Contents lists available at ScienceDirect







CrossMark

journal homepage: www.elsevier.com/locate/ynicl

# 

F.Ph.S. Fischmeister <sup>a,b</sup>, I. Höllinger <sup>a,b</sup>, N. Klinger <sup>a,b</sup>, A. Geissler <sup>a,b</sup>, M.C. Wurnig <sup>a,b</sup>, E. Matt <sup>a,b</sup>, J. Rath <sup>a,b</sup>, S.D. Robinson <sup>b,c</sup>, S. Trattnig <sup>b,c</sup>, R. Beisteiner <sup>a,b,\*</sup>

<sup>a</sup> Study Group Clinical fMRI, Department of Neurology, Medical University of Vienna, Austria

<sup>b</sup> High Field MR Center, Medical University of Vienna, Austria

<sup>c</sup> Department of Biomedical Imaging and Image-guided Therapy, Medical University of Vienna, Austria

### ARTICLE INFO

Article history: Received 23 August 2013 Received in revised form 11 September 2013 Accepted 23 September 2013 Available online 30 September 2013

Keywords: Skull-stripping Normalization Lesion Functional MRI Clinical brain mapping Patients

## ABSTRACT

Establishing a reliable correspondence between lesioned brains and a template is challenging using current normalization techniques. The optimum procedure has not been conclusively established, and a critical dichotomy is whether to use input data sets which contain skull signal, or whether skull signal should be removed. Here we provide a first investigation into whether clinical fMRI benefits from skull stripping, based on data from a presurgical language localization task. Brain activation changes related to deskulled/not-deskulled input data are determined in the context of very recently developed (New Segment, Unified Segmentation) and standard normalization approaches. Analysis of structural and functional data demonstrates that skull stripping improves language localization in MNI space — particularly when used in combination with the New Segment normalization technique.

© 2013 The Authors. Published by Elsevier Inc. All rights reserved.

### 1. Introduction

Precise and valid spatial normalization into a common space across all subjects is one of the key components in group analysis of structural and functional neuroimaging data (Brett et al., 2002). In recent years a wealth of algorithms and methods have been developed to account for and correct inter-subject variability in healthy subjects' brains (for a recent review and comparison of algorithms see Klein et al., 2009, 2010). Most normalization methods use automated algorithms to minimize the difference between a subjects' image and a standardized template by applying linear and nonlinear transforms. The establishment of a reliable and robust correspondence between subjects' brains and a template is difficult, however, when there are inherent contrast differences between the two. Disparate B<sub>0</sub> signal dropout, B<sub>1</sub> inhomogeneity and differing tissue contrast can arise from acquisition at different field strengths or from the use of different measurement parameters. The situation becomes particularly problematic in the normalization of lesioned brains, since focal brain lesions or loss of brain tissue resulting from stroke, tumors, or surgery may lead to a lack of correspondence between patient images and standardized templates due to biased normalizations or overfitting (Brett et al., 2001). The impact of such a lack of correspondence in patients' brains to templates on the analysis of functional imaging data has been highlighted in a large body of work (Beisteiner et al., 2010; Crinion et al., 2007; Gartus et al., 2007; Hoeksma et al., 2005; Tahmasebi et al., 2009; Vandenbroucke et al., 2004; Yassa and Stark, 2009). Most clinical studies apply normalization techniques implemented in SPM and, until recently, the SPM standard normalization approach was most popular. However, the Unified Segmentation Model approach (Ashburner and Friston, 2005) constitutes a significant advance in normalization quality. Unified Segmentation attempts to capture all aspects of an anatomical image using a probabilistic framework with tissue prior maps (TPMs) and thus enables tissue classification, bias correction due to signal inhomogeneities, and nonlinear image registration in one model. Crinion et al. (2007) demonstrated that Unified Segmentation produces significantly better and more reliable anatomical co-localization than any of the conventional normalization approaches which employ cost-function masking (CFM) to deal with pathologies (Brett et al., 2001). More recently, Andersen et al. (2010) showed that for larger lesions resulting, for example, from strokes, the benefit of the Unified Segmentation Model can be further increased when used in addition to CFM rather than instead of it. The most recent development is the New Segment toolbox (SPM manual, FIL Group), introduced into SPM as a "work in progress" package. It utilizes the Unified Segmentation algorithm with an improved registration model and an extended set of tissue probability maps.

A critical factor not well investigated is the influence of skullstripping or scalp editing to remove non-brain areas before normalizing brains, although this is relevant to all the normalization techniques. Skull stripping may improve the robustness of the registration process,

 $<sup>\</sup>stackrel{\star}{\sim}$  This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

<sup>\*</sup> Corresponding author at: Study Group Clinical fMRI, Department of Neurology, High Field MR Center, Medical University of Vienna, Währingergürtel 18-20, A-1090 Vienna, Austria.

<sup>2213-1582/\$ –</sup> see front matter © 2013 The Authors. Published by Elsevier Inc. All rights reserved. http://dx.doi.org/10.1016/j.nicl.2013.09.007

since high resolution structural images contain considerable amounts of non-brain tissue such as eyeballs, bone, skin, and other tissues while the template images either do not, or only do to a certain extent. For voxel based morphometry (VBM) Fein et al. (2006) and Acosta-Cabronero et al. (2008) have already demonstrated that misregistrations of individual brains to a common template could be reduced by using brain-extracted images as initial input data sets. Despite these results, no investigations to date have examined the possible benefits of skull stripping as a postprocessing tool for clinical fMRI. Here, we provide the first detailed structural and functional investigation into whether or not skull-stripping (in the context of 3 different normalization approaches) influences the localization of brain function in a cohort of pathological brains which is typical for clinical functional diagnostics.

# 2. Materials and methods

## 2.1. Patients and paradigm

Patients referred for functional localization of language-related areas as part of presurgical evaluation were selected from a pool of data acquired on a 3 Tesla TIM Trio system (Siemens, Erlangen, Germany) according to the following criteria: (1) localization of the tumor, lesion or epileptic focus within the left hemisphere in the vicinity of the Broca or Wernicke area without any previous surgical excision, (2) the patients were right handed and older than 18 years of age, (3) patients were in a good general state of health with no unrelated clinical symptoms and good cooperation at the time of measurement and (4) there was unequivocal left hemispheric language dominance according to the local clinical fMRI report generated on individual non-normalized fMRI data (Foki et al., 2008), which served as functional gold standard in this study.

36 patients (22 male, 14 female, mean age 42.5 years) fulfilling the above criteria were included in this study (see Table 1). These patients and four healthy subjects (2 male, 2 female, mean age 33.75 years) were subdivided into four equally sized groups according to the extent of the lesion (calculated from the lesion mask). These groups were no-lesion (comprising healthy subjects and epileptic patients), small-lesion, medium-lesion and large-lesion (see Fig. 1). These subgroups were formed to assess the effects of lesion size on normalization differences related to skull-stripping.

Participants performed a simple overt language paradigm developed for a comprehensive test of all language components (Foki et al., 2008; Gartus et al., 2009). It consisted of 20 runs, each lasting 140 s. Each run comprised 3 active blocks alternating with 4 rest blocks, with each block lasting for 20 s. During the active phases, two German sentences were presented to the participants visually (for 10 s each). These sentences consisted of 4 words – the stem of a sentence – presented word by word, followed by two verbs displayed one above the other, constituting a correct and an incorrect possible ending of the sentence. The incorrect verbs were either grammatically wrong or semantically unsuitable. While reading the sentence out loud, subjects were explicitly required to choose the word which forms a correct German sentence.

The study was approved by the ethics committee of the Medical University of Vienna. All patients gave written informed consent.

# 2.2. fMRI acquisition

Images were acquired with a 3 Tesla TIM Trio system (Siemens, Erlangen, Germany) using a 32 channel head RF coil and a head fixation helmet (Edward et al., 2000). Functional MRI data were acquired using single-shot gradient-recalled EPI with 34 axial slices ( $1.8 \times 1.8$  mm inplane resolution, 3 mm slice thickness, matrix size of  $128 \times 128$ , a FOV of 230 mm, echo time (TE) 35 ms, repetition time (TR) 2500 ms and GRAPPA acceleration factor 2), aligned to the anterior and posterior commissures. Two dummy/preparation scans were prefaced each run to ensure quasi-equilibrium in longitudinal magnetization. High-

#### Table 1

Patient characteristics including sex, age, diagnosis and lesion size in cm<sup>3</sup>. Note that cases 31 to 36 were classified as controls since clinical evaluation showed no structural or functional finding except for epilepsy.

