



Harnessing neo-domestication of wild pigmented rice for enhanced nutrition and sustainable agriculture

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Abstract

Advances in precision gene editing have enabled the rapid domestication of wild crop relatives, a process known as neo-domestication. During domestication, breeding rice for maximum productivity under optimal growth conditions reduced genetic diversity, eliminating variants for stress tolerance and grain nutrients. Wild rice varieties have rich genetic diversity, including variants for disease resistance, stress tolerance, and grain nutritional quality. For example, the grain of pigmented wild rice has abundant antioxidants (anthocyanins, proanthocyanidins, and flavonoids), but low yield, poor plant architecture, and long life cycle limit its cultivation. In this review, we address the neo-domestication of wild pigmented rice, focusing on recent progress, CRISPR-Cas editing toolboxes, selection of key candidate genes for domestication, identifying species with superior potential via generating genomic and multi-omics resources, efficient crop transformation methods and highlight strategies for the promotion and application pigmented rice. We also address critical outstanding questions and potential solutions to enable efficient neo-domestication of wild pigmented rice and thus enhance food security and nutrition.

Introduction

Approximately 10,000–12,000 years ago, our ancestors shifted from hunting and gathering to farming, fundamentally transforming food acquisition. Early farmers selected and altered many wild plant species, resulting in domesticated cultivars tailored to their needs. This process, known as domestication syndrome, led to crops with larger grains, higher yields, more upright growth, non-shattering seeds, increased apical dominance, coordinated flowering, and loss of seed dormancy (Doebley 2006; Gross and Olsen 2010). Comprehensive demographic analysis indicates that domestication has occurred in approximately 2,500 species, with only 250 species undergoing extensive trait domestication (Meyer and Purugganan 2013). Among these, major staple crops like rice (*Oryza sativa*), maize (*Zea mays*), and wheat (*Triticum aestivum*) exhibit the highest productivity and

cultivation areas globally (Jeyasri et al. 2021; Nutan et al. 2020).

Rice is the primary food source for more than half of the global population, sustaining both affluent and impoverished communities (Song et al. 2025). The domestication of rice occurred independently in Asia and Africa (Meyer et al. 2016; Wing et al. 2018). Asian-cultivated rice (*O. sativa*) was domesticated from *O. rufipogon* in Asia around 9,000 years ago, resulting in two subspecies: *O. sativa* ssp. *japonica* and ssp. *indica* (Chen et al. 2019a; Gross and Zhao 2014). African-cultivated rice (*O. glaberrima*) was domesticated from *O. barthii* in West Africa approximately 3,000 years ago (Meyer and Purugganan 2013). Extensive domestication in diverse geographical regions has produced rice varieties with distinct plant characteristics, grain qualities, and cultivation patterns (Meyer and Purugganan 2013). However, rice domestication has primarily focused on productivity and grain quality, often compromising beneficial traits related to nutrition, biotic and abiotic stress tolerance (Fig. 1a). Additionally, domesticated rice requires high inputs of water, fertilizer, and agrochemicals. With the global population projected to reach 10 billion by 2050, developing crops with enhanced nutritional value and increased tolerance to biotic and abiotic stress is crucial for food security (Acosta-Motos et al. 2024). This necessitates a significant boost in crop productivity to meet future

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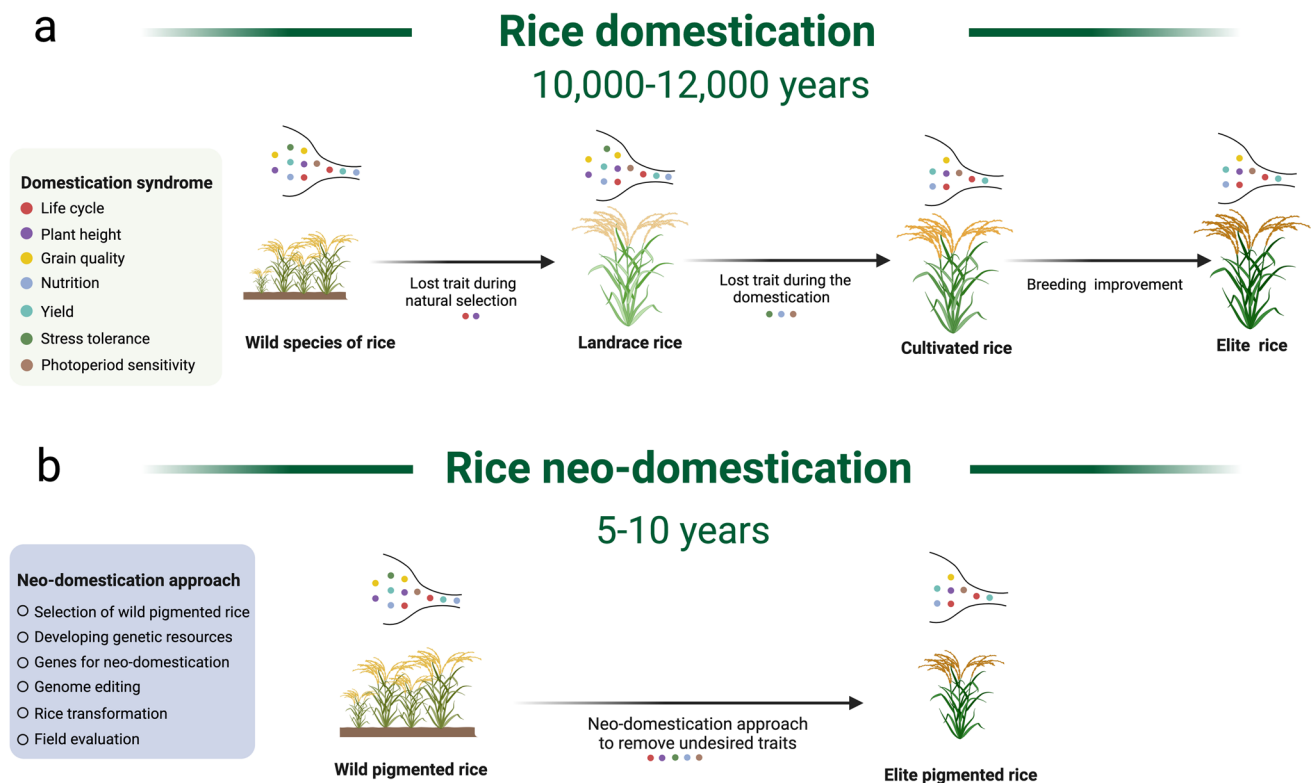


Fig.1 Strategies for traditional and neo-domestication approaches for the domestication of wild vs. pigmented rice. **a** Representation of the domestication process and subsequent loss of genetic variation. This illustration depicts the domestication process, highlighting the gradual loss of useful genetic variation that has resulted from selective breeding and the selection of a limited number of alleles. Over time, as humans have selected specific desirable traits such as increased yield or uniformity, the genetic diversity in cultivated rice populations has decreased, leading to a reduction in the pool of available alleles. **b** Neo-domestication approaches for wild pigmented rice.

This illustration depicts the proposed and currently practiced neo-domestication approaches for wild pigmented rice, aimed at addressing future agricultural challenges. These approaches focus on expediting the selection and domestication of elite pigmented rice varieties, while retaining genetic diversity and the associated elite traits. By leveraging advanced breeding techniques and genomic tools, these neo-domestication strategies offer promising avenues for developing improved pigmented rice varieties capable of both meeting evolving agricultural needs and preserving genetic diversity.

demands. Looking toward wild rice, particularly pigmented varieties, are nutritious and adapted to various climatic conditions, making them a potential solution for nutrition and food security in sustainable agriculture.

Pigmented rice varieties, such as black, red, and brown rice, have a rich history of cultivation and consumption, traditionally black rice is reserved for Chinese royalty and is often referred to as "forbidden rice" or "imperial rice" (Oikawa et al. 2015). These varieties are now gaining attention for their potential to improve human health and enhance food security due to their superior nutritional profiles compared to white rice (Sedeek et al. 2023). Pigmented rice exhibits potent antioxidant properties owing to its high content of anthocyanins and proanthocyanidins that help reduce oxidative stress and inflammation, which are linked to chronic diseases such as cardiovascular diseases, diabetes, and certain cancers (Table 1) (Avinash et al. 2024; Laokuldilok et al. 2011). Anthocyanins, responsible for the red, purple, and blue hues, exhibit significant antioxidant activity,

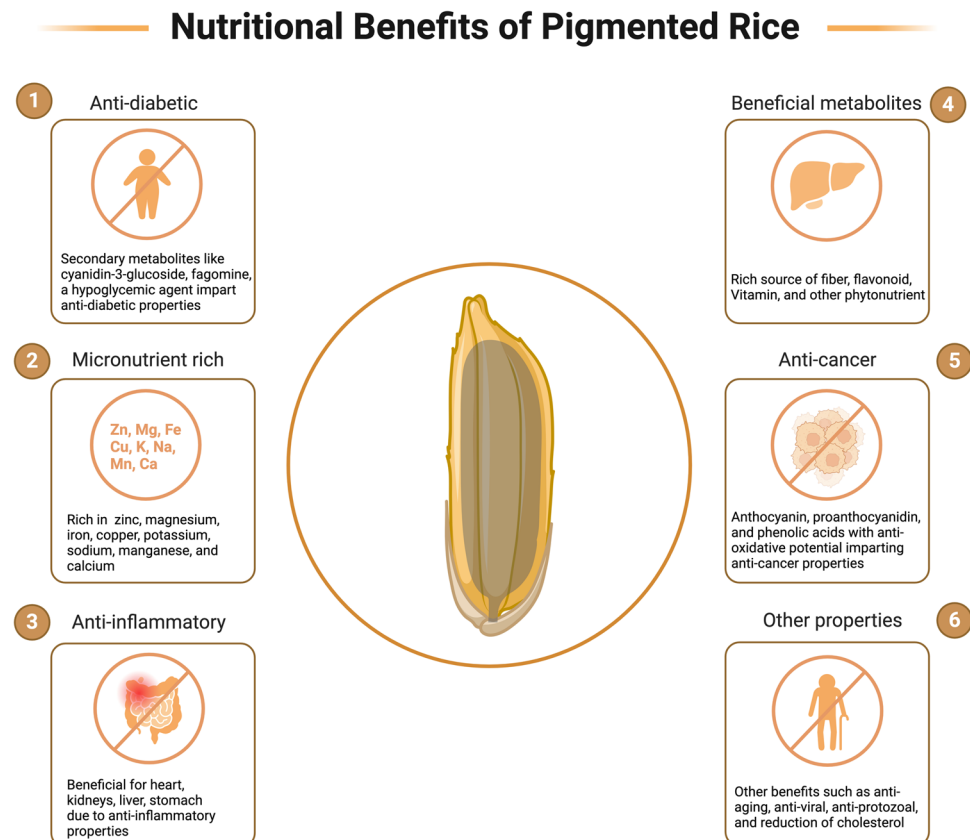
neutralizing free radicals, reducing oxidative stress, and lowering the risk of chronic diseases (Avinash et al. 2024; Laokuldilok et al. 2011). Their anti-inflammatory properties aid in mitigating chronic inflammation, a key factor in conditions like arthritis and cardiovascular diseases (Fig. 2) (Callcott et al. 2019; Pomilio et al. 2024). Additionally, anthocyanins enhance endothelial function, reduce blood pressure, prevent platelet aggregation, and potentially protect against neurodegenerative diseases like Alzheimer's and Parkinson's (Sivasinprasasn et al. 2024; Yadav and Appukuttan 2019). Proanthocyanidins protect cells from oxidative damage, support cardiovascular health by improving blood vessel function, lowering blood pressure, and reducing LDL cholesterol levels (Qi et al. 2023; Sivasinprasasn et al. 2024). Their anti-inflammatory and anticancer activities enhance their health benefits, contributing to the management of conditions like arthritis and the inhibition of cancer cell growth and tumor spread (Fig. 2) (Qi et al. 2023; Sivasinprasasn et al. 2024; Yadav and Appukuttan 2019). Black rice

Table 1 Comparison of the average metabolite abundance among pigmented rice varieties

Nutrient component (Average relative abundance)	Pigmented rice			Human health benefits
	Black (n = 24)	Red (n = 35)	Brown (n = 4)	
Carbohydrates	3.446	1.815	1.276	The main body energy source
Amino acid	1.468	1.158	1.456	Synthesis of body protein and hormones, essential in cell repair and growth
Secondary metabolites	6.642	0.957	0.663	Antioxidant and anti-inflammatory properties, modulate the body health, and maintaining biological processes
Lipids	2.550	0.974	2.206	Energy storage, cell membrane building, and integrity
Cofactors	1.441	1.492	1.206	Crucial for enzyme activities in metabolic pathways
Hormones	1.276	1.256	1.562	Regulate body functions
Flavonoids	12.259	0.904	0.569	Antioxidant properties, and reduces risk of chronic diseases
Peptides	1.906	0.692	0.769	Needed for muscle growth and repair, and slow down the aging process
Anthocyanin	177.998	0.410	0.058	Antioxidant properties, and reduces risk of chronic diseases
Proanthocyanidins	1.527	4.437	0.336	Antioxidant properties that protect the heart and cardiovascular system
Terpenoids	2.501	1.114	0.827	Have anticancer, antimicrobial, anti-inflammatory, antioxidant, and antiallergic properties
Vitamin B1	1.771	0.804	2.876	Needed for skin, hair, muscles, and brain health, essential for nerve function
Vitamin B2	1.911	0.866	0.563	Support various cellular functions, body growth and development
Vitamin B6	6.199	5.169	4.355	Helps make red blood cells, and improve immune function
Vitamin C	1.053	0.963	1.019	Antioxidant properties and lower the risk for some cancers
Vitamin E	5.254	5.098	4.748	Antioxidant properties, and protects vitamin A and certain lipids from damage

Pigmented rice measurements were taken from Sedeek et al. (2023)

Fig. 2 The importance of pigmented rice in providing a variety of nutraceutical benefits for human health. The importance of pigmented rice varieties lies in their diverse range of health benefits, disease resistance, and abiotic stress tolerances. These rice varieties contain natural compounds that have been proven to positively impact cardiovascular health, regulate blood sugar levels, boost the immune system, and provide antioxidant protection. They also possess anti-inflammatory properties and are rich in essential nutrients



contains higher levels of vitamins, fiber, and iron compared to non-pigmented counterparts, supporting immune function and digestive health (Tables 1 and 2) (Sedeek et al. 2023; Tiozon et al. 2023a). Red rice consumption has been associated with reduced cholesterol levels and better cardiovascular health due to monacolin K, a naturally occurring statin. The high fiber content in pigmented rice varieties aids in glycemic control and contains the antidiabetic compound fagomine, making them beneficial for individuals with diabetes (Sedeek et al. 2023; Zhao et al. 2024).

Pigmented rice is also a major source of amino acids, secondary metabolites, lipids, cofactors, hormones, flavonoids, phenylpropanoids, terpenoids, and vitamins like B1, B2, B6, C, and E that play crucial roles in maintaining human health and preventing various diseases (Table 1). Amino acids support muscle growth, immune function, and overall metabolism (Laokuldilok et al. 2011). Secondary metabolites, including flavonoids, phenylpropanoids, and terpenoids, act as antioxidants, anti-inflammatory agents, and antimicrobial compounds, preventing chronic diseases such as cancer, heart disease, and neurodegenerative disorders (Laokuldilok et al. 2011; Qi et al. 2023; Sivasinprasasn et al. 2024). Phytosterols in pigmented rice inhibit cholesterol absorption and regulate undesirable lipoprotein levels. Carotenoids like lutein and zeaxanthin, predominantly found in the bran, contribute to reducing cholesterol levels and protecting eye health (Mbanjo et al. 2020; Melini et al. 2019). Lipids support brain function, reduce inflammation, and improve heart health (Deng et al. 2013). Cofactors, including vitamins and minerals, are necessary for enzyme

function and metabolic processes, enhancing metabolic efficiency, supporting energy production, and promoting overall health (Tiozon et al. 2023a). Vitamins such as B1, B2, B6, C, and E are essential for various bodily functions, including metabolism, skin, eye health, brain function, immune support, and protection against oxidative stress (Mbanjo et al. 2020; Melini et al. 2019). Pigmented rice varieties also contain an abundance of metal ions, such as potassium, sodium, calcium, iron, zinc, manganese, magnesium, copper, cobalt, selenium, chromium, silver, and molybdenum, contributing to their nutritional value and health benefits (Table 2). These elements and secondary metabolites play vital roles in improving human health by acting as antitumor, cholesterol-lowering, immunosuppressant, antiprotozoal, anthelmintic, antiviral, and anti-aging agents (Vaishnav and Demain 2011) (Fig. 2). The rich nutrient profile of pigmented rice contributes to its numerous health benefits.

In terms of food security, pigmented rice is often more resilient to environmental stresses, both biotic and abiotic, reducing the risk of crop failure and ensuring a stable food supply. Moreover, wild pigmented rice, with its distinctive agronomic traits and specific farming practices, offers numerous advantages for farmers compared to cultivated white rice (Table 3). However, it also exhibits negative traits such as a long life cycle, low yield, and higher plant height, which make it more prone to lodging (Sedeek et al. 2024, 2023) (Table 3). Addressing these negative traits is crucial for the successful integration of pigmented rice into mainstream agriculture. The neo-domestication approach, leveraging advancements in biotechnology like precise genome

Table 2 Metal ion profiling of the pigmented rice

Metal ion content	Pigmented rice			Human health significance
	Black (n=24) µg/g	Red (n=35).µg/g	Brown (n=4) µg/g	
Potassium (K)	5573	5438	5296	Maintain fluid and electrolyte balance, and supports cardiovascular health
Sodium (Na)	98.9	107.7	71.4	Maintain fluid and electrolyte balance, supports nerve and cognitive function
Calcium (Ca)	235	237	260	Bone health, muscle contraction, and osteoporosis prevention
Iron (Fe)	47.3	29.2	30.1	Hemoglobin formation and prevents anemia
Zinc (Zn)	46.2	50.8	50.9	Essential for growth and recovery, and immune system function
Manganese (Mn)	67.0	48.2	35.7	Brain function, energy metabolism, protein synthesis, and Improve heart health
Magnesium (Mg)	2203	2365	2266	Muscle activity, protein, fat, and carbohydrate metabolism
Copper (Cu)	8.5	8.0	12.0	Electron carrier, and supports immune and brain function
Cobalt (Co)	0.117	0.140	0.025	Part of vitamin B12 and essential for cell function and nervous system
Selenium (Se)	0.208	0.430	0.000	Antioxidant, reduces inflammation, and enhances immunity
Chromium (Cr)	3.070	1.384	0.764	Carbohydrate metabolism, and controlling glucose level
Silver (Ag)	0.009	0.006	0.003	Antimicrobial properties
Molybdenum (Mo)	2.823	2.176	4.246	a key component of several important enzymes, and support the respiratory system

Pigmented rice measurements were taken from Sedeek et al. (2023)

Table 3 Agronomic trait comparison between cultivated white rice and wild pigmented rice

Agronomic trait	Cultivated white rice	Wild Pigmented Rice
Growth Conditions	Optimal in warm, temperate climates	Adaptable to diverse climates, often found in harsher environments
Soil Requirements	Prefers well-drained, fertile soils	Tolerant of various soil types, often grows in less fertile soils
Water Requirements	Requires consistent, abundant water	Require less or frequent irrigation
Plant Height	Typically 80–120 cm	Variable, shorter or highly tall
Maturity Period	90–150 days	120–180 days, varies with environment
Yield	High yield, often 4–8 tons/ha	Lower yield, generally 1–4 tons/ha
Pest and Disease Resistance	Moderate resistance to common pests	High resistance to pests and diseases
Nutrient Requirements	Standard nutrient requirements	Low nutrient requirements, can grow in poor soils
Harvesting Method	Mechanized and efficient	Often harvested manually due to uneven growth
Post-Harvest Processing	Standard milling processes	Traditional or manual milling, often more labor-intensive
Climate Adaptability	Limited to specific climate ranges	Highly adaptable, can grow in a range of climates
Environmental Impact	Moderate, depends on agricultural practices	Generally lower due to traditional farming methods
Sustainability	Dependent on modern agricultural practices	Often cultivated using traditional, sustainable practices

editing with CRISPR/Cas9, offers solutions by incorporating desirable traits while eliminating or minimizing undesirable characteristics in rice. These advanced tools can rapidly transform wild pigmented rice into commercially viable cultivars with enhanced agronomic and nutritional traits (Fig. 1b) (Anilkumar et al. 2022; Varshney et al. 2021). In addition, genetic diversity within pigmented rice cultivars offers significant potential for breeding programs aimed at developing new varieties with enhanced nutritional qualities and greater adaptability to changing climatic conditions. However, transferring beneficial traits from pigmented rice to modern cultivated rice is challenging due to complex genetic interactions and linkage drag (Meyer and Purugganan 2013). This resilience and adaptability make pigmented rice a vital crop for supporting sustainable agricultural practices and contributing to global food security. Additionally, the cultural significance and traditional value of pigmented rice in many regions can drive consumer acceptance and market demand, further promoting its cultivation and consumption. Therefore, focusing on pigmented rice not only addresses nutritional deficiencies and health concerns but also supports agricultural sustainability and food security. Given its role in sustainable agriculture, pigmented rice presents a compelling case for increased focus and research.

