



REVIEW

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Targeting p53 by small molecules in hematological malignancies

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Abstract

p53 is a powerful tumor suppressor and is an attractive cancer therapeutic target. A breakthrough in cancer research came from the discovery of the drugs which are capable of reactivating p53 function. Most anti-cancer agents, from traditional chemo- and radiation therapies to more recently developed non-peptide small molecules exert their effects by enhancing the anti-proliferative activities of p53. Small molecules such as nutlin, RITA, and PRIMA-1 that can activate p53 have shown their anti-tumor effects in different types of hematological malignancies. Importantly, nutlin and PRIMA-1 have successfully reached the stage of phase I/II clinical trials in at least one type of hematological cancer. Thus, the pharmacological activation of p53 by these small molecules has a major clinical impact on prognostic use and targeted drug design. In the current review, we present the recent achievements in p53 research using small molecules in hematological malignancies. Anticancer activity of different classes of compounds targeting the p53 signaling pathway and their mechanism of action are discussed. In addition, we discuss how p53 tumor suppressor protein holds promise as a drug target for recent and future novel therapies in these diseases.

Keywords: Hematological malignancies, Leukemia, Lymphoma, Myeloma, p53, Nutlin, RITA, PRIMA-1, MIRA-1, Apoptosis

Introduction

p53, 'guardian of the genome', was the first tumor suppressor gene to be identified in 1979. p53 functions to eliminate and inhibit the proliferation of abnormal cells, thereby preventing tumor development [1-4]. The human p53 gene is located on chromosome 17p and consists of 11 exons and 10 introns [5]. The central role of p53 in the cells suggests that the loss of p53 function may have severe consequences. The p53 function is lost in an estimated 50% of human cancers by mutations or deletions in p53 gene [6]. The frequency of mutation in p53 is, however, lower in hematological cancers than in solid tumors [7-12]. For example, TP53 is mutated in 10-20% of cases of chronic lymphocytic leukemia (CLL) [7,8], 3-8% of cases of acute myeloid leukemia (AML) [8], less than 3% in acute lymphoblastic leukemia (ALL) [9], and 10-12% cases of multiple myeloma (MM) [10-12]. Importantly, in hematological malignancies, deletion/mutation of p53 is

associated with high risk i.e., more aggressive disease, worse overall survival and resistance to therapies [7-13]. In the presence of wild type p53 other mechanisms may affect the expression and activity of p53 which include elevated expression of the negative regulators of p53, murine double minute 2 (MDM2) [14-20]. MDM2 is transcriptionally activated by the binding of p53 to a p53-responsive element within its gene. It then binds to the N-terminal region of p53, thereby preventing p53 from interacting with the transcriptional machinery and inducing its degradation [15,18,20-23].

There are evidences that many anti-cancer drugs induce apoptosis through multiple pathways that are at least in part dependent upon p53 activation [16-23]. Attempts have been made to develop strategies based on the small molecules to specifically modulate the activity of p53 proteins. These approaches can be classified into two categories: those that aim at modulating the activity of wild-type p53 (Figure 1A) and those that aim at restoring wild-type functions in cells expressing mutant p53 (Figure 1B). The small molecules have been identified by either cellular or protein assays [20-23]. The cellular approach involves screening to identify compounds which can cause tumor cell death. An advantage