(cm <sup>3</sup> )       Case 1     Female     45     Low grade glioma, left temporal     44.72       Case 2     Male     43     Tumor of unknown origin, left postcentral     236.29       Case 3     Male     32     Oligodendroglioma, left insular cortex     253.21       Case 4     Male     68     Tumor of unknown origin, left postcentral     65.12       Case 5     Male     44     Astrocytoma grade II, left temporal     564.20       Case 7     Male     50     Glioma grade II, left temporal cortex     201.71       Case 1     Male     38     Astrocytoma grade II, left temporal cortex     195.32       Case 10     Male     38     Astrocytoma grade II, left temporal cortex     102.01       Case 11     Female     30     Cavernous hemangioma, left frontal     2.28       Case 11     Female     31     Cavernous hemangioma, left temporal cortex     102.01       Case 13     Female     37     Oligoastrocytoma grade II, left opercular     271.05       Case 14     Male     53     Low grade glioma, left frontal     2.28	Case number	Sex	Age	Diagnosis	Lesion size
Case 1Fenale45Low grade glioma, left temporal44.72Case 2Male43Tumor of unknown origin, left postcentral236.29Case 4Male32Oligodendroglioma, left insular cortex65.12Case 5Male34Astrocytoma grade II, left fronto-temporal516.69Case 6Male50Gioma grade II, left temporal cortex504.20Case 7Male50Gioma grade II, left temporal cortex201.71Case 7Male51Gioma, left frontal cortex201.71Case 10Male38Astrocytoma grade II, left temporal cortex195.32Case 11Female33Astrocytoma grade II, left temporal cortex102.01Case 13Female54Tumor of unknown origin, left portular cortex102.01Case 14Male55Astrocytoma grade II, left opercular cortex102.01Case 15Female7Oligoastrocytoma grade II, left opercular cortex102.01Case 16Female7Oligoastrocytoma grade II, left opercular cortex144.60Case 17Female7Iuror of unknown origin, left temporal cortex144.60Case 18Male37Low grade glioma, left frontal274.19Case 20Male38Low grade glioma, left frontal274.19Case 21Male52Tumor of unknown origin, left frontal50.2Case 22Male34Coverous hemangioma, left frontal50.2Case 23Male <t< td=""><td></td><td></td><td></td><td></td><td>(cm<sup>3</sup>)</td></t<>					(cm <sup>3</sup> )
Case 2Male43Tumor of unknown origin, left postcentral236.29Case 3Male32Oligodendroglioma, left insular cortex253.21Case 4Male68Tumor of unknown origin, left postcentral516.69Case 5Male44Astrocytoma grade II, left fronton-temporal516.69Case 6Male45Astrocytoma, left frontal-temporal cortex504.90Case 7Male50Glioma grade II, left temporal cortex201.71Case 0Male38Astrocytoma grade I, left frontotemporal24.82Case 10Male38Astrocytoma grade I, left frontotemporal24.82Case 11Female33Astrocytoma grade I, left parietal147.24Case 12Female30Cavernous hemangioma, left frontal22.8Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female37Oligoastrocytoma grade II, left opercular271.08Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female37Low grade glioma, left frontal274.19Case 20Male38Low grade glioma, left frontal274.19Case 21Male52Tumor of unknown origin, left frontoal274.19Case 22Male34Cavernous hemangioma, left frontal274.19Case 23Male21Astrocytoma, left postcentral48.98 <trr<tr>Case 24Male52<td< td=""><td>Case 1</td><td>Female</td><td>45</td><td>Low grade glioma, left temporal</td><td>44.72</td></td<></trr<tr>	Case 1	Female	45	Low grade glioma, left temporal	44.72
Case 3Male32Oligodendroglioma, left insular cortex253.21Case 4Male68Tumor of unknown origin, left postcentral561.2Case 5Male34Astrocytoma grade II, left fronto-temporal564.20Case 7Male50Glioma grade II, left temporal cortex504.90Case 8Female40Astrocytoma, left temporal cortex201.71Case 10Male51Glioma, left frontal cortex195.32Case 11Female33Astrocytoma grade II, left frontotemporal24.82Case 12Female54Tumor of unknown origin, left parietal147.24Case 13Female30Astrocytoma grade II, left opercular cortex102.01Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female37Low grade glioma, left temporal cortex414.60Case 19Male37Low grade glioma, left frontal274.19Case 20Male38Low grade glioma, left frontal74.19Case 21Male52Tumor of unknown origin, left frontal49.58Case 23Male37Low grade glioma, left frontal74.19Case 24Male37Tumor of unknown origin, left fronto-central49.58Case 25Female37 </td <td>Case 2</td> <td>Male</td> <td>43</td> <td>Tumor of unknown origin, left postcentral</td> <td>236.29</td>	Case 2	Male	43	Tumor of unknown origin, left postcentral	236.29
Case 4Male68Tumor of unknown origin, left postcentral65.12Case 5Male34Astrocytoma grade II, left fronto-temporal516.69Case 6Male45Astrocytoma, left frontal-temporal504.20Case 7Male50Glioma grade II, left temporal cortex504.90Case 8Female40Astrocytoma, left temporal cortex201.71Case 10Male38Astrocytoma grade II, left temporal cortex201.71Case 11Female33Astrocytoma grade II, left frontotemporal24.82Case 12Female54Tumor of unknown origin, left parietal147.24Case 13Female50Cavernous hemangioma, left frontal2.28Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Uimor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female49Glioma grade III, left temporal cortex414.60Case 19Male69Tumor of unknown origin, left frontal7.186Case 21Male52Tumor of unknown origin, left frontal274.19Case 23Male21Astrocytoma, left postcentral49.58Case 24Male37Tumor of unknown origin, left fronto-temporal49.58Case 23Male21Astrocytoma, left postcentral49.58Case 24Male	Case 3	Male	32	Oligodendroglioma, left insular cortex	253.21
Case 5Male34Astrocytoma grade II, left fronto-temporal516.69Case 6Male45Astrocytoma, left frontal-temporal564.20Case 7Male50Glioma grade II, left temporal cortex540.22Case 8Female40Astrocytoma, left temporal cortex201.71Case 10Male38Astrocytoma grade II, left temporal cortex195.32Case 11Female33Astrocytoma grade II, left temporal cortex195.32Case 11Female34Astrocytoma grade II, left parietal147.24Case 12Female54Tumor of unknown origin, left parietal147.24Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female39Glioma grade glioma, left temporal cortex146.07Case 20Male38Low grade glioma, left frontal7.186Case 21Male52Tumor of unknown origin, left frontal274.19Case 22Male37Tumor of unknown origin, left frontotemporal49.58Case 23Male37Tumor of unknown origin, left frontotemporal49.58Case 24Male37Tumor of unknown origin, left frontal1.75Case 25Female7Tumor of unknown origin, left frontal1.75Case 26 <td>Case 4</td> <td>Male</td> <td>68</td> <td>Tumor of unknown origin, left postcentral</td> <td>65.12</td>	Case 4	Male	68	Tumor of unknown origin, left postcentral	65.12
Case 6Male45Astrocytoma, left frontal-temporal564.20Case 7Male50Glioma grade II, left temporal cortex504.90Case 8Female40Astrocytoma, left temporal cortex201.71Case 9Male51Glioma, left frontal cortex201.71Case 10Male38Astrocytoma grade II, left temporal cortex195.32Case 11Female33Astrocytoma grade II, left frontotemporal24.82Case 12Female54Tumor of unknown origin, left parietal147.24Case 13Female65Astrocytoma grade II, left opercular cortex102.01Case 14Male65Astrocytoma grade II, left opercular cortex173.55Case 16Female27Tumor of unknown origin, left temporal cortex414.60Case 17Female49Glioma grade II, left temporal cortex414.60Case 18Male37Low grade glioma, left frontal274.19Case 20Male54Tumor of unknown origin, left frontal274.19Case 21Male52Tumor of unknown origin, left frontal274.19Case 22Male37Tumor of unknown origin, left frontotemporal405.58Case 25Female60Tumor of unknown origin, left frontotemporal405.58Case 26Male45Tumor of unknown origin, left frontotemporal405.58Case 27Female50Tumor of unknown origin, left frontot-etemporal405.58Ca	Case 5	Male	34	Astrocytoma grade II, left fronto-temporal	516.69
Case 7Male50Glioma grade II, left temporal cortex504.90Case 8Female40Astrocytoma, left temporoparietal540.22Case 9Male31Glioma, left frontal cortex201.71Case 10Male38Astrocytoma grade II, left fromtotemporal24.82Case 11Female33Astrocytoma grade II, left frontotemporal24.82Case 12Female54Tumor of unknown origin, left parietal147.24Case 13Female65Astrocytoma grade II, left opercular cortex102.01Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left temporoparietal46.73Case 17Female37Low grade glioma, left temporal cortex414.60Case 19Male69Tumor of unknown origin, left frontal7.186Case 21Male52Tumor of unknown origin, left frontal274.19Case 22Male34Cavernous hemangioma, left fronto-central48.98Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left fronto-central148.98Case 25Female60Tumor of unknown origin, left fronto-central148.98Case 25Female61Tumor of unknown origin, left frontal1.1.75Case 2	Case 6	Male	45	Astrocytoma, left frontal-temporal	564.20
Case 8Female40Astrocytoma, left temporoparietal540.22Case 9Male51Glioma, left frontal cortex201.71Case 10Male38Astrocytoma grade II, left temporal cortex195.32Case 11Female54Tumor of unknown origin, left parietal147.24Case 12Female54Tumor of unknown origin, left parietal147.24Case 13Female50Cavernous hemangioma, left frontal2.28Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female37Low grade glioma, left temporal cortex414.60Case 18Male37Low grade glioma, left frontal2.09.93Case 20Male38Low grade glioma, left frontal274.19Case 23Male52Tumor of unknown origin, left frontotal274.19Case 24Male52Tumor of unknown origin, left frontotal49.58Case 25Female60Tumor of unknown origin, left fronto-central148.98Case 26Male45Tumor of unknown origin, left fronto-temporal49.58Case 27Female75Tumor of unknown origin, left fronto-temporal400.59Case 28Male45Tumor of unknown origin, left fronto-temporal49.58Case 29<	Case 7	Male	50	Glioma grade II, left temporal cortex	504.90
Case 9Male51Glioma, left frontal cortex201.71Case 10Male38Astrocytoma grade II, left temporal cortex195.32Case 11Female33Astrocytoma grade II, left frontotemporal24.82Case 12Female54Tumor of unknown origin, left parietal147.24Case 13Female50Cavernous hemangioma, left frontal2.28Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female49Glioma grade III, left temporal cortex414.60Case 18Male57Tumor of unknown origin, left temporo-parietal209.93case 20Male38Low grade glioma, left frontal7.186Case 21Male52Tumor of unknown origin, left frontal274.19Case 23Male51Astrocytoma, left postcentral5.02Case 24Male57Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left frontal11.75Case 26Male45Tumor of unknown origin, left fronto-central148.98Case 27Female75Tumor of unknown origin, left frontal11.75Case 28Male45Tumor of unknown origin, left frontal11.75Case 29Femal	Case 8	Female	40	Astrocytoma, left temporoparietal	540.22
Case 10Male38Astrocytoma grade II, left temporal cortex195.32Case 11Female33Astrocytoma grade II, left frontotemporal24.82Case 12Female54Tumor of unknown origin, left parietal147.24Case 13Female30Cavernous hemangioma, left frontal2.28Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female49Glioma grade III, left temporaparietal46.73Case 18Male37Low grade glioma, left temporal cortex414.60Case 21Male52Tumor of unknown origin, left frontal274.19Case 22Male38Cavernous hemangioma, left frontal274.19Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left frontot-central148.98Case 26Male45Tumor of unknown origin, left frontot-central148.98Case 27Female75Tumor of unknown origin, left frontot-central400.59Case 28Male45Tumor of unknown origin, left frontot-central148.98Case 29Female75Tumor of unknown origin, left frontot-central	Case 9	Male	51	Glioma, left frontal cortex	201.71
Case 11Female33Astrocytoma grade II, left frontotemporal24.82Case 12Female54Tumor of unknown origin, left parietal147.24Case 13Female30Cavernous hemangioma, left frontal2.28Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female49Glioma grade III, left temporoparietal46.73Case 18Male37Low grade glioma, left frontal209.93Case 20Male69Tumor of unknown origin, left temporoparietal274.19Case 21Male52Tumor of unknown origin, left frontal274.19Case 23Male34Cavernous hemangioma, left frontal274.19Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left frontot-central14.89Case 24Male37Tumor of unknown origin, left frontal11.75Case 25Female60Tumor of unknown origin, left frontot-central14.89Case 26Male35Tumor of unknown origin, left frontal11.75Case 27Female75Tumor of unknown origin, left frontal11.75Case 28Male45Tumor of unknown origin, left frontot-central14.89<	Case 10	Male	38	Astrocytoma grade II, left temporal cortex	195.32
Case 12Female54Tumor of unknown origin, left parietal147.24Case 13Female30Cavernous hemangioma, left frontal2.28Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female49Glioma grade III, left temporoparietal46.73Case 18Male37Low grade glioma, left temporoparietal209.93Case 19Male69Tumor of unknown origin, left frontal274.19Case 20Male38Low grade glioma, left frontal274.19Case 21Male52Tumor of unknown origin, left frontotemporal49.58Case 23Male31Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontot-central148.98Case 25Female60Tumor of unknown origin, left frontot-central148.98Case 26Male45Tumor of unknown origin, left frontot-central148.98Case 27Female55Tumor of unknown origin, left frontot-central400.59Case 28Male45Tumor of unknown origin, left frontot-central400.59Case 29Female55Tumor of unknown origin, left frontot-central51.56Case 31Female19Temporal lobe epilepsy left-	Case 11	Female	33	Astrocytoma grade II, left frontotemporal	24.82
Case 13Female30Cavernous hemangioma, left frontal2.28Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female49Glioma grade III, left temporoparietal46.73Case 18Male37Low grade glioma, left temporoparietal209.93Case 20Male38Low grade glioma, left frontal274.19Case 21Male52Tumor of unknown origin, left frontal274.19Case 23Male54Cavernous hemangioma, left basal ganglia36.74Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left fronto-central148.98Case 25Female75Tumor of unknown origin, left fronto-temporal400.59Case 26Male45Tumor of unknown origin, left fronto-temporal400.59Case 27Female75Tumor of unknown origin, left fronto-temporal400.59Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female19Temporal lobe epilepsy left-Case 31Female19Temporal lobe epilepsy left-Case 33Male20Temporal lobe epilepsy left-Case 34Female<	Case 12	Female	54	Tumor of unknown origin, left parietal	147.24
