

## Decreased Renal Accumulation of Biotinylated Chimeric Monoclonal Antibody-Neocarzinostatin Conjugate after Administration of Avidin

Eigo Otsuji,<sup>1</sup> Toshiharu Yamaguchi, Kazuhito Yamamoto, Hiroomi Matsumura, Hiroshi Tsuruta, Yoshihiro Yata, Hiroshi Nishi, Kazuma Okamoto, Kazuya Kitamura and Toshio Takahashi

First Department of Surgery, Kyoto Prefectural University of Medicine, Kawaramachi Hirokoji, Kamigyo-ku, Kyoto 602

Murine monoclonal antibodies (mAbs) such as A7 administered to humans induce a human anti-mouse antibody response. Moreover, because Fab fragments of mAbs are able to penetrate target tumors easily, they may be more suitable than intact mAb to be carriers of anticancer agents such as neocarzinostatin (NCS), which are rapidly inactivated in the blood. To address these problems, chimeric A7 Fab fragment-NCS conjugate (chA7Fab-NCS) was produced. However, large amounts of <sup>125</sup>I-labeled chA7Fab-NCS accumulate in the kidney and can lead to renal dysfunction. To decrease renal accumulation of chA7Fab-NCS, chA7Fab was biotinylated and administered with a subsequent injection of avidin. Human pancreatic carcinoma-bearing nude mice were injected with <sup>125</sup>I-labeled biotinylated chA7Fab-NCS with or without subsequent administration of avidin. The accumulation of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS in tissue samples was measured at appropriate time intervals. <sup>125</sup>I-labeled biotinylated chA7Fab-NCS was cleared more rapidly from the blood and the kidney with the administration of avidin than without it. There was no difference between tumor accumulation in these groups. The tumor/blood ratio of radioactivity of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS was significantly higher with subsequent administration of avidin than without avidin. The administration of biotinylated chA7Fab-NCS followed by avidin may enhance safety and permit the administration of larger doses of NCS without the subsequent development of renal failure. A larger amount of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS was retained in the liver and spleen with the subsequent administration of avidin than without avidin.

Key words: Pancreatic cancer — Chimeric antibody — Biotinylated antibody — Avidin-biotin complex

The development of hybridoma technology offers the possibility of an effective means of delivering chemotherapy to cancer cells. A number of mAbs have been linked to a variety of antitumor drugs and cytotoxins in attempts to increase the effectiveness of chemotherapy.<sup>1,2)</sup> We generated the mAb A7, and covalently conjugated it to the antitumor antibiotic NCS (A7-NCS).<sup>3)</sup> The conjugate A7-NCS has been used clinically to treat patients with colorectal and pancreatic carcinomas.<sup>4)</sup> However, murine mAbs such as A7, when administered to humans, induce a HAMA response<sup>5-7)</sup> that may reduce the tumor localization of the mAb and lead to an anaphylactic reaction. In general, because Fab fragments of mAbs are

able to penetrate target tumors easily, they may be more suitable than intact mAbs as carriers of anticancer agents, such as NCS, which are rapidly inactivated in the blood. To solve these problems, chA7Fab has been produced using recombinant DNA techniques as a new carrier of NCS.<sup>8)</sup> In an experiment using pancreatic cancer-bearing nude mice, <sup>125</sup>I-labeled chA7Fab-NCS accumulated in tumors earlier than <sup>125</sup>I-labeled A7-NCS, and significantly larger amounts of <sup>125</sup>I-labeled chA7Fab-NCS had accumulated in the tumors 1 h after injection, a time when NCS is still active.<sup>9,10)</sup> However, a relatively large amount of <sup>125</sup>I-labeled chA7Fab-NCS accumulated in the kidney, as previously observed with chA7Fab.<sup>10)</sup> Because renal dysfunction is a major side effect in chemotherapy with NCS in humans, renal accumulation of the chA7Fab-NCS conjugate needs to be minimized.

Avidin has a high binding affinity for biotin, a 244 D vitamin found in low concentrations in tissues and in blood.<sup>11)</sup> The bond formation is completed within 15 min, and once formed, is extremely stable.<sup>12)</sup> The strong affinity of the avidin-biotin system has drawn the attention of several researchers working on background reduction in imaging applications of mAbs.<sup>13-15)</sup> The avidin-biotin

<sup>1</sup> To whom requests for reprints should be addressed.

Abbreviations: mAb, monoclonal antibody; NCS, neocarzinostatin; HAMA, human anti-mouse antibody; chA7Fab, Fab fragments of chimeric A7 Fab fragments; chA7Fab-NCS, chimeric A7 Fab fragment-NCS conjugate; FBS, fetal bovine serum; EDTA, ethyldiaminetetraacetic acid; PBS, phosphate buffer solution; PDP, 3-(2-pyridyldithio)propionate; DTT, dithiothreitol; SPDP, N-succinimidyl-3-(2-pyridyldithio)propionate; %ID/g, % injected dose of radioactivity/g; SD, standard deviation.

system provides an extremely rapid increase in the contrast between blood and target tissue. In the present study, the biotinylated chA7Fab-NCS conjugate was administered to nude mice with pancreatic cancer. Thirty min later, a time when NCS is still active, avidin was injected and the kinetics of accumulation of biotinylated chA7Fab-NCS in various tissues were examined. Accumulation with avidin injection was compared to accumulation without avidin injection.

