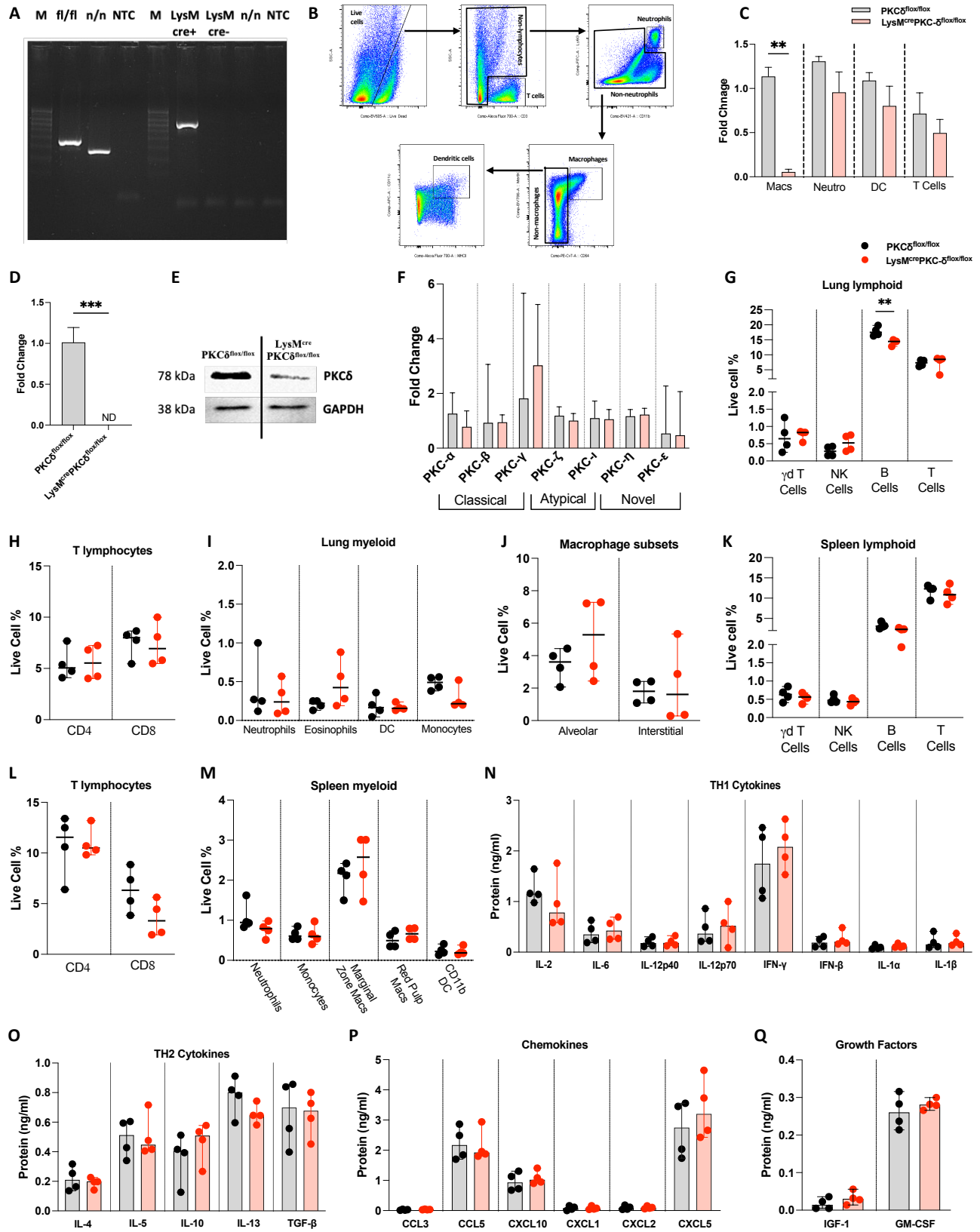