Case 14Male65Astrocytoma grade II, left opercular cortex102.01Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female49Glioma grade III, left temporarietal46.73Case 18Male37Low grade glioma, left temporal cortex414.60Case 19Male69Tumor of unknown origin, left temporo-parietal209.93case 20Male52Tumor of unknown origin, left frontal274.19Case 23Male52Tumor of unknown origin, left frontal5.02Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left frontot-central148.98Case 26Male45Tumor of unknown origin, left frontotal11.75Case 27Female75Tumor of unknown origin, left frontal205.98Case 28Male45Tumor of unknown origin, left frontal6.15Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 33Male20Temporal lobe epilepsy left-Case 34Female49Temporal lobe epilepsy left-Case 35Male35Healthy partici	Case 13	Female	30	Cavernous hemangioma, left frontal	2.28
Case 15Female27Tumor of unknown origin, left temporal cortex173.55Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female49Glioma grade II, left temporoparietal46.73Case 18Male37Low grade glioma, left temporoparietal209.93Case 19Male69Tumor of unknown origin, left temporoparietal209.93Case 20Male38Low grade glioma, left frontal7.186Case 21Male52Tumor of unknown origin, left frontal274.19Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left frontot-central148.98Case 26Male37Tumor of unknown origin, left frontot-central148.98Case 27Female75Tumor of unknown origin, left fronto-temporal400.59Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left fronto-temporal400.59Case 31Male31Low grade glioma, left insular cortex96.15Case 32Male31Low grade glioma, left insular cortex96.15Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male	Case 14	Male	65	Astrocytoma grade II, left opercular cortex	102.01
Case 16Female37Oligoastrocytoma grade II, left opercular271.08Case 17Female49Glioma grade III, left temporoparietal46.73Case 18Male37Low grade glioma, left temporal cortex414.60Case 19Male69Tumor of unknown origin, left temporo-parietal209.93Case 20Male38Low grade glioma, left frontal7.186Case 21Male52Tumor of unknown origin, left frontal274.19Case 22Male34Cavernous hemangioma, left basal ganglia36.74Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left frontot-central11.75Case 26Male45Tumor of unknown origin, left frontot-central148.98Case 27Female75Tumor of unknown origin, left frontot-central400.59Case 28Male45Tumor of unknown origin, left frontot-temporal400.59Case 29Female55Tumor of unknown origin, left precentral51.56Case 31Female19Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male21Temporal lobe epilepsy left-Case 34Female43Temporal lobe ep	Case 15	Female	27	Tumor of unknown origin, left temporal cortex	173.55
Case 17Female49Glioma grade III, left temporoparietal46.73Case 18Male37Low grade glioma, left temporal cortex414.60Case 19Male69Tumor of unknown origin, left temporo-parietal209.93cortexcortexcortex274.19Case 20Male52Tumor of unknown origin, left frontal274.19Case 21Male52Tumor of unknown origin, left frontal5.02Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left frontot-central148.98Case 26Male45Tumor of unknown origin, left frontot-central148.98Case 27Female75Tumor of unknown origin, left frontot-cemporal400.59Case 28Male45Tumor of unknown origin, left frontot-temporal40.59Case 29Female55Tumor of unknown origin, left proto-temporal400.59Case 30Male31Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male32Temporal lobe epilepsy left-Case 35Male32Temporal lobe epilepsy left-C	Case 16	Female	37	Oligoastrocytoma grade II, left opercular	271.08
Case 18Male37Low grade glioma, left temporal cortex414.60Case 19Male69Tumor of unknown origin, left temporo-parietal cortex209.93Case 20Male38Low grade glioma, left frontal7.186Case 21Male52Tumor of unknown origin, left frontal7.419Case 22Male34Cavernous hemangioma, left basal ganglia36.74Case 23Male37Tumor of unknown origin, left frontotemporal49.58Case 24Male37Tumor of unknown origin, left fronto-central148.98Case 25Female60Tumor of unknown origin, left frontoto-central148.98Case 26Male45Tumor of unknown origin, left frontot-central11.75Case 27Female75Tumor of unknown origin, left fronto-temporal400.59Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left fronto-temporal400.59Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male32Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male32Tempora	Case 17	Female	49	Glioma grade III, left temporoparietal	46.73
Case 19Male69Tumor of unknown origin, left temporo-parietal209.93 cortexCase 20Male38Low grade glioma, left frontal7.186Case 21Male52Tumor of unknown origin, left frontal274.19Case 22Male34Cavernous hemangioma, left basal ganglia36.74Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left fronto-cemtral148.98Case 25Female60Tumor of unknown origin, left fronto-cemtral148.98Case 26Male45Tumor of unknown origin, left fronto-cemtral206.98Case 27Female75Tumor of unknown origin, left fronto-temporal400.59Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left fronto-temporal51.56Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 33Male20Temporal lobe epilepsy left-Case 34Female41Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male32Temporal lobe epilepsy left-Case 38Female43Healthy participant-	Case 18	Male	37	Low grade glioma, left temporal cortex	414.60
cortexCase 20Male38Low grade glioma, left frontal7.186Case 21Male52Tumor of unknown origin, left frontal274.19Case 22Male34Cavernous hemangioma, left basal ganglia36.74Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left fronto-central148.98Case 26Male45Tumor of unknown origin, left fronto-temporal206.98Case 27Female75Tumor of unknown origin, left fronto-temporal400.59Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left prote-temporal61.56Case 29Female55Tumor of unknown origin, left protecntral51.56Case 30Male31Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 32Male20Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant </td <td>Case 19</td> <td>Male</td> <td>69</td> <td>Tumor of unknown origin, left temporo-parietal</td> <td>209.93</td>	Case 19	Male	69	Tumor of unknown origin, left temporo-parietal	209.93
Case 20Male38Low grade glioma, left frontal7.186Case 21Male52Tumor of unknown origin, left frontal274.19Case 22Male34Cavernous hemangioma, left basal ganglia36.74Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontomportal49.58Case 25Female60Tumor of unknown origin, left fronto-central148.98Case 25Female75Tumor of unknown origin, left fronto-temporal296.98Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left precentral51.56Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 32Male20Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female43Temporal lobe epilepsy left-Case 35Male35Healthy participant-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female43Healthy participant-Case 36Female30Healt				cortex	
Case 21Male52Tumor of unknown origin, left frontal274.19Case 22Male34Cavernous hemangioma, left basal ganglia36.74Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left fronto-central11.75Case 26Male45Tumor of unknown origin, left fronto-central296.98Case 27Female75Tumor of unknown origin, left fronto-temporal400.59Case 28Male45Tumor of unknown origin, left precentral51.56Case 29Female55Tumor of unknown origin, left precentral51.56Case 30Male31Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 32Male20Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male32Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male32Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male32Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35<	Case 20	Male	38	Low grade glioma, left frontal	7.186
Case 22Male34Cavernous hemangioma, left basal ganglia36.74Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left frontot-central148.98Case 26Male45Tumor of unknown origin, left frontot-central148.98Case 27Female75Tumor of unknown origin, left temporal cortex296.98Case 28Male45Tumor of unknown origin, left temporal cortex400.59Case 29Female55Tumor of unknown origin, left precentral51.56Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 33Male20Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male32Temporal lobe epilepsy left-Case 38Female43Healthy participant-Case 39Female30Healthy participant-Case 39Female30Healthy participant-Case 30Male27Healthy participant-	Case 21	Male	52	Tumor of unknown origin, left frontal	274.19
Case 23Male21Astrocytoma, left postcentral5.02Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left fronto-central148.98Case 26Male45Tumor of unknown origin, left frontol11.75Case 27Female75Tumor of unknown origin, left fronto-central10.59Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left precentral51.56Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male32Temporal lobe epilepsy left-Case 36Female43Temporal lobe epilepsy left-Case 36Female43Temporal lobe epilepsy left-Case 37Male32Temporal lobe epilepsy left-Case 38Female43Healthy participant-Case 39Female43Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 22	Male	34	Cavernous hemangioma, left basal ganglia	36.74
Case 24Male37Tumor of unknown origin, left frontotemporal49.58Case 25Female60Tumor of unknown origin, left fronto-central148.98Case 26Male45Tumor of unknown origin, left frontotal11.75Case 27Female75Tumor of unknown origin, left fronto-temporal296.98Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left fronto-temporal400.59Case 23Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 33Male20Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female49Temporal lobe epilepsy left-Case 37Male32Temporal lobe epilepsy left-Case 36Female43Healthy participant-Case 37Male35Healthy participant-Case 39Female43Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 23	Male	21	Astrocytoma, left postcentral	5.02
Case 25Female60Tumor of unknown origin, left fronto-central148.98Case 26Male45Tumor of unknown origin, left frontal11.75Case 27Female75Tumor of unknown origin, left temporal cortex296.98Case 28Male45Tumor of unknown origin, left temporal cortex400.59Case 29Female55Tumor of unknown origin, left proto-temporal400.59Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 33Male20Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female33Healthy participant-Case 39Female30Healthy participant-Case 30Male27Healthy participant-	Case 24	Male	37	Tumor of unknown origin, left frontotemporal	49.58
Case 26Male45Tumor of unknown origin, left frontal11.75Case 27Female75Tumor of unknown origin, left temporal cortex296.98Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left protecntral51.56Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 33Male20Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female30Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 25	Female	60	Tumor of unknown origin, left fronto-central	148.98
Case 27Female75Tumor of unknown origin, left temporal cortex296.98Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left precentral51.56Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 32Male20Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female30Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 26	Male	45	Tumor of unknown origin, left frontal	11.75
Case 28Male45Tumor of unknown origin, left fronto-temporal400.59Case 29Female55Tumor of unknown origin, left precentral51.56Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 33Male20Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male21Temporal lobe epilepsy left-Case 36Female42Temporal lobe epilepsy left-Case 37Male32Temporal lobe epilepsy left-Case 38Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female30Healthy participant-Case 40Male27Healthy participant-	Case 27	Female	75	Tumor of unknown origin, left temporal cortex	296.98
Case 29Female55Tumor of unknown origin, left precentral51.56Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 32Male20Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female43Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 28	Male	45	Tumor of unknown origin, left fronto-temporal	400.59
Case 30Male33Low grade glioma, left insular cortex96.15Case 31Female19Temporal lobe epilepsy left-Case 32Male20Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female30Healthy participant-Case 40Male27Healthy participant-	Case 29	Female	55	Tumor of unknown origin, left precentral	51.56
Case 31Female19Temporal lobe epilepsy left-Case 32Male20Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female43Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 30	Male	33	Low grade glioma, left insular cortex	96.15
Case 32Male20Temporal lobe epilepsy left-Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female30Healthy participant-Case 40Male27Healthy participant-	Case 31	Female	19	Temporal lobe epilepsy left	-
Case 33Male21Temporal lobe epilepsy left-Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female30Healthy participant-Case 40Male27Healthy participant-	Case 32	Male	20	Temporal lobe epilepsy left	-
Case 34Female47Temporal lobe epilepsy left-Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female43Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 33	Male	21	Temporal lobe epilepsy left	-
Case 35Male49Temporal lobe epilepsy left-Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female43Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 34	Female	47	Temporal lobe epilepsy left	-
Case 36Female32Temporal lobe epilepsy left-Case 37Male35Healthy participant-Case 38Female43Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 35	Male	49	Temporal lobe epilepsy left	-
Case 37Male35Healthy participant-Case 38Female43Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 36	Female	32	Temporal lobe epilepsy left	-
Case 38Female43Healthy participant-Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 37	Male	35	Healthy participant	-
Case 39Female30Healthy participant-Case 40Male27Healthy participant-	Case 38	Female	43	Healthy participant	-
Case 40 Male 27 Healthy participant –	Case 39	Female	30	Healthy participant	-
	Case 40	Male	27	Healthy participant	-

