

Association between *IL-1B* (-511)/*IL-1RN* (VNTR) polymorphisms and type 2 diabetes: a systematic review and meta-analysis

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ABSTRACT

Interleukin-1 (IL-1) plays an essential role in the immune pro-inflammatory process, which is regarded as one of many factors in the development of type 2 diabetes mellitus (T2DM). Several case-control studies have illustrated the association of the IL-1B (-511) (rs16944, Chr 2:112,837,290, C/T Intragenic, Transition Substitution) and IL-1RN (VNTR) (gene for IL-1 receptor antagonist, IL-1RA, 86 bp tandem repeats in intron 2) polymorphisms with T2DM risk. However, the results were inconsistent and inconclusive. We performed a meta-analysis (registry number: CRD42021268494) to assess the association of the IL-1B (-511) and IL-1RN (VNTR) polymorphisms with T2DM risk, Random-effects models were applied to calculate the pooled ORs (odds ratios) and 95% CIs (confidence intervals) to test the strength of the association in the overall group and subgroups stratified by ethnicity, respectively. Between-study heterogeneity and publication bias were evaluated by the Q-test, I² statistic, Harbord test, and Peters test accordingly. Sensitivity analyses were also performed. A total of 12 publications evaluating the association of IL-1B (-511) and IL-1RN (VNTR) polymorphisms with the risk of T2DM development were included. The meta-analysis showed that IL-1RN (VNTR) was related to the increasing development of T2DM risk in the recessive model (OR = 1.62, 95% CI [1.09–2.42], $P_{het} = 0.377$, $P_z = 0.018$) and in the homozygous model (OR = 2.02, 95% CI [1.07–3.83], $P_{het} = 0.085$, $P_z = 0.031$), and the IL-1RN 2* allele was found a significant association with evaluated T2DM risk in all ethnicities (OR = 2.08, 95% CI [1.43–3.02], P_{het} < 0.001, P_z < 0.001) and in EA (OR = 2.01, 95% CI [1.53–2.66], P_{het} = 0.541, P_z < 0.001). Moreover, stratification by ethnicity revealed that IL-1B (-511) was associated with a decreased risk of T2DM in the dominant model (OR=0.76, 95% CI [0.59-0.97], $P_{het} = 0.218$, $P_z = 0.027$) and codominant model (OR = 0.73, 95% CI [0.54–0.99], P_{het} = 0.141, P_z = 0.040) in the East Asian (EA) subgroup. Our results suggest that the IL-1RN 2* allele and 2*2* homozygous polymorphism are strongly associated with increasing T2DM risk and that

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the IL-1B (-511) T allele polymorphism is associated with decreasing T2DM risk in the EA subgroup.

Subjects Diabetes and Endocrinology, Epidemiology, Medical Genetics

Keywords IL-1B (-511), IL-1RN (VNTR), Polymorphism, Type 2 diabetes mellitus,

Meta-analysis

INTRODUCTION

The latest data from the 9th edition of the IDF Diabetes Atlas show that approximately 463 million adults (20–79 years) have diabetes, and by 2045, the number will rise to 700 million. Type 2 diabetes mellitus (T2DM) is the most prevalent type of diabetes (>90% of diabetes). The proportion of T2DM patients is increasing in most countries, especially in low- and middle-income countries such as China and India (https://www.idf.org/aboutdiabetes/what-is-diabetes/facts-figures.html). Growing evidence suggests that the risk of T2DM is associated with various factors, such as genetics, ethnicity, environment, and lifestyle. In particular, genetic susceptibility seems to play an essential role in the pathogenesis of T2DM (*Hivert, Vassy & Meigs*, 2014).

T2DM occurs when β -cell function progressively deteriorates and fails to compensate for insulin resistance, partly due to the demise of pancreatic β-cells through apoptosis (*Donath* & Halban, 2004). The chronic and low-grade inflammation in metabolic organs including the liver, brain, pancreas and adipose tissue is known as metaflammation (metabolisminduced inflammation), which can be widely observed in T2DM (Hotamisligil, 2006; Hotamisligil, 2017). Increasing evidence has shown that metaflammation plays a vital role in the development of T2DM and its cardiovascular complications (Hotamisligil, 2017) and pro-inflammatory cytokines, especially interleukin-1\(\beta\) (IL-1\(\beta\)), play a key role in this process (Dinarello, Donath & Mandrup-Poulsen, 2010; Fève & Bastard, 2009; Sathyapalan & Atkin, 2011). Moreover, the increased level of IL-1β in human pancreatic cells due to the elevated glucose concentrations and decreased level of IL-1 receptor antagonist (IL-1Ra) in islets of T2DM patients result in impaired insulin secretion, decreased cell proliferation, and apoptosis of β-cells (Böni-Schnetzler & Donath, 2013). A previous observational study demonstrated that a combined elevation of IL-1β and IL-6 was associated with a roughly threefold increased risk of T2DM, and IL-1β might induce insulin resistance via activating the IκB kinaseβ (Spranger et al., 2003). Furthermore, CANTOS and TRACK trials have evaluated the role of IL-1 inhibition in the treatment of several inflammatory disorders, such as coronary artery disease, T2DM and rheumatoid arthritis (Qamar & Rader, 2012; Ruscitti et al., 2019). According to the result from CANTOS trials, canakinumab (a fully humanized monoclonal antibody) could selectively block IL-1β, and its efficacy in suppressing both levels of inflammatory markers and clinical symptoms in patients with autoinflammatory disease such as T2DM and impaired glucose tolerance has been observed in this trial (Aday & Ridker, 2018; Rissanen et al., 2012). Therefore, the genetic polymorphisms that regulate the expression levels of IL-1β and IL-1Ra might have an essential impact on the interindividual differences in T2DM.

Consisting of three linked genes and mapping to chromosome 2q13-24, IL-1 cytokine genes encode IL-1α, IL-1β and IL-1Ra. Both IL-1α and IL-1β are pro-inflammatory cytokines, while IL-1Ra can inhibit inflammation by competing for receptor binding (*Dinarello*, 2000). In the promoter region, *IL-1B* (-511) (rs16944, Chr 2:112,837,290, C/T Intragenic, Transition Substitution) has a bi-allelic polymorphism at position -511, representing the C/T transition (Kristiansen et al., 2000). IL-1RN (VNTR) (gene for IL-1 receptor antagonist, IL-1RA, 86 bp tandem repeats in intron 2) has five different alleles: 1^* (four repeats), 2^* (two repeats), 3^* (five repeats), 4^* (three repeats) and 5^* (six repeats) (Tarlow et al., 1993). IL-1RN 1* (four repeats) and IL-1RN 2* (two repeats) are the most common, whereas the others occur much less frequently (<5%) (Santtila, Savinainen & Hurme, 1998). Many genetic association studies have been performed to estimate the relationship of the IL-1B (-511) and IL-1RN (VNTR) polymorphisms with T2DM risk (Achyut et al., 2007; Muktabhant et al., 2013). Several results suggest that there is no relationship between them (Borilova Linhartova et al., 2019; Muktabhant et al., 2013), whereas Achyut et al., (2007) showed that both the IL-1B (-511) and IL-1RN (VNTR) polymorphisms were associated with susceptibility to T2DM as well as complications, and he indicated that T2 (IL-1 β -511T/IL-1 RN^*2) haplotype was associated with a roughly twofold increased risk of T2DM. Considering the small sample sizes (n < 600) and varying population characteristics (ethnic differences) in different studies, the results are conflicting. To illustrate the potential association of the IL-1B (-511) and IL-1RN (VNTR) polymorphisms with T2DM risk, we conducted a meta-analysis including 12 reported publications.

