Review Article

HMBG1 as a Driver of Inflammatory and Immune Processes in the Pathogenesis of Ocular Diseases

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High-mobility group box 1 (HMGB1) is a nuclear protein that can also act as an extracellular trigger of inflammation, proliferation, and migration in eye diseases. It induces signaling pathways by binding to the receptor for advanced glycation end products (RAGE) and Toll-like receptors (TLRs) 2, 4, and 9. This proinflammatory activity is considered to be important in the pathogenesis of a wide range of ocular diseases resulting from hemodynamic changes, presence of neovascular endothelial cells, secretion of intraocular immune factors or inflammation, and apoptosis of retinal cell layers. Further work is needed to elucidate in detail how HMGB1 contributes to ocular disease and how its damaging activity can be modulated. In this review, we summarize current knowledge on HMGB1 as a ligand that can evoke inflammation and immune responses in ocular diseases.

1. Introduction

The eye is a relatively independent and closed system in which various tissues and cells interact in complex ways [1]. Many eye diseases have been described, such as glaucoma, keratitis, diabetic retinopathy, and age-related macular degeneration, and these diseases often arise as a result of pathogenic immune responses against eye autoantigens or as a local manifestation of a systemic autoimmune response. Although development of new drugs and innovations in surgical methods has led to effective treatment of ocular diseases, how these diseases occur and progress remains poorly understood. Several studies indicate that ocular diseases have a close relationship with autoimmune reactions and inflammatory responses [2-5], and a key protein in such processes is high-mobility group box 1 (HMGB1) [6-8]. The present review examines the role of HMGB1 in eve diseases.

HMGB1 is a DNA-binding nuclear protein discovered over 30 years ago [9–11]. Normally, this protein exists in the cell nucleus; when it is released outside the cell, it becomes an immune-inflammatory factor [12–14]. It can be released from damaged cells or secreted by activated immune cells such as macrophages, dendritic cells (DCs), and natural killer cells [15–18]. It may induce signaling pathways by binding to the receptor for advanced glycation end products (RAGE) and Toll-like receptors (TLRs) 2, 4, and 9 [19, 20].

HMGB1 is closely related to many inflammatory diseases, such as ischemia of liver and kidney, hepatitis, arthritis, stroke, ischemia of liver and kidney, sepsis, and systemic lupus erythematosus [21–25]. In this review, our focus is the important role of HMGB1 in inflammatory immune eye diseases, including keratitis, uveitis, dry eye, diabetic retinopathy, and retinal degeneration. Inhibition of this protein may be an effective new treatment for patients with immune-inflammatory eye disease. We suggest that more research with both animal and human models is needed to confirm that HMGB1 has the therapeutic potential suggested by initial studies.

2. HMGB1 Structure

HMGB1 was identified as a nonhistone chromatin-binding protein about 30 years ago [9–11]. This protein is abundantly

expressed in nearly all eukaryotic cells [26]. It acts as a damage-associated molecular pattern (DAMP) molecule [27]. HMGB1 is a member of the high-mobility group (HMG) chromosomal protein family [28]. HMG chromosomal proteins are divided into the three superfamilies HMGB, HMGN, and HMGA [29]. Human HMGB1 is an alarmin encoded by a single gene that is located on chromosome 13q12 [30]. In most cell types, HMGB1 is mainly located in the nucleus under physiological conditions. When cells are stimulated or suffer injury or death, HMGB1 translocates from inside to outside the cell [31, 32]. HMGB1 contains 215 amino acids arranged into two DNA-binding domains (HMG A box and HMG B box) and one C-terminal acidic tail, which contains a stretch of approximately 30 glutamic and aspartic acid residues [27, 33]. The HMG A and B boxes can bind to DNA and participate in DNA folding and twisting [34-36]. The HMG B box causes macrophages to secrete additional proinflammatory cytokines; in fact, the B box on its own can trigger the same processes as full-length HMGB1 [27, 37]. The A box can antagonize cytokine activity [38, 39]. The spatial arrangement of A and B boxes was regulated by C-terminal acidic tail. [10, 40, 41].

Each HMGB1 domain interacts with different receptors, and these interactions regulate the biological activity of extracellar HMGB1. TLR4 is binding with residues 89–108 of HMGB1 [42, 43], while RAGE is binding with residues 150–184 of HMGB1 [44]. Within the C-terminal acidic tail (residues 186–215), residues 201–205 exert antiinflammatory activity [45]. Two nuclear localization sequences in HMGB1 may stabilize the chromatin structure and regulate gene transcription.

3. Expression of HMGB1 in Eye Disease

Numerous studies suggest that HMGB1 may contribute to eye disease by acting as an inflammatory cytokine. In a model of retinal ischemia reperfusion injury and inflammation, HMGB1 expression is upregulated in retinal pigment epithelium, retinal endothelial cells, ganglion cells, Müller cells, astrocytes, and photoreceptors [46]. Chronic epithelial cell damage to the cornea or lacrimal glands triggers HMGB1 secretion, which initiates an inflammatory cycle [47]. HMGB1 levels are elevated in the cytoplasm and extracellular space in eye disease. All these studies indicate that HMGB1 is abundantly expressed in ocular diseases, which reflects its role in contributing to these disorders.