This review outlines current progress and presents a comprehensive plan for the neo-domestication of wild pigmented rice. Key questions addressed include: What we learn from domestication, what are the successful examples of CRISPR neo-domestication in crop species, and how can these be applied to wild pigmented rice? What CRISPR-based molecular suites are available for generating various types of genetic edits? Which domestication genes are amenable to CRISPR engineering in wild pigmented rice, and what potential traits can be improved through gene editing? How should appropriate wild-type varieties of pigmented

rice be selected based on phenotypic characteristics, nutritional values, and resilience to biotic and abiotic factors? What is the amenability of wild pigmented rice varieties to tissue culture, transformation, and regeneration, and what genomic, metabolomic, and sequencing resources are available or needed? How can making pigmented rice available and affordable in low-income areas provide health benefits and solve local nutritional problems for both producers and consumers? How does the neo-domestication of wild pigmented rice impact human health and food security, and why is a multipronged approach necessary for its successful implementation?

CRISPR breeding and editing toolbox for neo-domestication

Lessons from traditional domestication and milestones in neo-domestication

Throughout history, phenotype-targeted selection has served as the fundamental basis of crop domestication and advancement. At the initiation of domestication, various crops underwent convergent alterations in phenotypic traits associated with the domestication syndrome (Doebley 2006; Meyer and Purugganan 2013; Pickersgill 2018). These significant morphological changes seem to have been primarily caused by a restricted number of large-effect QTLs (Doebley 2006; Lenser and Theissen 2013; Meyer and Purugganan 2013). The identification of genes responsible for domestication in current crops has led to intriguing discoveries concerning the nature of the relevant mutations. Meyer and Purugganan (2013) analyzed 60 characterized domestication genes and found that 28 genes had loss-of-function mutations, such as frameshifts and splicing defects resulting in

premature truncations of translated proteins. Additionally, cis-regulatory mutations were found in 15 out of 60 genes, and 10 domestication genes showed either loss-of-function or cis-regulatory mutations. These findings revealed that fundamental aspects of human-oriented crop domestication are focused on loss-of-function mutations or mutations that alter the expression of key genes. Similar studies by Gross and Olsen (2010) and Osterberg et al. (2017) supported this hypothesis. Moreover, a recent study by Chen et al. (2020) revealed that among 17 domestication-related rice QTLs with clear origins, the alleles selected at 11 of the 17 QTLs (65%) are loss-of-function mutations, 4 (23%) are cis-regulatory mutations, and 2 (12%) involve both loss-of-function mutations and cis-regulatory mutations. These reports suggest that loss-of-function mutations have played a more critical role than cis-regulatory mutations in rice domestication.

Building upon these foundational insights, researchers have harnessed domestication knowledge to enhance crop improvement through neo-domestication utilizing CRISPR-based breeding techniques. Two pioneering studies employed CRISPR-Cas tools in wild tomato (*Solanum pimpinellifolium*), to enhance fruit quality and yield while retaining disease resistance and salt tolerance traits (Li et al. 2018; Zsögön et al. 2018). To optimize these plants for high density planting and urban agriculture, key domestication genes associated with compact plant architecture, fruit size, and nutrient content were targeted (Li et al. 2018; Zsögön et al. 2018). These studies involved editing coding sequences, cis-regulatory regions, and upstream open reading frames of genes such as *SELF-PRUNING 5G* (for floral induction), *FASCIATED*, *CLAVATA3*, *WUSCHEL* and *FRUIT WEIGHT 2.2* (for fruit size), *SELF-PRUNING* (for determinate growth habit), *OVATE* (for fruit shape), *MULTIFLORA* (for fruit number), *LYCOPENE BETA CYCLASE* (for lycopene content), and *GDP-L-GALACTOSE PHOSPHORYLASE* (for vitamin C content) (Li et al. 2018; Zsögön et al. 2018). The resulting mutants exhibited improved yield, fruit traits, and plant architecture in a single generation through a single transformation event. Similarly, Rodríguez-Leal et al. (2017) targeted the *CLAVATA-WUSCHEL* signaling pathway in promoter regions and generated lines with varying gene expression levels to enhance fruit size and number. Transformation efficiency varied significantly across genotypes, with success rates ranging from approximately 10–40% in *Agrobacterium*-mediated transformation and often lower in direct protoplast editing approaches due to regeneration challenges. Gene-editing success determined by Sanger sequencing, amplicon deep sequencing, or PCR-RE assays, demonstrated a mutagenesis efficiency of 50–90%, with biallelic or homozygous edits observed at frequencies above 30% in regenerated lines. Notably, off-target effects were minimized through the use of highly specific gRNAs and bioinformatics predictions, but genetic mosaicism in early

regenerants posed limitations, requiring multiple cycles of screening to identify stable homozygous edits. Key challenges encountered included low transformation efficiency in certain accessions, unintended phenotypic trade-offs, and genetic compensation effects that influenced the predictability of phenotypic outcomes. Future improvements necessitate optimizing transformation protocols tailored to wild germplasm, refining multiplex editing strategies to achieve precise allelic variations, and integrating novel base editing or prime editing technologies to enhance efficiency and specificity.

In parallel, CRISPR-Cas has been applied to African-cultivated rice (*Oryza glaberrima*), particularly the landrace Kabre, which is known for its environmental stress tolerance but suffers from lower yield and susceptibility to lodging. Targeting domestication loci such as *HIGH TILLER DWARF 1* (*HTD1*) for plant height and *GRAIN SIZE* (*GS3*), *GRAIN WIDTH* (*GW2*), and *GRAIN NUMBER* (*Gn1a*) for grain quality resulted in considerable improvements in grain yield and reduced lodging (Lacchini et al. 2020). Moreover, targeting *HTD1*, *Gn1a*, *GS3*, and *GW2* via *Agrobacterium*-mediated rice transformation in 13 screened accessions, 61.5% (8/13) regenerated plantlets under optimized hormone conditions with Kabre prioritized for its vigor. Multiplex editing generated 76 T₀ mutants lines with variable editing efficiency, including 3 *gn1a* single, 19 *gs3* single, 38 *gn1a/gs3* double, and 16 *gn1a/gs3/gw2* triple mutants, though *GW2* efficiency was low (6.25% homozygosity) (Lacchini et al. 2020). Expanding this approach to polyploid systems, which offer advantages like genome buffering and environmental resilience, studies in the perennial allotetraploid wild rice *O. alta* (CCDD genome) targeted six domestication-related traits, including seed shattering (*OsqSH1*), awn length (*OsAN-1*), plant height (*OsSD1*), seed size (*OsGS3*) and long heading date (*OsDTH7* and *OsGhd7*). Edited lines exhibited reduced shattering, shorter awns, earlier flowering, reduced plant height, and higher yield (Yu et al. 2021). These studies demonstrate a potential path forward for creating stress-resistant rice by combining genomics knowledge of cultivated crops and rapid genetic change via genome editing (Yu et al. 2021). Despite these advances, both studies highlight persistent bottlenecks such as off-target effects, variable transformation and multiplex efficiency, and genetic heterogeneity emphasizing the need for scalable protocols for non-regenerating landraces and field validation to translate CRISPR gains into sustainable agricultural outcomes.

Similarly, orphan crop groundcherry (*Physalis pruinosa*), CRISPR was used to overcome undesirable traits such as sprawling growth habits and small fruits, leading to enhanced fruit density and improved plant architecture (Lemmon et al. 2018). In potato, knocking out the self-incompatibility gene *S-RNase* allowed the neo-domestication of potato into an

inbred-line-based diploid crop, presenting a promising alternative to traditional clonal propagation of tetraploid potato (Ye et al. 2018). Major technical challenges included tissue culture recalcitrance in *Physalis*, low regeneration rates, unintended pleiotropic effects on plant morphology, and genetic mosaicism in early-generation mutants, necessitating multiple propagation cycles to obtain stable lines. Additionally, in potato, overcoming linkage drag associated with deleterious alleles remained a key limitation, requiring further genomic selection and backcrossing strategies. Future improvements should focus on optimizing regeneration protocols, enhancing multiplex editing for polygenic trait modification, and integrating base or prime editing to minimize off-target effects and maximize precise allele replacement. These studies highlight CRISPR's transformative potential for neo-domestication via precisely targeting for loss of functions or modifying specific gene expression in wild pigmented rice, which enables the rapid development of varieties with enhanced traits, contributing to food security and sustainability.

CRISPR editing toolbox

Genome-editing tools like CRISPR-Cas are invaluable for generating allelic variations in crops mimicking natural genetic diversity and enabling targeted trait improvement (Ahmad et al. 2021; Ali et al. 2020; Butt et al. 2020a; Doudna and Charpentier 2014). These tools use specific nucleases to create double-strand breaks (DSBs) in the DNA at precise genomic locations. These breaks are repaired via homology-directed repair (HDR), which requires a donor template, or through the error-prone non-homologous end-joining (NHEJ) pathway (Sedeek et al. 2019; Zafar et al. 2020). Traditional nucleases like zinc-finger nucleases, transcription activator-like effector nucleases (TALENs), and mega-nucleases have been used in plant gene editing but are limited by their complex protein engineering requirements, reducing their versatility and applicability for gene engineering (Mahfouz and Li 2011). In contrast, CRISPR-Cas has emerged as the leading genome-editing tool due to its simplicity, high efficiency, versatility, and cost-effectiveness. It is capable of targeting a wide range of genomic sites, enabling precise editing with a high degree of accuracy in domesticating wild relatives of crops (Chen et al. 2021a; Fernie and Gutierrez-Marcos 2019; Gasparini et al. 2021; Khan et al. 2019; Zsögön et al. 2018). Furthermore, efficient Cas nucleases like Cas9, Cas12a, Cas12b, and Cas12f, along with advanced CRISPR variants such as base, prime, and twin prime editors, facilitate precise gene insertion, deletion, or base substitution, offering powerful tools for neo-domestication of wild pigmented rice (Fig. 3) (Hua et al. 2019; Veillet et al. 2019).

Utilizing efficient CRISPR-Cas nucleases for trait improvement in wild pigmented rice

CRISPR-Cas9 nuclease is primarily and widely adopted for generating targeted loss-of-function mutations by deleting regulatory regions that suppress desirable traits, thereby improving crop performance (Sedeek et al. 2019). Cas9 mainly generates double-stranded breaks (DSBs) at specific genomic locations having G-rich PAM sequence (NGG) and repaired through non-homologous end joining (NHEJ) pathway (Fig. 3a). Moreover, Cas9 nucleases are a remarkably efficient wide-range of crops traits improvement but having certain limitation such as dependency on G-rich PAM sequences (NGG) and its relatively large size (~4.2 kb) poses challenges for delivery into plant cells (Sedeek et al. 2019). Additionally, while NHEJ efficiently facilitates knockouts, the homologous-directed repair (HDR) pathway, necessary for precise DNA integrations, remains inefficient in plants, restricting its broader applicability in genome editing. The advancement in Class 2 systems Cas12 family of nucleases, such as Cas12a, Cas12b, and Cas12f, address these limitations along with more efficient and flexible genome editing (Fig. 3b). Unlike Cas9 which requires NGG PAM sequences, Cas12a and Cas12b target T-rich PAM sequences, significantly expanding the range of editable genomic sites (Garcia-Doval and Jinek 2017; Tang and Fu 2018) (Fig. 3b). Cas12a generates staggered DSBs, which enhance the efficiency of NHEJ-based knock-in strategies, while its shorter guide RNA (~43 bp) facilitates simpler delivery compared to Cas9's sgRNA (~100 bp) (Moon et al. 2019). Cas12a-mediated genome editing has been successfully utilized in rice and soybean (Xu et al. 2017). Furthermore, Cas12b is smaller in size compared to Cas9 and Cas12a, making it a promising CRISPR system for genome editing in crops. In rice, Cas12b recognizes VTTV PAMs, with a preference for ATTV and GTTG PAMs, additionally, Cas12b has been used in *Arabidopsis thaliana* to induce mutations, including creating large deletions at multiple loci without observed off-target effects (Wu et al. 2020). Unlike Cas12a and b, Cas12f works as an RNA-guided DNA nuclease, specifically cleaving single-stranded DNA (ssDNA) in a target-specific manner (Fig. 3b) (Harrington et al. 2018; Khan et al. 2019). Unlike other Class 2 systems, Cas12f does not require a specific PAM sequence to target and cleave ssDNA. Instead, factors such as seed region, secondary structure, thermodynamic stability, and cofactors help recognize the target sequence (Harrington et al. 2018). Cas12f is the smallest functional CRISPR system known, at only one-third the size of Cas9. The CRISPR-Cas12f system shows promising applications in crop defense against ssDNA viruses and mobile genetic elements. It has been successfully utilized to confer field resistance against ssDNA viruses, such as those from the *Geminiviridae* and *Nanoviridae* families (Khan et al. 2019), significantly expanding the application of the CRISPR system in crop genomes. In summary, the

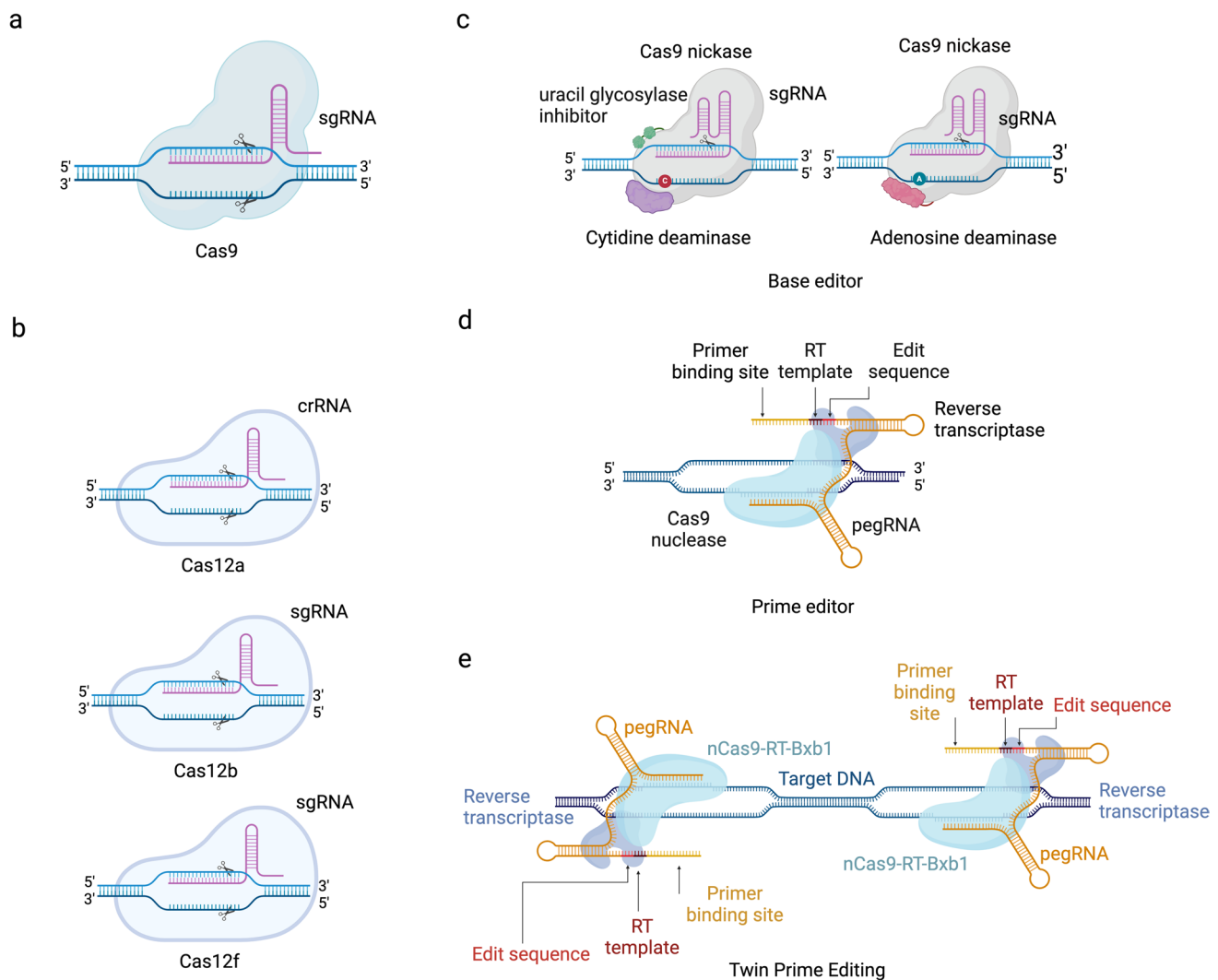


Fig. 3 Different genome editing tools for neo-domestication of pigmented rice. **a** Guided by single-guide RNA (sgRNA), the CRISPR-Cas9 system can generate site-specific double-strand breaks. **b** Other than Cas9, Cas nucleases such as Cas12a, Cas12b, and Cas12f have been harnessed for RNA-guided genome editing. Cas12a: works like Cas9 but allows editing of additional genomic regions having AT-rich PAM sites. Cas12b works like Cas9 but allows editing of additional genomic regions having AT-rich PAM sites and a smaller size than Cas12a, which is a Class 2 Type V-B nuclease. Cas12f is a highly compact Class 2 Type V nuclease that functions as an RNA-guided DNA nuclease and specifically cleaves single-stranded DNA in a tar-

get-specific manner. **c** The base editor method enables precise base conversions without double-strand breaks by fusing Cas9 nickase with cytidine deaminase or adenosine deaminase. Cytosine base editors (CBEs) and adenine base editors (ABEs) can convert C:G to T:A or A:T to G:C, respectively. **d** The prime editor method uses engineered reverse transcriptase with a Cas9 H840 nickase and a prime editing guide RNA (pegRNA). The 5' overhang is cleaved by exonucleases and 3' sequences are ligated to the DNA strand. **e** The prime editor method for useful for larger genomic rearrangements, such as inserting entire genes or removing large genomic segments with high precision

Cas12 family of nucleases provides a robust toolkit for the neo-domestication of pigmented rice, e.g., *OsSPL14* can be targeted to enhance yield and drought tolerance by modulating tillering and grain size (Miura et al. 2010), while *IPA1* can be edited to optimize plant architecture and increase grain production (Liu et al. 2019). Genes involved in anthocyanin biosynthesis, such as *OsMYB2* and *OsDFR*, can be modified to enhance pigmentation and antioxidant properties, improving

nutritional value (Nie et al. 2024; Yang et al. 2021) in the neo-domestication of wild pigmented rice.