resolution T1-weighted MR images were acquired using a 3D MPRAGE sequence (TE = 3.02 ms, TR = 2190 ms, inversion time (TI) = 1300 ms) with a matrix size of  $250 \times 250 \times 256$ , with isometric voxels with a nominal side length of 0.9 mm, flip angle of 9° and a GRAPPA acceleration factor of 2.

## 2.3. Image preprocessing

First, binary masks delineating lesions in original unprocessed anatomical T1 images were defined manually in the native space of each patient using MRIcron (Rorden and Brett, 2000). Although it has been repeatedly shown that the quality of the mask has limited influence on the normalization results, tumor boundaries were outlined as precisely as possible by experienced clinical fMRI experts (FF, RB) (Andersen et al., 2010; Brett et al., 2001). Lesion masks generated in this way were smoothed with an 8 mm FWHM Gaussian filter as recommended by Brett et al. (2001) and constrained so as not to extend beyond the brain.

In a separate step, brain extracted images, i.e. the deskulled anatomical images, were obtained using FSL's (Software library of the Oxford Centre for Functional MRI of the Brain (FMRIB): http://www.fmrib.ox. ac.uk/fsl/) brain extraction tool (BET2; Smith, 2002) followed by manual removal of residual non-brain areas, again using MRIcron. To this end, a mask was drawn capturing residual non-brain areas including bone, fat, and meninges and added to the brain mask resulting from BET2. The



Fig. 1. Histogram of lesion size across the three lesioned brain groups. The numbers on the abscissa correspond to the patient numbers listed in Table 1.

amount of manual editing needed was comparable for the four lesion groups. This combined mask was applied to individual T1 scans, resulting in clean deskulled anatomical images.

Image processing, involving the different normalization pipelines, preprocessing and statistical analysis of the functional data was performed using SPM8 (Software library by the members & collaborators of the Wellcome Trust Centre for Neuroimaging (Functional Imaging Laboratory Group); http://fil.ion.ulc.ac.uk/spm) and largely followed the steps described by Crinion et al. (2007). Default parameters were chosen for all analysis steps - except where noted in the following description - to keep the normalization and analysis procedures as close as possible to that used in current practice. Normalization of the structural and functional images involved two steps. Step I: generation of a common spatial starting point; ensuring that images had the same rotation and origin as the MNI template by applying an affine 3D rigid-body transformation. Step II: standard SPM normalization (Ashburner and Friston, 1999), Unified Segmentation normalization (Ashburner and Friston, 2005) and New Segment normalization (SPM manual, FIL Group) using skulled and deskulled input data sets.