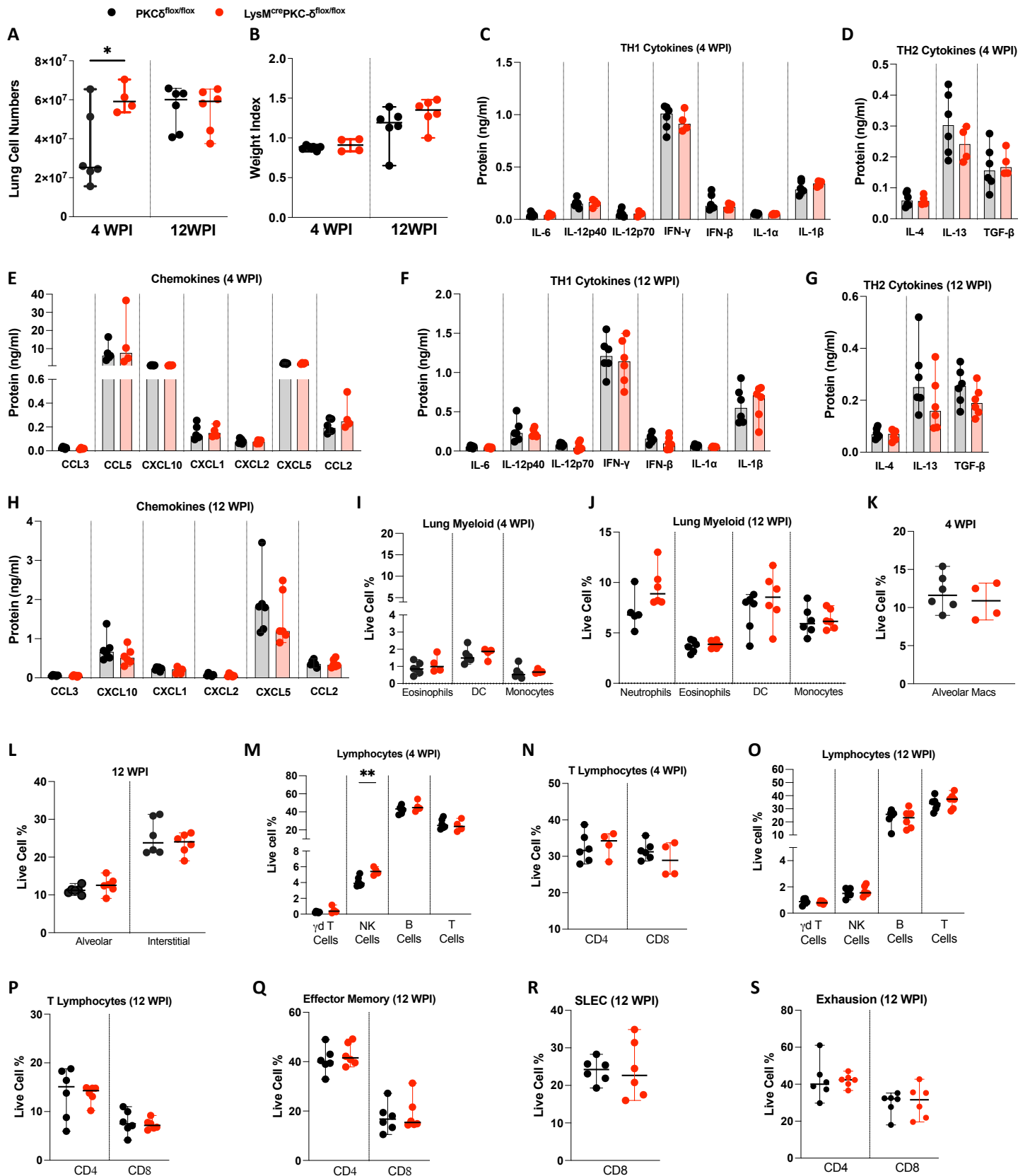