Applications of base, prime, and twin prime editing and other emerging technologies for trait improvement in wild pigmented rice

Base editing, prime editing, and twin prime editing represent advanced CRISPR-based techniques that offer precise and

efficient methods for domain knock-in or knockout in genetic research (Fig. 3c–e). These tools are invaluable for the neo-domestication of wild pigmented rice, primarily focusing on crop improvement.

Base editors are ideal for introducing precise point mutations to enhance or knockout gene function. They enable the conversion of one base pair into another without creating double-strand breaks, making them useful for precise point mutations to correct or introduce single nucleotide polymorphisms (Fig. 3c) (Komor et al. 2016). Base editing has been used to introduce point mutations in the ALS gene, resulting in herbicide-resistant plants without off-target effects, ensuring stable trait modifications without double-strand breaks (Shimatani et al. 2017). Prime editing can introduce more complex genetic changes, such as correcting loss-of-function mutations or inserting beneficial alleles. Prime editors offer greater precision by allowing targeted insertions, deletions, and all 12 possible base-to-base conversions without needing double-strand breaks (Fig. 3d). This system uses a reverse transcriptase to copy an RNA template directly into the DNA (Anzalone et al. 2019). Butt et al. (2020b) successfully edited 3 genes using prime editing in rice for herbicide tolerance and increase in yield they targeted *ACETOLACTATE SYNTHASE* (*OsALS*), *IDEAL PLANT ARCHITECTURE 1* (*OsIPA*) and *TEOSINTE BRANCHED 1* (*OsTBI*). Twin prime editing builds on the prime editing principle and is useful for larger genomic rearrangements, such as larger insertions, deletion of genomic region, and introduction of new biosynthetic pathways with high precision (Fig. 3e) (Anzalone et al. 2022). Although its application in plants remains underexplored, twin prime editing could revolutionize the neo-domestication of wild rice by allowing the precise integration of beneficial alleles while preserving genetic diversity. By leveraging the precision and versatility of base editing, prime editing, and twin prime editing, researchers can introduce beneficial traits, improve yield, and enhance stress resistance in pigmented rice varieties.

Harnessing emerging technologies for the neo-domestication of pigmented rice extends beyond advanced CRISPR-Cas systems to include innovative platforms like Retron Library Recombineering (RLR) and the Cas-CLOVER system. This platform allows for the simultaneous introduction of numerous genomic variants, creating pooled and barcoded variant libraries that can be analyzed via targeted deep sequencing, thus surpassing the scale and specificity of traditional CRISPR-Cas methodologies (Orozco-Arias et al. 2022). Successful applications in crops such as *Musa* spp. demonstrate its potential (Thomson et al. 2019; Tripathi et al. 2023). Moreover, Cas-CLOVER can facilitate the development of transgene-free genome-edited crops through the piggyBac transposon system, which introduces targeted double-strand breaks and excises T-DNA without altering the insertion site, thereby maintaining genome integrity

(Madison et al. 2022). Furthermore, recent advancements in epigenome editing and synthetic biology approaches can be harnessed to improve traits in pigmented rice varieties. Epigenome editing, using tools like CRISPR/dCas9 fused with epigenetic modifiers, allows for precise and reversible regulation of gene expression by modifying DNA or histone marks without altering the genetic code which can further harness for upregulate beneficial genes, such as those involved in stress response and nutrient biosynthesis, thereby enhancing crop resilience and nutritional content (Ghoshal et al. 2021; Goell and Hilton 2021). Additionally, synthetic biology techniques like gene drives offer a powerful method for rapidly spreading desirable traits, such as disease resistance, through rice populations, accelerating the neo-domestication process (Berr et al. 2015; Bewick et al. 2017). These innovations represent a significant leap forward in the genetic improvement of rice, particularly in adapting to environmental challenges and meeting nutritional needs.

Balancing coding vs non-coding DNA editing for neo-domestication of wild pigmented rice

When contemplating the neo-domestication of wild-pigmented rice, both coding and non-coding DNA editing offer substantial and unique advantages. Coding DNA editing directly manipulates genes, altering their function or expression to excise negative regulators impacting yield, nutrition, stress tolerance, and other agronomically desirable traits. This method has proven efficacious in engendering significant phenotypic transformations, notwithstanding the occasional pleiotropic effects that may arise (Chen et al. 2019b; Zhang et al. 2020). Coding DNA editing through CRISPR/Cas9-mediated loss of function mutation has shown significant potential in improving key agronomic and quality traits in *Oryza sativa*, providing a foundation for the neo-domestication of wild pigmented rice through targeted gene modifications. For example, knocking out of *GS3*, a positive regulator of grain weight and size, has resulted in elevated grain length, enhancing yield potential (Wang et al. 2018; Yuyu et al. 2020). Modifications in Amino Acid Permease 6 (*AAP6*) and *AAP10*, which regulate grain growth and production, have improved eating and cooking quality, making rice more appealing to consumers (Wang et al. 2020). Additionally, loss-of-function mutations in *BADH2*, which plays a role in synthesizing aromatic compounds like 2-acetyl-1-pyrroline, have produced aromatic rice variants, significantly increasing market value (Ashokkumar et al. 2020). So, this knowledge of gene functions, along with rational multiplexing of 4–6 pivotal genes for various traits, can help in fastening the process of neo-domestication of pigmented wild rice.

In contrast, non-coding DNA editing focuses on natural sequence variations within promoter regions, 3' and 5'

UTRs, enhancers, silencers, and transposable elements, allowing for the fine-tuning of gene expression without altering protein-coding sequences. Various studies show that non-coding DNA editing has remarkable potential for improving rice traits through targeted modifications in diverse genomic regions. For example, editing the promoter of the *xa13* gene enhanced bacterial blight resistance without compromising fertility, as partial promoter deletions suppressed pathogen-induced gene expression while maintaining normal yield (Li et al. 2020). Modifications in the 3' UTR of *TAWAWA1*, particularly within conserved non-coding sequences downstream of the gene, increased panicle branching and improved yield traits, illustrating the role of UTRs in fine-tuning gene expression (Lv et al. 2023; Yoshida et al. 2013). Enhancer editing, such as the insertion of the *nDart1* transposon upstream of *miR156d*, produced the gain-of-function mutant *Bushy dwarf tiller 1*, characterized by increased tillering and dwarfism, highlighting the importance of enhancers in trait development (Hayashi-Tsugane et al. 2015; Nishimura et al. 2019). Similarly, studies on the *Ac* transposon demonstrated its utility in insertional mutagenesis for identifying silencer elements that regulate gene expression in rice (Castanera et al. 2023; Greco et al. 2003). These advancements in non-coding DNA editing provide a powerful framework for fine-tuning gene expression and optimizing agronomic traits in rice. By harnessing these insights, researchers can precisely modify regulatory elements in wild pigmented rice, enabling its neo-domestication while preserving its genetic diversity and enhancing its nutritional, environmental, and economic value. By judiciously choosing coding and non-coding editing approaches, the advantages of genetic editing can be maximized, augmenting desirable traits while maintaining genetic diversity and stability.

Domestication genes amenable to CRISPR breeding in wild pigmented rice

The genus *Oryza sativa* presents a rich genetic pool, and insights gleaned from domestication-associated genes can pave the way for the neo-domestication of wild pigmented rice. During the domestication of today's cultivated rice, a series of changes were marked in their morphological traits, physiological characteristics, and ecological adaptability. These changes mainly occurred through either altered or loss-of-function mutations in specific traits governing genes (Meyer and Purugganan 2013). These changes include genes for seed shattering, seed dormancy, awn length, plant height, tiller and panicle numbers, heading date, grain number, size, and weight (Fig. 4). Here we summarized a plethora of candidate genes that played a major role in rice domestication and some newly identified gene which can further harness

for neo-domestication of pigmented wild rice through CRISPR.

Seed dispersal loss due to shattering has played a crucial role in the domestication process of major cereal crops (Fig. 4). The occurrence of seed shattering in rice is governed by two primary QTLs: *shattering4* (*SH4*)/*Shattering1* (*SHA1*) and *Seed shattering in chromosome 1* (*qSH1*). A SNP positioned 12 kb upstream of *qSH1* significantly impacts its expression at the separation layer, leading to the loss of seed shattering (Konishi et al. 2006). The loss of seed shattering during the domestication of Asian and African rice can be attributed to two independent causative SNPs located in the coding region of *SH4* (Li et al. 2006; Lin et al. 2007). Further research has unveiled complex gene regulatory network beyond *SH4* and *qSH1* involved in seed shattering, such as *SH5*, *SHAT1*, *SSH1*, *SH1*, *OSH15*, *CPL1*, *GRF4*, and *NPC1* (Qin et al. 2010; Subudhi et al. 2013). These loci are linked to regulatory pathways like auxin signaling, cytokinin levels, and certain transcription factors that fine-tune abscission zone (AZ) development and seed dispersal. Through a detailed understanding of the AZ regulatory network with precisely edited genes like *SH4*, *qSH1*, and other related genes using CRISPR it is possible to enhance non-shattering traits in wild pigmented rice while preserving desirable characteristics like nutrition and stress tolerance. Furthermore, these facilitate easier harvesting and increase the commercial viability of wild pigmented rice for supporting sustainable agriculture. Furthermore, the reduction of seed dormancy played a major role in cereal crop domestication. Wild and African-cultivated rice varieties exhibit more dormancy than Asian-cultivated rice (Sarila and Swamy 2005; Vaughan et al. 2008). Rice seed dormancy is primarily regulated by two genes, *SEED DORMANCY 4* (*Sdr4*) and *qSD7-1/Rc* during rice domestication (Sugimoto et al. 2010; Wang et al. 2018). *Sdr4* is regulated by the *OsVPI* transcription factor, and *qSD7-1* encodes basic helix-loop-helix (bHLH) transcription factors, both integrated ABA signaling and biosynthesis pathways; transcriptomic analysis of knockout mutants revealed they regulate multiple metabolic, hormonal pathways and regulatory network (Chen et al. 2023; Sugimoto et al. 2010). By leveraging this knowledge and precisely targeting these genes could fine-tune seed dormancy in pigmented rice, promoting synchronized germination. Similarly, reduction or elimination of awns has been pivotal in rice domestication (Fig. 4). Three genes associated with the transition from long awns to short or no awns have been successfully cloned and characterized: *Awn-1* (*An-1*), *Awn-2* (*An-2*)/*LONG AND BARBED AWN1* (*LABA1*), and *GRAIN NUMBER, GRAIN LENGTH AND AWN DEVELOPMENT* (*GAD1*)/*REGULATOR OF AWN ELONGATION 2* (*RAE2*). *An-1* and *An-2* show additive effects on awn length, with *An-1* promoting awn formation and *An-2* enhancing awn elongation (Bessho-Uehara et al. 2023; Gu et al. 2015;

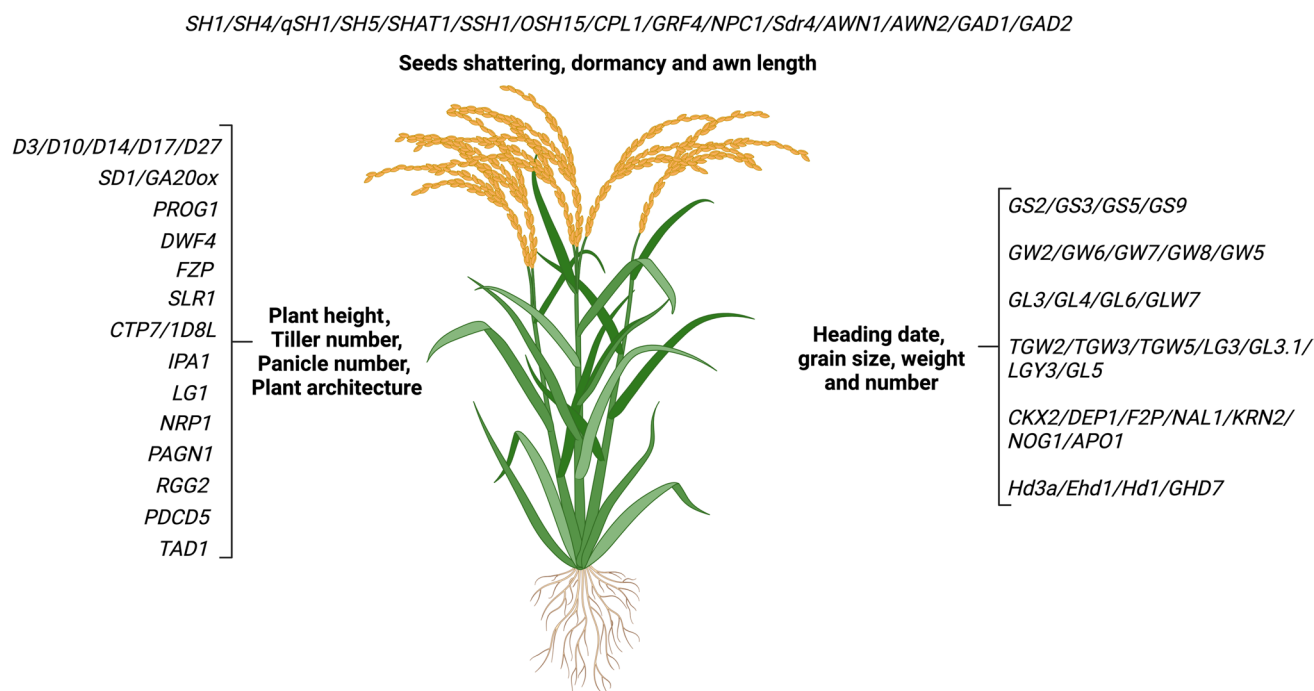


Fig. 4 Knowledge of the genetic pool to determine which domesticated genes and genes are associated with the improvement of traits is of utmost importance. This illustration depicts the important genes involved in domestication and improvement of traits in pigmented rice, including seed shattering, dormancy, awn structure, flowering

time, grain characteristics, plant height and architecture, and tiller number. These genes have great potential both to rapidly facilitate the domestication and further improvement of pigmented rice and to enhance its cultivation

Jin et al. 2016; Luo et al. 2013). Recently, an intricate regulatory network governing awn length revealed that hormonal pathways including auxin and gibberellin signaling are critical for determining awn length and overall spikelet architecture (Yuan et al. 2020). For example, *GAD2/EPFL2* encodes small peptides belonging to the EPIDERMAL PATTERNING FACTOR-LIKE (EPFL) peptide family and modulates awn growth through receptor-like kinases, and this gene undergoes positive selection during rice domestication (Xiong et al. 2022). Understanding these networks provides a foundation for fine-tuning awn traits using CRISPR in wild pigmented rice.

Rice plant architecture has been a prime target for domestication because shorter plants with more tillers produce more grain and resist lodging (Fig. 4). Foremost, a gene is *PROSTRATE GROWTH 1 (PROG1)*, which encodes a Cys2/His2-type zinc-finger transcription factor shown transition from prostrate to erect growth in rice, enhancing lodging resistance (Jin et al. 2008; Tan et al. 2008; Wu et al. 2018). Similarly, *IDEAL PLANT ARCHITECTURE 1 (IPA1)*, which regulates plant height and tiller number, altering IPA1 expression can lead to an ideal plant architecture with fewer tillers and sturdier stems, enhancing yield and resistance to lodging (Jiao et al. 2010). PROG1 and IPA1 encode transcription factors that act as central hubs in regulatory circuits

modulating growth patterns and plant morphology. Recent studies have shown that IPA1 is regulated post-transcriptionally by miR156 and miR529, which modulate its expression to balance tiller number and stem robustness, optimizing yield and lodging resistance. This highlights IPA1 as a master regulator integrating hormonal and environmental signals into developmental programs (Miura et al. 2010). *LIGULELESS1 (LG1)*, encoding a SQUAMOSA promoter-binding transcription factor, regulates panicle architecture, targeting an SNP located 11 kb upstream of *LG1* affects its expression, resulting in compact panicles (Ishii et al. 2013; Zhu et al. 2013). Gene expression network analysis suggests that LG1 modulates auxin signaling pathways by influencing the expression of auxin transporters and biosynthetic genes, further integrating hormonal regulation into panicle morphology. Another gene, *FRIZZY PANICLE (FZP)*, increases the number of secondary branches per panicle and enhances grain yield during rice domestication (Huang et al. 2018). Transcriptional repression of FZP by TAWAWA1 (TAW1) forms a feedback loop limiting excessive branching, ensuring optimal panicle architecture. The semi-dwarfing allele sd-1, found at the SD-1 locus, has a major role in gibberellic acid (GA). This gene became a major contributor to the green revolution (Ashikari et al. 2005), SD-1 loss functions resulting in rice plants with shorter, thicker culms associated with

increased harvest index, lodging resistance, and improved response to nitrogen fertilizer (Ashikari et al. 2005; Hedden 2003; Monna et al. 2002; Spielmeyer et al. 2002). *SD1* also regulates downstream genes in the GA signaling pathway, interacting with DELLA proteins such as *SLENDER RICE 1 (SLR1)*, which mediate growth responses. *SLR1*, regulated by the F-box protein *GIBBERELLIN INSENSITIVE DWARF2 (GID2)*, is essential for GA signaling, and mutations result in a dwarf plant phenotype (Ikeda et al. 2001; Sasaki et al. 2003). *DWARF4 (DWF4)* has a role in the BR homeostasis loss function of these genes, resulting in a more erect leaf phenotype and a higher yield compared to wild-type plants (Sakamoto and Matsuoka 2006). Strigolactones (SLs), a class of phytohormones derived from carotenoids, are also essential for plant architecture. SLs repress branching, and their function is highly conserved in monocots (Gomez-Roldan et al. 2008). In rice, genes like *DWARF3 (D3)*, *D10*, *D14*, *D17 (HTD1)*, and *D27* are involved in SL biosynthesis or signaling. Loss of function of these genes results in a SL-insensitive dwarf phenotype and an increased number of tillers (Arite et al. 2007, 2009; Ishikawa et al. 2005; Lin et al. 2009; Zhou et al. 2013). Additionally, some transcription factor genes like *NEGATIVE REGULATOR OF PHOTOSYNTHESIS 1 (NRP1)* negatively regulate the overall height, yield, and biomass. Knockout of these genes shows increased plant height, panicle number, and biomass compared to wild-type plants (Chen et al. 2021b). CRISPR-Cas9 knockout of *PLANT ARCHITECTURE AND GRAIN NUMBER 1 (PAGN1)* results in plants with increased height, tiller number, and grains per panicle (Yan et al. 2022). *RICE G-PROTEIN Γ SUBUNIT 2 (RGG2)*, encoding a type B heterotrimeric G-protein, negatively impacts plant height (Miao et al. 2019). Genes that influence senescence, such as *PROGRAMMED CELL DEATH 5 (PDCD5)*, also negatively regulate plant height and grain yield (Dong et al. 2021). The tiller number and tillering pattern determine the panicle number. Genes like *TILLERING AND DWARF 1 (TAD1)*, encoding an anaphase-promoting complex (APC/C) subunit of E3 ligase, affect the tiller number, mutant of *tad1* plants exhibit decreased height but increased tillers, resulting in higher yield (Xu et al. 2012). These genes are part of broader transcriptional and hormonal networks integrating environmental and developmental signals, with significant implications for neo-domestication. Furthermore, these analyses facilitate the identification of previously uncharacterized regulatory elements, such as non-coding RNAs and cis-regulatory regions, which can provide additional layers of control for trait refinement through CRISPR technology.