MATERIALS AND METHODS

This systematic review was conducted following the Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) statement guidelines (*Moher et al.*, 2009). A protocol was registered before commencing this review on PROSPERO (CRD42021268494).

The search strategy

The online databases we searched included PubMed, Web of Science, CNKI (China National Knowledge Infrastructure) and Wanfang. We identified relevant articles reporting the association of *IL-1RN* (VNTR) and *IL-1B* (*-511*) polymorphisms with the risk of T2DM in the medical literature through the end of September 12, 2020. This study used a combination of the following search terms: "Type 2 Diabetes Mellitus" or "T2DM" or "interleukin-1" or "IL-1" or "polymorphism" or "genetics" or "association". In addition, we also conducted a hand search to identify relevant data in references from retrieved articles. Juan Jiao and Zhaoping Wang performed the Search Strategy. In case of disagreement, it will be settled by a third assessor's evaluation and discussed until a consistent result was reached.

The inclusion and exclusion criteria

All studies included in this analysis met the following criteria: (1) the study revealed the relationship of *IL-1B* (-511) and *IL-1RN* (VNTR) with T2DM risk; (2) the design

was a "case-control study"; (3) the research subjects were humans with T2DM; (4) the publication language was English or Chinese; and (5) the study provided sufficient data to estimate ORs (odds ratios) and 95% CIs (confidence intervals). The major exclusion criteria were as follows: (1) duplicate data; (2) randomly chosen controls; (3) non-research articles; and (4) insufficient reporting of data.

Data extraction

All data were systematically reviewed and extracted by two investigators (ZPW and DNG) according to a standardized form, and then all of the following information was collected in an electronic database: the first author's name, year of publication, country of origin, ethnicity, total number of cases/controls, genotyping method, diagnostic criteria of T2DM, genotypic frequencies and language of the report. Disagreement was settled by a third assessor's evaluation and discussed until a consistent conclusion was agreed.

Assessment of quality score

We assessed the quality score of identified publications based on the Newcastle Ottawa Scale (NOS) (http://www.ohri.ca/programs/clinical_epidemiology/oxford.asp). And the NOS involved "selection", "comparability", and "estimation of outcomes or exposures". The scores ranged from 0 to 9, and the score of "high quality study" was ≥ 6 (detailed scores see Table S1) (*Lo, Mertz & Loeb, 2014*). And the STREGA (Strengthening the REporting of Genetic Association) system was also performed to assessed the methodological quality of all included studies, which includes 22 items with scoring from 0 to 22 (detailed scores see Table S2) (*Little et al., 2009*). And the score of high quality was 18–22; the score of moderate-high quality was 13–17; the score of low quality was 0–12 (*Duan et al., 2018*).

Data analysis

We conducted this analysis and entered data with Stata software, version 15.0 (Stata Corp., College Station, TX, USA). We used original genotypic distribution data without any adjustment to measure the strength of the association between IL-1B (-511)/IL-1RN and T2DM risk by ORs (odds ratios) with 95% CIs (confidence intervals) under dominant, recessive, additive (overdominant+codominant) and homozygous models. The distributions of genotype frequencies of controls were all consistent with Hardy-Weinberg Equilibrium (HWE) (P > 0.05, Table 1). Between-study heterogeneity among the studies was tested with Cochran's O and the I^2 test statistics. I^2 was calculated based on the formula $I^2 = 100\% \times (Q-df)/Q$. Heterogeneity was regarded as significant when P_{het} < 0.1 or $I^2 > 50\%$. For those with significant heterogeneity, we performed stratified analyses to examine the statistical significance of the difference in ORs according to ethnicity (East Asian, South Asian, North African and Caucasian). We used the randomeffects model (DerSimonian and Laird method) which was combined by applying inverse variance-weighted meta-analysis to calculate the pooled OR and 95% CI, with $P_z < 0.05$ considered statistically significant (DerSimonian & Laird, 1986). When the heterogeneity was significant, sensitivity analyses were conducted to evaluate the influence of each single study by omitting one study at a time and checking the pool effect size for the remainder of the studies. Finally, the Harbord test and Peters test were used when fewer than ten

Table 1 Characteristics of Identified Studies on IL-1B (-511)/IL-1RN(VNTR) polymorphism and the risks of T2DM.

Author	Year	Country/Ethnicity	Diagnostic criteria	Genotyping method	Number		Language	IL-1 gene	QA	HWE
					Cases	Controls				
Zhang Jian	2004	China/EA	NA	PCR-RFLP	106	198	Chinese	IL-1RN	7	1.00
Zhang Ping-an	2004	China/EA	WHO, 1999	PCR-RFLP	106	247	Chinese	IL-1RN	8	0.70
Zhou Jian-zhong	2010	China/EA	WHO, 1999	PCR-RFLP	72	97	Chinese	IL-1RN	7	0.23
Petra Borilova Linhartova	2019	Czech Republic/Caucasian	NA	PCR-RFLP	380	212	English	IL-1RN	8	1.00
Alexandra I. F. Blakemore	1995	North of England/Caucasian	WHO, NA	PCR	117	248	English	IL-1RN	7	0.61
B.R Achyut	2006	North Indian/SA	WHO, 1999	PCR-RFLP	200	223	English	IL-1RN IL-1B (-511)	8	0.100.16
Liu Chang	2014	China/EA	WHO, 1999	PCR-RFLP	583	366	Chinese	IL-1B (-511)	7	1.00
Cao Yong	2013	China/EA	WHO, 1999	PCR-RFLP	268	263	Chinese	IL-1B (-511)	8	0.06
Lin Neng-bo	2016	China/EA	WHO, 1999	PCR-RFLP	286	327	Chinese	IL-1B (-511)	8	0.08
Natalie E. Doody	2017	North Indian/SA	WHO, 1999	PCR-RFLP	202	203	English	IL-1B (-511)	8	0.87
Benja Muktabhant	2013	Thailand/SEA	NA	PCR-RFLP	90	30	English	IL-1B (-511)	7	0.16
Safaa I. Tayel	2018	Egypt/NA ₁	ADA, NA	TaqMan	50	30	English	IL-1B (-511)	8	1.00

Notes.