# 2.3.1. Step I (see Fig. 2)

The estimation of different parameter sets to transform the data to MNI space. First, to account for residual small-scale motion, motion correction parameters were calculated for the functional images using the individual mean EPI image as the reference image. To minimize interpolation errors, these parameters were calculated but not applied to the individual images at this stage, i.e. images were not resliced or resampled in this step. Secondly, both deskulled and not-deskulled structural T1 images were transformed to the individual mean EPI image, calculating EPI-space transformation parameters and then transforming to MNI space using affine 3D rigid-body transformation with the standard SPM T1 template as reference. MNI transformation parameters were thereby generated. These two parameter sets (motion correction parameters and MNI transformation parameters) were then combined to generate a combined transformation which was applied to the functional EPI data. The same procedure was applied to the structural and lesion mask images by combining EPI-space and MNI transformation parameters. Although this is not usually required at this stage of the data analysis, all data sets were resliced then resampled to  $2\times 2\times 2$  mm voxel size for the functional data and  $1\times 1\times 1$  for the anatomical data. This step resulted in a common starting point for the subsequent normalization pipelines and was conducted to exclude any confounding effects. Among these are possible distortions resulting from prior non-applied transformations, e.g. Unified Segmentation required the images to be in the approximate position of the MNI space before starting the normalization while standard normalization does not.

To check for possible differences between skulled and deskulled images introduced by the linear transformations of Step I, we performed two analyses. (1) Comparison of skulled with deskulled T1 images after registration of T1 to the mean EPI. (2) Comparison of skulled with deskulled T1 images after Step I had been completed (i.e. after generation of a uniform starting-point for all 6 normalizations). This was done by calculating DICE similarity indices (Dice, 1945) for the skulled/deskulled T1 images. These provide a direct measure of the structural differences between skulled and deskulled T1 at stages (1) and (2). DICE calculations were performed separately for the 4 different lesion groups and with the approach described below (section "Evaluation of structural differences between normalized and template images"). The comparison of skulled with deskulled T1 images was carried out with the deskulled image serving as the reference and the skulled image as the template.

### 2.3.2. Step II (see Fig. 3)

The default parameters implemented in SPM8 were used for the standard normalization scheme, except for the specification of the template image. Since the MNI152 template provided by SPM8 contains scalp, skull, and meninges, the brain extracted and the standard MNI152 templates provided by FSL were used as references (Fein et al., 2006). For consistency with the SPM template, both FSL templates were smoothed using an 8 mm FWHM Gaussian kernel and then used as reference images to normalize the stripped and non-stripped individual brains. For the Unified Segmentation Model, all parameters including the probabilistic prior maps were left unchanged (following Andersen et al., 2010; Crinion et al., 2007). In accordance with these studies, the number of Gaussians for the "other prior map" (see above) was left unchanged, i.e. it was assumed that the number of different intensity distributions within this tissue map would not be changed by stripping off the skull. Both approaches were conducted with cost-function masking to weight brain lesions appropriately (Andersen et al., 2010) for the three lesioned brain groups. The New Segment algorithm does not support cost function masking but is designed to ignore voxels with a value of zero, which is essentially identical to a cost-function masking approach (personal communication with John Ashburner, FIL methods group). Therefore, lesioned neuronal areas within the anatomical images were first discarded by setting their voxel-values to zero and the resulting "cleaned" stripped and non-





estimation of parameters
application of estimated parameters

Fig. 2. Flow chart delineating the preprocessing steps undertaken to achieve a common starting point for the 6 normalization pipelines, i.e. estimation steps to transform the data into MNI space. See text for further details.

stripped anatomical images were then submitted to New Segment normalization using the default parameters as provided by the authors of the toolbox (SPM manual, FIL Group). Again, this "cleaning" of the anatomical images was only conducted for the lesioned brain group.

This estimation procedure yielded six sets of transformation parameters corresponding to skulled and deskulled data sets submitted to either standard normalization, Unified Segmentation or New Segment. In all cases these transformation parameters were applied to the structural images, the lesion mask, and the functional data if appropriate, i.e. transformation parameters obtained from the New Segment approach were applied to the original, not "cleaned" structural images.

## 2.4. Analysis of structural data

2.4.1. Evaluation of intensity differences between normalized and template images

To assess the general quality of the different normalizations the mean square error (MSE) of intensities was calculated between

# Step II: General approach conducted for each of the normalization pipelines



Fig. 3. Flow chart delineating the normalization steps illustrating the general approach conducted for each of the six normalization pipelines. See text for further details.

averaged volumes (mean intensities across all patients) and the brain extracted MNI152 template provided by FSL (Hellier et al., 2003) which we used as the template for normalization in this study. Previous literature has shown that MSE values are useful as comprehensive indicator of general normalization quality and provide a robust statistical measure of intensity similarities (c.f. Razlighi et al. (2013); Ripollés et al. (2012)). The value of MSE is always positive, and is defined such that zero represents the ideal but practically unlikely gold standard of identical image intensities. Since this measure assumes identical MR scanner calibration, all image intensities were scaled to a maximum of one. The averaged brain volume across all patients (one for each of the 4 normalizations), was calculated as the weighted mean for each voxel excluding the individual lesioned brain areas as defined by the lesion mask after normalization for the three lesioned brain groups. Subsequently, the MSE was calculated as the mean squared difference between this weighted averaged image and the reference separately

for each normalization using only voxels belonging to the brain of the reference image. That is, voxels belonging to the skull, for example, were left out.

# 2.4.2. Evaluation of structural differences between normalized and template images

To assess the quality of the various normalization approaches in more detail, differences between normalized brains and the MNI152 template were assessed using a second approach — the DICE Similarity Index (DSI; Dice, 1945). This index measures the overlap between template and individual normalized brain, separately for whole brain, gray matter (GM) and white matter (WM). This index indicates how well the group of normalized images fits to the template and is within the range 0 (no overlap) to 1 (perfect agreement), meaning perfect alignment or similarity. This measure has also repeatedly been used to quantify normalization quality (e.g. Klein et al. (2009); Ripollés et al.

(2012)). To this end, the normalized anatomical images resulting from each pipeline as well as the brain extracted MNI152 template were segmented using the "New Segment" approach implemented in SPM8 (SPM manual, FIL Group). The rationale for re-segmentation was the fact that only 2 of the 3 normalizations (Unified Segmentation and New Segment) provide segmented tissue maps. These were generated prior to normalization. In order to avoid bias in further analysis towards one or the other approach, we decided to run a segmentation at this point for all normalization routes, not only for standard normalization. The DSI was then calculated for the whole brain as well as for WM and GM for each normalization separately using the segmented MNI152 template as a reference. Results were compared using random effect analyses of variance (RFX-ANOVA) with the within subject factors Skull (skulled/deskulled) and Normalization (standard/unified/new segment) and the between subject factor Group (no-lesion, smalllesion, medium-lesion, large-lesion). These ANOVAs were calculated separately first for the whole brain, disregarding tissue types, and then for the two tissue types of interest (gray matter, white matter) resulting from the re-segmentation.

Finally, a visual inspection of all brains was performed by two of the authors (RB, FF) evaluating every patients' normalized brain from all 6 pipelines with a focus on the pipelines with the largest DICE difference (see Fig. 5). This was carried out to identify poor normalization and segmentation results and to ensure that DICE values (see below) corresponded to visible outcomes.

# 2.5. Analysis of functional data

Following normalization using the six pipelines, all functional images were spatially smoothed using a Gaussian kernel (FWHM = 5 mm). For single subject analysis, statistical parametric maps were calculated separately for each run using a General Linear Model that included a single regressor representing the activity phase, convolved with a canonical hemodynamic response function. Six nuisance regressors, corresponding to the motion realignment parameters were also included in the model to regress out residual motion artifacts. For this single subject analysis standard default parameters were used, i.e. the model included a high-pass filter of 128 s as well as an AR(1) term. The resulting statistical maps for the regressor of interest were combined across all runs to form one contrast image representing language-related activations.

In order to address activation differences between the different normalization pipelines a random effects repeated measures  $2 \times 3 \times 4$ ANOVA was calculated with the within subject factors Skull (skulled/ deskulled) and Normalization (standard normalization, Unified Segmentation, New Segment) and the between subject factor Group (nolesion, small-lesion, medium-lesion, large-lesion). For the calculation of this model a repeated measures GLM with partitioned error variances (in which between-subject and within-subject error terms are modeled separately) was used, allowing between-subject and within-subject effects to be tested within one model.

Statistical parametric maps were thresholded using a voxel-wise p < 0.001. Since our primary interest was in clinically relevant effects, all data were masked exclusively for an extended temporoparietal ROI (including Wernicke's area) and an extended inferior frontal ROI (including Broca's area) using automated anatomical labeling (AAL; Tzourio-Mazoyer et al., 2002) and the Wake Forest University PickAtlas (WFU; Maldjian et al., 2003). In addition, an individual neuroanatomical assessment of functional localization was performed. Statistical t-maps were overlaid onto the warped individual anatomical image and onto the MNI152 template for visual inspection of functional clusters (Wernicke and Broca) and ROI peak activation (peak t-value) in relation to individual neuroanatomy was evaluated by two of the authors (RB, FF, see Fig. 8). For this, the patients' independent (non-normalized) clinical fMRI results, which are used in pre-surgical planning (Beisteiner

#### Table 2

MSE coefficients for the different normalization pipelines separately for the four different lesion size groups (no-lesion, small-lesion, medium-lesion, large-lesion).