Generally, domesticated rice varieties have larger and heavier grains compared to wild rice species (Fig. 4). Numerous QTLs associated with natural variations in grain characteristics have been functionally characterized. These include *GRAIN SIZE 2 (GS2)*, *GS3*, *GS5*, *GS9*, *GRAIN*

WIDTH 2 (GW2), *GW5*, *GW6*, *GW7/GRAIN LENGTH 7 (GL7)*, *GW8*, *GL4*, *GL6*, *GRAIN LENGTH AND WIDTH 7 (GLW7)*, *THOUSAND-GRAIN WEIGHT 2 (TGW2)*, *TGW3*, *TGW6*, *LARGE GRAIN (LG3)*, *GL3.1*, *GRAIN YIELD QTL (qLGY3)*, and *LARGE GRAIN QTL (qGL5)* (Fan et al. 2006; Hu et al. 2015; Ishimaru et al. 2013; Li et al. 2011; Liu et al. 2018; Qi et al. 2012; Qiao et al. 2021; Ruan et al. 2020; Shi et al. 2020; Song et al. 2007; Wang et al. 2019, 2015a, 2012; Weng et al. 2008; Ying et al. 2018; Yu et al. 2017; Zeng et al. 2013). Many of these genes encode proteins involved in regulatory signaling pathways such as the ubiquitin–proteasome pathway, G-protein signaling, phytohormone signaling, and transcriptional regulation. For instance, *GW2* encodes a predicted RING-type protein with E3 ubiquitin ligase activity, promoting cell proliferation and increasing the size of the spikelet hull through the ubiquitin–proteasome pathway (Song et al. 2007). *GS3* regulates grain length and weight through G-protein signaling (Fan et al. 2006). *TGW6* has indole-3-acetic acid (IAA)-glucose hydrolase activity that positively regulates grain yield by acting on the source organs such as starch (Ishimaru et al. 2013). *TGW3* interacts with *AUXIN RESPONSE FACTOR 4 (ARF4)* to regulate grain length and weight (Hu et al. 2018), and *GL3.1/qGL3* controls grain length and yield through interactions with *GSK3* that modulate BR signaling (Qi et al. 2012). *GW5* is a positive regulator of BR signaling and enhances grain yield in rice (Li et al. 2011). *GS9* interacts with *OFPI4* and *OFPI8* to regulate grain morphology and is a regulatory target of *GSK2* kinase (Liu et al. 2018). *GW6* encodes a GA-regulated GAST family protein that positively influences grain width (Shi et al. 2020), and *GW8* encodes the *SPL16* transcription factor, which controls grain width via transcriptional regulation of *GW7* (Wang et al. 2015b, 2012). Similarly, *GLW7*, *GS2*, and *GL6* are all transcriptional regulators contributing to grain size determination (Hu et al. 2015; Wang et al. 2019). The QTL *qLGY3* affects rice grain yield and encodes the transcription factor *MADS1*, which contains the MADS domain and is a crucial effector downstream of the G-protein $\beta\gamma$ dimer. The variable splicing protein of *MADS1* leads to longer grains and improves the quality and yield of rice grains (Liu et al. 2018). *LG3* is a transcription factor in the ERF family and positively regulates rice grain length without affecting grain quality (Yu et al. 2017). Loss of function of the QTL *qGL5 (AUX3)* results in more significant grain length and weight (Qiao et al. 2021). The transcription factor *ARF6* binds directly to the auxin response elements of the *AUX3* promoter and regulates grain length by altering longitudinal expansion and auxin distribution and content in glume cells (Qiao et al. 2021). These studies suggest that all these qTL are interconnected through regulatory hubs and co-expression modules that integrate hormonal signaling (e.g., auxins, brassinosteroids, and gibberellins) and epigenetic regulation and provide actionable insights for precise

genetic interventions using CRISPR to enhance grain yield and quality in rice.

Several genes regulating grain number in cultivated rice include *Gn1a/CKX2*, *DEP1*, *FZP*, *NAL1*, *KRN2*, *NOG1*, and *APO1* (Fig. 4). Among these, *CKX2*, encoding a cytokinin oxidase/dehydrogenase, has been identified as a major QTL for grain number (Ashikari et al. 2005). Natural variations in *CKX2* result in reduced expression or loss of function, elevating the cytokinin level and increasing grain number and secondary panicle branching (Rashid et al. 2024). Use of *DENSE AND ERECT PANICLE 1 (DEP1)* that encodes G protein γ subunit, which regulates the architecture of dense and erect panicles (Huang et al. 2009). *FRIZZY PANICLE (FZP)*, encoding an AP2/ERF domain transcription factor, and natural variations in the regulatory region of *FZP* lead to decreased expression and consequently elevated grain number (Komatsu et al. 2003; Xu and Sun 2021). Similarly, natural variations in *NARROW LEAF 1 (NAL1)* influence grain number (Ren et al. 2021). Knocking out *KERNEL ROW NUMBER (KRN2)* encodes WD40 proteins and increases grain number and secondary branching, which results in increased grain yield by ~8%, without any observable trade-offs in other agronomic traits (Chen et al. 2022). *NUMBER OF GRAINS 1 (NOG1)* encodes an enoyl-CoA hydratase/isomerase, and the insertion of 12 base pairs into the promoter region of *NOG1* elevates its expression, subsequently increasing grain number (Huo et al. 2017). *ABERRANT PANICLE ORGANIZATION 1 (APO1)*, which plays a role in the transition from rachis branch meristem to spikelet meristem and positively regulates the grain number per panicle (Yin et al. 2021). Gene expression network analysis reveals that these genes are part of interconnected pathways involving cytokinin signaling, transcription factor activity, and metabolic regulation. Integrating such insights with tools like CRISPR allows precise manipulation of key nodes, to optimize grain number without compromising other agronomic traits.

Day length affects flowering time for many plants, and this restricts where plants can be grown to specific latitudes. It has an important role in rice domestication in different geological places around the globe. In-depth investigations of the core genes involved in the regulation of flowering pathways have established a strong correlation between the expression of *Heading Date 3a (Hd3a)* and flowering time in 64 cultivated rice varieties collected from various regions worldwide (Takahashi et al. 2009). The observed variations in *Hd3a* expression can be partly attributed to the allelic diversity in *Heading Date 1 (Hd1)* as well as to the expression levels of *Early Heading Date 1 (Ehd1)*. Additionally, polymorphisms in *GRAIN NUMBER*, *PLANT HEIGHT*, and *HEADING DATE 7 (Ghd7)* have been associated with flowering time and the geographical distribution of rice cultivation in China (Fig. 4) (Xue et al. 2008). These genes form a

complex regulatory network integrating photoperiodic signals with developmental pathways. Gene expression network analyses have revealed that interactions between *Hd1*, *Ehd1*, *Hd3a*, and *Ghd7* are modulated by environmental cues like day length and temperature, as well as epigenetic factors such as histone modifications and DNA methylation. Understanding these networks provides insights into the molecular mechanisms of photoperiodic adaptation and enables the targeted manipulation of flowering time using tools like CRISPR. Such interventions could enhance the adaptability of rice to changing climates and expand its cultivation to new geographic regions.

Understanding traditional knowledge, genetic resources, and biotechnological potential of wild pigmented rice

Identification of pigmented rice varieties exhibiting superior potential

The starting materials are crucial for neo-domestication, and the first step is to select elite materials that exhibit superior potential for further improvement (Cang et al. 2016; Goraloglia et al. 2021). The ideal germplasm for neo-domestication will have: (1) disease resistance, (2) tolerance to abiotic stress, (3) adaptive growth habit, (4) genetic diversity, (5) ease of breeding and genetic transformation, (6) nutritionally rich, (7) cultural acceptance, and (8) easy regulatory compliance.

Furthermore, efficient use of germplasm resources via integrating knowledge from genetics, genomics, genetic engineering, and computational biology, will also facilitate the selection of superior wild pigmented rice varieties for neo-domestication (Huang et al. 2022; Sedeek et al. 2023; Zhang et al. 2023a). Germplasm offers a reservoir of genetic diversity that is essential for selecting elite pigmented rice varieties (Huang et al. 2022). In addition, genomics plays a pivotal role in understanding the genetic basis underlying desirable pigmented rice traits. Whole-genome sequencing enables the identification of key genes associated with ideal traits and the molecular mechanisms governing these traits (Zhang et al. 2023b). Molecular markers such as single nucleotide polymorphisms (SNPs) and simple-sequence repeats (SSRs) allow the precise delineation of genomic regions linked to desirable traits, facilitating marker-assisted selection that can expedite breeding efforts (Kurniasih et al. 2019; Singh et al. 2020). Population genomics offers valuable insights into the evolutionary dynamics that have shaped pigmented rice diversity by elucidating genetic diversity patterns and population structures (Huang et al. 2022). Association mapping studies leverage genomic data to discover correlations between genetic markers and phenotypic

traits, thereby pinpointing candidate genes responsible for yield, and nutritional properties. Genetic studies, including analysis of quantitative trait loci (QTLs) and trait mapping, contribute to understanding the genetic architecture of pigmented rice traits, guiding the selection of parental lines harboring favorable alleles for trait improvement (Mbanjo et al. 2023). Furthermore, phylogeny-driven pharmacological screening holds considerable promise in refining the selection criteria for the creation of elite pigmented rice varieties, while offering deeper insights into their translational applications (Bartlett et al. 2023). Translating foundational knowledge from extensively studied related species can offer valuable insights into developing newly domesticated plant varieties. Additionally, indigenous knowledge systems play a significant role in selecting elite pigmented rice for neo-domestication, as ethnobotanical records offer valuable insights into cultural significance, traditional uses, and ecological significance (Ramirez-Villegas et al. 2022). Hence, indigenous and scientific knowledge must be integrated to select appropriate starting materials for the neo-domestication of pigmented rice.

Multi-omics resources for wild and cultivated pigmented rice to unveil key gene regulatory network and complex biological processes

Genomic resources for wild and cultivated pigmented rice remain limited but are essential for uncovering the genetic basis of pigmentation, yield, and nutritional quality. Although various japonica and indica rice varieties have been sequenced, however, high-quality reference genomes for pigmented varieties remain scarce. Expanding these resources is crucial for identifying key genes and pathways that can enhance precision breeding in pigmented rice (Qin et al. 2021; Shang et al. 2022). Recent efforts have aimed to bridge this gap by constructing comprehensive genomic datasets. For instance, rice pan-genome encompassing 251 accessions of cultivated and wild Asian and African rice species has been aimed to identify novel haplotypes to improve crop potential (Shang et al. 2022). Additionally, Qin et al. (2021) assembled 31 high-quality genomes from a diverse rice collection, providing valuable insights into genetic diversity. More specifically, Sedeek et al. (2023) generated genome assemblies of five pigmented rice varieties, evaluating genetic variation in 51 varieties by resequencing 46 additional ones revealed significant genetic variation. Furthermore, Xie et al. (2023) analyzed approx. 5 k rice accessions globally, including 2,794 with red or black pericarps, shedding light on the evolutionary trajectory of red landraces originating from wild rice. These studies collectively enhance our understanding of the genetic foundation of pigmented rice and provide essential resources for its precision breeding.

Building on these genomic resources, multi-omics approaches offer a deeper understanding of trait development and genetic regulation in pigmented rice, which is crucial for its neo-domestication. Multi-omics data, including genomes, transcriptomes, metabolomes, proteomes, and epigenomes, are crucial for understanding complex traits in crops. Modern crops, show higher gene expression levels and alternative splicing than their wild ancestors, reprogramming the transcriptome during adaptation (Chen et al. 2018; Liu et al. 2015). However, multi-omics also reveals a consistent reduction in allelic diversity across domesticated crops, with major crops containing only a fraction of the genetic diversity found in wild gene pools (Fernie et al. 2006). For example, RNA sequencing (RNA-seq) data from six rice varieties identified genes associated with grain pigmentation, nutrition, and aromatic properties (Zainal-Abidin and Mohamed-Hussein 2020). Additionally, epigenomics studies provides insights into partial heritability in domestication, revealing differential DNA methylation patterns in promoter regions of key transcription factors such as MYB and bHLH shows without selective sweeps, and the causal relationship between genetic variation and resultant phenotypes, offering valuable perspectives on the molecular mechanisms underlying crop domestication (Meyer and Purugganan 2013; Razifard et al. 2020; Schmitz et al. 2013).

To translate these insights into actionable breeding strategies, advanced data integration approaches are essential for accelerating the neo-domestication of wild pigmented rice. Co-expression network analysis methods such as weighted gene co-expression network analysis (WGCNA) facilitate the identification of hub genes that regulate complex traits, including those involved in anthocyanin biosynthesis and stress adaptation (Langfelder and Horvath 2008). Furthermore, combining quantitative trait loci (QTL) mapping with genome-wide association studies (GWAS) has proven effective in identifying genomic regions controlling flavonoid biosynthesis and nutritional properties in pigmented rice. For instance, the QTL GS3.1 has been identified as a regulator of grain size by influencing the metabolic flux between flavonoid and lignin biosynthesis pathways (Zhang et al. 2021). This QTL encodes a MATE transporter that directs p-coumaric acid toward flavonoid biosynthesis, affecting grain size without compromising stress tolerance (Zhang et al. 2021). Furthermore, multi-layered regulatory network analysis incorporating transcription factor binding sites, chromatin accessibility, and metabolomic profiling can delineate key regulatory nodes governing secondary metabolism (Liu et al. 2023; Zhu et al. 2023). Machine learning-driven integration of transcriptomics and metabolomics data has further improved the identification of biomarkers for precision breeding, enabling the selection of pigmented rice with enhanced antioxidant profiles (Zhang et al. 2023a).

Metabolomics, used in QTL studies, investigates genetic determinants of quality traits, although studies explicitly on pigmented rice are limited. Nonetheless, recent efforts have begun addressing this gap. Recently, in pigmented rice, the multi-omic resources disseminated by Sedeek et al. (2023) have furnished an all-encompassing array of genomic, metabolomic, and nutrient data for pigmented rice. A similar metabolomics study was carried out by Zhang et al. (2023b) and Tiozon et al. (2023b) shows how black and red rice have high levels of the flavonoids anthocyanin and proanthocyanidin. Additionally, Chen et al. (2019a) conducted a study to comprehensively characterize the intricate molecular mechanisms operating in the biosynthetic pathway of flavonoids in red, white, and black rice cultivars. Using RNA sequencing of caryopses and isobaric tags for relative and absolute quantitation (iTRAQ) analysis, they performed a detailed analysis of mRNA content and expressed proteins in various colored rice varieties. Specifically, for the red, white, and black rice cultivars, 3417, 329, and 227 distinct genes were discovered, respectively (Chen et al. 2019b). Moreover, proteomic analysis has further strengthened our understanding of these metabolic pathways. Proteomic analysis of these colored rice varieties revealed the presence of 13,996 distinct peptides corresponding to 3,916 proteins. Notably, a total of 32 genes that play a role in the biosynthesis pathway of flavonoids were identified. These findings underscore the importance of integrating high-throughput multi-omics approaches, particularly through gene regulatory network modeling and predictive analytics, to advance precision breeding strategies for pigmented rice.

To enhance the integration of multi-omics datasets, bioinformatics pipelines serve as pivotal tools for constructing gene regulatory networks (GRNs) that elucidate the interactions between genes, metabolites, and regulatory elements. These networks are developed by correlating genomic, transcriptomic, and metabolomic datasets using advanced computational approaches, including weighted gene co-expression network analysis (WGCNA), machine learning algorithms, and Bayesian inference techniques. For instance, metabolite–gene correlation networks have been instrumental in linking flavonoid biosynthesis genes with anthocyanin accumulation, a critical trait in pigmented rice. For example, Zhang and Wong (2022) demonstrated that these networks can identify key enzymes, such as glycosyltransferases, that modulate pigment modification, presenting viable genetic targets for precision breeding strategies. Similarly, integrating transcriptomic and proteomic data derived from RNA-seq and iTRAQ analyses (Chen et al. 2019a) with QTL mapping has unveiled epistatic interactions and regulatory nodes that govern pigmentation mechanisms. These insights are essential for guiding the selection and modification of wild rice germplasm to develop improved pigmented rice varieties with enhanced nutritional and

agronomic traits. Beyond identifying regulatory elements, multi-omics approaches also facilitate the functional validation of key pigmentation-related genes. Transcriptome-wide association studies (TWAS) and chromatin immunoprecipitation sequencing (ChIP-seq) have been pivotal in identifying regulatory elements involved in anthocyanin biosynthesis in pigmented rice. These studies have highlighted the central role of the MYB-bHLH-WD40 (MBW) complex in pigment accumulation. Within this complex WD40 repeat gene *OsTTG1* has been identified as a crucial component directly influencing anthocyanin production in rice (Yang et al. 2021). Experimental validation using CRISPR-Cas9 knockout lines of *OsTTG1* demonstrated that disrupting these genes reduced flavonoid accumulation and simultaneously altered responses to oxidative stress, revealing functional redundancy and compensatory pathways that could be leveraged for neo-domestication (Yang et al. 2021). Beyond pigmentation, research also indicates that transcription factors such as NAC and DREB play significant roles in abiotic stress responses in rice. For instance, overexpression of the NAC transcription factor gene *OsNAC022* has been shown to enhance drought and salt tolerance in rice plants (Hong et al. 2016). Similarly, DREB transcription factors have been associated with improved stress tolerance, with studies demonstrating that overexpression of *DREB1B* in tobacco confers enhanced resistance to various abiotic stresses (Gutha and Reddy 2008). While direct interactions between the MBW complex and stress-responsive transcription factors like NAC and DREB in rice remain to be fully elucidated, the involvement of these factors in both anthocyanin biosynthesis and stress responses suggests potential regulatory crosstalk. To further refine neo-domestications strategies, pathway enrichment analyses, such as Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG), augment multi-omics data integration by identifying metabolic and biosynthetic pathways associated with pigmentation and nutritional enhancement. Additionally, multi-omics-based genome-wide association studies (GWAS) have further revealed co-localization of anthocyanin biosynthesis loci with yield-related traits, such as grain size and starch metabolism, where key regulators like *OsC1* and *OsDFR* influence both flavonoid accumulation and agronomic performance demonstrating the feasibility of breeding strategies that optimize both pigmentation and crop productivity (Zheng et al. 2019). Further aiding in neo-domestication, visualization platforms like Cytoscape enable the exploration of GRNs, facilitating the identification of hub genes responsible for phenotypic variation in pigmented rice. Studies show that linked the upregulation of key enzymes, including phenylalanine ammonia-lyase and chalcone synthase, to increased flavonoid biosynthesis in colored rice cultivars (Zhang et al. 2023b). Integrating these findings into neo-domestication strategies highlights the potential of

multiplex genome editing and precision breeding to optimize pigmentation, stress tolerance, and yield-related traits in wild rice germplasm, ultimately accelerating the development of nutritionally enriched and climate-resilient rice cultivars.

