

HIV vaccination: turning the spotlight on effector memory T cells as mucosal gatekeepers

Hilde Cheroutre

Address: La Jolla Institute for Allergy and Immunology, 9420 Athena Circle, La Jolla, CA 92037, USA

Email: hilde@liai.org

F1000 Biology Reports 2009, 1:89 (doi:10.3410/B1-89)

This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (<http://creativecommons.org/licenses/by-nc/3.0/legalcode>), which permits unrestricted use, distribution, and reproduction in any medium, for non-commercial purposes provided the original work is properly cited. You may not use this work for commercial purposes.

The electronic version of this article is the complete one and can be found at: <http://F1000.com/Reports/Biology/content/1/89>

Abstract

The accumulating failures in HIV vaccine development demonstrate that the immunization approaches used so far are insufficient to reproduce the naturally occurring immunity that controls the virus in long-term non-progressors, HIV controllers, and continuously exposed sex workers. They also underscore the desperate need for new approaches in the design of more effective vaccination protocols. Recent findings might have brought us closer to that goal by providing proof of concept for a novel preventative HIV vaccine by establishing CD8 effector memory T cells within the mucosal sites of transmission.

Introduction and context

When HIV was first discovered in 1983 [1,2], the race was on to develop an effective vaccine. Twenty-six years later, despite enormous efforts, we have still not reached the finish line. The recent failure of the STEP phase 2b clinical vaccination trial [3,4] using relatively innocuous adenovirus serotype 5 (Ad-5) vectors containing HIV T cell epitopes has caused many to question the ability of cellular immunity to contribute to an effective HIV vaccine.

Retroviruses that cause AIDS in primates, including HIV and its close relative, the simian immunodeficiency virus (SIV), belong to the family of lentiviruses (or 'slow' viruses). They are known for inducing clinical disease only after a prolonged period of relatively stable chronic infection, during which time the immune system slowly but steadily declines. However, the infection kinetics clearly consist of a two-phased process. The initial, acute phase is characterized by an exceedingly high viral replication rate accompanied by a massive loss of memory CCR5⁺CD4 T cells, particularly those that reside within the gastrointestinal tract [5-9]. To a lesser extent, macrophages and some dendritic cells that express CD4 and CCR5 are also affected. This is followed by a chronic phase of variable but usually incomplete virus control, which eventually develops into AIDS [10].

The frequent mutations of the replicating virus which lead to incredible genomic heterogeneity impose an enormous challenge for the design of an effective vaccine. Perhaps more important in this context is the ability of the virus to specifically target immune cells, thereby increasingly breaking down the immune system itself and inevitably leading to the development of AIDS.

Major recent advances

Back to the drawing board

It is not certain why the STEP trial failed, and why it even increased the risk of HIV infection in those individuals with pre-existing Ad-5-specific immunity. One possibility is that the expansion of Ad-5-specific CD4 memory T cells could have created a permissive environment for HIV infection [11]. Nevertheless, two recent studies could not confirm a correlation between the numbers of expanded Ad-5-specific T cells and an increased risk of HIV infection in vaccinees with pre-existing Ad-5 immunity [12,13]. This suggests that other flaws undermined the success of the Ad-5/HIV vaccination strategy and that new concepts need to be considered in order to reach the finish line of successful vaccination.

Several studies have demonstrated that CD8⁺ cytotoxic T cell (CTL) responses play a crucial role in the initial

containment and early control of HIV or SIV replication [10,14-16]. In primary infections, a potent anti-HIV or SIV specific CTL response is detected at a time when a humoral response is weak or nonexistent, suggesting that CD8⁺ effector cells are key controllers of early viremia [15,16]. Nevertheless, SIV-macaque models demonstrated that measuring cellular immune responses to an SIV vaccine, using *in vitro* assays on blood samples, does not faithfully predict the efficacy of the vaccine. Low levels of protection against SIV were observed in the face of rather vigorous vaccine-induced immune responses [17]. Also, live, attenuated SIV infection provided potent protection against wild-type SIV, whereas only weak anti-SIV immune responses were measured *in vitro* [18,19], suggesting that vaccine-induced protection against HIV/SIV might be critically dependent on localized, likely mucosal-specific immune responses *in vivo*.

