



Emergence and Evolution of Unique Plasmids Harboring *bla*_{IMP-70} and *bla*_{CTX-M-253} in Multidrug-Resistant *Providencia rettgeri*

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ABSTRACT Although the prevalence of carbapenem-resistant Enterobacterales remains low in Japan, these bacteria are a growing problem worldwide, owing to their multidrug resistance phenotype. We isolated a multidrug-resistant *Providencia rettgeri* strain, NR1418, harboring a rare $bla_{\rm IMP}$ variant, $bla_{\rm IMP-70}$, a novel $bla_{\rm CTX-M}$ variant, designated $bla_{\rm CTX-M-253}$, and $bla_{\rm MOX-1}$. This strain is resistant to β -lactams, amikacin, levofloxacin, and colistin. Genomic analysis revealed that NR1418 carries two plasmids, designated pNR1418-1 and pNR1418-2. The pNR1418-1 plasmid harbors $bla_{\rm CTX-M-253}$, $bla_{\rm TEM-1}$, and $bla_{\rm MOX-1}$, while the pNR1418-2 plasmid harbors $bla_{\rm IMP-70}$, which is located in a class 1 integron. Both plasmids exhibit high similarities with the plasmid of the *P. rettgeri* isolate BML2526, which also harbors $bla_{\rm IMP-70}$ and was identified in the same region of Japan as NR1418 at a different point in time. This indicates the possibility of the emergence and evolution of IMP-70producing *P. rettgeri* and suggests that the plasmid of BML2526 may have occurred following recombination of the two plasmids harbored by NR1418. Further, $bla_{\rm IMP-70}$ and $bla_{\rm CTX-M-253}$ were found on unique plasmids, indicating that they likely evolved through mutations and recombination.

IMPORTANCE Although *Providencia rettgeri* is an opportunistic pathogen, its intrinsic resistance to colistin and tigecycline makes the treatment of carbapenem-resistant *P. rettgeri* challenging. We isolated a multidrug-resistant *P. rettgeri* strain which harbored a rare bla_{IMP} variant, $bla_{IMP-70'}$ a novel bla_{CTX-M} variant, $bla_{CTX-M-253'}$, and bla_{MOX-1} from a urinary sample obtained in Osaka, Japan. We investigated its genetic structure and evaluated the evolution of the plasmids carrying these genes. We show that $bla_{IMP-70'}$, $bla_{CTX-M-253'}$, and bla_{MOX-1} are present on unique plasmids and that they have high similarities to the plasmid of another IMP-70-producing *P. rettgeri* isolate that was identified as being from the same location. The evolution of plasmids through mutations and recombination may play a role in the development and spread of multidrug resistance.

KEYWORDS *Providencia rettgeri*, multidrug-resistant Enterobacterales, β -lactamase, IMP-70, CTX-M-253, plasmid

P rovidencia rettgeri, a member of Enterobacterales, is an emerging opportunistic pathogen that is often associated with urinary tract infections. The spread of carbapenem-resistant Enterobacterales (CRE) is a growing problem worldwide, as they exhibit multidrug resistance. *P. rettgeri* is usually susceptible to most antibiotics; however, it is intrinsically resistant to colistin and tigecycline, making the treatment of carbapenem-resistant *P. rettgeri* challenging. Recently, carbapenem-resistant *P. rettgeri* has been reported in several countries (1–3). NDM-1-positive *P. rettgeri* plays a major role in the spread of antibiotic resistance in Latin America (4). In Japan, the prevalence of CRE remains low (0.31% in 2020) (5), and the IMP-type metallo- β -lactamase is the most common carbapenemase (6). Although carbapenem-resistant

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TABLE 1 Susceptibilities of Providencia	<i>rettaeri</i> NR1418 harborin	a blaimp 70, blacty M 252	, blatem 1, and blamov	1 and its transformants