Moreover, along with the high-quality reference genome and multi-omics resources, population genetic analyses should be expanded to include more accessions of pigmented rice to better understand their evolutionary dynamics and identify beneficial alleles. Furthermore, integrating indigenous knowledge systems and scientific research will enrich the selection criteria for elite pigmented rice varieties, considering cultural significance, traditional uses, and ecological importance (Ramirez-Villegas et al. 2022). By leveraging these integrative multi-omics data and generating additional ones, researchers can more precisely identify key regulatory genes and pathways for neo-domestication of wild pigmented rice via keeping its genetic diversity and nutritional values intact.

Amenability of wild pigmented rice varieties to tissue culture, transformation, and regeneration

Tissue culture techniques, including callus induction, somatic embryogenesis, and organogenesis, have been successfully applied to wild rice species, albeit with varying degrees of efficiency. Recent studies showed that a few wild rice and some of cultivated pigmented rice varieties possess the inherent ability to regenerate from explants, a critical trait for genetic transformation and subsequent plant development (Sedeek et al. 2023; Wang et al. 2019).

Transformation efficiency in wild pigmented rice can be enhanced through the optimization of *Agrobacterium*-mediated transformation protocols. Advances in vector design, such as the use of binary vectors with strong promoters and selectable marker genes, have improved the integration and expression of cisgenes in pigmented rice (Molina-Risco et al. 2021; Sedeek et al. 2023; Susanto et al. 2020). Additionally, the application of biolistic methods, although less common, has shown promise in delivering transgenes into recalcitrant genotypes (Gantait et al. 2022; Kausch et al. 2019). Regeneration of transgenic plants from transformed tissues is a critical step in developing genetically modified varieties. Wild pigmented rice varieties, such as Chaling common wild rice (*Oryza rufipogon*), have shown efficient regeneration capabilities, producing fertile plants capable of setting seeds and passing on introduced traits to subsequent generations. Additionally, optimized *Agrobacterium*-mediated transformation protocols, specifically utilizing scutellum tissue of embryos from mature seeds, these wild rice varieties can achieve high transformation and regeneration efficiencies. For example, the regeneration system developed

for Chaling common wild rice resulted in transformation efficiencies between 87–94%, with subsequent regeneration of fertile plants that could reproduce and retain introduced traits across generations (Xiang et al. 2022). The regeneration efficiency of rice varieties, including wild pigmented types, can be significantly enhanced by optimizing the composition of the regeneration medium. Key factors influencing regeneration rates include the concentration and type of growth hormones (such as auxins and cytokinins) and the choice of gelling agents. For example, specific combinations of kinetin, naphthalene acetic acid, and agarose have been shown to improve the regeneration frequency in various rice cultivars (Susanto et al. 2020; Xiang et al. 2022). Overall, the amenability of wild pigmented rice varieties to tissue culture, transformation, and regeneration is promising, making them suitable candidates for genetic improvement through biotechnological approaches. Continued refinement of these techniques and further exploration of genotype-specific responses will enhance the efficiency of producing improved pigmented rice varieties with desirable traits.

Additionally, recent advancements in genetic transformation technologies, including *Agrobacterium*-mediated transformation, have seen significant improvements through the overexpression of morphogenic factors such as *BBM*, *WUS2*, and *GRF4/GIF1*, which enhance somatic embryogenesis and regeneration processes (Debernardi et al. 2020; Lowe et al. 2016). These advancements have been successfully applied in crops like *Zea mays*, *Sorghum bicolor*, and *Oryza sativa* L., showing enhanced transformation efficiency (Debernardi et al. 2020; Lowe et al. 2016). Additionally, novel systems such as the ternary vector system with type III secretion to repress host defenses have further increased transformation success rates (Raman et al. 2022). Direct transformation methods, including biolistic delivery, nanotubes, virus transfection, and PEG- Ca^{2+} mediated methods, bypass the need for DNA insertion by delivering Cas protein and sgRNA as RNP complexes, facilitating precise genetic modifications without chromosomal integration (Chen et al. 2022; Doyle et al. 2023).

Moreover, innovative techniques like the cut-dip-budding delivery method for tuberous and woody species, and RNA virus-mediated delivery into the apical meristem, present efficient alternatives for genetic transformation (Cao et al. 2023; Ellison et al. 2020). The integration of nanotechnology with CRISPR technologies is also being explored, offering transgene-free genome transformation capabilities by leveraging the unique properties of nanoparticles for precise cargo delivery (Demirer et al. 2019). This recent advancement can be further utilized to speed up the process of neo-domestication of pigmented rice.

Promotion and application of neo-domestication of wild pigmented rice

Addressing nutritional needs in low-income areas is essential due to the pressing challenges of food security, hidden hunger, and the growing population. According to the FAO 2022 report on "The State of Food Security and Nutrition in the World 2022," over 820 million people worldwide suffer from hunger, and malnutrition remains a significant issue in many low-income countries ((FAO, 2022). Hidden hunger, which refers to micronutrient deficiencies that occur even when caloric intake is sufficient, affects over 2 billion people globally, particularly in many parts of Africa and Asia (Muthayya et al. 2013). With the global population projected to reach 9.7 billion by 2050, increasing the demand for nutritious and sustainable food sources is imperative (United Nations, 2019).

Recent studies highlight the potential health benefits of pigmented rice (Chen et al. 2017; Sedeek et al. 2023). Increasing the availability and affordability of pigmented rice can thus provide a valuable dietary supplement to populations suffering from hidden hunger. By integrating pigmented rice into the diets of these populations, multiple nutritional deficiencies can be addressed simultaneously. The economic impact of promoting pigmented rice cultivation and consumption extends beyond nutritional benefits. For producers, particularly smallholder farmers, pigmented rice varieties offer an opportunity to diversify crops and increase income. These varieties often fetch higher market prices due to their perceived health benefits and unique culinary qualities (Gunaratne et al. 2013; Sompong et al. 2011). Additionally, it fosters cultural pride, reduces healthcare burdens by alleviating diet-related diseases, and empowers marginalized groups through targeted training and market access programs (Goswami et al. 2023; Horgan et al. 2018). These multifaceted benefits contribute to enhanced livelihoods, environmental sustainability, and inclusive development. Consequently, promoting pigmented rice can enhance the livelihoods of farmers and stimulate local economics. For example, in Thailand, initiatives to cultivate black rice (*hom nil*) have improved farmers' incomes and enhanced local food security by providing a nutritious alternative to conventional rice (Boonjung and Fukai 1996; Suwannaporn and Linne-mann 2008). In China, the long-standing cultivation of black rice has been further boosted by recent government initiatives. Farmers in regions like Guangxi and Yunnan have benefited from higher incomes due to the increased demand for black rice, known for its high antioxidant content and health benefits (Hu et al. 2003; Liu et al. 2022; Wang et al. 2024). In Indonesia, black rice cultivation is gaining popularity, particularly in regions like Bali and

Java. The Indonesian government has implemented programs to support farmers through financial aid and agricultural extension services, helping them improve their cultivation practices, leading to higher yields and better-quality rice (Sitaresmi et al. 2023). In India, integrating red rice into public food distribution systems has helped mitigate iron deficiency anemia among vulnerable populations and shown the dual benefits of health improvement and economic support (Nandeep et al. 2024). In India, a study by Goswami et al. (2023) revealed that the return on investment for black rice cultivation is significantly higher compared to conventional rice, with one rupee invested in black rice yielding a return of 2.15 rupees. This shift not only improves the economic status of the farmers but also contributes to better health outcomes in the region.

Overall, making pigmented rice available and affordable is essential to provide significant health benefits and solve local problems for those in low-income areas. By addressing both nutritional deficiencies and economic challenges, promoting pigmented rice cultivation and consumption stands to benefit producers and consumers alike. This integration contributes to sustainable agricultural practices and enhanced food security, addressing the urgent need to support vulnerable populations.

Conclusions and perspectives

The neo-domestication of wild-pigmented rice offers a transformative opportunity to enhance human health and food security. A multipronged approach is imperative for the successful neo-domestication of wild pigmented rice. This approach requires the integration of insights from previous rice domestication studies, the identification and deployment of efficient CRISPR tools and target genes for precise genome editing, the application of advanced knowledge systems for the selection of elite germplasm, the development and utilization of comprehensive genomic resources, the identification of genotypes amenable to tissue culture and genetic transformation, and the implementation of effective breeding strategies. This comprehensive strategy enhances the agronomic performance and promotes pigmented rice for sustainable agriculture that ultimately results in eradicating hunger and malnutrition.

Despite its immense potential for neo-domestication, pigmented rice still has several crucial questions and challenges that must be addressed. One such question revolves around the efficient introduction of desired genes associated with domestication while preserving the genetic diversity of pigmented rice. Additionally, the identification of optimal combinations of genes and traits to focus on wild pigmented rice is essential for enhancing its nutritional value, yield, and ability to withstand environmental stressors. Furthermore,

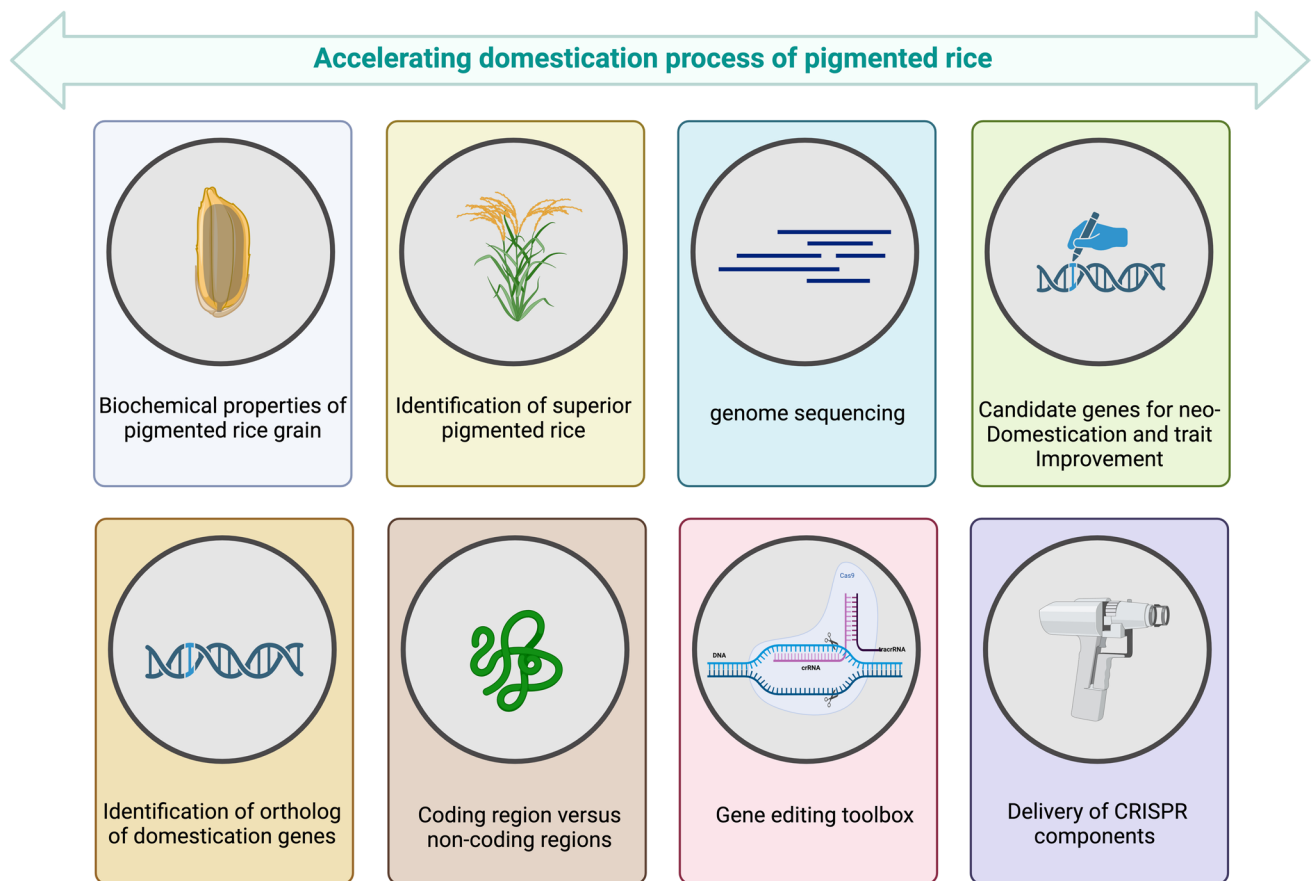


Fig. 5 An overview of the strategic plan outlining the essential measures necessary for the efficient advancement of the entire procedure of neo-domestication pertaining to wild pigmented rice

determining how genome editing tools should be utilized for coding versus non-coding genome sequences in pigmented rice is of utmost importance. Identifying the most efficient genome editing tools and techniques for precise and targeted modifications in pigmented rice and determining the most effective methods for delivering editing components into pigmented rice cells to achieve the desired trait modifications are crucial aspects. Furthermore, extensive educational and communication efforts outside the scientific community are necessary to clarify distinctions between genome editing and traditional genetic modification, address regulatory uncertainties, and promote societal acceptance of neo-domestication approaches in pigmented rice.

Future directions should focus on specific goals for the next decade regarding research, policy, and implementation. Research efforts should aim to expand genomic resources for pigmented rice, facilitating comprehensive trait improvement. Policy initiatives should support the integration of advanced biotechnological tools in breeding programs and ensure regulatory frameworks that enable the rapid adoption of neo-domesticated varieties. Strategies should promote the cultivation and consumption of pigmented rice

in low-income areas, leveraging its nutritional benefits to address hidden hunger and improve public health outcomes. By pursuing these goals, the neo-domestication of wild pigmented rice can make a substantial contribution to sustainable agriculture and global food security. In summary (Fig. 5 roadmap for neo-domestication), genome editing and other emerging technologies provide formidable tools in the endeavor to speed up neo-domestication of pigmented rice and to develop novel crop varieties that can withstand the challenges posed by climate change. By embracing these advancements, we can envision a future in which agriculture assumes a pivotal role in promoting human health, environmental sustainability, and food security for future generations.

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Declarations

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References

- Acosta-Motos JR, Franco-Navarro JD, Gómez-Bellot MJ, Álvarez S (2024) Editorial: crop resistance mechanisms to alleviate climate change-related stress. *Front Plant Sci*. <https://doi.org/10.3389/fpls.2024.1368573>
- Ahmad S, Tang LQ, Shahzad R, Mawia AM, Rao GS, Jamil S, Wei C, Sheng ZH, Shao GN, Wei XJ et al (2021) CRISPR-based crop improvements: a way forward to achieve zero hunger. *J Agric Food Chem* 69:8307–8323. <https://doi.org/10.1021/acs.jafc.1c02653>
- Ali Z, Shami A, Sedeek K, Kamel R, Alhabsi A, Tehseen M, Hassan N, Butt H, Kababji A, Hamdan SM, Mahfouz MM (2020) Fusion of the Cas9 endonuclease and the VirD2 relaxase facilitates homology-directed repair for precise genome engineering in rice. *Commun Biol* 3(1):44. <https://doi.org/10.1038/s42003-020-0768-9>
- Anilkumar C, Sunitha NC, Harikrishna D, Bhat N, Ramesh S (2022) Advances in integrated genomic selection for rapid genetic gain in crop improvement: a review. *Planta* 256:87. <https://doi.org/10.1007/s00425-022-03996-y>
- Anzalone AV, Randolph PB, Davis JR, Sousa AA, Koblan LW, Levy JM, Chen PJ, Wilson C, Newby GA, Raguram A et al (2019) Search-and-replace genome editing without double-strand breaks or donor DNA. *Nature* 576:149–157. <https://doi.org/10.1038/s41586-019-1711-4>
- Anzalone AV, Gao XD, Podracky CJ, Nelson AT, Koblan LW, Raguram A, Levy JM, Mercer JAM, Liu DR (2022) Programmable deletion, replacement, integration and inversion of large DNA sequences with twin prime editing. *Nat Biotechnol* 40:731–740. <https://doi.org/10.1038/s41587-021-01133-w>
- Arite T, Iwata H, Ohshima K, Maekawa M, Nakajima M, Kojima M, Sakakibara H, Kyojuka J (2007) DWARF10, an RMS1/MAX4/DAD1 ortholog, controls lateral bud outgrowth in rice. *Plant J* 51:1019–1029. <https://doi.org/10.1111/j.1365-3113.2007.03210.x>
- Arite T, Umehara M, Ishikawa S, Hanada A, Maekawa M, Yamaguchi S, Kyojuka J (2009) d14, a strigolactone-insensitive mutant of rice, shows an accelerated outgrowth of tillers. *Plant Cell Physiol* 50:1416–1424. <https://doi.org/10.1093/pcp/pcp091>
- Ashikari M, Sakakibara H, Lin SY, Yamamoto T, Takashi T, Nishimura A, Angeles ER, Qian Q, Kitano H, Matsuoka M (2005) Cytokinin oxidase regulates rice grain production. *Science* 309:741–745. <https://doi.org/10.1126/science.1113373>
- Ashokkumar S, Jaganathan D, Ramanathan V, Rahman H, Palaniswamy R, Kambale R, Muthurajan R (2020) Creation of novel alleles of fragrance gene OsBADH2 in rice through CRISPR/Cas9 mediated gene editing. *PLoS ONE* 15:e0237018. <https://doi.org/10.1371/journal.pone.0237018>
- Avinash G, Sharma N, Prasad KR, Kaur R, Singh G, Pagidipala N, Thulasinathan T (2024) Unveiling the distribution of free and bound phenolic acids, flavonoids, anthocyanins, and proanthocyanidins in pigmented and non-pigmented rice genotypes. *Front Plant Sci*. <https://doi.org/10.3389/fpls.2024.1324825>
- Bartlett ME, Moyers BT, Man J, Subramaniam B, Makunga NP (2023) The power and perils of de novo domestication using genome editing. *Annu Rev Plant Biol* 74:727–750. <https://doi.org/10.1146/annurev-arplant-053122-030653>
- Berr A, Shafiq S, Pinon V, Dong A, Shen W-H (2015) The trxG family histone methyltransferase SET DOMAIN GROUP 26 promotes flowering via a distinctive genetic pathway. *Plant J* 81:316–328. <https://doi.org/10.1111/tpj.12729>
- Bessho-Uehara K, Masuda K, Wang DR, Angeles-Shim RB, Obara K, Nagai K, Murase R, Aoki S, Furuta T, Miura K et al (2023) Regulator of Awn Elongation 3, an E3 ubiquitin ligase, is responsible for loss of awns during African rice domestication. *Proc Natl Acad Sci* 120(4):e2207105120. <https://doi.org/10.1073/pnas.2207105120>
- Bewick AJ, Niederhuth CE, Ji L, Rohr NA, Griffin PT, Leebens-Mack J, Schmitz RJ (2017) The evolution of CHROMOMETHYLASES and gene body DNA methylation in plants. *Genome Biol* 18:65. <https://doi.org/10.1186/s13059-017-1195-1>
- Boonjung H, Fukai S (1996) Effects of soil water deficit at different growth stages on rice growth and yield under upland conditions. 2. Phenology, biomass production and yield. *Field Crop Res* 48:47–55. [https://doi.org/10.1016/0378-4290\(96\)00039-1](https://doi.org/10.1016/0378-4290(96)00039-1)
- Butt H, Zaidi SSEA, Hassan N, Mahfouz M (2020a) CRISPR-based directed evolution for crop improvement. *Trends Biotechnol* 38:236–240. <https://doi.org/10.1016/j.tibtech.2019.08.001>
- Butt H, Rao GS, Sedeek K, Aman R, Kamel R, Mahfouz M (2020b) Engineering herbicide resistance via prime editing in rice. *Plant Biotechnol J* 18:2370–2372. <https://doi.org/10.1111/pbi.13399>
- Callcott ET, Blanchard CL, Snell P, Santhakumar AB (2019) The anti-inflammatory and antioxidant effects of acute consumption of pigmented rice in humans. *Food Funct* 10:8230–8239. <https://doi.org/10.1039/C9FO02455G>
- Cang FA, Wilson AA, Wiens JJ (2016) Climate change is projected to outpace rates of niche change in grasses. *Biol Lett* 12(9):20160368. <https://doi.org/10.1098/rsbl.2016.0368>
- Cao X, Xie H, Song M, Lu J, Ma P, Huang B, Wang M, Tian Y, Chen F, Peng J et al (2023) Cut-dip-budding delivery system enables genetic modifications in plants without tissue culture. *Innovation (Camb)* 4:100345. <https://doi.org/10.1016/j.xinn.2022.100345>
- Castanera R, Morales-Díaz N, Gupta S, Purugganan M, Casacuberta JM (2023) Transposons are important contributors to gene expression variability under selection in rice populations. *Elife* 12:RP86324. <https://doi.org/10.7554/eLife.86324>
- Chen L, Huang Y, Ming X, Cheng Z, Zheng J (2017) Proteomic analysis reveals coordinated regulation of anthocyanin biosynthesis through signal transduction and sugar metabolism in black rice leaf. *Int J Mol Sci* 18(12):2722. <https://doi.org/10.3390/ijms18122722>
- Chen Q, Han Y, Liu H, Wang X, Sun J, Zhao B, Li W, Tian J, Liang Y, Yan J et al (2018) Genome-wide association analyses reveal