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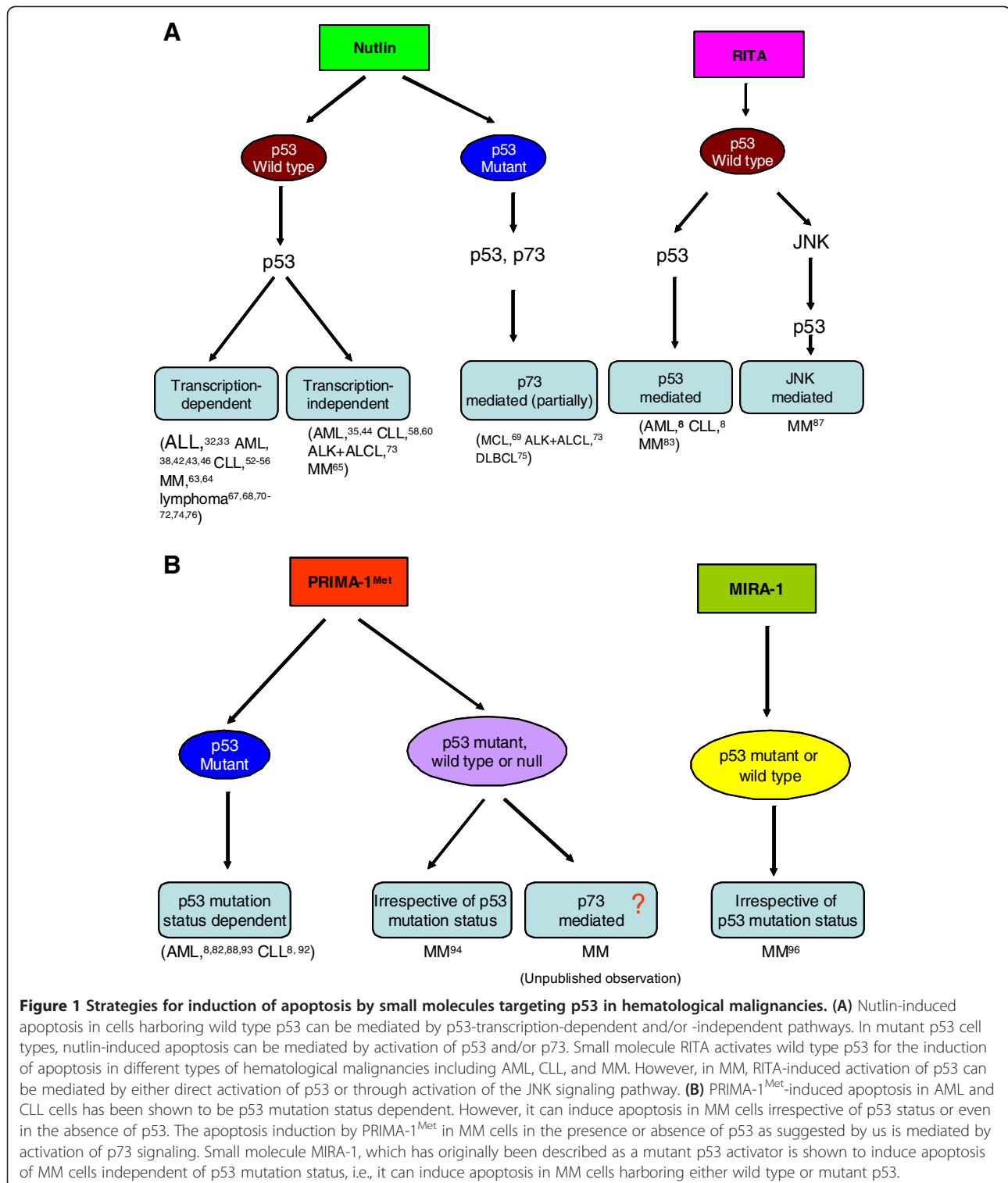


Figure 1 Strategies for induction of apoptosis by small molecules targeting p53 in hematological malignancies. (A) Nutlin-induced apoptosis in cells harboring wild type p53 can be mediated by p53-transcription-dependent and/or -independent pathways. In mutant p53 cell types, nutlin-induced apoptosis can be mediated by activation of p53 and/or p73. Small molecule RITA activates wild type p53 for the induction of apoptosis in different types of hematological malignancies including AML, CLL, and MM. However, in MM, RITA-induced activation of p53 can be mediated by either direct activation of p53 or through activation of the JNK signaling pathway. **(B)** PRIMA-1^{Met}-induced apoptosis in AML and CLL cells has been shown to be p53 mutation status dependent. However, it can induce apoptosis in MM cells irrespective of p53 status or even in the absence of p53. The apoptosis induction by PRIMA-1^{Met} in MM cells in the presence or absence of p53 as suggested by us is mediated by activation of p73 signaling. Small molecule MIRA-1, which has originally been described as a mutant p53 activator is shown to induce apoptosis of MM cells independent of p53 mutation status, i.e., it can induce apoptosis in MM cells harboring either wild type or mutant p53.

of this approach is that the compounds identified e.g., nutlin, RITA (Reactivation of p53 and induction of tumor cell apoptosis) and PRIMA-1 (p53 reactivation and induction of massive apoptosis) have a desired biological outcome such as apoptosis and rarely display

genotoxicity [24-27]. However, it is difficult to elucidate their exact molecular mechanism for apoptosis. On the other hand, a protein based approach can identify compounds e.g., CP-31398 that directly affect a target protein. But the compounds may be toxic or may not have

adequate bioavailability [27-31]. A number of small molecules with activities fitting within these two categories have been identified and some of those have already progressed to advanced preclinical development or early-phase (phase I/II) clinical trials (Table 1). In this review we will describe all of these aspects of targeting p53 in hematological malignancies.