Supplementary Fig 1.



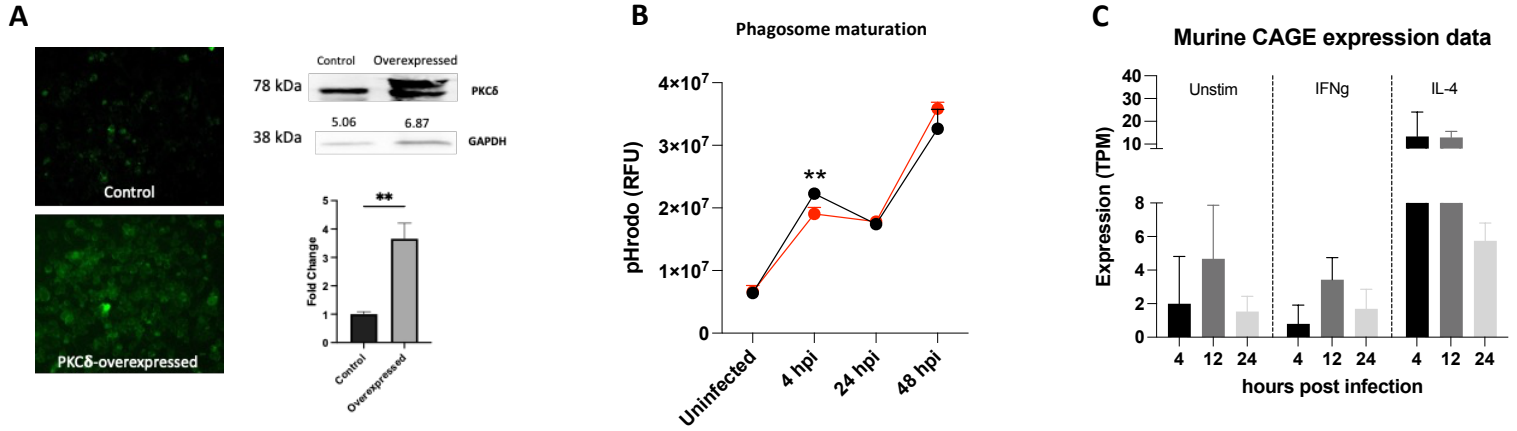
Supplementary Figure 1. $\text{LysM}^{\text{cre}}\text{PKC}\delta^{\text{flox/flox}}$ mice are indistinguishable from $\text{PKC}\delta^{\text{flox/flox}}$ littermate control at the naive state. [A] Integrity of loxP-floxed $\text{PKC}\delta$ and presence of LysM^{cre} was confirmed in the genomic DNA extracted from the tail cuts of $\text{PKC}\delta^{\text{flox/flox}}$ and $\text{LysM}^{\text{cre}}\text{PKC}\delta^{\text{flox/flox}}$ mice. Representative agarose gel demonstrates the marker (m), wild-type (n/n), floxed $\text{PKC}\delta$ (fl/fl), $\text{LysM}^{\text{cre}}\text{PKC}\delta^{\text{flox/flox}}$ (LysM^{cre} +), $\text{PKC}\delta^{\text{flox/flox}}$ (LysM^{cre} -), non-template control (NTC). [B] Gating strategy for lung sorted immune cells and confirmation of $\text{PKC}\delta$ deletion in macrophages by qRT-PCR. [C] $\text{PKC}\delta$ expression was determined in sorted immune cell populations by qRT-PCR. Expression levels are normalized to the endogenous housekeeping gene *Hprt*. [D-E] Deletion of $\text{PKC}\delta$ in bone-marrow derived macrophages was confirmed by qRT-PCR and western blot analysis. *HPRT* and *GAPDH* were used as a normalizing control in qRT-PCR and Western blot respectively. [F] Protein kinase C isoform-specific primers were used for gene expression analysis in $\text{PKC}\delta$ deficient bone-marrow derived macrophages ($\text{LysM}^{\text{cre}}\text{PKC}\delta^{\text{flox/flox}}$) as compared to the control ($\text{PKC}\delta^{\text{flox/flox}}$) by qRT-PCR. [G-M] Various lymphoid and myeloid immune populations were detected by flow cytometry in the lung [G-J] and spleen [K-M] in both $\text{PKC}\delta^{\text{flox/flox}}$ and $\text{LysM}^{\text{cre}}\text{PKC}\delta^{\text{flox/flox}}$ mice. [N-Q] Determining cytokine levels in lung homogenates collected from $\text{PKC}\delta^{\text{flox/flox}}$ and $\text{LysM}^{\text{cre}}\text{PKC}\delta^{\text{flox/flox}}$ mice. All data shown mean $\pm$ SD and is representative of two independent experiments with n=4 mice/group. Statistical analyses were performed using an unpaired student t-test. Asterisks are defining significance compared to the control group as: **p < 0.01, ***p < 0.001.