4. HMGB1 and Diabetic Retinopathy

Dysfunction of retinal endothelial cells, damage to the blood-retinal barrier, ischemia, and retinal neo-vascularization are the characters of diabetic retinopathy at different stages [48]. Inflammation factor-mediated damage to the retina and optic nerve are key factors in the pathogenesis of diabetic retinopathy, which appears to involve continuous, low-level inflammation [49]. HMGB1 helps drive this process by acting as a late inflammatory factor [50].

In patients with proliferative diabetic retinopathy (PDR), HMGB1 levels in vitreous humor increased significantly [51]. The expression level of HMGB1 in vitreous body of patients with active PDR is significantly higher than that of inactive PDR patients [52]. HMGB1 may interact with vascular adhesion protein 1 (VAP-1), 8-hydroxy-2'-deoxyguanosine (8-OHdG), and heme oxygenase-1 (HO-1) in contributing to the pathogenesis of inflammation and angiogenesis in PDR [53].

HMGB1 is involved in the pathogenesis of diabetic retinopathy through the combination of RAGE and TLR9 [53]. In BV2 cells, a signaling pathway mediated by HMGB1, RAGE, TLR4, p38, and nuclear factor kappa light chain enhancer of activated B cells (NF- κ B) activates matrix metalloprotease-9 in response to high glucose levels [54]. In RGC-5 cells, short interfering RNA against HMGB1 obviously downregulates expression of TLR4 and NF- κ B mRNA in response to high glucose [55]. Treating human retinal endothelial cells with high glucose activates HMGB1 to trigger signaling by ERK1/2 and NF- κ B, which in turn induces inflammatory responses and break vascular barrier of retinal [56].

The presence of diabetes in rats or intravitreal injection of HMGB1 in normal rats significantly upregulates the levels of mRNAs encoding SDF-1/CXCL12 receptor CXCR4, as well as levels of HIF-1, EGR-1, TK-2, and CXCL12/CXCR4 proteins in the chemokine axis [57]. In diabetic retinopathy, HMGB1 is closely related not only to the apoptosis of nerve cells but also to neovascularization; the latter involves STAT-3 as a downstream effector of HMGB1 to drive angiogenesis [58]. In human retinal microvascular endothelial cells, HMGB1 significantly upregulates levels of IL-1 β and reactive oxygen species, as well as expression of NADPH oxidase 2 (Nox2), caspase-3, and PARP-1 [59]. HMGB1 exerts these proinflammatory and angiogenic effects via TNF α and VEGF [57].

Together, these studies demonstrate that HMGB1 helps to drive diabetic retinopathy in two ways. HMGB1 binds to TLR4 and RAGE to trigger nerve cell death. At the same time, HMGB1 triggers release of proangiogenic factors to induce pathological neovascularization (Figure 1).

5. HMGB1 and Uveitis

Inflammation of the iris, ciliary body, vitreous body, retina, or choroid is a typical feature of uveitis. It refers to a group of intraocular inflammatory diseases that arise without a clear infectious trigger and is a leading cause of visual impairment and legal blindness, particularly in the younger population. It contributes to 10–15% of all cases of blindness (defined as central visual acuity of 1/10 or less in the better eye) in developed countries. Such blindness arises mainly as a result of macular edema, ocular hypertonia, or retinal ischemia.

Topical and systemic anti-inflammatory and immunosuppressive medications can completely eliminate inflammatory signs such as keratic precipitates, aqueous flare, miosis, cortical (equatorial) cataracts, vitreal opacification, and lesions of the fundus or optic nerve head. Inflammation can recur after such therapy.

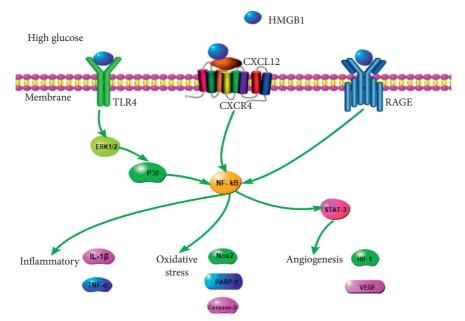


FIGURE 1: HMGB1 signal by binding to TLR4, CXCR4, and RAGE to moderate BV2 cells, RGC cells, retinal endothelial cells, and retinal microvascular endothelial cells function in PDR. In these cells, HMGB1 binds TLR4 and activates ERK1/2-P38-NF- κ B signal pathway. HMGB1 also binds RAGE and upregulated the expression of NF- κ B. In human retinal microvascular endothelial cells, HMGB1 binds CXCR4 to activate NF- κ B/STAT-3 signal pathway. These three signaling has been implicated in cell inflammatory, oxidative stress, and angiogenesis.

Uveitis is closely related to the immune response associated with T cells. Indeed, experimental autoimmune uveitis is induced by autogenous reactive T-cell injection in rodents [60]. linked to the ability of HMGB1 to activate the NF- κ B signaling pathway and thereby stimulate the secretion of inflammatory cytokines (Figure 2).

