

Chinese Pharmaceutical Association Institute of Materia Medica, Chinese Academy of Medical Sciences

Acta Pharmaceutica Sinica B

www.elsevier.com/locate/apsb www.sciencedirect.com



REVIEW

Benzimidazole and its derivatives as cancer therapeutics: The potential role from traditional to precision medicine

Yeuan Ting Lee, Yi Jer Tan, Chern Ein Oon*

Institute for Research in Molecular Medicine (INFORMM), Universiti Sains Malaysia, 11800 Minden, Penang, Malaysia

Received 31 May 2022; received in revised form 11 August 2022; accepted 8 September 2022

KEY WORDS

Benzimidazole derivatives; Targeted therapy; Anticancer; Precision medicine **Abstract** Cancer is the second leading cause of mortality globally which remains a continuing threat to human health today. Drug insensitivity and resistance are critical hurdles in cancer treatment; therefore, the development of new entities targeting malignant cells is considered a high priority. Targeted therapy is the cornerstone of precision medicine. The synthesis of benzimidazole has garnered the attention of medicinal chemists and biologists due to its remarkable medicinal and pharmacological properties. Benzimidazole has a heterocyclic pharmacophore, which is an essential scaffold in drug and pharmaceutical development. Multiple studies have demonstrated the bioactivities of benzimidazole and its derivatives as potential anticancer therapeutics, either through targeting specific molecules or non-gene-specific strategies. This review provides an update on the mechanism of actions of various benzimidazole derivatives and the structure-activity relationship from conventional anticancer to precision healthcare and from bench to clinics.

© 2023 Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

*Corresponding author. Tel.: +60 4653 4879.

https://doi.org/10.1016/j.apsb.2022.09.010



E-mail address: chern.oon@usm.my (Chern Ein Oon).

Peer review under responsibility of Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences.

^{2211-3835 © 2023} Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Benzimidazole derivatives as anticancer agents

The rapid escalation of cancer incidence and the incremental mortality rate have made cancer a global burden. The development of tumour resistance, drug toxicities, cancer recurrence, and the low success rate of drug development reaching clinical trials are limiting factors that aggravate the challenges in cancer treatment. One of the focuses on improving treatment efficacy and survival rate of cancer patients is the search for new classes of anticancer drugs. The traditional 'one-size-fits-all' approach using conventional non-targeting agents is toxic to healthy cells and may not benefit all patients. As cancer is a significant focus of the precision medicine initiative, personalized therapeutic approaches aimed to maximize outcomes based on individual variability in genetic profile, lifestyle, and environmental factors are widely gaining acceptance¹. Therefore, targeted therapy provides the foundation of precision medicine to allow tailored treatment targeting specific oncogenic markers in cancers.

Among the anticancer drugs discovered in recent years, various benzimidazole derivatives have gained attention in anticancer agent development due to their diverse biological activities and clinical applications. The unique core structure of benzimidazole and its minimal toxicity property has made it an excellent scaffold in anticancer drug development. Benzimidazole (also known as 1H-benzimidazole, 1,3-benzodiazole, benzoglyoxaline, iminazole, and imidazole) is an aromatic organic compound that contains a benzene ring fused to an imidazole ring at 4,5-position to form a bicyclic ring^{2,3}. Historically, benzimidazole (*i.e.*, 2,6dimethylbenzimidazole) was first synthesized by Hoebrecker, followed by Ladenberg and Wundt in the 1870s². Benzimidazole has a molecular weight of 118.14 g/mol and appears as white tabular crystals. Benzimidazole contains a hydrogen atom attached to nitrogen in the 1-position and can form a tautomer upon interaction with aprotic solvents, such as water or the existence of more than one benzimidazole molecule². Nonetheless, substitution at position N will prohibit the tautomerism process³. Benzimidazole is a weak base with a pK value at 5.3 and 12.3 for pK_{a1} and pK_{a2} , respectively⁴. Therefore, the benzimidazole ring is highly stable and can withstand extreme conditions such as being heated under pressure up to 270 °C in a concentrated sulphuric acid solution or vigorous treatment with hot hydrochloric acid or with alkalis⁴.

Benzimidazole is an important biologically active heterocyclic compound that serves as one of the top ten most frequently employed five-membered nitrogen heterocycles among the US Food and Drug Administration (FDA) approved drugs⁵. The electron-rich nitrogen heterocycles of benzimidazole could readily accept or donate protons and easily allow the formation of diverse weak interactions, offering an advantage for it to bind with a broad spectrum of therapeutic targets, thereby exhibiting wide-ranging pharmacological activities⁶. Mounting evidence has reported a vast pharmacological profile of benzimidazole and its derivatives, substitution at the 1, 2, 5 and/or 6-positions, in multiple categories of therapeutic agents with unique properties, including anti-microbial⁷⁻¹⁰, anti-tuberculosis^{11–13}, anti-viral^{14–16}, anti-ulcer^{17,18}, anti-inflammatory^{19–21}, anti-diabetic, anti-convulsant^{22,23}, anti-hypertensive²⁴, and anti-malarial^{25–27}. There is also increasing evidence highlighting the prospect of benzimidazole derivatives as anticancer agents, particularly in the advancement of precision medicine, which will be discussed in this review.

Benzimidazoles have a structure that resembles naturally occurring purine nucleotides, which allows them to easily contact the biopolymers within the living system²⁸. The benzimidazole pharmacophore can form hydrogen bonds, amide-ring and aromatic-ring interactions, hydrophobic interactions, van der Waals forces, polar contact and pi-bonds with the targets (Table 1, Fig. 1). Benzimidazole and its derivatives are reported to play key roles as topoisomerase inhibitors, DNA intercalation and alkylating agents, androgen receptor antagonists, poly(ADP-ribose) polymerase (PARP) inhibitors, protein kinase inhibitors, dihydrofolate reductase inhibitors, and microtubule inhibitors^{29,30}. Benzimidazole derivatives have also been reported to act as epigenetic regulators with demonstrated promising anticancer activities³¹⁻³⁴. Examples of benzimidazole-based drugs that have gained approval clinically encompass Binimetinib (NCT04965818 and NCT03170206), Bendamustine (NCT04217317 and NCT04510636), Selumetinib (NCT02768766), Abemaciclib (NCT04003896 and NCT0404-0205), Veliparib (NCT02723864 and NCT01434316), Dovitinib (NCT01635907), Pracinostat (NCT03848754), Galeterone (NCT-04098081) and Nazartinib (NCT02335944 and NCT02108964)³⁵. This review article summarises the recent literature on benzimidazole derivatives exhibiting anticancer properties based on different mechanisms.

Available X-ray crystal structures with root-mean-square deviation (RMSD) ≤ 2.5 Å from each category were selected for analysis. In addition, ligand—receptor interactions between the benzimidazole moiety and target receptors are presented based on analyses from Protein Data Bank in Europe (PDBe) database (Table 1, Fig. 1).

1.1. Traditional non-oncogene targeting anticancer agents

1.1.1. Topoisomerase inhibitors

DNA topoisomerase is an important ubiquitous enzyme associated with genomic integrity involving DNA replication, transcription, recombination, and chromatin remodelling³⁶. Type I topoisomerase (Topo I) cleaves only one strand of the DNA molecule. In contrast, type II topoisomerase (Topo II) functions by cutting both strands of the double-stranded DNA molecule³⁷. Cells mainly utilize this enzyme to maintain the chromosome segregation and topology of DNA. In general, these topoisomerase inhibitors work as catalytic inhibitors to suppress topoisomerase activity, to activate enzyme activity towards bona fide or surrogate substrate; or convert topoisomerase into a toxic enzyme in the cells^{38,39}. Much effort has been channelled into developing topoisomerase inhibitors or topoisomerase poisons using the benzimidazole scaffold in cancer cells. Therefore, targeting DNA topoisomerases to prevent cancer replicative immortality has been one of the research interests in anticancer drug development for several decades (Fig. 2).

The flexible nature of the bis-benzimidazole ring allows the high-affinity binding of its derivatives to the DNA, which leads to the change in DNA conformation and inhibits the formation of the cleavable complex⁴⁰. Bielawski and colleagues have reported the use of bis-benzimidazole derivatives with chloroalkyl and bromoalkyl moieties in developing the Topo I and Topo II inhibitors, compounds 1–4. These compounds inhibit DNA synthesis by interacting with the GC base pair at the DNA minor groove, leading to the irreversible inhibition of proliferation in the MDA-MB-231 estrogen receptor-negative human breast cancer cells. The four compounds 1–4 have been found to hamper the incorporation of [³H]thymidine into the DNA at 48 ± 2, 38 ± 2, 25 ± 3 and 17 ± 3 µmol/L, respectively, resulting in the reduction of the MDA-MB-231 cell viability⁴⁰. Oksuzoglu et al.⁴¹ have cleverly

Category	PDB ID	RMSD (Å)	Compd.	Receptor	Interaction
DNA intercalation and	2B3E	1.36	DBN	DNA	Hydrogen, polar, carbon-pi, aromatic
alkylating agents	453D	1.80	E96	DNA	Van der Waals, hydrogen, polar, carbon-pi,
	11/7/	1 77	DID	DNA	pi—pi Van den Waste bedressen gelen seiten mi
	IVZK	1.//	DIR	DNA	van der waals, hydrogen, polar, carbon-pi,
	442D	1.60	IB	DNA	Van der Waals hydrogen bond polar
	1120	1.00	12	2101	carbon-pi
PARP inhibitors	3KJD	1.95	78P	PARP-2	Hydrogen, polar, carbon-pi, pi-pi, donor-
					pi, amide-ring, hydrophobic
	7AAC	1.59	78P	PARP-1	Van der Waals, hydrogen, polar, carbon-pi,
					pi—pi, donor–pi, amide-ring, aromatic,
	5WS1	1.00	7110	DA DD 1	Nan der Waals hydrogen polar carbon-pi
	54431	1.90	109	IARI-I	ni-ni donor-ni amide ring aromatic
					hydrophobic
	6NRJ	1.65	KYJ	PARP-1	Hydrogen, polar, carbon-pi, donor-pi,
					aromatic, hydrophobic
Kinase inhibitors	1ZOH	1.81	K44	CK2	Van der Waals, hydrogen, polar, carbon-pi,
		1.07		CTT A	methionine-sulfur-pi, hydrophobic
	4KWP	1.25	EXX	CK2	Van der Waals, hydrogen, polar, carbon-pi,
	3H30	1.56	RE7	CK2	Van der Waals hydrogen polar carbon-pi
	51150	1.50	KI Z	CK2	methionine-sulfur-pi
	4DSU	1.70	BZI	GTPase KRas	Hydrogen, polar, carbon-pi
	3DA6	2.00	BZ9	MAPK10	Van der Waals, hydrogen, polar, carbon-pi,
					methionine-sulfur-pi
	3EWH	1.60	K11	VEGFR2	Polar, carbon-pi
	5KGD	1.98	6SL	Ser/Thr protein kinase Pim 1	Hydrogen, polar, carbon-pi
Androgen receptor inhibitors	2YLO	2.5	YLO	Androgen receptor	Van der Waals, polar, carbon-pi, pi-pi
	4HLW	2.5	17W	Androgen receptor	Polar, pi—pi
Epigenetic modulators	7KBG	1.26	WBD	HDAC2	Van der Waals, hydrogen, polar, pi-pi,
	6DOE	1.69	U61		hydrophobic Van dar Waala hydrogan polar donor ni
	UDQI	1.08	H01	KDMJA	hydrophobic
	5PHK	1.25	BZI	KDM4D	Van der Waals, hydrogen, polar, covalent.
					hydrophobic
	6G5X	1.78	MY7	KDM4A	Van der Waals, hydrogen, carbon-pi, pi-pi,
					donor-pi, cation-pi, aromatic, covalent,
					metal complex

 Table 1
 The interaction between the benzimidazole pharmacophore of compounds and respective target receptors.

synthesized eighteen derivatives of 2,5-disubstituted-benzoxazole and benzimidazole with demonstrated Topo I and Topo II enzymatic activities. Among the eighteen compounds, 2-phenoxymethylbenzimidazole (compound 17) contained the benzimidazole moiety and exhibited the most potency in inhibiting DNA Topo I enzymatic activities, with IC₅₀ values at 14.1 μ mol/L. MH1 is another potent topoisomerase inhibitor, synthesis of 2,5-disubstituted benzimidazoles, that binds to DNA at the minor groove in leukemia cells (Molt4 cells), inhibiting the conversion of supercoiled DNA to circular DNA, leading to G2/M arrest and apoptosis⁴². Gao et al.⁴³ synthesized a benzimidazole-acridine derivative, compound 8I, with reported strong cytotoxic effects against K562 leukaemia and HepG-2 hepatocellular carcinoma cells at 2.68 and 8.11 µmol/L, respectively. This compound functions as Topo I inhibitor and promotes cell death in K562 cells through the intrinsic apoptotic pathway. In another study, Li et al.⁴⁴ demonstrated a panel of benzimidazole-rhodanine conjugates to possess strong antiproliferative activity against human lymphoma, acute leukaemia, human cervical, breast, lung, and prostate cancer cells. Among the 35 synthesized benzimidazole-rhodanine conjugates, compounds **8g** and **8j** exhibited the best Topo II inhibitory activity at 10 μ mol/L. Compounds **8g** and **8j** act as non-intercalative Topo II inhibitors that bind to the ATP-binding site of the Topo II enzyme to block the enzymatic activity. The presence of the benzyl and electron donor groups on the compound revealed a significant impact on Topo II inhibitory activity.

1.1.2. DNA intercalation and alkylating agents

Protein-DNA binding interactions are implicated in various cancer pathways and often require the direct or indirect contact of protein at the major and/or minor grooves of DNA^{45,46}. The interactions play a significant role in DNA replication, gene transcription, chromosomal packaging, and DNA repair^{47,48}. However, the interruption of protein-DNA interactions by the DNA intercalator could alter the DNA conformation and the biological functions of DNA-binding proteins such as transcription factors, topoisomerase, polymerase and DNA repair proteins⁴⁹. Therefore, DNA intercalators are widely accepted in cancer chemotherapy⁵⁰. Drugs targeting protein-DNA interaction work either through covalent or non-covalent binding⁵¹. The former involves metallointercalators



Figure 1 Key interactions between the benzimidazole pharmacophore of benzimidazole derivatives and target receptors. Representative structures illustrating key interactions between the benzimidazole moiety of various compounds with (a) kinases (PDB ID: 3DA6), (b) PARP proteins (PDB ID: 7AAC), (c) androgen receptors (PDB ID: 2YLO), (d) epigenetic regulators (PDB ID: 7KBG) and (e) DNA (PDB ID: 1VZK). Structures and interactions were obtained from the PDBe database and visualized using PyMOL 2.0 software. Receptors are presented as cartoons, and residue side chains are presented as licorice according to the elements: oxygen is red, nitrogen is blue, and sulfur is yellow. Crystal waters are presented as red spheres. Benzimidazole compounds are presented as green licorice.