## MATERIALS AND METHODS

**Cell line and tumor xenografts** The pancreatic carcinoma cell line HPC-YS<sup>16)</sup> was established from a ductal cell adenocarcinoma of the human pancreas and was obtained from Dr. N. Yamaguchi (Research Institute of Neurology and Geriatrics, Kyoto Prefectural University of Medicine). Cultured HPC-YS cells were maintained in RPMI 1640 media supplemented with 10% FBS (Flow Laboratories, Inc., Rockville, MD) and harvested after a brief treatment with EDTA, washed in PBS and resuspended in PBS. Approximately  $5 \times 10^6$  viable cells were injected subcutaneously into the left flank of athymic 8-week-old male nude mice (BALB/C, *nu/nu*) (SLC Co., Shizuoka). Tumor masses were detected 7 days after inoculation in all mice injected with HPC-YS cells.

**Preparation of chA7Fab-NCS** ChA7Fab was produced using recombinant DNA techniques as previously described.<sup>10)</sup> The NCS conjugation procedures have been described by Yamaguchi *et al.*<sup>8)</sup> Briefly, chA7Fab and NCS were mixed separately with SPDP, and chA7Fab conjugated to PDP (chA7Fab-PDP) and NCS-PDP were collected. NCS-PDP was thiolated with DTT, then chA7Fab-PDP was combined with NCS-SH, and the mixture was loaded onto a Q-Sepharose column. Elution was conducted using a linear gradient from 0 to 200 mM NaCl in 0.02 M Tris-HCl buffer. The conjugation ratio used was 1 mol of NCS per mol of chA7Fab, i.e., 4.5 mg of chA7Fab was bound to 1 mg of NCS.

**Preparation of radiolabeled chA7Fab-NCS** Radiolabeling of chA7Fab-NCS with <sup>125</sup>I (IMS 30, Amersham Japan, Ltd., Tokyo) was performed using the chloramine-T method.<sup>17)</sup> Iodinated chA7Fab-NCS was separated from excess reactants by gel filtration on a Sephadex G-25 column. ChA7Fab-NCS was labeled with <sup>125</sup>I to a specific activity of 1.0  $\mu$ Ci/ $\mu$ g.

**Biotinylation of <sup>125</sup>I-labeled chA7Fab-NCS** Three hundred micrograms of NHS-Biotin (Pierce 23225, Rockford, IL) in dimethyl sulfoxide was added to 200  $\mu$ g of <sup>125</sup>I-labeled chA7Fab-NCS in 50 mM sodium bicarbonate buffer, pH 8.5. Following incubation on ice for two hours, the preparations were incubated for 30 min at room temperature. Biotinylated <sup>125</sup>I-labeled chA7Fab-NCS was separated from unreacted biotin by gel filtra-

tion on a Sephadex G-25 column. The biotinylation ratio used was 2 mol of biotin per mol of chA7Fab-NCS.

**Binding activity of biotinylated chA7Fab-NCS to HPC-YS cells** The binding activity of biotinylated chA7Fab-NCS was measured by using a competitive radioimmunoassay with HPC-YS cells. Aliquots of HPC-YS cells ( $5 \times 10^5$ ) were incubated with <sup>125</sup>I-labeled chA7Fab-NCS ( $1 \times 10^5$  cpm) in the presence of 4-fold serially diluted chA7Fab-NCS or biotinylated chA7Fab-NCS in PBS at 37°C for 60 min. The concentration of chA7Fab-NCS or biotinylated chA7Fab-NCS ranged from  $(1/4)^9 \times 10^{-5}$  to  $1.0 \times 10^{-5}$  mol/liter. After incubation, the cell pellets were subjected to  $\gamma$ -scintillation counting, and the percent inhibition was calculated as compared to the control.