6. HMGB1 and AMD

In models of experimental autoimmune uveoretinitis, RAGE is upregulated in inflammatory cells, including macrophages infiltrating the anterior chamber, vitreous cavity, retina, subretinal space, and choroid [61]. HMGB1 activates RAGE, promoting the release of TNF- α , ICAM-1, VCAM-1, and IL-6. HMGB1 also induces CXCL12 release by infiltrating effector T cells [62]. Release of intraocular HMGB1 and CXCL12 at one day after cell transfer may attract inflammatory T cells [63]. Elevated HMGB1 levels may subsequently be associated with the participation of T cells and the retina lesion [62]. Autoimmune uveitis is activated by IRBP-specific T cells [64]. These kinds of T cells were interacted with substantive cells including Müller cells, DCs, astrocytes, and microglia. These cells produce HMGB1 via Fas/FasL-mediated signaling. RIP2 is also involved in HMGB1 release which is deeply related with Fas in uveitis [65]. This suggests the active involvement of the HMGB1-Fas-RIP2 axis in uveitis.

HMGB1 plays an important role in Behcet's disease, one of the forms of uveitis. Although this disease involves elevated HMGB1 levels in serum, these serum levels do not appear to be associated with disease activity or specific organ damage [66]. Neutrophils help drive Behcet's disease, and expression of proinflammatory cytokines such as TNF- α , IFN- γ , and IL-6 in neutrophils is enhanced by extracellular HMGB1, which acts via the NF- κ B pathway [6]. These findings suggest that pathogenesis of Behcet's disease is AMD is a disease of macular degeneration, which is the main cause of irreversible central vision loss in the elderly in developed countries [67]. This vision loss is thought to involve degeneration of retinal pigment epithelial cells and the overlying photoreceptors, which rely on the retinal pigment epithelium for trophic support.

In a culture model of AMD in which retinal pigment epithelial cells are treated with NaIO₃, HMGB1 is released from the nucleus [68]. HMGB1 is also released from the nucleus and secreted by retinal pigment epithelial cells in response to oxidative stress [69]. Depleting HMGB1 from the culture medium inhibits the induction of TNF- α production by retinal pigment epithelial cells [70]. Treating wild-type retinal pigment epithelial cells with poly(I:C) and Z-VAD induces the secretion of substantial amounts of HMGB1, which is completely blocked by deleting the Rip3 gene from the epithelial cells [70]. The release of HMGB1 from necrotic retinal pigment epithelial cells can increase the generation of IL-6 and TNF- α in macrophages and release of these cytokines from retinal pigment epithelial cells; the cytokine release is inhibited by deleting the *Rip3* gene [71]. In another culture model of AMD in which retinal pigment epithelial cells are treated with poly(I:C) and Z-VAD, NF- κ B signaling is directly modulated by HMGB1 and receptorinteracting protein kinase 3 (Rip3) [72].

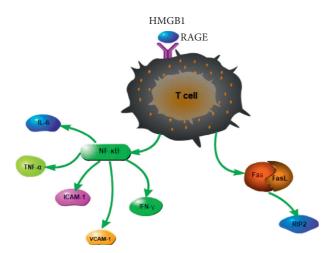


FIGURE 2: HMGB1 signals by binding to RAGE to activate T-cell cytokine release and Fas/FasL signal pathway in uveitis. In uveoretinitis, HMGB1 binds RAGE and activates NF- κ B, promoting the release of TNF- α , IL-6, ICAM-1, and VCAM-1. HMGB1 also can activate Fas/FasL-mediated signaling. In Behcet's disease, expression of proinflammatory cytokines such as TNF- α , IL-6, and IFN- γ in neutrophils is enhanced by extracellular HMGB1, which acts via the NF- κ B pathway.

7. HMGB1 and Glaucoma

Acute glaucoma is an ophthalmic emergency that progresses to blindness if untreated. Obstruction of the drainage of the aqueous humour as a result of narrowing or closure of the anterior chamber angle causes high intraocular pressure, which damages the optic nerve and triggers retinal ischemic reperfusion injury and retinal ganglion cell death. In rats, HMGB1 has been linked to glaucoma induced by elevated intraocular pressure [73]. Treating cocultures of retinal ganglion cells and glia with HMGB1 causes ganglion cell death [74].

HMGB1 significantly upregulates cleaved caspase-8 and activates the canonical NLRP3-inflammasome, which induces the processing of IL-1 β via pathways dependent on caspase-1 and caspase-8 and leads to retinal ganglion cell death and retinal damage [75]. In a model of retinal IR, HMGB1 appears to upregulate expression of the p65 subunit of p-NF- κ B and activate the NF- κ B pathway [75]. RAGE and TLR4 mediate the ability of HMGB1 to activate proinflammatory pathways and enhance retinal damage. Treating ganglion cell layer neurons with anti-HMGB1 antibody improves their survival more than knocking out either RAGE or TLR4 expression [73].

8. HMGB1 and Corneal Diseases

One of the most general reasons of microbial keratitis is *pseudomonas aeruginosa* which is widespread in patients with prolonged contact lenses and impaired immunity. Impaired healing of corneal wounds resulting from alkaline burn is also a major cause of impaired vision, and strategies for treatment should be improved in order to avoid

keratoplasty. These two kinds of corneal diseases are refractory: so far, there is no effective treatment to cure them. HMGB1 is involved in the pathogenesis of both diseases, and it may have potential as a therapeutic target [76].