Anti-tumor activities of nutlin in hematological malignancies

Of the small molecules that inhibit the protein-protein interaction between p53 and MDM2, the first reported was nutlins [24]. Nutlin is a nongenotoxic compound which binds to the p53-binding pocket in the MDM2 protein, thus releasing p53 from the negative control of MDM2 leading to effective p53 stabilization and activation in cancer cells with wild type but not mutant or deleted p53 [24,30,31]. Since its discovery, nutlin has been one of the most investigated small molecules in the field of cancer therapy and has shown considerable promise

Table 1 Small molecules used for targeting p53 in various hematological malignancies

Hematological malignancy	Small molecule	Potential target
Leukemia		
ALL	Nutlin	MDM2 ³²⁻³⁴
	RITA	p53 ⁸²
	JNJ-26854165	p53, E2F1 ⁹⁹
AML	*Nutlin	MDM2 ^{35,38,42,43,46}
	MI-219	MDM2 ⁴⁸
	RITA	p53 ⁸
	*PRIMA-1	p53 ^{8,82,88,93}
	JNJ-26854165	p53, E2F1 ⁹⁹
CLL	Nutlin	MDM2 ^{54-56,58-60}
	RITA	p53 ⁸
	PRIMA-1	p53 ^{8,92}
CML	Nutlin	MDM2 ^{61,62}
	MI-219	MDM2 ⁶¹
Lymphoma		
HL	Nutlin	MDM2 ^{67,68}
MCL	Nutlin	MDM2 ⁶⁹⁻⁷²
Birkitt's lymphoma	Nutlin	MDM2 ⁷⁶
B Cell lymphoma	MI-219	MDM2 ⁷⁴
Follicular lymphoma	MI-319	MDM2 ⁷⁸
Myeloma		
	Nutlin	MDM2 ⁶³⁻⁶⁵
	RITA	p53 ⁸⁴ , JNK ⁸⁷
	PRIMA-1 ^{Met}	p53, p73 ⁹⁴
	MIRA-1	p53 ⁹⁶
	Halofuginone	p53 ⁹⁷

*in phase I/II clinical trials.^{91,101}.

in this area. In pre-clinical studies, nutlin alone or in combination with chemotherapeutic drugs has displayed increasing potential for the treatment of blood malignancies [32-79].

Preclinical studies of nutlin in ALL and AML

Nutlin induced cytotoxic and apoptotic response in both ALL [32-34] and AML [35-49] cells including the cell lines and/or patient samples with little effect on normal CD34+ hematopoietic progenitor cells [35]. Nutlin-mediated killing of ALL cells harboring wild type p53 and over-expressing MDM2 is clinically very much significant since all patients with leukemic cells over-expressing MDM2 are usually resistant to conventional therapy and have a poor prognosis [32]. Nutlin-induced apoptosis in AML cells can be mediated by both p53-transcription-dependent and -independent pathways [35] (Figure 1A). Moreover, nutlin displayed synergistic responses in AML cells with several anti-leukemic agents including a Bcl2 antagonist (ABT-737) [36], MEK inhibitors (PD98059) [37] and (AZD6244) [38], recombinant TRAIL [39,40]; FI-700, an FLT3 inhibitor [41], a vitamin D metabolite (1-25D) [42], HDAC inhibitor (valproic acid) [43], PI3K/mTOR inhibitor (PI-103) [44], perifosine (an Akt inhibitor) [49], and sorafenib, a second generation protein kinase inhibitor which is in phase I clinical trial [45].