**Kinetics of <sup>125</sup>I-labeled chA7Fab-NCS accumulation after administration of avidin** The kinetics of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS accumulation after the administration of avidin was investigated in athymic nude mice bearing HPC-YS tumors. Fourteen days after tumor cell inoculation, 32 mice were divided into two groups. Sixteen mice from each group were injected with 0.7  $\mu$ Ci of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS in 100  $\mu$ l of PBS containing chA7Fab-NCS (chA7Fab, 2.25 mg/kg; NCS, 500  $\mu$ g/kg). Each of the mice was injected intravenously with 30  $\mu$ g of streptavidin (Sigma, St. Louis, MO) in 100  $\mu$ l of 50 mM HEPES in 5% mannitol buffer, pH 7.4 or the same solution without streptavidin 30 min after the injection of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS. The mice were killed 0.5, 6, 12, or 24 h after the injection of avidin. The tumors, blood, and normal organs (heart, lung, liver, spleen, pancreas, colon, and kidney) were weighed. The radioactivity in each tissue was then measured using a  $\gamma$ -scintillation counter (Auto-Gamma 5000, Packard, Meriden, CT). The results from the different tissues were expressed as cpm/g and compared with each other. To compare the kinetics of <sup>125</sup>I-labeled biotinylated chA7Fab accumulation in each group, the results were presented as %ID/g. Student's *t* test was used to check for statistically significant differences between groups. To compare the specific localization of the probes in the tumor to that in the blood, the ratio of radioactivity in the tumor to that in the blood was calculated.

In a separate experiment, we examined the kinetics of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS accumulation in the blood, kidney, liver, and spleen at 5 min intervals for 30 min after avidin administration. The results are presented as %ID/g of each tissue.

## RESULTS

**Binding activity of biotinylated chA7Fab-NCS to HPC-YS cells** The binding of biotinylated chA7Fab-NCS to

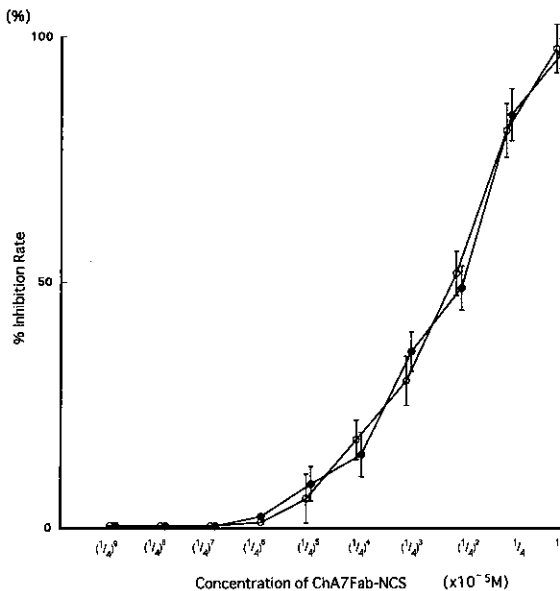


Fig. 1. The binding activity of biotinylated chA7Fab-NCS to HPC-YS cells was compared to that of chA7Fab in a competitive radioimmunoassay. Biotinylated chA7Fab retained a binding activity nearly identical to that of chA7Fab. O, biotinylated chA7Fab-NCS; ●, chA7Fab-NCS; point, mean; bar, SD.

HPC-YS cells was compared to that of chA7Fab-NCS using a competitive radioimmunoassay. The results indicate that biotinylated chA7Fab-NCS retained a binding activity nearly identical to that of chA7Fab-NCS (Fig. 1). **Kinetics of  $^{125}\text{I}$ -labeled chA7Fab-NCS accumulation after administration of avidin** The blood accumulation of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS with and without the administration of avidin decreased linearly with time.  $^{125}\text{I}$ -Labeled biotinylated chA7Fab-NCS was cleared more rapidly from the blood with the administration of avidin than without it, and significant differences were observed at 0.5, 6, and 12 h after the injection of avidin. In contrast, the tumor clearance pattern of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS with the subsequent administration of avidin was similar to that when avidin was not administered. As for normal tissues,  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS was cleared earlier from the kidney with the subsequent administration of avidin than without avidin. Significant differences were observed between the two groups at 0.5, 6, and 12 h after injection ( $P < 0.05$ ). A larger amount of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS was retained in the liver and spleen with the subsequent administration of avidin compared to that without avidin. Significant differences were observed between the two groups at 0.5 and 6 h after injection ( $P < 0.05$ ) (Fig. 2). The tumor tissue/blood ratio of radioac-