T2DM, Type 2 diabetes mellitus; EA, East Asian; SA, South Asian; SEA, South East Asian; NA, Not Available (the diagnosis of T2DM was originally based on the presence of clinical symptoms and biochemical); NA1, North African; ADA, American Diabetes Association (fasting blood glucose ≥126 mg/dL or 2 h blood glucose after overload with 75 g of glucose ≥ 200 mg/dL in oral glucose tolerance test (OGTT) or glycated hemoglobin (HbA1c) ≥6.5 in patients with classic symptoms of hyperglycemia); WHO, World Health Organization (fasting glucose levels [7.0 mmol/L or 126 mg/dL] after a minimum 12-h fast or 2-h post glucose level (oral glucose tolerance test or 2-h OGTT) [11.1 mmol/L or 200 mg/dL] on more than one occasion); QA, Quality Assessment; HWE, Hardy-Weinberg Equilibrium.

articles were included, as they are more sensitive for assessing publication bias, with P < 0.1 assuming that the bias was statistically significant.

RESULTS

Included studies

Figure 1 shows the detailed process of selecting and assessing eligible studies. We identified 450 publications via the initial keyword search. After screening, 12 publications met our inclusion criteria (Achyut et al., 2007; Blakemore et al., 1996; Borilova Linhartova et al., 2019; Cao et al., 2013; Doody et al., 2017; Lin et al., 2016; Liu et al., 2014; Muktabhant et al., 2013; Tayel et al., 2018; Zhang, Xiao & Li, 2004a; Zhang et al., 2004b; Zhou et al., 2010), and 438 studies were excluded for containing duplicate data, missing critical data, and being review papers or studies that were not related to T2DM. Table 1 illustrates the characteristics of all included publications in this meta-analysis. In terms of polymorphisms and diseases, six identified case-control studies, comprising 981 cases and 1,225 controls, regarded IL-1RN (VNTR) to evaluate its association with T2DM risk (Achyut et al., 2007; Blakemore et al., 1996; Borilova Linhartova et al., 2019; Zhang, Xiao & Li, 2004a; Zhang et al., 2004b; Zhou et al., 2010), and seven identified case-control studies, including 1,679 cases and 1,442 controls, tested the association between *IL-IB* (-511) and T2DM risk (Achyut et al., 2007; Cao et al., 2013; Doody et al., 2017; Lin et al., 2016; Liu et al., 2014; Muktabhant et al., 2013; Tayel et al., 2018). Stratified by ethnicity, six publications were performed in East Asian (EA) populations (Cao et al., 2013; Lin et al., 2016; Liu et al., 2014; Zhang, Xiao & Li, 2004a; Zhang et al., 2004b; Zhou et al., 2010), two studies were undertaken in South Asian (SA) populations (Achyut et al., 2007; Doody et al., 2017), two studies were conducted in Caucasian populations (Blakemore et al., 1996; Borilova Linhartova et al., 2019), and the remaining studies were completed in Southeast Asian (SA) (Muktabhant et al., 2013) and North African (NA₁) populations (*Tayel et al.*, 2018).

Individual polymorphism meta-analysis

The genotypic distributions of the *IL-1B* (*-511*) and *IL-1RN* (VNTR) polymorphisms are shown in Table 2. Table 3 shows the meta-analysis results for these two polymorphisms. For the *IL-1B* (*-511*) polymorphism, seven studies with 1,679 cases and 1,442 controls were included in the meta-analysis. The results of the pooled analysis suggest that the *IL-1B* (*-511*) polymorphism is not significantly associated with T2DM risk in all study subjects under the dominant model (OR = 0.84, 95% CI [0.57–1.25], $P_{het} = 0.001$, $P_z = 0.395$) (Table 3 and Fig. 2A), recessive model (OR = 0.89, 95% CI [0.61–1.31], $P_{het} < 0.001$, $P_z = 0.561$) (Table 3), and homozygous model (OR = 0.82, 95% CI [0.48–1.39], $P_{het} < 0.001$, $P_z = 0.451$) (Table 3). In the stratification analyses by ethnicity, we found that there was a significant association between the *IL-1B* (*-511*) polymorphism and decreasing T2DM risk in the EA population under the dominant model (OR = 0.76, 95% CI [0.59–0.97], $P_{het} = 0.218$, $P_z = 0.027$) (Table 3 and Fig. 2A) and codominant model (OR = 0.73, 95% CI [0.54–0.99], $P_{het} = 0.141$, $P_z = 0.040$) (Table 3), but no association was observed under the recessive model (OR = 0.97, 95% CI [0.79–1.20], $P_{het} = 0.589$, $P_z = 0.789$) or homozygous model (OR = 0.80, 95% CI [0.62–1.03], $P_{het} = 0.571$, $P_z = 0.081$). In

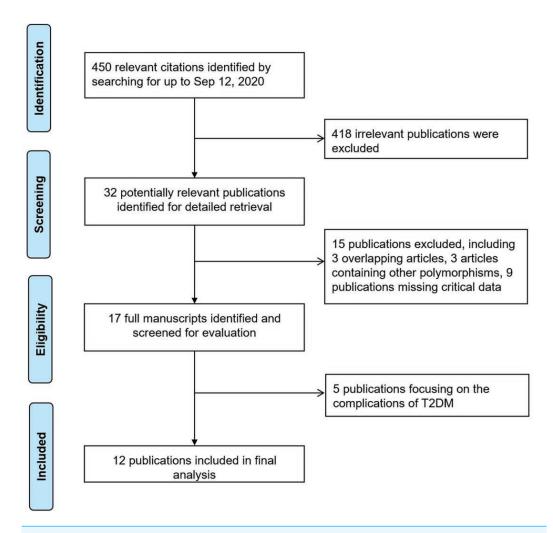


Figure 1 Flowchart of study identification process.

other ethnic groups, no association between the *IL-1B* (-511) polymorphism and T2DM risk was found. Between-study heterogeneity was found in the dominant, recessive and homozygous models ($P_{het} = 0.001$, $P_{het} < 0.001$, $P_{het} < 0.001$) (Table 3).