In *P. aeruginosa*-induced keratitis, elevated levels of extracellular HMGB1 exacerbate inflammation by stimulating neutrophils via RAGE and TLR pathways [77]. Attenuating HMGB1-mediated disease progression can help treat keratitis. In an animal model of bacterial keratitis, silencing HMBG1 reduces expression of IL-1 β , MIP-2, and TNF- α by inhibition of NF- κ B as well as the receptors TLR4 and RAGE [78]. Silencing HMGB1 also downregulates CXCL12 and CXCR4 expression. Silencing HMGB1 in a mouse model of *P. aeruginosa*-induced keratitis increases levels of anti-inflammatory cytokines, reduces levels of inflammatory cytokines and neutrophil infiltration, and improves prognosis [79].

HMGB1 also plays an important role in corneal healing and corneal neovascularization. There are two neovascularization processes. One of these processes is the production of endothelial cells from existing blood vessels. The other process is initiated by bone-derived progenitor cells, which then drive endothelial progenitor cells to target sites and into the vascular endothelium. Endothelial cells are also bone-derived progenitor cells. Then, the progenitor cells differentiate into endothelial cells and support the construction of new blood vessels. HMGB1 accelerates corneal neovascularization via TLR4- and RAGE-mediated signaling pathways by activating macrophages, stimulating the secretion of proangiogenic factors and promoting TLR4-induced recruitment of endothelial progenitor cells [80]. Of the three HMBG1 receptors (TLR2, TLR4, and RAGE), HMBG1 appears to act via TLR4 to drive corneal neovascularization. Nuclear HMGB1 interacts with TLR2, which is present in human and rat corneal epithelial cells, but this receptor has no effect on secreted HMGB1 [81]. Silencing TLR2 does not significantly affect corneal neovascularization [81]. Alkaline burns upregulating RAGE and TLR4 in mouse cornea with similar kinetics, but the upregulation of RAGE is small [47] (Figure 3).

9. Conclusion and Perspectives

Although tremendous advances have been made in the study of the structure, release patterns, and receptors of HMGB1, the intracellular signaling pathways that it modulates are still poorly understood, especially its interaction with TLRs and RAGE. This is important to understand in order to clarify how HMGB1 contributes to ocular diseases and develop better therapeutic strategies. HMGB1 antagonists have shown significant therapeutic effects in animal models, but more extensive trials are needed in animals and ultimately in humans. Future studies should also address whether posttranslational modifications affect HMGB1-mediated signaling and whether other carrier or binding proteins assist HMGB1 in modulating cytokine activity in ocular diseases.

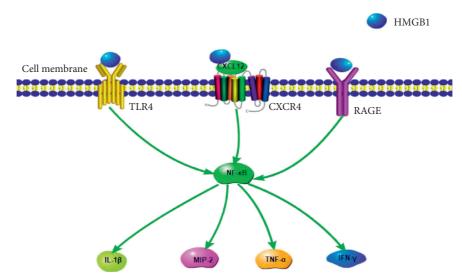


FIGURE 3: Signaling pathways downstream of RAGE, TLR4, and CXCR4 mediate the effects of HMGB1 in corneal diseases. RAGE signals, TLR4 signals, and CXCR4 signals promote the activation of NF- κ B. After the activation of NF- κ B, the inflammatory cytokines such as IL-1 β , MIP-2, TNF- α , and IFN- γ are released.

This work may solidify HMGB1 as a novel therapeutic approach.

Disclosure

Yi Liu and Guo-Bin Zhuang are co-first authors, and they contributed equally to this work.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this article.

Authors' Contributions

Xuezhi Zhou and Yi Liu conceived and designed the study. Yi Liu and Guo-bin Zhuang wrote this paper. Xuezhi Zhou reviewed and edited the manuscript. All authors read and approved the manuscript.

References

- R. Barea, L. Boquete, M. Mazo, and E. Lopez, "System for assisted mobility using eye movements based on electrooculography," *IEEE Transactions on Neural Systems and Rehabilitation Engineering*, vol. 10, no. 4, pp. 209–218, 2002.
- [2] D. A. Jabs and J. T. Rosenbaum, "Guidelines for the use of immunosuppressive drugs in patients with ocular inflammatory disorders: recommendations of an expert panel," *American Journal of Ophthalmology*, vol. 131, no. 5, p. 679, 2001.
- [3] M. A. Toscano, A. G. Commodaro, J. M. Ilarregui et al., "Galectin-1 suppresses autoimmune retinal disease by promoting concomitant Th2- and T regulatory-mediated antiinflammatory responses," *Journal of Immunology*, vol. 176, no. 10, pp. 6323–6332, 2006.
- [4] V. L. Perez and R. R. Caspi, "Immune mechanisms in inflammatory and degenerative eye disease," *Trends in Immunology*, vol. 36, no. 6, pp. 354–363, 2015.