Supplementary Fig 2.



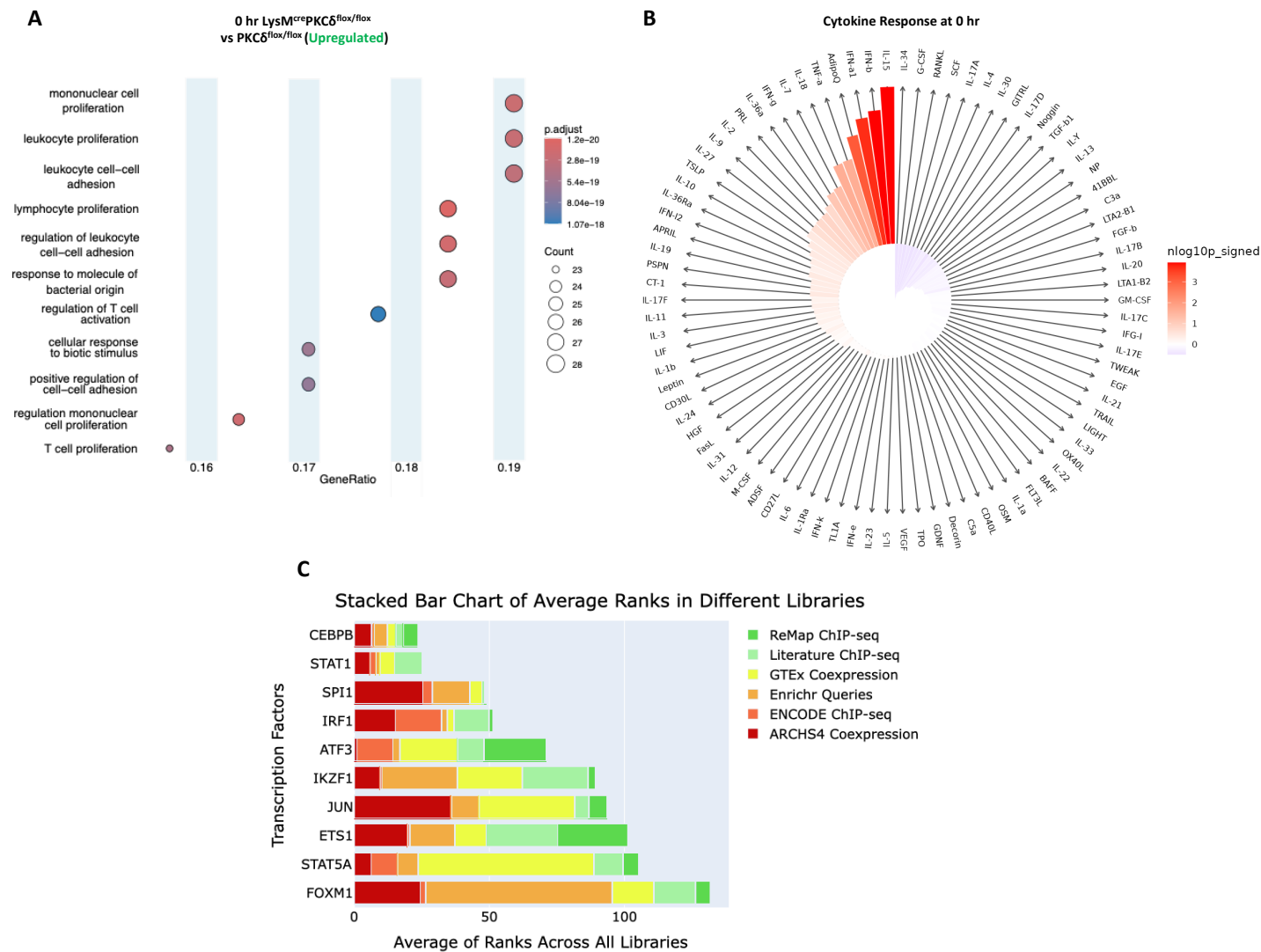
Supplementary Figure 2. Lung cell numbers, cytokine profiling, and immune cell phenotyping of LysM^{cre}PKC δ ^{flox/flox} mice during acute (4WPI) and chronic (12 WPI) *Mtb* infection. [A] Lung cell numbers and [B] weight index were determined at acute and chronic stage of *Mtb* infection. [C-H] Determining cytokine and chemokine levels in lung homogenates collected from PKC δ ^{flox/flox} and LysM^{cre}PKC δ ^{flox/flox} mice at acute and chronic stage of *Mtb* infection. [I-S] Various myeloid and lymphoid immune cell populations were detected by flow cytometry at acute and chronic stage of *Mtb* infection. All data shown mean $\pm$ SD and is representative of two independent experiments with n=4-6 mice/group. Statistical analyses were performed using an unpaired student t-test. Asterisks are defining significance compared to the control group as: *p < 0.05, **p < 0.01.