For the *IL-1RN* (VNTR) polymorphism, six publications were involved in the metanalysis (980 cases and 1,225 controls). Overall, a significantly increased T2DM risk was found to be associated with 2^* allele in all ethnicities (OR = 2.08, 95% CI [1.43–3.02], $P_{het} < 0.001$, $P_z < 0.001$) and in EA (OR = 2.01, 95% CI [1.53–2.66], $P_{het} = 0.541$, $P_z < 0.001$) (Table 3). Pooling data also revealed that this polymorphism is strongly associated with an increasing T2DM risk in the recessive model (OR = 1.62, 95% CI [1.09–2.42], $P_{het} = 0.377$, $P_z = 0.018$) (Table 3 and Fig. 3A) and homozygous model (OR = 2.02, 95% CI [1.07–3.83], $P_{het} = 0.085$, $P_z = 0.031$) (Table 3 and Fig. 3B), but there was no association in the dominant model (OR = 1.35, 95% CI [0.79–2.31], $P_{het} < 0.001$, $P_z = 0.275$) (Table 3). Between-study heterogeneity was found in the dominant model ($P_{het} < 0.001$) (Table 3).

Table 2 The IL-1 polymorphism distribution in cases and controls. Table 2 shows the genotypic distributions of the *IL-1B* (-511) and *IL-1RN* (*VNTR*) polymorphisms.

Gene			Case					Control	l	
IL-1B (-511)	C	T	CC	TC	TT	C	T	CC	TC	TT
Cao Yong	302	270	78	129	61	332	322	52	147	64
Liu Chang	634	532	171	292	120	385	347	101	183	82
B.R Achyut	108	292	9	90	101	175	271	29	117	77
Lin Neng-bo	302	270	85	132	69	332	322	76	180	71
Safaa I. Tayel	55	45	16	23	11	21	39	4	13	13
Natalie E. Doody	186	218	40	106	56	133	273	21	91	91
Benja Muktabhant	90	90	23	44	23	33	27	11	11	8
IL-1RN	1*	2*	1*/1*	1*/2*	2*/2*	1*	2*	1*/1*	1*/2*	2*/2*
B.R Achyut	227	234	60	107	20	340	92	138	64	14
Zhang Jian	192	34	88	16	2	360	36	163	34	1
Zhang Ping-an	192	34	88	16	2	450	44	204	42	1
Zhou Jian-zhong	107	71	36	35	1	153	41	58	37	2
Alexandra I. F. Blakemore	154	106	55	44	18	372	124	141	90	17
Petra Borilova Linhartova	569	289	220	129	31	303	121	108	87	17

Sensitivity and publication bias analysis

We next performed sensitivity analysis on the association between IL-1 β (-511) and T2DM. The results showed that Achyut's study had an influence on the pooled OR in the IL-1 β (-511)-dominant model (OR = 0.71, 95% CI [0.53–0.96]) and homozygous model (OR = 0.65, 95% CI [0.44–0.97]) (Figs. 2B, 2D) but did not affect the pooled OR in the recessive model (Fig. 2C). Achyut's study was performed in a SA population; thus, there was no influence on the pooled OR in the EA population. Due to the lack of heterogeneity in IL-1RN recessive and homozygous models, sensitivity analysis was not performed.

The results of the Harbord test and Peters test showed no publication bias for *IL-1B* (-511) in the dominant model ($P_{har} = 0.759$ and $P_{pet} = 0.881$, respectively) (Fig. 4A). Moreover, no evidence of publication bias was found for *IL-1RN* (VNTR) in the recessive ($P_{har} = 0.498$ and $P_{pet} = 0.495$, respectively) or homozygous models ($P_{har} = 0.634$ and $P_{pet} = 0.481$, respectively) (Figs. 4B, 4C).

DISCUSSION

The *IL-1B* (-511) and *IL-1RN* (VNTR) polymorphisms have been reported to be related to the pathogenesis of T2DM (*Ehses et al.*, 2009; *Masters et al.*, 2010), and many case-control studies have illustrated the association of *IL-1B* (-511) and *IL-1RN* (VNTR) polymorphisms with T2DM risk. However, each individual study may not have been powerful, causing their results to be controversial. Given the above, we conducted the present meta-analysis to derive a more precise evaluation of the association of *IL-1B* (-511) and *IL-1RN* (VNTR) polymorphisms with T2DM risk. We found that (i) the *T* allele of *IL-1B* (-511) was related with a decreased T2DM risk in the EA subgroup; (ii) significantly elevated risk of developing T2DM was observed to be associated with the 2* allele in all ethnicities and

Table 3 Meta-analysis of the relationship of the IL-1 polymorphisms with the risks of T2DM.

Gene	No. studies	Allele	comparison		Genetic model comparison				
		OR (95% CI)	P_{het}	P_z	OR (9	P_{het}	P_z		
IL-1B (-511)									
Total	7	0.89 (0.68–1.17)	< 0.001	0.406	Dominant	0.84 (0.57–1.25)	0.001	0.395	
					Recessive	0.89 (0.61–1.31)	< 0.001	0.561	
					Overdominant	0.92 (0.74–1.14)	0.065	0.422	
					Codominant	0.85 (0.6–1.21)	0.011	0.375	
					Codominant#	0.96 (0.69-1.34)	0.003	0.803	
					Homozygote	0.82 (0.48-1.39)	< 0.001	0.451	
EA	3	0.89 (0.79-1.01)	0.606	0.072	Dominant	0.76 (0.59-0.97)	0.218	0.027	
					Recessive	0.97 (0.79-1.20)	0.589	0.789	
					Overdominant	0.82 (0.64-1.04)	0.162	0.094	
					Codominant	0.73 (0.54-0.99)	0.141	0.040	
					Codominant#	1.07 (0.86–1.34)	0.385	0.531	
					Homozygote	0.80 (0.62-1.03)	0.571	0.081	
SA	2	1.00 (0.33-2.98)	< 0.001	0.997	Dominant	1.20 (0.18–7.84)	< 0.001	0.853	
SEA	1	1.22 (0.68–2.20)	_	0.503	Dominant	1.69 (0.70-4.07)	_	0.245	
NA	1	0.44 (0.23-0.85)	_	0.015	Dominant	0.33 (0.10-1.10)	_	0.070	
IL-1RN									
Total	6	2.08 (1.43-3.02)	< 0.001	< 0.001	Dominant	1.35 (0.79–2.31)	< 0.001	0.275	
					Recessive	1.62 (1.09–2.42)	0.377	0.018	
					Overdominant	1.20 (0.72–1.99)	< 0.001	0.485	
					Codominant	1.28 (0.73-2.24)	< 0.001	0.398	
					Codominant#	1.38 (0.89–2.13)	0.351	0.155	
					Homozygote	2.02 (1.07-3.83)	0.085	0.031	
EA	3	2.02 (1.53–2.66)	0.541	< 0.001	Dominant	1.11 (0.78–1.59)	0.525	0.558	
					Recessive	2.30 (0.57–9.25)	0.470	0.243	
					Overdominant	1.05 (0.72–1.54)	0.337	0.790	
					Codominant	1.06 (0.74–1.53)	0.371	0.749	
					Codominant#	2.25 (0.53–9.55)	0.356	0.270	
					Homozygote	2.42 (0.60–9.78)	0.554	0.216	
SA	1	3.81 (2.84–5.11)	_	<0.001	Dominant	3.75 (2.48–5.67)	_	< 0.001	
Caucasian	2	1.60 (0.998–2.58)	0.02	0.051	Dominant	1.04 (0.54–2.02)	0.017	0.901	

Notes.