		MIC (µg/mL) ^a													
Strains	eta-Lactamase genes	PIP	TZP	CPD	стх	CTX+CLA	CAZ	FEP	CMZ	ATM	IPM	MEM	LVX	AMK	CST
P. rettgeri NR1418	bla _{IMP-70} , bla _{CTX-M-253} , bla _{MOX-1} , bla _{TEM-1}	>256	32	>256	>256	>256	>256	>256	>256	32	>256	>256	32	256	>256
<i>E. coli</i> pNR1418-1/ TOP10	bla _{CTX-M-253} , bla _{MOX-1} , bla _{TEM-1}	>256	4	256	128	16	8	2	16	4	0.25	≤0.06	≤0.06	2	0.5
<i>E. coli</i> pNR1418-2/ TOP10	bla _{IMP-70}	2	2	>256	128	64	>256	32	128	≤0.06	4	8	≤0.06	32	0.25
E. coli TOP10		2	2	1	≤0.06	≤0.06	0.25	≤0.06	1	≤0.06	0.25	≤0.06	≤0.06	2	0.5

^aAntibiotics: PIP, piperacillin; TZP, piperacillin-tazobactam; CPD, cefpodoxime; CTX, cefotaxime; CLA, clavulanic acid; CAZ, ceftazidime; FEP, cefepime; CMZ, cefmetazole; ATM, aztreonam; IPM, imipenem; MEM, meropenem; LVX, levofloxacin; AMK, amikacin; CST, colistin.

P. rettgeri has been infrequently reported, there are a few reports of IMP-producing *P. rettgeri* (7–9). In this study, we isolated a multidrug-resistant *P. rettgeri* strain, NR1418, harboring a rare $bla_{\rm IMP}$ variant, $bla_{\rm IMP-70}$, which has only been reported in Japan (9). This isolate also carries a novel $bla_{\rm CTX-M}$ variant, $bla_{\rm CTX-M-253}$, and $bla_{\rm MOX-1}$. We further investigated the genetic structure of *P. rettgeri* coharboring $bla_{\rm IMP-70}$, $bla_{\rm CTX-M-253}$, and $bla_{\rm MOX-1}$. Using genomic analysis and evaluated the evolution of the plasmids carrying these genes.

NR1418 was isolated from a urinary sample from a catheterized patient at a hospital in Osaka, Japan, in 2015. NR1418 was identified as *P. rettgeri* using matrix-assisted laser desorption ionization time-of-flight mass spectrometry (VITEK MS). Antimicrobial susceptibilities to antibiotics, excluding colistin, were determined using the agar dilution method; the susceptibility to colistin was determined using the broth microdilution method, according to the guidelines of the Clinical and Laboratory Standards Institute (10). NR1418 was resistant to all antibiotics tested, including β -lactams, amikacin, levofloxacin, and colistin (Table 1). The results of the carbapenemase inactivation method (11, 12) and the modified carbapenemase inactivation method (13) were positive, indicating that NR1418 produces carbapenemase.

β-Lactamase genes were identified using PCR and Sanger sequencing (14, 15), which revealed that NR1418 harbors bla_{IMP-70} , bla_{TEM-1} , bla_{MOX-1} , and a novel CTX-M β-lactamase gene, $bla_{CTX-M-253}$ (GenBank accession no. LC670768). CTX-M-253 is identical to $bla_{CTX-M-2}$, except for a single amino acid substitution, Ala80Val. This substitution is associated with higher structural stability and catalytic activity in CTX-M-55, which is a CTX-M-15 variant resulting from the same substitution (16). The role of Ala80Val in increasing resistance or stability has been reported for other β-lactamases, as well (17, 18), and the substitution may play a role in CTX-M-253. However, further investigation is needed to determine the actual changes in the structure and activity of CTX-M-253 compared to those of CTX-M-2. IMP-70 (GenBank accession no. LC348383) is an IMP variant that we registered in 2017. IMP-70 differs from IMP-10 by a single amino acid substitution, Phe69Val, and from IMP-1 by an additional substitution, Val49Phe.

The transferability of the β -lactamase genes was tested using filter mating experiments with *Escherichia coli* J53 as the recipient strain (19). However, the genes were not transferred by conjugation. Instead, they were introduced into *E. coli* TOP10 (Thermo Fisher Scientific, Waltham, MA, USA) by transformation. Two types of transformants were obtained. One transformant harbored $bla_{CTX-M-253}$, bla_{TEM-1} , and bla_{MOX-1} (pNR1418-1/TOP10), while the other transformant harbored bla_{IMP-70} (pNR1418-2/TOP10). It was also found that pNR1418-1/TOP10 was resistant to piperacillin, cefotaxime, and cefpodoxime and was inhibited by clavulanic acid. Further, pNR1418-2/TOP10 was resistant to cephems and carbapenems.

