the uterus; its effects on other tissues, including the craniofacial skeleton, have not been investigated.²⁶ Our data suggest that CC may have a greater effect on calvarial development

than EH. This may be due to the greater biological activity of the zuclomiphene isomer in CC. Another possibility is that those animals treated with EH at 6.5 mm SL developed more severe calvarial abnormalities and died prior to two months. Further research is needed to explore this effect in depth.

Interestingly, we observed that the timing of drug administration had a significant impact on survivability, with later exposure more likely to be lethal to the developing larvae. This may be due to effects on other organ systems at later critical stages. For example, between 6.2 to 6.6 mm SL, the fin rays and swim-bladder (an organ critical to controlling buoyancy) are developing.²⁷ We speculate that disruption of these critical structures by CC and EH could have led to poor feeding, growth, and, ultimately, survival. Necropsies and additional dose-response experiments should shine further light on the mechanisms at play.

In addition to effects on survivability, both CC and EH had impacts on body growth. This is in keeping with previous human and mouse studies demonstrating decreased fetal growth with CC exposure.^{17,28} The observed intrauterine growth restriction in animals and humans with CC exposure has been attributed to anti-estrogenic effects on the endometrium causing reduced blood flow or directly on embryogenesis. We speculate that the anti-estrogenic activity of CC has similar effects in zebrafish, as estrogen signaling has been shown to be critical to early zebrafish cardiac and brain development.^{29,30}

The most striking observation of our study is that CC or EH exposure is associated with fusion of the interfrontal suture, a structure that typically remains patent throughout the life of the zebrafish. Aberrations in signaling pathways known to induce craniosynostosis in humans also result in suture fusion in zebrafish, rendering them excellent for modeling of craniosynostosis.³¹ While previous clinical registry studies have examined an association between CC and all types of isolated craniosynostosis,^{13,14} our animal study suggests that the interfrontal suture is more susceptible to disruption by CC than other sutures. We speculate that this is due to signaling pathways unique to interfrontal suture formation compared to other sutures, a hypothesis supported by recent transcriptome analysis of the interfrontal suture in mice.²⁵ As in zebrafish, under normal circumstances the anterior interfrontal suture never fuses in mice.³² Previous work in mice has pointed to a role of estrogen receptor signaling in posterofrontal suture fusion. James et al. showed that estrogen receptor- α is upregulated at the time of suture fusion and that application of the estrogen receptor antagonist fulvestrant can maintain suture patency in postnatal mice.³³ The apparent discrepancy between this latter observation and our own finding that an estrogen receptor antagonist induces suture fusion may be attributable to species- and stage-specific differences in estrogen activity.

While zebrafish have increasingly been used to study craniofacial development over the last 30 years owing to their sequence homology with the human genome, rapidity of embryogenesis, and transparency allowing for visualization of embryogenesis, there are limitations to this animal model.^{34,35}