CI, confidence interval; IL-1, interleukin-1; OR, odds ratio; P_{het} , P-value for heterogeneity; P_z , P-value for overall effect; SA, South Asian; SEA, South East Asian; EA, East Asian; NA, North Africa.

For IL1B (-511) polymorphism: dominant (TT + TC vs CC), recessive (TT vs TC + CC), overdominant (CT vs CC + TT), codominant (CT vs CC), codominant (TT vs TT) and homozygote (TT vs TT).

For IL1RN (VNTR) polymorphism: dominant (2*2* + 2*1* vs 1*1*), recessive (2*2* vs 2*1* + 1*1*), overdominant (2*1* vs 1*1* + 2*2*), codominant (2*1* vs 1*1* + 2*2*), codominant (2*1* vs 1*1* + 2*2*) and homozygote (2*2* vs 1*1*).

EA subgroup, and its carriers of the *IL-1RN* (VNTR) polymorphism had an increased risk of developing T2DM among all ethnicities; (iii) compared to the 1*/1* homozygotes, the 2*/2* homozygotes of the *IL-1RN* (VNTR) polymorphism had an increased risk of T2DM development; and (iv) no significant association was observed between the *IL-1RN* (VNTR) polymorphism and T2DM risk in the EA subgroup. The results of sensitivity

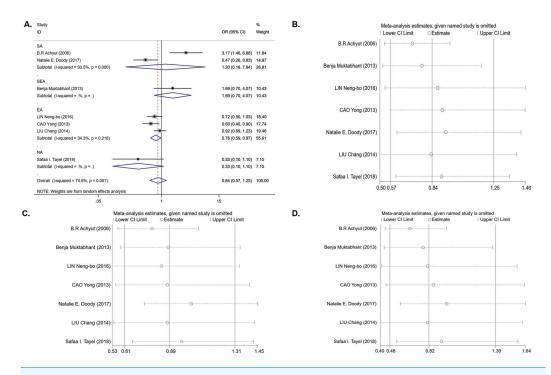
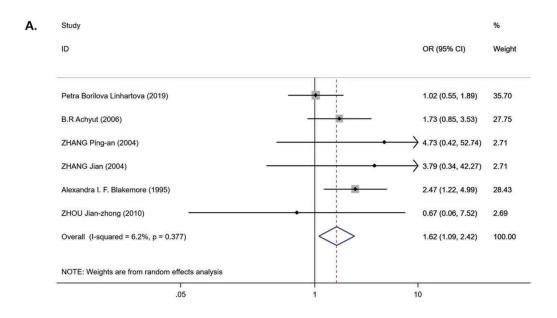


Figure 2 An overall forest plot of IL-1B (-511) associated with T2DM risk in the dominant model (TT + TC vs CC) and sensitivity analyses. (A) Forest plot of IL-1B (-511) associated with T2DM in the dominant model; (SA: South Asian, SEA: South East Asian, EA: East Asian, NA: North Africa). (B) Sensitivity analysis for TT + TC vs CC associated with T2DM in the overall meta-analysis. (C) Sensitivity analysis for TT vs TC + CC associated with T2DM in the overall meta-analysis. (D) Sensitivity analysis for TT vs TC associated with T2DM in the overall meta-analysis.

analysis showed that Achyut's study affected the pooled ORs in the IL-1 β (-511) dominant and homozygous models. However, Achyut's study did not influence the pooled OR in the EA subgroup after stratification by ethnicity. Therefore, the result of the association between *IL-1B* (-511) and the decreased risk of T2DM in the dominant model in the EA subgroup was convincing.

First, IL-1 β was considered a mediator of fever, and rapidly, it was found that IL-1 β induced innate immunity to defend against pathogens. However, chronic overexpression of IL-1 β has been related to multiple immune diseases, including T2DM (*Mandrup-Poulsen*, 1996). Eventually, it was suggested that IL-1 β can induce the inflammatory microenvironment of islets, leading to impaired insulin secretion, decreased cell proliferation and apoptosis of β -cells, ultimately contributing to the development of T2DM (*Rhodes*, 2005). Donath, M Y et al. extensively described the development of islet inflammation in GK rats, suggesting that the expression of pro-inflammatory cytokines IL-1 β and others (IL-6, TNF α) was increased in islets, while elevated expression levels of many chemokines (CXCL1/KC, MCP-1, MIP-1 α) and infiltration of immune cells in islets were observed (*Ehses et al.*, 2009). In all cases, macrophage infiltration is increased in the islet inflammatory process (*Ehses et al.*, 2007). Moreover, two independent studies have now indicated that the numbers of islet-related CD68+ cells are increased in T2DM patients



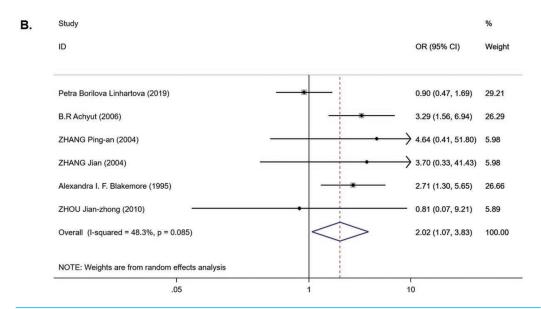


Figure 3 Forest plot of meta-analysis on IL-1RN (VNTR) and the risk of T2DM in the recessive model (2*2* vs 2*1* + 1*1*) and homozygous model (2*2* vs 1*1*). (A) Forest plot of IL-1RN (VNTR) associated with T2DM in the recessive model. (B) Forest plot of IL-1RN (VNTR) associated with T2DM in the homozygous model.