Cytotoxic response of nutlin in CLL and chronic myeloid leukemia (CML)

Studies have shown that selective p53 activation with nutlin variably induced apoptosis in both low- and high-risk subtypes of B-CLL [50-60] and CML [61,62] patient cells. Notably, nutlin induced cytotoxicity toward B-CLL cells at concentrations that were less toxic toward normal B lymphocytes, peripheral blood mononuclear cells (PBMCs), and bone marrow (BM) hematopoietic progenitor cells [54]. In addition to conventional p53 transcriptional pathway, nutlin also induced apoptosis in CLL cells by mitochondrial pathway (Figure 1A) [58,60].

Anti-myeloma activity of nutlin

We and others have demonstrated potent anti-myeloma activity of nutlin by molecular and functional analysis of the p53 pathway in MM cell lines, primary MM patient samples and in the cells of the bone marrow microenvironment [63-65]. We explored the molecular mechanisms for nutlin-induced apoptosis in MM cells and provided the evidence for association of both p53-transcription-dependent and -independent pathways (Figure 1A) [65]. Our study supports the concept that the transcriptional and mitochondrial functions of p53 are equally important for nutlin-triggered apoptosis, perhaps depending on cancer cell types and their local micro-environments [65,66].

Anti-tumor activity of nutlin in lymphoma and adult T-cell leukemia (ATL)

Among different lymphoma models, nutlin has been shown effective in inducing wild type p53-dependent apoptosis of Hodgkin's lymphoma (HL) [67,68], mantle cell lymphoma (MCL) [69-72], ALK-positive anaplastic large cell lymphoma (ALCL) [73], B-cell lymphoma (BCL) [74,75], Burkitt's and follicular lymphoma [76-78], and adult T cell leukemia [79]. Interestingly, when combined with geldanamycin (an HSP90 inhibitor) nutlin exerted its apoptotic activity in both p53 wild type and mutant HL cells since geldanamycin-induced apoptosis in HL cells was p53-independent [68]. MDM2 inhibition by nutlin successfully induced intrinsic mitochondrial apoptotic activation through increased expression of Noxa in refractory MCL cells, which had limited sensitivity to bortezomib alone. The Nutlin/bortezomib combination enhanced Noxa protein expression in mutant p53 cells but not in wild type p53 MCL cells [71]. Similar to our observations in MM [65], nutlin-induced apoptosis in ALCL cells involved both p53-mediated transcriptional and non-transcriptional mechanisms [71,73]. Recently, by both *in vitro* and *in vivo* evidence Drakos et al. demonstrated that nutlin induced cell cycle arrest and apoptosis in DLBCL cells with functional p53, t(14;18)(q32;q21) translocation, and Bcl2 over-expression [75]. Importantly, combined treatment with nutlin and doxorubicin synergistically inhibited the growth of ALCL or DLBCL cells harboring either wild type or mutant p53 [73,75]. These studies also demonstrated that nutlin induced increased expression of p73 in MCL, ALCL, or BCL cells harboring mutant p53 [72,73,75]. Activation of p53 by nutlin resulted in both cellular senescence and apoptosis in ATL-related cell lines harboring wild type p53 suggesting that cellular senescence might be an important event in p53-dependent cell death in ATL cells [79].

Targeting p53 by RITA

RITA (also known as NSC 652287) was identified through a screening assay based on a library. Upon binding to p53, RITA reactivates it and induces apoptosis by disrupting the interaction with MDM2 [25,30]. Although the IC₅₀ values for RITA vary depending on tumor cell type, growth inhibition is clearly more effective in wild type p53-expressing cells [25,30,80-87].

Anti-leukemic activity of RITA

Among hematological malignancies, anti-tumor activity of RITA was first described in a panel of CLL and AML patient samples [8]. This study described a constitutive activation of the p53 pathway leading to cell cycle arrest and apoptosis by RITA in CLL and AML cells harboring wild type p53 [8]. However, RITA acted synergistically with fludarabine in CLL cells irrespective of p53 status

and with PRIMA-1 in AML cells with or without p53 deletion [8].