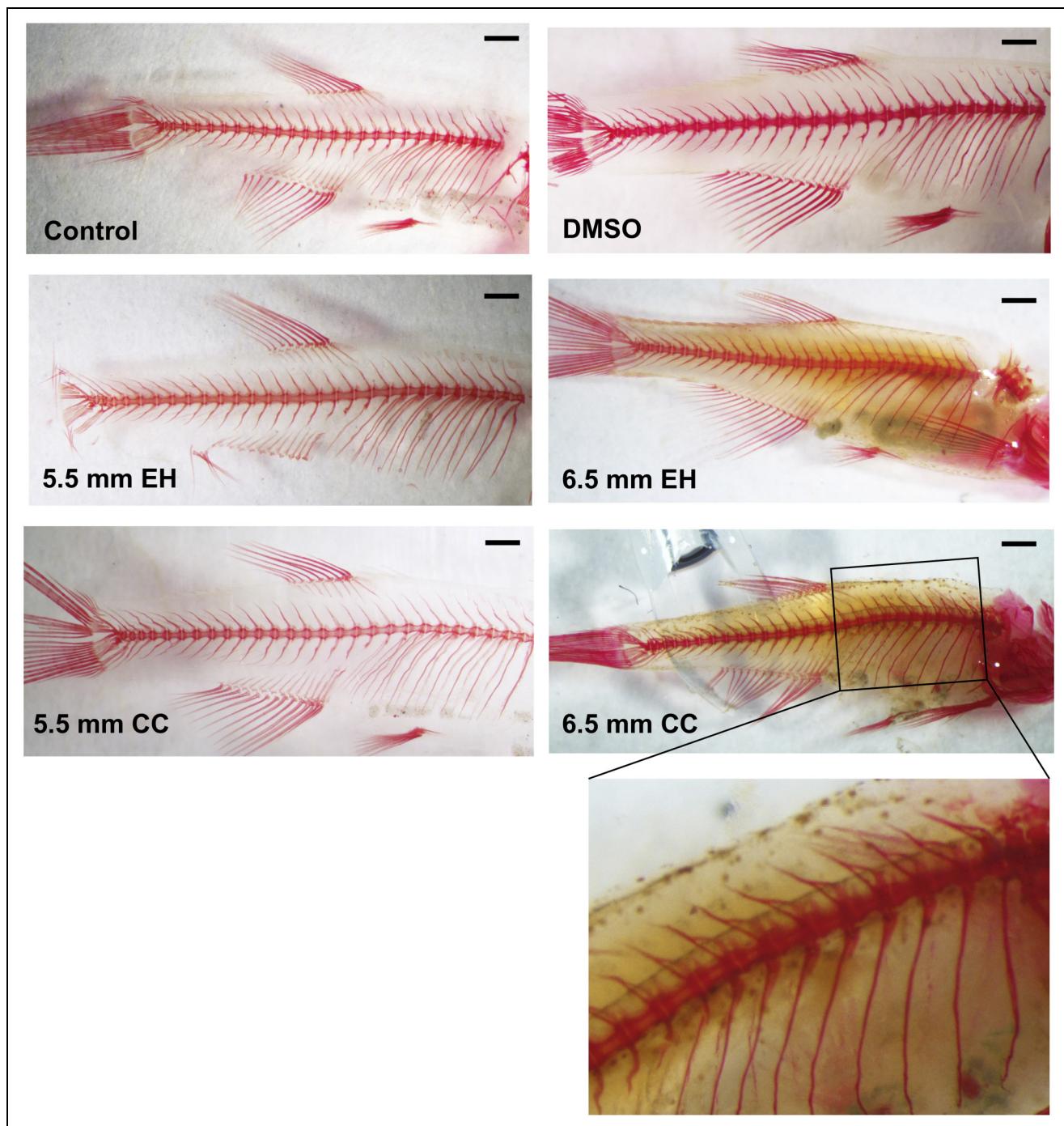


Figure 3. Representative Alizarin red stain of vertebral and rib morphology in pooled surviving fish from each treatment group. At increased magnification, no vertebral fusions are observed (inset). Bar = 1 mm.

Unlike mice and humans, zebrafish cranial development occurs at an earlier stage post-fertilization. As in rodents, but unlike humans, zebrafish sutures remain patent throughout life. Finally, zebrafish bones flanking the coronal sutures derive from mesoderm, whereas in mammals coronal sutures mark the boundary between anterior neural crest bones and posterior mesoderm bones. Such species differences may limit the relevance of our findings to humans.

Despite the limitations of small sample size, this preliminary study points to novel avenues of investigation. Future work will focus on dose-response experiments and necropsies to identify the cause of early lethality noted in the 6.5 mm SL experimental group. While this preliminary study focused on whole-mount staining to specifically examine suture development, we plan to use three-dimensional microcomputed tomography to assess craniofacial morphology in greater depth. Additionally,

while we tested the effect an estrogen receptor antagonist on suture patency, we did not examine estrogen receptor expression, either through immunostaining or in situ hybridization, during zebrafish cranial suture development. Finally, we will examine whether exogenous 17-β estradiol can maintain interfrontal suture patency in fish treated with CC or EH. By further characterizing the expression of estrogen receptors and ligands in the cranial skeleton of zebrafish, we hope to provide essential insights into the signaling pathways that govern the development of metopic craniostenosis.

Ethics Approval

Approval for this work was received from the Mount Saint Vincent University-Saint Mary's University Joint Animal Care Committee (protocol number 16-03). Informed consent was not applicable.