(*e.g.*, cisplatin) which form a dual-inhibition mode (*i.e.*, intercalation and metal coordination) that leads to irreversible binding and ultimately induces cancer cell death. On the contrary, non-covalent binding is reversible in which intercalation, groove binding or electrostatic interactions take place between two adjacent base pairs of duplex DNA, resulting in the inhibition of nucleic acid synthesis and consequently impair cancer cell replication. The mechanism involved either intercalation, grooves binding or electrostatic interactions^{47,50,52,53}. Most benzimidazole derivatives act as DNA minor groove binders (MGBs), particularly at the AT-rich sequences (Fig. 3).

Yamori et al.⁵² have synthesized a compound known as MS-247, (2-[[*N*-[1-methyl-2-[5-[*N*-[4-[*N*,*N*-bis(2-chloroethyl) amino] phenyl]] carbamoyl]-1*H*-benzimidazol-2-yl] pyrrol-4-yl] carbamoyl] ethyldimethylsulfonium di-*p*-toluenesulfonate) that exhibited antitumour activity in 39 cancer cell lines and 17 tumour xenografts of the lung, colon, stomach, breast, and ovarian cancers. MS-247 consists of a netropsin-like moiety and an alkylating residue in the structure to facilitate the DNA minor groove binding and DNA alkylation properties, respectively. MS-247 attaches to the AT-rich sites of the DNA minor groove to inhibit DNA synthesis *via* the formation of DNA–DNA interstrand crosslinks (ICL). This ultimately leads to cell cycle blockage at the G2/M phase and apoptosis induction^{52,54}.

Bisbenzimodazole compound **5**, another widely studied DNA MGB, is a symmetrical bis-benzimidazole, 2,2'-di-[[(3,5-dimethyl-4-methoxy)pyrid-2-yl]methylenethio]-5,5'-bis-1H,1'H-benzimid-azole synthesized by Joubert et al.⁵⁵ as a dimeric bis-benzimidazole molecule that recognises ten base pair ([A'T]₄-[G'C]-[A'T]₄ mo-tifs) in the DNA sequence. Interestingly, it is less sensitive on the

human ovarian carcinoma cells (IC₅₀ > 10 µmol/L range) as compared to two other benzimidazole derivatives (IC₅₀ <<1 µmol/L), which, however, displayed no known role as minor groove binders and quick uptake by the tumour cell. The central nitrogen in the $-N^+H(Me)$ - linker of compound 5 was predicted to have the highest binding affinity towards the DNA B-helix. Compound **8** is another bis-benzimidazole derivative with an omeprazole thioether group connected to the 2-position benzimidazole, demonstrating MGB function. Compound **8** exhibited cytotoxic effects on ovarian cancer cells at IC₅₀ = 2.95 µmol/L but was least effective on cervical and gastric cancer cells (IC₅₀ = or > 50 µmol/L)⁵⁶. Similarly, Compound **16** with pyrid-4yl substitute that Yang and colleagues⁵⁶ synthesized is another MGB also demonstrated a potent antitumour effect at IC₅₀ = 2.81, 32.4 and 11.0 µmol/L on SKOV-3 ovarian, HeLa cervical and BGC-823 gastric cancer cells respectively.

Bendamustine hydrochloride comprises a benzimidazole heterocyclic ring, mechlorethamine, N-substituted methyl group and butyric acid substitution. The bendamustine compound can form intra- and interstrand crosslinks between DNA bases, impede DNA replication, deter transcription and repair, and disrupt the DNA matrix function during DNA replication⁵⁷. Bendamustine is an alkylating agent which causes DNA breakage. It is more potent than other DNA-alkylating agents such as cisplatin, cyclophosphamide, or carmustine, highlighting the role of the benzimidazole ring in enhancing the anticancer properties of bendamustine compared to other conventional 2-chloroethylamine alkylators⁵⁸. In 2008, the US Food and Drug Administration (FDA) has approved bendamustine hydrochloride (TREANDA®) as a DNA alkylating agent targeting chronic lymphocytic leukaemia (CLL) and B-cell non-Hodgkin's lymphoma (NHL)⁵⁷. The molecular



Figure 2 Benzimidazole derivatives as topoisomerase inhibitors.

mechanism of Bendamustine in cancer includes the induction of S-phase mitotic arrest, mitotic catastrophe and the activation of p53-mediated apoptosis⁵⁸. However, this drug may cause pyrexia, nausea, and vomiting in CLL patients. In addition, headache, fatigue, weight loss, vomiting, diarrhoea, pyrexia, rash, constipation, anorexia, cough, dyspnea, and stomatitis were also commonly observed in patients diagnosed with non-Hodgkin's lymphoma^{59,60}.

The benzimidazole ring in synthesized compounds has been shown to increase anticancer potency selectively in leukaemia^{61,62}. Compound 4f (NSC: 761982/1) uses bendamustine and chlorambucil as templates. Compound 4f consists of a methylene linker which joins the benzimidazole and oxadiazole. Compound 4f is cytotoxic on leukaemia, melanoma, ovarian, prostate, breast, colon, central nervous system, and non-small cell lung cancer cells⁶³. The presence of oxadiazole conjugate was found to enhance the antiproliferative effects compared to thiadiazole, triazolo-thiadiazines and triazolo-thiadiazoles⁶³. Compound 4c is a pyrrolobenzodiazepine (PBD)-conjugated benzimidazole derivative that has demonstrated remarkable DNA-binding affinity at GI₅₀ value and potent inhibition of cancer cell growth with concentration less than 10 nmol/L in the 60 cancer cell lines screen of NCI, comprising of leukaemia, melanoma, lung, ovarian, colon, renal, breast, prostate, and CNS cancers⁶⁴. In 2013, Al-Mudaris and colleagues⁶⁵ successfully synthesized the benzyl vanillin (2-(benzyloxy)-3methoxybenzaldehyde) analogues, 2MP, 2-(2-benzyloxy-3methoxyphenyl)-1H-benzimidazole) and 2XP (N-1-(2-benzyloxy-3-methoxybenzyl)-2-(2-benzyloxy-3-methoxyphenyl)-1H-benzimidazole), and screened them for anticancer potentials. The addition of benzimidazole moiety to the side chain has improved the

DNA binding affinity, and enhanced the cytotoxic effect of **2MP** and **2XP** compared to their parent structure. However, the anticancer activity was not shown in **2MP** but in **2XP**. In addition, **2XP** exhibited anti-proliferative capabilities towards HL60 leukaemia cancer cells and induced G2/M phase cell cycle arrest and apoptosis in these cells.

1.1.3. Microtubule inhibitors

Microtubules are major cytoskeleton components composed of α and β -tubulin subunits⁶⁶. Microtubules help maintain cellular shape development, motility, and facilitate cell-cell communication, mitosis and cell division⁶⁶. Hence, the dynamic instability of microtubules would inevitably result in a continuous rapid turnover, which is critical for mitosis⁶⁷. Microtubule inhibitors, also known as antimitotic drugs, function by suppressing the spindle-microtubule dynamics, leading to the partial attachment of microtubules to chromosomes at their kinetochores, and the incomplete formation of metaphase spindles⁶⁸. Treatment with such inhibitors would arrest mitosis at the metaphase-anaphase transition and induce apoptotic cell death^{66,67,69}. This strategy has been successful with the employment of clinical drugs such as paclitaxel and Vinca alkaloids in cancers such as Hodgkin and non-Hodgkin lymphomas, breast, colon, cervical, ovarian and testicular carcinoma⁶⁹⁻⁷¹. However, toxicity and multidrug resistance are limiting factors in the development of microtubule targeting agents (MTAs)⁷². The paclitaxel site, Vinca and Colchicine domains are the three main binding sites targeted by MTAs through microtubule stabilization and destabilization^{66,72}. The benzimidazole derivatives function as a microtubule inhibitor are summarized in Fig. 4.



Figure 3 Benzimidazole derivatives as DNA intercalation and alkylating agents.

Methyl 2-(5-fluoro-2-hydroxyphenyl)-1*H*-benzo[*d*]imidazole-5-carboxylate (MBIC) is a microtubule inhibitor that consists of 2hydroxyl and 5-fluoro substitution in the aryl ring, and a methyl ester group giving a strong cytotoxic effect against breast cancer cells^{73,74}. The MBIC was reported to induce mitosis and stimulates mitochondria-dependent intrinsic apoptotic cell death in cervical cancer⁷⁵. Cervical cancer cells treated with MBIC exhibited G2-M phase arrest followed by cancer cell death. MBIC was also reported to inhibit microtubule polymerisation, through the up-regulation of cyclin B1, cyclin-dependent kinase 1 (CDK1), and budding uninhibited by benzimidazole-related 1 (BubR1) proteins and down-regulation of the Aurora B protein, indicating the occurrence of mitotic arrest. Besides, MBIC also demonstrated cytotoxicity against a panel of hepatocellular carcinoma cells *via* reactive oxygen species (ROS)-mediated activation of the JNK signalling cascade⁷⁶.



Figure 4 Benzimidazole derivatives used as microtubule inhibitors.

Nocodazole (methyl [5-(2-thienylcarbonyl)-1*H*-benzimidazol-2-yl]carbamate) functions as a tubulin destabiliser that disrupts the microtubule formation in prometaphase cells and hampers the ability of the microtubules to attach to the chromosome kinetochores, leading to cell cycle arrest, and eventually cell death^{77–79}. The benzimidazole core is substituted at position 2 by a (methoxycarbonyl) amino group and position 5 by a 2-thienoyl group. A study conducted by Blajeski and co-workers demonstrated that selected breast cancer cells (*i.e.*, MCF-7, MDA-MB-231, MDA-MB-436, MDA-MB-453, HS0578T, SKBr3, and ZR-75-10) to be susceptible to mitotic arrest when treated with nocodazole. These breast cancer cells undergo p21-associated cell cycle arrest at G1 and G2 phases post-treatment with 1 μ mol/L Nocodazole⁷⁸.

Compound **5** is an imidazo[1,5-a]pyridine-benzimidazole hybrid that showed significant cytotoxic activity against the 60 human cancer cell lines with GI₅₀ values ranging from 0.43 to 7.73 µmol/L while displaying no cytotoxicity in normal human embryonic kidney (HEK-293) cells⁸⁰. The cytotoxic effects of compound **51** on cancer cells are attributed to its binding pattern to the colchicine binding site of the tubulin. The presence of the methoxy phenyl group improved the binding affinity to the colchicine binding site and further enhanced its antiproliferative activity. Compound 51 binds to the colchicine binding site to suppress tubulin polymerisation by 71.27% at the IC₅₀ value of 1.71 µmol/L. This treatment concentration eventually resulted in ROS production and mitochondrial-dependent cell death of MCF-7 breast cancer cells. In addition, compound 51 also inhibits tubulin polymerisation and the p13/AKT pathway to retard breast tumour growth.

1.2. Target-specific modalities as the basis of precision medicine

1.2.1. Kinase inhibitors

Kinases are enzymes that are imperative in mediating the execution of the signalling cascade in regulating molecular functions and biological processes, including growth, proliferation, differentiation, and apoptosis. The human kinome comprises approximately 535 protein kinases, which can be classified into tyrosine kinases, serine/threonine kinases, and tyrosine kinase-like enzymes⁸¹. Tyrosine kinases (TKs) encompassed receptor tyrosine kinases (RTKs) (*e.g.*, EGFR, FGFR, PDGFR) and non-receptor tyrosine kinases (NRTKs) (*e.g.*, ABL, FBK, SRC). BRAF/MEK/ ERK, CDKs, PI3K/AKT/mTOR are serine/threonine kinases. In living cells, a defect in cellular kinase phosphorylation can eventually lead to the constitutive activation of kinases to promote the development of malignancy and tumour progression^{81,82}. The benzimidazole scaffold has been widely employed as a template for synthesizing kinase inhibitors (Fig. 5).

TIBI or 4,5,6,7-tetraiodobenzimidazole is an example of an ATP-competitive inhibitor targeting protein kinase CK2 inhibitor derived from 5,6-dichlorobenzimidazole $1-\beta$ -D-ribofuranoside (DRB) scaffold^{83,84}. The substitution of four iodine groups on the benzene ring demonstrated a better binding affinity to the CK2 ATP-binding pocket than tetrabromo and tetrachloro substitutions, which possess an IC₅₀ value of 0.023 µmol/L⁸³. Koronkiewicz and colleagues⁸⁵ have previously reported that TIBI exerted proapoptotic and cytostatic effects on promyelocytic leukaemia cell line through inhibition of CK2 activity. Inhibition of CK2 by TIBI was then found to interfere with another protein kinase, Rio1, which is highly expressed in glioblastoma cells⁸⁴. The inhibition of CK2

has been reported to negatively affect the Rio1 atypical serine kinase, resulting in the perturbation of ribosome biogenesis, cell cycle progression, and chromosome maintenance⁸⁴. The specific interaction between Rio1 and CK2 resulted in the similar susceptibility of the two protein kinases to TIBI, creating another cross-link between the enzymes.

Dovitinib ((3E)-4-amino-5-fluoro-3-[5-(4-methylpiperazine-1yl)-1,3-dihydrobenzimidazol-2-ylidene]quinolin-2-one), initially known as CHIR-258 or TKI258, is an orally bioavailable lactate salt of a benzimidazole-quinolinone compound which function as a multitargeted growth factor receptor kinase inhibitor⁸⁶. Dovotinib binds to class III (FLT3/c-Kit) with IC₅₀ of 1-2 nmol/L, class IV (fibroblast growth factor receptor 3, FGFR1/3) and class V (vascular endothelial growth factor receptor, VEGFR1-4) receptor tyrosine kinases (RTKs) with IC₅₀ of 8-13 nmol/L^{86,87}. Treatment of hepatocellular carcinoma (SK-HEP1) cells with dovitinib resulted in G2/M phase arrest, inhibition of cell proliferation, blockage of bFGF-induced cell motility, and promoted apoptosis⁸⁸. Furthermore, dovitinib potently suppressed lung tumour growth, metastasis and significantly prolonged mouse survival⁸⁸. Besides, dovotinib was also found to hamper the leukaemia K562 cancer cells growth by functioning as Topo I and II inhibitors⁸⁹. The FDA has recently approved Dovitinib in a premarket approval application for a companion test in renal cell carcinoma (RCC) patients⁹⁰.

Selumetinib (ARRY-142886 or AZD6244) [6-(4-bromo-2chloro-phenylamino)-7-fluoro-3-methyl-3H-benzimidazole-5carboxylic acid (2-hydroxy-ethoxy)-amide], is a non-ATPcompetitive, mitogen-activated protein kinase (MEK) inhibitor targeting MEK1 and MEK2 (IC₅₀ = 14 nmol/L). The binding of Selumetinib at the MEK1/2 allosteric inhibitor binding site disrupts the interaction of both ATP and substrate and the assessment of the ERK activation loop^{91,92}. Selumetinib can inhibit oncogenic downstream effects of the Raf-MEK-ERK signalling pathway, demonstrating potent anticancer effects in vitro and in vivo, and is currently under phase I and II trials for pancreatic cancer, melanoma, non-small cell lung cancer and differentiated thyroid cancer⁹³⁻⁹⁵. Although selumetinib has been investigated for the treatment of several types of cancer, it is currently approved only for the treatment of neurofibromatosis type 1 (NF1) in paediatric patients ≥ 2 years who have symptomatic, inoperable plexiform neurofibromas⁹⁶.