	No- lesion	Small- lesion	Medium- lesion	Large- lesion
Skulled brains: Standard Normalization	0.1003	0.3044	0.2999	0.2892
Deskulled brains: Standard Normalization	0.0462	0.2108	0.2408	0.2302
Skulled brains: Unified Segmentation	0.1032	0.1881	0.3113	0.2625
Deskulled brains: Unified Segmentation	0.0360	0.1428	0.2800	0.2253
Skulled brains: New Segment Deskulled brains: New Segment	0.1068 0.0332	0.1120 0.0373	0.1173 0.0276	0.1304 0.0656

et al., 2000) and which have been verified via intraoperative cortical stimulation (see Roessler et al. (2005b)), served as a gold standard.

To check whether brain activation changes more when the lesion is closer to activation, we tested effects of "lesion-to-activation-distance" on normalization differences within the Wernicke area. For this we calculated the Euclidian distance between the lesion (border of the lesion mask) and the peak activation for every patient on original non-transformed functional EPI data. This generated the "lesion-toactivation-distance". Since the main focus of our study was on differences between skulled/deskulled input data, we then checked the influence of "lesion-to-activation-distance" on "differences in peak voxel location" between skulled and deskulled data by calculating corresponding correlations for all 3 normalizations.

Based on the hypothesis that within subject differences in normalization quality will also lead to differences in the MNI localization of the peak activation, we correlated the maximum DICE difference (gray

## Table 3a

Mean DICE coefficients and results from the repeated-measures ANOVA using the factors Group (no-lesion, small-lesion, medium-lesion, and large-lesion), Normalization (Standard Normalization, Unified Segmentation, New Segmentation) and Skull (deskulled, skulled images) separately for whole-brain, gray-matter and white matter. Only significant effects with their corresponding mean DICE coefficients (standard error is given in brackets) are given.

Whole-brain analysis:
Main effect Skull: F = 1141.209; df = 1,36; p < 0.000
Deskulled: 0.857 (0.002); skulled: 0.786 (0.002)
Main effect Group: F = 5.258; df = 3,36; p < 0.004
No-lesion: 0.830 (0.003); small: 0.820 (0.003); medium: 0.819 (0.003);
large: 0.817 (0.003)
Interaction effect Skull $\times$ Group: F = 8.590; df = 3,36; p < 0.000
Deskulled: No-lesion: 0.875 (0.005); small: 0.853 (0.005); medium: 0.851
(0.005); large: 0.849 (0.005)
Skulled: No-lesion: 0.786 (0.001); small: 0.788 (0.001); medium: 0.787
(0.001); large: 0.785 (0.001)
Gray-matter analysis:
Main effect Skull: F = 127.169; df = 1,36; p < 0.000
Deskulled: 0.857 (0.002); skulled: 0.786 (0.002)
Main effect Normalization: $F = 7.207$ ; $df = 2,72$ ; $p < 0.001$
Normalization: 0.747 (0.004); Unified Segmentation: 0.764 (0.008);
New Segment: 0.779 (0.002)
Interaction effect Normalization $\times$ Skull: F = 7.859; df = 2,72; p < 0.001
Deskulled: Normalization: 0.771 (0.005); Unified Segmentation: 0.781 (0.09);
New Segmentation: 0.788 (0.004)
Skulled: Normalization: 0.723 (0.004); Unified Segmentation: 0.746 (0.007);
New Segmentation: 0.752 (0.002)
Interaction effect Normalization $\times$ Skull $\times$ Group: F = 3.387; df = 6,36; p > 0.005
See Table 3b for details on mean DICE coefficients.
White-matter analysis:
Main effect Skull: $F = 234.189$ ; $df = 1,36$ ; $p < 0.000$
Deskulled: 0.682 (0.003); skulled: 0.653 (0.003)
Interaction Normalization $\times$ Skull: F = 14.312; df = 2,72; p < 0.000
Deskulled: Normalization: 0.679 (0.002); Unified Segmentation: 0.679 (0.007);
New Segmentation: 0.687 (0.004)
Skulled: Normalization: 0.642 (0.003); Unified Segmentation: 0.656 (0.006);
New Segmentation: 0.661 (0.003)

#### Table 3b

DICE coefficients for the different normalization pipelines separately for whole-brain (WB), gray-matter segmentation (GM) and white-matter segmentation (WM) (see also Fig. 4).

Structure	Deskulled brains: Normalization	Skulled brains: Normalization	Deskulled brains: Unified Segmentation	Skulled brains: Unified Segmentation	Deskulled brains: New Segment	Skulled brains: New Segment
No-lesion Group:						
Whole brain analysis	0.8659	0.7825	0.8690	0.7881	0.8898	0.7870
Gray-matter segmentation	0.7371	0.7072	0.7832	0.7432	0.7887	0.7597
White-matter segmentation	0.6809	0.6453	0.6716	0.6501	0.6924	0.6719
Small-lesion Group:						
Whole brain analysis	0.8506	0.7889	0.8551	0.7869	0.8528	0.7868
Gray-matter segmentation	0.7776	0.7269	0.7784	0.7373	0.7984	0.7507
White-matter segmentation	0.6845	0.6515	0.6813	0.6570	0.6967	0.6679
Medium-lesion Group:						
Whole brain analysis	0.8513	0.7868	0.8519	0.7870	0.8499	0.7869
Gray-matter segmentation	0.7968	0.7284	0.7907	0.7541	0.7940	0.7502
White-matter segmentation	0.6808	0.6364	0.6884	0.6617	0.6926	0.6617
Large-lesion Group:						
Whole brain analysis	0.8447	0.7843	0.8560	0.7873	0.8454	0.7822
Gray-matter segmentation	0.7706	0.7281	0.7735	0.7509	0.7712	0.7476
White-matter segmentation	0.6699	0.6334	0.6761	0.6550	0.6681	0.6431

matter) and the corresponding Euclidian distance of peak activations and tested whether the resulting Pearson's r was positive and significantly different from zero. To this end, we quantified the largest from all pairwise DICE differences per participant and calculated the Euclidian distance between the peak activations within Wernicke area of those two corresponding normalization pipelines.

# 3. Results

# 3.1. Structural analysis

## 3.1.1. Structural T1 differences within postprocessing Step I

There was very good congruence between skulled and deskulled T1 images after registration to EPI and the entirety of Step I (all DICE coefficients >0.98 for all analyses and lesion groups). This indicates that the structural differences described below are introduced during *normaliza-tion* (Step II) of the skulled/deskulled images. DICE results were also confirmed via subject-wise visual inspection of overlaid images (skulled overlaid on deskulled).

# 3.1.2. Evaluation of intensity differences between normalized and template images

The mean squared error in intensities revealed a general improvement in the quality of the normalization for skull-stripping (for details, see Table 2). The mean MSE value for deskulled images was 0.13 and the mean MSE for skulled images was 0.19. In addition, the New Segment normalization clearly outperformed the 2 other normalization techniques. The size of brain lesions also affected results. Normalization quality was worse with larger brain lesions. For no-lesion/small-lesion the mean MSE value was 0.12, for medium-lesion/large-lesion it was 0.21.

# 3.1.3. Evaluation of structural differences between normalized and template images

DICE coefficients used to assess the quality of the different approaches were submitted to ANOVAs. Detailed results are shown in Tables 3a and 3b and depicted in Figs. 4 and 5. The dominant finding was a significant improvement of template congruence for the skull-stripped images in every tissue category (whole brain, gray matter, white matter). Further parts of the analysis (main effects and interactions) indicated that template congruence was worse with older normalization techniques and larger brain lesions. All findings could be confirmed by the visual qualitative control (see Fig. 5).

#### 3.2. Functional analysis

Reliable task related activations were found within Wernicke and Broca AAL regions and other brain areas as described previously (Foki et al., 2008; Gartus et al., 2009). Detailed results of the  $2 \times 3 \times 4$  RFX-ANOVA are shown in Table 4 and illustrated in Figs. 6-8. Concerning general pipeline dependent localization effects, the 6 different normalization pipelines shifted the Wernicke peak more than 1 cm within the MNI space (group data, Fig. 7). The ANOVA generated 3 significant results: a main effect Skull, a main effect Normalization and an interaction Skull  $\times$  Group. Skull stripping specifically affected the cortex adjacent to Wernicke's core area, which is located in the posterior superior temporal gyrus. Skulled input data showed larger activations in inferior parietal cortex and in the anterior superior temporal gyrus - both outside of the classical Wernicke core (Fig. 7B, D). Analysis of the significant Skull  $\times$  Group effect (again in inferior parietal cortex) indicated that the skulled > deskulled differences are primarily driven by the medium and large lesion groups.



Fig. 4. DICE-values for the different normalization pipelines, for whole-brain (WB), graymatter segmentation (GM) and white-matter segmentation (WM) separately for the six normalization pipelines and across the four lesion size groups. For detailed values separated by lesion size see Tables 3a and 3b.