Consistent with the importance of the local response, and further underscoring the central, early contribution of CD8 T cells, are the results from nonhuman primates demonstrating that depletion of CD8 T cells *in vivo* resulted in enhanced AIDS viral replication during the acute phase, increased loss of CCR5⁺CD45RA⁻CD4 T cells in the intestine, reduced humoral responses and, ultimately, in an accelerated lethal outcome of the disease in the infected monkeys [20-26].

The efficacy of virus-specific CD8 CTLs to control viral replication and/or prevent AIDS led to the concept that an effective vaccine should lead to the induction of a strong and early viral-specific CD8 immune response that, although unable to provide sterilizing protection, ought to be able to control the viral load and delay or prevent the onset of disease, as well as reduce the potential for secondary transmission.

A major step forward: providing proof of concept

Unlike naïve T lymphocytes and central memory T cells (T_{CMs}), effector memory T cells (T_{EMs}) have the ability to reside long-term in non-lymphoid tissues, such as the gut mucosa, and to provide immediate effector function, without the pre-requisite of an initial proliferation step [27]. This suggests that the generation of vaccine-induced mucosal HIV-specific T_{EMs} might provide front-line protection to prevent the initial early phase of HIV infection.

An important recent study by Hansen *et al.* [28] provides direct support for this new paradigm. They demonstrated that Rhesus macaques inoculated with rhesus cytomegalovirus (RhCMV) vectors containing an SIV Gag, Rev-Tat-Nef fusion protein, and Env were protected from progressive wild-type SIV infection after repeated

limiting-dose intrarectal challenges. Moreover, the protected animals that were repeatedly infected via the mucosal route showed no evidence of systemic infection months later, suggesting that the infection was contained locally, well before extensive viral replication or systemic spreading of the virus. Interestingly, and in contrast to the CMV vector-based protection that generated a high frequency of SIV-specific T_{EMs}, an Ad-5-boost vaccine regimen, using the same limiting-dose intrarectal challenge protocol, instead induced a strong T_{CM}-biased immune response that did not provide protection against SIV challenge.

Future directions

Turning the spotlight on T_{EMs} as the mucosal frontline defenders

Earlier SIV/macaque model studies also showed that partial successful SIV protection was reached when immunization was performed via the intranasal or oral route [29,30], further stressing the issue that the induction of mucosal immunity might be key for a successful HIV/AIDS vaccination strategy. Most HIV infections are acquired across a mucosal barrier, and the mucosal immune system forms the main HIV reservoir, the most important target for HIV replication, and it endures the major early T cell losses [5,6,8]. The depletion of mucosal T cell responses is probably the result of direct infection of T cells and immune exhaustion due not only to chronic activation by HIV, but also to activation by intestinal bacteria. The rapid and severe depletion of mucosal CCR5⁺CD4 memory T cells [5,6,8] results in a considerable translocation of bacteria across the mucosal barrier [31]. Especially critical is the loss of Th17 CD4 memory cells, a major contributor to impaired barrier function [32]. Together these processes contribute to the induction of chronic systemic immune activation and T cell exhaustion.

In order to generate protective immunity, future HIV vaccines will probably have to include the induction of viral-specific T_{EM} populations at relevant mucosal entry surfaces as one of the absolute goals. A successful protective immune response will need to be able to sense a low dose of viral epitopes endogenously produced by naturally infected cells, and to eliminate those infected cells before extensive viral replication and the loss of mucosal CD4 memory T cells occurs. This may require specific immunization strategies other than systemic immunization, and the development of more accurate assays to distinguish mucosal HIV-specific T cell responses from those of peripheral cells. But most of all, we need to understand and identify the mechanisms and factors that can direct vaccine-induced differentiation and maintenance of highly effective and long-lived

mucosal T_{EM}s that can contribute to form the first line of defense and destroy the pathogen before it gets a chance to destroy its host.

Abbreviations

Ad-5, adenovirus serotype 5; CCR5, chemokine (C-C motif) receptor 5; CD4, cluster of differentiation 4; CMV, cytomegalovirus; CTL, cytotoxic T cell; RhCMV, rhesus cytomegalovirus; SIV, simian immunodeficiency virus; T_{CM}, central memory T cell; T_{EM}, effector memory T cell.

Competing interests

The author declares that she has no competing interests.

Acknowledgements

I wish to thank M Cheroutre for her contribution and Y Huang for helpful discussions and for providing supportive experimental data. This work was supported by a National Institutes of Health RO1 grant: AI064584 (HC, YH). This is manuscript 1210 from the La Jolla Institute for Allergy and Immunology.