Binimetinib [5-((4-bromo-2-fluorophenyl)amino)-4-fluoro-N-(2-hydroxyethoxy)-1-methyl-1H-benzimidazole-6-carboxamide] is another non-ATP-competitive, high selectivity inhibitor toward MEK1/2 with IC₅₀ at 12 nmol/L^{97,98}. Remarkably, Binimetinib was highly selective, thus no off-target inhibition was detected across 220 other serine/threonine and tyrosine kinases^{97,98}. Inhibition of the MEK1/2 led to the suppression of ERK activity which ultimately promoted cancer cell apoptosis and hampered tumour growth in tumour xenograft^{97,99}. Binimetinib gained approval by the FDA in 2018 to be used in combination with Encorafenib to treat metastatic cutaneous melanoma with the BRAF V600E or V600K mutation^{98,100}.

Abemaciclib [*N*-[5-[(4-ethylpiperazin-1-yl)methyl]-pyridin-2yl]-*N*'-[5-fluoro-4-(7-fluoro-2-methyl-3-methylethyl-3*H*-benzimidazol-5-yl)-pyrimidin-2-yl]-amine] is a cyclin-dependent kinase (CDK) inhibitor that has gained the FDA approval to be used as an adjuvant with endocrine therapy to treat breast cancers¹⁰¹. Abemaciclib selectively targets CDK4 (IC₅₀: 2 nmol/L) and CDK6 (IC₅₀:10 nmol/L) and is competitively bound to the ATP binding site of the enzymes¹⁰². Inhibition of CDK4/6 suppressed Rb



Figure 5 Benzimidazole derivatives as protein kinase inhibitors.

phosphorylation, led to G1 arrest, and impeded cancer cell proliferation¹⁰³.

Nazartinib [EGF816; (R,E)-N-(7-chloro-1-(1-[4-(dimethylamino)but-2-enoyl]azepan-3-yl)-1H-benzo[d]imidazole-2-yl)-2methylisonicotinamide] is a third-generation, irreversible, mutantselective EGFR TKI that potently targets EGFR-activating mutations¹⁰⁴. Nazartinib comprises dimethylamino crotonamide, which is known as the optimal group for a number of covalent pan-EGFR inhibitors, a racemic 3-substituted azepane linker, and a chloro substituent at the benzene ring of the benzimidazole nucleus contribute to the better solubility, oral bioavailability, selectivity and high affinity towards EFGR¹⁰⁵. Nazartinib suppresses the EGFR signalling and MAPK pathway, promoting cell cycle arrest and apoptosis and tumour regression in xenograft models¹⁰⁶. Nazartinib is currently under phase I/II clinical trial in patients with EGFR-mutant non-small-cell lung carcinoma¹⁰⁴.

Compound 56q (CPL304110) was identified as a selective and potent pan-FGFR inhibitor for FGFR1, 2, 3 with IC₅₀ of 0.75, 0.50, 3.05 nmol/L respectively, whereas IC₅₀ of 87.90 nmol/L for FGFR4. Due to its favourable pharmacokinetic profile, low toxicity and potent antitumour activity in vivo, compound 56g is currently under evaluation in phase I clinical trial for the treatment of bladder, gastric and squamous cell lung cancers (01FGFR2018; NCT04149691)¹⁰⁷. Compound **12n** is a 6,7-dimethoxy-*N*-(2phenyl-1*H*-benzo[*d*]imidazole-6-yl)quinolin-4-amine derivatives with para-tert-butyl substituent to the phenyl ring, targeting the c-Met tyrosine kinase with an IC₅₀ value of 0.030 \pm 0.008 μ mol/L. Their findings suggested the substituent with lipophilic properties favours the inhibitory activity due to the ability to form a hydrophobic interaction with c-Met kinase. The anticancer potential of compound 12n is also well demonstrated in A549 non-small cell lung, MCF-7 breast, and MKN-45 stomach cancer cells at IC_{50} concentrations of 7.3 \pm 1.0, 6.1 \pm 0.6, and 13.4 \pm 0.5 µmol/ L respectively¹⁰⁸. Compounds **18** and **27** are rapidly accelerated fibrosarcoma (RAF) kinase inhibitors, with the reported IC₅₀ values of 0.002 and 0.014 µmol/L against the B-RAF^{V600E}oncogenic protein is strikingly elevated in skin and colon cancers¹⁰⁹. The RAF oncogene positively correlates with cancer cellular processes, including proliferation, invasion, metastasis and cell survival. Both compounds can inhibit the proliferation of the SK-MEL-28 melanoma cells harbouring the B-RAF^{V600E} proteins (half-maximal effective concentration, $EC_{50} = 4.6$ and 2.3 µmol/L) to consequently inhibit the phosphorylation of extracellular-signal-regulated kinase (ERK). Both compounds 18 and 27 possess a methyl group at the benzimidazole NH but varied in the substituent at the R₂ position (4-bromo substituent in compound 18, whereas 3-(4-pyridyl) in compound 27)¹⁰⁹. Ramurthy and colleagues demonstrated the presence of methyl group at benzimidazole NH to enhance the binding affinity towards RAF kinase compared to the unmethylated analogous. In this study, compound 27 demonstrated significant impediment of the ERK pathway, particularly in B-RAF^{V600E} in HT29 colon mouse tumour xenograft model with an oral administration of 30 and 100 (mg/kg)/day for 28 days. However, significant body weight loss was observed as a notable side effect at a high dosage $(100 \text{ mg/kg/day})^{109}$.

Compound **5a** (2-chloro-N-(2-p-tolyl-1H-benzo[d]imidazole-5-yl)acetamide) is a 2-aryl benzimidazole which is known as a multi-target RTK inhibitor against EGFR, VEGFR-2 and PDGFR¹¹⁰. The presence of Part II: new candidates of pyrazole– benzimidazole conjugates as checkpoint kinase 2 (CHK2) inhibitors. In a separate study by Chu and colleagues¹¹¹, compound 5a was reported to block the erythroblastic leukaemia viral oncogene homologue (ErbB) full family of transmembrane receptor tyrosine kinases (RTKs). These RTKs include the epidermal growth factor receptor (EGFR) and human epidermal growth factor receptor 2 (HER2), which are markedly overexpressed in breast cancer. In addition, compound 5a also upregulates the death receptor 5 (DR5) through the activation of the c-Jun N-terminal kinase (JNK) signalling pathway in EGFRpositive, HER2-positive and EGFR/HER2-double-negative breast cancer cells (i.e., MDA-MB-468, BT-549, MDA-MB-231, HCC1937, T-47D, BT-474, MDA-MB-453, ZR-75-1 and MCF-7 cells). Inhibition of the EFGR and HER2 expression may subsequently obstruct the downstream activation of PI3K/AKT and MEK/ERK pathways. Conversely, the activation of DR5 protein expression resulted in breast cancer cell death¹¹¹. Compound 8d with carbamoylhydrazone substituent was another candidate from the library of benzimidazole derivatives synthesized by Galal et al.¹¹², acting as a checkpoint kinase 2 (CHK2) inhibitor to inhibit the cell cycle progression in ER + MCF7 breast, HeLa cervical, HepG2 hepatocellular cancer cell lines. In addition, the compound 8d significantly shifted the cell cycle distribution from 8% in the S phase to 51% in the G2/M phase when used in combination with doxorubicin in a breast cancer model, indicating its potency as a cytostatic agent. However, compound 8d displayed toxicity in vitro and in vivo and showed a median lethal dose (MD₅₀) at 500 mg/kg body weight in mice.

The 1,2,3-triazolyl linked 2-aryl benzimidazole derivatives, compounds 10c (p-chlorophenyl-substituted 1,2,3-triazolyl N-isopropylamidine) and 11f (benzyl-substituted 1,2,3-triazolyl imidazoline) demonstrated remarkable anti-proliferative activity and induced apoptosis and primary necrosis in non-small cell lung cancer (A549) cells at IC50 concentrations: 0.05 and 0.07 µmol/L¹¹³. The 1,2,3-triazolyl demonstrated a high binding affinity towards different kinases and its combination with benzimidazole increased the cytostatic selectivity against nonsmall cell lung cancer cells. The substituent of imidazoline in compound **11f** demonstrated better proliferative activity than the amidine substituent in compound 10c. The presence of PhOCH₂ linker contributes to the high selectivity towards A549 cells and increased activity on CFPAC-1 and HeLa cells. Moreover, the p-substitution with CIPh further enhanced the potency of compound 10c towards A549 cells. Nevertheless, both compounds showed a remarkable reduction of sphingosine kinase 1 (SK1) and p38 mitogen-activated protein kinase (p38 MAPK) proteins expressed in abundance in lung carcinoma. Besides, compound 11f also downregulated the cyclin-dependent kinase 9 (CDK9) protein expression, delineating its function in cell cycle arrest. In silico study further confirmed their inhibition activity against p38 MAPK activities¹¹³.

1.2.2. Poly(ADP-ribose) polymerase-1 (PARP-1) inhibitors

Poly(ADP-ribose) polymerase-1 (PARP-1), also known as poly(ADP-ribose) synthetase or poly(ADP-ribose) transferase, is a nuclear enzyme with a crucial role in executing an immediate DNA damage response (DDR) to DNA damage and promoting DNA repair. DNA damage is classified as single-strand breaks or double-strand breaks. If these damages are not repaired, chronic DDR signalling may trigger cancer cell death or cellular senescence¹¹⁴. Cancers of the liver, breast, skin, lung, and ovary have been reported to rely on PARP-mediated DNA repair for survival¹¹⁵. PARP-mediated DDR includes base excision repair (BER), mismatch repair (MMR), nucleotide excision repair

(NER), non-homologous end joining (NHEJ) or homologous recombination (HR)^{116–118}. PARP maintains genome integrity and facilitates cell survival¹¹⁹. Inhibition of PARP-1 results in genomic instability and accumulation of damaged cells in cell cycle arrest, eventually leading to apoptosis¹¹⁹. Synthetic lethality (SL) refers to cell death due to simultaneous deficiencies in expressing two or more genes¹²⁰. SL has been detected in HR-deficient cancer cells bearing defective DDR upon inhibition of PARP¹¹⁵. SL can also transpire as a consequence of gene perturbation using small molecule inhibitors. For instance, PARP inhibitor has been shown to induce synthetic lethality in HR-deficient breast cancer cells harbouring the *BRCA1* and *BRCA2* gene mutations by exploiting the defect in DNA repair machinery in these cells. The PARP inhibitors with benzimidazole moiety are illustrated in Fig. 6.

Many commercially available PARP inhibitors, including Olaparib, Rucaparib, Niraparib, Veliparib, and Talazoparib, are being studied in various cancer clinical trials^{121,122}. Among them, Rucuparib and Veliparib are benzimidazole carboxamide-derived PARP inhibitors. The benzimidazole-carboxamide scaffold provides an advantage with its low molecular weight and high intrinsic potency towards the active binding site of PARP by forming hydrogen bonds and π -stacking interactions with the nicotinamide binding site of PARP-1^{121,123}. Rucaparib (RubracaTM) is a tricyclic benzimidazole carboxamide derivative targeting PARP-1, -2, and -3 with IC₅₀ of 0.8, 0.5 and 28 nmol/L respectively^{124,125}. Rucaparib is the first PARP inhibitor tested in clinical trials, having entered phase III trial on solid tumour^{124,126-128}. The FDA has approved it¹²⁹ and the European Medicine Agency (EMA)¹³⁰ to be used as a therapeutic agent to treat ovarian, fallopian tube, and peritoneal cancers, as well as advanced solid tumours with evidence of germline or somatic BRCA mutation. Veliparib (2-[(R)-2-methylpyrrolidin-2-yl]-1Hbenzimidazole-4-carboxamide) has an inhibitory constant of 5.2 and 2.9 nmol/L against PARP-1 and PARP-2 respectively¹³¹. Veliparib has been tested preclinically in combination with temozolomide in a panel of tumours, including lymphoma, glioma, melanoma, colon, breast, ovarian, lung, prostate and pancreatic cancers¹³². Remarkably, the study revealed significant inhibition of PARP in all xenograft tumours. Veliparib is currently in phase II and phase III clinical trials for breast, lung, prostate, glioma, pancreatic, colorectal and ovarian cancers¹³¹. The relatively low molecular weight and high intrinsic potency of Veliparib have made it attractive in cancer therapy¹²¹. However, Veliparib exhibits the least potency to trap PARP-1 on DNA with relatively low cytotoxic and reduced stability of PARP–DNA complexes in cancer cells as compared to other PARP-1 inhibitors used in the clinics (Talazoparib >> Olaparib = Rucaparib >> Veliparib)^{133,134}.

Compounds 5cj and 5cp, also known as 2-(1-(3-(4-chloroxyphenyl)-3-oxo-propyl)pyrrolidine-3-yl)-1H-benzo[d] imidazole-4-carboxamide, are PARP inhibitor synthesized using Veliparib as a template with substitution of alkyl side chain with an aromatic ring attached to the nitrogen atom in the five-member cyclic amine, and a phenyl group attached to the terminal of the side chain with the purpose to improve membrane permeability. Both compounds 5cj and 5cp displayed an inhibition potency with IC₅₀ values at 3.9 and 3.6 nmol/L for PARP-1 as well as 4.2 and 3.2 nmol/L for PARP-2, respectively. These two compounds have demonstrated better cytotoxicity against breast and pancreatic cancer cells as compared to Veliparib and Olaparib¹²¹. The IC₅₀ of compound 5cj towards MDA-MB-436 breast and CAPAN-1 pancreatic cancer cells are at 17.4 and 11.4 µmol/L, respectively. On the other hand, compound 5cj displayed an IC₅₀ of 19.8 and 15.5 µmol/L against the breast and pancreatic cancer cell lines¹²¹. In an elegant study, Lee et al.¹²³, reported compound **24**, with an oxadiazole moiety at the 2-phenyl group of benzimidazole scaffold, inhibits H₂O₂-induced poly(ADP-ribosyl)ation in C-41 cervical carcinoma cells (EC₅₀ = 3.7 nmol/L) and exhibits an excellent in vivo oral efficacy in the B16F10 murine melanoma



Rucaparib ICso Cell free assay: PARP1:0.0008 µmol/L PARP2:0.0005 µmol/L PARP3: 0.028 µmol/L Ovarian cancer cells: 2.5 - > 15 mmol/L



IC₅₀ Cell free assay: PARP1: 0.0038 µmol/L PARP2: 0.0042 µmol/L Pancreatic cancer cells: 11.4 µmol/L Breast cancer cells: 17.4 µmol/L

Compound 5cj



Veliparib

I C₅₀ Cell free assay: PARP1: 0.0052 μmol/L PARP2: 0.0029 μmol/L Ovarian cancer cells: 85.5 μmol/L Pancreatic cancer cells: 83.0 μmol/L Breast cancer cells: 130.1 μmol/L



Compound 5cp

ICso Cell free assay: PARP1: 0.0036 µmol/L PARP2: 0.0032 µmol/L Pancreatic cancer cells: 15.5 µmol/L Breast cancer cells: 19.8 µmol/L



Compound 24

IC₅₀ Cell free assay: PARP1: 0.001 μmol/L Pancreatic cancer cells: 15.5 μmol/L EC₅₀ Cervical cancer cells: 0.0037 μmol/L

Compound **27** ICso Cell free assay: PARP1: 0.018 µmol/L PARP2: 0.042 µmol/L Breast cancer cells: 0.92 µmol/L

Figure 6 Benzimidazole derivatives used as PARP inhibitor.

model. Compound **27** (6-fluoro-2-(4,5,6,7-tetrahydrothieno[2,3-*c*] pyridin-2-yl)-1*H*-benzo[*d*]imidazole-4-carboxamide) which contains a 4,5,6,7-tetrahydrothienopyridin-2-yl moiety and a methyl group at N1 of benzimidazole core, revealed better potency against PARP-1 and PARP-2 with the IC₅₀ of 18 and 42 nmol/L respectively compared to other analogues¹³⁵. Compound **27** was cytotoxic towards cells harbouring BRCA2 mutants with an IC₅₀ of 0.92 µmol/L, and significantly impeded tumour growth in the BRCA-1-mutated MDA-MB-436 breast xenograft tumour model.