(Ehses et al., 2007; Richardson et al., 2009). In addition, data obtained from laser-captured β -cells from T2DM patients have shown that evaluated expression levels of IL-1 β and chemokines possibly led to this immune cell infiltration (Böni-Schnetzler et al., 2008). Since it can regulate various inflammatory processes, any change in the level of IL-1 β in blood or tissue possibly affects these processes. Larsen et al. suggested that blocking IL-1 β is a potential therapy in the treatment of T2DM (Larsen et al., 2007). The IL-1B (-511) C

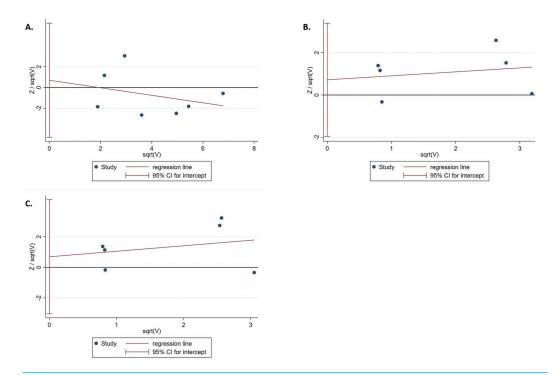


Figure 4 Harbord test for publication bias testing. (A) Publication bias of IL-1B (-511) associated with T2DM in the dominant model ($TT + TC \ vs \ CC$); (B) Publication bias of IL-1RN (VNTR) associated with T2DM in the recessive model ($2^*2^* \ vs \ 2^*1^* + 1^*1^*$); (C) Publication bias of IL-1RN (VNTR) associated with T2DM in the homozygous model($2^*2^* \ vs \ 1^*1^*$).

allele is associated with higher expression of IL-1 β and with severe inflammation in the liver, while the (-511) T allele is associated with lower levels of IL-1 β (*Hirankarn et al.*, 2006; *Vishnoi et al.*, 2008). In this meta-analysis, we found that the *IL-1B* (-511) T allele had a decreased T2DM risk in the EA subgroup, suggesting an influence of the *IL-1B* (-511) polymorphism system in this ethnic group.

The *IL-1RN* 2* allele (two repeats) is associated with increased IL-1Ra levels (*Danis et al.*, 1995), which can compete with an inhibitor of IL-1β by binding to the IL-1 receptor. The 2* allele significantly increases the IL-1β secretion *in vitro* and balances the expression of IL-1β and IL-1Ra (*Santtila, Savinainen & Hurme, 1998*), and their ratio (IL-1Ra/IL-1β) determines the severity of inflammation. It has been shown that the *IL-1RN* 2* allele is associated with a low ratio (IL-1Ra/IL-1β), thereby inducing a longer and more severe proinflammation (*Witkin, Gerber & Ledger, 2002*). The frequency of the 2* allele is increased in inflammatory conditions or autoimmune diseases (*Tountas et al., 1999*; *Van der Paardt et al., 2002*). Consistently, this meta-analysis suggested that the *IL-1RN* 2* allele and 2*2* homozygotes increased T2DM risk, which indicated that the *IL-1RN* 2* allele and 2*2* homozygote polymorphism play critical roles in the development of T2DM.

Although we have devoted considerable efforts and resources to testing the potential association of the *IL-1B* (-511) and *IL-1RN* (VNTR) polymorphisms with T2DM risk, this analysis still has some limitations. First, significant heterogeneity was found in pooled

analyses between the *IL-1B* (*-511*) polymorphism and T2DM risk. Due to the limited number of included publications, we conducted further ethnic stratification analysis and meta-regression to identify the exact sources of between-study heterogeneity. The meta-regression results showed that ethnicity was not the source of heterogeneity (both values of tau² were not much different (0.34 *vs* 0.40)). Some possible relevant factors (genotyping method, sex) may lead to heterogeneity. However, subgroup analysis can significantly reduce heterogeneity. Second, the literature articles were primarily published in English or Chinese; thus, some eligible publications may be missing, causing some bias. Third, the publications included have some confounders, such as age, sex, diet, and exercise, whereas we used only the raw data to conduct pooled analysis; therefore, we were unable to control these possible confounders or test the potential gene-environment interactions. Fourth, the number of included publications was limited; thus, additional studies with large sample sizes and a wider variety of ethnicities are needed in the future to evaluate the association.

In conclusion, our meta-analysis first indicated that the IL-1B (-511) T allele polymorphism is associated with decreased T2DM risk in the EA population and that the IL-1RN 2* allele and 2*2 * homozygote polymorphism are strongly associated with increased T2DM risk. Further well-designed studies including different ethnicities with large sample sizes are needed to verify this conclusion.

Abbreviations

T2DM type 2 diabetes mellitus

IL-1 Interleukin-1

IL-1Ra IL-1 receptor antagonist

IL-1β interleukin-1β

HWE Hardy–Weinberg Equilibrium

OR odds ratios

CIs confidence intervals

EA East Asian

ADDITIONAL INFORMATION AND DECLARATIONS

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Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Juan Jiao and Zhaoping Wang conceived and designed the experiments, performed the
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 drafts of the paper, and approved the final draft.
- Yanfei Guo, Xiuqing Huang, Liang Sun, Xiaoquan Zhu and Qi Zhou analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Jie Liu and Ze Yang conceived and designed the experiments, authored or reviewed drafts of the paper, and approved the final draft.
- Xiaolin Ni and Danni Gao performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Huiping Yuan conceived and designed the experiments, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

The raw data is available in Tables 1 and 2.

Supplemental Information

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REFERENCES

Achyut BR, Srivastava A, Bhattacharya S, Mittal B. 2007. Genetic association of interleukin-1beta (-511C/T) and interleukin-1 receptor antagonist (86 bp repeat) polymorphisms with Type 2 diabetes mellitus in North Indians. *Clinica Chimica Acta* 377:163–169 DOI 10.1016/j.cca.2006.09.012.

Aday AW, Ridker PM. 2018. Antiinflammatory therapy in clinical care: the CANTOS trial and beyond. *Frontiers in Cardiovascular Medicine* **5**:62 DOI 10.3389/fcvm.2018.00062.