The whole-genome of NR1418 was sequenced using MiSeq (Illumina) and MinION (Oxford Nanopore), and a hybrid *de novo* assembly was performed with Unicycler v0.4.8 (20). The genome sequences were annotated using DFAST v1.5.0 (21) and corrected manually using BLAST (https://blast.ncbi.nlm.nih.gov/Blast.cgi). Resistance genes were analyzed using ResFinder 4.1 (22). The assembled genome consisted of three circular contigs: a 4,371,793 bp chromosome, a 172,709 bp plasmid, pNR1418-1 (GenBank accession no. AP025670), and a 128,012 bp plasmid, pNR1418-2 (GenBank accession no. AP025671). The average depth was 260x. An average nucleotide identity (ANI) analysis performed using JSpeciesWS

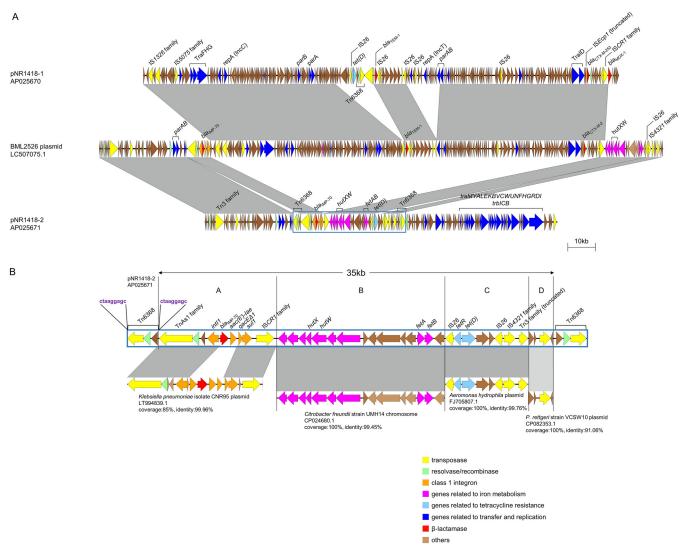


FIG 1 Genetic environments of plasmids harbored by IMP-70-producing *Providencia rettgeri*. (A) BLAST comparison of pNR1418-1, pNR1418-2, and the plasmid of BML2526. The comparisons were visualized with Easyfig version 2.2.2 (25). The plasmid of BML2526 consists of regions identical to parts of pNR1418-1 and pNR1418-2. Dark gray bars indicate nucleotide identity >99% in a window of 3000 bp. The 35 kb region flanked by Tn6368 in pNR1418-2 is shown inside a blue box, and its genetic structures are described in panel B. (B) Schematic representation of the 35 kb region flanked by Tn6368 in pNR1418-2. Structures with high similarity to each region are shown in the bottom row. Region A is identical to a structure identified in a plasmid of *Klebsiella pneumoniae* isolate, except for the gene cassettes in the class 1 integron. Structures highly similar to regions B through D can be found in other strains. Transposases are seen at the border of every region. The 9 bp TSD pattern flanking Tn6368 is shown in purple. Dark and light gray bars indicate nucleotide identity >99% and >90%, respectively, in a window of 1000 bp.

(23) with *P. rettgeri* DSM 1131 (GenBank accession no. ACCl02000000), *P. rettgeri* Dmel1 (GenBank accession no. AJSB01000000), *P. rustigianii* NCTC11802 (GenBank accession no. UGTY01000000), *P. alcalifaciens* 205/92 (GenBank accession no. JALD01000000), and *P. stuartii* INSRA21868 (GenBank accession no. LGYB01000000) as reference genomes showed ANI values (ANIb) of 98.48, 91.23, 77.55, 77.49, and 77.21%, respectively. Thus, NR1418 was identified as *P. rettgeri* through ANI analysis, as well. The identification of plasmid replicons using PlasmidFinder (24) showed that pNR1418-1 hosts IncC and IncT replicons, whereas pNR1418-2 harbors a col3M replicon.