1.2.3. Androgen receptor (AR) antagonists

Androgen receptor (AR) is also known as nuclear receptor subfamily 3, group C, member 4 (NR3C4). It belongs to the steroid receptor family that plays an essential role in regulating gene transcription in response to testosterone and dihydrotestosterone (DHT) binding¹³⁶. AR is a critical prognostic marker used commonly for diagnosing breast and prostate cancers. Approximately 53%–99% of breast cancer¹³⁷ and 80%–90% of prostate cancer¹³⁸ have been reported to depend on AR for tumour initiation and progression. Increased expression of AR proteins is also associated with poor response to endocrine treatment¹³⁹. AR has many vital functions, including the stimulation of prostate-specific antigen (PSA) secretion, regulation of lipid metabolism, and growth promotion of castration-resistant prostate cancer (CRPC)¹⁴⁰. AR antagonists that disrupt the AR/SRC association can activate the human epidermal growth factor receptor (i.e., HER2, HER3) or phosphoinositide 3-kinase (PI3K), to inhibit androgen-induced cell proliferation in breast cancer¹⁴¹. ARs provide excellent therapeutic targets in managing estrogen receptorpositive breast cancer to prevent recurrence and spread¹⁴¹. However, very little is known about the use of benzimidazole-derived AR antagonists to treat cancer; hence this opens a new avenue for further investigation. Below are some benzimidazole derivatives employed in cancer research (Fig. 7).

Galeterone (3β -hydroxy-17-(1*H*-benzimidazole-1-yl)androsta-5,16-diene) functions by blocking androgen binding to AR. Galeterone, formerly known as VN/124-1, has shown that the presence of benzimidazole moieties in the steroidal C-17 benzoazoles structure enhanced the antagonist effect against AR^{142,143}. The anticancer effect is orchestrated through CYP17 lyase inhibition, AR antagonism, or induction of AR degradation



Figure 7 Benzimidazole derivatives used as androgen receptor antagonist.

which in turn disrupts androgen signalling^{144,145}. Galeterone specifically targets two highly homologous deubiquitinating enzymes (i.e., USP12 and USP46). Both enzymes are implicated in the positive regulation of the AR-AKT-MDM2-p53 signalling pathway. Interestingly, treatment with Galeterone can effectively inhibit cancer cell growth in androgen-sensitive, and rogenindependent anti-androgen resistant and androgen-negative prostate cancer cell lines¹⁴⁵. With an IC₅₀ value of 384 nmol/L in human prostate PC3AR cells, Galeterone significantly inhibited the growth of androgen-dependent LAPC4 human prostate tumour xenograft¹⁴². Galeterone was previously investigated in phase III clinical trials to investigate its efficacy in cancer patients with splice variant 7 (AR-V7) demonstrated resistance to AR treatment^{146,147}. Regrettably, the primary endpoint of radiographic progression-free survival was not met, demonstrating a lack of clinically meaningful benefits to the patients; hence the trial was terminated by the Data Safety Monitoring Board¹⁴⁸.

Munuganti et al.¹⁴⁹ have synthesized a series of 2-((2phenoxyethyl)thio)-1*H*-benzimidazole and 2-((2-phenoxyethyl) thio)-1H-indole, compound **32**, that target the binding function 3 (BF3) protein in AR. Compound 32 is a BF3-specific inhibitor with the corresponding IC50 of 4.2 µmol/L, which causes a conformation change in the BF3 receptor. The sulphur atom in compound 32 is essential for the binding to AR receptor and inhibitory activity. The replacement of sulphur atom with other atoms such as nitrogen, carbon, sulfinyl, and sulfonyl has reduced activity and binding affinity. Besides, the presence of a methyl group at the meta-position of benzene rings enhances the ligand binding by forming a hydrophobic interaction with BF3. Compound 32 decreased prostate-specific antigen (PSA) levels in LNCaP and Enzalutamide-resistant pancreatic cancer cells and Enzalutamide-resistant cell line with the reported IC₅₀ of 4.3 µmol/L¹⁴⁹. Compound 1 (5,6-dichloro-benzimidazole derivatives) is a hydroxyl analogue with trifluoromethyl group having demonstrated antagonistic effect in rat prostate xenograft model with $ID_{50} = 0.15$ mg/day. The 2-(2,2,2-trifluoro-ethyl)benzimidazole 4 has shown a potent efficacy towards AR and the substitution of the OH group in compound 1 further improves the oral bioavailability (PO). The compound has a high PO (132%) and a half-life of 14.7 h¹⁵⁰. Ng et al.¹⁵¹ further extended the study by synthesizing the N-substituted 2-(2,2,2-trifluoroethyl)-5,6dichloro-benzimidazoles with benzyl, N-aceto, and ethylene aryl ether group, using the 5,6-dichloro-benzimidazole scaffold. Benzyl substitution with bromine at the 4th position gave rise to the 4-bromobenzyl analogue, compound 17, which has an ID_{50} of 0.13 mg/day in the rat and is more potent than another known antiandrogen drug, bicalutamide. Conversely, the N-aceto and ethylene aryl ether derivatives exhibited no improvement in the antagonistic activity in prostate cancer cells.

1.2.4. Aromatase inhibitors

Aromatase is a cytochrome P450 (CYP19A1) enzyme that synthesises estrogen from either androstenedione or testosterone^{152,153}. Aromatase was overexpressed in breast cancer and fuel the tumour progression and metastasis in estrogen receptor- α (ER α) positive breast cancers¹⁵³. Aromatase inhibitors lower estrogen levels by halting the enzyme activity of aromatase in estrogen biosynthesis¹⁵⁴. Therefore, aromatase inhibition is one of the effective current therapeutic strategies for controlling estrogen-dependent breast cancer (Fig. 8i).

A library of benzimidazole-triazolothiadiazine derivatives has been synthesized and exhibited aromatase inhibition activities¹⁵⁵.

Among the compounds, compound **5e** revealed the potent inhibitory effect of aromatase with an IC₅₀ of 0.032 µmol/L in breast cancer cells¹⁵⁵. Compound **5e** carries a 4-cyanophenyl substituent at the fourth position of the phenyl ring, which contributes to its inhibitory activity¹⁵⁵. Another potent aromatase inhibitor, a 4benzylpiperidine derivative: compound **2g** inhibits aromatase and exhibits cytotoxic effects against breast and lung cancer cells with an IC₅₀ of 0.024 and 0.071 µmol/L, respectively¹⁵⁶. The presence of chlorine substituent on the benzene ring enhanced the interaction with the aromatase active site by forming a halogen bond with the Gln215 amino group within the enzyme¹⁵⁶.

1.2.5. Dihydrofolate reductase (DHFR) inhibitors

Dihydrofolate reductase (DHFR) is a metabolic enzyme that regulates cell proliferation by catalysing the reduction of dihydrofolate to tetrahydrofolate by NADPH to produce active folate required for the *de novo* synthesis of purines, amino acids, and thymidylate (TMP)¹⁵⁷. The up-regulation of DHFR has been described in different cancers, for example, human glioma¹⁵⁸, ovarian¹⁵⁹, and acute lymphoblastic leukaemia (ALL)¹⁶⁰. High DHFR expression is also associated with poor survival and drug resistance^{159,160}. Inhibition of DHFR debilitates tetrahydrofolate production, which subsequently affects cell growth and proliferation, and ultimately drives cell death in various cancers including leukaemia, osteosarcoma, hepatocellular carcinoma and lung cancer¹⁵⁷.

Compound **6b** from Singla et al.¹⁶¹ is the most active DHFR inhibitor with an IC₅₀ of 1.05 μ mol/L (Fig. 8ii). It is a regioisomeric hybrid of *s*-triazine and benzimidazole moieties, substituting primary and secondary amines. Compound **6b** harbours nucleophilic displacement at the primary and secondary amines. This attribute enhances its anticancer effects with a median growth inhibition (GI₅₀) in the range of 3.31–5.61 μ mol/L in melanoma, colon, breast, and ovarian cancer cells. Besides performing as a DHFR inhibitor, compound **6b** has also displayed a strong DNA interaction, showing its potential to act as a DNA intercalator¹⁶¹.

1.3. Epigenetic modulators

Cancer is a consequence of accumulative genetic mutations in concert with epigenetic alterations that lead to alteration in various cellular events, including cell proliferation, invasion, senescence, and apoptosis^{162,163}. Epigenetic heterogeneity in the tumour could

also lead to various diagnostic and treatment outcomes even in cancer patients with the same tumour stage and grade. However, the application of benzimidazole moiety in the development of epigenetic regulators, is scarce. Here we report the reported application of benzimidazole moiety as histone deacetylases (HDACs) and demethylase inhibitors (Fig. 9).

HDACs describe a group of histone-silencing enzymes that remove the acetyl group from histones, and maintain the steadystate level of lysine acetylation level of histone and non-histone proteins¹⁶⁴. HDACs are implicated in gene silencing transcription repression, in which the aberrant expression may lead to malignancy^{165,166}. HDAC can be divided into four classes comprising class I HDACs (yeast Rpd3-like proteins: HDAC1-3, and HDAC8), class II HDACs with a single deacetylase domain at the C-terminus (yeast Hda1-like proteins: HDAC4-7, and HDAC9-10), class 3 HDACs (yeast silent information regulator 2 (Sir2)like proteins: Sirtuin 1–7), and class IV $(HDAC11)^{162}$. Given the potent antitumour effects of HDAC inhibitors (HDACi) to reverse the epigenetic changes in cancer by restoring the balance of histone acetylation¹⁶⁴. HDACi, thus, is a potent inducer of growth arrest and apoptosis and inhibit the differentiation of malignant cells¹⁶⁷.

Compound 5x is an HDAC2 inhibitor, one of the N-hydroxy-3-[3-(1-substituted-1H-benzoimidazol-2-yl)-phenyl]-acrylamide analogues, which displayed effective anticancer activities in vitro and *in vivo* in nanomolar range³¹. Compound 5x induced the hyperacetylation of histone H3 and H4 as well as p21 activity, which led to tumour suppression in HCT116 colorectal cancer and PC3 prostate xenograft models³¹. Pracinostat (SB939) dialkyl benzimidazole competitive histone deacetylase inhibitor targeting Class I, II and IV HDACs¹⁶⁸. Combination treatment of Pracinostat with azacytidine has shown synergistic effects against acute mveloid leukaemia (AML) in phase II clinical trial, with reported tolerable safety and efficacy. However, common adverse effects such as infection, thrombocytopenia and febrile neutropenia were present in patients¹⁶⁹. Regrettably, the phase III clinical trial of Pracinostat with azacytidine in AML was discontinued because the treatment outcome was unlikely to meet the primary endpoint of overall survival¹⁷⁰.

Sirtuins are a family of class III histone deacetylases which rely on nicotinamide adenine dinucleotide (NAD⁺) to function^{171,172}. Sirtuin 1–7 perform mainly as lysine deacetylase and/ or mono-ADP-ribosyltransferase on both histone and non-histone



Figure 8 Benzimidazole derivatives as (i) aromatase inhibitor and (ii) dihydrofolate reductase (DHFR) inhibitor.

proteins^{173,174}. Accumulating evidence has demonstrated the crucial role of sirtuin in cancer initiation and progression, thereby making sirtuins a target of interest in anticancer therapy^{173,175–178}. BZD9L1 is a newly discovered sirtuin inhibitor with autofluorescence and anticancer properties (Fig. 7)¹⁷⁹. BZD9L1 is a diversified 1,2-disubstituted benzimidazole analogue targeting both SIRT1 (IC₅₀: 42.9 µmol/L) and SIRT2 (IC₅₀: 9 µmol/L) proteins but showed higher inhibitory potency towards SIRT2 protein¹⁷⁹. The substitution of the piperidinyl group, which is a fundamental and strong electron-donating side chain, at the phenyl ring builds the foundation for SIRT inhibitory activity. This piperidinyl side-chain stabilises the benzimidazole moiety and allows a stronger interaction with the active site of SIRT protein¹⁷⁹. BZD9L1 has demonstrated comparable SIRT1 and/or SIRT2 inhibitory effects to commercially available sirtuin inhibitors such as AGK-2, EX527, and Tenovin-6. A functional study by Tan et al. has revealed its anticancer capabilities either as stand-alone or in combination with 5-fluorouracil (5-FU), the firstline chemotherapy for colorectal cancer, in vitro and in vivo^{32,33}. Tan and colleagues have also reported the ability of BZD9L1 to impede colorectal cancer cell viability, proliferation, migration, invasion, and induce apoptosis in vitro at the IC₅₀ concentration of 16.82 and 20.11 µmol/L in HCT116 and HT-29 colorectal cancer cell lines respectively³². Combining BZD9L1 with 5-FU further enhanced the treatment efficiency by hindering colorectal tumour growth through increased cell survival, cell cycle arrest, apoptosis, senescence and micronucleated³³. Furthermore, BZD9L1 was predicted to modulate the p53-dependent signalling pathways to exert cell death in CRC cells¹⁸⁰.

DNA demethylation removes the methyl group from cytosine by ten-eleven translocation methylcytosine dioxygenases (TET) to produce 5-formylcytosine and 5-carboxylcytosine through the oxidation process¹⁸¹. The histone lysine demethylase (KDM) subfamily contains the catalytic Jumanji C domain and utilizes Fe²⁺ as a cofactor and α -ketoglutarate [α -KG or 2-oxoglutarate (2OG)] as a co-substrate to remove methyl groups from histone lvsine residues^{34,182}. KDMs are overexpressed in various cancers, including breast, prostate, pancreatic, lung, colorectal, liver, glioma, gynaecological, oesophageal and lymphatic cancers, resulting unaggressive phenotypes^{34,183,184}. Compound **15** is a benzimidazole pyrazole-based inhibitor of recombinant lysine demethylase 4E (KDM4E), modified from its parental structure CBN209350 by introducing a non-polar aromatic ringside chain at the scaffold R₂ position, that resulted in 10-fold improved potency relative to the original HTS hit³⁴. Crystallography data showed that compound 15 competes with the enzyme for Fe^{2+} binding leading to the removal of active-site iron, thereby inhibiting KDM4E activity. Treatment of LnCaP and DU145 prostate cancer cells with compound 15 also revealed a significant reduction of the H3K9me3 compared to untreated cells³⁴.