- the importance of alternative splicing in diversifying gene function and regulating phenotypic variation in maize. *Plant Cell* 30:1404–1423. <https://doi.org/10.1105/tpc.18.00109>
- Chen EW, Huang XH, Tian ZX, Wing RA, Han B (2019a) The genomics of species provides insights into rice domestication and heterosis. *Annu Rev Plant Biol* 70(70):639–665. <https://doi.org/10.1146/annurev-arplant-050718-100320>
- Chen F, Yang Y, Li B, Liu Z, Khan F, Zhang T, Zhou G, Jinxiang T, Shen J, Yi B, Tingdong F, Dai C, Ma C (2019) Functional analysis of M-locus protein kinase revealed a novel regulatory mechanism of self-incompatibility in *Brassica napus* L. *Int J Mol Sci* 20(13):3303. <https://doi.org/10.3390/ijms20133303>
- Chen Z, Li XX, Lu HW, Gao Q, Du HL, Peng H, Qin P, Liang CZ (2020) Genomic atlases of introgression and differentiation reveal breeding footprints in Chinese cultivated rice. *J Genet Genomics* 47:637–649. <https://doi.org/10.1016/j.jgg.2020.10.006>
- Chen FM, Zheng GY, Qu MN, Wang YJ, Lyu MJA, Zhu XG (2021a) Knocking out negative regulator of photosynthesis 1 increases rice leaf photosynthesis and biomass production in the field. *J Exp Bot* 72:1836–1849. <https://doi.org/10.1093/jxb/eraa566>
- Chen QY, Li WY, Tan LB, Tian F (2021b) Harnessing knowledge from maize and rice domestication for new crop breeding. *Mol Plant* 14:9–26. <https://doi.org/10.1016/j.molp.2020.12.006>
- Chen YH, Lu J, Yang X, Huang LC, Zhang CQ, Liu QQ, Li QF (2023) Gene editing of non-coding regulatory DNA and its application in crop improvement. *J Exp Bot* 74:6158–6175. <https://doi.org/10.1093/jxb/erad313>
- Debernardi JM, Tricoli DM, Ercoli MF, Hayta S, Ronald P, Palatnik JF, Dubcovsky J (2020) A GRF–GIF chimeric protein improves the regeneration efficiency of transgenic plants. *Nat Biotechnol* 38(11):1274–1279. <https://doi.org/10.1038/s41587-020-0703-0>
- Demirer GS, Zhang H, Goh NS, González-Grandío E, Landry MP (2019) Carbon nanotube-mediated DNA delivery without transgene integration in intact plants. *Nat Protoc* 14:2954–2971. <https://doi.org/10.1038/s41596-019-0208-9>
- Deng G-F, Xu X-R, Zhang Y, Li D, Gan R-Y, Li H-B (2013) Phenolic compounds and bioactivities of pigmented rice. *Crit Rev Food Sci Nutr* 53:296–306. <https://doi.org/10.1080/10408398.2010.529624>
- Doebley J (2006) Plant science - unfallen grains: how ancient farmers turned weeds into crops. *Science* 312:1318–1319. <https://doi.org/10.1126/science.1128836>
- Dong S, Dong X, Han X, Zhang F, Zhu Y, Xin X, Wang Y, Hu Y, Yuan D, Wang J, Huang Z (2021) OsPDCD5 negatively regulates plant architecture and grain yield in rice. *Proc Natl Acad Sci* 118(29):e2018799118. <https://doi.org/10.1073/pnas.2018799118>
- Doudna JA, Charpentier E (2014) The new frontier of genome engineering with CRISPR–Cas9. *Science* 346(6213):1258096. <https://doi.org/10.1126/science.1258096>
- Doyle B, Cardi T, Bakhsh A, Nicolai A, Bathi K (2023) Novel delivery methods for CRISPR-based plant genome editing. In pp. 41. https://doi.org/10.1007/978-3-031-46150-7_3
- Ellison EE, Nagalakshmi U, Gamo ME, Huang PJ, Dinesh-Kumar S, Voytas DF (2020) Multiplexed heritable gene editing using RNA viruses and mobile single guide RNAs. *Nature plants* 6(6):620–624. <https://doi.org/10.1038/s41477-020-0670-y>
- Fan CH, Xing YZ, Mao HL, Lu TT, Han B, Xu CG, Li XH, Zhang QF (2006) GS3, a major QTL for grain length and weight and minor QTL for grain width and thickness in rice, encodes a putative transmembrane protein. *Theor Appl Genet* 112:1164–1171. <https://doi.org/10.1007/s00122-006-0218-1>
- (FAO), F.a.A.O.o.t.U.N. (2022) The State of Food Security and Nutrition in the World 2022. FAO.
- Fernie AR, Gutierrez-Marcos J (2019) From genome to phenome: genome-wide association studies and other approaches that bridge the genotype to phenotype gap. *Plant J* 97:5–7. <https://doi.org/10.1111/tpj.14219>
- Fernie AR, Tadmor Y, Zamir D (2006) Natural genetic variation for improving crop quality. *Curr Opin Plant Biol* 9:196–202. <https://doi.org/10.1016/j.pbi.2006.01.010>
- Gantait S, Mukherjee E, Jogam P, Babu KH, Jain SM, Suprasanna P (2022) Chapter 13 - Improving crops through transgenic breeding—Technological advances and prospects. In: Chandra Rai A, Kumar A, Modi A, Singh M (eds) *Advances in Plant Tissue Culture*. Academic Press, JHuhuh
- Garcia-Doval C, Jinek M (2017) Molecular architectures and mechanisms of Class 2 CRISPR-associated nucleases. *Curr Opin Struct Biol* 47:157–166. <https://doi.org/10.1016/j.sbi.2017.10.015>
- Gasparini K, dos Reis Moreira J, Peres LEP, Zsögön A (2021) De novo domestication of wild species to create crops with increased resilience and nutritional value. *Curr Opin Plant Biol* 60:102006. <https://doi.org/10.1016/j.pbi.2021.102006>
- Ghoshal B, Picard CL, Vong B, Feng S, Jacobsen SE (2021) CRISPR-based targeting of DNA methylation in *Arabidopsis thaliana* by a bacterial CG-specific DNA methyltransferase. *Proc Natl Acad Sci* 118:e2125016118. <https://doi.org/10.1073/pnas.2125016118>
- Goell JH, Hilton IB (2021) CRISPR/Cas-based epigenome editing: advances, applications, and clinical utility. *Trends Biotechnol* 39:678–691. <https://doi.org/10.1016/j.tibtech.2020.10.012>
- Gomez-Roldan V, Fermas S, Brewer PB, Puech-Pagès V, Dun EA, Pillot JP, Letisse F, Matusova R, Danoun S, Portais JC et al (2008) Strigolactone inhibition of shoot branching. *Nature* 455:189–U122. <https://doi.org/10.1038/nature07271>
- Goraloglia GS, Redick TP, Strauss SH (2021) Gene editing in tree and clonal crops: progress and challenges. *In Vitro Cellular & Dev Biol-Plant* 57:683–699. <https://doi.org/10.1007/s11627-021-10197-x>
- Goswami K, Thapa DB, Sandilya J, Deka N (2023) An assessment of economic profitability of black rice (*Oryza sativa* L. indica) production in Assam, India. *J Appl Res Med Aromat Plants* 34:100488. <https://doi.org/10.1016/j.jarmap.2023.100488>
- Greco R, Ouwerkerk PBF, de Kam RJ, Sallaud C, Favalli C, Colombo L, Guiderdoni E, Meijer AH, Hoge JHC, Pereira A (2003) Transpositional behaviour of an Ac/Ds system for reverse genetics in rice. *Theor Appl Genet* 108(1):10–24. <https://doi.org/10.1007/s00122-003-1416-8>
- Gross BL, Olsen KM (2010) Genetic perspectives on crop domestication. *Trends Plant Sci* 15:529–537. <https://doi.org/10.1016/j.tplants.2010.05.008>
- Gross BL, Zhao ZJ (2014) Archaeological and genetic insights into the origins of domesticated rice. *Proc Natl Acad Sci USA* 111:6190–6197. <https://doi.org/10.1073/pnas.1308942110>
- Gu BG, Zhou TY, Luo JH, Liu H, Wang YC, Shangguan YY, Zhu JJ, Li Y, Sang T, Wang ZX et al (2015) An-2 encodes a cytokinin synthesis enzyme that regulates awn length and grain production in rice. *Mol Plant* 8:1635–1650. <https://doi.org/10.1016/j.molp.2015.08.001>
- Gunaratne A, Wu K, Li D, Bentota A, Corke H, Cai Y-Z (2013) Antioxidant activity and nutritional quality of traditional red-grained rice varieties containing proanthocyanidins. *Food Chem* 138:1153–1161. <https://doi.org/10.1016/j.foodchem.2012.11.129>
- Gutha LR, Reddy AR (2008) Rice DREB1B promoter shows distinct stress-specific responses, and the overexpression of cDNA in tobacco confers improved abiotic and biotic stress tolerance. *Plant Mol Biol* 68:533–555. <https://doi.org/10.1007/s11103-008-9391-8>
- Harrington LB, Burstein D, Chen JS, Paez-Espino D, Ma E, Witte IP, Cofsky JC, Kyrpides NC, Banfield JF, Doudna JA (2018) Programmed DNA destruction by miniature CRISPR–Cas14

- enzymes. *Science* 362(6416):839–842. <https://doi.org/10.1126/science.aav4294>
- Hayashi-Tsugane M, Maekawa M, Tsugane K (2015) A gain-of-function bushy dwarf tiller 1 mutation in rice microRNA gene miR156d caused by insertion of the DNA transposon nDart1. *Sci Rep* 5:14357. <https://doi.org/10.1038/srep14357>
- Hedden P (2003) The genes of the green revolution. *Trends Genet* 19:5–9. [https://doi.org/10.1016/S0168-9525\(02\)00009-4](https://doi.org/10.1016/S0168-9525(02)00009-4)
- Hong Y, Zhang H, Huang L, Li D, Song F (2016) Overexpression of a stress-responsive NAC transcription factor gene ONAC022 improves drought and salt tolerance in rice. *Front Plant Sci* 7:4. <https://doi.org/10.3389/fpls.2016.00004>
- Horgan FG, Kudavidanage EP, Weragodaarachchi A, Ramp D (2018) Traditional ‘maavee’ rice production in Sri Lanka: environmental, economic and social pressures revealed through stakeholder interviews. *Paddy Water Environ* 16:225–241. <https://doi.org/10.1007/s10333-017-0604-0>
- Hu C, Zawistowski J, Ling W, Kitts DD (2003) Black rice (*Oryza sativa* L. indica) pigmented fraction suppresses both reactive oxygen species and nitric oxide in chemical and biological model systems. *J Agric Food Chem* 51:5271–5277. <https://doi.org/10.1021/jf034466n>
- Hu J, Wang YX, Fang YX, Zeng LJ, Xu J, Yu HP, Shi ZY, Pan JJ, Zhang D, Kang SJ et al (2015) A rare allele of GS2 enhances grain size and grain yield in rice. *Mol Plant* 8:1455–1465. <https://doi.org/10.1016/j.molp.2015.07.002>
- Hu Z, Lu SJ, Wang MJ, He H, Sun L, Wang H, Liu XH, Jiang L, Sun JL, Xin X et al (2018) A novel QTL qTGW3 encodes the GSK3/SHAGGY-like kinase OsGSK5/OSSK41 that interacts with OsARF4 to negatively regulate grain size and weight in rice. *Mol Plant* 11:736–749. <https://doi.org/10.1016/j.molp.2018.03.005>
- Hua K, Zhang J, Botella JR, Ma C, Kong F, Liu B, Zhu JK (2019) Perspectives on the application of genome-editing technologies in crop breeding. *Mol Plant* 12:1047–1059. <https://doi.org/10.1016/j.molp.2019.06.009>
- Huang XZ, Qian Q, Liu ZB, Sun HY, He SY, Luo D, Xia GM, Chu CC, Li JY, Fu XD (2009) Natural variation at the DEP1 locus enhances grain yield in rice. *Nat Genet* 41:494–497. <https://doi.org/10.1038/ng.352>
- Huang YY, Zhao SS, Fu YC, Sun HD, Ma X, Tan LB, Liu FX, Sun XY, Sun HY, Gu P et al (2018) Variation in the regulatory region of FZP causes increases in secondary inflorescence branching and grain yield in rice domestication. *Plant J* 96:716–733. <https://doi.org/10.1111/tpj.14062>
- Huang XH, Huang SW, Han B, Li JY (2022) The integrated genomics of crop domestication and breeding. *Cell* 185:2828–2839. <https://doi.org/10.1016/j.cell.2022.04.036>
- Huo, X., Wu, S., Zhu, Z.F., Liu, F.X., Fu, Y.C., Cai, H.W., Sun, X.Y., Gu, P., Xie, D.X., Tan, L.B., et al. (2017). *NOG1* increases grain production in rice. *Nature Communications* 8ARTN 1497. <https://doi.org/10.1038/s41467-017-01501-8>.
- Ikeda A, Ueguchi-Tanaka M, Sonoda Y, Kitano H, Koshioka M, Futsuhara Y, Matsuoka M, Yamaguchi J (2001) Slender rice, a constitutive gibberellin response mutant, is caused by a null mutation of the SLR1 gene, an ortholog of the height-regulating gene GAI/RGA/RHT/D8. *Plant Cell* 13:999–1010. <https://doi.org/10.1105/tpc.13.5.999>
- Ishii T, Numaguchi K, Miura K, Yoshida K, Thanh PT, Htun TM, Yamasaki M, Komeda N, Matsumoto T, Terauchi R et al (2013) Regulates a closed panicle trait in domesticated rice. *Nat Genet* 45:462–465. <https://doi.org/10.1038/ng.2567>
- Ishikawa S, Maekawa M, Arite T, Takamure I, Koyozuka J (2005) Suppression of tiller bud activity in tillering dwarf mutants of rice. *Plant Cell Physiol* 46:S192
- Ishimaru K, Hirotsu N, Madoka Y, Murakami N, Hara N, Onodera H, Kashiwagi T, Ujiiie K, Shimizu B, Onishi A et al (2013) Loss of function of the IAA-glucose hydrolase gene TGW6 enhances rice grain weight and increases yield. *Nat Genet* 45(6):707–711. <https://doi.org/10.1038/ng.2612>
- Jeyasri R, Muthuramalingam P, Satish L, Pandian SK, Chen J-T, Ahmar S, Wang X, Mora-Poblete F, Ramesh M (2021) An overview of abiotic stress in cereal crops: negative impacts, regulation, biotechnology and integrated omics. *Plants* 10(7):1472. <https://doi.org/10.3390/plants10071472>
- Jiao Y, Wang Y, Xue D, Wang J, Yan M, Liu G, Dong G, Zeng D, Lu Z, Zhu X et al (2010) Regulation of OsSPL14 by OsmiR156 defines ideal plant architecture in rice. *Nat Genet* 42:541–544. <https://doi.org/10.1038/ng.591>
- Jin J, Huang W, Gao JP, Yang J, Shi M, Zhu MZ, Luo D, Lin HX (2008) Genetic control of rice plant architecture under domestication. *Nat Genet* 40:1365–1369. <https://doi.org/10.1038/ng.247>
- Jin J, Hua L, Zhu ZF, Tan LB, Zhao XH, Zhang WF, Liu FX, Fu YC, Cai HW, Sun XY et al (2016) Encodes a secreted peptide that regulates grain number, grain length, and awn development in rice domestication. *Plant Cell* 28:2453–2463. <https://doi.org/10.1105/tpc.16.00379>
- Kausch AP, Nelson-Vasilchik K, Hague J, Mookkan M, Quemada H, Dellaporta S, Fragoso C, Zhang ZJ (2019) Edit at will: Genotype independent plant transformation in the era of advanced genomics and genome editing. *Plant Sci* 281:186–205. <https://doi.org/10.1016/j.plantsci.2019.01.006>
- Khan MZ, Zaidi SSEA, Amin I, Mansoor S (2019) A CRISPR way for fast-forward crop domestication. *Trends Plant Sci* 24:293–296. <https://doi.org/10.1016/j.tplants.2019.01.011>
- Komatsu M, Chujo A, Nagato Y, Shimamoto K, Koyozuka J (2003) is required to prevent the formation of axillary meristems and to establish floral meristem identity in rice spikelets. *Development* 130:3841–3850. <https://doi.org/10.1242/dev.00564>
- Komor AC, Kim YB, Packer MS, Zuris JA, Liu DR (2016) Programmable editing of a target base in genomic DNA without double-stranded DNA cleavage. *Nature* 533(7603):420–424. <https://doi.org/10.1038/nature17946>
- Konishi S, Izawa T, Lin SY, Ebana K, Fukuta Y, Sasaki T, Yano M (2006) An SNP caused loss of seed shattering during rice domestication. *Science* 312:1392–1396. <https://doi.org/10.1126/science.1126410>
- Kurniasih NUR, Susandarini R, Susanto F, Nuringtyas T, Jenkins G, Purwestri YA (2019) Characterization of indonesian pigmented rice (*Oryza sativa*) based on morphology and Single nucleotide polymorphisms. *Biodiversitas* 20:1208–1214. <https://doi.org/10.13057/biodiv/d200437>
- Lacchini E, Kiegle E, Castellani M, Adam H, Jouannic S, Gregis V, Kater MM (2020) CRISPR-mediated accelerated domestication of African rice landraces. *Plos One* 15:e0229782. <https://doi.org/10.1371/journal.pone.0229782>
- Langfelder P, Horvath S (2008) WGCNA: an R package for weighted correlation network analysis. *BMC Bioinform* 9:559. <https://doi.org/10.1186/1471-2105-9-559>
- Laokuldilok T, Shoemaker CF, Jongkaewwattana S, Tulyathan V (2011) Antioxidants and antioxidant activity of several pigmented rice brans. *J Agric Food Chem* 59:193–199. <https://doi.org/10.1021/jf103649q>
- Lemmon ZH, Reem NT, Dalrymple J, Soyk S, Swartwood KE, Rodriguez-Leal D, Van Eck J, Lippman ZB (2018) Rapid improvement of domestication traits in an orphan crop by genome editing. *Nature Plants* 4:766–770. <https://doi.org/10.1038/s41477-018-0259-x>
- Lenser T, Theissen G (2013) Molecular mechanisms involved in convergent crop domestication. *Trends Plant Sci* 18:704–714. <https://doi.org/10.1016/j.tplants.2013.08.007>