Anti-tumor activity of RITA in MM and MCL

Anti-tumor activity of RITA in MM cells was first described by our group in 2010 [83]. Our *in vitro* studies demonstrated that RITA displayed potent anti-myeloma activities in MM cells harboring wild type p53 without killing normal cells [83]. The *in vitro* observation was further confirmed in xenograft mouse model of MM where we have demonstrated significant inhibition of tumor growth and prolongation of survival in mice bearing MM tumors [84,85]. RITA was initially thought to bind with amino terminal domain of p53, inducing a conformational change of the protein and increasing its half life and its accumulation in tumor cells. However, the results of a recent nuclear magnetic resonance (NMR) study indicated that RITA might affect p53 function by other mechanisms, not involving binding to its N-terminal, such as interaction with other binding proteins and cofactors [86]. In keeping with this theory, most recently we provided the evidence that RITA targeted c-Jun N-terminal Kinase (JNK) for the induction of apoptosis in MM cells suggesting that RITA might function as a multi-target molecule [87] (Figure 1A). Further studies are needed to identify the specific binding targets for RITA.

Interestingly, study by Jones et al. provided the evidence that continuous exposure of MCL and MM models to two different MDM2 inhibitors MI-63 and nutlin resulted in p53 point mutations as a mechanism of acquired drug resistance, and that RITA might overcome this resistance by restoring p53 function [81]. This study, therefore, suggests simultaneous restoration of p53 function and MDM2 inhibition as a rational strategy for clinical translation. In support of this, we showed that RITA in combination with nutlin displayed synergistic cytotoxic response in MM cells [83]. The combination of RITA with MI-63 resulted in synergistic response in both MCL and MM cell lines resistant to MI-63 or nutlin [81]. In addition, our studies showed that RITA exerted synergistic response in combination with current chemotherapeutic agents such as doxorubicin or dexamethasone or with the JNK activator 2-Cyano-3,12-dioxooleana-1,9-dien-28 oic Acid (CDDO) [87].

Other small molecules targeting p53-MDM2 interaction

Among the other small molecule MDM2 inhibitors examined in hematological malignancies are MI-63, MI-219, and MI-319 [48,61,69,74,78,81]. MI-63 showed synergistic response with gemcitabine in MCL cells [69]. A most recent study demonstrated that MM or MCL cell lines resistant to either MI-63 or nutlin showed cross-resistant to the other, and were less sensitive to bortezomib, doxorubicin, cisplatin, and melphalan, but not to RITA. Exposure to resistant cells (MM and MCL) to RITA induced cell

cycle arrest and activation of p53-transcriptional targets, supporting a restoration of p53 activity. Combination of RITA and MI-63 showed re-sensitization of resistant MM or MCL cells to MI-63 [81]. Similar to nutlin, MI-219 binds to MDM2 with a high affinity, activated the p53 pathway and selectively inhibited cell growth in cancer cell lines with wild type p53. Anti-tumor activities of MI-219 have been described in AML [48], CML [61], and B-cell lymphoma [74]. Since MI-219/319 achieved an excellent oral bioavailability, it was tested in mouse xenograft models of human follicular lymphoma where MI-319 treatment resulted in inhibition of tumor growth and prolongation of survival [78]. These results suggest that MI-63, MI-219 or MI-319 may be considered as a promising cancer therapy with possible future clinical applications.

Restoration of wild type conformation of mutant p53

Targeting mutant p53 by small molecules appears as an even greater challenge than activating wild type p53 in a tumor cells. Various strategies for reconstitution of wild-type p53 function in tumors have been successfully developed and some have even reached the clinic. Screening approaches have led to the identification of small molecules which can restore p53 function in tumor cells [21-23,26,27]. Here we will focus on two initially reported mutant p53 reactivating drugs (PRIMA-1/PRIMA-1^{Met}, and MIRA-1) and describe their anti-tumor activities in hematological malignancies.

PRIMA-1

PRIMA-1 is a low-molecular weight compound that can restore wild-type conformation of mutant p53 and specific DNA binding, consequently triggering apoptosis in tumour cells carrying mutant p53 [21,26,27,88]. Both unfolded mutant p53 and unfolded wild type p53 can be refolded by PRIMA-1 [89]. The identification of PRIMA-1 as an anti-tumor agent goes back to 2002 in a Swedish Lab where it was found that PRIMA-1 had preferential growth inhibitory activity on different type of human cancer cell lines carrying mutant p53 [21,26,27,90]. This distinguishes PRIMA-1 from anticancer drugs commonly used in treatment of malignant disease [90]. It takes about 10 years for this drug to come into its successful clinical trial in 2012 [91] in a hope to make it a promising anti-cancer drug.