References

- Barre-Sinoussi F, Chermann JC, Rey F, Nugeyre MT, Chamaret S, Gruest J, Dauguet C, Axler-Blin C, Vezinet-Brun F, Rouzioux C, Rozenbaum W, Montagnier L: **Isolation of a T-lymphotropic retrovirus from a patient at risk for acquired immune deficiency syndrome (AIDS)**. *Science* 1983, **220**:868-71.
 - Gallo RC, Sarin PS, Gelmann EP, Robert-Guroff M, Richardson E, Kalyanaraman VS, Mann D, Sidhu GD, Stahl RE, Zolla-Pazner S, Leibowitch J, Popovic M: **Isolation of human T-cell leukemia virus in acquired immune deficiency syndrome (AIDS)**. *Science* 1983, **220**:865-7.
 - Sekaly RP: **The failed HIV Merck vaccine study: a step back or a launching point for future vaccine development?** *J Exp Med* 2008, **205**:7-12.
 - Lu S: **Human versus HIV: round 2 defeat in AIDS vaccine development**. *Expert Rev Vaccines* 2008, **7**:151-3.
 - Veazey RS, DeMaria M, Chalifoux LV, Shvets DE, Pauley DR, Knight HL, Rosenzweig M, Johnson RP, Desrosiers RC, Lackner AA: **Gastrointestinal tract as a major site of CD4+ T cell depletion and viral replication in SIV infection**. *Science* 1998, **280**:427-31.
 - Brenchley JM, Schacker TW, Ruff LE, Price DA, Taylor JH, Beilman GJ, Nguyen PL, Khoruts A, Larson M, Haase AT, Douek DC: **CD4+ T cell depletion during all stages of HIV disease occurs predominantly in the gastrointestinal tract**. *J Exp Med* 2004, **200**:749-59.
- F1000 Factor 6.0 *Must Read*
Evaluated by Sarah Rowland-Jones 21 Dec 2004
- Brenchley JM, Price DA, Douek DC: **HIV disease: fallout from a mucosal catastrophe?** *Nat Immunol* 2006, **7**:235-9.
 - Mehandru S, Poles MA, Tenner-Racz K, Horowitz A, Hurlley A, Hogan C, Boden D, Racz P, Markowitz M: **Primary HIV-1 infection is associated with preferential depletion of CD4+ T lymphocytes from effector sites in the gastrointestinal tract**. *J Exp Med* 2004, **200**:761-70.
 - Li Q, Duan L, Estes JD, Ma ZM, Rourke T, Wang Y, Reilly C, Carlis J, Miller CJ, Haase AT: **Peak SIV replication in resting memory**

CD4+ T cells depletes gut lamina propria CD4+ T cells. *Nature* 2005, **434**:1148-52.

F1000 Factor 6.6 *Must Read*
Evaluated by James Crowe 19 May 2005, Luis J Montaner 19 May 2005, Sarah Rowland-Jones 19 May 2005