Statement of Financial Interest

The authors have no financial interests or conflicts-of-interest to disclose

Availability of Data and Materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Authors' Contributions

TF-O performed the animal treatments, collected survival data, and performed histological examinations. MB developed the concept, analyzed and interpreted the data and wrote the manuscript. All authors read and approved the final manuscript.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work was supported by the Nova Scotia Health Research Foundation (grant number Development Grant (#2014-15)).

ORCID iD

Michael Bezuhy  <https://orcid.org/0000-0002-7356-5147>

REFERENCES

1. Di Rocco, F, Arnaud, E, Meyer, P, Sainte-Rose, C, & Renier, D. Focus session on the changing "epidemiology" of craniostenosis (comparing two quinquennia: 1985-1989 and 2003-2007) and its impact on the daily clinical practice: a review from Necker Enfants Malades. *Childs Nerv Syst.* 2009;25(7):807-811.
2. Fisher, DC, Kornrumpf, BP, Couture, D, Glazier, SS, Argenta, LC, & David, LR. Increased incidence of metopic suture abnormalities in children with positional plagiocephaly. *J Craniofac Surg.* 2011;22(1):89-95.
3. Kolar, JC. An epidemiological study of nonsyndromal craniostenoses. *J Craniofac Surg.* 2011;22(1):47-49.
4. Kweldam, CF, van der Vlugt, JJ, & van der Meulen, JJ. The incidence of craniostenosis in the Netherlands, 1997-2007. *J Plast Reconstr Aesthet Surg.* 2011;64(5):583-588.
5. Lee, HQ, Hutson, JM, Wray, AC, et al. Changing epidemiology of nonsyndromic craniostenosis and revisiting the risk factors. *J Craniofac Surg.* 2012;23(5):1245-1251.
6. van der Meulen, J, van der Hulst, R, van Adrichem, L, et al. The increase of metopic synostosis: a pan-European observation. *J Craniofac Surg.* 2009;20(2):283-286.
7. Selber, J, Reid, RR, Chike-Obi, CJ, et al. The changing epidemiologic spectrum of single-suture synostoses. *Plast Reconstr Surg.* 2008;122(2):527-533.
8. Rasmussen, SA, Yazdy, MM, Carmichael, SL, Jamieson, DJ, Canfield, MA, & Honein, MA. Maternal thyroid disease as a risk factor for craniostenosis. *Obstet Gynecol.* 2007;110(2 Pt 1):369-377.
9. Ardinger, HH, Atkin, JF, Blackston, RD, et al. Verification of the fetal valproate syndrome phenotype. *Am J Med Genet.* 1988;29(1):171-185.
10. Kallen, B, & Robert-Gnansia, E. Maternal drug use, fertility problems, and infant craniostenosis. *Cleft Palate Craniofac J.* 2005;42(6):589-593.
11. Chandra, A, Martinez, GM, Mosher, WD, Abma, JC, & Jones, J. Fertility, family planning, and reproductive health of U.S. Women: data from the 2002 national survey of family growth. *Vital Health Stat.* 2005;23(25):1-160.
12. Abma, JC, Chandra, A, Mosher, WD, Peterson, LS, & Piccinino, LJ. Fertility, family planning, and women's Health: new data from the 1995 national survey of family growth. *Vital Health Stat.* 1997;23(19):136-137.
13. Reehuis, K, Honein, MA, Shaw, GM, & Romitti, PA. Fertility treatments and craniostenosis: California, Georgia and Iowa, 1993-1997. *Pediatrics.* 2003;111(5 Pt 2):1163-1166.
14. Reehuis, J, Honein, MA, Schieve, LA, et al. Use of clomiphene citrate and birth defects, national birth defects prevention study, 1997-2005. *Hum Reprod.* 2011;26(2):451-457.
15. Mikkelson, TJ, Kroboth, PD, Cameron, WJ, Dittert, LW, Chungi, V, & Manberg, PJ. Single-dose pharmacokinetics of clomiphene citrate in normal volunteers. *Fertil Steril.* 1986;46(3):392-396.
16. Young, SL, Opsahl, MS, & Fritz, MA. Serum concentrations of enclomiphene and zuclomiphene across consecutive cycles of clomiphene citrate therapy in anovulatory infertile women. *Fertil Steril.* 1999;71(4):639-644.
17. Dziadek, M. Preovulatory administration of clomiphene citrate to mice causes fetal growth retardation and neural tube defects (exencephaly) by an indirect maternal effect. *Teratology.* 1993;47(4):263-273.
18. Sheehan-Rooney, K, Pálinkášová, B, Eberhart, JK, & Dixon, MJ. A cross-species analysis of Satb2 expression suggests deep conservation across vertebrate lineages. *Dev Dyn.* 2010;239(12):3481-3491.
19. Quarto, N, & Longaker, MT. The zebrafish (*danio rerio*): a model system for cranial suture patterning. *Cells Tissues Organs.* 2005;181(2):109-118.