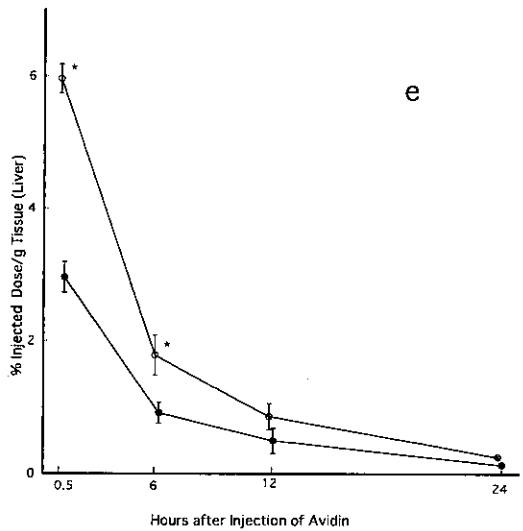
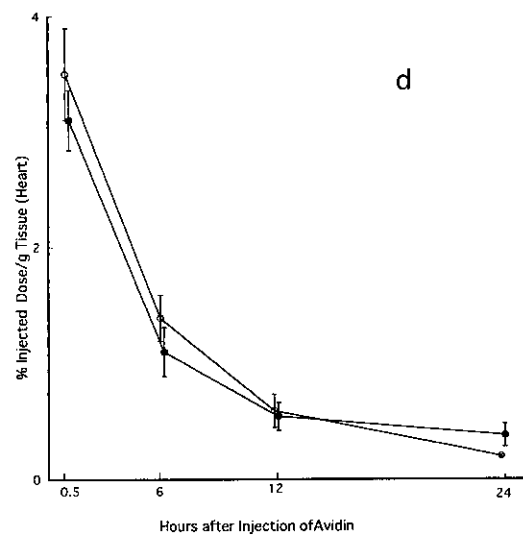
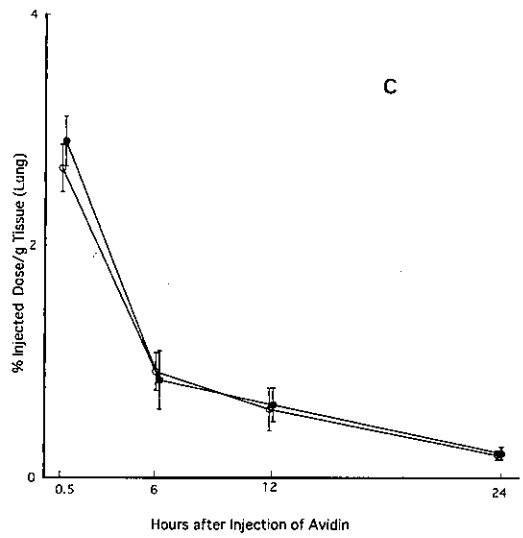
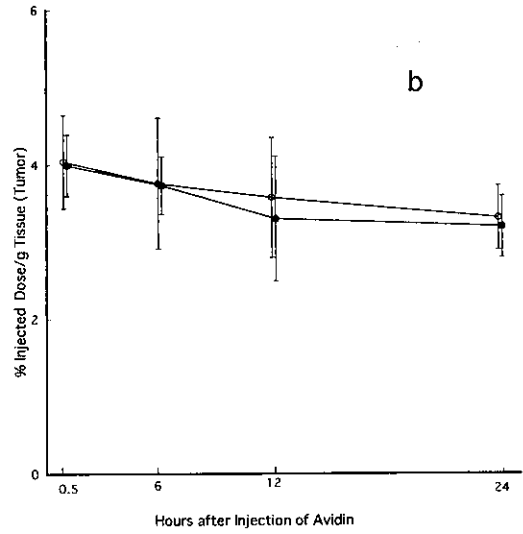
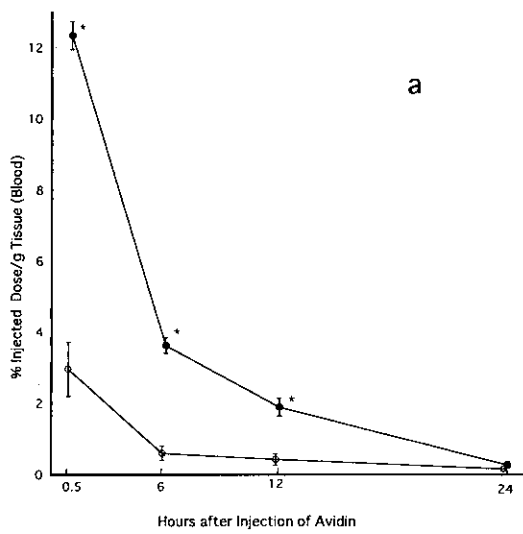
tivity of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS was significantly higher with a subsequent administration of avidin than that without avidin (Fig. 3). In the separate experiment, the concentration of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS in the blood and the kidney decreased rapidly in the first 5 min after avidin administration and that in the spleen and the liver increased rapidly in the first 5 min after avidin administration (Fig. 4).

## DISCUSSION

HAMA production should decrease when NCS conjugated to a human/mouse chimeric mAb is administered to humans because the origin of the Fc portion of the chimeric mAb, which is the most immunopotent region of intact mAbs, is human.<sup>18)</sup> In this study we used chA7Fab-NCS in order to target NCS to human pancreatic carcinoma.

In general, the variable fragment of mAbs has the ability to leave the vascular space rapidly and to penetrate target tumor tissue.<sup>19)</sup> Thus, the Fab fragment of mAb A7 may be able to deliver a large amount of a short-acting anticancer drug such as NCS<sup>20)</sup> directly into the tumor. In a previous study, the antitumor effect of chA7Fab-NCS on the growth of the human pancreatic carcinoma grafted into nude mice was compared with that of A7-NCS, and the results suggested that only chA7Fab-NCS can completely suppress the tumor growth.<sup>21)</sup> However, more  $^{125}\text{I}$ -labeled chA7Fab-NCS than  $^{125}\text{I}$ -labeled A7-NCS accumulated in the kidney<sup>9)</sup> because most of the intravenously injected chA7Fab-NCS was cleared via the kidney. Because renal failure is a serious side effect of NCS therapy, renal accumulation of chA7Fab-NCS is a complication of chemotherapy with this conjugate.