- Letvin NL, Walker BD: **Immunopathogenesis and immunotherapy in AIDS virus infections**. *Nat Med* 2003, **9**:861-6.
 - Perreau M, Pantaleo G, Kremer EJ: **Action of a dendritic cell-T cell axis by Ad5 immune complexes creates an improved environment for replication of HIV in T cells**. *J Exp Med* 2008, **205**:2717-25.
- F1000 Factor 3.0 *Recommended*
Evaluated by Lindsey Hutt-Fletcher 06 Nov 2008
- O'Brien KL, Liu J, King SL, Sun YH, Schmitz JE, Lifton MA, Hutnick NA, Betts MR, Dubey SA, Goudsmit J, Shiver JW, Robertson MN, Casimiro DR, Barouch DH: **Adenovirus-specific immunity after immunization with an Ad5 HIV-1 vaccine candidate in humans**. *Nat Med* 2009, **15**:873-5.
- F1000 Factor 6.4 *Must Read*
Evaluated by Thomas Hope 01 Sep 2009, Sheena McCormack 12 Oct 2009
- Hutnick NA, Carnathan DG, Dubey SA, Cox KS, Kierstead L, Ratcliffe SJ, Robertson MN, Casimiro DR, Ertl HC, Betts MR: **Baseline Ad5 serostatus does not predict Ad5 HIV vaccine-induced expansion of adenovirus-specific CD4+ T cells**. *Nat Med* 2009, **15**:876-8.
- F1000 Factor 6.0 *Must Read*
Evaluated by Sheena McCormack 12 Oct 2009
- Belyakov IM, Kuznetsov VA, Kelsall B, Klinman D, Moniuszko M, Lemon M, Markham PD, Pal R, Clements JD, Lewis MG, Strober W, Franchini G, Berzofsky JA: **Impact of vaccine-induced mucosal high-avidity CD8+ CTLs in delay of AIDS viral dissemination from mucosa**. *Blood* 2006, **107**:3258-64.
 - Belyakov IM, Isakov D, Zhu Q, Dzutsev A, Berzofsky JA: **A novel functional CTL avidity/activity compartmentalization to the site of mucosal immunization contributes to protection of macaques against simian/human immunodeficiency viral depletion of mucosal CD4+ T cells**. *J Immunol* 2007, **178**:7211-21.
- F1000 Factor 3.2 *Recommended*
Evaluated by Matthew Dolan 02 Apr 2008, Scott Sieg 28 May 2008
- Daucher M, Price DA, Brenchley JM, Lamoreaux L, Metcalf JA, Rehm C, Nies-Kraske E, Urban E, Yoder C, Rock D, Gumkowski J, Betts MR, Dybul MR, Douek DC: **Virological outcome after structured interruption of antiretroviral therapy for human immunodeficiency virus infection is associated with the functional profile of virus-specific CD8+ T cells**. *J Virol* 2008, **82**:4102-14.
 - Pantaleo G: **HIV-1 T-cell vaccines: evaluating the next step**. *Lancet Infect Dis* 2008, **8**:82-3.
 - Vogel TU, Reynolds MR, Fuller DH, Vielhuber K, Shipley T, Fuller JT, Kunstman KJ, Sutter G, Marthas ML, Erfle V, Wolinsky SM, Wang C, Allison DB, Rud EW, Wilson N, Montefiori D, Altman JD, Watkins DI: **Multispecific vaccine-induced mucosal cytotoxic T lymphocytes reduce acute-phase viral replication but fail in long-term control of simian immunodeficiency virus SIVmac239**. *J Virol* 2003, **77**:13348-60.
 - Casimiro DR, Wang F, Schleif WA, Liang X, Zhang ZQ, Tobery TW, Davies ME, McDermott AB, O'Connor DH, Fridman A, Bagchi A, Tussey LG, Bett AJ, Finnefrock AC, Fu TM, Tang A, Wilson KA, Chen M, Perry HC, Heidecker GJ, Freed DC, Carella A, Punt KS, Sykes KJ, Huang L, Ausensi VI, Bachinsky M, Sadasivan-Nair U, Watkins DI, Emini EA, et al.: **Attenuation of simian immunodeficiency virus SIVmac239 infection by prophylactic immunization with dna and recombinant adenoviral vaccine vectors expressing Gag**. *J Virol* 2005, **79**:15547-55.
 - Matano T, Shibata R, Siemon C, Connors M, Lane HC, Martin MA: **Administration of an anti-CD8 monoclonal antibody interferes with the clearance of chimeric simian/human**

immunodeficiency virus during primary infections of rhesus macaques. *J Virol* 1998, **72:164-9.**

21. Schmitz JE, Kuroda MJ, Santra S, Sasseville VG, Simon MA, Lifton MA, Racz P, Tenner-Racz K, Dalesandro M, Scallon BJ, Ghayeb J, Forman MA, Montefiori DC, Rieber EP, Letvin NL, Reimann KA: **Control of viremia in simian immunodeficiency virus infection by CD8+ lymphocytes.** *Science* 1999, **283**:857-60.
22. Jin X, Bauer DE, Tuttleton SE, Lewin S, Gettie A, Blanchard J, Irwin CE, Safrit JT, Mittler J, Weinberger L, Kostrikis LG, Zhang L, Perelson AS, Ho DD: **Dramatic rise in plasma viremia after CD8(+) T cell depletion in simian immunodeficiency virus-infected macaques.** *J Exp Med* 1999, **189**:991-8.
23. Kawada M, Tsukamoto T, Yamamoto H, Takeda A, Igarashi H, Watkins DI, Matano T: **Long-term control of simian immunodeficiency virus replication with central memory CD4+ T-cell preservation after nonsterile protection by a cytotoxic T-lymphocyte-based vaccine.** *J Virol* 2007, **81**:5202-11.