2. Repurposing FDA-approved benzimidazole-derived drugs for cancer treatment

Drug repositioning is a valuable alternative strategy in drug discovery due to establishing well-documented pharmacokinetic, pharmacodynamic, and safety profiles of drug candidates, which may speed up the traditional process of approval and utilization in clinics¹⁸⁵. Intriguingly, many benzimidazole anthelmintic drugs have been reprofiled as anticancer agents; these examples include fenbendazole (FBZ), carbendazim (CBZ), oxibendazole, mebendazole (MBZ), albendazole (ABZ), and parbendazole (Fig. 10)^{185,186}. For instance, FBZ, and MBZ possess the ability to bind with the colchicine binding site, which can lead to the reduction of angiogenesis and multidrug-resistance in cancer cells^{187,188}. Moreover, parbendazole, oxibendazole, FBZ and MBZ have also reduced pancreatic cancer cell viability in the nanomolar range¹⁸⁹. Furthermore, Parbendazole possesses a linear



Figure 9 Benzimidazole derivatives used as epigenetic modulator.

alkylic side-chain that provides a strong antiproliferative benefit and results in the drastic inhibition of pancreatic cancer cell growth, survival, migration and induced DNA damage response, cell cycle arrest, and apoptosis¹⁸⁹. In addition, combination treatment of parbendazole with gemcitabine also enhanced the treatment effects in pancreatic cancer cell lines¹⁸⁹.

An investigation of the anticancer activities of these compounds on Kirsten rat sarcoma 2 viral oncogene homolog (KRAS)-wildtype and KRAS-mutant lung cancer in vitro and in vivo has revealed that lung cancer cells harbouring the KRAS mutation showed increased sensitivity towards benzimidazole derivatives. Methiazole and FBZ were used as TK inhibitors and showed enhanced inhibitory effects on KRAS-mutant lung cancer model in vitro and in vivo, inhibiting RAS signalling-related TK and suppressing the PI3K/AKT and RAF/MEK/ERK pathway, causing inhibition of proliferation and inhibition of apoptosis³⁰. From the study, methiazole was cytotoxic towards cancer cell lines harbouring KRAS mutation with an IC50 of 1.9 and 0.6 µmol/L in A549 and H23 lung cancer cells, respectively, while an $IC_{50} > 40 \ \mu mol/L$ was reported in KRAS wildtype cancer cells, without affecting the normal lung epithelial cells. On the other hand, FBZ exhibited higher sensitivity in KRAS mutant than KRAS wildtype cells (i.e., IC₅₀ in KRAS-mutant A-549: 1.5 µmol/L; H-23: 0.4 µmol/L, and KRAS-wildtype: H-1650: 6.2 µmol/L, H-2228: 7.8 µmol/L) but slightly cytotoxic towards normal lung epithelial cells³⁰.

Methyl 2-benzimidazolecarbamate (CBZ) is a well-known fungicide with a high binding affinity toward mammalian tubulin (K_d , 42.8 ± 4.0 µmol/L) in the MCF7 human breast cancer cells¹⁹⁰. CBZ has also been highlighted to hamper cell

proliferation in B16 melanoma (IC₅₀ = $8.5 \mu mol/L$) and HT-29 colon carcinoma (IC₅₀ = 9.5 μ mol/L) cell lines¹⁹¹. The anticancer potential of CBZ has also been shown to extend to the human breast (MCF-7, MX-1), pancreas (Panc-1, MiaPaCa), colon (ht-29), lung (A549, SK-MES), prostate (DU145), and murine (B16, p388) tumour models^{191,192}. CBZ functions by interfering with the formation of microtubules to instigate cell cycle arrest and apoptosis^{191,192}. CBZ was reported to have no interaction with the colchicine binding site but suppressed the spindle microtubule dynamic at interphase, resulting in mitotic arrest and cancer cell death¹⁹⁰. Another popular benzimidazolederived veterinary drug, FBZ (methyl N-(6-phenylsulfanyl-1Hbenzimidazol-2-yl) carbamate) can induce microtubule depolymerising activity in non-small cell lung carcinoma cells, ensuing in early G2/M arrest accompanied by p53-mediated cell death in both *in vitro* and *in vivo* studies⁷². In addition, oral treatment of FBZ in lung cancer xenografts demonstrated tumour vascularisation reduction and tumour growth inhibition⁷². Furthermore, FBZ has been shown to interfere with cellular microtubule polymerisation-impeded lymphoma tumour growth in vivo when used in combination with supplemented vitamins¹⁹³.

MBZ demonstrated a wide range of anticancer mechanisms by inhibiting tubulin polymerization, angiogenesis, pro-survival pathways, matrix metalloproteinases, and multi-drug resistance protein transporters in various cancer cell lines and xenograft models, which is extensively reviewed by Guerini and colleagues¹⁹⁴. However, MBZ has demonstrated low toxicity but possesses poor oral bioavailability (17%–20% of the dose absorption)¹⁹⁴. There is no evidence of MBZ in a clinical trial for cancer treatment, but many studies on MBZ alone or in



Methiazole

Lung cancer cells: 0.6-1.9 µmol/L



Fenbendazole

I C₆₀ Lung cancer cells: 0.4-1.5 μmol/L Pancreatic cancer cells: 2.66-3.26 μmol/L Paraganglioma cells: 0.10-0.15 μmol/L Colorectal cancer cells: 0.02-0.78 μmol/L



Mebendazole IC₅₀ Paraganglioma cells: 0.01-0.19 µmol/L Pancreatic cancer cells: 0.08-0.40 µmol/L Colorectal cancer cells: 0.08-1.26 µmol/L

H₃C S NH O CI

Albendazole

IC₆₀ Paraganglioma cells: 0.05-0.29 μmol/L Pancreatic cancer cells: 0.10-0.19 μmol/L Colorectal cancer cells: 0.17-0.28 μmol/L



Carbendazim

I C₅₀ Melanoma cells: 8.5 μmol/L Colorectal cancer cells: 9.5 μmol/L



Parbendazole

IC₆₀ Paraganglioma cells: 0.04-0.17 μmol/L Pancreatic cancer cells: 0.03-0.58 μmol/L Colorectal cancer cells: 0.01-0.57 μmol/L



Oxibendazole

IC₅₀ Paraganglioma cells: 1.79-3.29 μmol/L Pancreatic cancer cells: 1.07-1.45 μmol/L Colorectal cancer cells: 0.01-0.96 μmol/L

Figure 10 Repurposed benzimidazole derivatives as anticancer agents.

combination treatments have been registered for anticancer clinical trials¹⁹⁴.

Both ABZ and FBZ work well as antimitotic agents and regulators of ATP production $^{72,195-197}$, and have therefore been repositioned as anticancer candidates. ABZ has been reported to hinder cell proliferation in leukaemia, and breast, ovarian and liver malignancies¹⁹⁸⁻²⁰⁰. In addition, the role of ABZ in driving cells towards intrinsic apoptosis in Epothilone-paclitaxel resistant leukemic cells has been documented. The mechanism contributing to this event was the upregulation of cleaved caspase-3 and cytochrome C, and the downregulation of BCL-2 anti-apoptotic proteins tightly controlled by the p53 cellular gatekeeper²⁰¹. A recent study by Castro et al.²⁰⁰, also reported that ABZ could suppress breast cancer cell proliferation in vitro, and trigger p53dependent apoptosis in mouse Ehrlich mammary tumour cells in vivo. Another study by Dogra and colleagues⁷² reported that FBZ could disrupt microtubule dynamics through p53 activation in vitro and in vivo in human NSCLC cells. These findings corroborate the latest study by Mrkvová and colleagues²⁰², in which the inhibition of MDM2 and MDMX by ABZ and FBZ eventually led to p53 activation. In this study, Mrkvová et al.²⁰³ highlighted the ability of ABZ and FBZ to inhibit MDM2 and MDMX, through which a subsequent increase in p53 protein (by 2.5 and 1.3-folds respectively) was observed. MDM2 and MDMX are known as p53 regulators, which negatively regulate and abrogate p53 activities by promoting p53 proteasome-mediated degradation. The inhibition of p53 prevents cancer cells from cell cycle arrest and thus promotes cell survival.

3. Conclusions and future perspectives

Benzimidazole is a promising compound for anticancer either through target-specific or non-oncogene-specific targeting. Numerous benzimidazole-derived drugs have recently gained approval by the FDA, delineating the remarkable potential of benzimidazole scaffolds to be employed as anticancer agents. The advancement of science and technology has revolutionized healthcare and necessitated the development of more precise targeted drugs. The emergence of benzimidazole-derived PARP inhibitor: Rucaparib was redefined and approved by the FDA as one of the personalized medicines in 2020²⁰⁴. With the unique structure of benzimidazole core that exhibits a broad spectrum of bioactivities, it could potentially mimic naturally occurring nucleotides to interrupt biological processes in cancer and thereby potentially hijack multiple cellular processes at any one time. The enormous literature on the imperative roles of benzimidazole derivatives in targeting cancers also supports the use of benzimidazole scaffold in the transition from conventional to precision medicine.

Cancers are associated with multiple dysfunctions in genes or proteins in which multitargeted drugs may have an added advantage over non-target-specific drugs. Some benzimidazole drugs have been shown to target multiple biomarkers, which may improve the treatment response and result in the synergistic impediment of tumour growth. Although evidence has acknowledged the promising outcomes of multi-targeted therapies, the underlying possibility of inducing antagonist effect on tumour progression and offtarget toxicity cannot be ruled out. Additional downstream analyses need to be undertaken to investigate further the molecular mechanism and pathway *in vitro* and *in vivo*. Inevitably, problems like toxicity, drug-resistant and poor bioavailability have become the limiting factors in the development of small molecules in general. Thus, lead optimization is needed to generate drug candidates that are highly specific, less toxic, and with improved bioavailability. Due to the exorbitant costs in drug discovery and development, it may be reasonable to repurpose the currently available benzimidazole drugs to be developed in the cancer field as increasing evidence has demonstrated the therapeutic switching of benzimidazole drugs used to treat other diseases to demonstrate anticancer effects. This review could provide insight into the broad application of benzimidazole in targeting cancer, which may open up new avenues for benzimidazole anticancer drug development toward precision medicine.

Acknowledgments

We acknowledge the Ministry of Higher Education Malaysia for the Fundamental Research Grant Scheme with the Project Code: FRGS/1/2021/SKK06/USM/02/7 for supporting this work. We would also like to thank Lim Wei Khei from the School of Pharmacy and Biomedical Sciences at Curtin University, Australia, for proofreading the manuscript.

Author Contributions

Yeuan Ting Lee drafted the manuscript. Yi Jer Tan performed the structural analysis for ligand-target interaction. Chern Ein Oon reviewed and edited the manuscript. All authors read and approved the final manuscript.

Conflicts of interest

The authors have no conflicts of interest to declare.

References

- 1. Hodson R. Precision medicine. Nature 2016;537:S49.
- 2. Wright JB. The chemistry of the benzimidazoles. *Chem Rev* 1951;48: 397–541.
- Keri RS, Hiremathad A, Budagumpi S, Nagaraja BM. Comprehensive review in current developments of benzimidazole-based medicinal chemistry. *Chem Biol Drug Des* 2015;86:19–65.
- Singh PK, Silakari O. Benzimidazole: journey from single targeting to multitargeting molecule. In: Silakari O, editor. *Key heterocycle cores for designing multitargeting molecules*. Amsterdam: Elsevier; 2018.
- Vitaku E, Smith DT, Njardarson JT. Analysis of the structural diversity, substitution patterns, and frequency of nitrogen heterocycles among U.S. FDA approved pharmaceuticals. *J Med Chem* 2014;57: 10257–74.
- Gaba M, Mohan C. Development of drugs based on imidazole and benzimidazole bioactive heterocycles: recent advances and future directions. *Med Chem Res* 2016;25:173–210.
- El-masry HA, Fahmy HH, Ali Abdelwahed HS. Synthesis and antimicrobial activity of some new benzimidazole derivatives. *Molecules* 2000;5:1429–38.
- Ansari KF, Lal C. Synthesis, physicochemical properties and antimicrobial activity of some new benzimidazole derivatives. *Eur J Med Chem* 2009;44:4028–33.
- **9.** Padalkar VS, Borse BN, Gupta VD, Phatangare KR, Patil VS, Umape PG, et al. Synthesis and antimicrobial activity of novel 2-substituted benzimidazole, benzoxazole and benzothiazole derivatives. *Arab J Chem* 2016;**9**:S1125–30.
- Tahlan S, Kumar S, Ramasamy K, Lim SM, Shah SAA, Mani V, et al. Design, synthesis and biological profile of heterocyclic

benzimidazole analogues as prospective antimicrobial and antiproliferative agents. *BMC Chemistry* 2019;**13**:50.