- [5] E. J. Lee, B. R. Brown, E. E. Vance et al., "Mincle activation and the syk/card9 signaling axis Are central to the development of autoimmune disease of the eye," *Journal of Immunology*, vol. 196, no. 7, pp. 3148–3158, 2016.
- [6] M. Magna and D. S. Pisetsky, "The role of HMGB1 in the pathogenesis of inflammatory and autoimmune diseases," *Molecular Medicine*, vol. 20, pp. 138–146, 2014.
- [7] H. Yang, H. Wang, S. S. Chavan, and U. Andersson, "High mobility group box protein 1 (HMGB1): the prototypical endogenous danger molecule," *Molecular Medicine*, vol. 21, no. 1, pp. S6–S12, 2015.
- [8] S. Martinotti, M. Patrone, and E. Ranzato, "Emerging roles for HMGB1 protein in immunity, inflammation, and cancer," *ImmunoTargets and Therapy*, vol. 4, pp. 101–109, 2015.
- [9] N. M. Agalave and C. I. Svensson, "Extracellular high-mobility group box 1 protein (HMGB1) as a mediator of persistent pain," *Molecular Medicine*, vol. 20, pp. 569–578, 2015.
- [10] D. Musumeci, G. N. Roviello, and D. Montesarchio, "An overview on HMGB1 inhibitors as potential therapeutic agents in HMGB1-related pathologies," *Pharmacology & Therapeutics*, vol. 141, no. 3, pp. 347–357, 2014.
- [11] Q. Chen, X. Guan, X. Zuo, J. Wang, and W. Yin, "The role of high mobility group box 1 (HMGB1) in the pathogenesis of kidney diseases," *Acta Pharmaceutica Sinica B*, vol. 6, no. 3, pp. 183–188, 2016.
- [12] F. C. Wang, J. X. Pei, J. Zhu et al., "Overexpression of HMGB1 A-box reduced lipopolysaccharide-induced intestinal inflammation via HMGB1/TLR4 signaling in vitro," World Journal of Gastroenterology, vol. 21, no. 25, pp. 7764–7776, 2015.
- [13] S. Tancharoen, T. Tengrungsun, T. Suddhasthira et al., "Overexpression of receptor for advanced glycation end products and high-mobility group box 1 in human dental pulp inflammation," *Mediators of Inflammation*, vol. 2014, Article ID 754069, 13 pages, 2014.
- [14] W. Wan, L. Cao, R. Khanabdali, B. Kalionis, X. Tai, and S. Xia, "The emerging role of HMGB1 in neuropathic pain: a potential therapeutic target for neuroinflammation," *Journal of Immunology Research*, vol. 2016, Article ID 6430423, 9 pages, 2016.

- [15] H. Saidi, M. Bras, P. Formaglio et al., "HMGB1 is involved in IFN-alpha production and TRAIL expression by HIV-1exposed plasmacytoid dendritic cells: impact of the crosstalk with NK cells," *PLoS Pathogens*, vol. 12, no. 2, Article ID e1005407, 2016.
- [16] C. Semino, G. Angelini, A. Poggi, and A. Rubartelli, "NK/iDC interaction results in IL-18 secretion by DCs at the synaptic cleft followed by NK cell activation and release of the DC maturation factor HMGB1," *Blood*, vol. 106, no. 2, pp. 609– 616, 2005.
- [17] L. Ulloa and D. Messmer, "High-mobility group box 1 (HMGB1) protein: friend and foe," *Cytokine & Growth Factor Reviews*, vol. 17, no. 3, pp. 189–201, 2006.
- [18] S. Zhang, J. Zhong, P. Yang, F. Gong, and C. Y. Wang, "HMGB1, an innate alarmin, in the pathogenesis of type 1 diabetes," *International Journal of Clinical and Experimental Pathology*, vol. 3, no. 1, pp. 24–38, 2009.
- [19] H. E. Harris, U. Andersson, and D. S. Pisetsky, "HMGB1: a multifunctional alarmin driving autoimmune and inflammatory disease," *Nature Reviews Rheumatology*, vol. 8, no. 4, pp. 195–202, 2012.
- [20] U. Andersson and H. E. Harris, "The role of HMGB1 in the pathogenesis of rheumatic disease," *Biochimica et Biophysica Acta (BBA)—Gene Regulatory Mechanisms*, vol. 1799, no. 1-2, pp. 141–148, 2010.
- [21] S. L. Yu, C. K. Wong, and L. S. Tam, "The alarmin functions of high-mobility group box-1 and IL-33 in the pathogenesis of systemic lupus erythematosus," *Expert Review of Clinical Immunology*, vol. 9, no. 8, pp. 739–749, 2013.
- [22] L. M. Bellussi, S. Cocca, L. Chen, F. M. Passali, C. Sarafoleanu, and D. Passali, "Rhinosinusal inflammation and high mobility group box 1 protein: a new target for therapy," *ORL*, vol. 78, no. 2, pp. 77–85, 2016.
- [23] H. Wang, O. Bloom, M. Zhang et al., "HMG-1 as a late mediator of endotoxin lethality in mice," *Science*, vol. 285, no. 5425, pp. 248–251, 1999.
- [24] B. Wittemann, G. Neuer, H. Michels, H. Truckenbrodt, and F. A. Bautz, "Autoantibodies to nonhistone chromosomal proteins HMG-1 and HMG-2 in sera of patients with juvenile rheumatoid arthritis," *Arthritis & Rheumatism*, vol. 33, no. 9, pp. 1378–1383, 1990.
- [25] L. M. Ayer, R. L. Rubin, G. H. Dixon, and M. J. Fritzler, "Antibodies to HMG proteins in patients with drug-induced autoimmunity," *Arthritis & Rheumatism*, vol. 37, no. 1, pp. 98–103, 1994.
- [26] L. Campana, L. Bosurgi, and P. Rovere-Querini, "HMGB1: a two-headed signal regulating tumor progression and immunity," *Current Opinion in Immunology*, vol. 20, no. 5, pp. 518–523, 2008.
- [27] M. E. Bianchi and A. A. Manfredi, "High-mobility group box 1 (HMGB1) protein at the crossroads between innate and adaptive immunity," *Immunological Reviews*, vol. 220, no. 1, pp. 35–46, 2007.
- [28] J. R. Klune, R. Dhupar, J. Cardinal, T. R. Billiar, and A. Tsung, "HMGB1: endogenous danger signaling," *Molecular Medicine*, vol. 14, no. 7-8, pp. 476–484, 2008.
- [29] W. Huang, Y. Tang, and L. Li, "HMGB1, a potent proinflammatory cytokine in sepsis," *Cytokine*, vol. 51, no. 2, pp. 119–126, 2010.
- [30] N. K. Sun and C. C. Chao, "The cytokine activity of HMGB1-extracellular escape of the nuclear protein," *Chang Gung Medical Journal*, vol. 28, no. 10, pp. 673–682, 2005.
- [31] H. Schierbeck, P. Lundback, K. Palmblad et al., "Monoclonal anti-HMGB1 (high mobility group box chromosomal protein