F1000 Factor 3.0 *Recommended*

Evaluated by James Tartaglia 14 May 2007

24. Amara RR, Ibegbu C, Villinger F, Montefiori DC, Sharma S, Nigam P, Xu Y, McClure HM, Robinson HL: **Studies using a viral challenge and CD8 T cell depletions on the roles of cellular and humoral immunity in the control of an SHIV-89.6P challenge in DNA/MVA-vaccinated macaques.** *Virology* 2005, **343**:246-55.
25. Schmitz JE, Johnson RP, McClure HM, Manson KH, Wyand MS, Kuroda MJ, Lifton MA, Khunkhun RS, McEvers KJ, Gillis J, Piatak M, Lifson JD, Grosschupff G, Racz P, Tenner-Racz K, Rieber EP, Kuus-Reichel K, Gelman RS, Letvin NL, Montefiori DC, Ruprecht RM, Desrosiers RC, Reimann KA: **Effect of CD8+ lymphocyte depletion on virus containment after simian immunodeficiency virus SIVmac251 challenge of live attenuated SIVmac239delta3-vaccinated rhesus macaques.** *J Virol* 2005, **79**:8131-41.
26. Veazey RS, Acierno PM, McEvers KJ, Baumeister SH, Foster GJ, Rett MD, Newberg MH, Kuroda MJ, Williams K, Kim EY, Wolinsky SM, Rieber EP, Piatak M Jr, Lifson JD, Montefiori DC, Brown CR, Hirsch VM, Schmitz JE: **Increased loss of CCR5+ CD45RA- CD4+ T cells in CD8+ lymphocyte-depleted Simian immunodeficiency virus-infected rhesus monkeys.** *J Virol* 2008, **82**:5618-30.

F1000 Factor 3.0 *Recommended*

Evaluated by Leo Stamatatos 23 May 2008

27. Sallusto F, Lenig D, Forster R, Lipp M, Lanzavecchia A: **Two subsets of memory T lymphocytes with distinct homing potentials and effector functions.** *Nature* 1999, **401**:708-12.
28. Hansen SG, Vieville C, Whizin N, Coyne-Johnson L, Siess DC, Drummond DD, Legasse AW, Axthelm MK, Oswald K, Trubey CM, Piatak M Jr, Lifson JD, Nelson JA, Jarvis MA, Picker LJ: **Effector memory T cell responses are associated with protection of rhesus monkeys from mucosal simian immunodeficiency virus challenge.** *Nat Med* 2009, **15**:293-9.

F1000 Factor 4.8 *Must Read*

Evaluated by Lindsey Hutt-Fletcher 01 Apr 2009, Sarah Rowland-Jones 02 Jun 2009

29. Tsukamoto T, Yuasa M, Yamamoto H, Kawada M, Takeda A, Igarashi H, Matano T: **Induction of CD8+ cells able to suppress CCR5-tropic simian immunodeficiency virus SIVmac239 replication by controlled infection of CXCR4-tropic simian-human immunodeficiency virus in vaccinated rhesus macaques.** *J Virol* 2007, **81**:1640-9.
30. Patterson LJ, Malkevitch N, Venzon D, Pinczewski J, Gomez-Roman VR, Wang L, Kalyanaraman VS, Markham PD, Robey FA, Robert-Guroff M: **Protection against mucosal simian immunodeficiency virus SIV(mac251) challenge by using replicating adenovirus-SIV multigene vaccine priming and subunit boosting.** *J Virol* 2004, **78**:2212-21.
31. Li Q, Estes JD, Duan L, Jessurun J, Pambuccian S, Forster C, Wietgreffe S, Zupancic M, Schacker T, Reilly C, Carlis JV, Haase AT: **Simian immunodeficiency virus-induced intestinal cell apoptosis is the underlying mechanism of the regenerative enteropathy of early infection.** *J Infect Dis* 2008, **197**:420-9.
32. Raffatellu M, Santos RL, Verhoeven DE, George MD, Wilson RP, Winter SE, Godinez I, Sankaran S, Paixao TA, Gordon MA, Kolls JK, Dandekar S, Baumler AJ: **Simian immunodeficiency virus-induced mucosal interleukin-17 deficiency promotes Salmonella dissemination from the gut.** *Nat Med* 2008, **14**:421-8.

F1000 Factor 3.0 *Recommended*

Evaluated by Jonathan Heeney 02 Jun 2008

F1000 Factor 3.2 *Recommended*

Evaluated by Matthew Dolan 02 Apr 2008, Scott Sieg 28 May 2008