The association constant of the avidin-biotin bond is  $10^{15} \text{ M}^{-1}$ ,<sup>12)</sup> and as such, is  $10^6$ -fold greater than most antigen-antibody interactions. The bond formation is completed within 15 min, and once formed, is extremely stable. The *in vitro* biotinylation of proteins has been well-described,<sup>12)</sup> and any type of immunoglobulin can be easily biotinylated. ChA7Fab-NCS was biotinylated without loss of antibody-antigen binding activity and was injected into nude mice. Biotinylated chA7Fab-NCS bound to avidin forms a large aggregate, because avidin has four binding sites for biotin. As shown in Figs. 2 and 3, the accumulation of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS in the spleen and the liver rapidly increased with the subsequent administration of avidin and a significantly greater amount of biotinylated chA7Fab-NCS was retained in the spleen and the liver in the group of mice injected with avidin compared to those which were not injected with avidin. The avidin-biotinylated mAb-NCS aggregate which formed in the serum were



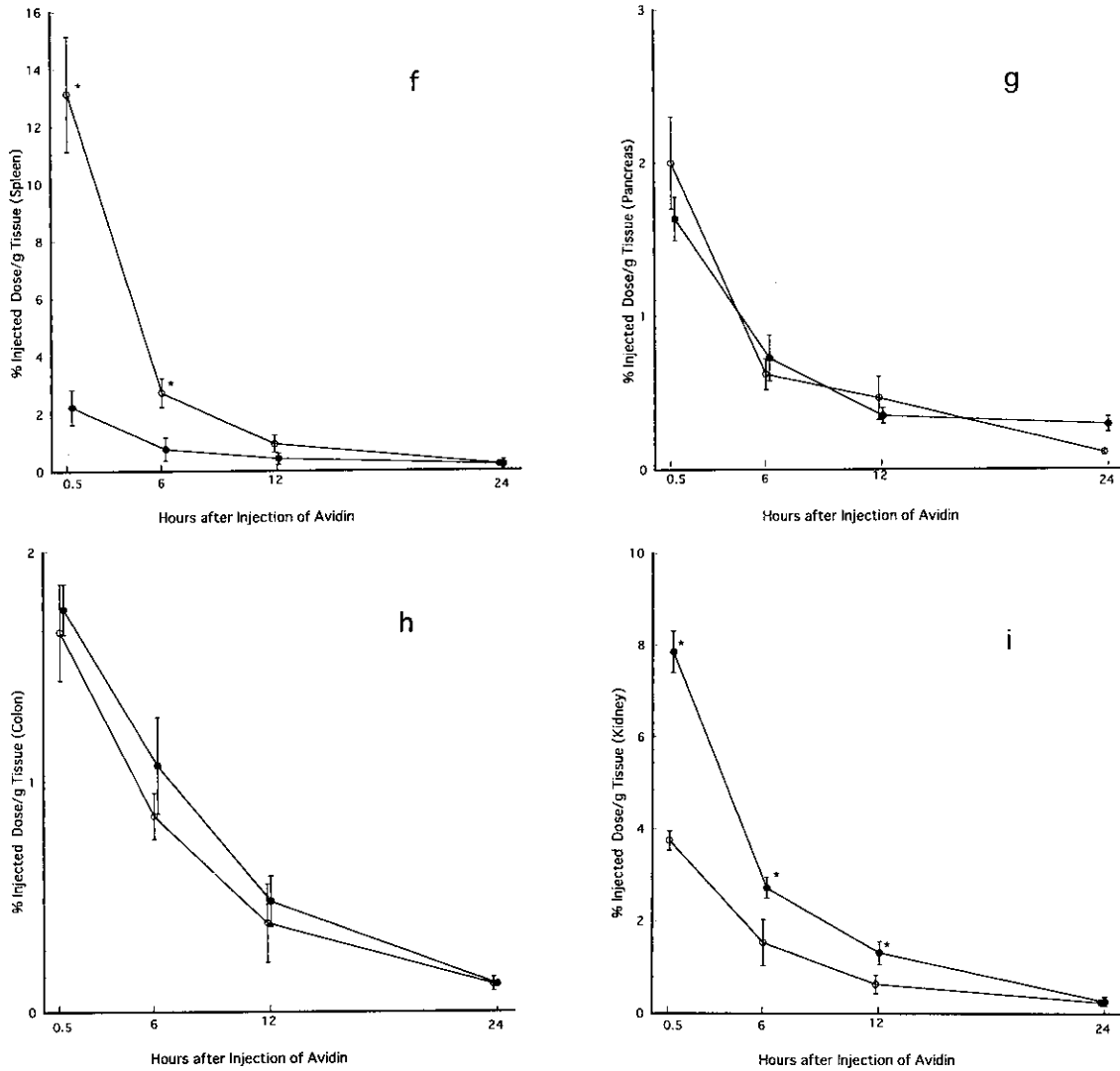


Fig. 2. The accumulation of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS in the blood (a), tumor (b), and normal tissues (c, lung; d, heart; e, liver; f, spleen; g, pancreas; h, colon; i, kidney) of mice with or without the subsequent administration of avidin. The mice received an injection of avidin in  $100\ \mu\text{l}$  of  $50\ \text{mM}$  HEPES in  $5\%$  mannitol buffer or the same solution without avidin 30 min after the injection of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS. The pattern of tumor concentration of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS with the subsequent administration of avidin was similar to that without administration of avidin. The concentrations of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS in the blood and kidney were significantly lower following the administration of avidin than without its administration at 0.5, 6, and 12 h after the injection of avidin. The accumulation of  $^{125}\text{I}$ -labeled biotinylated chA7Fab-NCS in the spleen and the liver was higher following the administration of avidin than it was without the administration of avidin 1 and 6 h after the avidin injection. ○, with the subsequent administration of avidin; ●, no avidin administration; point, mean; bar, SD; \* $P < 0.05$ .

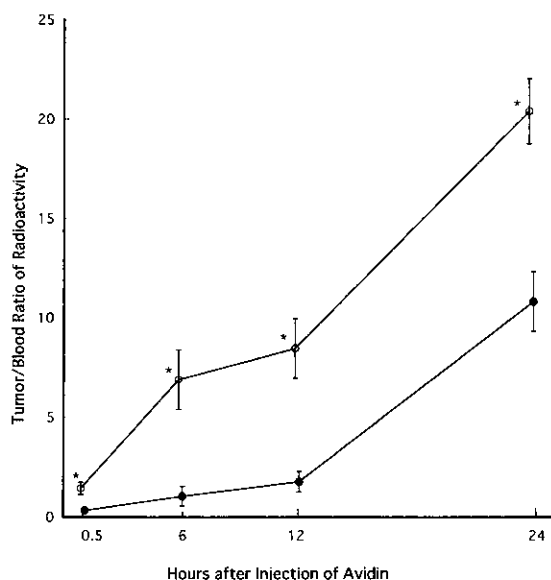