1) antibody protection in two experimental arthritis models," *Molecular Medicine*, vol. 17, no. 9-10, pp. 1039–1044, 2011.

- [32] A. Tsung, S. Tohme, and T. R. Billiar, "High-mobility group box-1 in sterile inflammation," *Journal of Internal Medicine*, vol. 276, no. 5, pp. 425–443, 2014.
- [33] I. E. Dumitriu, P. Baruah, A. A. Manfredi, M. E. Bianchi, and P. Rovere-Querini, "HMGB1: guiding immunity from within," *Trends in Immunology*, vol. 26, no. 7, pp. 381–387, 2005.
- [34] N. T. Sebastian, E. M. Bystry, N. A. Becker, and L. J. Maher III, "Enhancement of DNA flexibility in vitro and in vivo by HMGB box A proteins carrying box B residues," *Biochemistry*, vol. 48, no. 10, pp. 2125–2134, 2009.
- [35] K. T. Bauerle, E. Kamau, and A. Grove, "Interactions between N- and C-terminal domains of the Saccharomyces cerevisiae high-mobility group protein HMO1 are required for DNA bending," *Biochemistry*, vol. 45, no. 11, pp. 3635–3645, 2006.
- [36] M. E. Churchill, J. Klass, and D. L. Zoetewey, "Structural analysis of HMGD-DNA complexes reveals influence of intercalation on sequence selectivity and DNA bending," *Journal of Molecular Biology*, vol. 403, no. 1, pp. 88–102, 2010.
- [37] D. S. Pisetsky, H. Erlandsson-Harris, and U. Andersson, "High-mobility group box protein 1 (HMGB1): an alarmin mediating the pathogenesis of rheumatic disease," *Arthritis Research & Therapy*, vol. 10, no. 3, p. 209, 2008.
- [38] H. Yang, M. Ochani, J. Li et al., "Reversing established sepsis with antagonists of endogenous high-mobility group box 1," *Proceedings of the National Academy of Sciences*, vol. 101, no. 1, pp. 296–301, 2004.
- [39] J. P. Bidwell, J. Yang, and A. G. Robling, "Is HMGB1 an osteocyte alarmin?," *Journal of Cellular Biochemistry*, vol. 103, no. 6, pp. 1671–1680, 2008.
- [40] H. Wang, T. Lei, J. Liu, M. Li, H. Nan, and Q. Liu, "A nuclear factor of high mobility group box protein in Toxoplasma gondii," *PLoS One*, vol. 9, no. 11, Article ID e111993, 2014.
- [41] Q. Wang, M. Zeng, W. Wang, and J. Tang, "The HMGB1 acidic tail regulates HMGB1 DNA binding specificity by a unique mechanism," *Biochemical and Biophysical Research Communications*, vol. 360, no. 1, pp. 14–19, 2007.
- [42] R. Chen, W. Hou, Q. Zhang, R. Kang, X. G. Fan, and D. Tang, "Emerging role of high-mobility group box 1 (HMGB1) in liver diseases," *Molecular Medicine*, vol. 19, no. 1, pp. 357–366, 2013.
- [43] S. A. Lee, M. S. Kwak, S. Kim, and J. S. Shin, "The role of high mobility group box 1 in innate immunity," *Yonsei Medical Journal*, vol. 55, no. 5, pp. 1165–1176, 2014.
- [44] H. Rauvala and A. Rouhiainen, "RAGE as a receptor of HMGB1 (Amphoterin): roles in health and disease," *Current Molecular Medicine*, vol. 7, no. 8, pp. 725–734, 2007.
- [45] G. P. Sims, D. C. Rowe, S. T. Rietdijk, R. Herbst, and A. J. Coyle, "HMGB1 and RAGE in inflammation and cancer," *Annual Review of Immunology*, vol. 28, no. 1, pp. 367– 388, 2010.
- [46] G. Dvoriantchikova, E. Hernandez, J. Grant, A. R. Santos, H. Yang, and D. Ivanov, "The high-mobility group box-1 nuclear factor mediates retinal injury after ischemia reperfusion," *Investigative Opthalmology & Visual Science*, vol. 52, no. 10, pp. 7187–7194, 2011.
- [47] S. Yang, T. S. Yang, F. Wang, and S. B. Su, "High-mobility group box-1-Toll-Like receptor 4 axis mediates the recruitment of endothelial progenitor cells in alkali-induced corneal neovascularization," *International Immunopharmacology*, vol. 28, no. 1, pp. 450–458, 2015.