- Blakemore AI, Cox A, Gonzalez AM, Maskil JK, Hughes ME, Wilson RM, Ward JD, Duff GW. 1996. Interleukin-1 receptor antagonist allele (IL1RN*2) associated with nephropathy in diabetes mellitus. *Human Genetics* 97:369–374 DOI 10.1007/bf02185776.
- Borilova Linhartova P, Poskerova H, Tomandlova M, Bartova J, Kankova K, Fassmann A, Izakovicova Holla L. 2019. Interleukin-1 gene variability and plasma levels in Czech patients with chronic periodontitis and diabetes mellitus. *International Journal of Dentistry* 2019:6802349 DOI 10.1155/2019/6802349.
- **Böni-Schnetzler M, Donath MY. 2013.** How biologics targeting the IL-1 system are being considered for the treatment of type 2 diabetes. *British Journal of Clinical Pharmacology* **76**:263–268 DOI 10.1111/j.1365-2125.2012.04297.x.
- Böni-Schnetzler M, Thorne J, Parnaud G, Marselli L, Ehses JA, Kerr-Conte J, Pattou F, Halban PA, Weir GC, Donath MY. 2008. Increased interleukin (IL)-1beta messenger ribonucleic acid expression in beta -cells of individuals with type 2 diabetes and regulation of IL-1beta in human islets by glucose and autostimulation. *Journal of Clinical Endocrinology and Metabolism* 93:4065–4074 DOI 10.1210/jc.2008-0396.
- Cao Y, Wan Q, Tang XP, Song XQ, Chen Z, Chen F. 2013. The relationship between interleukin -1β- 511 (C/T) gene polymorphism and type 2 diabetes mellitus. *Immunological Journal* 29:503–506.
- **Danis VA, Millington M, Hyland VJ, Grennan D. 1995.** Cytokine production by normal human monocytes: inter-subject variation and relationship to an IL-1 receptor antagonist (IL-1Ra) gene polymorphism. *Clinical and Experimental Immunology* **99**:303–310 DOI 10.1111/j.1365-2249.1995.tb05549.x.
- **DerSimonian R, Laird N. 1986.** Meta-analysis in clinical trials. *Controlled Clinical Trials* 7:177–188 DOI 10.1016/0197-2456(86)90046-2.
- **Dinarello CA. 2000.** The role of the interleukin-1-receptor antagonist in blocking inflammation mediated by interleukin-1. *New England Journal of Medicine* **343**:732–734 DOI 10.1056/nejm200009073431011.
- Dinarello CA, Donath MY, Mandrup-Poulsen T. 2010. Role of IL-1 beta in type 2 diabetes. *Current Opinion in Endocrinology Diabetes and Obesity* 17:314–321 DOI 10.1097/MED.0b013e32833bf6dc.
- **Donath MY, Halban PA. 2004.** Decreased beta-cell mass in diabetes: significance, mechanisms and therapeutic implications. *Diabetologia* **47**:581–589 DOI 10.1007/s00125-004-1336-4.
- Doody NE, Dowejko MM, Akam EC, Cox NJ, Bhatti JS, Singh P, Mastana SS. 2017. The Role of TLR4, TNF- α and IL-1 β in Type 2 Diabetes Mellitus development within a North Indian population. *Annals of Human Genetics* **81**:141–146 DOI 10.1111/ahg.12197.
- **Duan L, Xue Z, Ji H, Zhang D, Wang Y. 2018.** Effects of CYP2R1 gene variants on vitamin D levels and status: a systematic review and meta-analysis. *Gene* **678**:361–369 DOI 10.1016/j.gene.2018.08.056.
- Ehses JA, Lacraz G, Giroix MH, Schmidlin F, Coulaud J, Kassis N, Irminger JC, Kergoat M, Portha B, Homo-Delarche F, Donath MY. 2009. IL-1 antagonism reduces

- hyperglycemia and tissue inflammation in the type 2 diabetic GK rat. *Proceedings of the National Academy of Sciences of the United States of America* **106**:13998–14003 DOI 10.1073/pnas.0810087106.
- Ehses JA, Perren A, Eppler E, Ribaux P, Pospisilik JA, Maor-Cahn R, Gueripel X, Ellingsgaard H, Schneider MK, Biollaz G, Fontana A, Reinecke M, Homo-Delarche F, Donath MY. 2007. Increased number of islet-associated macrophages in type 2 diabetes. *Diabetes* 56:2356–2370 DOI 10.2337/db06-1650.
- **Fève B, Bastard JP. 2009.** The role of interleukins in insulin resistance and type 2 diabetes mellitus. *Nature Reviews Endocrinology* **5**:305–311 DOI 10.1038/nrendo.2009.62.
- Hirankarn N, Kimkong I, Kummee P, Tangkijvanich P, Poovorawan Y. 2006.

 Interleukin-1 beta gene polymorphism associated with hepatocellular carcinoma in hepatitis B virus infection. *World Journal of Gastroenterology* 12:776–779

 DOI 10.3748/wjg.v12.i5.776.
- **Hivert MF, Vassy JL, Meigs JB. 2014.** Susceptibility to type 2 diabetes mellitus–from genes to prevention. *Nature Reviews Endocrinology* **10**:198–205 DOI 10.1038/nrendo.2014.11.
- **Hotamisligil GS. 2006.** Inflammation and metabolic disorders. *Nature* **444**:860–867 DOI 10.1038/nature05485.
- **Hotamisligil GS. 2017.** Inflammation, metaflammation and immunometabolic disorders. *Nature* **542**:177–185 DOI 10.1038/nature21363.
- Kristiansen OP, Pociot F, Johannesen J, Bergholdt R, Dinarello CA, Nerup J, Mandrup-Poulsen T. 2000. Linkage disequilibrium testing of four interleukin-1 gene-cluster polymorphisms in Danish multiplex families with insulin-dependent diabetes mellitus. *Cytokine* 12:171–175 DOI 10.1006/cyto.1999.0546.
- Larsen CM, Faulenbach M, Vaag A, Vølund A, Ehses JA, Seifert B, Mandrup-Poulsen T, Donath MY. 2007. Interleukin-1-receptor antagonist in type 2 diabetes mellitus. *New England Journal of Medicine* 356:1517–1526 DOI 10.1056/NEJMoa065213.
- **Lin NB, Tang XP, Cao Y, Wan Q. 2016.** Association between interleukin-1β-511 (C/T) gene polymorphism and early diabetic kidney disease. *Chinese Journal of Diabetes* 407–411 DOI 10.3969/j.issn.1006-6187.2016.05.005.
- Little J, Higgins JP, Ioannidis JP, Moher D, Gagnon F, von Elm E, Khoury MJ, Cohen B, Davey-Smith G, Grimshaw J, Scheet P, Gwinn M, Williamson RE, Zou GY, Hutchings K, Johnson CY, Tait V, Wiens M, Golding J, Van Duijn C, McLaughlin J, Paterson A, Wells G, Fortier I, Freedman M, Zecevic M, King R, Infante-Rivard C, Stewart A, Birkett N. 2009. Strengthening the reporting of genetic association studies (STREGA): an extension of the STROBE statement. *PLOS Medicine* 6:e22 DOI 10.1371/journal.pmed.1000022.
- **Liu C, Wang YX, Wu ZX, Zhao Y, Chi XM. 2014.** Study on the correlation between IL-1β Rs16944 gene polymorphism and Type 2 Diabetes Mellitus in Han People of Liaoning Province. *Journal of Clinical Research* **3**:426–428 DOI 10.3969/j.issn.1671-7171.2014.03.004.