Anti-leukemic activity of PRIMA-1/ PRIMA-1^{Met}

PRIMA-1 was initially tested on 60 human tumor cell lines including lymphoma tumor cell lines [26,90]. A few years later, Nahi et al. reported the effect of PRIMA-1 in leukemic cells from CLL and AML patients with or without p53 deletion [88,92]. There were no obvious differences in cytotoxic response of PRIMA-1 between hemizygous p53 deleted and non-deleted CLL samples

[92]. However, PRIMA-1 was more cytotoxic to AML cells with hemizygous p53 deletion/mutation [88]. Several studies including ours showed that normal hematopoietic cells were relatively resistant to PRIMA-1 in the concentrations used to kill tumor cells [92-94]. PRIMA-1 has been shown to display synergistic or additive response in combination with fludarabine in CLL [92] and AML [93]. The methylated analog of PRIMA-1, PRIMA-1^{Met}, has even greater potency [27], leading to its development as a candidate therapeutic drug under the code name APR246 which is in phase I/II clinical trial [91].

Anti-myeloma activity of PRIMA-1^{Met}

The therapeutic concept is that PRIMA-1^{Met} may selectively rescue mutant p53 and induce apoptosis in cancer cells, leaving wild-type p53 in normal cells mostly unaffected [26]. However, so far there is little information on how PRIMA-1^{Met} affects p53 in cancer cells with no mutation. At first sight, PRIMA-1^{Met} activates wild type p53 appears to contradict claims that the drug is specific for mutant p53 [26,27]. However, there are good biochemical reasons to propose that the drug operates as an all-around rescuer of inactive p53, independent of p53 mutation status. In some cancer cells, p53 protein activity may be disrupted by other mechanisms with functional consequences equivalent to mutation. How such a functionally disrupted p53 may react to PRIMA-1^{Met} is unknown. In the meantime, the recent studies including our preliminary results added a new dimension to the potential of PRIMA-1^{Met} as a therapeutic drug by showing that it induced apoptosis in cells bearing wild type p53 or even in the absence of p53 (Figure 1B) [92-94]. Thus, the therapeutic usage of PRIMA-1^{Met} may be extended for treatment of the vast majority of tumors with a broader spectrum.

Anti-tumor activity of MIRA-1

Using a similar method as described for PRIMA-1, Bykov et al. identified a novel class of molecules that are structurally distinct from PRIMA-1. MIRA-1, the first of these molecules to be tested, induces apoptosis in cells containing mutant p53 with even higher potency than PRIMA-1 [95]. The reactivation of mutant p53 by MIRA-1 has been demonstrated by studies revealing the induction of expression p53-target genes such as p21, MDM2 and Puma in solid tumor cell lines. Therefore MIRA-1 and its structural analogs are postulated to act by shifting the equilibrium between the native and unfolded conformations of p53 toward the native conformation, leading to the restoration of p53-mediated transactivation of target genes and the induction of p53-dependent apoptosis. We have examined anti-tumor activities of MIRA-1 in MM cell lines and patients samples. The results of our preliminary studies showed that anti-myeloma

activity of MIRA-1 was independent of p53 status (Figure 1B) [96].

Other miscellaneous small molecules targeting p53 in hematological malignancies

Recent *in vitro* and *in vivo* studies by Leiba et al. showed that the small molecule halofuginone hydrobromide (HF), a synthetic derivative of quinazoline alkaloid, triggered growth inhibition in both MM cell lines and patient samples. In addition, HF enhanced cytotoxicity of conventional (melphalan, dexamethasone, and doxorubicin) and novel anti-MM (such as lenalidomide) agents [97]. Similar to nutlin, CLL cells with p53 deletion was less sensitive to the small molecule Bcl2 inhibitor ABT-737 than the cells without p53 deletion [98]. JNJ-26854165, a tryptamine derivative has been shown to block the proteasomal degradation of p53 and induce apoptosis in both wild type and mutant p53 expressing leukemia cell lines [99]. Due to its broad anti-tumour activity, JNJ-26854165 is being assessed in a phase I trial as an oral agent for advanced solid tumors [96]. An Aurora kinase inhibitor MLN8237 has been shown effective in killing myeloma cells *in vitro* and *in vivo* [100].