- [48] T. A. Ciulla, A. G. Amador, and B. Zinman, "Diabetic retinopathy and diabetic macular edema: pathophysiology, screening, and novel therapies," *Diabetes Care*, vol. 26, no. 9, pp. 2653–2664, 2003.
- [49] A. P. Adamis and A. J. Berman, "Immunological mechanisms in the pathogenesis of diabetic retinopathy," *Seminars in Immunopathology*, vol. 30, no. 2, pp. 65–84, 2008.
- [50] S. Gardella, C. Andrei, D. Ferrera et al., "The nuclear protein HMGB1 is secreted by monocytes via a non-classical, vesiclemediated secretory pathway," *EMBO Reports*, vol. 3, no. 10, pp. 995–1001, 2002.
- [51] A. M. El-Asrar, M. I. Nawaz, D. Kangave et al., "High-mobility group box-1 and biomarkers of inflammation in the vitreous from patients with proliferative diabetic retinopathy," *Molecular Vision*, vol. 17, pp. 1829–1838, 2011.
- [52] A. M. El-Asrar, L. Missotten, and K. Geboes, "Expression of high-mobility groups box-1/receptor for advanced glycation end products/osteopontin/early growth response-1 pathway in proliferative vitreoretinal epiretinal membranes," *Molecular Vision*, vol. 17, pp. 508–518, 2011.
- [53] A. M. Abu El-Asrar, K. Alam, M. Garcia-Ramirez et al., "Association of HMGB1 with oxidative stress markers and regulators in PDR," *Molecular Vision*, vol. 23, pp. 853–871, 2017.
- [54] S. H. Zhu, B. Q. Liu, M. J. Hao et al., "Paeoniflorin suppressed high glucose-induced retinal microglia MMP-9 expression and inflammatory response via inhibition of TLR4/NFkappaB pathway through upregulation of SOCS3 in diabetic retinopathy," *Inflammation*, vol. 40, no. 5, pp. 1475– 1486, 2017.
- [55] J. J. Lee, C. C. Hsiao, I. H. Yang et al., "High-mobility group box 1 protein is implicated in advanced glycation end products-induced vascular endothelial growth factor A production in the rat retinal ganglion cell line RGC-5," *Molecular Vision*, vol. 18, pp. 838–850, 2012.
- [56] S. Jiang and X. Chen, "HMGB1 siRNA can reduce damage to retinal cells induced by high glucose in vitro and in vivo," *Drug Design, Development and Therapy*, vol. 11, pp. 783–795, 2017.
- [57] A. M. Abu El-Asrar, G. Mohammad, M. I. Nawaz, and M. M. Siddiquei, "High-mobility group box-1 modulates the expression of inflammatory and angiogenic signaling pathways in diabetic retina," *Current Eye Research*, vol. 40, no. 11, pp. 1141–1152, 2015.
- [58] G. Mohammad, D. Jomar, M. M. Siddiquei, K. Alam, and A. M. Abu El-Asrar, "High-mobility group box-1 protein mediates the regulation of signal transducer and activator of transcription-3 in the diabetic retina and in human retinal muller cells," *Ophthalmic Research*, vol. 57, no. 3, pp. 150–160, 2017.
- [59] G. Mohammad, K. Alam, M. I. Nawaz, M. M. Siddiquei, A. Mousa, and A. M. Abu El-Asrar, "Mutual enhancement between high-mobility group box-1 and NADPH oxidasederived reactive oxygen species mediates diabetes-induced upregulation of retinal apoptotic markers," *Journal of Physiology and Biochemistry*, vol. 71, no. 3, pp. 359–372, 2015.
- [60] H. Shao, H. J. Kaplan, and D. Sun, "Major histocompatibility complex molecules on parenchymal cells of the target organ protect against autoimmune disease," *Chemical Immunology and Allergy*, vol. 92, pp. 94–104, 2007.
- [61] T. Watanabe, H. Keino, Y. Sato, A. Kudo, H. Kawakami, and A. A. Okada, "High mobility group box protein-1 in experimental autoimmune uveoretinitis," *Investigative Opthalmology & Visual Science*, vol. 50, no. 5, pp. 2283–2290, 2009.