- Li CB, Zhou AL, Sang T (2006) Rice domestication by reducing shattering. *Science* 311:1936–1939. <https://doi.org/10.1126/science.1123604>
- Li YB, Fan CC, Xing YZ, Jiang YH, Luo LJ, Sun L, Shao D, Xu CJ, Li XH, Xiao JH et al (2011) Natural variation in GS5 plays an important role in regulating grain size and yield in rice. *Nat Genet* 43:1266. <https://doi.org/10.1038/ng.977>
- Li XH, Garvey M, Kaplan I, Li BP, Carrillo J (2018) Domestication of tomato has reduced the attraction of herbivore natural enemies to pest-damaged plants. *Agric for Entomol* 20:390–401. <https://doi.org/10.1111/afe.12271>
- Li C, Li W, Zhou Z, Chen H, Xie C, Lin Y (2020) A new rice breeding method: CRISPR/Cas9 system editing of the Xa13 promoter to cultivate transgene-free bacterial blight-resistant rice. *Plant Biotechnol J* 18:313–315. <https://doi.org/10.1111/pbi.13217>
- Lin ZW, Griffith ME, Li XR, Zhu ZF, Tan LB, Fu YC, Zhang WX, Wang XK, Xie DX, Sun CQ (2007) Origin of seed shattering in rice (*Oryza sativa* L.). *Planta* 226:11–20. <https://doi.org/10.1007/s00425-006-0460-4>
- Lin H, Wang RX, Qian Q, Yan MX, Meng XB, Fu ZM, Yan CY, Jiang B, Su Z, Li JY et al (2009) DWARF27, an iron-containing protein required for the biosynthesis of strigolactones, regulates rice tiller bud outgrowth. *Plant Cell* 21:1512–1525. <https://doi.org/10.1105/tpc.109.065987>
- Liu H, Wang X, Warburton ML, Wen W, Jin M, Deng M, Liu J, Tong H, Pan Q, Yang X et al (2015) Genomic, transcriptomic, and phenomic variation reveals the complex adaptation of modern maize breeding. *Mol Plant* 8:871–884. <https://doi.org/10.1016/j.molp.2015.01.016>
- Liu Q, Han R, Kun W, Zhang J, Ye Y, Wang S, Chen J, Pan Y, Li Q, Xiaopeng X, Zhou J, Tao D, Yuejin W, Xiangdong F (2018) G-protein $\beta\gamma$ subunits determine grain size through interaction with MADS-domain transcription factors in rice. *Nat Commun* 9(1):852. <https://doi.org/10.1038/s41467-018-03047-9>
- Liu M, Shi Z, Zhang X, Wang M, Zhang L, Zheng K, Liu J, Hu X, Di C, Qian Q et al (2019) Inducible overexpression of Ideal Plant Architecture1 improves both yield and disease resistance in rice. *Nature Plants* 5:389–400. <https://doi.org/10.1038/s41477-019-0383-2>
- Liu C, Wang Y, Ma X, Cui D, Han B, Xue D, Han L (2022) Traditional agricultural management of Kam Sweet Rice (*Oryza sativa* L.) in southeast Guizhou Province China. *J Ethnobiol Ethnomed* 18:30. <https://doi.org/10.1186/s13002-022-00528-6>
- Liu Y, Singh SK, Pattanaik S, Wang H, Yuan L (2023) Light regulation of the biosynthesis of phenolics, terpenoids, and alkaloids in plants. *Commun Biol* 6:1055. <https://doi.org/10.1038/s42003-023-05435-4>
- Lowe K, Wu E, Wang N, Hoerster G, Hastings C, Cho MJ, Seelange C, Lenderts B, Chamberlin M, Cushatt J et al (2016) Morphogenic regulators baby boom and Wuschel improve monocot transformation. *Plant Cell* 28:1998–2015. <https://doi.org/10.1105/tpc.16.00124>
- Luo JH, Liu H, Zhou TY, Gu BG, Huang XH, Shangguan YY, Zhu JJ, Li Y, Zhao Y, Wang YC et al (2013) An-1 encodes a basic helix-loop-helix protein that regulates awn development, grain size, and grain number in rice. *Plant Cell* 25:3360–3376. <https://doi.org/10.1105/tpc.113.113589>
- Lv Y, Zhang X, Yanjuan H, Liu S, Yin Y, Wang X (2023) BOS1 is a basic helix–loop–helix transcription factor involved in regulating panicle development in rice. *Front Plant Sci*. <https://doi.org/10.3389/fpls.2023.1162828>
- Madison BB, Patil D, Richter M, Li X, Tong M, Cranert S, Wang X, Martin R, Xi H, Tan Y et al (2022) Cas-CLOVER is a novel high-fidelity nuclease for safe and robust generation of T(SCM)-enriched allogeneic CAR-T cells. *Mol Ther Nucleic Acids* 29:979–995. <https://doi.org/10.1016/j.omtn.2022.06.003>
- Mahfouz MM, Li L (2011) TALE nucleases and next generation GM crops. *GM Crops* 2:99–103. <https://doi.org/10.4161/gmcr.2.2.17254>
- Mbanjo EGN, Kretschmar T, Jones H, Ereful N, Blanchard C, Boyd LA, Sreenivasulu N (2020) The genetic basis and nutritional benefits of pigmented rice grain. *Front Genet* 11:229. <https://doi.org/10.3389/fgene.2020.00229>
- Mbanjo EGN, Pasion EA, Jones H, Carandang S, Misra G, Ignacio JC, Kretschmar T, Sreenivasulu N, Boyd LA (2023) Unravelling marker trait associations linking nutritional value with pigmentation in rice seed. *Plant Genome*. <https://doi.org/10.1002/tpg2.20360>
- Melini V, Panfili G, Fratianni A, Acquistucci R (2019) Bioactive compounds in rice on Italian market: pigmented varieties as a source of carotenoids, total phenolic compounds and anthocyanins, before and after cooking. *Food Chem* 277:119–127. <https://doi.org/10.1016/j.foodchem.2018.10.053>
- Meyer RS, Purugganan MD (2013) Evolution of crop species: genetics of domestication and diversification. *Nat Rev Genet* 14:840–852. <https://doi.org/10.1038/nrg3605>
- Meyer RS, Choi JY, Sanches M, Plessis A, Flowers JM, Amas J, Dorph K, Barretto A, Gross B, Fuller DQ et al (2016) Domestication history and geographical adaptation inferred from a SNP map of African rice. *Nat Genet* 48:1083. <https://doi.org/10.1038/ng.3633>
- Miao J, Yang ZF, Zhang DP, Wang YZ, Xu MB, Zhou LH, Wang J, Wu SJ, Yao YL, Du X et al (2019) Mutation of RGG2, which encodes a type B heterotrimeric G protein γ subunit, increases grain size and yield production in rice. *Plant Biotechnol J* 17:650–664. <https://doi.org/10.1111/pbi.13005>
- Miura K, Ikeda M, Matsubara A, Song X-J, Ito M, Asano K, Matsuoka M, Kitano H, Ashikari M (2010) OsSPL14 promotes panicle branching and higher grain productivity in rice. *Nat Genet* 42:545–549. <https://doi.org/10.1038/ng.592>
- Molina-Risco M, Ibarra O, Faion-Molina M, Kim B, Septiningsih EM, Thomson MJ (2021) Optimizing Agrobacterium-Mediated Transformation and CRISPR-Cas9 Gene Editing in the tropical japonica Rice Variety Presidio. *Int J Mol Sci* 22:10909
- Monna L, Kitazawa N, Yoshino R, Suzuki J, Masuda H, Maehara Y, Tanji M, Sato M, Nasu S, Minobe Y (2002) Positional cloning of rice semidwarfing gene, sd-1: rice “green revolution gene” encodes a mutant enzyme involved in gibberellin synthesis. *DNA Res* 9:11–17. <https://doi.org/10.1093/dnares/9.1.11>
- Moon SB, Kim DY, Ko JH, Kim JS, Kim YS (2019) Improving CRISPR genome editing by engineering guide RNAs. *Trends Biotechnol* 37:870–881. <https://doi.org/10.1016/j.tibtech.2019.01.009>
- Muthayya S, Rah JH, Sugimoto JD, Roos FF, Kraemer K, Black RE (2013) The global hidden hunger indices and maps: an advocacy tool for action. *PLoS ONE* 8:e67860. <https://doi.org/10.1371/journal.pone.0067860>
- Nandee ER, Mahajan H, Mummadi MK, Sairam C, K, V., Kadiyam, J., Meshram, I., Pagidoju, S., Reddy, V.R., Panda, H., et al (2024) Implementation, delivery, and utilization of iron fortified rice supplied through public distribution system across different states in India: an exploratory mixed-method study. *PLOS Global Public Health* 4:e0003533. <https://doi.org/10.1371/journal.pgph.0003533>
- Nie S, Huang W, He C, Wu B, Duan H, Ruan J, Zhao Q, Fang Z (2024) Transcription factor OsMYB2 triggers amino acid transporter OsANT1 expression to regulate rice growth and salt tolerance. *Plant Physiol*. <https://doi.org/10.1093/plphys/kiae559>
- Nishimura H, Himi E, Eun C-H, Takahashi H, Qian Q, Tsugane K, Maekawa M (2019) Transgenerational activation of an autonomous DNA transposon, Dart1-24, by 5-azaC treatment in rice. *Theor Appl Genet* 132:3347–3355. <https://doi.org/10.1007/s00122-019-03429-7>

- Nutan KK, Rathore RS, Tripathi AK, Mishra M, Pareek A, Singla-Pareek SL (2020) Integrating the dynamics of yield traits in rice in response to environmental changes. *J Exp Bot* 71:490–506. <https://doi.org/10.1093/jxb/erz364>
- Oikawa T, Maeda H, Oguchi T, Yamaguchi T, Tanabe N, Ebana K, Yano M, Ebitani T, Izawa T (2015) The birth of a black rice gene and its local spread by introgression. *Plant Cell* 27:2401–2414. <https://doi.org/10.1105/tpc.15.00310>
- Orozco-Arias, S., Candamil-Cortes, M.S., Jaimes, P.A., Valencia-Castrillon, E., Tabares-Soto, R., Isaza, G., and Guyot, R. (2022). Automatic curation of LTR retrotransposon libraries from plant genomes through machine learning. *J Integr Bioinform* 1910.1515/jib-2021–0036.
- Osterberg JT, Xiang W, Olsen LI, Edenbrandt AK, Vedel SE, Christiansen A, Landes X, Andersen MM, Pagh P, Sandoe P et al (2017) Accelerating the domestication of new crops: feasibility and approaches. *Trends Plant Sci* 22:373–384. <https://doi.org/10.1016/j.tplants.2017.01.004>
- Pickersgill, B. (2018). Parallel vs. Convergent Evolution in Domestication and Diversification of Crops in the Americas. *Frontiers in Ecology and Evolution* 6 56. <https://doi.org/10.3389/fevo.2018.00056>.
- Pomilio AB, Szewczuk NA, Duchowicz PR (2024) Dietary anthocyanins balance immune signs in osteoarthritis and obesity – update of human in vitro studies and clinical trials. *Crit Rev Food Sci Nutr* 64:2634–2672. <https://doi.org/10.1080/10408398.2022.2124948>
- Qi P, Lin YS, Song XJ, Shen JB, Huang W, Shan JX, Zhu MZ, Jiang LW, Gao JP, Lin HX (2012) The novel quantitative trait locus GL3.1 controls rice grain size and yield by regulating Cyclin-T1;3. *Cell Res* 22:1666–1680. <https://doi.org/10.1038/cr.2012.151>
- Qi Q, Chu M, Yu X, Xie Y, Li Y, Du Y, Liu X, Zhang Z, Shi J, Yan N (2023) Anthocyanins and Proanthocyanidins: Chemical Structures, Food Sources, Bioactivities, and Product Development. *Food Rev Intl* 39:4581–4609. <https://doi.org/10.1080/87559129.2022.2029479>
- Qiao JY, Jiang HZ, Lin YQ, Shang LG, Wang M, Li DM, Fu XD, Geisler M, Qi YH, Gao ZY et al (2021) A novel miR167a-OsARF6-OsAUX3 module regulates grain length and weight in rice. *Mol Plant* 14:1683–1698. <https://doi.org/10.1016/j.molp.2021.06.023>
- Qin Y, Kim S-M, Zhao X, Jia B, Lee H-S, Kim K-M, Eun M-Y, Jin I-D, Sohn J-K (2010) Identification for quantitative trait loci controlling grain shattering in rice. *Genes & Genomics* 32:173–180. <https://doi.org/10.1007/s13258-009-0879-7>
- Qin, P., Lu, H.W., Du, H.L., Wang, H., Chen, W.L., Chen, Z., He, Q., Ou, S.J., Zhang, H.Y., Li, X.Z., et al. (2021). Pan-genome analysis of 33 genetically diverse rice accessions reveals hidden genomic variations. *Cell* 184:3542–+. <https://doi.org/10.1016/j.cell.2021.04.046>.
- Raman V, Rojas CM, Vasudevan B, Dunning K, Kolape J, Oh S, Yun J, Yang L, Li G, Pant BD et al (2022) Agrobacterium expressing a type III secretion system delivers *Pseudomonas* effectors into plant cells to enhance transformation. *Nat Commun* 13:2581. <https://doi.org/10.1038/s41467-022-30180-3>
- Ramirez-Villegas J, Khoury CK, Achicanoy HA, Diaz MV, Mendez AC, Sosa CC, Kehel Z, Guarino L, Abberton M, Aunario J et al (2022) State of ex situ conservation of landrace groups of 25 major crops. *Nature Plants* 8:491. <https://doi.org/10.1038/s41477-022-01144-8>
- Rashid A, Achary VMM, Abdin MZ, Karippadakkam S, Parmar H, Panditi V, Prakash G, Bhatnagar-Mathur P, Reddy MK (2024) Cytokinin oxidase2-deficient mutants improve panicle and grain architecture through cytokinin accumulation and enhance drought tolerance in indica rice. *Plant Cell Rep* 43:207. <https://doi.org/10.1007/s00299-024-03289-6>
- Razifard H, Ramos A, Della Valle AL, Bodary C, Goetz E, Manser EJ, Li X, Zhang L, Visa S, Tieman D et al (2020) Genomic evidence for complex domestication history of the cultivated tomato in Latin America. *Mol Biol Evol* 37:1118–1132. <https://doi.org/10.1093/molbev/msz297>
- Ren MM, Huang MH, Qiu HY, Chun Y, Li L, Kumar A, Fang JJ, Zhao JF, He H, Li XY (2021) Genome-wide association study of the genetic basis of effective tiller number in rice. *Rice* 14:56. <https://doi.org/10.1186/s12284-021-00495-8>
- Rodríguez-Leal D, Lemmon ZH, Man J, Bartlett ME, Lippman ZB (2017) Engineering quantitative trait variation for crop improvement by Genome Editing. *Cell* 171:470. <https://doi.org/10.1016/j.cell.2017.08.030>
- Ruan BP, Shang LG, Zhang B, Hu J, Wang YX, Lin H, Zhang AP, Liu CL, Peng YL, Zhu L et al (2020) Natural variation in the promoter of TGW2 determines grain width and weight in rice. *New Phytol* 227:629–640. <https://doi.org/10.1111/nph.16540>
- Sakamoto T, Matsuoka M (2006) Characterization of constitutive photomorphogenesis and dwarfism homologs in rice (*Oryza sativa* L.). *J Plant Growth Regul* 25:245–251. <https://doi.org/10.1007/s00344-006-0041-6>
- Sarla N, Swamy BPM (2005) *Oryza glaberrima*: a source for the improvement of *Oryza sativa*. *Curr Sci* 89:955–963
- Sasaki A, Itoh H, Gomi K, Ueguchi-Tanaka M, Ishiyama K, Kobayashi M, Jeong DH, An G, Kitano H, Ashikari M et al (2003) Accumulation of phosphorylated repressor for gibberellin signaling in an F-box mutant. *Science* 299:1896–1898. <https://doi.org/10.1126/science.1081077>
- Schmitz RJ, Schultz MD, Urich MA, Nery JR, Pelizzola M, Libiger O, Alix A, McCosh RB, Chen HM, Schork NJ et al (2013) Patterns of population epigenomic diversity. *Nature* 495:193–198. <https://doi.org/10.1038/nature11968>
- Sedek KEM, Mahas A, Mahfouz M (2019) Plant genome engineering for targeted improvement of crop traits. *Front Plant Sci*. <https://doi.org/10.3389/fpls.2019.00114>
- Sedek K, Zuccolo A, Fornasiero A, Weber AM, Sanikommu K, Sampathkumar S, Rivera LF, Butt H, Mussurova S, Alhabsi A et al (2023) Multi-omics resources for targeted agronomic improvement of pigmented rice. *Nature Food* 4:366–371. <https://doi.org/10.1038/s43016-023-00742-9>
- Sedek K, Mohammed N, Zhou Y, Zuccolo A, Sanikommu K, Kantharajappa S, Al-Bader N, Tashkandi M, Wing RA, Mahfouz MM (2024) Multitrait engineering of Hassawi red rice for sustainable cultivation. *Plant Sci* 341:112018. <https://doi.org/10.1016/j.plantsci.2024.112018>
- Shang LG, Li XX, He HY, Yuan QL, Song YN, Wei ZR, Lin H, Hu M, Zhao FL, Zhang C et al (2022) A super pan-genomic landscape of rice. *Cell Res* 32:878–896. <https://doi.org/10.1038/s41422-022-00685-z>
- Shi CL, Dong NQ, Guo T, Ye WW, Shan JX, Lin HX (2020) A quantitative trait locus GW6 controls rice grain size and yield through the gibberellin pathway. *Plant J* 103:1174–1188. <https://doi.org/10.1111/tpj.14793>
- Shimatani Z, Kashojiya S, Takayama M, Terada R, Arazoe T, Ishii H, Teramura H, Yamamoto T, Komatsu H, Miura K, Ezura H, Nishida K, Ariizumi T, Kondo A (2017) Targeted base editing in rice and tomato using a CRISPR-Cas9 cytidine deaminase fusion. *Nat Biotechnol* 35(5):441–443. <https://doi.org/10.1038/nbt.3833>
- Singh RK, Prasad A, Muthamilarasan M, Parida S K, Prasad M (2020) Breeding and biotechnological interventions for trait improvement: status and prospects. *Planta*. <https://doi.org/10.1007/s00425-020-03465-4>