Fig. 3. The tumor tissue/blood ratio of radioactivity of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS with subsequent injection of avidin was compared to that without avidin injection. The tumor tissue/blood ratio of radioactivity of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS was higher with the subsequent administration of avidin than that without avidin ( $P < 0.05$ ). ○, with the subsequent administration of avidin; ●, no avidin administration; point, mean; bar, SD; \* $P < 0.05$ .

thought to be trapped by the reticuloendothelial system. In contrast, the concentration of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS in the blood and the kidney decreased rapidly following the administration of avidin, and a significantly smaller amount of biotinylated chA7Fab-NCS was retained in the blood and kidney in the group of mice injected with avidin compared with those not injected with avidin. These results suggest that avidin administration may potentially decrease renal damage associated with biotinylated chA7Fab-NCS therapy.

Marked accumulation of biotinylated chA7Fab-NCS was seen in the spleen. Marshall *et al.* have reported the decreased splenic accumulation of <sup>125</sup>I-labeled antibody using a galactosylated form of streptavidin in tumor imaging.<sup>22)</sup> However, the liver accumulation of <sup>125</sup>I-labeled antibody increased because of the binding of galactose and asialoglycoprotein receptor of hepatocytes. ChA7Fab-NCS was not galactosylated in the present study to avoid further accumulation of chA7Fab-NCS in the liver. The splenic accumulation of biotinylated chA7Fab conjugated to NCS is unlikely to be toxic to patients, because there have been no reports of adverse effects of NCS on splenic function. Although a relatively

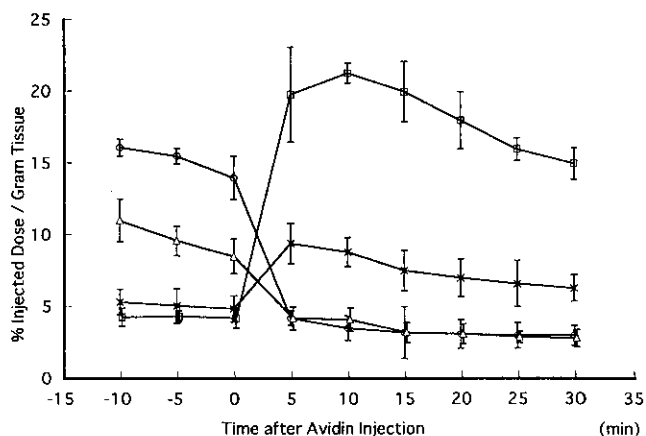


Fig. 4. The kinetics of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS accumulation in the blood, kidney, liver, and spleen were examined at 5 min intervals for 30 min after avidin administration. The concentration of <sup>125</sup>I-labeled biotinylated chA7Fab-NCS in the blood and the kidney decreased rapidly in the first 5 min after avidin administration and that in the spleen and the liver increased rapidly in the first 5 min after avidin administration. ○, blood; △, kidney; ×, liver; □, spleen; point, mean; bar, SD.

large amount of biotinylated chA7Fab-NCS also accumulated in the liver, NCS has been reported to be inactivated rapidly in the liver,<sup>20)</sup> which may minimize its toxicity. Moreover, because biotinylated chA7Fab-NCS is rapidly cleared from the liver and the spleen, the adverse effects may be minimized.

