MITOGENOME ANNOUNCEMENT

OPEN ACCESS

Complete chloroplast genome sequence of Acacia crassicarpa (Fabaceae)

Xinjian Yue^{a,b}, Yuyun Yu^a (), Wei Gao^c, Shipin Chen^a, Zebin Weng^d and Gongfu Ye^c

^aCollege of Forestry, Fujian Agriculture and Forestry University, Fuzhou, People's Republic of China; ^bForest Inventory and Planning Institute of Fujian Province, Fuzhou, People's Republic of China; ^cFujian Academy of Forestry Science, Fuzhou, People's Republic of China; ^dFujian Provincial Department of Forestry, Fuzhou, People's Republic of China

ABSTRACT

Acacia crassicarpa (Fabaceae), a nitrogen-fixing tree species, is critically important for coastal protection in southeast China. In this study, we report the complete chloroplast genome sequence of *A. crassicarpa*, with a length of 176,493 bp. It contains a pair of inverted repeats (IR 39,851 bp), a large single-copy region (LSC 91,869 bp), and a small single-copy region (SSC 4,922 bp). The complete genome comprises 138 genes, including 93 protein-coding genes, 37 tRNA, and 8 rRNA genes. Our phylogenetic analysis reveals that *A. crassicarpa* is closely related to *A. podalyriifolia* and *A. dealbata*.

ARTICLE HISTORY Received 15 March 2021 Accepted 12 June 2021

Taylor & Francis

Taylor & Francis Group

KEYWORDS *Acacia crassicarpa;* chloroplast genome; phylogeny; Fabaceae

Acacia crassicarpa A.Cunn. ex Benth. 1842 (Fabaceae), a nitrogen-fixing tree species, is native to Australia, Papua New Guinea and Indonesia (Moran et al. 1989; Sulistyono et al. 2020). Acacia crassicarpa has been a critically important tree species for coastal protection forest in southeast China since the 1980s (Lin 2019). The wood of A. crassicarpa has many commercial uses, including the production of fiber, pulp, construction, shipbuilding, among others (Ling 1996). Few studies have focused on the genome of A. crassicarpa with only its microRNA identified to date (Liu 2015). However, comprehensive understanding of the chloroplast genome of A. crassicarpa is still lacking. Research into the chloroplast genome can not only improve its function, but also enhance our understanding of its biology and biodiversity. Here, we report the complete chloroplast genome sequences of A. crassicarpa, and reveal the phylogenetic relationships to related species in Fabaceae.

Fresh leaves of *A. crassicarpa* was collected, and dried into silica gel immediately, in Dongshan county (23°38'21.22"N, 117°24'22.17"E) in Fujian Province, China. The voucher specimen (Acr_yyy) was deposited in the Herbarium, College of Forestry, Fujian Agriculture and Forestry University (Xinjian Yue, yxinj03@126.com). Total genomic DNA was extracted from leaf tissue samples preserved in silica gel using the CTAB method (Doyle 1987). The obtained DNA was fragmented to construct a paired-end library with an insert-size of 500 bp and the genome sequencing were performed using Illumina Hiseq Xten platform, with approximately 10 GB of data generated. Illumina data were filtered by SOAPnuke (parameter: -n 0.01 -l 20 -q 0.3 -A0.25 –cutAdaptor -Q 2 -G

-polyX50 -minLen 150). The chloroplast genome of *A. crassicarpa* was then assembled using the GetOrganelle pipe-line (https://github.com/Kinggerm/GetOrganelle) by recruiting plastid-like reads. Final reads were viewed and edited by Bandage (Wick et al., 2015). The assembled chloroplast genome annotation was based on the comparison with *A. dealbata* by Geneious v.11.1.5 (Kearse et al. 2012). The annotation results were drawn with the online tool OGDRAW (http://ogdraw.mpimp-golm.mpg.de/) (Marc et al. 2013).

The complete chloroplast genome sequence of *A. crassicarpa* (GeneBank accession: MW649002) was 176,493 bp in length. It contained a large single-copy (LSC) region of 91,869 bp, a small single-copy (SSC) region of 4,922 bp, and a pair of inverted repeats (IR) regions of 39,851 bp. The complete chloroplast genome was comprised of 138 genes, with 93 protein-coding genes, 37 tRNA genes, and 8 rRNA genes. The GC contents of the LSC, SSC, and IR regions individually, and of the complete genome as a whole, are 33.1%, 29.4%, 38.3%, and 35.3%, respectively.