- 11. Kumar K, Awasthi D, Lee SY, Zanardi I, Ruzsicska B, Knudson S, et al. Novel trisubstituted benzimidazoles, targeting Mtb FtsZ, as a new class of antitubercular agents. *J Med Chem* 2011;**54**: 374–81.
- Yoon YK, Ali MA, Choon TS, Ismail R, Wei AC, Kumar RS, et al. Antituberculosis: synthesis and antimycobacterial activity of novel benzimidazole derivatives. *BioMed Res Int* 2013;2013:926309.
- Desai NC, Shihory NR, Kotadiya GM, Desai P. Synthesis, antibacterial and antitubercular activities of benzimidazole bearing substituted 2-pyridone motifs. *Eur J Med Chem* 2014;82:480–9.
- Zou R, Ayres KR, Drach JC, Townsend LB. Synthesis and antiviral evaluation of certain disubstituted benzimidazole ribonucleosides. J Med Chem 1996;39:3477–82.
- Tonelli M, Simone M, Tasso B, Novelli F, Boido V, Sparatore F, et al. Antiviral activity of benzimidazole derivatives. Ii. Antiviral activity of 2-phenylbenzimidazole derivatives. *Bioorg Med Chem* 2010;18: 2937–53.
- 16. Vausselin T, Séron K, Lavie M, Mesalam AA, Lemasson M, Belouzard S, et al. Identification of a new benzimidazole derivative as an antiviral against hepatitis C virus. *J Virol* 2016;90:8422.
- Noor A, Qazi NG, Nadeem H, Khan AU, Paracha RZ, Ali F, et al. Synthesis, characterization, anti-ulcer action and molecular docking evaluation of novel benzimidazole-pyrazole hybrids. *Chem Cent J* 2017;11:85.
- Radhamanalan R, Alagumuthu M, Nagaraju N. Synthesis and drug efficacy validations of racemic-substituted benzimidazoles as antiulcer/antigastric secretion agents. *Future Med Chem* 2018;10: 1805–20.
- Arora RK, Kaur N, Bansal Y, Bansal G. Novel coumarinbenzimidazole derivatives as antioxidants and safer antiinflammatory agents. *Acta Pharm Sin B* 2014;4:368–75.
- 20. Sharma R, Bali A, Chaudhari BB. Synthesis of methanesulphonamido-benzimidazole derivatives as gastro-sparing antiinflammatory agents with antioxidant effect. *Bioorg Med Chem Lett* 2017;27:3007–13.
- Can NÖ, Çevik UA, Sağlık BN, Özkay Y, Atlı Ö, Baysal M, et al. Pharmacological and toxicological screening of novel benzimidazole-morpholine derivatives as dual-acting inhibitors. *Molecules* 2017;22:1374.
- Shingalapur RV, Hosamani KM, Keri RS, Hugar MH. Derivatives of benzimidazole pharmacophore: synthesis, anticonvulsant, antidiabetic and DNA cleavage studies. *Eur J Med Chem* 2010;45:1753–9.
- 23. El Bakri Y, Anouar EH, Marmouzi I, Sayah K, Ramli Y, El Abbes Faouzi M, et al. Potential antidiabetic activity and molecular docking studies of novel synthesized 3.6-dimethyl-5-oxo-pyrido[3,4-f][1,2,4] triazepino[2,3-a]benzimidazole and 10-amino-2-methyl-4-oxo pyrimido[1,2-a]benzimidazole derivatives. J Mol Model 2018;24:179.
- 24. Zhang Y, Xu J, Li Y, Yao H, Wu X. Design, synthesis and pharmacological evaluation of novel no-releasing benzimidazole hybrids as potential antihypertensive candidate. *Chem Biol Drug Des* 2015;**85**: 541–8.
- Torres-Gómez H, Hernández-Núñez E, León-Rivera I, Guerrero-Alvarez J, Cedillo-Rivera R, Moo-Puc R, et al. Design, synthesis and *in vitro* antiprotozoal activity of benzimidazole-pentamidine hybrids. *Bioorg Med Chem Lett* 2008;18:3147–51.
- 26. Toro P, Klahn AH, Pradines B, Lahoz F, Pascual A, Biot C, et al. Organometallic benzimidazoles: synthesis, characterization and antimalarial activity. *Inorg Chem Commun* 2013;35:126–9.
- 27. Okombo J, Brunschwig C, Singh K, Dziwornu GA, Barnard L, Njoroge M, et al. Antimalarial pyrido[1,2-*a*]benzimidazole derivatives with mannich base side chains: synthesis, pharmacological evaluation, and reactive metabolite trapping studies. ACS Infect Dis 2019;5:372–84.
- Kamanna K. Synthesis and pharmacological profile of benzimidazoles. In: Marinescu M, editor. *Chemistry and applications of benzimidazole and its derivatives*. London: IntchOpen; 2019.

- 29. Shrivastava N, Naim MJ, Alam MJ, Nawaz F, Ahmed S, Alam O. Benzimidazole scaffold as anticancer agent: synthetic approaches and structure—activity relationship. *Arch Pharmazie* 2017;350: e201700040.
- **30.** Shimomura I, Yokoi A, Kohama I, Kumazaki M, Tada Y, Tatsumi K, et al. Drug library screen reveals benzimidazole derivatives as selective cytotoxic agents for KRAS-mutant lung cancer. *Cancer Lett* 2019;**451**:11–22.
- Bressi JC, Jong Rd, Wu Y, Jennings AJ, Brown JW, O'Connell S, et al. Benzimidazole and imidazole inhibitors of histone deacetylases: synthesis and biological activity. *Bioorg Med Chem Lett* 2010; 20:3138–41.
- Tan YJ, Lee YT, Yeong KY, Petersen SH, Kono K, Tan SC, et al. Anticancer activities of a benzimidazole compound through sirtuin inhibition in colorectal cancer. *Future Med Chem* 2018;10:2039–57.
- 33. Tan YJ, Lee YT, Petersen SH, Kaur G, Kono K, Tan SC, et al. BZD9L1 sirtuin inhibitor as a potential adjuvant for sensitization of colorectal cancer cells to 5-fluorouracil. *Ther Adv Med Oncol* 2019; 11:1758835919878977.
- **34.** Carter DM, Specker E, Małecki PH, Przygodda J, Dudaniec K, Weiss MS, et al. Enhanced properties of a benzimidazole benzylpyrazole lysine demethylase inhibitor: mechanism-of-action, binding site analysis, and activity in cellular models of prostate cancer. *J Med Chem* 2021;**64**:14266–82.
- Haider K, Yar MS. Advances of benzimidazole derivatives as anticancer agents: bench to bedside. In: Kendrekar P, Adimule V, editors. *Benzimidazole*. London: IntechOpen; 2022.
- Champoux JJ. DNA topoisomerases: structure, function, and mechanism. Annu Rev Biochem 2001;70:369–413.
- 37. Lodish H, Berk A, Zipursky SL, Matsudaira P, Baltimore D, Darnell J. The role of topoisomerases in DNA replication. In: Osheroff N, Bjornsti MA, editors. DNA topoisomerase protocols: Volume II: enzymology and drugs. Totowa: Humans Press; 2001.
- McClendon AK, Osheroff N. DNA topoisomerase ii, genotoxicity, and cancer. *Mutat Res* 2007;623:83–97.
- **39.** Delgado JL, Hsieh CM, Chan NL, Hiasa H. Topoisomerases as anticancer targets. *Biochem J* 2018;**475**:373–98.
- 40. Bielawski K, Wolczynski S, Bielawska A. Inhibition of DNA topoisomerase i and ii, and growth inhibition of MDA-MB-231 human breast cancer cells by bis-benzimidazole derivatives with alkylating moiety. *Pol J Pharmacol* 2004;56:373–8.
- Oksuzoglu E, Tekiner-Gulbas B, Alper S, Temiz-Arpaci O, Ertan T, Yildiz I, et al. Some benzoxazoles and benzimidazoles as DNA topoisomerase i and ii inhibitors. *J Enzym Inhib Med Chem* 2008;23:37–42.
- 42. Hegde M, Kumar KSS, Thomas E, Ananda H, Raghavan SC, Rangappa KS. A novel benzimidazole derivative binds to the DNA minor groove and induces apoptosis in leukemic cells. *RSC Adv* 2015;5:93194–208.
- 43. Gao C, Li B, Zhang B, Sun Q, Li L, Li X, et al. Synthesis and biological evaluation of benzimidazole acridine derivatives as potential DNA-binding and apoptosis-inducing agents. *Bioorg Med Chem* 2015;23:1800–7.
- 44. Li P, Zhang W, Jiang H, Li Y, Dong C, Chen H, et al. Design, synthesis and biological evaluation of benzimidazole–rhodanine conjugates as potent topoisomerase ii inhibitors. *MedChemComm* 2018;9:1194–205.
- 45. Gromiha MM, Nagarajan R. Computational approaches for predicting the binding sites and understanding the recognition mechanism of protein–DNA complexes. In: Gromiha MM, editor. *Protein interactions: computational methods, analysis and applications.* Danvers: World Scientific Publishing; 2020.
- 46. Bhaduri S, Ranjan N, Arya DP. An overview of recent advances in duplex DNA recognition by small molecules. *Beilstein J Org Chem* 2018;14:1051–86.
- Wong KC. A novel approach to predict core residues on cancerrelated DNA-binding domains. *Cancer Inf* 2016;15:1–7.
- Thirumal Kumar D, Mendonca E, Priyadharshini Christy J, George Priya Doss C, Zayed H. A computational model to predict the

structural and functional consequences of missense mutations in *O*⁶methylguanine DNA methyltransferase. In: Donev R, editor. *Advance in protein chemistry and structural biology*. Elsevier; 2019.

- **49.** Goftar MK, Kor NM, Kor ZM. Dna intercalators and using them as anticancer drugs. *Int J Adv Biol Biom Res* 2014;**2**:811–22.
- Soni A, Khurana P, Singh T, Jayaram B. A DNA intercalation methodology for an efficient prediction of ligand binding pose and energetics. *Bioinformatics* 2017;33:1488–96.
- Waring MJ. DNA modification and cancer. *Annu Rev Biochem* 1981; 50:159–92.
- 52. Yamori T, Matsunaga A, Sato S, Yamazaki K, Komi A, Ishizu K, et al. Potent antitumor activity of MS-247, a novel DNA minor groove binder, evaluated by an *in vitro* and *in vivo* human cancer cell line panel. *Cancer Res* 1999;**59**:4042.
- Rehman S, Sarwar T, Husain M, Ishqi H, Tabish M. Studying noncovalent drug–DNA interactions. Arch Biochem Biophys 2015;576: 49–60.
- Matsuba Y, Edatsugi H, Mita I, Matsunaga A, Nakanishi O. A novel synthetic DNA minor groove binder, MS-247: antitumor activity and cytotoxic mechanism. *Cancer Chemother Pharmacol* 2000;46:1–9.
- 55. Joubert A, Sun XW, Johansson E, Bailly C, Mann J, Neidle S. Sequence-selective targeting of long stretches of the DNA minor groove by a novel dimeric bis-benzimidazole. *Biochemistry* 2003;42: 5984–92.
- 56. Yang YH, Cheng MS, Wang QH, Nie H, Liao N, Wang J, et al. Design, synthesis, and anti-tumor evaluation of novel symmetrical bis-benzimidazoles. *Eur J Med Chem* 2009;44:1808–12.
- Gandhi V, Burger JA. Bendamustine in B-cell malignancies: the new 46-year-old kid on the block. *Clin Cancer Res* 2009;15:7456.
- 58. Leoni LM, Bailey B, Reifert J, Bendall HH, Zeller RW, Corbeil J, et al. Bendamustine (treanda) displays a distinct pattern of cytotoxicity and unique mechanistic features compared with other alkylating agents. *Clin Cancer Res* 2008;14:309–17.
- **59.** Leong H, Bonk ME. Bendamustine (treanda) for chronic lymphocytic leukemia: a brief overview. *P T* 2009;**34**:73–6.
- Cephalon Treanda® (bendamustine hydrochloride). Available from: https://www.accessdata.fda.gov/drugsatfda_docs/label/2008/ 022303lbl.pdf.
- Rashid M, Husain A, Mishra R. Synthesis of benzimidazoles bearing oxadiazole nucleus as anticancer agents. *Eur J Med Chem* 2012;54: 855–66.
- **62.** Husain A, Rashid M, Shaharyar M, Siddiqui AA, Mishra R. Benzimidazole clubbed with triazolo-thiadiazoles and triazolo-thiadiazines: new anticancer agents. *Eur J Med Chem* 2013;**62**:785–98.
- 63. Rashid M, Husain A, Mishra R, Karim S, Khan S, Ahmad M, et al. Design and synthesis of benzimidazoles containing substituted oxadiazole, thiadiazole and triazolo-thiadiazines as a source of new anticancer agents. *Arab J Chem* 2015;12:3203–24.
- **64.** Kamal A, Praveen Kumar P, Sreekanth K, Seshadri BN, Ramulu P. Synthesis of new benzimidazole linked pyrrolo[2,1-*c*][1,4]benzodiazepine conjugates with efficient DNA-binding affinity and potent cytotoxicity. *Bioorg Med Chem Lett* 2008;**18**:2594–8.
- 65. Al-Mudaris ZA, Majid ASA, Ji D, Al-Mudarris BA, Chen SH, Liang PH, et al. Conjugation of benzylvanillin and benzimidazole structure improves DNA binding with enhanced antileukemic properties. *PLoS One* 2013;8:e80983.
- Jordan MA, Wilson L. Microtubules as a target for anticancer drugs. Nat Rev Cancer 2004;4:253–65.
- Jordan MA, Wilson L. Microtubule dynamics. In: Fojo T, editor. *The role of microtubules in cell biology, neurobiology, and oncology.* Totowa, NJ: Humana Press; 2008.
- Azarenko O, Okouneva T, Singletary KW, Jordan MA, Wilson L. Suppression of microtubule dynamic instability and turnover in MCF7 breast cancer cells by sulforaphane. *Carcinogenesis* 2008;29: 2360-8.
- Bates D, Eastman A. Microtubule destabilising agents: far more than just antimitotic anticancer drugs. Br J Clin Pharmacol 2017;83: 255–68.

- **70.** Wang TH, Popp DM, Wang HS, Saitoh M, Mural JG, Henley DC, et al. Microtubule dysfunction induced by paclitaxel initiates apoptosis through both c-Jun M-terminal kinase (JNK)-dependent and -independent pathways in ovarian cancer cells. *J Biol Chem* 1999;**274**:8208–16.
- Gascoigne KE, Taylor SS. Cancer cells display profound intra- and interline variation following prolonged exposure to antimitotic drugs. *Cancer Cell* 2008;14:111–22.
- 72. Dogra N, Kumar A, Mukhopadhyay T. Fenbendazole acts as a moderate microtubule destabilizing agent and causes cancer cell death by modulating multiple cellular pathways. *Sci Rep* 2018;8: 11926.
- 73. Karthikeyan C, Solomon VR, Lee H, Trivedi P. Synthesis and biological evaluation of 2-(phenyl)-3*H*-benzo[*d*]imidazole-5-carboxylic acids and its methyl esters as potent anti-breast cancer agents. *Arab J Chem* 2017;10:S1788–94.
- 74. Hasanpourghadi M, Pandurangan AK, Karthikeyan C, Trivedi P, Mustafa MR. Mechanisms of the anti-tumor activity of methyl 2-(-5fluoro-2-hydroxyphenyl)-1*H*-benzo[*d*]imidazole-5-carboxylate against breast cancer *in vitro* and *in vivo*. *Oncotarget* 2017;8: 28840–53.
- 75. Hasanpourghadi M, Karthikeyan C, Pandurangan AK, Looi CY, Trivedi P, Kobayashi K, et al. Targeting of tubulin polymerization and induction of mitotic blockage by methyl 2-(5-fluoro-2hydroxyphenyl)-1*H*-benzo[*d*]imidazole-5-carboxylate (MBIC) in human cervical cancer hela cell. J Exp Clin Cancer Res 2016;35:58.
- 76. Dai X, Wang L, Deivasigamni A, Looi CY, Karthikeyan C, Trivedi P, et al. A novel benzimidazole derivative, MBIC inhibits tumor growth and promotes apoptosis *via* activation of ROS-dependent JNK signaling pathway in hepatocellular carcinoma. *Oncotarget* 2017;8: 12831–42.
- 77. Choi H, Fukui M, Zhu B. Role of cyclin B1/CDC2 up-regulation in the development of mitotic prometaphase arrest in human breast cancer cells treated with nocodazole. *PLoS One* 2011;6:e24312.
- **78.** Blajeski AL, Phan VA, Kottke TJ, Kaufmann SH. G₁ and G₂ cellcycle arrest following microtubule depolymerization in human breast cancer cells. *J Clin Investig* 2002;**110**:91–9.
- **79.** Soto JD. The comparative effectiveness of parp1 inhibitors alone or in combination against sporadic cancer. *FASEB J* 2011;**25**:620–3.
- 80. Kamal A, Rao AV, Nayak VL, Reddy NV, Swapna K, Ramakrishna G, et al. Synthesis and biological evaluation of imidazo[1,5-a]pyridine-benzimidazole hybrids as inhibitors of both tubulin polymerization and PI3K/ALT pathway. *Org Biomol Chem* 2014;12:9864–80.
- Zhong L, Li Y, Xiong L, Wang W, Wu M, Yuan T, et al. Small molecules in targeted cancer therapy: advances, challenges, and future perspectives. *Signal Transduct Targeted Ther* 2021;6:201.
- Paul MK, Mukhopadhyay AK. Tyrosine kinase-role and significance in cancer. Int J Med Sci 2004;1:101–15.
- Gianoncelli A, Cozza G, Orzeszko A, Meggio F, Kazimierczuk Z, Pinna LA. Tetraiodobenzimidazoles are potent inhibitors of protein kinase CK2. *Bioorg Med Chem* 2009;17:7281–9.
- Kubiński K, Masłyk M, Orzeszko A. Benzimidazole inhibitors of protein kinase CK2 potently inhibit the activity of atypical protein kinase Rio1. *Mol Cell Biochem* 2017;426:195–203.
- Koronkiewicz M, Zukowska M, Chilmonczyk Z, Orzeszko A, Kazimierczuk Z. Synthesis and proapoptotic properties of new casein kinase ii inhibitors. *Acta Pol Pharm* 2010;67:635–41.
- 86. Trudel S, Li ZH, Wei E, Wiesmann M, Chang H, Chen C, et al. Chir-258, a novel, multitargeted tyrosine kinase inhibitor for the potential treatment of t(4;14) multiple myeloma. *Blood* 2005;105:2941–8.
- 87. Azab AK, Azab F, Quang P, Maiso P, Sacco A, Ngo HT, et al. FGFR3 is overexpressed waldenstrom macroglobulinemia and its inhibition by dovitinib induces apoptosis and overcomes stroma-induced proliferation. *Clin Cancer Res* 2011;**17**:4389–99.
- Huynh H, Chow PK, Tai WM, Choo SP, Chung AY, Ong HS, et al. Dovitinib demonstrates antitumor and antimetastatic activities in xenograft models of hepatocellular carcinoma. *J Hepatol* 2012;56: 595–601.