Sung *et al.*<sup>23)</sup> have reported that avidin-biotin complex was formed in the tumor as well as the normal tissues. Saga *et al.*<sup>24)</sup> have also investigated the increased tumor localization of radiolabeled avidin after the injection of biotinylated antibody. Their results suggested that tumor accumulation and the tumor/blood radioactivity ratio of radiolabeled avidin in the pretargeted group were higher than those in the nontargeted group. In the present study, though the tumor/blood ratio of radioactivity was higher with than without avidin administration because the blood <sup>125</sup>I-labeled chA7Fab-NCS concentration decreased, tumor accumulation of <sup>125</sup>I-labeled chA7Fab-NCS was unchanged by administration of avidin. This discrepancy may be explained by the internalization of A7 antigen-antibody complex into the tumor cells and the decrease in the expression of cell-surface antigen due to antigenic modulation.<sup>25)</sup>

Because avidin-biotinylated mAb aggregates were formed in the serum, the immunogenicity of this complex may be enhanced. However, we did not observe any side effects in the animals at the avidin doses used in this

study. Other researchers also have reported that such doses of avidin are not toxic.<sup>13, 15, 26)</sup>

Based on these results, we conclude that biotinylated chA7Fab may be a suitable carrier of NCS and may show decreased renal toxicity when avidin is subsequently administered.