Clinical trials with small molecules targeting p53 in hematological malignancies

The first MDM2 inhibitor that entered clinical development is RG7112 (RO5045337), a member of the nutlin family, from Hoffmann-La Roche (*clinicaltrials.gov*; identifiers: NCT01164033, NCT01143740, NCT00623870, and NCT00559533). RG7112 was given to patients with acute or chronic relapsing or refractory leukemia orally every day for 10 days followed by 18 days of rest. Preliminary clinical data indicated that RG7112 appeared to be well tolerated in patients and showed initial evidence of clinical activity and a mechanism of action consistent with targeting the p53-MDM2 interaction [101].

PRIMA-1^{Met} (APR-246) has recently been tested on humans in a phase I/II study, which was conducted on 22 patients with advanced blood or prostate cancer. The patients received daily infusions of APR-246 for four days. Analysis of the cancer cells taken before and after treatment showed activation of the p53 leading to apoptosis of cancer cells. Ten patients could be evaluated as regards the development of their cancer, and in two of them there were signs of tumor regression [91].

Since tumor cells can acquire resistance to MDM2 inhibitors or current therapeutic agents through p53 mutation it is important to prevent the development of drug resistance and secondary cancer. The combination approaches has been proved successful in this regard as well as to reduce the side-effects of the drugs. A number of studies have demonstrated that nutlin may be used not just as a single agent but also in combination with

other anti-cancer agents to achieve better anti-tumor activity than alone. For example, *ex vivo* experiments using patient tumor samples have shown that nutlin synergizes with doxorubicin, chlorambucil, and fludarabine in B-CLL [30,50,51,55,62]; with doxorubicin and cytosine arabinoside in AML [30,35,36,42]; with doxorubicin and etoposide in HL [67,68]; with melphalan, bortezomib, and lexatumumab (a DR5 agonist) in MM [63,65,102,103]; and with bortezomib in MCL [71], and with doxorubicin in ALCL and DLBCL [73,75]. Moreover, synergistic response of fludarabine with nutlin, RITA or PRIMA-1^{Met} in CLL and/or AML is clinically significant because treatment with fludarabine has been shown to increase the complete remission rate, enhance progression-free survival, and increase the median duration of the clinical response as compared with single therapy [45,56,58]. Importantly, nutlin in combination with bortezomib synergistically contributes to apoptosis induction not only in wild type-p53-possessing MCL cells, but also in MCL with known negative prognostic factors that include p53 mutation, and bortezomib resistance [71]. These findings indicate potential therapeutic efficacy of the small molecules in combination with current chemotherapeutics for the treatment of chemorefractory hematological malignancies.

Conclusions and future directions

Although many of the cellular effects of the described small molecules require the presence of p53, the evidence of some p53-independent effects suggests that p53 may not be its only target. For example, the effect of nutlins initially seemed to be limited to cells harboring wild type p53, subsequent research revealed that nutlin also exerted its anti-cancer activities in p53-negative and p53-mutant human tumor cells through different mechanisms including activation of E2F1 or p73 in different types of cancers including hematological malignancies [48,72,104,105]. Similarly, the biological activity of RITA may be mediated by additional, still unknown, biochemical mechanisms. The global alteration of gene-expression profile rather than merely p53 targets following treatment of PRIMA-1^{Met} suggests other pathways may exist in PRIMA-1^{Met}-induced cell death in MM cells [94]. There is also evidence that PRIMA-1^{Met} can stabilize p53 family members in solid tumors as well as in MM and this may be part of its mechanism of action in mutant p53-expressing tumors [106,107]. Although the exact molecular mechanisms remain unclear, it is possible that PRIMA-1^{Met} can release p73 from an inactive complex with mutant p53 (our unpublished observation). Thus, at least in certain conditions, targeting of p73 or p63 might be an interesting approach to interfere with alternative tumor suppressor pathways [107]. Identifying the potent and selective target(s) for these small molecules will not only be important for understanding the precise

mechanisms of the action of the drugs but also provide the basis for improved drug design to preferentially kill cancer cells with only a limited toxicity towards normal cells.

Competing interests

The authors declare that they have no competing interests.

Authors' contribution

MNS and HC carried out the primary literature search and wrote the paper. LQ participated in the discussion and editing the manuscript. All authors read and approved the final manuscript.

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