The BLAST analysis showed that pNR1418-1 harbors $bla_{CTX-M-253}$, bla_{TEM-1} , and bla_{MOX-1} (Fig. 1A). A truncated ISEcp1 was found upstream of $bla_{CTX-M-253}$, and bla_{MOX-1} was located 6.9 kb downstream. The pNR1418-2 plasmid harbors $bla_{IMP-70'}$ located in a class 1 integron structure along with aac(6')-lae, encoding aminoglycoside resistance. Both pNR1418-1 and pNR1418-2 have tra genes, and pNR1418-2 also has trb genes; however, these plasmids were

not transferred by conjugation in this study. Further study is needed to determine if other factors, such as the compatibility of the recipient strain and the plasmids, may be involved. Both pNR1418-1 and pNR1418-2 had high similarities with the plasmid of the *P. rettgeri* isolate BML2526, which harbors bla_{IMP-70} (9). BML2526 harbors bla_{IMP-70} , $bla_{CTX-M-2}$, and bla_{TEM-1} , but it does not harbor bla_{MOX-1} , and bla_{IMP-70} is located in the same integron as pNR1418-2. In addition, the structure surrounding $bla_{CTX-M-2}$ is similar to that surrounding $bla_{CTX-M-253}$, indicating that $bla_{CTX-M-253}$ may have occurred as a result of a point mutation in $bla_{CTX-M-2}$.

The integron containing *bla*_{IMP-70} in pNR1418-2 is located within a region flanked by two copies of Tn6368. This approximately 35 kb region could be divided further into four regions according to its similarity with other reported structures (Fig. 1B). Region A includes the class 1 integron containing bla_{IMP-70} and has a Tn3-like element TnAs1 family transposase and an IS91-like element ISCR1 family transposase, at either end. Region B mostly consists of genes involved in iron metabolism and shares 99.45% identity with the chromosome of Citrobacter freundii strain UMH14 (GenBank accession no. CP024680.1). Region C has a tetracycline resistance gene, tet(D), as well as its transcriptional regulator, tetR, flanked by two copies of IS26 and followed by an IS4321 family transposase and a truncated Tn3 family transposase. Region D consists mostly of hypothetical proteins and shows the highest similarity to the P. rettgeri strain VCSW10 plasmid pVCSW10.2 (GenBank accession no. CP082353.1), with a shared identity of 91.06%. The remaining region of pNR1418-2 shows the highest similarity to the Pectobacterium carotovorum plasmid Drgb2 (GenBank accession no. KT351733.1), with a coverage of 62% and a shared identity of 96.97%. Analysis of target site duplication (TSD) patterns revealed a 9 bp TSD pattern (ctaaggagc) flanking Tn6368, located next to region A. As transposases of transposons and insertion elements are located at the border of each region, it is possible that the accumulation of several regions via transposable elements resulted in pNR1418-2. An exact match of region A was found in the plasmid of BML2526; however, an approximately 140 kb region similar to pNR1418-1 exists between sul1 and the ISCR1 family transposase. Regions similar to regions B and D follow; however, some parts of region C are missing, except for IS26, the IS4321 family transposase, and the truncated Tn3 family transposase. The region 18 kb upstream of region A is also identical to a region in BML2526. No copies of Tn6368 were found on the plasmid of BML2526. Interestingly, NR1418 and BML2526 were identified in Osaka, Japan, at different points in time; NR1418 was isolated in 2015, and BML2526 was isolated in 2018, indicating the possibility of the emergence and evolution of IMP-70-producing P. rettgeri. These findings suggest that the plasmid of BML2526 may have occurred following recombination of the two plasmids harbored by NR1418.

In conclusion, we characterized a multidrug-resistant *P. rettgeri* strain carrying multiple β -lactamases, including bla_{IMP-70} , bla_{TEM-1} , bla_{MOX-1} , and $bla_{CTX-M-253}$. These β -lactamases were found on unique plasmids, indicating that they likely evolved through mutations and recombination. Some NDM-1-positive plasmids harbored by *P. rettgeri* likely originated by a cointegration of plasmids, making them easier to disseminate among Enterobacteriaceae (4). These findings suggest that the cointegration of plasmids in *P. rettgeri* might not be unusual and that they may play a role in the transmission of clinically relevant β -lactamases. Although *P. rettgeri* is not a common clinical pathogen, we highlight its potential role in the spread of multidrug resistance and the importance of continued monitoring and surveillance of IMP-producing *P. rettgeri*.

Data availability. The assembled nucleotide sequences of NR1418 have been deposited in GenBank under the accession numbers AP025669–AP025671.

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We have no conflicts of interest to declare.