## REFERENCES

- 1) Apelgren, L. D., Zimmerman, D. L., Briggs, S. L. and Bumol, T. F. Antitumor activity of monoclonal antibody-Vinca alkaloid immunoconjugate LY203725 (KS1/4-4-desacetylvinblastine-3-carboxyhydrazide) in a nude mouse model of human ovarian cancer. *Cancer Res.*, **50**, 3540-3544 (1990).
- 2) Hurwitz, E., Levy, R., Maron, R., Wilchek, M., Arnon, R. and Sela, M. The covalent binding of daunomycin and adriamycin to antibodies, with retention of both drug and antibody activities. *Cancer Res.*, **35**, 1175-1181 (1975).
- 3) Fukuda, K. The study of targeting chemotherapy against gastrointestinal cancer. *Akita J. Med.*, **12**, 451-468 (1985).
- 4) Takahashi, T., Yamaguchi, T., Kitamura, K., Suzuyama, H., Honda, M. and Hashimoto, Y. Clinical application of monoclonal antibody-drug conjugates for immunotargeting chemotherapy of colorectal carcinoma. *Cancer*, **61**, 881-888 (1988).
- 5) LoBugli, A. F., Saleh, M., Peterson, L., Wheeler, R., Carrano, R., Huster, W. and Khazaeli, M. B. Phase 1 clinical trial of CO17-1A monoclonal antibody. *Hybridoma*, **5**, S117-S123 (1986).
- 6) Blottiere, H. M., Maurel, C. and Douillard, J. Y. Immune function of patients with gastrointestinal carcinoma after treatment with multiple infusions of monoclonal antibody 17.1A. *Cancer Res.*, **47**, 5238-5241 (1987).
- 7) Takahashi, T., Yamaguchi, T., Kitamura, K., Noguchi, A., Honda, M. and Otsuji, E. Follow-up study of patients treated with monoclonal antibody-drug conjugate: report of 77 cases with colorectal cancer. *Jpn. J. Cancer Res.*, **84**, 976-981 (1993).
- 8) Yamaguchi, T., Tsurumi, H., Kitamura, K., Otsuji, E., Miyagaki, T., Kotani, T. and Takahashi, T. Production, binding and cytotoxicity of human/mouse chimeric monoclonal antibody-neocarzinostatin conjugate. *Jpn. J. Cancer Res.*, **84**, 1190-1194 (1993).
- 9) Otsuji, E., Yamaguchi, T., Yamaoka, N., Taniguchi, K., Kato, M., Kotani, T., Kitamura, K., Takahashi, T. Biodistribution of neocarzinostatin conjugated to chimeric Fab fragments of the monoclonal antibody A7 in nude mice bearing human pancreatic cancer xenografts. *Jpn. J. Cancer Res.*, **85**, 530-535 (1994).
- 10) Otsuji, E., Yamaguchi, T., Yamaoka, N., Kotani, T., Kato, M. and Taniguchi, K. Biodistribution of murine and chimeric Fab fragments of the monoclonal antibody A7 in human pancreatic cancer. *Pancreas*, **10**, 265-273 (1995).
- 11) Mock, D. M. and DuBois, D. B. A sequential, solid-phase assay for biotin in physiologic fluids that correlates with expected biotin status. *Ann. Biochem.*, **153**, 272-278 (1986).
- 12) Green, N. M. Avidin. The use of [<sup>14</sup>C]biotin for kinetic studies and for assay. *Biochem. J.*, **89**, 585-591 (1963).
- 13) Hnatowich, D. J., Virzi, F. and Rusckowski, M. Investigations of avidin and biotin for imaging applications. *J. Nucl. Med.*, **28**, 1294-1302 (1987).
- 14) Sinitsyn, V. V., Mamontova, A. G., Checkneva, Y. Y., Shnyra, A. A. and Domogatsky, S. P. Rapid blood clearance of biotinylated IgG after infusion of avidin. *J. Nucl. Med.*, **30**, 66-69 (1989).
- 15) Ogihara, I., Sasaki, T., Toyama, H., Oda, K., Senda, M. and Nishigori, H. Rapid tumor imaging by active background reduction using biotin-bearing liposomes and avidin. *Cancer Res.*, **54**, 463-467 (1994).
- 16) Otsuji, E., Yamaguchi, Y., Yanamaguchi, N., Koyama, K., Imanishi, J., Yamaoka, N. and Takahashi, T. Expression of the cell surface antigen detected by the monoclonal antibody A7 in pancreatic carcinoma cell lines. *Surg. Today*, **22**, 351-356 (1992).
- 17) Hunter, W. M. and Greenwood, F. C. Preparation of iodine, <sup>131</sup>I-labelled human growth hormone of high specific activity. *Nature*, **194**, 495-496 (1962).
- 18) Spiegelberg, H. L. and Weigle, W. O. The catabolism of homologous and heterologous 7s gamma globulin fragments. *J. Exp. Med.*, **121**, 323-338 (1965).
- 19) Sutherland, R., Buchegger, F., Scheyer, M., Vacca, A. and Mach, J. P. Penetration and binding of radiolabeled anti-carcinoembryonic antigen monoclonal antibodies and their antigen binding fragments in human colon multicellular tumor spheroids. *Cancer Res.*, **47**, 1627-1633 (1987).
- 20) Fujita, H., Nakayama, N., Sawabe, T. and Kimura, K. *In vivo* distribution and inactivation of neocarzinostatin. *Jpn. J. Antibiot.*, **23**, 471-478 (1970).
- 21) Otsuji, E., Yamaguchi, T., Tsuruta, H., Yata, Y., Nishi, H., Okamoto, K., Kitamura, K. and Takahashi, T. Effects of neocarzinostatin-chimeric Fab conjugates on the growth of human pancreatic carcinoma xenografts. *Br. J. Cancer*, **73**, 1178-1182 (1996).
- 22) Marshall, D., Pedley, R. B., Melton, R. G., Boden, J. A., Boden, R. and Begent, R. H. J. Galactosylated streptavidin for improved clearance of biotinylated intact and F(ab')<sub>2</sub> fragments of an anti-tumour antibody. *Br. J.*

## ACKNOWLEDGMENTS

This work was supported in the part by a Grant-in-Aid from the Ministry of Health and Welfare of Japan and a grant from the Pancreatic Research Foundation of Japan.

(Received September 10, 1996/Accepted November 29, 1996)

- Cancer*, 71, 18–24 (1995).
- 23) Sung, C., Osdol, W. V., Saga, T., Neumann, R., Dedrick, R., Weinstein, J. Streptavidin distribution in metastatic tumors pretargeted with a biotinylated monoclonal antibody: theoretical and experimental pharmacokinetics. *Cancer Res.*, 54, 2166–2175 (1994).
- 24) Saga, T., Weinstein, N. W., Jeong, J. M., Heya, T., Lee, J. T., Le, N. and Paik, C. H. Two-step targeting of experimental lung metastasis with biotinylated antibody and radiolabeled streptavidin. *Cancer Res.*, 54, 2160–2165 (1994).
- 25) Yokota, T., Otsuji, E., Noguchi, A., Yamaguchi, T., Sawai, K., Takahashi, T. Antigenic modulation and internalization of monoclonal antibody to human colonic carcinoma cells detected by enzyme-linked immunosorbent assay. *Int. J. Cancer*, 44, 1095–1099 (1989).
- 26) Paganeeli, G., Perves, S., Siccardi, A. G., Rowlinson, G., Deleige, G., Chiolerio, F., Malcovati, M., Seassellati, G. A. and Epenetos, A. A. Intraperitoneal radio-localization of tumors pre-targeted by biotinylated monoclonal antibodies. *Int. J. Cancer*, 45, 1184–1189 (1990).