Supplementary Fig 3.



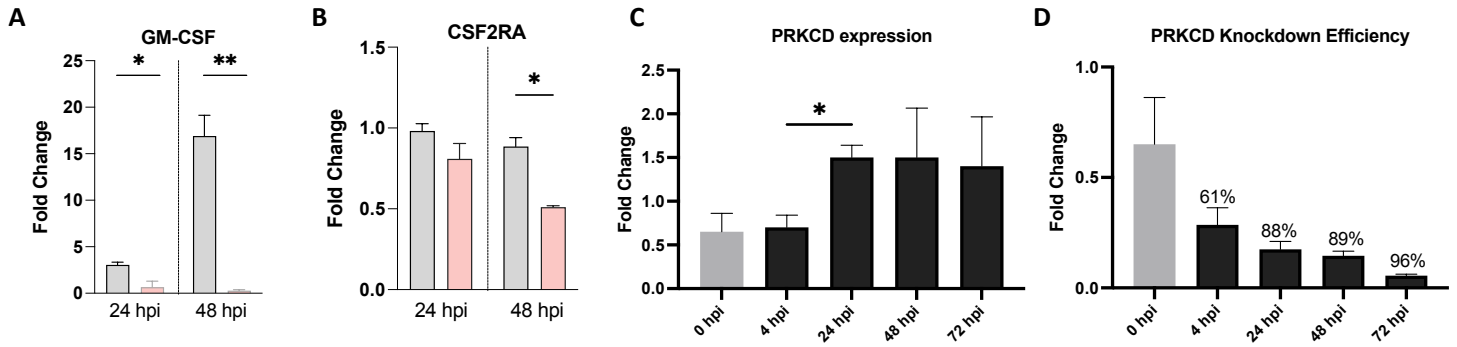
Supplementary Figure.3. PKC δ overexpression in RAW264.7 murine macrophages, determination of phagosome maturation in LysM^{cre}PKC δ ^{flox/flox} BMDMs and murine PKC δ expression during *Mtb* infection (adapted from FANTOM5 CAGE database). [A] Lentivirus-mediated overexpression of PKC δ in RAW264.7 murine macrophage cell line confirmed by ZOE fluorescence imaging, western blot, and qRT-PCR. [B] Determination of phagosome maturation (Relative fluorescence unit) based on pH change in the phagolysosomal compartment in bone-marrow-derived macrophages from LysM^{cre}PKC δ ^{flox/flox} and PKC δ ^{flox/flox} mice. [C] Determining PKC δ expression in classically (IFN γ -stimulated) or alternatively (IL-4 stimulated) activated murine macrophages utilizing publicly available FANTOM5 CAGE database. Asterisks are defining significance compared to the control group as: **p < 0.01.

Supplementary Fig 4.