- Sitaesmi T, Hairmansis A, Widyastuti Y, Susanto U, Wibowo BP, Widiastuti ML, Rumanti IA, Suwarno WB, Nugraha Y (2023) Advances in the development of rice varieties with better nutritional quality in Indonesia. *J Agri Food Res* 12:100602
- Sivasinprasasn S, Tocharus J, Mahatheeranont S, Nakrat S, Tocharus C (2024) Anthocyanin-rich fraction of black rice bran extract protects against amyloid β -Induced oxidative stress, endoplasmic reticulum stress, and neuronal apoptosis in SK-N-SH Cells. *Pharmaceuticals* 17:1039
- Sompong R, Siebenhandl-Ehn S, Linsberger-Martin G, Berghofer E (2011) Physicochemical and antioxidative properties of red and black rice varieties from Thailand, China and Sri Lanka. *Food Chem* 124:132–140. <https://doi.org/10.1016/j.foodchem.2010.05.115>
- Song XJ, Huang W, Shi M, Zhu MZ, Lin HX (2007) A QTL for rice grain width and weight encodes a previously unknown RING-type E3 ubiquitin ligase. *Nat Genet* 39:623–630. <https://doi.org/10.1038/ng2014>
- Song M, Xu L, Ge J, Zhang H, Zuo L, Jiang J, Ding Y, Xie Y, Wu F (2025) EARice10: a 10 m resolution annual rice distribution map of East Asia for 2023. *Earth Syst Sci Data* 17:661–683. <https://doi.org/10.5194/essd-17-661-2025>
- Spielmeier W, Ellis MH, Chandler PM (2002) Semidwarf (sd-1), “green revolution” rice, contains a defective gibberellin 20-oxidase gene. *Proc Natl Acad Sci U S A* 99:9043–9048. <https://doi.org/10.1073/pnas.132266399>
- Subudhi PK, Singh PK, DeLeon T, Parco A, Karan R, Biradar H, Cohn MA, Sasaki T (2013) Mapping of seed shattering loci provides insights into origin of weedy rice and rice domestication. *J Hered* 105:276–287. <https://doi.org/10.1093/jhered/est089>
- Sugimoto K, Takeuchi Y, Ebana K, Miyao A, Hirochika H, Hara N, Ishiyama K, Kobayashi M, Ban Y, Hattori T et al (2010) Molecular cloning of Sdr4, a regulator involved in seed dormancy and domestication of rice. *Proc Natl Acad Sci USA* 107:5792–5797. <https://doi.org/10.1073/pnas.0911965107>
- Susanto FA, Wijayanti P, Fauzia AN, Komalasari RD, Nuringtyas TR, Purwestri YA (2020) Establishment of a plant tissue culture system and genetic transformation for agronomic improvement of Indonesian black rice (*Oryza sativa* L.). *plant cell. Tissue Organ Culture (PCTOC)* 141:605–617. <https://doi.org/10.1007/s11240-020-01819-0>
- Suwannaporn P, Linnemann A (2008) Consumer Preferences and buying criteria in rice: a study to identify market strategy for Thailand jasmine rice export. *J Food Prod Market* 14:33–53. <https://doi.org/10.1080/10454440801986348>
- Takahashi Y, Teshima KM, Yokoi S, Innan H, Shimamoto K (2009) Variations in Hd1 proteins, Hd3a promoters, and Ehd1 expression levels contribute to diversity of flowering time in cultivated rice. *Proc Natl Acad Sci USA* 106:4555–4560. <https://doi.org/10.1073/pnas.0812092106>
- Tan L, Li X, Liu F, Sun X, Li C, Zhu Z, Fu Y, Cai H, Wang X, Xie D et al (2008) Control of a key transition from prostrate to erect growth in rice domestication. *Nat Genet* 40:1360–1364. <https://doi.org/10.1038/ng.197>
- Tang YY, Fu Y (2018) Class 2 CRISPR/Cas: an expanding biotechnology toolbox for and beyond genome editing. *Cell Biosci* 8:59. <https://doi.org/10.1186/s13578-018-0255-x>
- Thomson J, Oikeh S, Sithole-Niang I, Tripathi L (2019) Advanced genetic technologies for improving plant production. In pp. 161–169. <https://doi.org/10.4324/9780429401701-21>
- Tiozon RJN, Sartagoda KJD, Fernie AR, Sreenivasulu N (2023a) The nutritional profile and human health benefit of pigmented rice and the impact of post-harvest processes and product development on the nutritional components: a review. *Crit Rev Food Sci Nutr* 63:3867–3894. <https://doi.org/10.1080/10408398.2021.1995697>
- Tiozon RN, Sreenivasulu N, Alseekh S, Sartagoda KJD, Usadel B, Fernie AR (2023) Metabolomics and machine learning technique revealed that germination enhances the multi-nutritional properties of pigmented rice. *Commun Biol* 6:1000. <https://doi.org/10.1038/s42003-023-05379-9>
- Tripathi L, Ntui VO, Tripathi JN, Norman D, Crawford J (2023) A new and novel high-fidelity genome editing tool for banana using Cas-CLOVER. *Plant Biotechnol J* 21:1731–1733. <https://doi.org/10.1111/pbi.14100>
- United Nations, D.o.E.a.S.A., Population Division (2019). *World Population Prospects 2019*.
- Vaishnav P, Demain AL (2011) Unexpected applications of secondary metabolites. *Biotechnol Adv* 29:223–229. <https://doi.org/10.1016/j.biotechadv.2010.11.006>
- Varshney RK, Barmukh R, Roorkiwal M, Qi Y, Kholova J, Tuberosa R, Reynolds MP, Tardieu F, Siddique KHM (2021) Breeding custom-designed crops for improved drought adaptation. *Adv Genet* 2:e202100017. <https://doi.org/10.1002/ggn2.202100017>
- Vaughan DA, Lu BR, Tomooka N (2008) The evolving story of rice evolution. *Plant Sci* 174:394–408. <https://doi.org/10.1016/j.plantsci.2008.01.016>
- Veillet F, Perrot L, Chauvin L, Kermarrec MP, Guyon-Debast A, Chauvin JE, Nogu   F, Mazier M (2019) Transgene-free genome editing in tomato and potato plants using agrobacterium-mediated delivery of a CRISPR/Cas9 cytidine base editor. *Int J Mol Sci* 20:402. <https://doi.org/10.3390/ijms20020402>
- Wang SK, Wu K, Yuan QB, Liu XY, Liu ZB, Lin XY, Zeng RZ, Zhu HT, Dong GJ, Qian Q et al (2012) Control of grain size, shape and quality by *OsSPL16* in rice. *Nat Genet* 44:950. <https://doi.org/10.1038/ng.2327>
- Wang H, Studer AJ, Zhao Q, Meeley R, Doebley JF (2015) Evidence that the origin of naked kernels during maize domestication was caused by a single amino acid substitution in. *Genetics* 200:965. <https://doi.org/10.1534/genetics.115.175752>
- Wang SK, Li S, Liu Q, Wu K, Zhang JQ, Wang SS, Wang Y, Chen XB, Zhang Y, Gao CX et al (2015) The regulatory module determines grain shape and simultaneously improves rice yield and grain quality. *Nat Genet* 47:949. <https://doi.org/10.1038/ng.3352>
- Wang M, Li WZ, Fang C, Xu F, Liu YC, Wang Z, Yang R, Zhang M, Liu SL, Lu SJ et al (2018) Parallel selection on a dormancy gene during domestication of crops from multiple families. *Nat Genet* 50:1435. <https://doi.org/10.1038/s41588-018-0229-2>
- Wang AH, Hou QQ, Si LZ, Huang XH, Luo JH, Lu DF, Zhu JJ, Shang-guan YY, Miao JS, Xie YF et al (2019) The PLATZ transcription factor GL6 affects grain length and number in rice. *Plant Physiol* 180:2077–2090. <https://doi.org/10.1104/pp.18.01574>
- Wang S, Yang Y, Guo M, Zhong C, Yan C, Sun S (2020) Targeted mutagenesis of amino acid transporter genes for rice quality improvement using the CRISPR/Cas9 system. *The Crop Journal* 8:457–464. <https://doi.org/10.1016/j.cj.2020.02.005>
- Wang F, Sun T, Yu S, Liu C, Cheng Z, Xia J, Han L (2024) Ethnobotanical studies on rice landraces under on-farm conservation in Xishuangbanna of Yunnan Province. *Chin J Ethnobiol Ethnomed* 20:45. <https://doi.org/10.1186/s13002-024-00683-y>
- Weng JF, Gu SH, Wan XY, Gao H, Guo T, Su N, Lei CL, Zhang X, Cheng ZJ, Guo XP et al (2008) Isolation and initial characterization of GW5, a major QTL associated with rice grain width and weight. *Cell Res* 18:1199–1209. <https://doi.org/10.1038/cr.2008.307>
- Wenkang Chen L, Chen XZ, Yang N, Guo J, Wang M, Ji S, Zhao X, Yin P, Cai L et al (2022) Convergent selection of a WD40 protein that enhances grain yield in maize and rice. *Science* 375(6587):eabg7985. <https://doi.org/10.1126/science.abg7985>

- Wing RA, Purugganan MD, Zhang QF (2018) The rice genome revolution: from an ancient grain to Green Super Rice. *Nat Rev Genet* 19:505–517. <https://doi.org/10.1038/s41576-018-0024-z>
- Wu YZ, Zhao SS, Li XR, Zhang BS, Jiang LY, Tang YY, Zhao J, Ma X, Cai HW, Sun CQ et al (2018) Deletions linked to PROG1 gene participate in plant architecture domestication in Asian and African rice. *Nat Commun* 9:4157. <https://doi.org/10.1038/s41467-018-06509-2>
- Wu F, Qiao X, Zhao Y, Zhang Z, Gao Y, Shi L, Du H, Wang L, Zhang YJ, Zhang Y et al (2020) Targeted mutagenesis in *Arabidopsis thaliana* using CRISPR-Cas12b/C2c1. *J Integr Plant Biol* 62:1653–1658. <https://doi.org/10.1111/jipb.12944>
- Xiang Z, Chen Y, Chen Y, Zhang L, Liu M, Mao D, Chen L (2022) Agrobacterium-mediated high-efficiency genetic transformation and genome editing of chaling common wild rice (*Oryza rufipogon* Griff.) using scutellum tissue of embryos in mature seeds. *Front Plant Sci*. <https://doi.org/10.3389/fpls.2022.849666>
- Xie L, Wu D, Fang Y, Ye C, Zhu QH, Wei X, Fan L (2023) Population genomic analysis unravels the evolutionary roadmap of pericarp color in rice. *Plant Commun* 5:100778. <https://doi.org/10.1016/j.xplc.2023.100778>
- Xiong LL, Huang YY, Liu ZP, Li C, Yu H, Shahid MQ, Lin YH, Qiao XY, Xiao JY, Gray JE et al (2022) Small epidermal patterning FACTOR-like2 peptides regulate awn development in rice. *Plant Physiol* 190:516–531. <https://doi.org/10.1093/plphys/kiac278>
- Xu R, Sun CQ (2021) What happened during domestication of wild to cultivated rice. *Crop Journal* 9:564–576. <https://doi.org/10.1016/j.cj.2021.02.005>
- Xu C, Wang YH, Yu YC, Duan JB, Liao ZG, Xiong GS, Meng XB, Liu GF, Qian Q, Li JY (2012) Degradation of MONOCULM 1 by APC/CTAD1 regulates rice tillering. *Nat Commun* 3:750. <https://doi.org/10.1038/ncomms1743>
- Xu R, Qin R, Li H, Li D, Li L, Wei P, Yang J (2017) Generation of targeted mutant rice using a CRISPR-Cpf1 system. *Plant Biotechnol J* 15:713–717. <https://doi.org/10.1111/pbi.12669>
- Xue WY, Xing YZ, Weng XY, Zhao Y, Tang WJ, Wang L, Zhou HJ, Yu SB, Xu CG, Li XH et al (2008) Natural variation in Ghd7 is an important regulator of heading date and yield potential in rice. *Nat Genet* 40:761–767. <https://doi.org/10.1038/ng.143>
- Yadav S, Appukuttan JP (2019) Inhibition of LPS induced neurochemical imbalance and oxidative stress by pigmented and non-pigmented rice bran extracts. *J Food Biochem* 43:e12735. <https://doi.org/10.1111/jfbc.12735>
- Yan PW, Zhu Y, Wang Y, Ma FY, Lan DY, Niu F, Dong SQ, Zhang XW, Hu J, Liu SW et al (2022) A new RING finger protein, plant architecture and grain number 1, affects plant architecture and grain yield in rice. *Int J Mol Sci* 23:824. <https://doi.org/10.3390/ijms23020824>
- Yang X, Wang J, Xia X, Zhang Z, He J, Nong B, Luo T, Feng R, Wu Y, Pan Y et al (2021) OsTTG1, a WD40 repeat gene, regulates anthocyanin biosynthesis in rice. *Plant J* 107:198–214. <https://doi.org/10.1111/tpj.15285>
- Ye M, Peng Z, Tang D, Yang Z, Li D, Xu Y, Zhang C, Huang S (2018) Generation of self-compatible diploid potato by knockout of S-RNase. *Nat Plants* 4:651–654. <https://doi.org/10.1038/s41477-018-0218-6>
- Yin C, Zhu Y, Li X, Lin Y (2021) Molecular and genetic aspects of grain number determination in rice (*Oryza sativa* L.). *Int J Mol Sci* 22(2):728. <https://doi.org/10.3390/ijms22020728>
- Ying JZ, Ma M, Bai C, Huang XH, Liu JL, Fan YY, Song XJ (2018) qTGW12a, a major QTL that negatively modulates grain length and weight in rice. *Mol Plant* 11:750–753. <https://doi.org/10.1016/j.molp.2018.03.007>
- Yoshida A, Sasao M, Yasuno N, Takagi K, Daimon Y, Chen R, Yamazaki R, Tokunaga H, Kitaguchi Y, Sato Y et al (2013) TAWAWA1, a regulator of rice inflorescence architecture, functions through the suppression of meristem phase transition. *Proc Natl Acad Sci* 110:767–772. <https://doi.org/10.1073/pnas.1216151110>
- Yu JP, Xiong HY, Zhu XY, Zhang HL, Li HH, Miao JL, Wang WS, Tang ZS, Zhang ZY, Yao GX et al (2017) Contributing to rice grain length and yield was mined by Ho-LAMap. *Bmc Biology*. <https://doi.org/10.1186/s12915-017-0365-7>
- Yu H, Lin T, Meng XB, Du HL, Zhang JK, Liu GF, Chen MJ, Jing YH, Kou LQ, Li XX et al (2021) A route to de novo domestication of wild allotetraploid rice. *Cell* 184:1156. <https://doi.org/10.1016/j.cell.2021.01.013>
- Yuan Z, Persson S, Zhang D (2020) Molecular and genetic pathways for optimizing spikelet development and grain yield. *abioTECH* 1:276–292. <https://doi.org/10.1007/s42994-020-00026-x>
- Yuyu C, Aike Z, Pao X, Xiaoxia W, Yongrun C, Beifang W, Yue Z, Liaqat S, Shihua C, Liyong C et al (2020) Effects of GS3 and GL3.1 for grain size editing by CRISPR/Cas9 in rice. *Sci* 27:405–413. <https://doi.org/10.1016/j.rsci.2019.12.010>
- Zafar K, Sedeek KEM, Rao GS, Khan MZ, Amin I, Kamel R, Mukhtar Z, Zafar M, Mansoor S, Mahfouz MM (2020) Genome editing technologies for rice improvement: progress, prospects, and safety concerns. *Front Genome Ed* 2:5. <https://doi.org/10.3389/fgeed.2020.00005>
- Zainal-Abidin RA, Mohamed-Hussein ZA (2021) Computational analysis of rice transcriptomic and genomic datasets in search for SNPs involved in flavonoid biosynthesis. In: Ansari MR (ed) Recent Advances in Rice Research. IntechOpen. <https://doi.org/10.5772/intechopen.94876>
- Zeng YH, Zhu YS, Lian L, Xie HG, Zhang JF, Xie HA (2013) Genetic analysis and fine mapping of the pubescence gene GL6 in rice (*Oryza sativa* L.). *Chin Sci Bull* 58:2992–2999. <https://doi.org/10.1007/s11434-013-5737-y>
- Zhang T, Wong G (2022) Gene expression data analysis using Hellinger correlation in weighted gene co-expression networks (WGCNA). *Comput Struct Biotechnol J* 20:3851–3863. <https://doi.org/10.1016/j.csbj.2022.07.018>
- Zhang Y, Pribil M, Palmgren M, Gao C (2020) A CRISPR way for accelerating improvement of food crops. *Nat Food* 1:200–205. <https://doi.org/10.1038/s43016-020-0051-8>
- Zhang Y-M, Yu H-X, Ye W-W, Shan J-X, Dong N-Q, Guo T, Kan Y, Xiang Y-H, Zhang H, Yang Y-B et al (2021) A rice QTL GS3.1 regulates grain size through metabolic-flux distribution between flavonoid and lignin metabolons without affecting stress tolerance. *Commun Biol* 4:1171. <https://doi.org/10.1038/s42003-021-02686-x>
- Zhang JK, Yu H, Li JY (2023) domestication: retrace the history of agriculture to design future crops. *Curr Opin Biotechnol*. <https://doi.org/10.1016/j.copbio.2023.102946>
- Zhang LN, Cui D, Ma XD, Han B, Han LZ (2023) Comparative analysis of rice reveals insights into the mechanism of colored rice via widely targeted metabolomics. *Food Chem*. <https://doi.org/10.1016/j.foodchem.2022.133926>
- Zhao M, Zhai L, Tang Q, Ren J, Zhou S, Wang H, Yun Y, Yang Q, Yan X, Xing F et al (2024) Comparative metabolic profiling of different colored rice grains reveals the distribution of major active compounds and key secondary metabolites in green rice. *Foods* 13:1899
- Zheng J, Wu H, Zhu H, Huang C, Liu C, Chang Y, Kong Z, Zhou Z, Wang G, Lin Y et al (2019) Determining factors, regulation system, and domestication of anthocyanin biosynthesis in rice leaves. *New Phytol* 223:705–721. <https://doi.org/10.1111/nph.15807>
- Zhou F, Lin QB, Zhu LH, Ren YL, Zhou KN, Shabek N, Wu FQ, Mao HB, Dong W, Gan L et al (2013) D14-SCFD3-dependent degradation of D53 regulates strigolactone signalling. *Nature* 504:406. <https://doi.org/10.1038/nature12878>

- Zhu ZF, Tan LB, Fu YC, Liu FX, Cai HW, Xie DX, Wu F, Wu JZ, Matsumoto T, Sun CQ (2013) Genetic control of inflorescence architecture during rice domestication. *Nat Commun.* <https://doi.org/10.1038/ncomms3200>
- Zhu F, Wen W, Cheng Y, Alseekh S, Fernie AR (2023) Integrating multiomics data accelerates elucidation of plant primary and secondary metabolic pathways. *aBIOTECH* 4:47–56. <https://doi.org/10.1007/s42994-022-00091-4>
- Zsögön A, Čermák T, Naves ER, Notini MM, Edel KH, Weinl S, Freschi L, Voytas DF, Kudla J, Peres LEP (2018) De novo

domestication of wild tomato using genome editing. *Nat Biotechnol.* <https://doi.org/10.1038/nbt.4272>

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