To investigate the phylogenetic history of *A. crassicarpa*, 23 complete chloroplast genomes of Fabaceae and Magnoliaceae species were downloaded from NCBI and were aligned with MAFFT(Katoh and Standley 2013). A maximum-likelihood (ML) tree was constructed based on the 23 complete chloroplast genome sequences using the CIPRES Science Gateway web server (RAxML-HPC2 on XSEDE 8.2.12) (Miller et al. 2010) with 1000 bootstrap replicates. The phylogenetic analysis revealed that *A. crassicarpa* is closely related to *A. podalyriifolia* and *A. dealbata* (Figure 1).

CONTACT Gongfu Ye 🖾 yegongfu@126.com 💼 Fujian Academy of Forestry Sciences, Fuzhou, 350012, People's Republic of China

© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

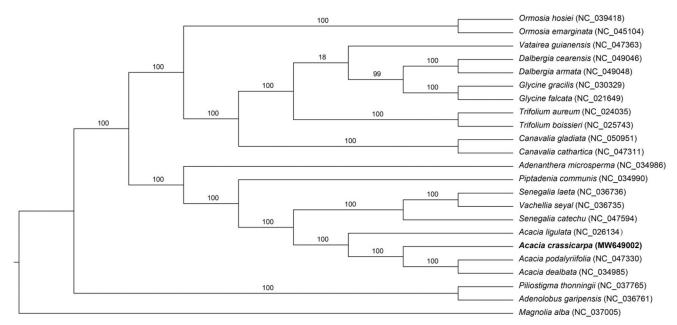


Figure 1. Maximum likelihood phylogenetic tree based on the complete chloroplast genome sequence of Acacia crassicarpa and other related species, with Magnolia alba representing the outgroup.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This work was supported by Fujian Provincial Department of Science and Technology under Grant [2018NZ0001-1].

ORCID

Yuyun Yu (i) http://orcid.org/0000-0002-8515-3019

Data availability statement

The genome sequence data that support the findings of this study are openly available in GenBank of NCBI at [https://www.ncbi.nlm.nih.gov] under the accession no.MW649002. The associated BioProject, SRA, and Bio-Sample numbers are PRJNA698470, SRX10001513, and SAMN17720400 respectively.

References

- Doyle J. 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. Phytochem Bull. 19(1):11–15.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, et al. 2012. Geneious Basic: an integrated and extendable desktop software platform for the

organization and analysis of sequence data. Bioinformatics. 28(12): 1647–1649.

- Lin Y. 2019. Nutrient resorption efficiency and stoichiometry of N and P on *Acacia crassicarpa* plantations. J Fujian Forest Sci Technol. 46(01): 11–16.
- Ling C. 1996. A multifunctional tree species—*Acacia crassicarpa*. Forest Guangxi. (04):9–9.
- Liu W. 2015. Identification and expression of microRNAs and their targets in *Acacia crassicarpa* organogenesis. Beijing: Beijing Forestry University.
- Marc L, Oliver D, Sabine K, Ralph B. 2013. OrganellarGenomeDRAW—a suite of tools for generating physical maps of plastid and mitochondrial genomes and visualizing expression data sets. Nucleic Acids Res. 41(W1):W575–W581.
- Miller MA, Pfeiffer WT, Schwartz T. 2010. Creating the CIPRES science gateway for inference of large phylogenetic trees. Gateway Computing Environments Workshop (GCE), 2010.
- Moran GF, Muona O, Bell JC. 1989. Breeding systems and genetic diversity in *Acacia auriculiformis* and *A. crassicarpa*. Biotropica. 21(3): 250–256.
- Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol Biol Evol. 30(4):772–780.
- Sulistyono E, Kkadan SK, Maretha MV, Souza Tavares WD, Sirait BA, Hanjelina Br Sinulingga NG, Tarigan M, Duran A. 2020. First report, morphological and molecular identification of Spodoptera species (Lepidoptera, Noctuidae) on *Acacia crassicarpa* (Fabaceae) in Sumatra, Indonesia. J Lepidopterists Soc. 74(3):176.
- Wick RR, Schultz MB, Justin Z, Holt KE. 2015. Bandage: interactive visualization of de novo genome assemblies. Bioinformatics. 31(20): 3350–3352.