REFERENCES

- Abdallah M, Balshi A. 2018. First literature review of carbapenem-resistant *Providencia*. New Microbes New Infect 25:16–23. https://doi.org/10.1016/j .nmni.2018.05.009.
- Tshisevhe VS, Lekalakala MR, Tshuma N, Janse van Rensburg S, Mbelle N. 2016. Outbreak of carbapenem-resistant *Providencia rettgeri* in a tertiary hospital. S Afr Med J 107:31–33. https://doi.org/10.7196/SAMJ.2016.v107.i1.12002.
- Marquez-Ortiz RA, Haggerty L, Sim EM, Duarte C, Castro-Cardozo BE, Beltran M, Saavedra S, Vanegas N, Escobar-Perez J, Petty NK. 2017. First complete *Providencia rettgeri* genome sequence, the NDM-1-producing clinical strain RB151. Genome Announc 5:e01472-16. https://doi.org/10.1128/genomeA.01472-16.
- Marquez-Ortiz RA, Haggerty L, Olarte N, Duarte C, Garza-Ramos U, Silva-Sanchez J, Castro BE, Sim EM, Beltran M, Moncada MV, Valderrama A, Castellanos JE, Charles IG, Vanegas N, Escobar-Perez J, Petty NK. 2017. Genomic epidemiology of NDM-1-encoding plasmids in Latin American clinical isolates reveals insights into the evolution of multidrug resistance. Genome Biol Evol 9:1725–1741. https://doi.org/10.1093/gbe/evx115.
- Japan Nosocomial Infections Surveillance. 2020. Public information January– December 2020 annual report. Nosocomial infection control surveillance testing department (inpatient specimens). (in Japanese). https://janis.mhlw.go.jp/ report/open_report/2020/3/1/ken_Open_Report_202000.pdf.
- Yonekawa S, Mizuno T, Nakano R, Nakano A, Suzuki Y, Asada T, Ishii A, Kakuta N, Tsubaki K, Mizuno S, Ogawa M, Yano H, Kasahara K, Mikasa K. 2020. Molecular and epidemiological characteristics of carbapenemase-producing *Klebsiella pneumoniae* clinical isolates in Japan. mSphere 5:e00490-20. https://doi.org/10 .1128/mSphere.00490-20.
- Shiroto K, Ishii Y, Kimura S, Alba J, Watanabe K, Matsushima Y, Yamaguchi K. 2005. Metallo-β-lactamase IMP-1 in *Providencia rettgeri* from two different hospitals in Japan. J Med Microbiol 54:1065–1070. https://doi.org/10.1099/ jmm.0.46194-0.
- Nishio H, Komatsu M, Shibata N, Shimakawa K, Sueyoshi N, Ura T, Satoh K, Toyokawa M, Nakamura T, Wada Y, Orita T, Kofuku T, Yamasaki K, Sakamoto M, Kinoshita S, Aihara M, Arakawa Y. 2004. Metallo-β-lactamase-producing gram-negative bacilli: laboratory-based surveillance in cooperation with 13 clinical laboratories in the Kinki region of Japan. J Clin Microbiol 42:5256–5263. https://doi.org/10.1128/JCM.42.11.5256-5263.2004.
- Iwata S, Tada T, Hishinuma T, Tohya M, Oshiro S, Kuwahara-Arai K, Ogawa M, Shimojima M, Kirikae T. 2020. Emergence of carbapenem-resistant *Providencia rettgeri* and *Providencia stuartii* producing IMP-type metallo-β-lactamase in Japan. Antimicrob Agents Chemother 64:e00382-20. https://doi.org/10.1128/ AAC.00382-20.
- Clinical and Laboratory Standards Institute. 2020. Performance Standards for antimicrobial susceptibility testing; 30th informational supplement. CLSI supplement M100. Clinical and Laboratory Standards Institute, Wayne, PA.
- Saito K, Nakano R, Suzuki Y, Nakano A, Ogawa Y, Yonekawa S, Endo S, Mizuno F, Kasahara K, Mikasa K, Kaku M, Yano H. 2017. Suitability of carbapenem inactivation method (CIM) for detection of IMP metallo-β-lactamase-producing *Enterobacteriaceae*. J Clin Microbiol 55:1220–1222. https://doi.org/10.1128/JCM.02275-16.
- van der Zwaluw K, de Haan A, Pluister GN, Bootsma HJ, de Neeling AJ, Schouls LM. 2015. The carbapenem inactivation method (CIM), a simple and low-cost alternative for the Carba NP test to assess phenotypic carbapenemase activity in gram-negative rods. PLoS One 10:e0123690. https:// doi.org/10.1371/journal.pone.0123690.