**Fig. 5.** Examples for misaligned brains. Patients with a large (top and middle row, cases 4 and 3) or a small (bottom row, case 25) difference in DICE indices. Most of the patients showed the largest DICE difference between standard normalization without skull-stripping and New Segment with skull-stripping. MNI slices z: -40 and z: +15 are shown. The MNI template is outlined in red. Note the considerable mismatch within ventricular planes (+15) in the top row and the mismatch within basal planes (-40) for case 3. Case 25 (bottom row) with similar DICE values for all 6 pipelines shows also similar brain alignments. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

The normalization techniques affected functional results in a similar way, generating significant differences adjacent to the Wernicke core. While Unified Segmentation and New Segment showed comparable functional signals, standard normalization generated much larger activation in inferior parietal cortex (supramarginal gyrus) and middle temporal gyrus – again both outside of the classical Wernicke core (Fig. 7A, C).

#### Table 4

Results of the 2 $\times$ 3 $\times$ 4 RFX-ANOVA (p = 0.001 uncorr): Anatomical regions with MN
coordinates and location of the peak-voxel within each cluster are given.

Anatomical region – location (area)	x, y, z (mm)	F
Main effect skull:		
Left anterior superior temporal gyrus	-55-64	23.746
Left inferior parietal cortex	-49 - 32  18	19.80
Main effect normalization:		
Left middle temporal gyrus	-47 - 48 22	14.179
Left supramarginal gyrus	-55 - 50 30	10.961
Interaction effect Skull $ imes$ Group		
Left supramarginal gyrus	-51 - 50 26	8.091

Note that all activations listed are significant at p < 0.001 uncorrected. Per cluster center (bold face) maximal 2 additional local maxima were listed > 8.0 mm apart.

Concerning the question of whether Wernicke activation changes more when the lesion is closer to activation, no significant correlation was found. The shift of peak activation between skulled and deskulled brains did not correlate with individual lesion-to-activation-distances (Wernicke ROI: Standard Normalization: r = -0.24, Unified Segmentation: r = -0.16, New Segment: r = 0.1). However, our hypothesis is that differences in DICE indices reflect localization differences of the peak activation (r = 0.28, p = 0.0402). This indicates that an increase in the deviation between brain and template also increases the shift of functional activations in MNI space.

# 3.3. Neuroanatomical assessment of individual fMRI activations

The changes in MNI coordinates of group activation clusters (Fig. 7) were further elucidated by a qualitative single subject analysis (Fig. 8) in which locations were compared with those established in the clinical patient reports (Beisteiner et al., 2000, 2008; Roessler et al., 2005a). This revealed that activation strength, cluster size and position of activation clusters relative to surrounding neuroanatomy were quite stable. In addition, the atypical "Wernicke activations" found for not-deskulled input data and standard normalization (inferior parietal, anterior



**Fig. 6.** One-sample t-test group results. Significant activation above a threshold of p < 0.001 uncorrected is overlaid on the brain extracted or the standard MNI152 templates provided by FSL. Note that the position of the activation cluster differs (c.f. slice 18 showing almost no activation for the Unified Segmentation Model with skull-stripping as indicated with a red circle) and the Wernicke peak-voxel is shifted between normalization pipelines >1 cm (indicated with an arrow, locations are given in MNI coordinates). Only slices covering the Wernicke area are shown. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

superior temporal, middle temporal) were not evident in the patient reports based on standard clinical thresholds. A secondary analysis of the group data (the details of which we do not report) confirmed these individual qualitative findings by demonstrating that group cluster sizes and group cluster t-values did not differ between the 6 normalization pipelines. However, as is also evident from the visual analysis of the DICE index differences (Fig. 5), the overall brain positions varied in MNI space depending on the normalization pipeline. The consequence of this finding is that the MNI coordinates of an activation cluster change despite keeping a rather stable position within the individual brain (Fig. 8). Correspondingly, MNI peak activation coordinates varied depending on the normalization pipeline — typically below 1 cm, but up to 4.6 cm with one outlier patient.

# 4. Discussion

Our study provides 2 major results: (1) structural analysis indicates that the most reliable MNI coordinates are achieved using *deskulled* 



**Fig. 7.** Main effects and contrasts of the 2 × 3 × 4 RFX-ANOVA: Activation differences found for the two main effects "normalization" (in left supramarginal gyrus and left middle temporal gyrus) (A) and "skull-stripping" (in left anterior superior temporal gyrus and left inferior parietal cortex) (B) are shown, rendered onto the SPM5 single-subject brain template. Contrast estimates for all significant brain areas are shown in panels C and D. Anatomical regions with MNI-coordinates and location of the peak-voxel within each cluster can be found in Table 4. All data are masked exclusively for Wernicke's area.

*input data* — particularly when combined with new normalization techniques (Unified Segmentation, New Segment). (2) As a consequence, the MNI coordinates of essential language activations may be partly misleading with skulled input data sets — particularly when combined with standard normalization. This specifically concerns parietal and temporal cortex.

In more detail, we found that the skulled brains, for which normalization quality was inferior (see Fig. 5) lead to misleading activation. This is shown by a significant skulled > deskulled activation in the left inferior parietal cortex  $(-49 - 32 \ 18 \ in \ Table 4, red circles \ in \ Fig. 6)$  and left anterior superior temporal gyrus (-55 - 64 in Table 4) - bothclearly outside the classical Wernicke core. The term "misleading" seems justified for 3 reasons: (1) the remote parietal and temporal activations were not seen with standard clinical thresholds (clinical patient reports), (2) the larger the mismatch between template and brain (which was largest with skulled data), the greater the change in the location of Wernicke activation, and (3) no pipeline changed Wernicke activations significantly in relation to local neuroanatomy (Fig. 8), but neuroanatomy changed in relation to the MNI coordinates (i.e. a brain-template mismatch occurred with skulled data). Therefore, the conclusion must be that normalization of skulled brains shifts part of the "correct" Wernicke activations to "wrong" MNI coordinates in the temporo-parietal cortex. A similar temporo-parietal effect was found for the standard normalization technique. Standard normalization generated a misleading activation increase in left supramarginal gyrus and left middle temporal gyrus outside the Wernicke core.

Details of the structural analysis revealed that the most important factor for improvement of the congruence between the MNI template and normalized brains (MSE values, DICE coefficients, Figs. 4 and 5) was skull-stripping. Further, the new normalization techniques outperformed standard normalization with New Segment proving to be the best approach. Evaluation of the procedures required for generation of a uniform starting-point of skulled/deskulled brains (Step I) indicated that the decisive differences between the 6 normalization pipelines were introduced during Step II.

With regard to lesion size effects, a systematic influence was found for the structural data: brains with larger lesions differed more significantly from the template than brains with smaller lesions. With the functional data, lesion size tended to increase the functional mislocalizations (larger parietal skulled > deskulled effects). The distance between brain lesion and brain activation however, did not significantly affect normalization quality.

Summarizing our functional and structural findings (Figs. 5–8), the primary cause of our activation differences is a differing quality of alignment between normalized brains and MNI template. This leads to shifts in activation clusters and peak activations (maximum 4.6 cm) within MNI space. Skull-stripping the input data is the most important factor in improving this. Clearly, the implicit skull-stripping step, already implemented in most normalization algorithms, does not produce results of the same quality as explicitly editing the input data. With standard normalization, implicit skull-stripping is realized by weighting of nonbrain voxels to exclude non-brain structures after an initial affine transformation but prior to nonlinear normalization. With Unified Segmentation (Ashburner and Friston, 2005) tissue probability maps for gray matter, white matter, cerebrospinal fluid (CSF) and a fourth map for the residuals are generated. The latter implicitly accounts for the skull and the scalp. However, in concordance with Fein et al. (2006) and Acosta-Cabronero et al.(2008) we found a clear benefit for normalization quality if deskulled data are used as primary input for the normalization process.

Interestingly, the left temporoparietal position of our functional differences corresponds to the left temporal differences found for different normalization algorithms in the work of Crinion et al. (2007), who also investigated language data but not skull-stripping effects. Their and our results indicate that temporal areas are a specific source of structural variability during the normalization process with current templates.

378



Fig. 8. Brain position and MNI coordinates of peak voxel location for a representative patient (case 6) resulting from the 6 normalization pipelines. Note that the Wernicke peak-voxel is located in the same neuroanatomical region, yet this region is shifted in the MNI space.

Besides choosing optimized postprocessing techniques it seems sensible to recommend that group studies, where critical activations are expected in temporal brain areas, include a series of single-patient analyses to check for internal consistency of the structural and functional data. A further issue of special clinical relevance concerns activations in other brain areas comprising essential cortex. MNI peak activation shifts of the size found here (>1 cm with the group data, >4 cm individually) may easily become critical. For example in primary sensorimotor cortex around the central sulcus, such a peak activation shift within MNI space may well decide between concluding that the main result of a study is primary motor activation or a primary sensory activation. Therefore, skull-stripping of the input data should become standard, but not only for clinical studies. It can be performed either as a separate step or by inclusion in standard analysis pipelines, as already suggested for non-human data (Budin et al., 2013). Our implementation of skullstripping with the BET2 software requires considerable manual postprocessing to obtain optimal removal of non-brain areas. Newer and potentially more accurate algorithms, such as the simplex mesh and histogram algorithm (SMHASS; Galdames et al., 2012), may be candidates for integration into fully automated routines.