- Hasinoff BB, Wu X, Nitiss JL, Kanagasabai R, Yalowich JC. The anticancer multi-kinase inhibitor dovitinib also targets topoisomerase i and topoisomerase ii. *Biochem Pharmacol* 2012;84:1617–26.
- Precision Oncology News. FDA accepts allarity therapeutics' PMA for dovitinib companion test. Available from: https://www. precisiononcologynews.com/cancer/fda-accepts-allaritytherapeutics-pma-dovitinib-companion-test#.YXfFh55BzIW.
- **91.** Yeh TC, Marsh V, Bernat BA, Ballard J, Colwell H, Evans RJ, et al. Biological characterization of ARRY-142886 (AZD6244), a potent, highly selective mitogen-activated protein kinase kinase 1/2 inhibitor. *Clin Cancer Res* 2007;**13**:1576–83.
- Roskoski Jr R. Properties of FDA-approved small molecule protein kinase inhibitors: a 2021 update. *Pharmacol Res* 2021;165:105463.
- 93. Davies BR, Logie A, McKay JS, Martin P, Steele S, Jenkins R, et al. AZD6244 (ARRY-142886), a potent inhibitor of mitogen-activated protein kinase/extracellular signal-regulated kinase kinase 1/2 kinases: mechanism of action *in vivo*, pharmacokinetic/pharmacodynamic relationship, and potential for combination in preclinical models. *Mol Cancer Therapeut* 2007;6:2209–19.
- **94.** Gao JH, Wang CH, Tong H, Wen SL, Huang ZY, Tang CW. Targeting inhibition of extracellular signal-regulated kinase kinase pathway with AZD6244 (ARRY-142886) suppresses growth and angiogenesis of gastric cancer. *Sci Rep* 2015;**5**:16382.
- 95. Brown SR, Hall A, Buckley HL, Flanagan L, Gonzalez de Castro D, Farnell K, et al. Investigating the potential clinical benefit of selumetinib in resensitising advanced iodine refractory differentiated thyroid cancer to radioiodine therapy (SEL-I-METRY): protocol for a multicentre UK single arm phase ii trial. *BMC Cancer* 2019;19:582.
- 96. Markham A, Keam SJ. Selumetinib: first approval. Drugs 2020;80: 931–7.
- **97.** Lee PA, Wallace E, Marlow A, Yeh T, Marsh V, Anderson D, et al. Abstract 2515: preclinical development of ARRY-162, a potent and selective MEK 1/2 inhibitor. *Cancer Res* 2010;**70**:2515.
- **98.** Tran B, Cohen MS. The discovery and development of binimetinib for the treatment of melanoma. *Expet Opin Drug Discov* 2020;**15**: 745–54.
- **99.** Woodfield SE, Zhang L, Scorsone KA, Liu Y, Zage PE. Binimetinibinhibits mek and is effective against neuroblastoma tumor cells with low NF1 expression. *BMC Cancer* 2016;**16**:172.
- 100. U.S. Food and Drug Administration. FDA approves encorafenib and binimetinib in combination for unresectable or metastatic melanoma with BRAF mutations. Available from: https://www.fda.gov/drugs/ resources-information-approved-drugs/fda-approves-encorafeniband-binimetinib-combination-unresectable-or-metastatic-melanomabraf.
- 101. U.S. Food and Drug Administration. FDA approves abemaciclib with endocrine therapy for early breast cancer. Available from: https:// www.fda.gov/drugs/resources-information-approved-drugs/fdaapproves-abemaciclib-endocrine-therapy-early-breast-cancer.
- 102. Corona SP, Generali D. Abemaciclib: a cdk4/6 inhibitor for the treatment of HR⁺/HER2⁻ advanced breast cancer. *Drug Des Dev Ther* 2018;12:321–30.
- 103. Gelbert LM, Cai S, Lin X, Sanchez-Martinez C, Del Prado M, Lallena MJ, et al. Preclinical characterization of the CDK4/6 inhibitor ly2835219: *in-vivo* cell cycle-dependent/independent anti-tumor activities alone/in combination with gemcitabine. *Invest N Drugs* 2014;**32**:825–37.
- 104. Tan DS, Leighl NB, Riely GJ, Yang JC, Sequist LV, Wolf J, et al. Safety and efficacy of nazartinib (EGF816) in adults with egfrmutant non-small-cell lung carcinoma: a multicentre, open-label, phase 1 study. *Lancet Respir Med* 2020;8:561–72.
- 105. Lelais G, Epple R, Marsilje TH, Long YO, McNeill M, Chen B, et al. Discovery of (*R,E*)-*N*-(7-chloro-1-(1-[4-(dimethylamino)but-2enoyl]azepan-3-yl)-1*H*-benzo[*d*]imidazole-2-yl)-2methylisonicotinamide (EGF816), a novel, potent, and WT sparing covalent inhibitor of oncogenic (L858r, ex19del) and resistant (T790M) EGFR mutants for the treatment of EGFR mutant nonsmall-cell lung cancers. *J Med Chem* 2016;**59**:6671–89.

- 106. Jia Y, Juarez J, Li J, Manuia M, Niederst MJ, Tompkins C, et al. EGF816 exerts anticancer effects in non-small cell lung cancer by irreversibly and selectively targeting primary and acquired activating mutations in the egf receptor. *Cancer Res* 2016;**76**:1591–602.
- 107. Yamani A, Zdżalik-Bielecka D, Lipner J, Stańczak A, Piórkowska N, Stańczak PS, et al. Discovery and optimization of novel pyrazolebenzimidazole CPL304110, as a potent and selective inhibitor of fibroblast growth factor receptors FGFR (1–3). *Eur Med Chem* 2021; 210:112990.
- 108. Zhang QW, Ye ZD, Shen C, Tie HX, Wang L, Shi L. Synthesis of novel 6,7-dimethoxy-4-anilinoquinolines as potent c-Met inhibitors. *J Enzym Inhib Med Chem* 2019;34:124–33.
- 109. Ramurthy S, Subramanian S, Aikawa M, Amiri P, Costales A, Dove J, et al. Design and synthesis of orally bioavailable benzimidazoles as raf kinase inhibitors. J Med Chem 2008;51:7049–52.
- 110. Li Y, Tan C, Gao C, Zhang C, Luan X, Chen X, et al. Discovery of benzimidazole derivatives as novel multi-target EGFR, VEGFR-2 and PDGFR kinase inhibitors. *Bioorg Med Chem* 2011;19:4529–35.
- 111. Rumpf T, Schiedel M, Karaman B, Roessler C, North BJ, Lehotzky A, et al. Selective SIRT2 inhibition by ligand-induced rearrangement of the active site. *Nat Commun* 2015;6:6263.
- 112. Galal SA, Khairat SHM, Ali HI, Shouman SA, Attia YM, Ali MM, et al. Part II: new candidates of pyrazole-benzimidazole conjugates as checkpoint kinase 2 (CHK2) inhibitors. *Eur J Med Chem* 2018; 144:859–73.
- 113. Bistrović A, Krstulović L, Harej A, Grbčić P, Sedić M, Koštrun S, et al. Design, synthesis and biological evaluation of novel benzimidazole amidines as potent multi-target inhibitors for the treatment of non-small cell lung cancer. *Eur J Med Chem* 2018;143:1616–34.
- Jackson SP, Bartek J. The DNA-damage response in human biology and disease. *Nature* 2009;461:1071–8.
- 115. Morales J, Li L, Fattah FJ, Dong Y, Bey EA, Patel M, et al. Review of poly (ADP-ribose) polymerase (PARP) mechanisms of action and rationale for targeting in cancer and other diseases. *Crit Rev Eukaryot Gene Expr* 2014;24:15–28.
- 116. Henning RJ, Bourgeois M, Harbison RD. Poly(ADP-ribose) polymerase (PARP) and parp inhibitors: mechanisms of action and role in cardiovascular disorders. *Cardiovasc Toxicol* 2018;18:493–506.
- 117. Cerrato A, Morra F, Celetti A. Use of poly ADP-ribose polymerase [PARP] inhibitors in cancer cells bearing DDR defects: the rationale for their inclusion in the clinic. J Exp Clin Cancer Res 2016;35:179.
- Schiewer MJ, Knudsen KE. Transcriptional roles of parp1 in cancer. Mol Cancer Res 2014;12:1069–80.
- 119. Ba X, Garg NJ. Signaling mechanism of poly(ADP-ribose) polymerase-1 (PARP-1) in inflammatory diseases. *Am J Pathol* 2011;**178**: 946-55.
- O'Neil NJ, Bailey ML, Hieter P. Synthetic lethality and cancer. *Nat Rev Genet* 2017;18:613–23.
- 121. Min R, Wu W, Wang M, Tang L, Chen D, Zhao H, et al. Discovery of 2-(1-(3-(4-chloroxyphenyl)-3-oxo-propyl) pyrrolidine-3-yl)-1*H*benzo[*d*]imidazole-4-carboxamide: a potent poly(ADP-ribose) polymerase (PARP) inhibitor for treatment of cancer. *Molecules* 2019;24: 1901.
- 122. Jiang X, Li W, Li X, Bai H, Zhang Z. Current status and future prospects of parp inhibitor clinical trials in ovarian cancer. *Cancer Manag Res* 2019;11:4371.
- 123. Tong Y, Bouska JJ, Ellis PA, Johnson EF, Leverson J, Liu X, et al. Synthesis and evaluation of a new generation of orally efficacious benzimidazole-based poly(ADP-ribose) polymerase-1 (PARP-1) inhibitors as anticancer agents. *J Med Chem* 2009;**52**:6803–13.
- 124. Shirley M. Rucaparib: a review in ovarian cancer. *Targeted Oncol* 2019;**14**:237–46.
- 125. Zanjirband M, Curtin N, Edmondson RJ, Lunec J. Combination treatment with rucaparib (Rubraca) and MDM2 inhibitors, Nutlin-3 and RG7388, has synergistic and dose reduction potential in ovarian cancer. *Oncotarget* 2017;8:69779–96.
- 126. Plummer R, Jones C, Middleton M, Wilson R, Evans J, Olsen A, et al. Phase I study of the poly(ADP-ribose) polymerase inhibitor,

AG014699, in combination with temozolomide in patients with advanced solid tumors. *Clin Cancer Res* 2008;**14**:7917–23.

- 127. Drew Y, Ledermann JA, Jones A, Hall G, Jayson GC, Highley M, et al. Phase II trial of the poly(ADP-ribose) polymerase (PARP) inhibitor AG-014699 in *BRCA* 1 and 2-mutated, advanced ovarian and/or locally advanced or metastatic breast cancer. *J Clin Oncol* 2011;**29**:3104.
- 128. Patsouris A, Tredan O, Campion L, Goncalves A, Arnedos M, Sablin MP, et al. An open-label, phase II study of rucaparib, a PARP inhibitor, in HER2⁻ metastatic breast cancer patients with high genomic loss of heterozygosity. J Clin Oncol 2018;36:TPS1112-TPS.
- U.S. Food and Drug Administration. Rubraca® (rucaparib) tablets, for oral use. Available from: http://www.accessdata.fda.gov/ drugsatfda_docs/label/2018/209115s003lbl.pdf.
- European Commission. Rubraca: summary of product characteristics. Available from: http://www.ema.europa.eu/documents/productinformation/rubraca-epar-product-information_en.pdf.
- 131. Wagner LM. Profile of veliparib and its potential in the treatment of solid tumors. *Onco Target Ther* 2015;8:1931–9.
- 132. Palma JP, Wang YC, Rodriguez LE, Montgomery D, Ellis PA, Bukofzer G, et al. ABT-888 confers broad *in vivo* activity in combination with temozolomide in diverse tumors. *Clin Cancer Res* 2009;15:7277–90.
- 133. Murai J, Huang SN, Das BB, Renaud A, Zhang Y, Doroshow JH, et al. Trapping of PARP1 and PARP2 by clinical PARP inhibitors. *Cancer Res* 2012;**72**:5588.
- 134. Pommier Y, O'Connor MJ, de Bono J. Laying a trap to kill cancer cells: PARP inhibitors and their mechanisms of action. *Sci Transl Med* 2016;8:362ps17.
- 135. Chen X, Huan X, Liu Q, Wang Y, He Q, Tan C, et al. Design and synthesis of 2-(4,5,6,7-tetrahydrothienopyridin-2-yl)-benzoimidazole carboxamides as novel orally efficacious poly(ADP-ribose) polymerase (PARP) inhibitors. *Eur J Med Chem* 2018;**145**:389–403.
- Davey RA, Grossmann M. Androgen receptor structure, function and biology: from bench to bedside. *Clin Biochem Rev* 2016;37:3–15.
- 137. Ricciardelli C, Bianco-Miotto T, Jindal S, Butler LM, Leung S, McNeil CM, et al. The magnitude of androgen receptor positivity in breast cancer is critical for reliable prediction of disease outcome. *Clin Cancer Res* 2018;24:2328.
- Heinlein CA, Chang C. Androgen receptor in prostate cancer. *Endocr Rev* 2004;25:276–308.
- 139. Bleach R, McIlroy M. The divergent function of androgen receptor in breast cancer; analysis of steroid mediators and tumor intracrinology. *Front Endocrinol* 2018;9:594.
- 140. Fujita K, Nonomura N. Role of androgen receptor in prostate cancer: a review. World J Men's Health 2019;37:288–95.
- 141. Giovannelli P, Di Donato M, Galasso G, Di Zazzo E, Bilancio A, Migliaccio A. The androgen receptor in breast cancer. *Front Endocrinol* 2018;9:492.
- 142. Handratta VD, Vasaitis TS, Njar VC, Gediya LK, Kataria R, Chopra P, et al. Novel C-17-heteroaryl steroidal CYP17 inhibitors/antiandrogens: synthesis, *in vitro* biological activity, pharmacokinetics, and antitumor activity in the LAPC4 human prostate cancer xenograft model. *J Med Chem* 2005;48:2972–84.
- 143. Schayowitz A, Sabnis G, Njar VC, Brodie AM. Synergistic effect of a novel antiandrogen, VN/124-1, and signal transduction inhibitors in prostate cancer progression to hormone independence *in vitro*. *Mol Cancer Ther* 2008;7:121–32.
- 144. Purushottamachar P, Ramalingam S, Njar VC. Development of benzimidazole compounds for cancer therapy. In: Marinescu M, editor. *Chemistry and applications of benzimidazole and its derivatives*. London: IntechOpen; 2019.
- 145. McClurg UL, Azizyan M, Dransfield DT, Namdev N, Chit NCTH, Nakjang S, et al. The novel anti-androgen candidate galeterone targets deubiquitinating enzymes, USP12 and USP46, to control prostate cancer growth and survival. *Oncotarget* 2018;9:24992–5007.