- [62] J. Yun, G. Jiang, Y. Wang et al., "The HMGB1-CXCL12 complex promotes inflammatory cell infiltration in uveitogenic T cell-induced chronic experimental autoimmune uveitis," *Frontiers in Immunology*, vol. 8, p. 142, 2017.
- [63] G. Jiang, D. Sun, H. Yang, Q. Lu, H. J. Kaplan, and H. Shao, "HMGB1 is an early and critical mediator in an animal model of uveitis induced by IRBP-specific T cells," *Journal of Leukocyte Biology*, vol. 95, no. 4, pp. 599–607, 2014.
- [64] G. Jiang, Y. Ke, D. Sun, G. Han, H. J. Kaplan, and H. Shao, "Reactivation of uveitogenic T cells by retinal astrocytes derived from experimental autoimmune uveitis-prone B10RIII mice," *Investigative Opthalmology & Visual Science*, vol. 49, no. 1, pp. 282–289, 2008.
- [65] G. Jiang, Y. Wang, J. Yun et al., "HMGB1 release triggered by the interaction of live retinal cells and uveitogenic T cells is Fas/FasL activation-dependent," *Journal of Neuroinflammation*, vol. 12, no. 1, p. 179, 2015.
- [66] A. W. de Souza, S. F. Perazzio, N. R. de Franca et al., "High mobility group box 1 serum levels are increased in Behcet's disease, but not associated with disease activity or disease manifestations," *Rheumatology (Oxford)*, vol. 54, no. 12, pp. 2151–2155, 2015.
- [67] D. S. Friedman, B. J. O'Colmain, B. Munoz et al., "Prevalence of age-related macular degeneration in the United States," *Archives of ophthalmology*, vol. 122, no. 4, pp. 564–572, 2004.
- [68] J. Hanus, C. Anderson, D. Sarraf, J. Ma, and S. Wang, "Retinal pigment epithelial cell necroptosis in response to sodium iodate," *Cell Death Discovery*, vol. 2, no. 1, article 16054, 2016.
- [69] J. Hanus, C. Anderson, and S. Wang, "RPE necroptosis in response to oxidative stress and in AMD," *Ageing Research Reviews*, vol. 24, pp. 286–298, 2015.
- [70] J. Hanus, H. Zhang, Z. Wang, Q. Liu, Q. Zhou, and S. Wang, "Induction of necrotic cell death by oxidative stress in retinal pigment epithelial cells," *Cell Death & Disease*, vol. 4, no. 12, p. e965, 2013.
- [71] A. Kaczmarek, P. Vandenabeele, and D. V. Krysko, "Necroptosis: the release of damage-associated molecular patterns and its physiological relevance," *Immunity*, vol. 38, no. 2, pp. 209–223, 2013.
- [72] Y. Murakami, H. Matsumoto, M. Roh et al., "Programmed necrosis, not apoptosis, is a key mediator of cell loss and DAMP-mediated inflammation in dsRNA-induced retinal degeneration," *Cell Death & Differentiation*, vol. 21, no. 2, pp. 270–277, 2014.
- [73] U. Andersson and K. J. Tracey, "HMGB1 is a therapeutic target for sterile inflammation and infection," *Annual Review* of *Immunology*, vol. 29, no. 1, pp. 139–162, 2011.
- [74] M. Schallenberg, V. Prokosch, and S. Thanos, "Regulation of retinal proteome by topical antiglaucomatous eye drops in an inherited glaucoma rat model," *PLoS One*, vol. 7, no. 7, Article ID e33593, 2012.
- [75] W. Chi, H. Chen, F. Li, Y. Zhu, W. Yin, and Y. Zhuo, "HMGB1 promotes the activation of NLRP3 and caspase-8 inflammasomes via NF-kappaB pathway in acute glaucoma," *Journal of Neuroinflammation*, vol. 12, no. 1, p. 137, 2015.
- [76] S. A. Ekanayaka, S. A. McClellan, R. P. Barrett, and L. D. Hazlett, "Topical glycyrrhizin is therapeutic for *Pseu*domonas aeruginosa keratitis," *Journal of Ocular Pharma*cology and Therapeutics, vol. 34, no. 3, pp. 239–249, 2018.
- [77] S. A. Ekanayaka, S. A. McClellan, R. P. Barrett, S. Kharotia, and L. D. Hazlett, "Glycyrrhizin reduces HMGB1 and bacterial load in *Pseudomonas aeruginosa* keratitis," *Investigative Opthalmology & Visual Science*, vol. 57, no. 13, pp. 5799–5809, 2016.

- [78] S. McClellan, X. Jiang, R. Barrett, and L. D. Hazlett, "Highmobility group box 1: a novel target for treatment of *Pseudomonas aeruginosa* keratitis," *Journal of Immunology*, vol. 194, no. 4, pp. 1776–1787, 2015.
- [79] L. D. Hazlett, S. A. McClellan, and S. A. Ekanayaka, "Decreasing HMGB1 levels improves outcome of *Pseudomonas aeruginosa* keratitis in mice," *Journal of Rare Diseases Research & Treatment*, vol. 1, no. 1, pp. 36–39, 2016.
- [80] S. Yang, L. Xu, T. Yang, and F. Wang, "High-mobility group box-1 and its role in angiogenesis," *Journal of Leukocyte Biology*, vol. 95, no. 4, pp. 563–574, 2014.
- [81] Q. Lin, X. P. Yang, D. Fang et al., "High-mobility group box-1 mediates toll-like receptor 4-dependent angiogenesis," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 31, no. 5, pp. 1024–1032, 2011.