In conclusion, we have shown that combining deskulled input data with the New Segment normalization technique generates the highest probability of achieving valid MNI coordinates for functional activations. The functional and structural variability described is relevant for functional conclusions in a clinical context and should also be considered when comparing MNI coordinates from different fMRI studies.

### Acknowledgement

This study was supported by the Austrian Science Fund (FWF P23611), a research cluster grant of the Medical University and University of Vienna (SO76100002) and "Vienna Advanced Clinical Imaging Center" (VIACLIC) project, funded by the Vienna Spots of Excellence Program of the Center of Innovation and Technology, City of Vienna (ZIT), Austria.

## References

- Acosta-Cabronero, J., Williams, G.B., Pereira, J.M.S., Pengas, G., Nestor, P.J., 2008. The impact of skull-stripping and radio-frequency bias correction on grey-matter segmentation for voxel-based morphometry. NeuroImage 39, 1654–1665.
- Andersen, S.M., Rapcsak, S.Z., Beeson, P.M., 2010. Cost function masking during normalization of brains with focal lesions: still a necessity? NeuroImage 53, 78–84.
- Ashburner, J., Friston, K.J., 1999. Nonlinear spational normalization using basis functions. Hum. Brain Mapp. 7, 254–266.
- Ashburner, J., Friston, K.J., 2005. Unified segmentation. NeuroImage 26, 839-851.
- Beisteiner, R., Lanzenberger, R., Novak, K., Edward, V., Windischberger, C., Erdler, M., Cunnington, R., Gartus, A., Streibl, B., Moser, E., Czech, T., Deecke, L., 2000. Improvement of presurgical patient evaluation by generation of functional magnetic resonance risk maps. Neurosci. Lett. 290, 13–16.
- Beisteiner, R., Drabeck, K., Foki, T., Geissler, A., Gartus, A., Lehner-Baumgartner, E., Baumgartner, C., 2008. Does clinical memory fMRI provide a comprehensive map of medial temporal lobe structures? Exp. Neurol. 213, 154–162.
- Beisteiner, R., Klinger, N., Höllinger, I., Rath, J., Gruber, S., Steinkellner, T., Foki, T., Geissler, A., 2010. How much are clinical fMRI reports influenced by standard postprocessing methods? An investigation of normalization and region of interest effects in the medial temporal lobe. Hum. Brain Mapp. 31, 1951–1966.

Brett, M., Leff, a.P., Rorden, C., Ashburner, J., 2001. Spatial normalization of brain images with focal lesions using cost function masking. NeuroImage 14, 486–500.

Brett, M., Johnsrude, I.S., Owen, A.M., 2002. The problem of functional localization in the human brain. Nat. Rev. Neurosci. 3, 243–249.

- Budin, F., Hoogstoel, M., Reynolds, P., Grauer, M., O'Leary-Moore, S.K., Oguz, I., 2013. Fully automated rodent brain MR image processing pipeline on a Midas server: from acquired images to region-based statistics. Front. Neuroinform. 7, 15.
- Crinion, J., Ashburner, J., Leff, A., Brett, M., Price, C., Friston, K.J., 2007. Spatial normalization of lesioned brains: performance evaluation and impact on fMRI analyses. NeuroImage 37, 866–875.
- Dice, L.R., 1945. Measures of the amount of ecologic association between species. Ecology 26, 297–302.
- Edward, V., Windischberger, C., Cunnington, R., Erdler, M., Lanzenberger, R., Mayer, D., Endl, W., Beisteiner, R., 2000. Quantification of fMRI artifact reduction by a novel plaster cast head holder. Hum. Brain Mapp. 11, 207–213.
- Fein, G., Landman, B., Tran, H., Barakos, J., Moon, K., Di Sclafani, V., Shumway, R., 2006. Statistical parametric mapping of brain morphology: sensitivity is dramatically increased by using brain-extracted images as inputs. NeuroImage 30, 1187–1195.
- Foki, T., Gartus, A., Geissler, A., Beisteiner, R., 2008. Probing overtly spoken language at sentential level: a comprehensive high-field BOLD-fMRI protocol reflecting everyday language demands. NeuroImage 39, 1613–1624.
- Galdames, F.J., Jaillet, F., Perez, C.a, 2012. An accurate skull stripping method based on simplex meshes and histogram analysis for magnetic resonance images. J. Neurosci. Methods 206, 103–119.
- Gartus, A., Geissler, A., Foki, T., Tahamtan, A.R., Pahs, G., Barth, M., Pinker, K., Trattnig, S., Beisteiner, R., 2007. Comparison of fMRI coregistration results between human experts and software solutions in patients and healthy subjects. Eur. Radiol. 17, 1634–1643.
- Gartus, A., Foki, T., Geissler, A., Beisteiner, R., 2009. Improvement of clinical language localization with an overt semantic and syntactic language functional MR imaging paradigm. AJNR Am. J. Neuroradiol. 30, 1977–1985.
- Hellier, P., Barillot, C., Corouge, I., Gibaud, B., Le Goualher, G., Collins, D.L., Evans, A., Malandain, G., Ayache, N., Christensen, G.E., Johnson, H.J., 2003. Retrospective evaluation of intersubject brain registration. IEEE Trans. Med. Imaging 22, 1120–1130.
- Hoeksma, M.R., Kenemans, J.L., Kemner, C., van Engeland, H., 2005. Variability in spatial normalization of pediatric and adult brain images. Clin. Neurophysiol. 116, 1188–1194.
- Klein, A., Andersson, J., Ardekani, B.a, Ashburner, J., Avants, B., Chiang, M.-C., Christensen, G.E., Collins, D.L., Gee, J., Hellier, P., Song, J.H., Jenkinson, M., Lepage, C., Rueckert, D., Thompson, P., Vercauteren, T., Woods, R.P., Mann, J.J., Parsey, R.V., 2009. Evaluation of 14 nonlinear deformation algorithms applied to human brain MRI registration. NeuroImage 46, 786–802.

- Klein, A., Ghosh, S.S., Avants, B., Yeo, B.T.T., Fischl, B., Ardekani, B.a, Gee, J.C., Mann, J.J., Parsey, R.V., 2010. Evaluation of volume-based and surface-based brain image registration methods. NeuroImage 51, 214–220.
- Maldjian, J.A., Laurienti, P.J., Kraft, R.A., Burdette, J.H., 2003. An automated method for neuroanatomic and cytoarchitectonic atlas-based interrogation of fMRI data sets. NeuroImage 19, 1233–1239.
- Razlighi, Q.R., Kehtarnavaz, N., Yousefi, S., 2013. Evaluating similarity measures for brain image registration. J. Vis. Commun. Image Represent. 24, 977–987.
- Ripollés, P., Marco-Pallarés, J., de Diego-Balaguer, R., Miró, J., Falip, M., Juncadella, M., Rubio, F., Rodriguez-Fornells, A., 2012. Analysis of automated methods for spatial normalization of lesioned brains. NeuroImage 60, 1296–1306.
- Roessler, K., Donat, M., Lanzenberger, R., Novak, K., Geissler, A., Gartus, A., Tahamtan, A.R., Milakara, D., Czech, T., Barth, M., Knosp, E., Beisteiner, R., 2005a. Evaluation of preoperative high magnetic field motor functional MRI (3 Tesla) in glioma patients by navigated electrocortical stimulation and postoperative outcome. J. Neurol. Neurosurg. Psychiatry 76, 1152–1157.
- Roessler, K., Donat, M., Lanzenberger, R., Novak, K., Geissler, a, Gartus, a, Tahamtan, a.R., Milakara, D., Czech, T., Barth, M., Knosp, E., Beisteiner, R., 2005b. Evaluation of preoperative high magnetic field motor functional MRI (3 Tesla) in glioma patients by navigated electrocortical stimulation and postoperative outcome. J. Neurol. Neurosurg. Psychiatry 76, 1152–1157.
- Rorden, C., Brett, M., 2000. Stereotaxic display of brain lesions. Behav. Neurol. 12, 191–200.
- Smith, S.M., 2002. Fast robust automated brain extraction. Hum. Brain Mapp. 17, 143–155.
- Tahmasebi, A.M., Abolmaesumi, P., Zheng, Z.Z., Munhall, K.G., Johnsrude, I.S., 2009. Reducing inter-subject anatomical variation: effect of normalization method on sensitivity of functional magnetic resonance imaging data analysis in auditory cortex and the superior temporal region. NeuroImage 47, 1522–1531.
- Tzourio-Mazoyer, N., Landeau, B., Papathanassiou, D., Crivello, F., Etard, O., Delcroix, N., Mazoyer, B., Joliot, M., 2002. Automated anatomical labeling of activations in SPM using a macroscopic anatomical parcellation of the MNI MRI single-subject brain. NeuroImage 15, 273–289.
- Vandenbroucke, M.W., Goekoop, R., Duschek, E.J., Netelenbos, J.C., Kuijer, J.P., Barkhof, F., Scheltens, P., Rombouts, S.A., 2004. Interindividual differences of medial temporal lobe activation during encoding in an elderly population studied by fMRI. NeuroImage 21, 173–180.
- Yassa, M.a, Stark, C.E.L., 2009. A quantitative evaluation of cross-participant registration techniques for MRI studies of the medial temporal lobe. NeuroImage 44, 319–327.