20. Kanther, M, Scalici, A, Rashid, A, Miao, K, Van Deventer, E, & Fisher, S. Initiation and early growth of the skull vault in zebrafish. *Mech Dev.* 2019;160:103578.
21. Topczewska, JM, Shoela, RA, Tomaszewski, JP, Mirmira, RB, & Gosain, AK. The morphogenesis of cranial sutures in zebrafish. *PLoS ONE.* 2016;11(11):e0165775.
22. Hamad, A, Kluk, M, Fox, J, Park, M, & Turner, JE. The effects of aromatase inhibitors and selective estrogen receptor modulators on eye development in the zebrafish (*danio rerio*). *Curr Eye Res.* 2007;32(10):819-827.
23. Maes, J, Verlooy, L, Buenafe, OE, de Witte, PA, Esguerra, CV, & Crawford, AD. Evaluation of 14 organic solvents and carriers for screening applications in zebrafish embryos and larvae. *PLoS One.* 2012;7(10):e43850.
24. Franz-Odendaal, TA, Ryan, K, & Hall, BK. Developmental and morphological variation in the teleost craniofacial skeleton reveals an unusual mode of ossification. *J Exp Zool B Mol Dev Evol.* 2007;308(6):709-721.
25. Holmes, G, Gonzalez-Reiche, AS, Lu, N, et al. Integrated transcriptome and network analysis reveals spatiotemporal dynamics of calvarial suturegenesis. *Cell Rep.* 2020;32(1):107871.
26. Rostami-Hodjegan, A, Lennard, MS, Tucker, GT, & Ledger, WL. Monitoring plasma concentrations to individualize treatment with clomiphene citrate. *Fertil Steril.* 2004;81(5):1187-1193.
27. Parichy, DM, Elizondo, MR, Mills, MG, Gordon, TN, & Engeszer, RE. Normal table of postembryonic zebrafish development: staging by externally visible anatomy of the living fish. *Dev Dyn.* 2009;238(12):2975-3015.
28. Forman, R, Gill, S, Moretti, M, Tulandi, T, Koren, G, & Casper, R. Fetal safety of letrozole and clomiphene citrate for ovulation induction. *J Obstet Gynaecol Can.* 2007;29(8):668-671.
29. Diamante, G, Menjivar-Cervantes, N, Leung, MS, Volz, DC, & Schlenk, D. Contribution of G protein-coupled estrogen receptor 1 (GPER) to 17 β -estradiol-induced developmental toxicity in zebrafish. *Aquat Toxicol.* 2017;186:180-187.
30. Hao, R, Bondesson, M, Singh, AV, et al. Identification of estrogen target genes during zebrafish embryonic development through transcriptomic analysis. *PLoS One.* 2013;8(11):e79020.
31. Laue, K, Pogoda, HM, Daniel, PB, et al. Craniosynostosis and multiple skeletal anomalies in humans and zebrafish result from a defect in the localized degradation of retinoic acid. *Am J Hum Genet.* 2011;89(5):595-606.
32. Sahar, DE, Longaker, MT, & Quarto, N. Sox9 neural crest determinant gene controls patterning and closure of the posterior frontal cranial suture. *Dev Biol.* 2005;280(2):344-361.
33. James, AW, Theologis, AA, Brugmann, SA, et al. Estrogen/estrogen receptor alpha signaling in mouse. Posteriorfrontal cranial suture fusion. *PLoS One.* 2009;4(9):e7120.
34. Morris-Kay, GM, & Wilkie, AO. Growth of the normal skull vault and its alteration in craniosynostosis: insights from human genetics and experimental studies. *J Anat.* 2005;207(5):637-653.
35. Cornille, M, Dambroise, E, Komla-Ebri, D, et al. Animal models of craniosynostosis. *Neurochirurgie.* 2019;65(5):202-209.