- Pierce VM, Simner PJ, Lonsway DR, Roe-Carpenter DE, Johnson JK, Brasso WB, Bobenchik AM, Lockett ZC, Charnot-Katsikas A, Ferraro MJ, Thomson RB, Jr, Jenkins SG, Limbago BM, Das S. 2017. Modified carbapenem inactivation method for phenotypic detection of carbapenemase production among *Enterobacteriaceae*. J Clin Microbiol 55:2321–2333. https://doi.org/10 .1128/JCM.00193-17.
- Dallenne C, Da Costa A, Decré D, Favier C, Arlet G. 2010. Development of a set of multiplex PCR assays for the detection of genes encoding important β-lactamases in Enterobacteriaceae. J Antimicrob Chemother 65: 490–495. https://doi.org/10.1093/jac/dkp498.
- Poirel L, Walsh TR, Cuvillier V, Nordmann P. 2011. Multiplex PCR for detection of acquired carbapenemase genes. Diagn Microbiol Infect Dis 70:119–123. https://doi.org/10.1016/j.diagmicrobio.2010.12.002.
- He D, Chiou J, Zeng Z, Liu L, Chen X, Zeng L, Chan EW, Liu JH, Chen S. 2015. Residues distal to the active site contribute to enhanced catalytic activity of variant and hybrid β-lactamases derived from CTX-M-14 and CTX-M-15. Antimicrob Agents Chemother 59:5976–5983. https://doi.org/10.1128/AAC.04920-14.
- Novais Â, Cantón R, Coque TM, Moya A, Baquero F, Galán JC. 2008. Mutational events in cefotaximase extended-spectrum β-lactamases of the CTX-M-1 cluster involved in ceftazidime resistance. Antimicrob Agents Chemother 52:2377–2382. https://doi.org/10.1128/AAC.01658-07.
- Borgianni L, Vandenameele J, Matagne A, Bini L, Bonomo RA, Frère JM, Rossolini GM, Docquier JD. 2010. Mutational analysis of VIM-2 reveals an essential determinant for metallo-β-lactamase stability and folding. Antimicrob Agents Chemother 54:3197–3204. https://doi.org/10.1128/AAC.01336-09.
- 19. Nakano R, Okamoto R, Nakano Y, Kaneko K, Okitsu N, Hosaka Y, Inoue M. 2004. CFE-1, a novel plasmid-encoded AmpC β -lactamase with an *ampR* gene originating from *Citrobacter freundii*. Antimicrob Agents Chemother 48:1151–1158. https://doi.org/10.1128/AAC.48.4.1151-1158.2004.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. Bioinformatics 34:1037–1039. https://doi.org/10.1093/bioinformatics/btx713.
- Bortolaia V, Kaas RS, Ruppe E, Roberts MC, Schwarz S, Cattoir V, Philippon A, Allesoe RL, Rebelo AR, Florensa AF, Fagelhauer L, Chakraborty T, Neumann B, Werner G, Bender JK, Stingl K, Nguyen M, Coppens J, Xavier BB, Malhotra-Kumar S, Westh H, Pinholt M, Anjum MF, Duggett NA, Kempf I, Nykäsenoja S, Olkkola S, Wieczorek K, Amaro A, Clemente L, Mossong J, Losch S, Ragimbeau C, Lund O, Aarestrup FM. 2020. ResFinder 4.0 for predictions of phenotypes from genotypes. J Antimicrob Chemother 75:3491–3500. https://doi.org/10 .1093/jac/dkaa345.
- Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a web server for prokaryotic species circumscription based on pairwise genome comparison. Bioinformatics 32:929–931. https://doi.org/10.1093/ bioinformatics/btv681.
- Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H. 2014. *In silico* detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. Antimicrob Agents Chemother 58:3895–3903. https://doi.org/10.1128/AAC.02412-14.
- Sullivan MJ, Petty NK, Beatson SA. 2011. Easyfig: a genome comparison visualizer. Bioinformatics 27:1009–1010. https://doi.org/10.1093/bioinformatics/ btr039.