- **Lo CK, Mertz D, Loeb M. 2014.** Newcastle-Ottawa Scale: comparing reviewers' to authors' assessments. *BMC Medical Research Methodology* **14**:45 DOI 10.1186/1471-2288-14-45.
- **Mandrup-Poulsen T. 1996.** The role of interleukin-1 in the pathogenesis of IDDM. *Diabetologia* **39**:1005–1029 DOI 10.1007/bf00400649.
- Masters SL, Dunne A, Subramanian SL, Hull RL, Tannahill GM, Sharp FA, Becker C, Franchi L, Yoshihara E, Chen Z, Mullooly N, Mielke LA, Harris J, Coll RC, Mills KHG, Mok KH, Newsholme P, Nunez G, Yodoi J, Kahn SE, Lavelle EC, O'Neill LAJ. 2010. Activation of the NLRP3 inflammasome by islet amyloid polypeptide provides a mechanism for enhanced IL-1 beta in type 2 diabetes. *Nature Immunology* 11:897–U1501 DOI 10.1038/ni.1935.
- **Moher D, Liberati A, Tetzlaff J, Altman DG. 2009.** Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLOS Medicine* **6**:e1000097 DOI 10.1371/journal.pmed.1000097.
- Muktabhant B, Duangjinda M, Sanchaisuriya P, Kananit S, Schelp FP. 2013. Association of the polymorphism of interleukin-1B (-511C/T) with type 2 diabetes mellitus patients with and without family history of a first degree relative with diabetes mellitus. *International Journal of Diabetes in Developing Countries* 33:236–240 DOI 10.1007/s13410-013-0155-x.
- **Qamar A, Rader DJ. 2012.** Effect of interleukin 1β inhibition in cardiovascular disease. *Current Opinion in Lipidology* **23**:548–553 DOI 10.1097/MOL.0b013e328359b0a6.
- **Rhodes CJ. 2005.** Type 2 diabetes-a matter of beta-cell life and death? *Science* **307**:380–384 DOI 10.1126/science.1104345.
- Richardson SJ, Willcox A, Bone AJ, Foulis AK, Morgan NG. 2009. Islet-associated macrophages in type 2 diabetes. *Diabetologia* 52:1686–1688 DOI 10.1007/s00125-009-1410-z.
- Rissanen A, Howard CP, Botha J, Thuren T. 2012. Effect of anti-IL-1β antibody (canakinumab) on insulin secretion rates in impaired glucose tolerance or type 2 diabetes: results of a randomized, placebo-controlled trial. *Diabetes, Obesity and Metabolism* 14:1088–1096 DOI 10.1111/j.1463-1326.2012.01637.x.
- Ruscitti P, Masedu F, Alvaro S, Airò P, Battafarano N, Cantarini L, Cantatore FP, Carlino G, D'Abrosca V, Frassi M, Frediani B, Iacono D, Liakouli V, Maggio R, Mulè R, Pantano I, Prevete I, Sinigaglia L, Valenti M, Viapiana O, Cipriani P, Giacomelli R. 2019. Anti-interleukin-1 treatment in patients with rheumatoid arthritis and type 2 diabetes (TRACK): a multicentre, open-label, randomised controlled trial. *PLOS Medicine* 16:e1002901 DOI 10.1371/journal.pmed.1002901.
- **Santtila S, Savinainen K, Hurme M. 1998.** Presence of the IL-1RA allele 2 (IL1RN*2) is associated with enhanced IL-1beta production in vitro. *Scandinavian Journal of Immunology* **47**:195–198 DOI 10.1046/j.1365-3083.1998.00300.x.
- **Sathyapalan T, Atkin SL. 2011.** Is there a role for immune and anti-inflammatory therapy in type 2 diabetes? *Minerva Endocrinologica* **36**:147–156.
- Spranger J, Kroke A, Möhlig M, Hoffmann K, Bergmann MM, Ristow M, Boeing H, Pfeiffer AF. 2003. Inflammatory cytokines and the risk to develop type 2 diabetes:

- results of the prospective population-based European Prospective Investigation into Cancer and Nutrition (EPIC)-Potsdam Study. *Diabetes* **52**:812–817 DOI 10.2337/diabetes.52.3.812.
- Tarlow JK, Blakemore AI, Lennard A, Solari R, Hughes HN, Steinkasserer A, Duff GW. 1993. Polymorphism in human IL-1 receptor antagonist gene intron 2 is caused by variable numbers of an 86-bp tandem repeat. *Human Genetics* 91:403–404 DOI 10.1007/bf00217368.
- Tayel SI, Fouda EAM, Elshayeb EI, Eldakamawy ARA, El-Kousy SM. 2018. Biochemical and molecular study on interleukin-1β gene expression and relation of single nucleotide polymorphism in promoter region with Type 2 diabetes mellitus. *Journal of Cellular Biochemistry* 119:5343–5349 DOI 10.1002/jcb.26667.
- Tountas NA, Casini-Raggi V, Yang H, Di Giovine FS, Vecchi M, Kam L, Melani L, Pizarro TT, Rotter JI, Cominelli F. 1999. Functional and ethnic association of allele 2 of the interleukin-1 receptor antagonist gene in ulcerative colitis. *Gastroenterology* 117:806–813 DOI 10.1016/s0016-5085(99)70338-0.
- Van der Paardt M, Crusius JB, García-González MA, Baudoin P, Kostense PJ, Alizadeh BZ, Dijkmans BA, Peña AS, van der Horst-Bruinsma IE. 2002. Interleukin-1beta and interleukin-1 receptor antagonist gene polymorphisms in ankylosing spondylitis. *Rheumatology* 41:1419–1423 DOI 10.1093/rheumatology/41.12.1419.
- Vishnoi M, Pandey SN, Choudhuri G, Mittal B. 2008. IL-1 gene polymorphisms and genetic susceptibility of gallbladder cancer in a north Indian population. *Cancer Genetics and Cytogenetics* 186:63–68 DOI 10.1016/j.cancergencyto.2008.05.004.
- Witkin SS, Gerber S, Ledger WJ. 2002. Influence of interleukin-1 receptor antagonist gene polymorphism on disease. *Clinical Infectious Diseases* 34:204–209 DOI 10.1086/338261.
- **Zhang PA, Li Y, Xu P, Wen ZY. 2004b.** Study on the correlation of Interleukin-1 Receptor antagonist genotype with NIDDM. *Journal of Clinical Research* 870–873.
- **Zhang J, Xiao Q, Li WN. 2004a.** Study of Interleukin-1 Receptor antagonist gene polymorphism in type 2 Diabetes mellitus. *Journal of Laboratory Medicine* 15–17.
- **Zhou JZ, Zhou Y, Zhong WB, Li HW. 2010.** Study on correlation between interleukin-1 receptor antagonist gene polymorphism and type 2 diabetes. *Clinical Journal of Medical Officers* **38**:235–237 DOI 10.3969/j.issn.1671-3826.2010.02.028.