- 146. Bastos DA, Antonarakis ES. Galeterone for the treatment of advanced prostate cancer: the evidence to date. *Drug Des Dev Ther* 2016;10:2289–97.
- 147. Kwegyir-Afful AK, Ramalingam S, Purushottamachar P, Ramamurthy VP, Njar VCO. Galeterone and VNPT55 induce proteasomal degradation of AR/AR-V7, induce significant apoptosis *via* cytochrome *c* release and suppress growth of castration resistant prostate cancer xenografts *in vivo. Oncotarget* 2015;6:27440–60.
- 148. McKay RR, Werner L, Fiorillo M, Roberts J, Heath EI, Bubley GJ, et al. Efficacy of therapies after galeterone in patients with castrationresistant prostate cancer. *Clin Genitourin Cancer* 2017;15:463–71.
- 149. Munuganti RSN, Leblanc E, Axerio-Cilies P, Labriere C, Frewin K, Singh K, et al. Targeting the binding function 3 (BF3) site of the androgen receptor through virtual screening. 2. Development of 2-((2-phenoxyethyl) thio)-1*H*-benzimidazole derivatives. *J Med Chem* 2013;56:1136-48.
- 150. Ng RA, Guan J, Alford VC, Lanter JC, Allan GF, Sbriscia T, et al. Synthesis and SAR of potent and selective androgen receptor antagonists: 5,6-dichloro-benzimidazole derivatives. *Bioorg Med Chem Lett* 2007;17:784–8.
- 151. Ng RA, Guan J, Alford VC, Lanter JC, Allan GF, Sbriscia T, et al. 2-(2,2,2-Trifluoroethyl)-5,6-dichlorobenzimidazole derivatives as potent androgen receptor antagonists. *Bioorg Med Chem Lett* 2007; 17:955–8.
- 152. Chumsri S, Howes T, Bao T, Sabnis G, Brodie A. Aromatase, aromatase inhibitors, and breast cancer. J Steroid Biochem Mol Biol 2011;125:13–22.
- 153. Molehin D, Rasha F, Rahman RL, Pruitt K. Regulation of aromatase in cancer. *Mol Cell Biochem* 2021;476:2449–64.
- **154.** Johnston SRD, Dowsett M. Aromatase inhibitors for breast cancer: lessons from the laboratory. *Nat Rev Cancer* 2003;**3**:821–31.
- 155. Acar Çevik U, Kaya Çavuşoğlu B, Sağlık BN, Osmaniye D, Levent S, Ilgın S, et al. Synthesis, docking studies and biological activity of new benzimidazole-triazolothiadiazine derivatives as aromatase inhibitor. *Molecules* 2020;25:1642.
- 156. Sağlık BN, Şen AM, Evren AE, Çevik UA, Osmaniye D, Çavuşoğlu BK, et al. Synthesis, investigation of biological effects and *in silico* studies of new benzimidazole derivatives as aromatase inhibitors. *Z Naturforsch, C: J Biosci* 2020;**75**:353–62.
- 157. Raimondi MV, Randazzo O, La Franca M, Barone G, Vignoni E, Rossi D, et al. DHFR inhibitors: reading the past for discovering novel anticancer agents. *Molecules* 2019;24:1140.
- 158. Zhao M, Tan B, Dai X, Shao Y, He Q, Yang B, et al. DHFR/TYMS are positive regulators of glioma cell growth and modulate chemosensitivity to temozolomide. *Eur J Pharmacol* 2019;863:172665.
- 159. Li Z, Wang Q, Zhang W, Yang Z, Li L. Cisplatin resistant effects of dihydrofolate reductase gene expression up-regulation in epithelial ovarian cancer. *Zhonghua Fu Chan Ke Za Zhi* 2015;50:854–60.
- 160. Organista-Nava J, Gómez-Gómez Y, Illades-Aguiar B, Rivera-Ramírez AB, Saavedra-Herrera MV, Leyva-Vázquez MA. Overexpression of dihydrofolate reductase is a factor of poor survival in acute lymphoblastic leukemia. *Oncol Lett* 2018;15:8405–11.
- 161. Singla P, Luxami V, Paul K. Triazine—benzimidazole hybrids: anticancer activity, DNA interaction and dihydrofolate reductase inhibitors. *Bioorg Med Chem* 2015;23:1691–700.
- **162.** Cheng Y, He C, Wang M, Ma X, Mo F, Yang S, et al. Targeting epigenetic regulators for cancer therapy: mechanisms and advances in clinical trials. *Signal Transduct Targeted Ther* 2019;**4**:62.
- 163. Lu Y, Chan YT, Tan HY, Li S, Wang N, Feng Y. Epigenetic regulation in human cancer: the potential role of epi-drug in cancer therapy. *Mol Cancer* 2020;19:79.
- 164. Hontecillas-Prieto L, Flores-Campos R, Silver A, de Álava E, Hajji N, García-Domínguez DJ. Synergistic enhancement of cancer therapy using HDAC inhibitors: opportunity for clinical trials. *Front Genet* 2020;11:578011.
- **165.** West AC, Johnstone RW. New and emerging HDAC inhibitors for cancer treatment. *J Clin Invest* 2014;**124**:30–9.

- **166.** Karagiannis D, Rampias T. HDAC inhibitors: dissecting mechanisms of action to counter tumor heterogeneity. *Cancers* 2021;**13**:3575.
- 167. Marks PA, Xu WS. Histone deacetylase inhibitors: potential in cancer therapy. J Cell Biochem 2009;107:600–8.
- 168. Quintás-Cardama A, Kantarjian HM, Ravandi F, Foudray C, Pemmaraju N, Kadia TM, et al. Very high rates of clinical and cytogenetic response with the combination of the histone deacetylase inhibitor pracinostat (SB939) and 5-azacitidine in high-risk myelodysplastic syndrome. *Blood* 2012;120:3821.
- 169. Garcia-Manero G, Abaza Y, Takahashi K, Medeiros BC, Arellano M, Khaled SK, et al. Pracinostat plus azacitidine in older patients with newly diagnosed acute myeloid leukemia: results of a phase 2 study. *Blood Adv* 2019;3:508–18.
- Terry M. Helsinn and mei discontinue phase III trial of pracinostat in aml. *BioSpace* 2020. Available from: https://www.biospace.com/ article/helsinn-and-mei-abandon-pracinostat-in-aml/.
- 171. Alhazzazi T, Kamarajan P, Verdin E, Kapila Y. SIRT3 and cancer: tumor promoter or suppressor?. *Biochim Biophys Acta* 2011;1816: 80–8.
- 172. Imai S, Armstrong CM, Kaeberlein M, Guarente L. Transcriptional silencing and longevity protein SIR2 is an NAD-dependent histone deacetylase. *Nature* 2000;403:795–800.
- 173. Saunders LR, Verdin E. Sirtuins: critical regulators at the crossroads between cancer and aging. *Oncogene* 2007;26:5489–504.
- 174. Martínez-Redondo P, Vaquero A. The diversity of histone versus nonhistone sirtuin substrates. *Genes Cancer* 2013;4:148-63.
- 175. Carafa V, Altucci L, Nebbioso A. Dual tumor suppressor and tumor promoter action of sirtuins in determining malignant phenotype. *Front Pharmacol* 2019;10:38.
- **176.** Bosch-Presegue L, Vaquero A. Sirtuins in stress response: guardians of the genome. *Oncogene* 2014;**33**:3764.
- 177. Carafa V, Nebbioso A, Altucci L. Sirtuins and disease: the road ahead. *Front Pharmacol* 2012;3:4.
- 178. Bosch-Presegué L, Vaquero A. The dual role of sirtuins in cancer. *Genes Cancer* 2011;2:648–62.
- 179. Yoon Y, Ali M, Wei A, Choon T, Oon C, Shirazi A, et al. Correction: discovery of a potent and highly fluorescent sirtuin inhibitor. *Med-ChemComm* 2015;6:2235.
- 180. Tan YJ, Lee YT, Mancera RL, Oon CE. BZD911 sirtuin inhibitor: identification of key molecular targets and their biological functions in HCT 116 colorectal cancer cells. *Life Sci* 2021;284:119747.
- **181.** Bohnsack JP, Pandey SC. Histone modifications, DNA methylation, and the epigenetic code of alcohol use disorder. In: Pandey SC, editor. *International review of neurobiology*. Elsevier; 2021.
- Kooistra SM, Helin K. Molecular mechanisms and potential functions of histone demethylases. *Nat Rev Mol Cell Biol* 2012;13: 297–311.
- 183. Lee DH, Kim GW, Jeon YH, Yoo J, Lee SW, Kwon SH. Advances in histone demethylase KDM4 as cancer therapeutic targets. *FASEB J* 2020;34:3461-84.
- 184. Kuo KT, Huang WC, Bamodu OA, Lee WH, Wang CH, Hsiao M, et al. Histone demethylase JARID1B/KDM5B promotes aggressiveness of non-small cell lung cancer and serves as a good prognostic predictor. *Clin Epigenet* 2018;10:107.
- 185. Florio R, Carradori S, Veschi S, Brocco D, Di Genni T, Cirilli R, et al. Screening of benzimidazole-based anthelmintics and their enantiomers as repurposed drug candidates in cancer therapy. *Pharmaceuticals* 2021;14:372.

- **186.** Choi HS, Ko YS, Jin H, Kang KM, Ha IB, Jeong H, et al. Anticancer effect of benzimidazole derivatives, especially mebendazole, on triple-negative breast cancer (TNBC) and radiotherapy-resistant TNBC *in vivo* and. *in vitro*. *Molecules* 2021;**26**:5118.
- 187. McLoughlin EC, O'Boyle NM. Colchicine-binding site inhibitors from chemistry to clinic: a review. *Pharmaceuticals* 2020;13:8.
- 188. Khattab M, Al-Karmalawy AA. Revisiting activity of some nocodazole analogues as a potential anticancer drugs using molecular docking and DFT calculations. *Front Chem* 2021;9:628398.
- 189. Florio R, Veschi S, di Giacomo V, Pagotto S, Carradori S, Verginelli F, et al. The benzimidazole-based anthelmintic parbendazole: a repurposed drug candidate that synergizes with gemcitabine in pancreatic cancer. *Cancers* 2019;11:2042.
- 190. Yenjerla M, Cox C, Wilson L, Jordan MA. Carbendazim inhibits cancer cell proliferation by suppressing microtubule dynamics. J Pharmacol Exp Therapeut 2009;328:390.
- 191. Hao D, Rizzo JD, Stringer S, Moore RV, Marty J, Dexter DL, et al. Preclinical antitumor activity and pharmacokinetics of methyl-2benzimidazolecarbamate (FB642). *Invest N Drugs* 2002;20:261–70.
- 192. Hammond LA, Davidson K, Lawrence R, Camden JB, Von Hoff DD, Weitman S, et al. Exploring the mechanisms of action of FB642 at the cellular level. J Cancer Res Clin Oncol 2001;127:301–13.
- 193. Gao P, Dang CV, Watson J. Unexpected antitumorigenic effect of fenbendazole when combined with supplementary vitamins. J Am Assoc Lab Anim Sci 2008;47:37–40.
- **194.** Guerini AE, Triggiani L, Maddalo M, Bonù ML, Frassine F, Baiguini A, et al. Mebendazole as a candidate for drug repurposing in oncology: an extensive review of current literature. *Cancers* 2019;**11**:1284.
- 195. Bertram G. Clinical pharmacology of the anthelmintic drugs. *Basic Clin Pharmacol* 1992:748.
- 196. Gavidia CM, Gonzalez AE, Barron EA, Ninaquispe B, Llamosas M, Verastegui MR, et al. Evaluation of oxfendazole, praziquantel and albendazole against cystic echinococcosis: a randomized clinical trial in naturally infected sheep. *PLoS Neglected Trop Dis* 2010;4:e616.
- 197. Duan Q, Liu Y, Rockwell S. Fenbendazole as a potential anticancer drug. *Anticancer Res* 2013;**33**:355–62.
- 198. Pourgholami MH, Woon L, Almajd R, Akhter J, Bowery P, Morris DL. *In vitro* and *in vivo* suppression of growth of hepatocellular carcinoma cells by albendazole. *Cancer Lett* 2001;165:43–9.
- 199. Khalilzadeh A, Wangoo KT, Morris DL, Pourgholami MH. Epothilone-paclitaxel resistant leukemic cells CEM/dEpoB300 are sensitive to albendazole: involvement of apoptotic pathways. *Biochem Pharmacol* 2007;74:407–14.
- 200. Castro LSEPW, Kviecinski MR, Ourique F, Parisotto EB, Grinevicius VMAS, Correia JFG, et al. Albendazole as a promising molecule for tumor control. *Redox Biol* 2016;**10**:90–9.
- 201. Fridman JS, Lowe SW. Control of apoptosis by p53. Oncogene 2003; 22:9030–40.
- 202. Mrkvová Z, Uldrijan S, Pombinho A, Bartůněk P, Slaninová I. Benzimidazoles downregulate mdm2 and MDMX and activate p53 in MDMX overexpressing tumor cells. *Molecules* 2019;24:2152.
- 203. Wade M, Wang YV, Wahl GM. The p53 orchestra: MDM2 and MDMX set the tone. *Trends Cell Biol* 2010;20:299–309.
- 204. Personalized Medicine Coalition. Personalized medicine at FDA: the scope & significance of progress in 2020. Available from: https://www. personalizedmedicinecoalition.org/Userfiles/PMC-Corporate/file/ PM_at_FDA_The_Scope_Significance_of_Progress_in_2020.pdf.