Supplementary Figure.4. Gene ontology, cytokine response (IREA), and ChEA analysis in the LysM^{cre}PKC $\delta^{\text{flox/flox}}$ BMDMs at naive state. [A] Horizontal dot plots (Ora) detailing the association of enriched Gene Ontology (GO) biological processes are shown between LysM^{cre}PKC $\delta^{\text{flox/flox}}$ and PKC $\delta^{\text{flox/flox}}$ BMDMs at 0 hr (uninfected) time point. [B] IREA cytokine enrichment plot showing the enrichment score (ES) for each of the cytokine response in LysM^{cre}PKC $\delta^{\text{flox/flox}}$ BMDMs at 0 hr (uninfected) time point. Bar length is representing the ES with darker red (enriched in LysM^{cre}PKC $\delta^{\text{flox/flox}}$ BMDMs) and darker blue (enriched in PKC $\delta^{\text{flox/flox}}$ BMDMs). [C] Horizontal bar chart representing the top ranked transcription factors at 0 hr (uninfected) time point according to their average integrated scores across all the libraries. All data shown are analysed and produced using R studio packages and appyters web-based software.

Supplementary Fig 5.



Supplementary Figure.5. mRNA expression of GM-CSF and CSF2RA in $\text{LysM}^{\text{cre}}\text{PKC}\delta^{\text{flo}/\text{flo}}$ BMDMs, mRNA expression of PKC δ in MDMs and siRNA mediated PKC δ knockdown efficiency in MDMs during *Mtb* infection. [A-B] mRNA expression of GM-CSF and CSF2RA in bone-marrow-derived macrophages from $\text{LysM}^{\text{cre}}\text{PKC}\delta^{\text{flo}/\text{flo}}$ and $\text{PKC}\delta^{\text{flo}/\text{flo}}$ mice during *Mtb* infection at indicated time points. [C] mRNA expression of PRKCD in human monocyte-derived macrophages at indicated time points during *Mtb* infection. [D] siRNA mediated knockdown efficiency of PRKCD in human monocyte-derived macrophages during *Mtb* infection. Asterisks are defining significance compared to the control group as: * $p < 0.05$, ** $p < 0.01$.

Table S1.

Gene	Accession number	Forward Primer (5'-3')	Reverse Primer (5'-3')
Mouse PKCδ	M69042	CTGGGTAACCTTAACAAGACC	CTGCTAAATAACATGTTCGGTCC
Mouse PKCϵ	M18331	CATCGATCTCTCGGGATCATCG	CGGTTGTCAAATGACAAGGCC
Mouse PKCη	M62980	AGCTAGCCGTCTTCCACGAGACG C	GGACGACGCAGGTGCACACTTG G
Mouse PKCθ	D11061	AGCTAGCCGTCTTCCACGAGACG C	GGACGACGCAGGTGCACACTTG G
Mouse caspase 1	NM_009807	ACAAGGCACGGGACCTATG	TCCCAGTCAGTCCTGGAAATG
Mouse caspase 11	NM_009807	AGAGGGCATGGAGTCAGAGA	GCCATGAGACATTAGCACCA
Mouse NLRP3	NM_145827	ATTACCCGCCCGAGAAAGG	TCGCAGCAAAGATCCACACAG
Mouse AIM2	NM_001013779	GTCACCAGTTCCTCAGTTGTG	CACCTCCATTGTCCCTGTTTTAT
Mouse IL-1β	NM_008361	GCTTCAGGCAGGCAGTATC	AGGATGGGCTCTTCTTCAAAG
Mouse IL-18	NM_008360	ACTTTGGCCGACTTCACTGT	GGGTTCACTGGCACTTTGAT
Mouse GM-CSF	XM_006532127	ACCACCTATGCGGATTTTCAT	TCATTACGCAGGCACAAAAC
Mouse CSF2RA	NM_009970	ACGTGGCGCGATGCAT	ACTTGTCAGTCTGCTGGGGAGTG
Mouse iNOS	NM_011198	AGCCCTCACCTACTTCCTG	CAATCTCTGCCTATCCGTCTC
Human PRKCD	L07860	CACCATCTTCCAGAAAGAACG	CTTGCCATAGGTCCCGTTGTTG